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**CHARACTERIZATION OF INSERTION SEQUENCE IS605 IN
*HALANAEROBIUM HYDROGENIFORMANS***

by

MICHAEL SADLER

A THESIS

**Presented to the Faculty of the Graduate School of the
MISSOURI UNIVERSITY OF SCIENCE AND TECHNOLOGY
In Partial Fulfilment of the Requirements for the Degree
MASTERS OF SCIENCE IN APPLIED AND ENVIRONMENTAL BIOLOGY**

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ABSTRACT

Insertion sequences are the smallest prokaryotic transposable elements. These genes play a significant evolutionary role by promoting genome plasticity. Insertion sequences are highly diverse elements that have largely been uncharacterized. As such, the ability to accurately identify, annotate, and infer genomic impact of insertion sequences is lacking. The study of new insertion sequences contributes knowledge to their annotation and evolution.

Halanaerobium hydrogeniformans is a unique organism with an abnormally high number of insertion sequences. A family of insertion sequences, IS200/605, showed several interesting distinctions from other elements in the genome, including severe open reading frame degradation, and was characterized in detail. This research uses bioinformatics tools to present an in depth characterization of a single insertion sequence family in *H. hydrogeniformans*. From these results insertion sequence activity can be inferred, including transposition capability, element interaction, and insertion sequence evolution.

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1. INTRODUCTION

1.1. TRANSPOSABLE ELEMENTS

Transposable elements are mobile DNA segments capable of excision and integration within their host genome. They carry a gene encoding a transposase, which is responsible for the transposition activity, and they can carry non-transposition-essential genes known as accessory or passenger genes [1]. For some time after they were first described [2], transposable elements were thought to be junk DNA or selfish genes with little benefit to their hosts. It is now known that transposable elements play an important role in increasing genetic diversity by promoting gene duplication, genomic rearrangements, and horizontal gene transfer [3]. Additionally, transposable elements have been shown to be the most abundant and ubiquitous genes in nature [4]. Transposable elements and their fossils (relics of transposable elements that have lost their ability to transpose) can represent a large portion of eukaryote genomes (80% in maize) but make up a relatively much smaller percentage of prokaryotic genomes [5].

Transposable elements are classified by structure and mechanisms of transposition, and can be grouped into 2 classes. Class 1 transposable elements are composed of retrotransposon and retroposons, have similar structure to mRNA and retroviruses, and are usually bound by long terminal repeats. This class of transposable element transpose via an RNA intermediate. Class 2 transposable elements are composed of insertion sequences and transposases, and transpose through DNA intermediates. They typically carry terminal inverted

repeats [6]. Many eukaryotic transposons are related to prokaryotic insertion sequences, and carry a variety of passenger genes [7].

1.2. INSERTION SEQUENCES

Insertion sequences are the smallest and simplest of prokaryotic transposable elements. Insertion sequences are highly diverse in structure and organization. Insertion sequences typically have an open reading frame (ORF), terminal inverted repeats, and direct repeats. Many insertion sequences also insert preferentially within their host genome. The differences between elements with regard to these features, as well as their catalytic mechanisms for transposition, are used to categorize insertion sequences into groups and families. There are 4 major groups and 29 distinct families. It is important to note that these are general insertion sequence characteristics, and do not apply to all insertion sequence families.

1.2.1. Organization. Insertion sequences are typically between 0.8 and 2.5 kb in size and carry a single open reading frame (ORF) required for transposition. Insertion sequence ORFs can be divided into structural domains that contribute to distinct functions. The N-terminal and C-terminal regions principally contain DNA-binding and catalytic domains, respectively. This orientation permits the coupling of synthesis and activity of the transposase [8], [9]. Further evidence of the purpose of this organization is that for a number of insertion sequence families, DNA-binding domains isolated from the catalytic domains bind more readily than the whole protein. This suggests that the C-

terminal inhibits DNA binding to a degree through steric masking and provides an explanation for the preference of many transposases to act in cis (which is the preference for transposases to mobilize the gene from which is was encoded) [10].

1.2.2. Terminal Inverted Repeats. With few exceptions, insertion sequences contain terminal inverted repeats (IR). These are generally imperfect IR of 10-40 bp in length near each terminus of the transposable element outside the ORF. Inverted repeats are short sequences that read the same 5' – 3' on each strand of DNA. The outermost base pairs are involved in strand cleavage and transfer during the transposition reaction, and the internal base pairs are recognized for transposase binding [11]. Additionally, endogenous insertion sequence promoters have been located in the terminal inverted repeat sequences upstream of the transposase gene which may provide a mechanism for auto regulation. Binding sites for host specific proteins have also been observed within or close to the IR that may also impact transposition activity or transposase expression [12].

1.2.3. Target Site Duplications. Another common feature to insertion sequences is a target site duplication that is generated on insertion. Staggered DNA cuts at the target site in the DNA backbone results in the duplication of the target DNA flanking the insertion sequence upon insertion. The target site duplication results in a direct repeat (DR). The size of the DR vary between families and elements, but typically range between 2-14 bp in length [13].

1.2.4. Target Sequence. Some insertion sequences have a regional preference for insertion sites, inserting within an AT or GC rich area. Other elements require a specific sequence ranging between 4-9 nt in length. Many Insertion sequences insert within or proximal to sequences that resemble their own terminal inverted repeats. These elements often transpose with an intermediate of an IR-IR junction (including members of IS30 and IS3 families) This processes can result in a cascade of transposition events and numerous insertion sequences located proximally to one another [12]. The general structure of an insertion sequence is represented in Figure 1.1.

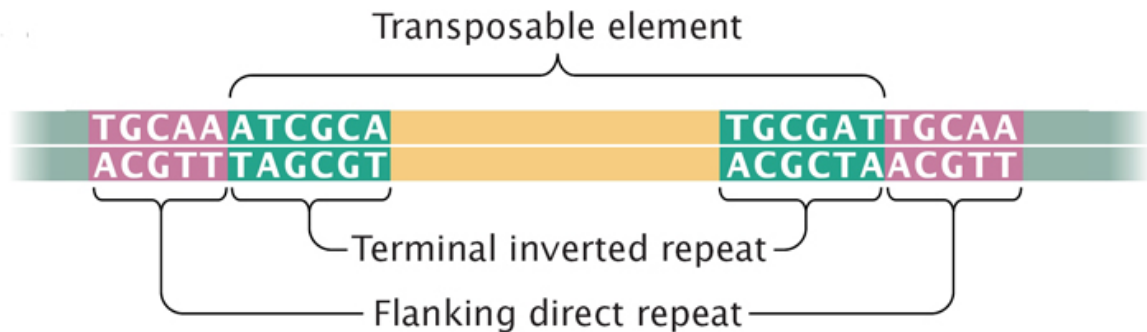


Figure 1.1. The general features of an insertion sequence containing inverted repeats and target site duplication (direct repeat) [14].

1.3. CATALYTIC CHEMISTRIES

Insertion sequences can be categorized into four groups based on their catalytic mechanisms for transposition. These groups are 1) the DDE, so called for the conserved catalytic DDE motif, 2) Y1, 3) S for their conserved tyrosine

and serine residues at the catalytic site, and 4) Y2 group that shows similarity to proteins involved in rolling circle replication.

1.3.1. DDE. The majority of identified insertion sequences fall into the DDE family of transposases. Within this group there are dominant family members, including the IS3 and IS5 [1]. DDE family members feature a triad of negatively charged catalytic residues D (asp) D (asp) E (glu) that are highly conserved. The distance between each conserved residue is variable between families and highly conserved within families containing the DDE motif. Two transposition steps are common to all reactions catalyzed by DDE family members. The first is DNA cleavage through hydrolysis and the second is the attack of target site DNA by the free 3'OH on the element end. However, these family members transpose via a double stranded intermediate. Generating the free dsDNA intermediate requires further processing of the second strand that is family specific [15]. The second strand is most commonly freed from its flanking DNA through the formation of a transient hairpin at the element end [16].

This DDE transposition mechanism has also been observed in host functions. For example, the RAG1/2 complex that catalyzes V(D)J recombination in developing lymphocytes is thought to have come from a domesticated transposase. RAG1 contains a highly conserved DDE motif [17].

1.3.2. Serine. Serine transposases are much less understood than their DDE counterparts. These transposases are encoded by the IS607 family of insertion sequences and show some similarity to serine recombinases that catalyze inversion of DNA segments [18]. Although characterized groups of

serine recombinases show an inversion of the typical DNA domain organization, serine transposases show the typical domain organization with DNA binding and catalytic domains in the N-terminus and C-terminus respectively [19]. In addition to the transposase, some IS607 family members also encode a second protein known as orfB or TnpB. This protein shows high sequence similarity to a protein encoded by members of the IS605 family. The TnpB protein is not required for IS607 transposition [20]. IS607 elements in *E.coli* systems have been shown to insert with very low target sequence specificity, which is atypical for reactions catalyzed by serine recombinases [21].

1.3.3. Y1. The Y1 transposases, which are among the smallest identified transposases (approximately 150aa in length), use a single catalytic tyrosine. These transposases are members of a greater superfamily of endonucleases known as the HUH (H = Histidine, U = hydrophobic) endonuclease family.

This HUH superfamily acts exclusively on ssDNA, catalyzing DNA breakage and formation of a 5' phosphotyrosine intermediate using the catalytic tyrosine residue and generating a free 3'OH at the cleavage site. Many HUH endonucleases recognize and bind DNA hairpin structures, cleaving ssDNA on either side of the stem or even within the hairpin structure itself [22]. These small hairpins can be identified and bound by the transposase through sequence specific recognition of the stem or loop, or through the recognition of structural irregularities in the stem [23].

Similar to the superfamily to which they belong, Y1 insertion sequences transpose via ssDNA intermediates and insert 3' to a conserved, element-

specific, penta- or tetranucleotide sequence. These Y1 family members also insert and excise preferentially from and into ssDNA [24], [25]. It is important to note that these transposable elements do not contain inverted repeats or generate target site duplications, common to the majority of identified insertion sequences.

1.3.4. Y2. Y2 insertion sequences, encoded by IS91, also fall within the HUH endonuclease superfamily. While Y1 transposases carry a single catalytic tyrosine, Y2 transposases carry two conserved tyrosine residues and appear to carry out transposition through a different mechanism. Y2 proteins also show similarities to proteins involved in rolling circle replication [1]. IS91 elements insert 3' to a conserved tetranucleotide sequence [26]. Relatively little is known about the transposition mechanisms of this family of insertion sequences compared to the more defined families.

See Figure 1.2 for the number of identified insertion sequences grouped by family and catalytic chemistry. See Figure 1.3 for the distribution of identified insertion sequences across prokaryotic genomes.

1.4. IS200/605

The IS200 and IS605 families of insertion sequences belong to the group of Y1 transposable elements briefly described in Section 1.3.3. The difference between these two families is that IS200 carries a single transposase gene (*tnpA*), while IS605 members encode a gene (*tnpB*) in addition to the *tnpA*. The *tnpB* gene is dispensable for transposition [25].

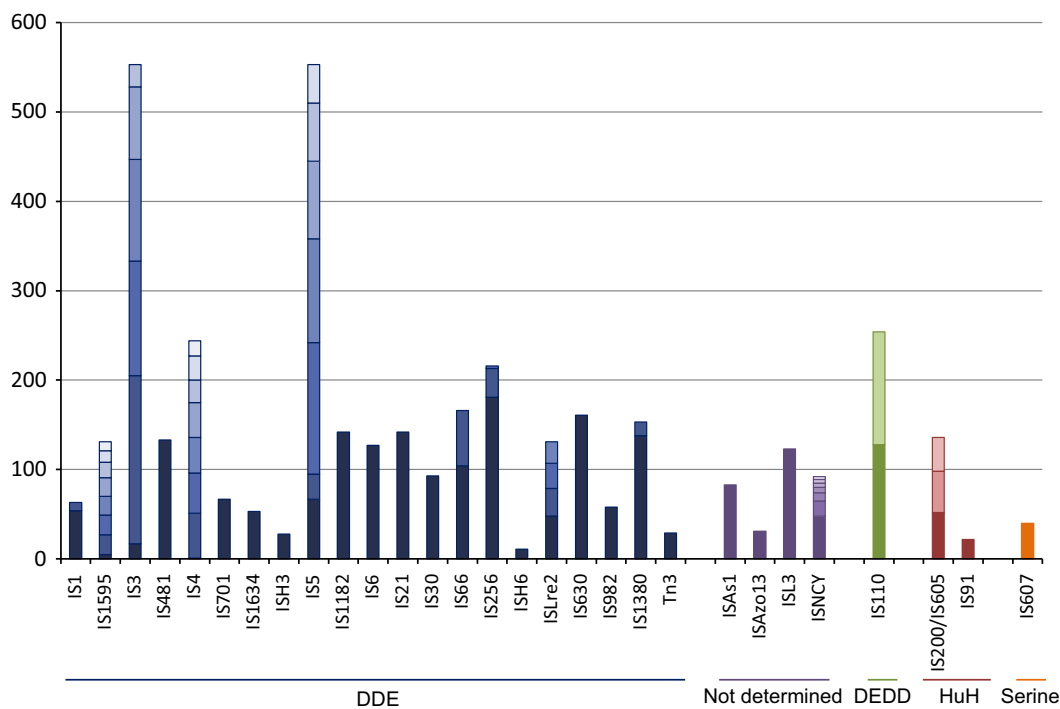


Figure 1.2. The number of identified insertion sequences grouped by catalytic chemistries and insertion sequence family [1]. DEDD represents a major subgroup of the DDE group. Shaded bars represent sub families

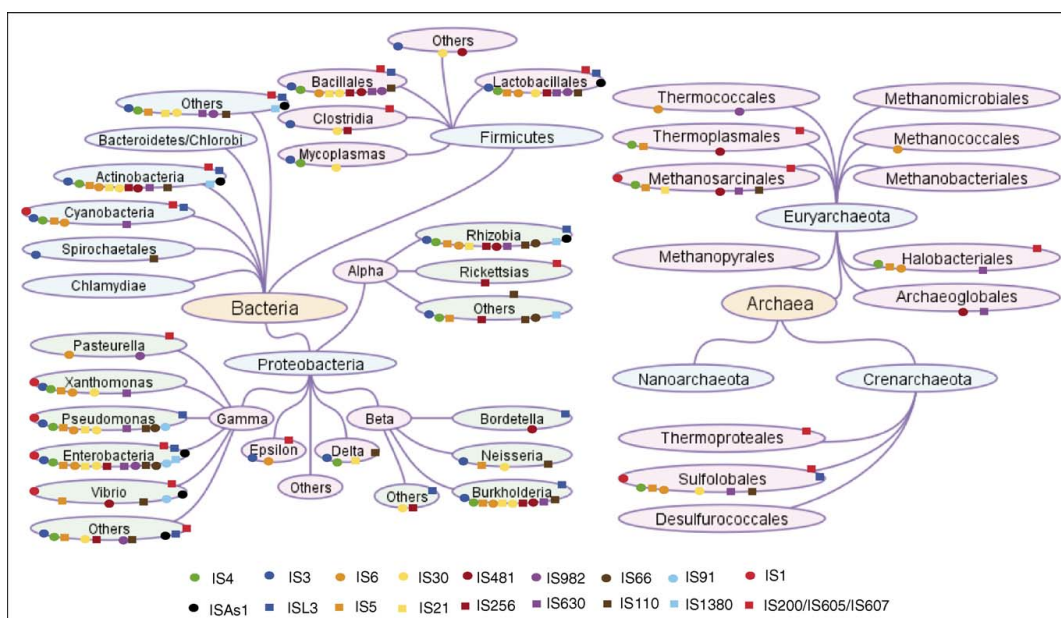


Figure 1.3. Distribution of identified insertion sequences families across prokaryotic genomes [5].

These elements do not contain terminal inverted repeats, nor do they generate target site duplications upon insertion. They contain secondary hairpin structure at both element ends that are necessary for transposition [24].

1.4.1. TnpA. The *tnpA* gene of the IS200 and IS605 families encodes the transposase. This protein contains the HUH motif and carries a single catalytic tyrosine. and inserts the element 3' to a specific tetra- or penta- nucleotide sequence [27].

The protein functions as an obligatory dimer [25]. For transposition to occur, each TnpA monomer binds an indispensable secondary hairpin structure present at each end of the element. In the well-characterized ISHp608 elements of the IS200/605 family, the sequence of these structures is the same at the left end (LE) and right end (RE) of the element [28], [24]. Transposition of the element occurs as circular ssDNA, and is strand specific [29]. The ability to differentiate between top and bottom strands in ISHp608 comes from a minor structural abnormality between the top and bottom strand [28]. Because this element transposes via a ssDNA intermediate, its transposition is coupled to the replication of host DNA, during which it has a preference for lagging strand template insertion [30]. These elements also have increased transposition rates with DNA repair mechanisms that produce large stretches of ssDNA [31].

The obligatory dimer of two TnpA proteins forms two functional conformations, a cis and a trans formation. In trans, the catalytic site is constructed of the HUH motif from one monomer, and the catalytic tyrosine from the other [32]. Conformation change from trans to cis results in strand breakage

and the formation of 2 phosphotyrosine bonds, The reverse conformation change results in the insertion of the element into the target site [23], [33], [34].

Insertion into a new location starts with target recognition. Recognition is a result of DNA-DNA interaction of a tetranucleotide sequence 5' to the LE hairpin structure and a target sequence. The target sequence is dependent on the sequence 5' to the hairpin structure. When the active tetranucleotide sequence of the element was altered, new insertion sites were targeted [32]. Insertion occurs without target site duplication, and element excision precisely seals donor DNA. This transposition does not require host cell DNA repair factors [25].

1.4.2. TnpB. An ORF, known as orfB or tnpB is often encoded proximal to the Y1 tnpA of IS200. When together, they represent the IS605 family. For these family members, hairpins necessary for element transposition are found external to the two ORFs. OrfB is approximately 1200 nt in length and is dispensable for transposition [25]. OrfB is located in successive, divergent, or overlapping orientation with respect to tnpA [24]. Until recently the function of TnpB was largely unknown. There is now evidence to suggest that TnpB plays a role in transposition regulation of IS200 and IS605 elements. TnpB has been shown to decrease both excision and insertion of TnpA, decreasing excision approximately 200 times more efficiently than insertion [35]. The mechanism of how TnpB inhibits transposition is unknown but it is speculated that TnpB protein could competitively bind the hairpin structures at either end of the element, or bind the TnpA protein itself. TnpB directly impacts the activity of TnpA and does not act through host mediated factors [35].

The TnpB polypeptide typically contains 3 domains, an N-terminal HTH, a central domain, and a C-terminal zinc finger. TnpB is most variable in the N-terminal and most conserved in the C-terminal domain. The inhibitory action of TnpB on TnpA transposition is strictly dependent on the integrity of the zinc finger domain [35]. Zinc fingers perform a broad range of functions, primarily as interaction modules binding to a wide variety of nucleic acids, proteins, and other molecules [36].

The TnpB protein has been associated with members of the IS607 family. This family utilizes a different mechanism of transposition than the IS200/605 families. When associated with the transposases of IS607, *tnpB* is dispensable for transposition [20]. Additionally, homologues of *tnpB* are found in diverse eukaryotic transposable elements [37].

Several reported elements encode *tnpB* as the only ORF. This has resulted in the labeling of TnpB as a putative transposase gene (IS1341, IS809, and IS1136). However, the evidence for TnpB mediated transposition is absent and it is likely that these elements are non-autonomous derivatives of IS605 or IS607 families. See Figure 1.4 for a representative structure of a IS605 family member with divergent ORFs.

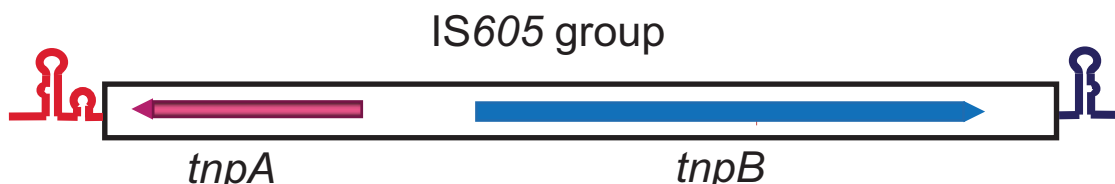


Figure 1.4. An illustration of an IS605 family member with divergent ORFs [1].

1.5. MINIATURE INVERTED REPEAT TRANSPOSABLE ELEMENTS

Miniature inverted repeat transposable elements (MITES) are partial copies of transposable elements that typically contain only the sequence or structures necessary for transposition. MITES can be impactful to host genomes as they can influence gene expression, alter mRNA stability, or influence transcription termination. In genomes without full length parent copies it can be extremely difficult to identify MITES, as they are often only present as short inverted repeat sequences [5]. MITES represent evidence of past insertion sequence activity and are important for understanding the evolution of insertion sequences within the host and the impact of insertion sequences on the genome.

1.6. INSERTION SEQUENCE ANNOTATION

Insertion sequences, their nonautonomous derivatives, and MITES, represent a substantial portion of bacterial and archeal genomes. Insertion sequences are highly diverse with respect to their transposases and element ends. Because of this diversity accurate insertion sequence identification and annotation is difficult. Transposase genes of insertion sequences are often mislabeled as integrases, recombinases, pseudogenes, and hypothetical proteins [38]. The element ends containing inverted repeat and direct repeats, which are smaller and more diverse than the proteins themselves, are typically overlooked. It is even more rare that MITES are identified.

The development of high throughput sequencing has led to the generation of thousands of complete genomes and metagenomes. With the sheer quantity

of insertion sequences and MITES, accurate identification and annotation requires more sophisticated methods than those currently available [39].

Several semi-automatic methods have been developed to aid in the identification of insertion sequences. Two of these are OASIS (Optimized Annotation System for Insertion Sequences) [40], and ISSaga (Insertion Sequence Semi-Automatic Genome Annotation) [38]. Enhanced methods to better visualize and organize these elements are being developed [41]. The underlying issue with these methods is that they rely on the quality of insertion sequence database libraries. While these methods expedite the identification of known insertion sequences, unique insertion sequences and MITES can be misidentified or completely overlooked. Even MITES of known and well characterized insertion sequences can be overlooked because they show such low similarity to the parent element, or a parent element may not be present in the genome being surveyed.

The inability to accurately identify insertion sequences and MITES leads to a severe bias towards characterized and complete copies of insertion sequences in surveys of insertion sequences across genomes [38]. Until more sophisticated methods are developed, or insertion sequence databases become more complete, a significant amount of manual curation is necessary when identifying and annotating insertion sequences and their MITES.

1.7. GENOMIC IMPACT OF TRANSPOSABLE ELEMENTS

Transposable elements were originally viewed as selfish DNA, serving little to no purpose to their host. However, it is now understood that transposable elements play a significant role in promoting genetic diversity, structure, and genomic plasticity [42].

1.7.1. Genomic Streamlining. Insertion sequences experience rapid expansion and loss within host genomes. This is accompanied by genomic rearrangement, and gene inactivation. With time, insertion sequences experience deletion that can be accompanied with deletion of host DNA, resulting in genome reduction. Insertion sequence degradation will lead to the development of non-functional, or non-autonomous elements, which are eventually cleared from the genome. The increase in transposable elements and reduction in genome size is most noted in new bacterial endosymbionts, and is only permissible with an increase in host dependence [5]. The relaxed selective pressure of new endosymbionts permits both the expansion of insertion sequences, and the ensuing genome reduction.

It has been observed that transposable element numbers increase in new bacterial endosymbionts compared to free living cells [43], and that genomic reduction is correlated with insertion sequence expansion. This is evident in comparing three *Bordetella* species *B. pertussis*, *B. parapertussis*, and *B. bronchiseptica*. The genome size of *B. bronchiseptica* is the largest of the three (5.34 Mb) and it harbors no insertion sequences, *B. parapertussis* has a reduced genome size (4.77 Mb) with over 100 insertion sequences, and *B. pertussis* with

the smallest genome size (4.1 Mb) has over 260 identified insertion sequences. The phylogeny of the of the organisms suggested that *B. bronchiseptica* was the ancestral species of the three [44].

1.7.2. Insertional Mutation. Insertion sequences can effect genomes through direct impedance by inserting into and disrupting genes. Insertion sequence mediated disruption in a *Rickettsi* species resulted in non-pathogenicity by insertion into virulence genes [45], as well as a metronidazole resistant *H. pylori* by insertion within a gene necessary for pro-drug activation [46].

1.7.3. Gene Expression. Although over 80% of genes in prokaryotic genomes encode proteins, not all insertion events cause a direct disruption. Insertion into intergenic regions can still impact the host genome. Some mobile elements carry endogenous transcriptional promoters [12], and their insertion leads to changes in expression of flanking genes. Insertion sequences can also change expression by activating or inactivating promoter or repressor genes [47].

1.7.4. Genomic Rearrangement. Insertion sequences also impact genomes through a variety of chromosomal architecture changes. This activity stems from the multiple copies of elements with high sequence similarity. Recombination between two IR of a single insertion sequence can result in an inversion. Direct inversion of elements carrying endogenous promoters has been shown to increase pathogenicity through phase variation in a number of organisms [48], [49]. Recombination can also occur between elements resulting in the inversion of the entire sequence between the elements, or in the deletion

of sequence between the elements [50]. Alternative transposition mechanisms can also result in inter-element sequence duplication [6].

1.8. INVASION – EXPANSION – EXTINCTION CYCLES

To a degree, insertion sequences provide a selective advantage to their host by increasing diversity and genomic plasticity [51]. However, insertion sequences are in general thought to be more damaging than beneficial to their host, and their persistence in genomes is questioned. It is hypothesized that insertion sequences undergo periodic invasion, expansion, and extinction cycles. These cycles are characterized by introduction to a new genome through horizontal gene transfer, expansion through replicative transposition, and extinction through unknown methods that eliminate, or degrade insertion sequences beyond recognition in a genome.

Insertion sequences have an extremely high, nearly identical, sequence similarity within genomes [52]. This unusually high sequence similarity is not due to evolutionary constraints, as insertion sequences between genomes show significant sequence divergence. Gene conversion, which is the homogenization of nearly identical sequences through recombination, has been proposed to be a mechanism for sequence conservation in obligate mutualistic endosymbionts [53]. However, evidence of gene conversion in insertion sequences of free living hosts is absent. Insertion sequences also show higher sequence conservation than gene duplicates, which would be subject to the same level of gene

conversion. Additionally, successful transposition rates of insertion sequences are higher than substitution rates [54].

This leads to the hypothesis that insertion sequences are newly acquired to most genomes, and that they invade and rapidly expand within a genome. The patchy distribution of insertion sequences across genomes of highly related strains [55], [56], supports this hypothesis, and also shows that insertion sequences are not sustainable within a genome. Insertion sequences are selected against over time through down regulation of transposition, excision, and the preference for the majority to act in cis. The expansion of insertion sequences is permitted only because of the temporary benefits they might provide through genomic rearrangement and transfer of beneficial genes. As such, their persistence in the environment is dependent on horizontal gene transfer [54].

1.9. HALANAEROBIUM HYDROGENIFORMANS

Halanaerobium hydrogeniformans is an extremophile isolated from a haloalkaline lake in Washington State. This organism has gained attention due to its unique metabolic capabilities and potential for industrial applications. After the sequence of the genome was determined, 2463 genes were annotated. Among them were 72 transposase genes belonging to eight insertion sequence families [57]. This puts the bacterial genome at approximately 3% transposable elements, which is higher than in most bacterial genomes [5]. Because transposable

elements are often misidentified, it was suspected that 72 transposable elements was a conservative estimate of the actual number encoded in the genome.

1.10. DATABASES AND BIOINFORMATIC TOOLS

1.10.1. NCBI. The National Center for Biotechnology Information (NCBI) was developed by the National Institutes of Health (NIH) after the need for computerized information processing in modern research was realized. NCBI's mission became "finding new approaches to deal with the volume and complexity of data in providing researchers with better access to analysis and computing tools to advance understanding of our genetic legacy and its role in health and disease." Data from the European Molecular Biology Laboratory (EMBL) and the DNA Database of Japan (DDBJ) is shared with NCBI. NCBI is also host for numerous automated DNA and protein tools such as blastp, blastn, RefSeq, and ORF-finder. NCBI also provides access to DNA and protein sequences, mapping, structural data, and phylogenetic outputs [58].

1.10.2. EBI. The European Bioinformatics Institute (EBI) is part of the EMBL and provides the most up-to-date and comprehensive range of basic research and computational biology tools for researchers in academia and industry. The data and much of the software from EBI can be downloaded and installed locally, or run via online servers. The tools provided span DNA/RNA alignments, molecular structures, protein sequences, families, and motifs, taxonomy, and systems pathways [59].

1.10.3. PFAM. Pfam is a database containing protein families. Protein families are sets of proteins that share regions of high amino acid sequence similarity that are generated from multiple sequence alignments and hidden Markov models. These conserved regions can be used in the prediction of protein functionality when compared to known proteins [60].

1.10.4. Phylogeny.fr. Phylogeny.fr provides free web based phylogenetic analysis tools for the non-specialist. It permits automated and semi-automated phylogenetic relationships to be constructed between nucleotide or protein sequences using a multiple alignment process and can provide a newick output for various tree viewers [61].

1.10.5. ISfinder. ISfinder is an online public database providing general features (size, target sequence, family, inverted repeat sequences) for insertion sequences isolated from bacteria and archaea. They rely on the scientific community to deposit sequences and information of characterized insertion sequences to enrich the database. ISfinder also provides a program ISbrowser that can be used to view identified and predicted insertion sequences in sequenced genomes [41], [62].

1.10.6. ISsaga. ISsaga is a tool of ISfinder that was developed to accurately identify and annotate insertion sequences with the use of a high-quality semi-automatic annotation system. This uses the ISfinder database to provide general prediction and annotation tools for potential insertion sequences in a genome. It provides genomic context of individual insertion sequences, visual display of genomic positions, and a small array of tools to find element

ends, target site duplications, and inverted repeats. Because the annotation accuracy of ISSaga is limited to the insertion sequence library of ISfinder, insertion sequences predicted by ISSaga have to be confirmed manually before being added to the ISfinder database [38], [63].

1.10.7. ExPASy. The Swiss Institute of Bioinformatics (SIB) has developed the Expert Protein Analysis System (ExPASy) web portal, offering access to numerous scientific resources, databases, and software tools. These tools are for areas of biology research including proteomics, genomics, phylogeny, structure, and more [64].

1.10.8. Sequence Alignment. Sequence alignments are made to determine the relatedness between two or more DNA or protein sequences. The services provided by the EBI offer programs for pairwise and multiple sequence alignment. Pairwise alignments are ideal for highlighting regions of similarity or dissimilarity that may confer a functional, structural, or evolutionary relationship between two sequences. These programs would include Needle, Stretcher, Water, Matcher, and LALIGN. The differences in these programs is that they utilize slightly different parameters to align sequences. Multiple sequence alignments are used to determine homology and evolutionary relatedness between sequences. These include Clustal Omega, an alignment program for three or more sequences [59].

1.10.9. Mfolds. DNA and RNA can contain secondary structure that is functional in a variety of biological processes. Mfolds and UNAFold are free web

based programs developed to identify possible secondary structure and predict under what conditions they might form [65].

1.10.10. Argo. Argo is a Java based genome browser developed by The Broad Institute for viewing and annotation of whole genomes. It displays the sequence and annotation of DNA tracks. Files can be uploaded in SAM/BAM, FASTA, Genbank, GFF, BLAST, BED, WIG, and Genscan formats. This program is useful in determining relative position to other genes, as well as extracting DNA and protein sequences for further phylogenetic or structural analysis [66].

1.11. SUMMARY

This thesis presents a detailed characterization of an IS200/605 family members within *H. hydrogeniformans*. This family was selected for detailed characterization because of the unique characteristics of Y1 transposases. Six Y1 elements were originally annotated in the genome. After investigation this number rose to 23 elements and 1 MITE. Many of the 605 elements were misidentified by insertion sequence annotation software, and exhibit unique disruptions and fragmentation not typically observed in insertion sequences. The phylogeny of these elements in comparison to their structural differences suggests recombination between elements is occurring. These elements differ from reported IS200/605 family members in that their element ends are unique, and do not share common sequence between the right and left ends. This work is a detailed survey of an IS605 family of elements not reported elsewhere and provides a look at how insertion sequences might degrade within host genomes.

2. MATERIALS AND METHODS

2.1. INSERTION SEQUENCE IDENTIFICATION

The *Halanaerobium hydrogeniformans* genome sequence is recorded at the National Center for Biotechnology Information (NCBI), accession number CP002304.1. All genes annotated as insertion sequence, transposase, and integrase were used for a BLAST search against Genbank to determine potential products. The results were used as a query against the ISfinder library to confirm insertion sequence identity. After confirmation, a representative open reading frame (ORF) from each different insertion sequence group was used for a BLAST search against the *H. hydrogeniformans* genome to identify partial insertion sequences that were annotated as pseudo or hypothetical genes. Insertion sequences in the genome were then identified with ISSaga to compare the identity results from manual and semi-automatic library based methods. ISSaga scans for insertion sequences in annotated genomes by comparing potential sequences against the ISfinder database. It then performs a blastn for replicons within the genome to identify partial elements or potential mobile elements not originally annotated.

The elements belonging to the Y1 family were chosen for further investigation due to the numerous members present in the genome. This family was also chosen because of its distinct characteristics and the significant sequence dissimilarities between their replicates. Dissimilar replicates are

inconsistent with reported high sequence similarity of insertion sequence between members of the same family within a bacterial genome [52].

The element families that were investigated in detail were given loci numbers for organization and further reference. Loci numbers were sorted 1-23 moving 5'-3' from the origin of replication on the + strand.

2.1.1. Element Ends. The ends of a Y1 insertion sequence extend beyond the ORF. The element ends are defined as the nucleotide sequences of the element outside the ORF. These were identified by extracting 1000 nucleotides 5' and 3' of each ORF and aligning to identify the extent of homology between elements.

2.1.2. MITES. Miniature Inverted Repeat Transposable Elements (MITES) were identified by querying the genome with the element ends. Identified ends were matched with their corresponding ORFs. Element ends without corresponding ORFs were marked as potential MITES and examined further.

2.2. GENOME BROWSER

The Argo Genome Browser was used to visualize the genome of *Halanaerobium hydrogeniformans*. The genome was uploaded into Argo in Genbank format. Genes of interest were marked and categorized for further use. Visualization of gene positions allowed for a preliminary survey for insertion sequence position and proximity patterns. The genome browser was used to extract nucleotide and conceptual protein sequences for phylogenetic and alignment uses [66].

2.3. BLAST

Chosen sequences are aligned against a target database using a Basic Local Alignment Search Tool (BLAST). Databases can be queried with protein or nucleotide sequences.

For blastp, a conceptual protein sequence is used to query a protein database. This is used to identify potential gene products and conserved domains. Megablast is used to query a nucleotide sequence for closely related sequences for identification, working best if sequences show a 95% or higher similarity. Megablast was used to identify insertion sequence replicates within the genome. Discontiguous megablast is similar to megablast but allowing for greater mismatches and is intended for sequences with low similarity and cross-species comparisons. Discontiguous megablast was used to search for insertion sequence replicates that were misidentified or not annotated. Blastn is slower than megablast and discontiguous megablast but allows a word-size of seven bases. This permits the comparison of short sequences with low similarity. Blastn was used to search for MITES and element fragments against the genome. These BLAST tools are available free for use at the National Center for Biotechnology Information (NCBI). Algorithm parameters for BLAST searches used are in Table 2.1

2.4. ALIGNMENTS

Alignments were made between two or more protein sequences or two or more nucleotide sequences. Alignments are useful in comparing sequence

similarity and structural differences. A number of alignment programs were used for pairwise and multiple sequence alignments. EMBOSS Needle and EMBOSS Stretcher utilize a Needleman-Wunsch algorithm to search for optimal global alignment between two sequences.

Table 2.1. Algorithm parameters for BLAST searches.

BLAST	Blastp	Megablast	Discontinuous Megablast	Blastn
Max Target Sequences	100	100	100	100
Expect Threshold	10	10	10	10
Word Size	6	28	11	11
Max Matches	0	0	0	0
Match/Mismatch	N/A	1, -2	2, -3	2, -3
Scoring Matrix	BLOSUM62	N/A	N/A	N/A
Gap Cost	Existence: 11 Extension: 1	Linear	Existence: 5 Extension: 2	Existence: 5 Extension: 2

Stretcher uses modifications that permit larger sequences to be globally aligned. LALIGN is a program for pairwise sequence alignment optimized for local alignment between two sequences [67]. Clustal Omega and Kalign are programs used to globally align multiple sequences [68]. All alignment programs are freely

available for use from the European Bioinformatics Institute. Parameters and options used for alignment programs are found in Table 2.2 and Table 2.3.

Table 2.2. Alignment options for Clustal Omega.

Program	Clustal Omega
Dealign Input Sequences	NO
Clustering Guide Tree	YES
Clustering Iterations	YES
Combined Iterations	0
Tree Iterations	Default
HMM Iterations	Default

2.5. OPEN READING FRAME DISRUPTION

Insertion sequences can insert within genes disrupting the ORF.

Automated identification of disrupted genes can be difficult. To identify if any of the Y1 insertion sequences inserted within a gene, 1000 nucleotides on either side of the insertion sequence (-1000/+1000) were extracted and spliced together. The 2000 nucleotide sequence frame was then searched with ORF Finder, a tool freely available for use from NCBI. Any ORF extending through position 0 (the middle of the extracted sequence) of the constructed ORF was

conceptually translated and subjected to a blastp search against the NCBI database to identify potential protein products.

Table 2.3 Alignment parameters and options for pairwise alignment programs.

Program	Kalign	Needle	Stretcher	Water	Matcher	LALIGN
Gap Open	80	10	16	10	16	-12
Gap Extension	3	0.5	4	0.5	4	-4
Terminal Gap	3	NA		NA	NA	NA
Bonus Score	0	NA	NA	NA	NA	NA
Matrix	N/A	DNAfull	DNAfull	DNAfull	DNAfull	(+ 5) / (- 4)
End Gap Penalty	NA	FALSE	NA	NA	NA	NA
End Gap Open	NA	10	NA	NA	NA	NA
End Gap Extension	NA	0.5	NA	NA	NA	NA
Alternatives Matrix	NA	NA	NA	NA	1	NA

2.6. PHYLOGENETIC ANALYSIS

Phylogenetic analysis was conducted with Phylogeny.fr. Extracted nucleotide sequences from insertion sequence ORF were input in FASTA format. Relationships of sequences were made using a MUSCLE sequence alignment without Gblock curation, and a maximum likelihood phylogenetic tree construction. Phylogenetic analysis was performed with the “one click” option for speed and alignment optimization [61], [69]. Mobile Elements in the genome showing significant deterioration were excluded from phylogenetic analysis, as

the nucleotide sequences of these elements were too short to construct an accurate phylogenetic relationship.

2.7. SECONDARY STRUCTURE IDENTIFICATION

External to the ORF are conserved insertion sequence ends. In Y1 elements these ends contain hairpin structures necessary for transposition. Regions of the element ends showing potential for hairpin formation were identified by aligning the element left and right end nucleotide sequence with its respective reverse complement. The pairwise alignment program LALIGN was used to scan for regions with emphasis on local alignment. Regions showing significant alignment to their reverse complement were visually identified and subsequently examined with Mfolds, a DNA folding program, to view the potential physical structures. Mfolds DNA folding form was used under default conditions [65], [70].

3. RESULTS

3.1. INSERTION SEQUENCE IDENTIFICATION

ISSaga identified 16 insertion sequence families in *Halanaerobium hydrogeniformans*. Initial observations reveal that these families are composed of few individual elements with varying levels in copy number.

Manual curation identified fewer families, with approximately the same number of total insertion sequences. Of note, ISSaga identified the presence of IS200/605, IS1341, and IS607 family members. In contrast, manual annotation resulted in the identification of one IS200 family member, and 22 IS605 family members. After detailed characterization, it was discovered that ISSaga misidentified these elements as there were no elements belonging to the IS1341 or IS607 families in *H. hydrogeniformans*. All misidentified elements showed high sequence similarities to the IS605 members. Table 3.1 presents the number of unique insertion sequences per family and the total number of elements belonging to that family as identified by ISSaga. Insertion sequence families IS1341, IS605, and IS607 are highlighted.

Detailed characterization of insertion sequences in *H. hydrogeniformans* was limited to the IS200 and IS605 family members. Each identified insertion sequence was given an independent locus number corresponding to its relative position to other detailed insertion sequences and the origin of replication. The elements are labeled locus 1-23 with increasing distance from the origin of

replication. The locus numbers for each element, as well as some of the elements characteristics which are further discussed, are outlined in Table 3.2

Table 3.1. Insertion sequences in *H. hydrogeniformans* as identified by ISsaga.

Family	Unique IS	Total IS
IS200_IS605_ssgf_IS1341	1	5
IS3_ssgf_IS407	1	3
IS3_ssgf_IS3	4	4
IS6	2	7
IS607	2	15
ISNCY_ssgf_IS1202	1	4
IS256	4	14
ISNCY	1	2
IS30	3	12
IS3_ssgf_IS150	3	16
IS200_IS605	2	4
IS1182	2	2
IS21	2	3
IS3_ssgf_IS51	1	8
IS3	1	8
IS110	1	1
Total	31	108

3.2. TnpA

There exist two different tnpA open reading frames (ORF). One belonging to an IS200 (locus 07), Accession number ADQ14068.1, in which it is the sole product of the insertion sequence.

Table 3.2. Characteristics of IS200 and IS605 elements in *H hydrogeniformans*.

Locus	tnpA type	tnpB	LE	RE	tnpA halsa	tnpB halsa	Leading/Lagging (tnpA)
1	Type 2	1B	consensus	type 2	Halsa_0245	Halsa_0244	Lead
2	Type 5	2C	consensus	type 1	N/A	Halsa_0258	Lead
3	Type 5	2C	consensus	type 1	N/A	Halsa_0296	Lag
4	Type 5	2A	consensus	type 2	N/A	Halsa_0322	Lead
5	Type 5	1A	consensus	type 1	N/A	Halsa_0445	Lead
6	Type 5	2A	consensus	type 2	N/A	Halsa_0509	Lag
7	IS200_TnpA	N/A	unknown	unknown	Halsa_0613	N/A	Lag
8	Type 5	1A	consensus	type 1	N/A	Halsa_0624	Lag
MITE	NA	NA	Hairpin	type 2	NA	NA	Lag
9	Type 1	2A	consensus	type 1	Halsa_0741	Halsa_0742	Lag
10	Type 5	1A	consensus	type 1	N/A	Halsa_0809	Lag
11	Type 5	3	consensus	type 1	N/A	Halsa_0886	Lag
12	Type 5	3	consensus	type 1	N/A	Halsa_1064	Lag
13	Type 3	2A*	consensus	type 1	Halsa_1089	Halsa_1090	Lag
14	Type 5	MISC	consensus	type 2	N/A	Halsa_1216	Lead
15	Type 4	3*	consensus	MISC	Halsa_1228	Halsa_1227	Lead
16	Type 5	1A	consensus	type 1	N/A	Halsa_1236	Lag
17	Type 5	2B	consensus	type 1	N/A	Halsa_1482	Lag
18	Type 5	2B*	consensus	type 1	N/A	Halsa_1629	Lead
19	Type 5	2B	consensus	type 1	N/A	Halsa_1739	Lag
20	Type 2	2B	consensus	type 1	Halsa_2178	Halsa_2179	Lead
21	Type 5	3	consensus	type 2	N/A	Halsa_2207	Lead
22	Type 5	3	consensus	type 1	N/A	Halsa_2220	Lead
23	Type 5	3	consensus	type 1	N/A	Halsa_2306	Lag

The other *tnpA* belonging to the IS605 members, accession number WP_013405283.1, of which there are 22 complete, partial, or fragmented copies. Each TnpA protein contains a single Y1_Tnp superfamily domain. These will be referred to as the IS200 *tnpA*, and the IS605 *tnpA*. A protein alignment of each TnpA type is shown in Figure 3.1.

```

#=====
#
# Aligned_sequences: 2
# 1: IS200_TnpA
# 2: IS605_TnpA
# Matrix: EBLOSUM62
# Gap_penalty: 12
# Extend_penalty: 2
#
# Length: 134
# Identity:      62/134 (46.3%)
# Similarity:   87/134 (64.9%)
# Gaps:         15/134 (11.2%)
# Score: 290
#
#=====
IS200_TnpA      1 MSNQLDSNRHAKYNLIYHLVVVTKFRKECISDNMYSDLNKHFKRLEGGK  50
  |...|::|. |::|:|. ||||:|:|:|. ||||:..|..:|. |. |::|.
IS605_TnpA      1 MDRDLNNNYHSVYSLQYHLVVI TKYRHECITFEMLEELEKIFTRLLKDKV  50

IS200_TnpA     51 CNLLEFGGEKDHIVMFSTPPQVQLSKVLNSLKTSTSR LIRRDYGDY LKD  100
  ||:| ||| ||| | | :|:|. | ||| ||| | | :|:|. | ||| | | :|:|. |..:|..:| |.
IS605_TnpA     51 CNVLEFGGEKDHSVHILFETPPQVQLSKLVN ILKTVSSRLIKKQYEHHLKK  100

IS200_TnpA     101 FYLK-----NISGQEVIVLCV FV----K      119
   :|. |           :..|.....:..: |
IS605_TnpA     101 YYWKPAFWSRSY CILSTGGAT IETIKKYIENQNK  134

```

Figure 3.1. IS200 TnpA and IS605 TnpA alignment.

3.2.1. IS200 *tnpA*. The IS200 *tnpA* ORF is 360nt long, consistent with other reported IS200 family members. Because this insertion sequence occurs without replicates, does not produce target site duplications, or contain inverted

repeats, the element ends could not be identified. An attempt to identify secondary structures was made by aligning the nucleotide sequence on either end of the ORF with each other and each end with its reverse complement (Appendix A). However, regions showing significant alignment could not be identified above background levels. Additionally, it is unknown if any sequence showing alignment was part of the element ends.

3.2.2. IS605 tnpA. There exist 5 sub-types of the IS605 tnpA, as characterized by ORF structural differences, for a total of 22 individuals. Each subtype has a complete or partial divergent tnpB ORF. The 5 subtypes are described below and visualized in Figure 3.2 where blocks and triangles indicate 5' and 3' orientation. Full sequence alignments of all IS605 tnpA types are found in Appendix B.

Type 1 IS605 TnpA is a single replicate at locus 09 and is 405nt in length. This is the only 605 tnpA that could produce a functional protein as types 2-5 show significant degradation in the ORF.

Type 2 IS605 tnpA has two replicates (loci 1 and 20). These ORFs align with the most 3' 234 nucleotides of type 1, and are the missing 171 nucleotides from the 5' end.

Type 3 IS605 tnpA exists as a single replicate at locus 13. Type 3 ORF is missing 171 nucleotides from the 5', 114 nucleotides from the 3' end, and aligns with the central 120 nucleotides of type 1.

Type 4 IS605 tnpA also occurs as a single replicate at locus 15, aligning with the most 3' 108 nucleotides of type 1.

Type 5 IS605 *tnpA* is the most commonly occurring with 17 individuals. It is also the most fragmented of the five types. Opposed to types 1-4, type 5 IS605 *tnpA* does not annotate as a pseudo or hypothetical gene by genomic annotation software or by ISSaga. This type is 122 nucleotides long, aligning with the most 5' 63 nucleotides and the most 3' 59 nucleotides of type 1.

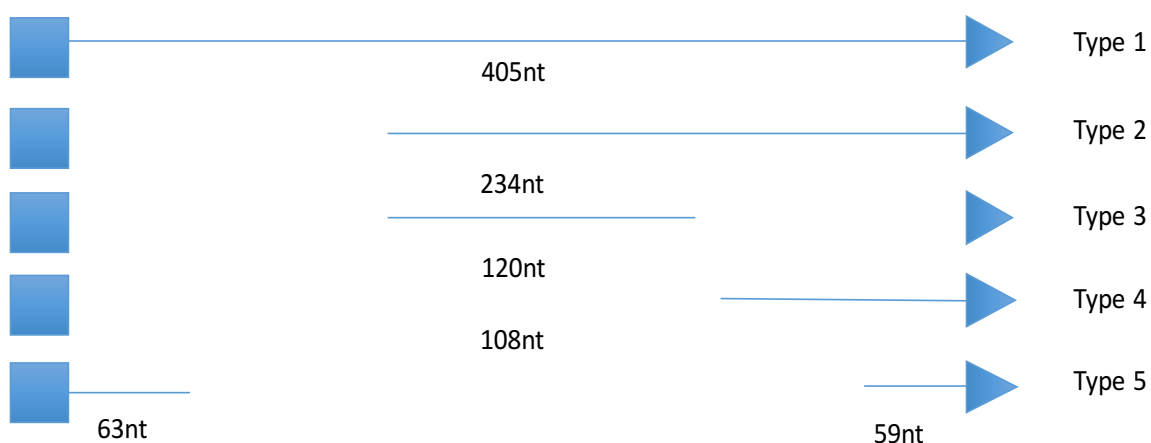


Figure 3.2. Relative IS605 *tnpA* sequence structures.

3.3. IS605 *tnpB*

There are 9 different 605 *tnpB* open reading frames present in the genome totaling 22 copies, each with a corresponding complete/partial/fragmented 605 *tnpA* (see table 3.2 for *tnpA*/*tnpB* pairings). These 9 different 605 *tnpB*s can be sorted into 3 primary groups and one miscellaneous group. These groups are described below and can be visualized in Figure 3.3 where blocks and triangles indicate 5' to 3' orientation. Full sequence alignments for all *tnpB* types are found in Appendix C

Type 1A tnpB has 4 replicates (loci 05, 08, 10, and 16). This ORF is 1254 nucleotides in length. This is not the most commonly occurring tnpB but it is the ORF most likely to produce a functional protein as types 2-3 and the miscellaneous group are sufficiently disrupted. This ORF encodes a protein containing three domains, a large ORFB_605 superfamily domain, a 605 central region, and a terminal Zn-ribbon binding domain. The element at locus 08 has inserted into and disrupted a sigma 54 interacting domain containing protein.

Type 1B tnpB is a single copy (locus 01) that aligns with type 1A ORFs. However, it contains a single nucleotide insertion at position 465 resulting in a frame shift and early translation termination.

Type2A tnpB has 3 replicates (loci 04, 06, and 09) and is 1382 nucleotides in length. These tnpB sequences align with type 1A ORFs with the exception of 2 additional 64 nucleotide inserts at position 433 and 1064. These inserts will be referred to as the left insert (LI) and right insert (RI) respectively.

Type 2A* tnpB is a single replicate (locus 13) and aligns with type 2A ORFs. It is classified as a type2A because it contains both LI and RI. It is denoted as a 2A* because it also is missing 173 nucleotides starting at nucleotide position 151.

Type 2B tnpB has 3 replicates (loci 17, 19, and 20) and has an ORF of 1318 nucleotides in length. This ORF aligns with type 2A tnpB but only contains the LI.

Type 2B* tnpB is a single replicate (locus 18) and has the same ORF and LI as type 2B ORFs. This element is denoted separately from type 2B because

the ORF is disrupted by an insertion sequence 2.6kb in length. This sequence was identified manually and by ISsaga as a IS21 family member. Extraction of this element reveals that the remainder of the ORF aligns with other type 2B ORFs. Interestingly, this putative IS21 mobile element occurs in 3 replicates and is proximal to an IS605, IS256, IS200, and IS3.

Type 2C *tnpB* occurs in 2 replicates (loci 02, and 03) and is 1318 nucleotides in length. This *tnpB* aligns with type 2A ORFs with the exception that it contains only the RI.

Type 3 *tnpB* has 5 replicates (loci 11, 12, 21, 22, and 23) and is 724 nucleotides in length. This element aligns with type 2A ORFs with the exception that it contains a hybrid insert (HI) at position 433 and is missing the 463 nucleotides that exist between the LI and RI of type 2A. These inserts are further discussed in Section 3.4.

Type 3* *tnpB* is a single replicate (locus 15) and is classified as a type 3 *tnpB* because of its hybrid insert and absence of an interior sequence. This element is denoted separately from other type 3 ORFs as it is in a more progressed state of deterioration than the other type3 *tnpBs*. It totals 499 nucleotides in length, lacking a 173 nucleotide sequence at position 146, and a 52 nucleotide sequence at position 422.

A single miscellaneous (MISC) *tnpB* ORF (locus 14) exists in the genome and is 172 nucleotides in length. This MISC *tnpB* ORF contains only the most 5' 102 nucleotides, and the most 3' 70 nucleotides of type 1A ORFs. Due to the

lack of internal sequence or inserts, this element cannot be confidently placed in any other group.

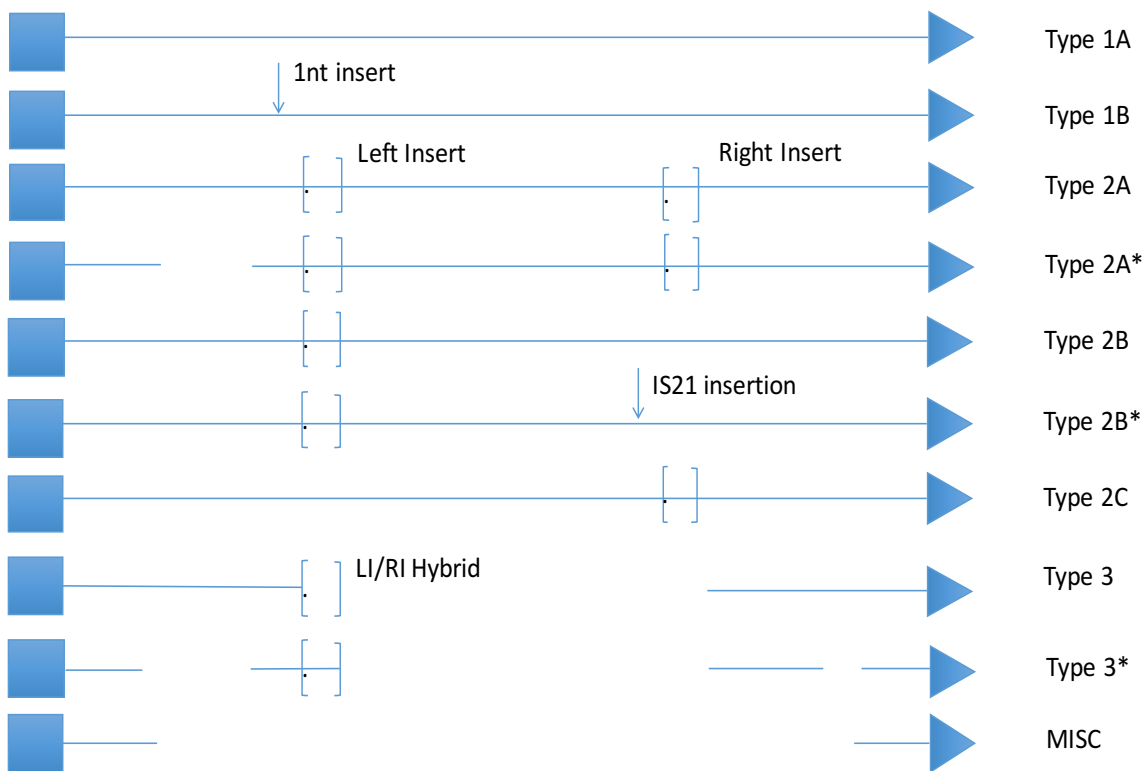


Figure 3.3. Relative IS605 tnpB sequence structures.

3.4. IS605 tnpA/tnpB INTER-ORF SPACE

The nucleotide sequence between the two divergent tnpA and tnpB ORFs is dependent on the IS605 tnpA ORF type present at each locus and varies on the tnpA end of the inter-ORF space. Figure 3.4 shows the nucleotide sequence alignments for the space between the ORFs. Each sequence is labeled with the IS605 tnpA ORF type it is present with. The inter-ORF sequence alignment for all loci is found in Appendix D.

```

Type_3      -----CTCCATTTTCTTTTTATAAGCAAACATATGTATGGTATAATTATAGTA      49
Type_4      -----ATTTTCTTTTTACAAGCAAACATATGTATGATATAATTATAGTA      45
Type_2      -----CTCCATTTTCTTTTTACAAGCAAACATATGTATGATATAATTATAGTA      49
Type_1      AAAAATCAAACCTCCATTTTCTTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Type_5      AAAAATCAAACCTCCATTTTCTTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
              *****
Type_3      GAATGGAGGTGAAAAATCA 68
Type_4      GGATGGAGGTGAAAAATCA 64
Type_2      GGATGGAGGTGAAAAATCA 68
Type_1      GGATGGAGGTGAAAAGTCA 79
Type_5      GGATGGAGGTGAAAAGTCA 79
              * *****

```

Figure 3.4. Inter-ORF sequence alignment.

3.5. ORF *tnpB* INSERTS

The inserts briefly discussed in Section 3.3 can be sorted into 3 groups using their location within the ORF and the most terminal 3 nucleotides on the 5' and 3' ends. The LI and RI are 64 nucleotides in length, while the HI is 67 nucleotides long. The structure of the three inserts are seen in Figure 3.5. All inserts share a common 61 nucleotide central region except where indicated. The LI however lacks a GCT sequence on its 3' end, and the RI insert lacks a TCA sequence on its 5' end. The hybrid insert contains both the TCA and GCT sequences. This hybrid pattern persists internal to the insert ends between 4 mismatched nucleotides that are a total of 9 nucleotides apart. These inserts disrupt the IS605 *tnpB* ORF resulting in a non-functional protein. Insert sequence alignment for all inserts is found in Appendix E.

3.6. ORF tnpB PHYLOGENY

IS605 tnpB ORFs were used for the analysis because they contain a larger sequence for alignment. Only ORF tnpB types 1-3 are included in the phylogeny.

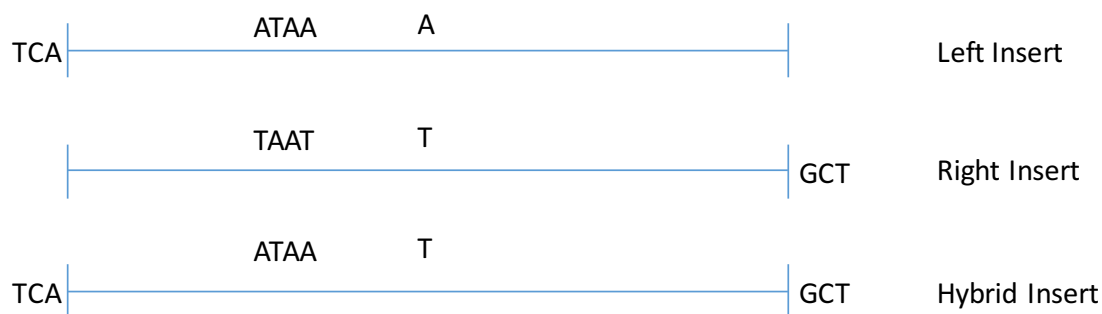


Figure 3.5 Relative insert sequence structure

Because of their more deteriorated state, types 2A*, 3* and MISC were excluded. The phylogenetic tree is located in Figure 3.6 and is labeled with the tnpB type and which locus it appears in (ex. T1A_05; Type 1A_Locus05). In the phylogenetic tree, we see that elements with structural similarities do not form a clade.

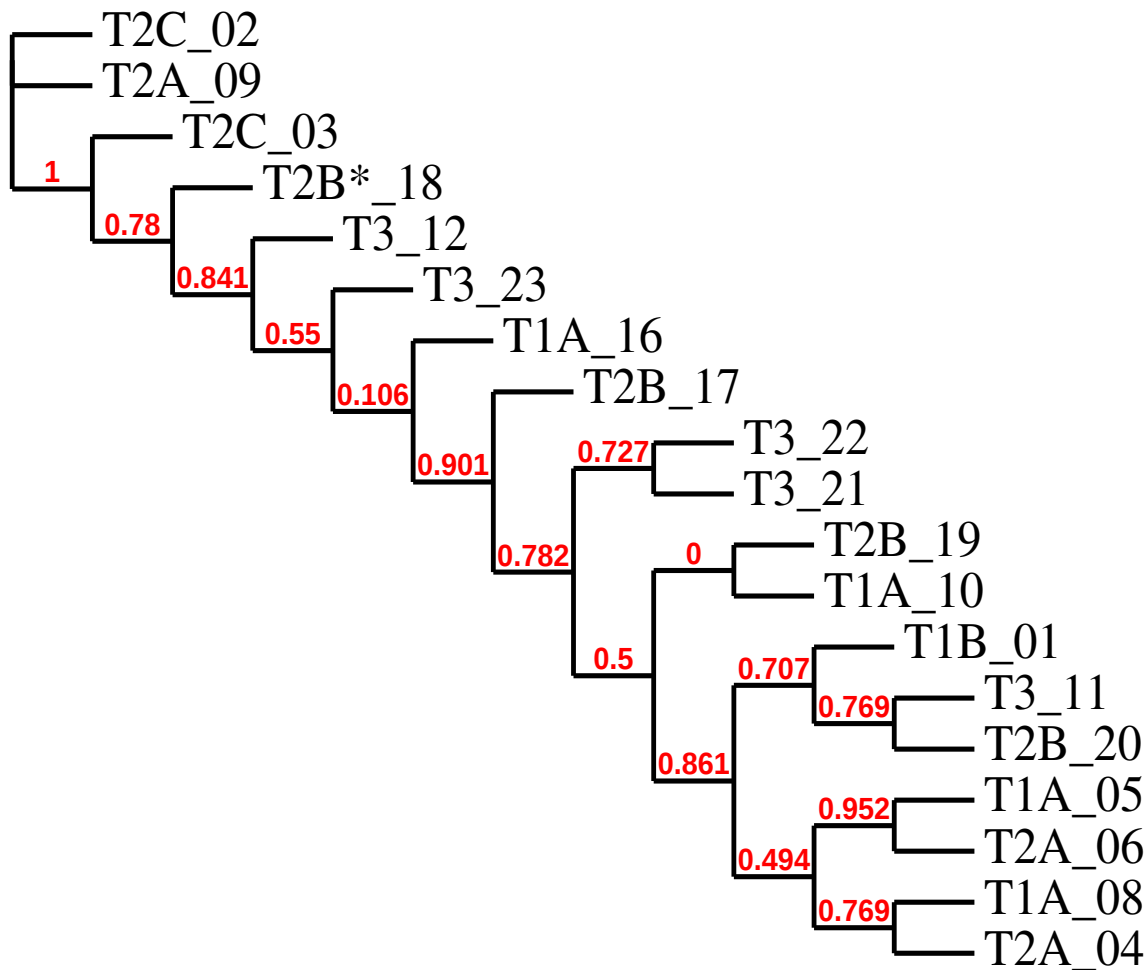


Figure 3.6. ORF *tnpB* phylogeny. Each *tnpB* is labeled with the type and locus number it appears in. Branches are labeled with branch support values. Branch length is ignored.

3.7. ELEMENT ENDS

The left end (LE) of the element, as defined as the sequence downstream of the IS605 *tnpA* ORF, is composed of a 60 nucleotide sequence for all but one of the 22 elements. The LE of locus 13 is missing 11 nucleotides on its *tnpA* end. The LE of the element begins with a TTTAT sequence (*tnpB* encoding strand)

which is consistent with IS200/605 family members. The left end sequence alignment for all loci is found in Appendix F.

The right end (RE) of the element, as defined as the sequence downstream of the 605 tnpB ORF, can be sorted into two groups and one miscellaneous based on the presence of a 28 nucleotide insert. Type 1, the consensus RE present for all elements unless otherwise stated, extends 132 nucleotides past the 3' end of the tnpB ORF. Type 2 is present at five loci (loci 01, 04, 06, 14, and 21). Type 2 RE contains a 28 nucleotide insert at position 99, and has a total length of 160 nucleotides. This 28 nucleotide insert does not show significant sequence similarity to the IS605 tnpB ORF inserts described in Section 3.4. The miscellaneous RE (locus 15) extends only 23 nucleotides past the 3' end of its respective tnpB ORF. Unlike the locus 18 tnpB disruption where the remainder of the element can be clearly identified beyond the putative IS21 family member, the remainder of the RE for locus 15 cannot be located. The right end sequence alignment for all loci is found in Appendix G.

3.8. HAIRPIN STRUCTURES

Both LE and RE sequences of the IS605 elements contain a hairpin structure required by IS200/605 family members for transposition.

3.8.1. Left End Structure. The LE has only one possible hairpin structure. It is composed of a 10 base pair stem, and 8 nucleotide loop starting 23 nucleotides from the 5' end of element (tnpB encoding strand). The LE sequence alignment highlighting the structure is seen in Figure 3.7. Figure 3.8 shows the

structure of the LE hairpin. The LE reverse complement alignments for identification of potential LE structure is found in Appendix H.

3.8.2. Right End Structures. The RE has 3 potential structures. Structures 1, 2, and 3, begin 52, 78, and 91 nucleotides from the 3' end of the tnpB ORF respectively. These structures form an imperfect stem with 8 out of 10, 9 out of 11, and 11 out of 13 base pairs with a 5, 7, and 8 nucleotide loop respectively. The 28 nucleotide insert present in the RE of 5 elements is inserted within structures 2 and 3, but not structure 1. The RE structures 1, 2, and 3 are in Figure 3.9, 3.10, and 3.11 respectively. The RE alignment of sequences highlighting these structures is shown in Figure 3.12. The RE reverse complement alignment for the identification of potential RE structures is found in Appendix I.

```
Locus09_LE      TTTATCTAAAACTGCCAAGAAAACTCCATCCAAGCTATGCATTGGGTGGAGATGAATTGG
                *****
```

Figure 3.7. Highlighted sequence of left end structure.

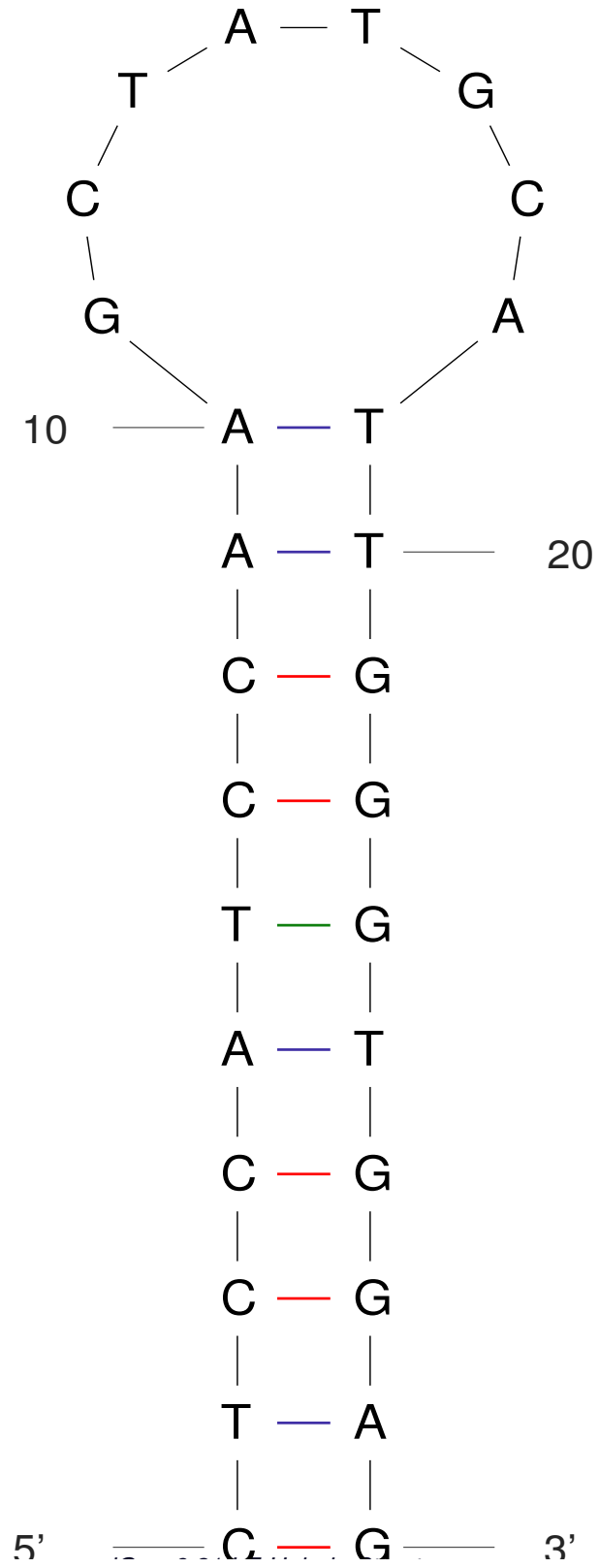


Figure 3.8. Left end hairpin structure.

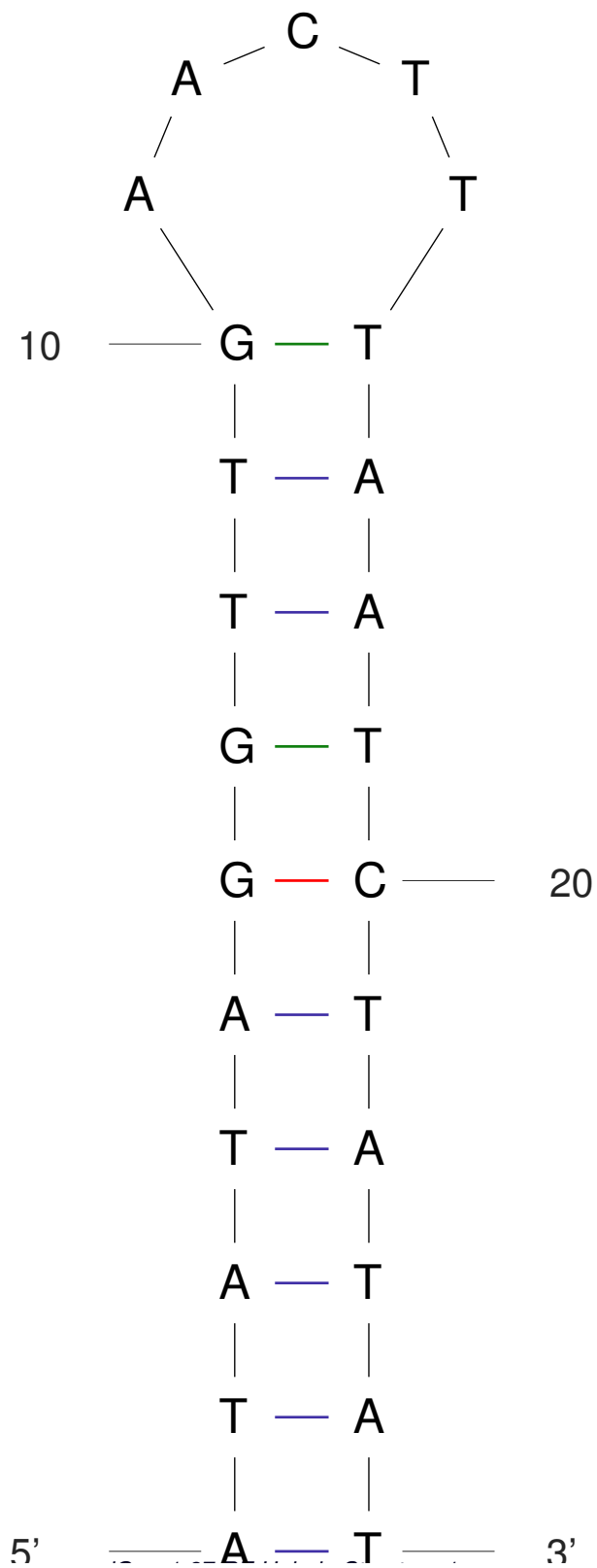


Figure 3.9. Right end hairpin structure 1.

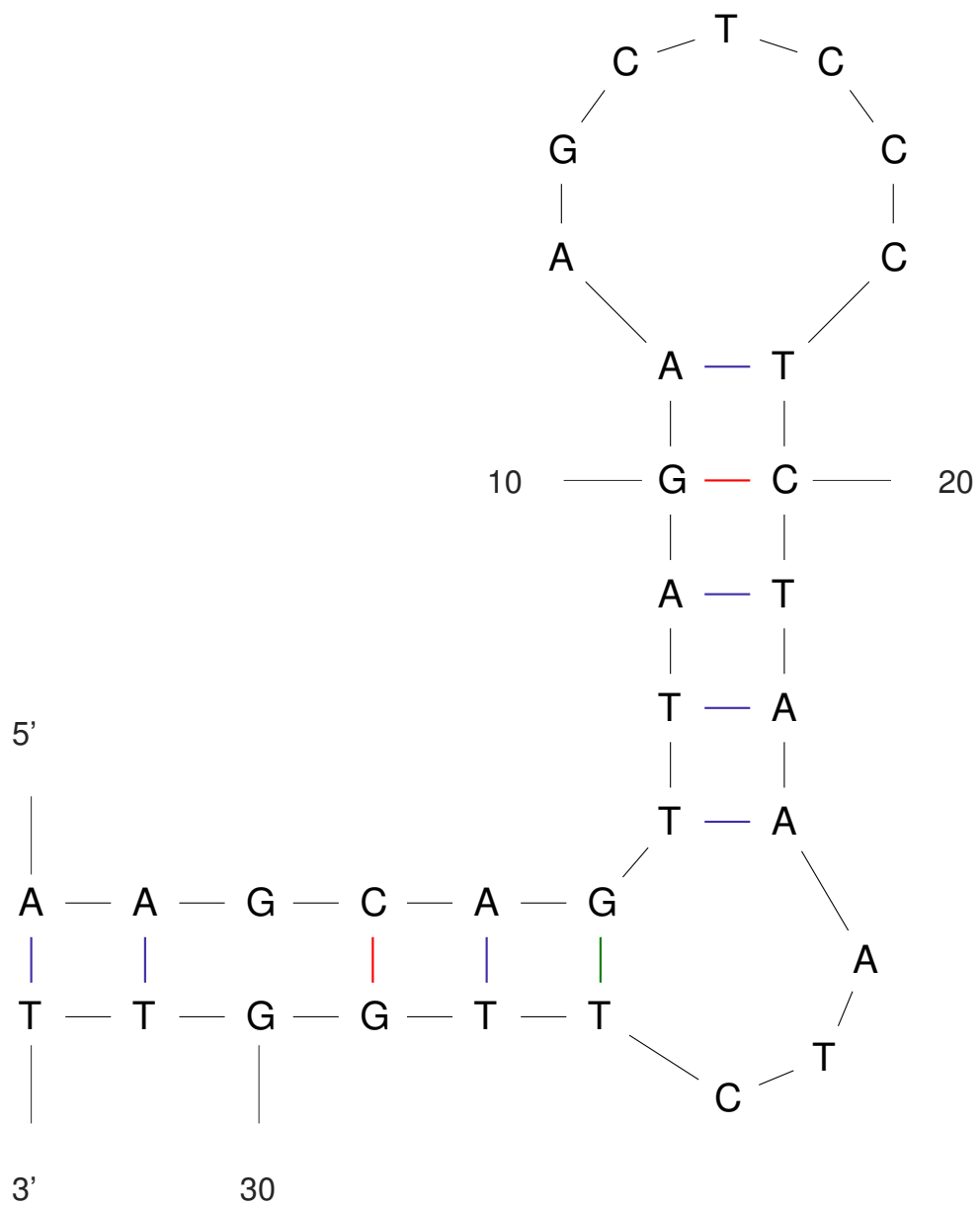


Figure 3.10. Right end hairpin structure 2.

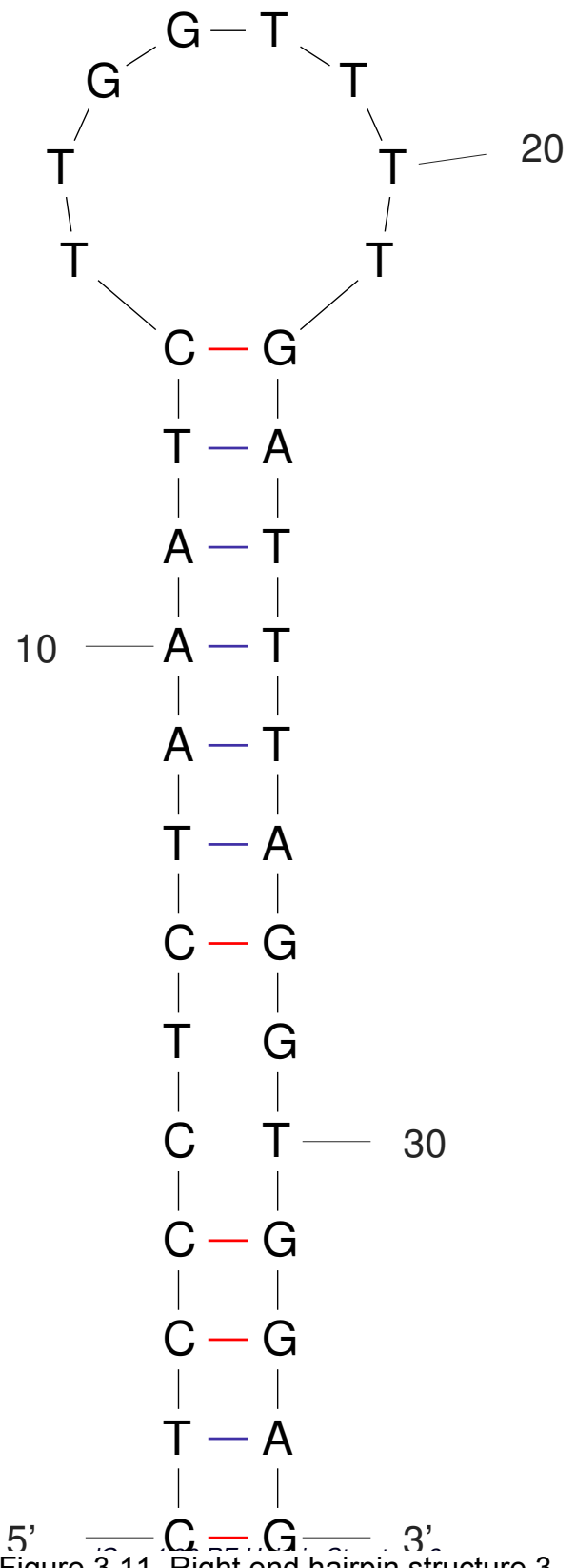


Figure 3.11. Right end hairpin structure 3.

```

Locus16_RE-1      ACTATTAGGAGCAAACCTTAAAAGCCAAACATCTTGTAACCTGACCTAGTAATATAGGTT
Locus16_RE-2      ACTATTAGGAGCAAACCTTAAAAGCCAAACATCTTGTAACCTGACCTAGTAATATAGGTT
Locus16_RE-3      ACTATTAGGAGCAAACCTTAAAAGCCAAACATCTTGTAACCTGACCTAGTAATATAGGTT
*****

Locus16_RE-1      GAACTTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGGTTTTGATTTAGGT
Locus16_RE-2      GAACTTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGGTTTTGATTTAGGT
Locus16_RE-3      GAACTTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGGTTTTGATTTAGGT
*****

Locus16_RE-1      GGAGAGGTTCAC
Locus16_RE-2      GGAGAGGTTCAC
Locus16_RE-3      GGAGAGGTTCAC
*****

```

Figure 3.12. Highlighted sequence of right end structures.

3.8.3. ORF tnpB Insert Structure. The 605 tnpB ORF inserts also contain a secondary hairpin structure. This structure is an imperfect stem with 7 out of 9 bp and a 5 nucleotide loop. The structure is shown in Figure 3.13. The insert sequence highlighting the structure is shown in Figure 3.14. The reverse complement alignments used to identify potential tnpB insert structure is found in Appendix J.

3.9. MINIATURE INVERTED REPEAT TRANSPOSABLE ELEMENTS

One IS605 MITE was identified within the genome. This MITE is approximately 271 nucleotides in length beginning at nucleotide position 843,418 in the genome and is closely located to locus 09. This MITE contains the last 58 nucleotides of tnpB ORF, no sequence of the 605 tnpA ORF, 28 nucleotides of the LE, and the entire 160 nucleotides of group 2 RE. The 28 nucleotides aligning with the LE contain the predicted secondary hairpin structure of the LE.

The LE, *tnpB* portion, and RE of the MITE are aligned with representatives in Figures 3.15, 3.16, and 3.17.

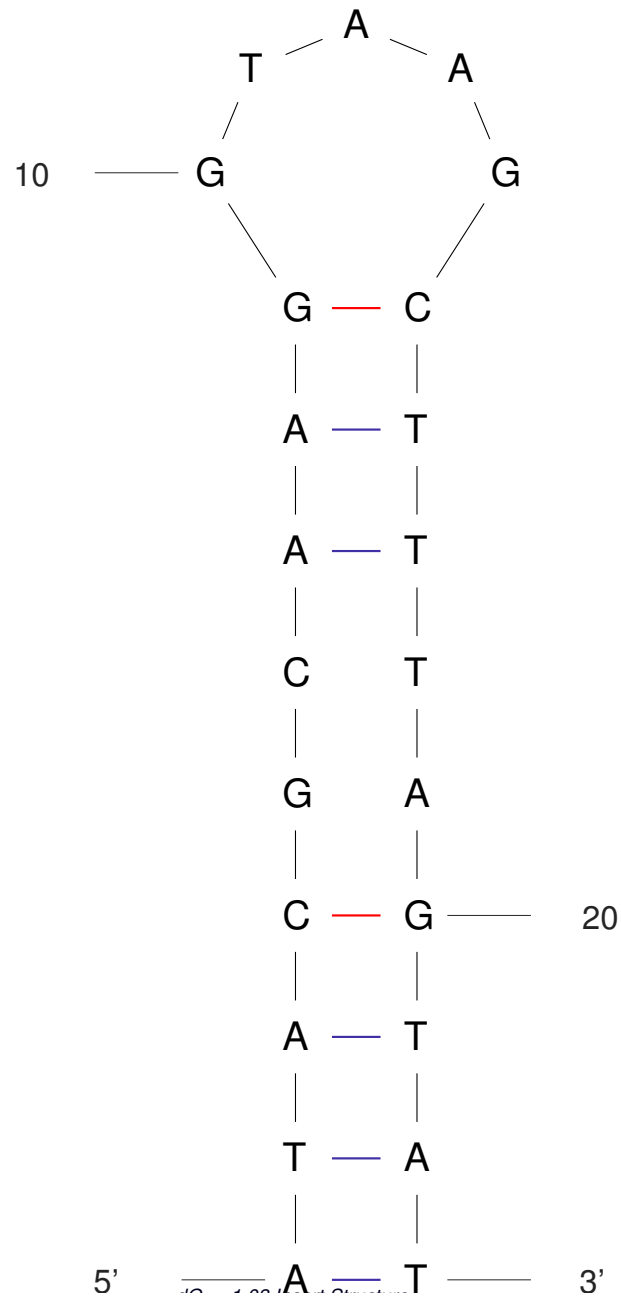


Figure 3.13. ORF *tnpB* insert hairpin structure.

```

Locus_22HI      TCACTAAAGCTTTTAAATTTATAATACGCAAGGTAAGCTTTAGTATGACCGTATTCGATTT 60
                *****
                GGCCGCT 67
                *****

```

Figure 3.14. Highlighted sequence of tnpB ORF insert structure.

```

MITE_LE          1 -----CTCCATCCAAGCTATGCATTGGGTGGA 27
                |||
Locus09_LE       1 TTTATCTAAAACTGCCAAGAAAACCTCCATCCAAGCTATGCATTGGGTGGA 50

MITE_LE          28 G----- 28
                |
Locus09_LE       51 GATGAATTGG 60

```

Figure 3.15. MITE LE alignment.

```

Locus16_tnpB    1151 ATATACTTCGTAAATACCATAACGATAAATGTATTCTCAGACCTATCAAA 1200
                |||
MITE_tnpB       1 -----TCAAA 5

Locus16_tnpB    1201 GAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATC 1250
                |||.|||
MITE_tnpB       6 GAGGTGAGAGATAATGGATTTGTGGCCAATCCTTCAAGATTAAGGGTATC 55

Locus16_tnpB    1251 CTAA 1254
                |||
MITE_tnpB       56 CTAA 59

```

Figure 3.16. MITE tnpB alignment.

```

MITE_RE          1 ACTATTAGGAGCAAAACTTAAAAGCCAAACATCTTGTAACCTGACCTAGT 50
                |||
Locus01_RE       1 ACTATTAGGAGCAAAACTTAAAAGCCAAACATCTTGTAACCTGACCTAGT 50

MITE_RE          51 AATATAGGTTGAACTTTAATCTATATGAAGCAGTTAGAAGCTCCATCCGA 100
                |||
Locus01_RE       51 AATATAGGTTGAACTTTAATCTATATGAAGCAGTTAGAAGCTCCATCCGA 100

MITE_RE          101 CGCGAAGCTAGTATCTCTTTTGATGCAAATCTTGGTTTTGATTTAGGTGG 150
                |||.|||
Locus01_RE       101 CGCGAAGCTAGCATCTCTTTTGATGCAAATCTTGGTTTTGATTTAGGTGG 150

MITE_RE          151 AGAGGTTAC 160
                |||
Locus01_RE       151 AGAGGTTAC 160

```

Figure 3.17. MITE RE alignment.

4. DISCUSSION

4.1. INSERTION SEQUENCE IDENTIFICATION

Y1 elements in *Halanaerobium hydrogeniformans* were chosen for detailed characterization because of their progressed stages of decay, their misidentification by the semi-automatic insertion sequence annotation program, and because Y1 elements do not have a strong preference for cis transposition (which is the preference for a transposase to act on the element that it was transcribed from). Many of the IS605 elements were identified as solo tnpB (IS1341) elements because partial IS605 tnpA sequences were not detected. Additionally, the most closely related tnpB in the ISfinder library (what ISSaga relies on for annotation), was a tnpB of an IS607, a serine transposase. This explains why many of the IS605's encoding only the tnpB were identified as a IS607 and suggests that for insertion sequences deposited in the ISfinder library, this IS605 is the closest relative to the IS607's or that other IS605 tnpB's have also been misidentified.

The misidentification of many of the IS605 elements in *H. hydrogeniformans* by ISSaga highlights the need for more developed automated insertion sequence annotation programs, and the importance of manual curation for the identification of insertion sequences. It also indicates the limits of library based annotation software.

4.2. ORF tnpB PHYLOGENY

A phylogenetic tree between the major tnpB ORFs was constructed to determine the order of transposition events of the IS605 elements. The tnpB ORF was chosen because it is the most consistent sequence between the 22 IS605 elements. ORFs of tnpB types 2A*, 3*, and MISC were not included in the phylogenetic analysis due to their further degraded state. To eliminate the effects of the inserts and the missing inter-insert sequence on the phylogeny, the inserts were manually removed and phylogeny was inferred without G-blocks curation.

It was hypothesized that elements sharing structural similarities (LI/RI/HI) would form a clade on the phylogenetic tree, and that it could be inferred when deletion and insertion events took place. If these elements were replicating without recombination, tnpB's with similar structure (LI/RI/HI) should form a clade. For example, all type 2A's would clade together, all type 2B's would clade together, and all type 2C's would clade together. Figure 3.6 however shows that tnpB ORFs sharing structural similarities do not form a clade. This strongly indicates that recombination between tnpB ORFs is occurring.

This evidence of insertion sequence recombination is contrary to past research. Insertion sequences were screened for evidence of recombination by searching for break points and for pairs of insertion sequence fragments showing more similarity to one another. The research concluded that there was no evidence of recombination or gene conversion [54]. It should be noted however, that the research excluded IS200/605 elements from the survey and was limited complete and annotated insertion sequences. Whereas the results presented

here contain elements that were originally annotated as pseudogenes but later manually identified as *tnpB* disrupted ORFs.

4.3. INVASION – EXPANSION – EXTINCTION CYCLES

Insertion sequences have a high sequence similarity within genomes [52]. There is a lack of evidence for recombination and gene conversion between elements [54]. Insertion sequences have a patchy distribution among genomes [56]. These observations have led to the generally accepted hypothesis that insertion sequences undergo invasion, expansion, and extinction cycles in their free living hosts.

Contrary to the DDE family of insertion sequences that show strong preference for cis action [9], Y1 elements do not. Strong cis action increases selective pressure against elements with disrupted, or otherwise non-functional protein encoding ORFs, because these elements have a reduced ability to replicate. Thus, Y1 elements with degraded ORFs encoding nonfunctional TnpA protein can still replicate, so long as they maintain the secondary structures necessary for transposition and there is at least one functional transposase encoded somewhere in the genome. This lack of cis preference has allowed for the observation of IS605 elements in various stages of degradation.

The present IS605 elements may be in the extinction phase of the insertion sequence cycle. Because they are not immediately selected against, elements with deletions and disruptions can accumulate in the genome and be observed. Other families of insertion sequences may degrade in similar ways

within their host genome, and their degraded states have not been observed because, unlike Y1 elements, their disrupted copies are removed from the genome.

Furthermore, recombination between insertion sequences may help explain the rapid extinction of elements in a genome. While ORF disruptions or fatal mutations may accumulate in one element, they can spread throughout replicates in the genome via recombination. This would reduce the number of elements with an intact transposase gene.

4.4. TYPE 5 tnpA

It is worth noting that the most commonly occurring 605 tnpA is type 5 (17 of the 22 elements). The abundance of IS605 elements containing a type 5 tnpA may be a result of increased rates of transposition relative to the other IS605 tnpA types. Presented here are two possibilities for an increased rate of transposition for elements containing a type 5 605 tnpA. Either size reduction increases transposition frequency, or the missing tnpA nucleotide sequence could have a regulatory function as well as encode a TnpA protein.

IS605 exclusively excises from, and preferentially inserts into ssDNA. This preference leads to a bias towards lagging strand template insertion when transposition is coupled with host replication [30]. As element size increases, the probability that both ends of the element exist as ssDNA decreases. Alternatively, as Okazaki fragment size increases, so does the probability that the element ends exist as ssDNA in the lagging strand template.

Thus, as element size decreases there is an increase in genome replication associated transposition events [30]. The 282 nucleotide size reduction of an element with a type 5 tnpA may increase the frequency of transposition by increasing the time spent in a ssDNA state during replication. It would be expected however that the size reduction of type 3 tnpB (530nt) would also increase the rate of transposition. The discrepancy in copy number of these elements, (6 type 3 tnpB vs 17 type 5 tnpA), does not support this. However, the accuracy of the phylogenetic tree is diminished by recombination events between elements, and it cannot be inferred which of the elements existed in the genome first or which has a higher relative replication rate.

A reduced element size increasing transposition frequency is an unlikely reason for the disproportional number of type 5 tnpA. This explanation relies on genome replication associated transposition and a preference for lagging strand template insertion. As seen in Table 3.2, there is no skew for or against insertion into the lagging strand template (10 of 22 tnpAs on leading strand)

The TnpB protein serves as a potential IS605 transposition regulatory protein and has been shown to inhibit IS605 excision and insertion. It is hypothesized that TnpB protein inhibits transposition by binding the terminal DNA hairpin structures or the TnpA protein itself. TnpB mediated transposition inhibition is dependent on the terminal Zn finger domain [35]. However, it has not been established what this domain interacts with. It is possible that the TnpB protein binds ssDNA of the IS605 tnpA ORF sequence, inhibiting TnpA binding or dimerization and preventing transposition. If the region of binding were missing

(Figure 3.2, type 5 tnpA) TnpB could not inhibit transposition and elements without this sequence would have an increased rate of transposition.

Alternatively, the disproportional number of type 5 tnpAs may be a relic of early formation after insertion sequence acquisition, and selective pressure against functional TnpA proteins. Without an accurate phylogenetic tree, it cannot be determined when this type of tnpA formed.

4.5. ORF tnpB INSERT

Left and right inserts (LI & RI) contain a common core 58 nucleotides and are distinguishable by their most 5' and 3' three nucleotides. All LI contain a TCA as the most 5' three nucleotides, while all RI contain a GCT as the most 3' three nucleotides. The hybrid insert (HI) is 67 nucleotides in length and contains both TCA and GCT trinucleotide sequences at the 5' and 3' ends of the insert as seen in Figure 3.5. This pattern indicates that a recombination event has occurred between a LI and a RI to form a hybrid insert.

This same hybrid pattern persists internal to the insert ends. The LI contains an ATAA and a A at nucleotide positions 20 and 33 respectively, while the RI contain a TAAT and T at these positions. The hybrid insert contains the ATAA and T at positions 20 and 33 indicating LI towards the 5' end and RI towards the 3' end. This suggests that the initiating endonuclease for recombination between these inserts has a higher affinity for the sequence between positions 20 and 33 of the insert.

These hybrid inserts (HI) are the product of recombination from a LI and a RI of either the same or different elements, (e.g. a single T2A that contains both a LI and RI, or a T2B and a T2C that contain a LI and a RI respectively). If the recombination event were to take place between a LI and RI of different elements, the results would be one element containing a hybrid insert with LI and RI characteristics at the 5' and 3' ends respectively, excluding the ORF regions between inserts (type 3 tnpB, Figure 3.3), and another element containing a HI with the LI and RI characteristics at the 3' and 5' ends respectively, with the sequence between the inserts being duplicated.

If the recombination took place between a LI and RI of the same element, only one product capable of transposition could be formed, that is a hybrid insert with LI and RI characteristics at the 5' and 3' ends, (type 3 tnpB, Figure 3.3).

No inserts were observed in the genome showing LI or RI characteristics at the 3' and 5' ends, nor were inter insert sequence duplications identified. It is hypothesized that all type 3 elements containing a HI are a result of recombination between a LI and a RI of a type 2A tnpB.

The LI and RI show high sequence similarity, indicating that that they originated from the same source. The differentiating three nucleotide sequence at either end suggests an imprecise excision of the insert before insertion into the IS605 tnpB ORF. The 64 nucleotide sequence of the insert, or any part of it, is not found in the genome outside a tnpB ORF.

The independent insertion of all the LI and RI to the same relative location within the tnpB ORF is unlikely. Their reoccurrence in tnpB ORF is thus likely a

result of two insertion events and the replication of those elements. As such it is also hypothesized that the presence of these inserts in the *tnpB* ORF does not impede transposition of the IS605 elements.

4.6. ELEMENT ENDS AND STRUCTURES

Element ends of IS200/605 family members contain hairpin structures indispensable for transposition. In characterized IS200/605 elements, left end (LE) and right end (RE) structures are the same for each element [32], [28].

The LE sequence for all IS605 elements in *H. hydrogeniformans* is highly conserved and stretches 60nt downstream of the *tnpA* ORF. The LE has the potential to form a single hairpin structure (Figure 3.7) but shows no sequence homology to the RE.

The RE of the IS605 elements is 132 nucleotides in length and has the potential to form 3 different hairpin structures (Figure 3.9, 3.10, and 3.11). Highlighted sequences of the RE structures (Figure 3.12) show that structures 2 and 3 have significant overlap, making them mutually exclusive. Structures 1 and 3 are separated by 14 nucleotides, so it may be possible to form both structures simultaneously. The base pairing in the stems of structures 1 and 2 however, are separated by a single nucleotide. It is not clear if structures 1 and 2 are exclusive or competitive, as a single nucleotide space may permit both structures to co-exist.

There has previously been speculation that the terminal hairpin structures of IS200/605 elements serve as a transcriptional terminator as well as

prevents ribosome binding. It has since been established that they play a mechanistic role in transposition [24]. Potentially competing and mutually exclusive structures may further serve a regulatory role by preventing the mechanistic hairpin structure from being bound by a TnpA monomer.

Competitive structures have been reported before, although in these instances it was clear which structures were mechanistic as only a single common structure was observed between the LE and the RE [24].

Of characterized IS200/605 elements, it is unknown whether the TnpA binds the terminal hairpins through structure recognition or DNA sequence recognition in the stem or loop of the structures [23]. This is the first known report to describe a characterized IS605 element that does not contain the same secondary structure at both the LE and RE. This difference in LE and RE structure, while maintaining transposable capability of the element, suggests that the hairpin is recognized from structure alone. However, there could be a short conserved sequence in both LE and RE structures recognized by TnpA.

Underlined in the highlighted LE structure sequence (Figure 3.8) and the highlighted RE structure 2 sequence (Figure 3.12) is a common AAGCT. This pentanucleotide sequence is presented in the hairpin loop in both structures. The sequence and location in the hairpin is the strongest similarity between any of the potential structures. This suggests that RE structure 2 is mechanistic, implying that RE structure 1 and 3 are potentially regulatory, and that TnpA recognizes a pentanucleotide sequence AAGCT in the loop of the hairpin structure.

At five loci, a 28 nucleotide long sequence has inserted into structure 2 and 3. This insert occurs immediately after nucleotide 21 of structure 2 and nucleotide 8 of structure 3 disrupting both structures. Because elements containing this RE insert have replicated (loci 01, 04, 06, 14, and 21), it is not completely preventing replication.

This supports the notion that RE structure 1 is the mechanistic structure. However, as the insert occurs toward the end of the structure, the AAGCT pentanucleotide sequence in the loop of structure 2 could still be presented. It is possible that the insert only reduces the affinity of TnpA for the RE structure 2.

Elements surveyed by Ronning [28] were shown to excise in a strand specific manner dependent on a secondary loop containing a T in the stem of the structure. RE structure 2 (Figure 3.10) contains a 3 nucleotide secondary loop containing a T. This secondary loop however is not present in the LE structure of elements described here.

Of note is the secondary structure of the tnpB ORF insert (Figure 3.13). In the reported strand, this hairpin contains an AAGCT pentanucleotide sequence in the loop. Additionally, an AAGGT sequence can be found in a more similar position compared to the pentanucleotide sequence in the LE structure and the RE structure 2. The complement strand hairpin also contains an AAGCT sequence in a similar position compared to the two structures. The implications of this observation are unknown at this time.

4.7. MITES

Bacterial MITES are typically difficult to identify as they are short elements and the parental transposable elements are often no longer present in the genome. A single IS605-related MITE was located in *H. hydrogeniformans*. This element is 247 nucleotides in length. It contains 28 nucleotides of the LE (Figure 3.15), the last 59 nucleotides of the IS605 tnpB ORF (Figure 3.16), and the entire 160 nucleotides of a RE containing the 28 nucleotide insert (Figure 3.17). These sequences occur in succession, without gaps. The 28 nucleotides of the LE contain the entire LE hairpin structure. The LE hairpin structure and an intact RE make it likely that this element is transposable.

4.8. CONCLUSIONS

Although only a single element contains an intact IS605 tnpA, all IS605 elements reported here contain intact hairpin structures and are likely capable of transposition by a TnpA acting in trans. Dissimilar RE and LE structure sequences suggest that hairpin recognition may be independent of hairpin sequence, although a conserved pentanucleotide sequence present in the hairpin loop is suggestive of a sequence specific recognition. Unique to our findings, the inserts in the tnpB ORF provide structural differences that can be used to infer recombination between insertion sequences. Because these Y1 elements do not rely on the integrity of their ORF for transposition, their detailed survey in a single genome provides a snapshot of how insertion sequences degrade during invasion-expansion-extinction cycles.

4.9. FUTURE DIRECTIONS

The results presented here explore interesting insertion sequence activity within *Halanaerobium hydrogeniformans*. However, they only provide a snapshot of activity. While there is evidence indicating element recombination, direct evidence for insertion sequence recombination is absent. Similarly, it is hypothesized that all the IS605 elements discussed are transposable due to their intact secondary structures. However, direct evidence of transposition is still needed. The LE and RE of the element do not share a common secondary structure sequence. It is unknown what commonality between the structures is essential for TnpA recognition. Future directions for research would address these issues.

Amplification and sequencing of the IS605 insertion sequences from a new sample of *H. hydrogeniformans* from the environment and comparison of the tnpB ORFs and flanking sequences could elucidate transposition and recombination hypotheses. TnpA and TnpB binding assays with elements containing mutated hairpin structures could help determine the functional sequences of the hairpins and the mechanism by which TnpB inhibits transposition

Immediate future research would include the comparison of the IS605 elements in *H. hydrogeniformans* to those in *Halanaerobium saccharolyticum*. *H. saccharolyticum* is the closest relative to *H. hydrogeniformans* and the sequence of its genome is currently being determined. Partial genome sequences from the project are available. Initial observations from the partial sequences show that *H.*

saccharolyticum contains a highly similar TnpA and TnpB to those described here (approximately 90% nucleotide similarity) in its genome in fragmented and partial copies. Interestingly, *Halanaerobium praevalens*, which is closely related to *H. hydrogeniformans* and *H. saccharolyticum*, does not contain any of the IS605 insertion sequences described in this work. Comparison of the elements and syntenic regions between *H. hydrogeniformans*, *H. saccharolyticum*, and *H. praevalens* would help us understand the origins of these elements, their continued activity in genomes, and the manner in which they decompose.

APPENDIX A.

IS200 ELEMENT ENDS AND REVERSE COMPLEMENTS

```

#=====
#
# Aligned_sequences: 2
# 1: IS200_RE
# 2: IS200_LE
# Matrix: EDNAFULL
# Gap_penalty: 10.0
# Extend_penalty: 0.5
#
# Length: 76
# Identity:      37/76 (48.7%)
# Similarity:    37/76 (48.7%)
# Gaps:          31/76 (40.8%)
# Score: 80.5
#
#
#=====

```

```

IS200_RE      22 TATTAGTTTACAAC TAAGTT--ATA-----TCATAATATAAA TATACTA      63
                |||||.|||          || .||          ||          |||.|.|.|
IS200_LE      1 TATTATTTT-----TTGCTTACCCGGGTC-----AATTTTCAA      33

IS200_RE      64 TAATATAACAATAGAGTCAATAATTT      89
                |          |||.|.|.|.|||          |||||
IS200_LE      34 T----TAATAAAAGGGTCA--AATTT      53

```

```

>>IS200_LE_RC (53 nt)
Waterman-Eggert score: 46; 68.0 bits; E(1) < 9.7e-18
55.3% identity (55.3% similar) in 47 nt overlap (53-7:2-48)

```

```

          50          40          30          20          10
IS200_ AAATTTGACCC TTTTATTAATTGAAAATTGACCCGGGTAAGCAAAAA
      :: ::  :: ::  : : :  :: :  :: ::  :  :: ::
IS200_ AATTTGACCC TTTTATTAATTGAAAATTGACCCGGGTAAGCAAAAA
          10          20          30          40

```

```

>--
Waterman-Eggert score: 46; 12.4 bits; E(1) < 0.41
90.9% identity (90.9% similar) in 11 nt overlap (53-43:24-34)

```

```

          50
IS200_ AAATTTGACCC
      ::: :::::
IS200_ AAAATTGACCC
          30

```

>--

Waterman-Eggert score: 31; 8.6 bits; E(1) < 1
87.5% identity (87.5% similar) in 8 nt overlap (11-4:46-53)

```

      10
IS200_ AAAAAATA
      ::: ::::
IS200_ AAATAATA
      50

```

>--

Waterman-Eggert score: 30; 8.3 bits; E(1) < 1
100.0% identity (100.0% similar) in 6 nt overlap (35-30:26-31)

```

IS200_ AATTGA
      :::::
IS200_ AATTGA
      30

```

>>IS200_LE_RC (53 nt)

Waterman-Eggert score: 50; 13.4 bits; E(1) < 0.23
65.0% identity (65.0% similar) in 40 nt overlap (1-39:15-53)

```

      10      20      30
IS200_ TATTATTTTTTGCTT-ACCCGGGTCAATTTCAATTAATA
      ::::: :: :: ::::: :: :: :::::
IS200_ TATTAATTGAAAATTGACCCGGGT-AAGCAAAAAATAATA
      20      30      40      50

```

>>IS200_RE_RC (101 nt)

Waterman-Eggert score: 78; 73.9 bits; E(1) < 5.6e-19
63.3% identity (63.3% similar) in 60 nt overlap (98-40:23-78)

```

      90      80      70      60      50
IS200_ CTTTATT-TGAAATTATTGACTCTATTGTTATATTATAGTATATTTATATTATGATATAA
      :: :::: : : ::::: : ::::: ::::: ::::: : : : : : :
IS200_ CTCTATTGTTATATTATAG----TATATTTATATTATGATATAACTTAGTTGTAAACTAA
      30      40      50      60      70

```

>--

Waterman-Eggert score: 66; 11.2 bits; E(1) < 0.99
66.7% identity (66.7% similar) in 63 nt overlap (92-34:19-79)

```

      90      80      70      60      50      40
IS200_ TTGAAATTATTGACTCTATTGT--TATATT-ATAGTAT-ATTTATATTATGATATAACTT
      ::::: ::::: : ::::: : ::::: ::::: ::::: ::::: ::::: : : : :
IS200_ TTGACTCTATTGT-TATATTATAGTATATTTATATTATGATATAACTTA-GTTGTAAACT

```

>>IS200_RE_RC (101 nt)
 Waterman-Eggert score: 116; 18.3 bits; E(1) < 0.03
 69.0% identity (69.0% similar) in 58 nt overlap (15-72:7-63)

```

      20      30      40      50      60      70
IS200_ TATAAAATATTAGTTTACAAC TAAGTTATATCATAATATAAA TATACTATAATATAAC
      :::  :  ::::  ::  ::  :::::  ::  ::::  ::::  ::  :::::  ::
IS200_ TATTTGAAATTA-TTGACTCTATTGTTATATTATAGTATATTTATATTATGATATAAC
      10      20      30      40      50      60

```

>--

Waterman-Eggert score: 92; 14.9 bits; E(1) < 0.28
 62.1% identity (62.1% similar) in 58 nt overlap (1-58:44-101)

```

      10      20      30      40      50
IS200_ TATATATCCTCTCCTATAAAATATTAGTTTACAAC TAAGTTATATCATAATATAAAATA
      :::  :::  :  :  ::::  ::  :  ::  :  :  ::  ::::  :  :  :::  :::
IS200_ TATTTATATTATGATATAA CTTAGTTGTAAACTAATATTTTATAGGAGAGGATATATA
      50      60      70      80      90      100

```

>--

Waterman-Eggert score: 90; 14.6 bits; E(1) < 0.33
 58.1% identity (58.1% similar) in 93 nt overlap (8-99:3-94)

```

      10      20      30      40      50      60
IS200_ CCTCTCCTATAAAATATT-AGTTTACAAC TAAGTTATATCATAATATAAA TATACTATAAA
      :::  :  :  ::::  ::::  :  :  ::  :  :  ::::  ::::  :  :  ::::  ::::
IS200_ CCTTTATTTGAAATTATTGACTCTATTGTTATATTATAGTATATTTATATTATGATATAAA
      10      20      30      40      50      60

```

APPENDIX B.

IS605 tnpA SEQUENCE ALIGNMENTS

CLUSTAL O(1.2.1) multiple sequence alignment

```

Locus13 ----- 0
Locus01 ----- 0
Locus20 ----- 0
Locus15 ----- 0
Locus09 ATGGATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATTAGTT 60

Locus13 ----- 0
Locus01 ----- 0
Locus20 ----- 0
Locus15 ----- 0
Locus09 GTAATTACAAAATACAGACATGAATGTATTACTTTTGAATGCTTGAAGAATTAGAAAA 120

Locus13 -----GGAGAAAA 9
Locus01 -----GGAGAAAA 9
Locus20 -----GGAGAAAA 9
Locus15 ----- 0
Locus09 ATATTCACCAGATTACTCAAGGACAAAGTTTGAATGTTCTAGAGTTTGGAGGAGAAAA 180

Locus13 GATCATGTGCATATCCTCTTTGAAAATCCACCTCAGGTACAATTATCTAAGTTAGTTAAT 69
Locus01 GATCATGTGCATATCCTCTTTGAAACTCCATCTCAGGTACAATTATCTAAGTTAGTTAAT 69
Locus20 GATCATGTGCATATCCTCTTTGAAACTCCACCTCAGGTACAATTATCTAAGTTAGTTAAT 69
Locus15 ----- 0
Locus09 GATCATGTGCATATCCTCTTTGAAACTCCACCTCAGGTACAATTATCTAAGTTAGTTAAT 240

Locus13 ATATTA AAAACTGTATCTTCAAGACTTATCAAAAAGCAATATGAACACCAT----- 120
Locus01 ATATTA AAAACTGTATCTTCAAGACTTATCAAAAAGCAATATGAACACCATCTGAAAAA 129
Locus20 ATATTA AAAACAGTATCTTCAAGACTTATCAAAAAGCAATATGAACACCATCTGAAAAA 129
Locus15 -----AAA 3
Locus09 ATATTA AAAACAGTATCTTCAAGACTTATCAAAAAGCAATATGAACACCATCTGAAAAA 300

Locus13 ----- 120
Locus01 TATTATFGGAAACCTGCTTTTTGGTCTAGAAGCTACTGCATTTTGTCTACTGGTGGTGCT 189
Locus20 TATTATFGGAAACCTGCTTTTTGGTCTAGAAGCTACTGCATTTTGTCTACTGGTGGTGCT 189
Locus15 TATTATFGGAAACCTGCTTTTTGGTCTAGAAGCTACTGCATTTTGTCTACTGGTGGTGCT 63
Locus09 TATTATFGGAAACCTGCTTTTTGGTCTAGAAGCTACTGCATTTTGTCTACTGGTGGTGCT 360

Locus13 ----- 120
Locus01 ACTATTGAGACAATTA AAAAGTATATTGAAAATCAGAATAAATAG 234
Locus20 ACTATTGAGACAATTA AAAAGTATATTGAAAATCAGAATAAATAG 234
Locus15 ACTATTGAGACAATTA AAAAGTATATTGAAAATCAGAATAAATAG 108
Locus09 ACTATTGAGACAATTA AAAAGTATATTGAAAATCAGAATAAATAG 405

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CLUSTAL O(1.2.1) multiple sequence alignment

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Locus_09      ATGGATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATTTAGTT      60
Locus_06      ATGAATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATTTAGTT      60
Locus_04      ATGGATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_03      ATGGATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_19      ATGAATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_10      ATGAATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_12      ATGAGTAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_02      ATGAATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_11      ATGAATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_18      ATGAATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_22      ATGAATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_17      ATGAATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_16      ATGAATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_08      ATGAATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_05      ATGAATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_23      ATGGATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_21      ATGGATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
Locus_14      ATGGATAGAGACTTAAATAACAATTATCATTCTGTTTATAGTCTACAATATCATT--ATT      58
                ***      *****      *****      *****      **

Locus_09      GTAATTACAAAATACAGACATGAATGTATTACTTTTGAAATGCTTGAAGAATTGAAAAA      120
Locus_06      -----      60
Locus_04      -----      58
Locus_03      -----      58
Locus_19      -----      58
Locus_10      -----      58
Locus_12      -----      58
Locus_02      -----      58
Locus_11      -----      58
Locus_18      -----      58
Locus_22      -----      58
Locus_17      -----      58
Locus_16      -----      58
Locus_08      -----      58
Locus_05      -----      58
Locus_23      -----      58
Locus_21      -----      58
Locus_14      -----      58

Locus_09      ATATTACCAGATTACTCAAGGACAAAGTTTGTAATGTTCTAGAGTTTGGAGGAGAAAAA      180
Locus_06      -----      60
Locus_04      -----      58
Locus_03      -----      58
Locus_19      -----      58
Locus_10      -----      58
Locus_12      -----      58
Locus_02      -----      58
Locus_11      -----      58
Locus_18      -----      58
Locus_22      -----      58
Locus_17      -----      58
Locus_16      -----      58

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Locus_08	-----	58
Locus_05	-----	58
Locus_23	-----	58
Locus_21	-----	58
Locus_14	-----	58
Locus_09	GATCATGTGCATATCCTCTTTGAAACTCCACCTCAGGTACAATTATCTAAGTTAGTTAAT	240
Locus_06	-----	60
Locus_04	-----	58
Locus_03	-----	58
Locus_19	-----	58
Locus_10	-----	58
Locus_12	-----	58
Locus_02	-----	58
Locus_11	-----	58
Locus_18	-----	58
Locus_22	-----	58
Locus_17	-----	58
Locus_16	-----	58
Locus_08	-----	58
Locus_05	-----	58
Locus_23	-----	58
Locus_21	-----	58
Locus_14	-----	58
Locus_09	ATATTAAAAACAGTATCTTCAAGACTTATCAAAAAGCAATATGAACACCATCTGAAAAA	300
Locus_06	-----	60
Locus_04	-----	58
Locus_03	-----	58
Locus_19	-----	58
Locus_10	-----	58
Locus_12	-----	58
Locus_02	-----	58
Locus_11	-----	58
Locus_18	-----	58
Locus_22	-----	58
Locus_17	-----	58
Locus_16	-----	58
Locus_08	-----	58
Locus_05	-----	58
Locus_23	-----	58
Locus_21	-----	58
Locus_14	-----	58
Locus_09	TATTATTGAAACCTGCTTTTTGGTCTAGAAGCTACTGCATTTTGTCTACTGGTGGTGCT	360
Locus_06	-----GTATCTTCTGGTGGTGCT	78
Locus_04	-----GTATCTTCTGGTGATGCT	76
Locus_03	-----GTATCTTCTGGTGATGCT	76
Locus_19	-----GTATCTTCTGGTGATGCT	76
Locus_10	-----GTATCTTCTGGTGATGCT	76
Locus_12	-----GTATCTTCTGGTGATGCT	76
Locus_02	-----GTATCTTCTGGTGGTGCT	76
Locus_11	-----GTATCTTCTGGTGGTGCT	76
Locus_18	-----GTATCTTCTGGTGGTGCT	76

Locus_22	-----GTATCTTCTGGTGGTGCT	76
Locus_17	-----GTATCTTCTGGTGGTGCT	76
Locus_16	-----GTATCTTCTGGTGGTGCT	76
Locus_08	-----GTATCTTCTGGTGGTGCT	76
Locus_05	-----GTATCTTCTGGTGGTGCT	76
Locus_23	-----GTATCTTCTGGTGGTGCT	76
Locus_21	-----GTATCTTCTGGTGGTGCT	76
Locus_14	-----GTATCTTCTGGTGGTGCT	76

* ** * ** * ** *

Locus_09	ACTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	405
Locus_06	ACTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	123
Locus_04	GCTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_03	GCTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_19	GCTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_10	GCTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_12	GCTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_02	ACTATTGAGACAATTAAAAAGTATATTGAAAATAAGAATAAATAG	121
Locus_11	ACTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_18	ACTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_22	ACTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_17	ACTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_16	ACTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_08	ACTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_05	ACTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_23	ACTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_21	ACTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121
Locus_14	ACTATTGAGACAATTAAAAAGTATATTGAAAATCAGAATAAATAG	121

APPENDIX C

IS605 tnpB SEQUENCE ALIGNMENTS

Type 1A/B

CLUSTAL O(1.2.1) multiple sequence alignment

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T1B_01      ATGCGATTATCATTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T1A_16      ATGCGATTATCATTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T1A_10      ATGCGATTATCATTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T1A_08      ATGCGATTATCATTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T1A_05      ATGCGATTATCATTAAATTC AACCCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
*****

T1B_01      GAATTAGCCTGGCATTGCTC TAAATTATATAATATAGTCAATTATCAGATTAAAAATAAT      120
T1A_16      GAATTAGCCTGGCATTGCTC TAAATTATATAATACAGTCAATTATCAGATTAAAAATAAT      120
T1A_10      GAATTAGCCTGGCATTAGTAA ACTATATAATACAGTCAATTATGAGGTTAAAAACAAT      120
T1A_08      GAATTAGCCTGGCATTGCTC TAAATTATATAATATAGTCAATTATCAGATTAAAAATAAT      120
T1A_05      GAATTAGCCTGGCATTGCTC TAAATTATATAATATAGTCAATTATCAGATTAAAAATAAT      120
*****

T1B_01      AAAGATGTAAAAGTTGTCT ATACTGAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
T1A_16      AAAGATGTAAAAGCTGTCT ATACTGAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
T1A_10      AAAGATGTAAAAGCTGTCT ATACTGAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
T1A_08      AAAGATGTAAAAGCTGTCT ATACTGAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
T1A_05      AAAGATGTAAAAGCTGTCT ATACTGAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
*****

T1B_01      GACTACCTTCACTCCCATA ACAGACAGCAGGCATTAAGCAGTTAGCTCAGGACTGGAAA      240
T1A_16      GACTACCTTCACTCCCATA ACAGACAGCAGGCATTAAGCAGTTAGTTCAGGACTGGAAA      240
T1A_10      GACTACCTTCACTCCCATA ACAGACAGCAGGCATTAAGCAGTTAGCTCAGGACTGGAAA      240
T1A_08      GACTACCTTCACTCCCATA ACAGACAACAGGCATTAAGCAGTTAGCTCAGGACTGGAAA      240
T1A_05      GACTACCTTCACTCCCATA ACAGACAGCAGGCATTAAGCAGTTAGCTCAGGACTGGAAA      240
*****

T1B_01      AGTTTTTTTAATTCCTCTC AAGATTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300
T1A_16      AGTTTTTTTAATTCCTCTC AAGATTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300
T1A_10      AGTTTTTTTAATTCCTCTC AAGATTATAAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA      300
T1A_08      AGTTTTTTTAATTCCTCTC AAGATTATAAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA      300
T1A_05      AGTTTTTTTAATTCCTCTC AAGATTATAAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA      300
*****

T1B_01      GGACCACCTAATTTTAAAC ATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
T1A_16      GGGTCACCTAATTTTAAAC ATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
T1A_10      GGATCACCTAATTTTAAAC ATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
T1A_08      GGGTCACCTAATTTTAAAC ATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
T1A_05      GGGTCACCTAATTTTAAAC ATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
**

T1B_01      GCTGTTAGAATTAGAGATA ACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA      420
T1A_16      GCTGTTAGAATTAGAGATA ACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA      420
T1A_10      GCTGTTAGAATTAAAGATA ACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA      420
T1A_08      GCTGTTAGGATTAGAGATA ACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA      420
T1A_05      GCTGTTAGAATTAAAGATA ACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA      420
*****

T1B_01      TATAATGTGAAAAGCTCTT AATTTGAGCTGCCTGAAAGCAGTTCAAAGCATTATAGATTT      480
T1A_16      TATAATGTGAAAGGCTCTT AATTTGAGCTGC-CTGAAGCAGTTCAAAGCATTATAGATTT      479
T1A_10      TATAATGTGAAAAGCTCTT AATTTGAGCTGC-CTGAAGCAGTTCAAAGCATTATAGATTT      479
T1A_08      TATAATGTGAAAGGCTCTT AATTTGAGCTGC-CTGAAGCAGTTCAAAGCATTATAGATTT      479

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T1A_05	TATAATGTGAAAAGCCTTAATTTTGAGCTGC-CTGAAGCAGTTCAAAGCATTATAGATTT ***** * * *****	479
T1B_01	AGATGCTGTCCAGCAGATAAAGATAAAGCAGGACCGTATTTCTAAAAGATGGTATCTACT	540
T1A_16	AGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCTCTT	539
T1A_10	AGATGCTGGCCAGCAGATAAAGATTAAGCAGGACCGCATTCTAAAAGATGGTATCTACT	539
T1A_08	AGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCTACT	539
T1A_05	AGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCTACT ***** ***** *	539
T1B_01	AATCATCTATAAAACCGAGGAAATAAAAGAAAATAATAACCTAACATAATGGCAGTTGA	600
T1A_16	AATTATCTACAAAGTTAAAGAGGCCAAAAGAAAGTAAGAAATCTAACATAATGGCAGTAGA	599
T1A_10	AATCATCTATAAGGTCGAGGAAATAAAAGAAAATAATAACCTAACATAATGGCAATAGA	599
T1A_08	AATCATCTATAAGACCGAGGAAATAAAAGAAAATAATAACCTAACATAATGGCAGTTGA	599
T1A_05	AATCATCTATAAGACCGAGGAAATAAAAGAAAATAATAACCTAACATAATGGCAGTAGA *** ** * * * ***** ** * **	599
T1B_01	TTTAGGCCCTTGATAATTTGGCTACTTTAACATTTAAAAACAATTCTGATTGTTATATTAT	660
T1A_16	TCTAGGCTTTGATAATTTGGCTACTTTAATATTTAAAAACAATTCTGATTGTTATATTAT	659
T1A_10	TCTAGGCTTTGATAATTTGGCTACTTTAACATTTAAAAACAATTCTGATTGTTATATTAT	659
T1A_08	TCTAGGCTTTGATAATTTGGCTACTTTAACATTTAAAAACAATTCTGATTGTTATATTAT	659
T1A_05	TCTAGGCTTTGATAATTTGGCTACTTTAACATTTAAAAACAATTCTGATTGTTATATTAT * **** *****	659
T1B_01	CAATGGTAAAACCTATTAATCCAAAAATCTTATTTTAATAAAGAAATGCCAGACTACA	720
T1A_16	CAATGGTAAAACCTATTAATCCAAAAATCTTATTTTAATAAAGAAATGCCAGACTACA	719
T1A_10	CAATGGTAAAACCTATTAATCCAAAAATCTTATTTTAATAAAGAAATGCCAGACTACA	719
T1A_08	CAATGGTAAAACCTATTAATCCAAAAATCTTATTTTAATAAAGAAATGCCAGACTACA	719
T1A_05	CAATGGTAAAACCTATTAATCCAAAAATCTTATTTTAATAAAGAAATGCCAGACTACA *****	719
T1B_01	AAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAAAATA	780
T1A_16	AAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAAAATA	779
T1A_10	AAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAAAATA	779
T1A_08	AAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAAAATA	779
T1A_05	AAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAAAATA *****	779
T1B_01	TCTGAGATTAAAGAGAAGAAATTATATTAGAGATTATCTCCATAAAGCTAGTTGCAAAAT	840
T1A_16	TCTGAGATTAAAGAGAAAAAATTATATTAGAGATTATCTCCATAAAGCTAGTTGCAAAAT	839
T1A_10	TCTGAGATTAAAGAGAAGAAATTATATTAGAGATTATCTCCATAAAGCTAGTTGCAAAAT	839
T1A_08	TCTGAGATTAAAGAGAAGAAATTATATTAGAGATTATCTCCATAAAGCTAGTTGCAAAAT	839
T1A_05	TCTGAGATTAAAGAGAAGAAATTATATTAGCAATTATCTCCATAAAGCTAGTTGCAAAAT ***** *****	839
T1B_01	AGTTGATTTAGCAATTGAAAATCAAGTAGAAACTATTGTAATTGGAGATATAAAAAATAT	900
T1A_16	AGTTGATTTAGCAATTGAAAATCAAGTAGAAACTATTGTAATTGGAGATATAAAAAATAT	899
T1A_10	AGTTGATTTAGCAATTGAAAATCAAGTAGAAACTATTGTAATTGGAGATATAAAAAATAT	899
T1A_08	AGTTGATTTAGCAATTGAAAATCAAGTAGAAACTATTGTAATTGGAGATATAAAAAATAT	899
T1A_05	AGTTGATTTAGCAATTGAAAATCAAGTAGAAACTATTGTAATCGGAGATATAAAAAATAT ***** *****	899
T1B_01	TAAACAATGCAGCAAACCTAAATCTTTTGTCCAAATACCAATCCAGAGATTAAAAAAT	960
T1A_16	TAAACAATGCAGCAAACCTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAAT	959
T1A_10	TAAAAAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAAT	959
T1A_08	TAAACAATGCAGCAATCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAAT	959
T1A_05	TAAACAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAAT	959

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**** *****
T1B_01      AATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTATAC      1020
T1A_16      AATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTATAC      1019
T1A_10      AATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTATAC      1019
T1A_08      AATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTATAC      1019
T1A_05      AATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTATAC      1019
*****

T1B_01      TTCTGGGTGTAGTTCAGTAGATCTGGAAAAATAAATAAAAAGTAACTATGATAAATCCAG      1080
T1A_16      TTCTGGGTGTAGTTCAGTAGATCTGGAAAAATAAATAAAAAGTAACTATGATAAATCCAG      1079
T1A_10      TTCTGGGTGTAGTTCAGTAGATCTGGAAAAATAAATAAAAAGTAACTATGATAAATCCAG      1079
T1A_08      TTCTGGGTGTAGTTCAGTAGATCTGGAAAAATAAATAAAAAGTAACTATGATAAATCCAG      1079
T1A_05      TTCTGGGTGTAGTTCAGTAGATCTGGAAAAATAAATAAAAAGTAACTATGATAAATCCAG      1079
*****

T1B_01      AAGAATTGCTAGAGGTCTCTTTAAAATAACGAGGGCCTATTAATTAATGCTGATCAGAA      1140
T1A_16      AAGAATTACTAGAGGTCTCTTTAAAATAACGAGGGCCTATTAATTAATGCTGATCAGAA      1139
T1A_10      AAGAATTACTAGAGGTCTCTTTAAAATAACGAGGGCCTATTAATTAATGCTGATCAGAA      1139
T1A_08      AAGAATTACTAGAGGTCTCTTTAAAATAACGAGGGCCTATTAATTAATGCTGATCAGAA      1139
T1A_05      AAGAATTACTAGAGGTCTCTTTAAAATAACGAGGGCCTATTAATTAATGCTGATCAGAA      1139
*****

T1B_01      TGGTAGTTTAAATATACTTCGTAAATACCATAACGATAAATGTATTCTCAGACCTATCAA      1200
T1A_16      TGGTAGTTTAAATATACTTCGTAAATACCATAACGATAAATGTATTCTCAGACCTATCAA      1199
T1A_10      TGGTAGCTTAAATATACTTCGTAAATACCATAACGATAAATGTATTCTCAGACCTATCAA      1199
T1A_08      TGGTAGCTTAAATATACTTCGTAAATACCATAACGATAAATGTATTCTCAGACCTATCAA      1199
T1A_05      TGGTAGCTTAAATATACTTCGTAAATACCATAAAGATAAATGTATTCTCAGACCTATCAA      1199
*****

T1B_01      AGAGGCAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCTAA      1255
T1A_16      AGAGGCAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCTAA      1254
T1A_10      AGAGGCAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCTAA      1254
T1A_08      AGAGGCAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCTAA      1254
T1A_05      AGAGGCAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCTAA      1254
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Type 2A

CLUSTAL O(1.2.1) multiple sequence alignment

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T1A_10      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T1A_05      ATGCGATTATCATTTAAATTC AACCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T1A_16      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T1A_08      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T2A_09      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T2A_06      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T2A_04      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
*****

T1A_10      GAATTAGCCTGGCATAATTAGTAAACTATATAATACAGTCAATTATGAGGTTAAAAACAAT      120
T1A_05      GAATTAGCCTGGCATTGCTCTAAATTATATAATATAGTCAATTATCAGATTAATAATAAT      120
T1A_16      GAATTAGCCTGGCATTGCTCTAAATTATATAATACAGTCAATTATCAGATTAATAATAAT      120
T1A_08      GAATTAGCCTGGCATTGCTCTAAATTATATAATATAGTCAATTATCAGATTAATAATAAT      120
T2A_09      GAATTAGCCTGGCATTGCTCTAAATTATATAATACAGTCAATTATCAGATTAATAATAAT      120
T2A_06      GAATTAGCCTGGCATTGCTCTAAATTATATAATACAGTCAATTATCAGATTAATAATAAT      120
T2A_04      GAATTAGCCTGGCATTGCTCTAAATTATATAATATAGTCAATTATCAGATTAATAATAAT      120
*****: ; *** ***** ***** * . ***** **

T1A_10      AAAGATATAAAAAGCTGTCTATACTGAATTAGAACTAGATATAAAAAAATAACTGGCATAAT      180
T1A_05      AAAGATGTA AAAAGCTGTCTATACTGAATTAGAACTAGATATAAAAAAATAACTGGCATAAT      180
T1A_16      AAAGATGTA AAAAGCTGTCTATACTGAATTAGAACTAGATATAAAAAAATAACTGGCATAAT      180
T1A_08      AAAGATGTA AAAAGCTGTCTATACTGAATTAGAACTAGATATAAAAAAATAACTGGCATAAT      180
T2A_09      AAAGATGTA AAAAGCTGTCTATACTGAATTAGAACTAGATATAAAAAAATAACTGGCATAAT      180
T2A_06      AAAGATGTA AAAAGCTGTCTATACTGAATTAGAACTAGATATAAAAAAATAACTGGCATAAT      180
T2A_04      AAAGATGTA AAAAGCTGTCTATACTGAATTAGAACTAGATATAAAAAAATAACTGGCATAAT      180
***** . *****

T1A_10      GACTACCTTCACTCCCAT AACAGACAGCAGGCATTAAGCAGTTAGCTCAGGACTGGAAA      240
T1A_05      GACTACCTTCACTCCCAT AACAGACAGCAGGCATTAAGCAGTTAGCTCAGGACTGGAAA      240
T1A_16      GACTACCTTCACTCCCAT AACAGACAGCAGGCATTAAGCAGTTAGTTCAGGACTGGAAA      240
T1A_08      GACTACCTTCACTCCCAT AACAGACAACAGGCATTAAGCAGTTAGCTCAGGACTGGAAA      240
T2A_09      GACTACCTTCACTCCCAT AACAGACAACAGGCATTAAGCAGTTAGCTCAGGACTGGAAA      240
T2A_06      GACTACCTTCACTCCCAT AACAGACAGCAGGCATTAAGCAGTTAGCTAAGGACTGGAAA      240
T2A_04      GACTACCTTCACTCCCAT AACAGACAACAGGCATTAAGCAGTTAGCTCAGGACTGGAAA      240
***** . *****

T1A_10      AGTTTTTTAATTCTCTCAAAGATTATAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA      300
T1A_05      AGTTTTTTAATTCTCTCAAAGATTATAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA      300
T1A_16      AGTTTTTTAATTCTCTCAAAGATTATAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300
T1A_08      AGTTTTTTAATTCTCTCAAAGATTATAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA      300
T2A_09      AGTTTTTTAATTCTCTCAAAGATTATAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA      300
T2A_06      AGTTTTTTAATTCTCTCAAAGATTATAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA      300
T2A_04      AGTTTTTTAATTCTCTCAAAGATTATAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA      300
*****: *****

T1A_10      GGATCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
T1A_05      GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
T1A_16      GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
T1A_08      GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
T2A_09      GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
T2A_06      GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
T2A_04      GGATCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
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T1A_10	GCTGTTAGAATTAAGATAACAAATTACTCTTATCCTTATCTAAAAAGATAACAATCTAAA	420
T1A_05	GCTGTTAGAATTAAGATAACAAATTACTCTTATCCTTATCTAAAAAGATAACAATCTAAA	420
T1A_16	GCTGTTAGAATTAGAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAGTCTAAA	420
T1A_08	GCTGTTAGGATTAGAGATAACAAATTACTCTTATCCTTATCTAAAAAGATAACAATCTAAA	420
T2A_09	GCTGTTAGAATTAAGATAACAAATTACTCTTATCCTTATCTAAAAAGATAACAATCTAAA	420
T2A_06	GCTGTTAGAATTAAGATAACAAATTACTCTTATCCTTATCTAAAAAGATAACAATCTAAA	420
T2A_04	GCTGTTAGGATTAGAGATAACAAATTACTCTTATCCTTATCTAAAAAGATAACAATCTAAA *****.****.*****.*****.*****.*****.*****.*****	420
T1A_10	TATAATGTGAAAG-----	433
T1A_05	TATAATGTGAAAA-----	433
T1A_16	TATAATGTGAAGG-----	433
T1A_08	TATAATGTGAAGG-----	433
T2A_09	TATAATGTGAAGGTCCTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA	480
T2A_06	TATAATGTGAAGGTCCTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA	480
T2A_04	TATAATGTGAAGGTCCTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA *****.*****.	480
T1A_10	-----CTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA	476
T1A_05	-----GCCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA	476
T1A_16	-----CTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA	476
T1A_08	-----CTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA	476
T2A_09	CCGTATTTCGATTTGGCCCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA	540
T2A_06	CCGTATTTCGATTTGGCCCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA	540
T2A_04	CCGTATTTCGATTTGGCCCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA *****.*****.*****.*****.*****.*****.*****.*****	540
T1A_10	TTTAGATGCTGGCCAGCAGATAAAGATTAAGCAGGACCGCATTTCTAAAAGATGGTATCT	536
T1A_05	TTTAGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCT	536
T1A_16	TTTAGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCT	536
T1A_08	TTTAGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCT	536
T2A_09	TTTAGATGCTGTCCAGCAGATAAAGATAAAGCAAGATCATATCTCTAAAAAATGGTATCT	600
T2A_06	TTTAGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCT	600
T2A_04	TTTAGATGCTTTACAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCT *****.*****.*****.*****.*****.*****.*****.*****	600
T1A_10	ACTAATCATCTATAAGGTCGAGGAAATAAAAAGAAAATAATAACCTAACATAATGGCAAT	596
T1A_05	ACTAATCATCTATAAGACCGAGGAAATAAAAAGAAAATAATAACCTAACATAATGGCAGT	596
T1A_16	CTTAATTATCTACAAAGTTAAAGAGGCCAAAAGAAAAGTAAGAAATCTAACATAATGGCAGT	596
T1A_08	ACTAATCATCTATAAGACCGAGGAAATAAAAAGAAAATAATAACCTAACATAATGGCAGT	596
T2A_09	CTTAATTATCTACAAAGTTAAAGAGGCCAAAAGAAAAGTAAGAAATCTAACATAATGGCAGT	660
T2A_06	CTTAATTATCTATAAGACCGAGGAAATAAAAAGAAAATAATAACCTAACATAATGGCAAT	660
T2A_04	ACTAATCGTCTATAAGAGCGAGGAAATAAAAAGAAAATAATAACCTAACATAATGGCAAT .****.****.***.***.***.***.*****.*****.*****.*****.*****.*****.*****	660
T1A_10	AGATCTAGGTCTTGATAAATTTGGCTACTTTAACATTTAAAAACAATCTGAGTGTTATAT	656
T1A_05	AGATCTAGGTCTTGATAAATTTGGCTACTTTAACATTTAAAAACAATCTGAGTGTTATAT	656
T1A_16	AGATCTAGGTCTTGATAAATTTGGCTACTTTAATATTTAAAAACAATCTGATTGTTATAT	656
T1A_08	TGATCTAGGTCTTGATAAATTTGGCTACTTTAACATTTAAAAACAATCTGATTGTTATAT	656
T2A_09	TGATTTAGGCCTTGATAAATTTAGCTGTACTAACATTTAAAGATAATCTGATTGTTATAT	720
T2A_06	AGATCTAGGTCTTGATAAATTTGGCTACTTTAACATTTAAAAACAATTTGATTGTTATAT	720
T2A_04	AGATCTAGGTCTTGATAAATTTGGCTACTTTAATATTTAAAAACAATCTGATTGTTATAT :***.***.*****.***.***.***.*****.*****.*****.*****.*****.*****	720
T1A_10	TATCAATGGTAAAACATTTAAATCCAAAAATCTTATTTTAAATAAAGAAATGCCAGACT	716
T1A_05	TATCAATGGTAAAACATTTAAATCCAAAAATCTTATTTTAAATAAAGAAATGCCAGACT	716

T1A_16	TATCAATGGTAAAAC TATTAATCCAAAAATCTTATTTTAATAAAGAAATTGCCAGACT	716
T1A_08	TATCAATGGTAAAAC TATTAATCCAAAAATCTTATTTTAATAAAGAAATTGCCAGACT	716
T2A_09	TATCAATGGTAAAAC TATTAATCCAAAAATCTTATTTTAATAAAGAAATTGCCAGACT	780
T2A_06	TATCAATGGTAAAAC TATTAATCCAAAAATCTTATTTTAATAAAGAAATTGCCAGACT	780
T2A_04	TATCAATGGTAAAAC TATTAATCCAAAAATCTTATTTTAATAAAGAAATTGCCAGACT *****	780
T1A_10	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	776
T1A_05	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	776
T1A_16	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	776
T1A_08	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	776
T2A_09	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	840
T2A_06	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	840
T2A_04	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA *****	840
T1A_10	ATATCTGAGATTAAGAGAAGAAATTATATTAGAGATTATCTCCATAAAGCTAGCTGCAA	836
T1A_05	ATATCTGAGATTAAGAGAAGAAATTATATTAGCAATTATCTCCATAAAGCTAGTTGCAA	836
T1A_16	ATATCTGAGATTAAGAGAAGAAATTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA	836
T1A_08	ATATCTGAGATTAAGAGAAGAAATTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA	836
T2A_09	ATATCTGAGATTAAGAGAAGAAATTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA	900
T2A_06	ATATCTGAGATTAAGAGAAGAAATTATATTAGCGATTATCTCCATAAAGCTAGTTGCAA	900
T2A_04	ATATCTGAGATTAAGAGAAGAAATTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA *****	900
T1A_10	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAAAATATTGTAATTGGAGATATAAAAAA	896
T1A_05	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAAAATATTGTAATCGGAGATATAAAAAA	896
T1A_16	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAAAATATTGTAATTGGAGATATAAAAAA	896
T1A_08	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAAAATATTGTAATTGGAGATATAAAAAA	896
T2A_09	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAAAATATTGTAATTGGAGATATAAAAAA	960
T2A_06	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAAAATATTGTAATCAGAGATATAAAAAA	960
T2A_04	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAAAATATTGTAATTGGAGATATAAAAAA *****	960
T1A_10	TATTAAAAAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAA	956
T1A_05	TATTAACAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAA	956
T1A_16	TATTAACAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAA	956
T1A_08	TATTAACAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAA	956
T2A_09	TATTAACAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAA	1020
T2A_06	TATTAACAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCAATCCAGAGATTAAAAAA	1020
T2A_04	TATTAACAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAA *****	1020
T1A_10	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1016
T1A_05	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1016
T1A_16	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1016
T1A_08	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1016
T2A_09	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1080
T2A_06	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1080
T2A_04	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA *****	1080
T1A_10	TACTTCTGGGTGTA-----	1030
T1A_05	TACTTCTGGATGTA-----	1030
T1A_16	TACTTCTGGATGTA-----	1030
T1A_08	TACTTCTGGATGTA-----	1030
T2A_09	TACTTCCGATGTACTAAAGCTTTTAAATTTTAAATTACGCAAGGAAAGCTTTAGTATGACC	1140

T2A_06	TACTTCTGGGTGACTAAAGCTTTTAATTTTAATTACGCAAGGTAAGCTTTAGTATGACC	1140
T2A_04	TACTTCTGGATGACTAAAGCTTTTAATTTTAATTACGCAAGGTAAGCTTTAGTATGACC ***** **.*****	1140
T1A_10	-----GTTTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA	1072
T1A_05	-----GTTTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA	1072
T1A_16	-----GTTTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA	1072
T1A_08	-----GTTTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA	1072
T2A_09	GTATTTCGATTTGGCCGCTTTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA	1200
T2A_06	GTATTTCGATTTGGCCGCTTTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA	1200
T2A_04	GTATTTCGATTTGGCCGCTTTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA *****	1200
T1A_10	AATCCAGAAGAATTACTAGAGGTCCTTTAAAATAACGAGGGCTATTAATTAATGCTG	1132
T1A_05	AATCCAGAAGAATTACTAGAGGTCCTTTAAAATAACGAGGGCTATTAATTAATGCTG	1132
T1A_16	AATCCAGAAGAATTACTAGAGGTCCTTTAAAATAACGAGGGCTATTAATTAATGCTG	1132
T1A_08	AATCCAGAAGAATTACTAGAGGTCCTTTAAAATAACGAGGGCTATTAATTAATGCTG	1132
T2A_09	AATCCAGAAGAATTACCAGAGGTCCTTTAAAATAACGAGGGCTATTAATTAATGCTG	1260
T2A_06	AATCCAGAAGAATTACCAGAGGTCCTTTAAAATAACGAGGGCTATTAATTAATGCTG	1260
T2A_04	AATCCAGAAGAATTACTAGAGGTCCTTTAAAATAACGAGGGCTATTAATTAATGCTG ***** **.*****	1260
T1A_10	ATCAGAATGGTAGCTTTAATATACTTCGTAATACCATAACGATAAATGTATTCTCAGAC	1192
T1A_05	ATCAGAATGGTAGCTTTAATATACTTCGTAATACCATAACGATAAATGTATTCTCAGAC	1192
T1A_16	ATCAGAATGGTAGCTTTAATATACTTCGTAATACCATAACGATAAATGTATTCTCAGAC	1192
T1A_08	ATCAGAATGGTAGCTTTAATATACTTCGTAATACCATAACGATAAATGTATTCTCAGAC	1192
T2A_09	ATCAGAATGGTAGCTTTAATATACTTCGTAATACCATAACGATAAATGTATTCTCAGAC	1320
T2A_06	ATCAGAATGGTAGCTTTAATATACTTCGTAATACCATAACGATAAATGTATTCTCAGAC	1320
T2A_04	ATCAGAATGGTAGCTTTAATATACTTCGTAATACCATAACGATAAATGTATTCTCAGAC ***** **.*****	1320
T1A_10	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1252
T1A_05	CCATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1252
T1A_16	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1252
T1A_08	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1252
T2A_09	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1380
T2A_06	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1380
T2A_04	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT * *****	1380
T1A_10	AA 1254	
T1A_05	AA 1254	
T1A_16	AA 1254	
T1A_08	AA 1254	
T2A_09	AA 1382	
T2A_06	AA 1382	
T2A_04	AA 1382 **	

Type 2A*

CLUSTAL O(1.2.1) multiple sequence alignment

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T2A_09      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T2A*_13     ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
*****

T2A_09      GAATTAGCCTGGCATTGCTCTAAAT TATAATAACAGTCAATTATCAGATTA AAAATAAT      120
T2A*_13     GAATTAGCCTGGCATTGCTCTAAAT TATAATAATAGTCAATTATCAGATTA AAAATAAT      120
*****

T2A_09      AAAGATGTAAAAGCTGCTTATACTGAATTAGAACTAGATATAAAAATAACTGGCATAAT      180
T2A*_13     AAAGATGTAAAAGCTGCTTATACTGAA-----                          147
*****

T2A_09      GACTACCTTCACTCCCAT AACAGACAACAGGCATTAAGCAGTTAGCTCAGGACTGGAAA      240
T2A*_13     -----                          147

T2A_09      AGTTTTTTTTTATCTCTCAAAGATTATAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA      300
T2A*_13     -----                          147

T2A_09      GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTTACCAATTTA      360
T2A*_13     -----TATGAACAGTAATCCCTGTGAAATAATTTTTACCAATTTA      187
*****

T2A_09      GCTGTTAGAATTAAGATAACAAAT TACTCTTATCCTTATCTAAAAAGATACAATCTAAA      420
T2A*_13     GCTGTTAGAATTAGAGATAACAAAT TACTCTTATCCTTATCTAAAAAGATACAGTCTAAA      247
*****.*****

T2A_09      TATAATGTGAAGGTCAC TAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA      480
T2A*_13     TATAATGTGAAGGTCAC TAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA      307
*****

T2A_09      CCGTATTCGATTTGGCCCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA      540
T2A*_13     CCGTATTCGATTTGGCCCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA      367
*****

T2A_09      TTTAGATGCTGTCCAGCAGATAAAGATAAAGCAAGATCATATCTCTAAAAAATGGTATCT      600
T2A*_13     TTTAGATGCTGTCCAACAGATAAAGATAAAGCAAGATCATATCTCTAAAAAATGGTATCT      427
*****.*****

T2A_09      CTTAAT TATCTACAAAGTTAAAGAGGCAAAGAAAGTAAGAAATCTAACATAATGGCAGT      660
T2A*_13     ACTAATCATCTACAAAGTTAAAGAGGCAAAGAAAGTAAGAAATCTAACATAATGGCAGT      487
.***

T2A_09      TGATTTAGGCCTTGATAACTTAGCTGTACTAACATTTAAAGATAATCTCGATTGTTATAT      720
T2A*_13     AGATCTAGGCTTTGATAATTTGGCTACTTTAACATTTAAAAACAATCTCGAGTGTATAT      547
:***

T2A_09      TATCAATGGTAAAAC TATTAATCCAAAAATCTTATTTTAATAAAGAAATTGCCAGACT      780
T2A*_13     TATCAATGGTAAAAC TATTAATCCAAAAATCTTATTTTAATAAAGAAATTGCCAGACT      607
*****

T2A_09      ACAAAGCATTAGAATTAGGCAGTTAGCTACCAGTAAATTAGAGATACTAACGAATAAA      840

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T2A*_13	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA *****	667
T2A_09	ATATCTGAGATTAAGAGAAGAAATATATTTAGAGATTATCTCCATAAAGCTAGTTGCAA	900
T2A*_13	ATATCTGAGATTAAGAGAAGAAATATATTTAGAGATTATATCCATAAAGCTAGTTGCAA *****	727
T2A_09	AATAGTTGATTTAGCAATGAAAATCAAGTAGAACTATTGTAATTGGAGATATAAAAAA	960
T2A*_13	AATAGTTGATTTAGCAATGAAAATCAAGTAGAACTATTGTAATCGGAGACATAAAAAA *****	787
T2A_09	TATTAACAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAA	1020
T2A*_13	TATTAACAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAA *****	847
T2A_09	ATTAATTGAATACAAAGTTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1080
T2A*_13	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA *****	907
T2A_09	TACTTCCGGATGTAATAAGCTTTTAATTTAATTACGCAAGGAAAGCTTTAGTATGACC	1140
T2A*_13	TACTTCCGGATGTAATAAGCTTTTAATTTAGTTACGCAAGGTAAGCTTTAGTATGACC *****	967
T2A_09	GTATTCGATTTGGCCGCTCTTCAGTAGATCTGGAAAAATAAATAAAAGTAACTATGATA	1200
T2A*_13	GTATTCGATTTGGCCGCTATTCAGTAGATCTGGAAAAATAAATAAAAGCAATTATGATA *****	1027
T2A_09	AATCCAGAAGAATTACCAGAGGTCCTTTAAACTAACGAGGGCTATTAATTAATGCTG	1260
T2A*_13	AATCCAGAAGAATTACTAGGGGTCCTTTAAACTAACGAGGGCTTATTAATTAATGCTG *****	1087
T2A_09	ATCAGAATGGTAGTTTTAATATACTTCGTAATACCATAACGATAAATGTATTCTCAGAC	1320
T2A*_13	ATCAGAATGGTAGTTTTAATATACTTCGTAATACCATAACGATAAATGTATTCTCAGAC *****	1147
T2A_09	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1380
T2A*_13	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT *****	1207
T2A_09	AA 1382	
T2A*_13	AA 1209 **	

Type 2B/2B*/2C

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T2C_03      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T2C_02      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T2A_09      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T2B_17      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T2B*_18     ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T2A_06      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T2A_04      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T2B_20      ATGCGATTATCATTTAAATTC AACCCATAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T2B_19      ATGCGATTATCATTTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
*****

T2C_03      GAATTAGCCTGGCATTGCTCT AAATTTATATAATACAGTCAATTATCAGATTAATAATAAT      120
T2C_02      GAATTAGCCTGGCATTGCTCT AAATTTATATAATACAGTCAATTATCAGATTAATAATAAT      120
T2A_09      GAATTAGCCTGGCATTGCTCT AAATTTATATAATACAGTCAATTATCAGATTAATAATAAT      120
T2B_17      GAATTAGCCTGGCATTGCTCT AAATTTATATAATACAGTCAATTATCAGATTAATAATAAT      120
T2B*_18     GAATTAGCCTGGCATTGCTCT AAATTTATATAATACAGTCAATTATCAGATTAATAATAAT      120
T2A_06      GAATTAGCCTGGCATTGCTCT AAATTTATATAATACAGTCAATTATCAGATTAATAATAAT      120
T2A_04      GAATTAGCCTGGCATTGCTCT AAATTTATATAATACAGTCAATTATCAGATTAATAATAAT      120
T2B_20      GAATTAGCCTGGCATTGCTCT AAATTTATATAATACAGTCAATTATCAGATTAATAATAAT      120
T2B_19      GAATTAGCCTGGCATTGCTCT AAATTTATATAATACAGTCAATTATCAGATTAATAATAAT      120
*****

T2C_03      AAAGATGTAAGCTGTCTATACT GAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
T2C_02      AAAGATGTAAGCTGTCTATACT GAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
T2A_09      AAAGATGTAAGCTGTCTATACT GAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
T2B_17      AAAGATGTAAGCTGTCTATACT GAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
T2B*_18     AAAGATGTAAGCTGTCTATACT GAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
T2A_06      AAAGATGTAAGCTGTCTATACT GAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
T2A_04      AAAGATGTAAGCTGTCTATACT GAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
T2B_20      AAAGATGTAAGCTGTCTATACT GAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
T2B_19      AAAGATGTAAGCTGTCTATACT GAATTAGAACTAGATATAAAAAATAACTGGCATAAT      180
*****

T2C_03      GACTACCTTCACTCCATAACAG ACAACAGGCATTAAGCAGTTAGCTAAGGACTGGAAA      240
T2C_02      GACTACCTTCACTCCATAACAG ACAACAGGCATTAAGCAGTTAGCTAAGGACTGGAAA      240
T2A_09      GACTACCTTCACTCCATAACAG ACAACAGGCATTAAGCAGTTAGCTAAGGACTGGAAA      240
T2B_17      GACTACCTTCACTCCATAACAG ACAACAGGCATTAAGCAGTTAGCTAAGGACTGGAAA      240
T2B*_18     GACTACCTTCACTCCATAACAG ACAACAGGCATTAAGCAGTTAGCTAAGGACTGGAAA      240
T2A_06      GACTACCTTCACTCCATAACAG ACAACAGGCATTAAGCAGTTAGCTAAGGACTGGAAA      240
T2A_04      GACTACCTTCACTCCATAACAG ACAACAGGCATTAAGCAGTTAGCTAAGGACTGGAAA      240
T2B_20      GACTACCTTCACTCCATAACAG ACAACAGGCATTAAGCAGTTAGCTAAGGACTGGAAA      240
T2B_19      GACTACCTTCACTCCATAACAG ACAACAGGCATTAAGCAGTTAGCTAAGGACTGGAAA      240
*****

T2C_03      AGTTTTTTTTTATCTCTCAAAG ATTTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300
T2C_02      AGTTTTTTTTTATCTCTCAAAG ATTTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300
T2A_09      AGTTTTTTTTTATCTCTCAAAG ATTTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300
T2B_17      AGTTTTTTTAAATCTCTCAAAG ATTTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300
T2B*_18     AGTTTTTTTAAATCTCTCAAAG ATTTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300
T2A_06      AGTTTTTTTAAATCTCTCAAAG ATTTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300
T2A_04      AGTTTTTTTAAATCTCTCAAAG ATTTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300
T2B_20      AGTTTTTTTAAATCTCTCAAAG ATTTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300
T2B_19      AGTTTTTTTAAATCTCTCAAAG ATTTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300

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T2C_03 GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA 360
T2C_02 GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA 360
T2A_09 GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA 360
T2B_17 GGGTCACCGAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATCTG 360
T2B*_18 GAGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA 360
T2A_06 GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA 360
T2A_04 GGATCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA 360
T2B_20 GGATCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA 360
T2B_19 GGATCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA 360
* *****

T2C_03 GCTGTTAGAATTAAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA 420
T2C_02 GCTGTTAGAATTAGAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA 420
T2A_09 GCTGTTAGAATTAAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA 420
T2B_17 GCTGTTAGAATTAAGATAATAAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA 420
T2B*_18 GCTGTTAGAATTAAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA 420
T2A_06 GCTGTTAGAATTAAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA 420
T2A_04 GCTGTTAGGATTAGAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA 420
T2B_20 GCTGTTAGAATTAGAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA 420
T2B_19 GCTGTTAGAATTAGAGATAATAAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA 420
*****

T2C_03 TATAATGTGAAG----- 432
T2C_02 TATAATGTGAAG----- 432
T2A_09 TATAATGTGAAGGTCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA 480
T2B_17 TATAATGTGAAGGTCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA 480
T2B*_18 TATAATGTGAAGGTCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA 480
T2A_06 TATAATGTGAAGGTCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA 480
T2A_04 TATAATGTGAAGGTCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA 480
T2B_20 TATAATGTGAAGGTCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA 480
T2B_19 TATAATGTGAAGGTCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA 480
*****

T2C_03 -----GCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA 476
T2C_02 -----GCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA 476
T2A_09 CCGTATTCGATTTGGCCCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA 540
T2B_17 CCGTATTCGATTTGGCCCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA 540
T2B*_18 CCGTATTCGATTTGGCC-TCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA 539
T2A_06 CCGTATTCGATTTGGCCCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA 540
T2A_04 CCGTATTCGATTTGGCCCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA 540
T2B_20 CCGTATTCGATTTGGCCCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA 540
T2B_19 CCGTATTCGATTTGGCCCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA 540
*****

T2C_03 TTTAGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCT 536
T2C_02 TTTAGATGCTGTCCAGCAGATAAAGATAAAGCAAGATCATATCTCTAAAAAATGGTATCT 536
T2A_09 TTTAGATGCTGTCCAGCAGATAAAGATAAAGCAAGATCATATCTCTAAAAAATGGTATCT 600
T2B_17 TTTAGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCT 600
T2B*_18 TTTAGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCT 599
T2A_06 TTTAGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCT 600
T2A_04 TTTAGATGCTTTACAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCT 600
T2B_20 TTTAGATGCTTTACAGCAGATAAAGGTTAAGCAGGACCGTATTTCTAAAAGATGGTATCT 600
T2B_19 TTTAGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGATTTCTAAAAGATGGTATCT 600
***** * ***** * ***** * * * * *****

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T2C_03	CTTAATTATCTACAAAGTTAAAGAGGCAAAGAAAGTAAGAAATCTAACATAATGGCAGT	596
T2C_02	CTTAATTATCTACAAAGTTAAAGAGGCAAAGAAAGTAAGAAATCTAACATAATGGCAGT	596
T2A_09	CTTAATTATCTACAAAGTTAAAGAGGCAAAGAAAGTAAGAAATCTAACATAATGGCAGT	660
T2B_17	ACTAATCATCTACAAAGTTAAAGAGGCAAAGAAATAAGAAATCTAACATAATGGCAGT	660
T2B*_18	CTTAATTATCTACAAAGTTAAAGAGGCAAAGAAAGTAAGAAATCTAACATAATGGCAGT	659
T2A_06	CTTAATTATCTATAAGACCGAGGAAAATAAGAAATAATAAACCCCTAACATAATGGCAAT	660
T2A_04	ACTAATCGTCTATAAGAGCGAGGAAAATAAGAAATAATAAACCCCTAACATAATGGCAAT	660
T2B_20	ACTAATTATCTATACGACCGAGGAAAATAAGAAATAATAAACCCCTAACATAATGGCAGT	660
T2B_19	ACTAATCATCTATAAGGTCGAGGAAAATAAGAAATAATAAATCTAACATAATGGCAGT **** * * * * * * * * * * *	660
T2C_03	TGATCTAGGTCTTGATAAATTTGGCTACTTTAACATTTAAAAACAATCTCGATTGTTATAT	656
T2C_02	TGATTTAGGCCTTGATAACTTAGCTGTACTAACATTTAAAGATAAATCTCGATTGTTATAT	656
T2A_09	TGATTTAGGCCTTGATAACTTAGCTGTACTAACATTTAAAGATAAATCTCGATTGTTATAT	720
T2B_17	AGATCTAGGTCTTGATAAATTTGGCTACTTTAACATTTAAAAACAATCTCGATTGTTATAT	720
T2B*_18	TGATCTAGGTCTTGATAAATTTGGCTACTTTAACATTTAAAAACAATCTCGATTGTTATAT	719
T2A_06	AGATCTAGGTCTTGATAAATTTGGCTACTTTAACATTTAAAAACAATTTTGATTGTTATAT	720
T2A_04	AGATCTAGGTCTTGATAAATTTGGCTACTTTAATATTTAAAAACAATCTCGATTGTTATAT	720
T2B_20	TGATCTAGGTCTTGATAAATTTGGCTACTTTAACATTTAAAAACAATCTCGATTGTTATAT	720
T2B_19	AGATATAGGTCTTGATAAATTTGGCTACTTTAACATTTAAAAACAATCTCGATTGTTATAT *** ***** ** * * * * * * * * * * *	720
T2C_03	TATCAATGGTAAAACCTATTTAAATCCAAAAATTCCTATTTTAAATAAGAAATGGCCAGACT	716
T2C_02	TATCAATGGTAAAACCTATTTAAATCCAAAAATTCCTATTTTAAATAAGAAATGGCCAGACT	716
T2A_09	TATCAATGGTAAAACCTATTTAAATCCAAAAATTCCTATTTTAAATAAGAAATGGCCAGACT	780
T2B_17	TATCAATGGTAAAACCTATTTAAATCCAAAAATTCCTATTTTAAATAAGAAATGGCCAGACT	780
T2B*_18	TATCAATGGTAAAACCTATTTAAATCCAAAAATTCCTATTTTAAATAAGAAATGGCCAGACT	779
T2A_06	TATCAATGGTAAAACCTATTTAAATCCAAAAATTCCTATTTTAAATAAGAAATGGCCAGACT	780
T2A_04	TATCAATGGTAAAACCTATTTAAATCTAAAAATTCCTATTTTAAATAAGAAATGGCCAGACT	780
T2B_20	TATCAATGGTAAAACCTATTTAAATCCAAAAATTCCTATTTTAAATAAGAAATGGCCAGACT	780
T2B_19	TATCAATGGTAAAACCTATTTAAATCCAAAAATTCCTATTTTAAATAAGAAATGGCCAGACT *****	780
T2C_03	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	776
T2C_02	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	776
T2A_09	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	840
T2B_17	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	840
T2B*_18	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	839
T2A_06	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	840
T2A_04	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	840
T2B_20	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA	840
T2B_19	ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA *****	840
T2C_03	ATATCTGAGATTTAAAGAGAAAAAATTTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA	836
T2C_02	ATATCTGAGATTTAAAGAGAAAAAATTTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA	836
T2A_09	ATATCTGAGATTTAAAGAGAAAAAATTTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA	900
T2B_17	ATATCTGAGATTTAAAGAGAAAAAATTTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA	900
T2B*_18	ATATCTGAGATTTAAAGAGAAAAAATTTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA	899
T2A_06	ATATCTGAGATTTAAAGAGAAAAAATTTATATTAGCGATTATCTCCATAAAGCTAGTTGCAA	900
T2A_04	ATATCTGAGATTTAAAGAGAAAAAATTTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA	900
T2B_20	ATATCTGAGATTTAAAGAGAAAAAATTTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA	900
T2B_19	ATATCTGAGATTTAAAGAGAAAAAATTTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA *****	900
T2C_03	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAACTATTGTAATCGGAGATATAAAAAA	896
T2C_02	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAACTATTGTAATCGGAGATATAAAAAA	896

T2A_09	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAACTATTGTAATTGGAGATATAAAAAA	960
T2B_17	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAACTATTGTAATTGGAGATATAAAAAA	960
T2B*_18	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAACTATTGTAATTGGAGATATAAAAAA	959
T2A_06	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAACTATTGTAATCAGAGATATAAAAAA	960
T2A_04	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAACTATTGTAATTGGAGATATAAAAAA	960
T2B_20	AATAGTTAATTTAGCAGTTGAAAATCAAGTAGAACTATTGTAATTGGAGATATAAAAAA	960
T2B_19	AATAGTTGATTTAGCAATTGAAAATCAAGTAGAACTATTGTAATCGGAGATATAAAAAA *****	960
T2C_03	TATTAACAATGCAGCAAACCTAAATCTTTTGTCCAAATACCGATTCAGAGATTAAAAAA	956
T2C_02	TATTAACAATGCAGCAAACCTAAATCTTTTGTCCAAATACCGATTCAGAGATTAAAAAA	956
T2A_09	TATTAACAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCGATTCAGAGATTAAAAAA	1020
T2B_17	TATTAACAATGCAGCAAACCTAAATCTTTTGTCCAAATACCGATTCAGAGATTAAAAAA	1020
T2B*_18	TATTAACAATGCAGCAAACCTAAATCTTTTGTCCAAATACCGATCA-GAGATTAAAAAA	1018
T2A_06	TATTAACAATGCAGCAAGCTTAAATCTTTTGTCCAAATACCAATCCAGAGATTAAAAAA	1020
T2A_04	TATTAACAATGCAGCAAACCTAAATCTTTTGTCCAAATACCGATTCAGAGATTAAAAAA	1020
T2B_20	TATTAACAATGCAGCAAACCTAAATCTTTTGTCCAAATACCGATTCAGAGATTAAAAAA	1020
T2B_19	TATTAACAATGCAGCAAACCTAAATCTTTTGTCCAAATACCGATTCAGAGATTAAAAAA *****	1020
T2C_03	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1016
T2C_02	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1016
T2A_09	ATTAATTGAATACAAAGTTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1080
T2B_17	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGGAATTGATGAAAGCTA	1080
T2B*_18	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1078
T2A_06	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1080
T2A_04	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1080
T2B_20	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA	1080
T2B_19	ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGAAATTGATGAAAGCTA *****	1080
T2C_03	TACTTCTGGGTGTACTAAAGCTTTTAAATTTTAAATTACGCAAGGTAAGCTTTAGTATGACC	1076
T2C_02	TACTTCTGGATGTACTAAAGCTTTTAAATTTTAAATTACGCAAGGTAAGCTTTAGTATGACC	1076
T2A_09	TACTTCCGGATGTACTAAAGCTTTTAAATTTTAAATTACGCAAGGAAAGCTTTAGTATGACC	1140
T2B_17	TACTTCTGGATGTA-----	1094
T2B*_18	TACTTCTGGGTGTA-----	1092
T2A_06	TACTTCTGGGTGTACTAAAGCTTTTAAATTTTAAATTACGCAAGGTAAGCTTTAGTATGACC	1140
T2A_04	TACTTCTGGATGTACTAAAGCTTTTAAATTTTAAATTACGCAAGGTAAGCTTTAGTATGACC	1140
T2B_20	TACTTCTGGGTGT-----	1093
T2B_19	TACTTCTGGATGT----- ***** ** **	1093
T2C_03	GTATTCGATTTGGCCGCTATTCAGTAGATCTGGAAAAAATAAATAAAAGTAACTATGATA	1136
T2C_02	GTATTCGATTTGGCCGCTATTCAGTAGATCTGGAAAAAATAAATAAAAGTAACTATGATA	1136
T2A_09	GTATTCGATTTGGCCGCTCTTCAGTAGATCTGGAAAAAATAAATAAAAGTAACTATGATA	1200
T2B_17	-----GTTTCAGTAGATCTGGAAAAAATAAATAAAAGTAACTATGATA	1136
T2B*_18	-----GTTTCAGTAGATCTGGAAAAAATAAATAAAAGTAACTATGATA	1134
T2A_06	GTATTCGATTTGGCCGCTATTCAGTAGATCTGGAAAAAATAAATAAAAGTAACTATGATA	1200
T2A_04	GTATTCGATTTGGCCGCTGTTTCAGTAGATCTGGAAAAAATAAATAAAAGTAACTATGATA	1200
T2B_20	-----AGTTTCAGTAGATCTGGAAAAAATAAATAAAAGTAACTATGATA	1136
T2B_19	-----AGTTTCAGTAGATCTGGAAAAAATAAATAAAAGTAACTATGATA *****	1136
T2C_03	AATCCAGAAGAATTACTAGAGGTCCTTTAAACTAACGAGGGCCTATTAATTAATGCTG	1196
T2C_02	AATCCAGAAGAATTACTAGAGGTCCTTTAAACTAACGAGGGCCTATTAATTAATGCTG	1196
T2A_09	AATCCAGAAGAATTACCAGAGGTCCTTTAAACTAACGAGGGCCTATTAATTAATGCTG	1260
T2B_17	AATCCAGAAGAATTACTAGAGGTCCTTTAAACTAACGAGGGCCTATTAATTAATGCTG	1196

T2B*_18	AATCCAGAAGAATTACTAGAGGTCTCTTTAAAAC TAACGAGGGCCTATTAATTAATGCTG	1194
T2A_06	AATCCAGAAGAATTACCAGAGGTCTCTTTAAAAC TAACGAGGGCCTATTAATTAATGCTG	1260
T2A_04	AATCCAGAAGAATTACTAGAGGTCTCTTTAAAAC TAACGAGGGCCTATTAATTAATGCTG	1260
T2B_20	AATCCAGAAGAATTACTAGAGGTCTCTTTAAAAC TAACGAGGGCCTATTAATTAATGCTG	1196
T2B_19	AATCCAGAAGAATTACTAGAGGTCTCTTTAAAAC TAACGAGGGCCTATTAATTAATGCTG	1196

T2C_03	ATCAGAATGGTAGTTTTAATATACTTCGTAAATATCATAACGATAAAATGTATTCTCAGAC	1256
T2C_02	ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC	1256
T2A_09	ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC	1320
T2B_17	ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC	1256
T2B*_18	ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC	1254
T2A_06	ATCAGAATGGTAGTTTTAATATACTTCGTAAATATCATAAGGATAAAATGTATTCTCAGAC	1320
T2A_04	ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC	1320
T2B_20	ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC	1256
T2B_19	ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC	1256

T2C_03	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1316
T2C_02	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1316
T2A_09	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1380
T2B_17	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1316
T2B*_18	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1314
T2A_06	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1380
T2A_04	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1380
T2B_20	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1316
T2B_19	CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT	1316

T2C_03	AA 1318	
T2C_02	AA 1318	
T2A_09	AA 1382	
T2B_17	AA 1318	
T2B*_18	AA 1316	
T2A_06	AA 1382	
T2A_04	AA 1382	
T2B_20	AA 1318	
T2B_19	AA 1318	
	**	

Type 3

CLUSTAL multiple sequence alignment by Kalign (2.0)

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T1A_16   ATGCGATTATCATTTAAATTCAAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT
T2A_09   ATGCGATTATCATTTAAATTCAAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT
T3_23    ATGCGATTATCATTTAAATTCAAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT
T3_22    ATGCGATTATCATTTAAATTCAAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT
T3_21    ATGCGATTATCATTTAAATTCAAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT
T3_12    ATGCGATTATCATTTAAATTCAAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT
T3_11    ATGCGATTATCATTTAAATTCAACCCATAAGCCATAAGCAATTAGTAATAATTAAT
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T1A_16   GAATTAGCCTGGCATTGCTCTAAATTATATAATACAGTCAATTATCAGATTAATAATAAT
T2A_09   GAATTAGCCTGGCATTGCTCTAAATTATATAATACAGTCAATTATCAGATTAATAATAAT
T3_23    GAATTAGCCTGGCATTGCTCTAAATTATATAATACAGTCAATTATCAGATTAATAATAAT
T3_22    GAATTAGCCTAGCATTGCTCTAAATTATATAATACAGTCAATTATCAGATTAATAATAAT
T3_21    GAATTAGCCTGGCATTGCTCTAAATTATATAATACAGTCAATTATCAGATTAATAATAAT
T3_12    GAATTAGCCTGGCATTGCTCTAAATTATATAATACAGTCAATTATCAGATTAATAATAAT
T3_11    GAATTAGCCTGGCATTGCTCTAAATTATATAATATAGTCAATTATCAGATTAATAATAAT
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T1A_16   AAAGATGTAAGCTGTCTATACTGAATTAGAACTAGATATAATAATAAAGCTGGCATAAT
T2A_09   AAAGATGTAAGCTGTCTATACTGAATTAGAACTAGATATAATAATAAAGCTGGCATAAT
T3_23    AAAGATGTAAGCTGTCTATACTGAATTAGAACTAGATATAATAATAAAGCTGGCATAAT
T3_22    AAAGATGTAAGCTGTCTATACTGAATTAGAACTAGATATAATAATAAAGCTGGCATAAT
T3_21    AAAGATGTAAGCTGTCTATACTGAATTAGAACTAGATATAATAATAAAGCTGGCATAAT
T3_12    AAAGATGTAAGCTGTCTATACTGAATTAGAACTAGATATAATAATAAAGCTGGCATAAT
T3_11    AAAGATGTAAGCTGTCTATACTGAATTAGAACTAGATATAATAATAAAGCTGGCATAAT
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T1A_16   GACTACCTTCACTCCCATAACAGACAGCAGGCATTAAGCAGTTAGCTCAGGACTGGAAA
T2A_09   GACTACCTTCACTCCCATAACAGACAGCAGGCATTAAGCAGTTAGCTCAGGACTGGAAA
T3_23    GACTACCTTCACTCCCATAACAGACAGCAGGCATTAAGCAGTTAGCTCAGGACTGGAAA
T3_22    GACTACCTTCACTCCCATAACAGACAGCAGGCATTAAGCAGTTAGCTCAGGACTGGAAA
T3_21    GACTACCTTCACTCCCATAACAGACAGCAGGCATTAAGCAGTTAGCTCAGGACTGGAAA
T3_12    GACTACCTTCACTCCCATAACAGACAGCAGGCATTAAGCAGTTAGCTCAGGACTGGAAA
T3_11    GACTACCTTCACTCCCATAACAGACAGCAGGCATTAAGCAGTTAGCTCAGGACTGGAAA
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T1A_16   AGTTTTTTTAAATTCTCTCAAAGATTATAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA
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T2A_09 AGTTTTTTTATTCTCTCAAAGATTATAAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA
T3_23 AGTTTTTTTAATTCACCTCAAAGATTATAAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA
T3_22 AGTTTTTTTAATTCACCTCAAAGATTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA
T3_21 AGTTTTTTTAATTCCTCTCAAAGATTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA
T3_12 AGTTTTTTTAATTCCTCTCAAAGATTATAAAAAAGAATCCTCAAAAATATAAGGGGCAGCCA
T3_11 AGTTTTTTTAATTCCTCTCAAAGATTATAAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA

T1A_16 GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTTACCAATTTA
T2A_09 GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTTACCAATTTA
T3_23 GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTTACCAATTTA
T3_22 GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTTACCAATTTA
T3_21 GGATCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTTACCAATCTG
T3_12 GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTTACCAATTTA
T3_11 GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTTACCAATTTA

T1A_16 GCTGTTAGAATTAGAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAGTCTAAA
T2A_09 GCTGTTAGAATTAAAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA
T3_23 GCTGTTAGAATTAGAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAGTCTAAA
T3_22 GCTGTTAGAATTAAAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA
T3_21 GCTATTAGAATTAAAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA
T3_12 GCTGTTAGAATTAAAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAATCTAAA
T3_11 GCTGTTAGAATTAGAGATAATAAATTACTCTTATCCTTATCTAAAAAGATACAGTCTAAA

T1A_16 TATAATGTGAAGG-----
T2A_09 TATAATGTGAAGGTCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGA
T3_23 TATAATGTGAAAAGCCTAAAGCTTTTAATTTATAA-----
T3_22 TATAATGTGAAGGTCACTAAAGCTTTTAATTTATAA-----
T3_21 TATAATGTGAAGGTCACTAAAGCTTTTAATTTATAA-----
T3_12 TATAATGTGAAAGTCACTAAAGCTTTTAATTTATAA-----
T3_11 TATAATGTGAAAGTCACTAAAGCTTTTAATTTATAA-----

T1A_16 -----CTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA
T2A_09 CCGTATTCGATTTGGCCCTCTTAATTTTGAGCTGCCTGAAGCAGTTCAAAGCATTATAGA
T3_23 -----
T3_22 -----
T3_21 -----
T3_12 -----
T3_11 -----

T1A_16 TTTAGATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCT
T2A_09 TTTAGATGCTGTCCAGCAGATAAAGATAAAGCAAGATCATATCTCAAAAATGGTATCT
T3_23 -----
T3_22 -----
T3_21 -----
T3_12 -----
T3_11 -----

T1A_16 CTTAATTATCTACAAAGTTAAAGAGGCAAAAGAAAGTAAGAAATCTAACATAATGGCAGT
T2A_09 CTTAATTATCTACAAAGTTAAAGAGGCAAAAGAAAGTAAGAAATCTAACATAATGGCAGT
T3_23 -----
T3_22 -----
T3_21 -----
T3_12 -----
T3_11 -----

T1A_16 AGATCTAGGTCTTGATAATTTGGCTACTTTAATATTTAAAAACAATTCTGATTGTTATAT
T2A_09 TGATTTAGGCCTTGATAACTTAGCTGTACTAACATTTAAAGATAATTCTGATTGTTATAT
T3_23 -----
T3_22 -----
T3_21 -----
T3_12 -----
T3_11 -----

T1A_16 TATCAATGGTAAAACTATTAAATCCAAAAATCTTATTTTAATAAAGAAATTGCCAGACT
T2A_09 TATCAATGGTAAAACTATTAAATCCAAAAATCTTATTTTAATAAAGAAATTGCCAGACT
T3_23 -----
T3_22 -----
T3_21 -----
T3_12 -----
T3_11 -----

T1A_16 ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA
T2A_09 ACAAAGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAA
T3_23 -----
T3_22 -----
T3_21 -----
T3_12 -----
T3_11 -----

T1A_16 ATATCTGAGATTAAAGAGAAAAAATTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA
T2A_09 ATATCTGAGATTAAAGAGAAAGAAATTATATTAGAGATTATCTCCATAAAGCTAGTTGCAA
T3_23 -----
T3_22 -----
T3_21 -----
T3_12 -----
T3_11 -----

T1A_16 AATAGTTGATTTAGCAATTGAAAATCAAGTAGAACTATTGTAATTGGAGATATAAAAAA
T2A_09 AATAGTTGATTTAGCAATTGAAAATCAAGTAGAACTATTGTAATTGGAGATATAAAAAA
T3_23 -----
T3_22 -----
T3_21 -----
T3_12 -----
T3_11 -----

T1A_16 TATTAACAATGCAGCAAACCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAA
T2A_09 TATTAACAATGCAGCAAAGCTTAAATCTTTTGTCCAAATACCGATCCAGAGATTAAAAAA
T3_23 -----
T3_22 -----
T3_21 -----
T3_12 -----
T3_11 -----

T1A_16 ATTAATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGGAATTGATGAAAGCTA
T2A_09 ATTAATTGAATACAAAGTTAAACTAAAAGGTATCAAAGTTGTTGGAATTGATGAAAGCTA
T3_23 -----
T3_22 -----
T3_21 -----
T3_12 -----

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T3_11 -----

T1A_16 TACTTCTGGATGTAGT-----
T2A_09 TACTTCCGGATGTACTAAAGCTTTTAATTTTAATTACGCAAGGAAAGCTTTAGTATGACC
T3_23 -----TACGCAAGGTAAGCTTTAGTATGACC
T3_22 -----TACGCAAGGTAAGCTTTAGTATGACC
T3_21 -----TACGCAAGGAAAGCTTTAGTATGACC
T3_12 -----TACGCAAGGAAAGCTTTAGTATGACC
T3_11 -----TACGCAAGGAAAGCTTTAGTATGACC

T1A_16 -----TCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA
T2A_09 GTATTTCGATTTGGCCGCTCTTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA
T3_23 GTATTTCGATTTGGCCGCTGTTTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA
T3_22 GTATTTCGATTTGGCCGCTGTTTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA
T3_21 GTATTTCGATTTGGCCGCTGTTTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA
T3_12 GTATTTCGATTTGGCCGCTATTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA
T3_11 GTATTTCGATTTGGCCGCTATTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGATA

T1A_16 AATCCAGAAGAATTACTAGAGGTCTCTTTAAACTAACGAGGGCCTATTAATTAATGCTG
T2A_09 AATCCAGAAGAATTACCAGAGGTCTCTTTAAACTAACGAGGGCCTATTAATTAATGCTG
T3_23 AATCCAGAAGAATTACTAGAGGTCTCTTTAAACTAACGAGGGCCTATTAATTAATGCTG
T3_22 AATCCAGAAGAATTACTAGAGGTCTCTTTAAACTAACGAGGGCCTATTAATTAATGCTG
T3_21 AATCCAGAAGAATTACTAGAGGTCTCTTTAAACTAACGAGGGCCTATTAATTAATGCTG
T3_12 AATCCAGAAGAATTACTAGAGGTCTCTTTAAATAAACGAGGGCCTATTAATTAATGCTG
T3_11 AATCCAGAAGAATTACTAGAGGTCTCTTTAAACTAACGAGGGCCTATTAATTAATGCTG

T1A_16 ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC
T2A_09 ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC
T3_23 ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC
T3_22 ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC
T3_21 ATCAGAATGGTAGTTTTAATATACTTCGTAAATATCATAACGATAAAATGTATTCTCAGAC
T3_12 ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC
T3_11 ATCAGAATGGTAGTTTTAATATACTTCGTAAATACCATAACGATAAAATGTATTCTCAGAC

T1A_16 CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT
T2A_09 CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT
T3_23 CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT
T3_22 CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT
T3_21 CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT
T3_12 CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT
T3_11 CTATCAAAGAGGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCT

T1A_16 AA
T2A_09 AA
T3_23 AA
T3_22 AA
T3_21 AA
T3_12 AA
T3_11 AA

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Type 3*

CLUSTAL O(1.2.1) multiple sequence alignment

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T3*_15      ATGCGATTATCATTTAAATCAAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
T3_11      ATGCGATTATCATTTAAATCAACCCATAAATTAAGCCATAAGCAATTAGTAATAATTAAT      60
*****

T3*_15      GAATTAGCCTGGCATATTAGTAAACTATATAATACAGTCAATTATCAGATTAATAATAAT      120
T3_11      GAATTAGCCTGGCATTGCCTCAAATATATAATATAGTCAATTATCAGATTAATAATAAT      120
*****: : ****

T3*_15      AAAGATGTAAAAGCTGTCTATACTGA-----                          146
T3_11      AAAGATGTAAAAGCTGTCTATACTGAATTAGAACTAGATATAATAATAACTGGCATAAT      180
*****

T3*_15      -----                          146
T3_11      GACTACCTTCACCTCCATAACAGACAGCAGGCATTAAAGCAGTTAGCTCAGGACTGGAAA      240

T3*_15      -----                          146
T3_11      AGTTTTTTTAAATCTCTCAAAGATTATAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA      300

T3*_15      -----ATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      187
T3_11      GGGTCACCTAATTTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA      360
*****

T3*_15      GCTGTTAGAATTAGAGATAACAAATTACTCTTATCCTTATCTAAAAAGATACAGTCTAAA      247
T3_11      GCTGTTAGAATTAGAGATAATAAATTACTCTTATCCTTATCTAAAAAGATACAGTCTAAA      420
*****

T3*_15      TATAATGTGAAGGTCCTAAAGCTTTTAAATTTATAATACGCAAGGTAAGCTTTAGTATGA      307
T3_11      TATAATGTGAAAGTCACTAAAGCTTTTAAATTTATAATACGCAAGGAAAGCTTTAGTATGA      480
*****.*

T3*_15      CTGTATTCGATTTGGCCGCTCTTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTTTGA      367
T3_11      CCGTATTCGATTTGGCCGCTATTCAGTAGATCTGGAAAAATAAAATAAAAGTAACTATGA      540
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T3*_15      TAAATCCAGAAGAATTTCCAGAGGTCCTTTAAAATAAGGAGGGCCTATTAATT-----      422
T3_11      TAAATCCAGAAGAATTACTAGAGGTCCTTTAAAATAACGAGGGCCTATTAATTAATGC      600
*****:*

T3*_15      -----AATGTATTCTCAG      435
T3_11      TGATCAGAATGGTAGCTTTAATATACTTCGTAAATACCATAACGATAAATGTATTCTCAG      660
*****

T3*_15      ATCTATCAAAGAGGCGAGAGATAATGGGTTTGTGGCCAATCCTTCAAGATTAAGGGTACC      495
T3_11      ACCTATCAAAGAGGCGAGAGATAATGGGTTTGTGGCCAATCCTTCAAGATTAAGGGTACC      720
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T3*_15      TAAA      499
T3_11      CTAA      724
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Type MISC

CLUSTAL O(1.2.1) multiple sequence alignment

T1A_16	ATGCGATTATCATTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT	60
MISC_14	ATACGATTATCATTAAATTC AAGCCTAAATTAAGCCATAAGCAATTAGTAATAATTAAT	60
	** *****	
T1A_16	GAATTAGCCTGGCATTGCTCTAAATTATATAATACAGTCAATTATCAGATTAAAAATAAT	120
MISC_14	GAATTAGCCTGGCATTGCTCTAAATTATATAATACAGTCAATTCTCAGACCTATCAAAGA	120
	***** * * *	
T1A_16	AAAGATGTAAAGCTGTCTATACTGAATTAGAACTAGATATAAAAATAACTGGCATAAT	180
MISC_14	GGCGAGAGAT-----	130
	** *	
T1A_16	GACTACCTTCACTCCCATAAACAGACAGCAGGCATTAAGCAGTTAGTTCAGGACTGGAAA	240
MISC_14	-----	130
T1A_16	AGTTTTTTAATCTCTCAAAGATTATAAAAAGAATCCTCAAAAATATAAGGGTCAGCCA	300
MISC_14	-----	130
T1A_16	GGGTCACCTAATTTAAACATATGAACAGTAATCCCTGTGAAATAATTTTACCAATTTA	360
MISC_14	-----	130
T1A_16	GCTGTTAGAATTAGAGATAACAAATTA CTCTTATCCTTATCTAAAAAGATACAGTCTAAA	420
MISC_14	-----	130
T1A_16	TATAATGTGAAGGCTCTTAATTTGAGCTGCCTGAAGCAGTTC AAAGCATTATAGATTTA	480
MISC_14	-----	130
T1A_16	GATGCTGTCCAGCAGATAAAGATTAAGCAGGACCGTATTTCTAAAAGATGGTATCTCTTA	540
MISC_14	-----	130
T1A_16	ATTATCTACAAAGTTAAAGAGGCAAAAGAAAGTAAGAAATCTAACATAATGGCAGTAGAT	600
MISC_14	-----	130
T1A_16	CTAGGCTTGATAATTTGGCTACTTTAATATTTAAAAACAATTC TGATTGTTATATATATC	660
MISC_14	-----	130
T1A_16	AATGGTAAAAC TATTAATCCAAAAATCTTATTTTAAATAAAGAAATGCCAGACTACAA	720
MISC_14	-----	130
T1A_16	AGCATTAGAATGAGGCAGTTAGCTACCAGTAAAATTAGAGATACTAAACGAATAAAATAT	780
MISC_14	-----	130
T1A_16	CTGAGATTAAGAGAAAAAATTATATTAGAGATTATCTCCATAAAGCTAGTTGCAAAATA	840

MISC_14	-----	130
T1A_16	GTTGATTTAGCAATTGAAAATCAAGTAGAACTATTGTAATTGGAGATATAAAAAATATT	900
MISC_14	-----	130
T1A_16	AAACAATGCAGCAAACCTAAATCTTTTGTCCAAATACCGATCCAGAGATAAAAAAATTA	960
MISC_14	-----	130
T1A_16	ATTGAATACAAAGCTAAACTAAAAGGTATCAAAGTTGTTGGAATTGATGAAAGCTATACT	1020
MISC_14	-----	130
T1A_16	TCTGGATGTAGTTCAGTAGATCTGGAAAAATAAATAAAAGTAACTATGATAAATCCAGA	1080
MISC_14	-----	130
T1A_16	AGAATTACTAGAGGTCTCTTTAAAATAACGAGGGCCTATTAATTAATGCTGATCAGAAT	1140
MISC_14	-----	130
T1A_16	GGTAGTTTTAATATACTTCGTAAATACCATAACGATAAATGTATTCTCAGACCTATCAAA	1200
MISC_14	-----	130
T1A_16	GAGCGAGAGATAATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCTAA	1254
MISC_14	-----AATGGATTTGTGGACAATCCTTCAAGATTAAGGGTATCCTAA	172

APPENDIX D.

tnpA/tnpB INTER-ORF SEQUENCE ALIGNMENT

CLUSTAL O(1.2.1) multiple sequence alignment

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Locus_13      -----CTCCATTTTCTTTTATAAGCAAACATATGTATGGTATAATTATAGTA      49
Locus_03      -AAAAATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      59
Locus_02      AAAGATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_18      AAAAATCAAACCTCCATGTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_16      AAAAATCAAACCTCCATGTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_08      AAAAATCAAACCTCCATGTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_09      AAAAATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_05      AAAGATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_10      AAAAATAAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_11      AAAGATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_12      AAAGATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_14      AAAGATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_17      AAAGATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_19      AAAGATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_20      -----CTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      49
Locus_15      -----ATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      45
Locus_01      -----CTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      49
Locus_23      AAAAATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_22      AAAAATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_21      AAAAATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_06      AAAAATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60
Locus_04      AAAAATCAAACCTCCATTTTCTTTTACAAGCAAACATATGTATGATATAATTATAGTA      60

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Locus_13      GAATGGAGGTGAAAAATCA      68
Locus_03      GGATGGAGGTGAAAAGTCA      78
Locus_02      GGATGGAGGTGAAAATTA      79
Locus_18      GGATGGAGGTGAAAAATCA      79
Locus_16      GGATGGAGGTGAAAAATCA      79
Locus_08      GGATGGAGGTGAAAAATCA      79
Locus_09      GGATGGAGGTGAAAAGTCA      79
Locus_05      GGATGGAGGTGAAAAATCA      79
Locus_10      GGATGGAGGTGAAAAATCA      79
Locus_11      GGATGGAGGTGAAAAATCA      79
Locus_12      GGATGGAGGTGAAAAATCA      79
Locus_14      GGATGGAGGTGAAAAATCA      79
Locus_17      GGATGGAGGTGAAAAATCA      79
Locus_19      GGATGGAGGTGAAAAATCA      79
Locus_20      GGATGGAGGTGAAAAATCA      68
Locus_15      GGATGGAGGTGAAAAATCA      64
Locus_01      GGATGGAGGTGAAAAATCA      68
Locus_23      GGATGGAGGTGAAAAATCA      79
Locus_22      GGATGGAGGTGAAAAATCA      79
Locus_21      GGATGGAGGTGAAAAATCA      79
Locus_06      GGATGGAGGTGAAAAATCA      79
Locus_04      GGATGGAGGTGAAAAATCA      79

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APPENDIX E.

IS605 tnpB ORF INSERT SEQUENCE ALIGNMENT

CLUSTAL O(1.2.1) multiple sequence alignment

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Locus_15HI      TCACTAAAGCTTTTAATTTATAATACGCAAGGTAAGCTTTAGTATGACTGTATTTCGATTT      60
Locus_22HI      TCACTAAAGCTTTTAATTTATAATACGCAAGGTAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_18LI      TCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_13LI      TCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_17LI      TCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_09LI      TCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_19LI      TCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_20LI      TCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_04LI      TCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_06LI      TCACTAAAGCTTTTAATTTATAATACGCAAGGAAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_21HI      TCACTAAAGCTTTTAATTTATAATACGCAAGGTAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_11HI      TCACTAAAGCTTTTAATTTATAATACGCAAGGTAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_12HI      TCACTAAAGCTTTTAATTTATAATACGCAAGGTAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_23HI      GCCTAAAGCTTTTAATTTATAATACGCAAGGTAAGCTTTAGTATGACCGTATTTCGATTT      60
Locus_02RI      ---CTAAAGCTTTTAATTTAATTACGCAAGGTAAGCTTTAGTATGACCGTATTTCGATTT      57
Locus_04RI      ---CTAAAGCTTTTAATTTAATTACGCAAGGTAAGCTTTAGTATGACCGTATTTCGATTT      57
Locus_06RI      ---CTAAAGCTTTTAATTTAATTACGCAAGGTAAGCTTTAGTATGACCGTATTTCGATTT      57
Locus_13RI      ---CTAAAGCTTTTAATTTAGTTACGCAAGGTAAGCTTTAGTATGACCGTATTTCGATTT      57
Locus_09RI      ---CTAAAGCTTTTAATTTAATTACGCAAGGAAAGCTTTAGTATGACCGTATTTCGATTT      57
Locus_03RI      ---CTAAAGCTTTTAATTTAATTACGCAAGGTAAGCTTTAGTATGACCGTATTTCGATTT      57

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Locus_15HI      GGCCGCT 67
Locus_22HI      GGCCGCT 67
Locus_18LI      GGCC--- 64
Locus_13LI      GGCC--- 64
Locus_17LI      GGCC--- 64
Locus_09LI      GGCC--- 64
Locus_19LI      GGCC--- 64
Locus_20LI      GGCC--- 64
Locus_04LI      GGCC--- 64
Locus_06LI      GGCC--- 64
Locus_21HI      GGCCGCT 67
Locus_11HI      GGCCGCT 67
Locus_12HI      GGCCGCT 67
Locus_23HI      GGCCGCT 67
Locus_02RI      GGCCGCT 64
Locus_04RI      GGCCGCT 64
Locus_06RI      GGCCGCT 64
Locus_13RI      GGCCGCT 64
Locus_09RI      GGCCGCT 64
Locus_03RI      GGCCGCT 64

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APPENDIX F.

IS605 LEFT END SEQUENCE ALIGNMENT

CLUSTAL O(1.2.1) multiple sequence alignment

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Locus_04      TTTATATAAAATGCCAAGAAAACCCATCCAAGCTATGCATTGGCTGGAGATGAATTGG      60
Locus_05      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTAGGTGGAGATGAATTGG      60
Locus_21      TTTATTTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTAGGTGGAGATGAATTGG      60
Locus_03      TTTATCTGAAACTGCCAAGAAAACCCATCCAAGCTATGCATTGGGTGGAGATGAATTGG      60
Locus_01      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGGTGGAGATGAATTGG      60
Locus_15      TTTATCTAAATATGCCAAGAAAACCCATCCAAGCTATGCATTGTGTGGAGATAAATTGG      60
Locus_11      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGGTGGAGATGAATTGG      60
Locus_12      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGGTGGAGATAAATTGG      60
Locus_14      TTTATCTAAAAC TGCCAAGAAAACCCCTCCAAGCTATGCATTGGGTGGAGATGAATTGG      60
Locus_16      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGCTGGAGATGAATTGG      60
Locus_17      TTTATCTAAAAC TGCTAAGAAAACCCATCCAAGCTATGCATTGGGTGGAGATAAATTGG      60
Locus_18      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGCTGGAGATGAATTGG      60
Locus_23      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGCTGGAGATGAATTGG      60
Locus_02      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGCTGGAGATGAATTGG      60
Locus_22      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGCTGGAGATGAATTGG      60
Locus_06      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGGTGGAGATGAATTGG      60
Locus_08      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGGTGGAGATGAATTGG      60
Locus_10      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGGTGGAGATGAATTGG      60
Locus_19      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGGTGGAGATGAATTGG      60
Locus_09      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGGTGGAGATGAATTGG      60
Locus_20      TTTATCTAAAAC TGCCAAGAAAACCCATCCAAGCTATGCATTGGGTGGAGATGAATTGG      60
Locus_13      TTTATCTAAAATGTCAAGAAAACCCATTCAAGCTATGCATTGGGGGA-----      49
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APPENDIX G.

IS605 RIGHT END SEQUENCE ALIGNMENT

CLUSTAL O(1.2.1) multiple sequence alignment

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Locus_01      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_21      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_14      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_04      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_06      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_02      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_12      GCTATTAGGAGCAAAATTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_13      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_17      GCTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_19      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_08      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_23      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_20      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_11      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_09      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_03      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_16      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_10      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_05      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_18      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
Locus_22      ACTATTAGGAGCAAACTTAAAAGCCAAACATCTTGTAAGCTGACCTAGTAATATAGGTT      60
                *****

Locus_01      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCATCCGACGCGAAGCTAGCATCTCTTT      120
Locus_21      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCATCCGACGCGAAGCTAGTATCTCTTT      120
Locus_14      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCATCCGACGCGAAGCTAGTATCTCTTT      120
Locus_04      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCATCCGACGCGAAGCTAGTATCTCTTT      120
Locus_06      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCATCCGACGCGAAGCTAGTATCTCTTT      120
Locus_02      GAACTTAATCTATATGAAGCAGTTAAAAGCTCCCTCTAAATCTTGTTTTGATTTAGG-      119
Locus_12      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGTTTTGATTTAGG-      119
Locus_13      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGATTTGATTTAGG-      119
Locus_17      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGTTTTGATTTAGG-      119
Locus_19      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCATCTAAATCTTGTTTTGATTTAGA-      119
Locus_08      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCATCTAAATCTTGTTTTGATTTAGA-      119
Locus_23      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGTTTTGATTTAGG-      119
Locus_20      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGTTTTGATTTAGG-      119
Locus_11      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGTTTTGATTTAGG-      119
Locus_09      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGTTTTGATTTAGG-      119
Locus_03      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGTTTTGATTTAGG-      119
Locus_16      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGTTTTGATTTAGG-      119
Locus_10      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGTTTTGATTTAGG-      119
Locus_05      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGTTTTGATTTAGG-      119
Locus_18      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCCTCTAAATCTTGTTTTGATTTAGG-      119
Locus_22      GAACTTAATCTATATGAAGCAGTTAGAAGCTCCATCTAAATCTTGTTTTGATTTAGG-      119
                *****

Locus_01      TGATGCAAATCTTGGTTTTGATTTAGGTGGAGAGGTTCCAC      160
Locus_21      TGATGCAAATCTTGGTTTTGATTTAGGTGGAGAGGTTCCAC      160
Locus_14      TGATGCAAATCTTGGTTTTGATTTAGGTGGAGAGGTTCCAC      160
Locus_04      TGATGCAAATCTTGGTTTTGATTTAGGTGGAGAGGTTCCAC      160
Locus_06      TGATGCAAATCTTGGTTTTGATTTAGGTGGAGAGGTTCCAC      160
Locus_02      -----TGGAGAGGTTCCAC      132
Locus_12      -----TGGAGAGGTTCCAC      132

```

Locus_13	-----TGGAGAGGTTAC	132
Locus_17	-----TGGAGAGGTTAC	132
Locus_19	-----TGGAGAGGTTAC	132
Locus_08	-----TGGAGAGGTTAC	132
Locus_23	-----TGGAGAGGTTAC	132
Locus_20	-----TGGAGAGGTTAC	132
Locus_11	-----TGGAGAGGTTAC	132
Locus_09	-----TGGAGAGGTTAC	132
Locus_03	-----TGGAGAGGTTAC	132
Locus_16	-----TGGAGAGGTTAC	132
Locus_10	-----TGGAGAGGTTAC	132
Locus_05	-----TGGAGAGGTTAC	132
Locus_18	-----TGGAGAGGTTAC	132
Locus_22	-----TGGAGAGGTTAC	132

APPENDIX H.

IS605 LEFT END REVERSE COMPLEMENT ALIGNMENT

>>>Locus_09LE, 60 nt vs lalign-I20160419-034638-0135-52459438-pg.bsequence library

>>Locus_09LE_RC (60 nt)

Waterman-Eggert score: 36; 88.8 bits; E(1) < 6.5e-24
88.9% identity (88.9% similar) in 9 nt overlap (60-52:16-24)

```

      60
Locus_ CCAATTCAT
      : : : : :
Locus_ CCAATGCAT
      20
    
```

>--
Waterman-Eggert score: 33; 10.9 bits; E(1) < 0.84
72.2% identity (72.2% similar) in 18 nt overlap (36-20:36-53)

```

      30
Locus_ AGCTTGGATGG-AGTTTT
      : : : : :
Locus_ AGTTTTCTTGGCAGTTTT
      40      50
    
```

>>Locus_09LE_RC (60 nt)

Waterman-Eggert score: 86; 26.4 bits; E(1) < 4.1e-05
65.2% identity (65.2% similar) in 46 nt overlap (15-60:1-46)

```

      20      30      40      50      60
Locus_ CCAAGAAACTCCATCCAAGCTATGCATTGGGTGGAGATGAATTGG
      : : : : : : : : : : : : : : : : : : : : : :
Locus_ CCAATTCATCTCCACCCAATGCATAGCTTGGATGGAGTTTTCTTGG
      10      20      30      40
    
```

>--
Waterman-Eggert score: 56; 17.6 bits; E(1) < 0.017
72.7% identity (72.7% similar) in 22 nt overlap (29-50:11-32)

```

      30      40      50
Locus_ TCCAAGCTATGCATTGGGTGGA
      : : : : : : : :
Locus_ TCCACCCAATGCATAGCTTGGA
      20      30
    
```

>--
Waterman-Eggert score: 34; 11.2 bits; E(1) < 0.78
69.2% identity (69.2% similar) in 26 nt overlap (23-47:14-38)

```

      30      40
Locus_ ACTCCATCCA-AGCTATGCATTGGGT
      : : : : : : : : : :
Locus_ ACCCAATGCATAGCT-TGGATGGAGT
      20      30
    
```

APPENDIX I.

IS605 RIGHT END REVERSE COMPLEMENT ALIGNMENT

Waterman-Eggert score: 39; 11.0 bits; E(1) < 1
73.3% identity (73.3% similar) in 15 nt overlap (32-18:114-128)

```

      30      20
Locus_ ATGTTTGGCTTTTAA
      : : : : : : : : : :
Locus_ AAGTTTGGCTCCTAA
      120

```

>>Locus_16RE_RC (132 nt)

Waterman-Eggert score: 82; 20.4 bits; E(1) < 0.013
64.4% identity (64.4% similar) in 59 nt overlap (63-120:13-70)

```

      70      80      90      100      110      120
Locus_ ACTTTAATCTATATGAAGCAGTTAGAACTCCCTCTAAAT-CTTGGTTTTGATTTAGGT
      :: : : : : : : : : : : : : : : : : : : : : : : : : : : : :
Locus_ ACCTAAATCAAAACCAAG-ATTTAGAGGGAGCTTCTAACTGCTTCATATAGATTAAAGT
      20      30      40      50      60      70

```

>--

Waterman-Eggert score: 82; 20.4 bits; E(1) < 0.013
68.4% identity (68.4% similar) in 38 nt overlap (41-78:46-83)

```

      50      60      70
Locus_ CTGACCTAGTAATATAGGTTGAACTTTAATCTATATGA
      :: : : : : : : : : : : : : : : : :
Locus_ CTAAGTCTTCATATAGATTAAAGTTCAACCTATATTA
      50      60      70      80

```

>--

Waterman-Eggert score: 62; 16.0 bits; E(1) < 0.23
64.7% identity (64.7% similar) in 34 nt overlap (91-124:9-42)

```

      100      110      120
Locus_ CTCCCTCTAAATCTTGGTTTTGATTTAGGTGGAG
      : : : : : : : : : : : : : : : :
Locus_ CTCCACCTAAATCAAAACCAAGATTTAGAGGGAG
      10      20      30      40

```

```
>>Locus_16RE_RC (132 nt)
  Waterman-Eggert score: 42; 146.7 bits; E(1) < 1.2e-40
  66.7% identity (66.7% similar) in 21 nt overlap (129-109:48-68)
```

```
      130      120      110
Locus_ AACCTCTCCACCTAAATCAAA
      :::  ::  ::  ::  ::  :::
Locus_ AACTGCTTCATATAGATTAAA
      50      60
```

```
>--
```

```
  Waterman-Eggert score: 39; 11.0 bits; E(1) < 1
  62.5% identity (62.5% similar) in 24 nt overlap (117-94:65-88)
```

```
      110      100
Locus_ TAAATCAAAAACCAAGATTTAGAGG
      ::::  ::::  :  :::  :::
Locus_ TAAAGTTCAACCTATATTACTAGG
      70      80
```

APPENDIX J.

IS605 tnpB ORF INSERT REVERSE COMPLEMENT ALIGNMENT


```
>>Locus_22HI_RC (67 nt)
Waterman-Eggert score: 35; 83.1 bits; E(1) < 4.2e-22
100.0% identity (100.0% similar) in 7 nt overlap (40-34:55-61)
```

```
40
Locus_ AAAGCTT
      :::::
Locus_ AAAGCTT
      60
```

```
>>Locus_22HI_RC (67 nt)
Waterman-Eggert score: 70; 19.0 bits; E(1) < 0.0087
63.4% identity (63.4% similar) in 41 nt overlap (3-43:25-65)
```

```
10      20      30      40
Locus_ ACTAAAGCTTTTAATTTATAATACGCAAGGTAAGCTTTAGT
      ::::: : :: : : : :::::
Locus_ ACTAAAGCTTACCTTGCGTATTATAAATTTAAAGCTTTAGT
      30      40      50      60
```

66.7% identity (66.7% similar) in 27 nt overlap (21-47:21-47)

```

          30          40
Locus_ TAATACGCAAGGTAAGCTTTAGTATGA
      : : : : : : : : : : : : : : : :
Locus_ TCATACTAAAGCTTACCTTGCGTATTA
          30          40

```

>--

Waterman-Eggert score: 40; 11.7 bits; E(1) < 0.74
100.0% identity (100.0% similar) in 8 nt overlap (6-13:55-62)

```

          10
Locus_ AAAGCTTT
      : : : : : :
Locus_ AAAGCTTT
          60

```

>--

Waterman-Eggert score: 38; 11.2 bits; E(1) < 0.85
76.9% identity (76.9% similar) in 13 nt overlap (11-23:45-57)

```

          20
Locus_ TTTTAATTTATAA
      : : : : : :
Locus_ TTATAAATTAATAA
          50

```

>--

Waterman-Eggert score: 34; 10.3 bits; E(1) < 0.97
62.9% identity (62.9% similar) in 35 nt overlap (22-54:14-46)

```

          30          40          50
Locus_ AATACG--CAAGGTAAGCTTTAGTATGACCGTATT
      : : : : : : : : : : : : : : : :
Locus_ AATACGGTCATACTAAAGCTTACCTTG--CGTATT
          20          30          40

```

>--

Waterman-Eggert score: 30; 9.3 bits; E(1) < 1
66.7% identity (66.7% similar) in 15 nt overlap (3-17:51-65)

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