Fall 2011

# Multiple Paternity of Caretta Caretta within the Northwestern Atlantic Ocean Population on Wassaw Island, Ga 

Jacob Lasala

Follow this and additional works at: https://digitalcommons.georgiasouthern.edu/etd

## Recommended Citation

Lasala, Jacob, "Multiple Paternity of Caretta Caretta within the Northwestern Atlantic Ocean Population on Wassaw Island, Ga" (2011). Electronic Theses and Dissertations. 757. https://digitalcommons.georgiasouthern.edu/etd/757

This thesis (open access) is brought to you for free and open access by the Graduate Studies, Jack N. Averitt College of at Digital Commons@Georgia Southern. It has been accepted for inclusion in Electronic Theses and Dissertations by an authorized administrator of Digital Commons@Georgia Southern. For more information, please contact digitalcommons@georgiasouthern.edu.

# MULTIPLE PATERNITY OF CARETTA CARETTA WITHIN THE NORTHWESTERN ATLANTIC OCEAN POPULATION ON WASSAW ISLAND, GA 

by<br>JACOB LASALA<br>(Under the Direction of David C. Rostal)


#### Abstract

Characterizing the mating system of a species is important for understanding demography and population dynamics and can contribute information to conservation efforts. Mating systems can impact the ecology, evolution, effective population size and genetic variability of a species. Polyandry, resulting in multiple paternity can influence the maintenance of genetic variation within a population. Within Testudines, the frequency of multiple paternity varies extensively among species (0-100\% of nests). Previous studies on the loggerhead turtle (Caretta caretta) have shown that within the large management unit of peninsular Florida, multiple paternity occurs in approximately $30 \%$ of nests. This study examines nests from the smaller and more endangered northern management unit. The primary objectives of this study are to determine if multiple paternity exists in Georgia's smaller nesting population and determine if the percentage of nests with multiple fathers differs significantly from previous studies. Secondary objectives are to compare the incidence of multiple paternity over multiple years, determine if multiple paternity varies over the course of the nesting period. Our final objectives were to determine the relationship between the number of fathers per nest and female size (straight carapace length), as well as hatching success and to determine how many males are actually contributing to this nesting population. Mothers and offspring (up to 20) were initially sampled from more than 90 nests over three nesting seasons on Wassaw Island,


GA (2008-2010). We found that multiple fathers contribute to $75 \%$ of nests over the three years. There is a difference in number of fathers per nest with relation to the year, but there is no relationship between the number of fathers per nest and the nesting period. There is a positive relationship between female size (SCL) and the number of fathers per nest. There is no relationship between the number of fathers contributing to a clutch and hatching success. Finally I found a total of 195 male genotypes over the course of the study, resulting in a sex ratio of 2.7 males per 1 nesting female. Every male genotype that I discovered only contributed to one nest over the three years samples, indicating there is a large number of males contributing to this nesting population.

INDEX WORDS: Caretta caretta, Mating system, Multiple paternity, Georgia, Barrier islands, Genotypes, Straight carapace length, Hatching success, Nest size, Sex ratio

MULTIPLE PATERNITY OF CARETTA CARETTA WITHIN THE NORTHWESTERN ATLANTIC OCEAN POPULATION ON WASSAW ISLAND, GA
by

## JACOB LASALA

B.A., University of Chicago, 2009<br>M.S., Georgia Southern University, 2011

A Thesis Submitted to the Graduate Faculty of Georgia Southern University in Partial Fulfillment of the Requirements for the Degree

## MASTER OF SCIENCE

STATESBORO, GEORGIA
2011

## JACOB LASALA

All Rights Reserved

# MULTIPLE PATERNITY OF CARETTA CARETTA WITHIN THE NORTHWESTERN 

 ATLANTIC OCEAN POPULATION ON WASSAW ISLAND, GAby

JACOB LASALA

Major Professor: David C. Rostal<br>Committee: John Scott Harrison<br>Denson McLain

Electronic Version Approved:
November 2011

## ACKNOWLEDGMENTS

I would like to thank the Caretta Research Project, who provided the core data sampling of my project. Specifically I would like to thank Kris Williams and Michael Frick. Kris' patience (with turtles, wind, people, weather and me) will always be the gold standard for how to approach research.

I would like to thank my primary advisor, Dr. David C. Rostal for his continual efforts to guide my work, mentoring, support (and lab meetings). Second, I would like to thank Dr. J. Scott Harrison for his patience and for the use of his lab space. I will always appreciate the knowledge he shared with me. I would also like to thank my third committee member, Dr. Denson McLain, for editing and providing me with moral support. He taught me the true value of constructive edits.

I would especially like to thank the people that provided me with help over the years, including: Mandy Moss, Amber Teare, Jessica Sparrow, Ketan Patel, Wade Carruth, Tiffany Klein, Leigh-Ann Colley, Matt Erickson, Matt Phillips, Jacob Heatherly, Michelle Carlson, Chris Evett, Brittany Arrington and Erica Johnson. A warm thanks to colleagues and friends (especially the GSU Beagles) who shared the journey. I would also like to thank Helen Worrell for her support and my parents for teaching me to finish what I start.

Funding for this project was provided by the Georgia Southern College of Graduate Studies Graduate Student Professional Development Fund (Spring 2010, Fall 2010) and the College of Science and Technology Academic Excellence Fund (Spring 2010). Travel support was provided by the Georgia Southern Student Government Association Travel Award and the Society for Integrative and Comparative Biology

Charlotte E Magnum Student Support Fund for travel to conferences where this research was presented. All work was done in accordance with animal care and use protocols approved by the Institutional Animal Care and Usage Committee at Georgia Southern University.

## TABLE OF CONTENTS

ACKNOWLEDGEMENTS ..... 6
LIST OF TABLES ..... 9
LIST OF FIGURES ..... 10
CHAPTER 1
MULTIPLE PATERNITY OF CARETTA CARETTA WITHIN THE NORTHWESTERN ATLANTIC OCEAN POPULATION ON WASSAWISLAND, GA: A LONGITUDINAL STUDY (2008-2010)11
Abstract ..... 11
Introduction ..... 12
Materials and Methods ..... 17
Results ..... 23
Discussion ..... 27
REFERENCES ..... 49
APPENDIX
APPENDIX A: Full Nest Data ..... 56

## LIST OF TABLES

Table 1: Studies of Multiple Paternity within sea turtles ............................................... 33
Table 2: Descriptive statistics of the five polymorphic tetra-nucleotide primers described by Shamblin et al., 2009 .................................................................................... 33

Table 3:. Descriptive table of multiple paternity by year, using GERUD, COLONY and
the average between the two programs ........................................................ 34
Table 4a:. Generalized Linear Model for GERUD, distribution gamma ....................... 35
Table 4b:. Generalized Linear Model for COLONY, distribution gamma ..................... 32

## LIST OF FIGURES

Figure 1: Graph showing the relationship between the programs GERUD and COLONY
and the three years analyzed. $\left(\mathrm{N}_{2008}=23, \mathrm{~N}_{2009}=21, \mathrm{~N}_{2010}=28\right) \ldots$

Figure 2: Relative contribution of fathers to clutches sired by multiple fathers from 2008
37
Figure 3: Relative contribution of fathers to clutches sired by multiple fathers from 2009

Figure 4: Relative contribution of fathers to clutches sired by multiple fathers from 2010

Figure 5: Difference between the average number of fathers by year ............................. 40
Figure 6: Relationship between female size (straight carapace length, cm ) and the number of fathers per nest .41

Figure 7: Relationship between the number of fathers per nest and the hatching success (\%) of each nest, using both programs .42

Figure 8: Relationship between female size (SCL, cm) and the number of eggs per nest
(nest size) ...................................................................................................... 43
Figure 9: Nesting period (early, middle and late) versus the average number of fathers
(a) 2008 \& (b) 2009
(c) 2010 \& (d) All 4 Combined ............................................................................ 45

Figure 10: Julian Nest Date compared to number of fathers per nest ............................. 46
Figure 11: Wassaw Island Multiple Paternity in regards to previous studies ................. 47
Figure 12: Relationship between siblings and paternal contributions ............................. 48

# MULTIPLE PATERNITY OF CARETTA CARETTA WITHIN THE NORTHWESTERN ATLANTIC OCEAN POPULATION ON WASSAW ISLAND, GA: <br> A LONGITUDINAL STUDY (2008-2010) 

Written with intent to submit as a manuscript to:<br>Molecular Ecology


#### Abstract

Characterizing the mating system of a species is important for understanding demography and population dynamics and can contribute information to conservation efforts. Mating systems can impact the ecology, evolution, effective population size and genetic variability of a species. Polyandry, the occurrence of a single female mating with multiple males can result in the multiple paternity of progeny arrays. Previous studies on the loggerhead turtle (Caretta caretta) have shown that within the large nesting population of peninsular Florida, multiple paternity occurs in approximately $30 \%$ of nests. This study tested nests from the smaller and more endangered northern nesting population for the presence of multiple paternal contributions. Mothers and up to 20 offspring were sampled from more than 90 nests over three nesting seasons on Wassaw Island, GA (2008-2010). We found that $75 \%$ of nests sampled had multiple fathers with an average of 2.65 fathers per nest. We determined that there was a difference in the number of fathers per nest with relation to the year, but not to the period the nest was laid. There was a positive relationship between the number of fathers per nest and female size (SCL), but there was no relationship between number of fathers and hatching success. Finally we found 195 paternal genotypes and determined that each individual only contributed to one nest over all three years.


## INTRODUCTION

The study presented here is the first examination of multiple paternity within loggerhead sea turtles (Caretta caretta) nesting on Georgia beaches. This is also the first study to look at the frequency of multiple paternity on a small nesting beach and will be the first study to examine the "full" nesting potential of this nesting beach: this research covers three years of nesting data, allowing for an assessment of the whole nesting population on this beach.

Loggerheads reach sexual maturity between 20 and 35 years of age (Federal Register, 2011). Every three years, females will travel to their mating sites between late March and early June, males are not restricted and can mate every year (Conant et al., 2009). Mating occurs in offshore wasters near to the nesting beaches several weeks before nesting (Limpus, 1985, Conant et al. 2009, etc). Males will occasionally fight one another for access to females (Schofield et al., 2006) and females can mate with more than one male and then store the sperm for up to a year (Uller and Olsson, 2008). Males return to their foraging grounds after mating, females will remain in the vicinity of the nesting beach (Limpus and Reed, 1985; Dodd, 1988, etc). Between late April and early September, females return to their natal nesting beaches. Females nest on continental beaches and barrier islands close to continental land masses (Dodd, 1985) Females will lay their first clutch and leave the beach, approximately two weeks (14 days) later the females return to nest again (Dodd, 1988; Conant et al., 2009 etc.); this process can be repeated for up to 7 (Dodd, 1988) or 8 (observed) clutches per female.

Determining how mating systems impact population ecology, genetics and evolution has long been of interest to ecology and evolutionary biologists (Wright, 1931;

Bjorndal et al. 1983; etc.). Direct observation of marine turtles mating has been a difficult task to accomplish in the wild, and has had limited success in closed and controlled settings (Moore et al., 2008). Advances in genetic techniques and analytical methods permit allow scientists to infer turtle mating systems that may be relevant to conservation policy.

In 1978 the loggerhead sea turtle was first listed as threatened in its circum-global distribution (Federal Register, 1978). In 2007 a petition circulated to ask that the Western North Atlantic Loggerhead be listed as endangered. In 2008, the "Recovery Plan for the Northwest Atlantic Population of the Loggerhead Sea Turtle (Caretta caretta)" highlighted five specific recovery units from Virginia to the Caribbean, including the Northern Gulf of Mexico (NMFS and USFWS, 2008). The primary goal of this recovery plan was to determine if the Northwest Atlantic loggerhead population was declining enough to warrant endangered status.

Currently there are 9 distinct population segments worldwide as defined by the National Marine Fisheries Service and the United States Fish and Wildlife Service (Federal Register, 2011). A distinct population segment is the smallest partition of a species allowed to be protected under the U.S. Endangered Species Act. A distinct population segment is granted to a population if it is both discrete and significant in relation to the rest of the species (Federal Register, 2011). Five of these 9 distinct population segments are now considered endangered, the remaining 4 distinct population segments are now considered threatened (including the Northwest Atlantic population).

Bowen et al. (2005) examined mitochondrial DNA, typically maternally inherited, suggested that nesting colonies across the southeastern United States had a strong
population structure (each beach was genetically distinctive when compared to one another). However, nuclear DNA (microsatellite markers: inherited from both parents) did not show a strong population structure on those same nesting beaches (Bowen et al., 2005). Bowen et al. suggested that the species mixed in the ocean and then as individuals grew, juveniles would group together depending on where they were initially hatched (ie. Western/Eastern North Atlantic etc.). In this model, adults would separate further, such that the loggerheads along the southeastern United States are all one population, but that breeding/nesting populations are distinctive nearer to shore.

Bowen et al.'s data indicates that this species' genetic structure along the Northwest Atlantic coast is driven by natal nesting (Bowen et al., 2005). Natal nesting is the process of nesting females returning to their natal nesting beaches to lay their eggs. In this model, gene flow is attributed to males, as they will move along the coast and not be restricted to the beaches they hatched on (Bowen et al., 2005, Federal Register, 2011). The Northwestern Atlantic population is genetically distinct from populations elsewhere in the Atlantic Ocean (Bowen et al., 2005; NMFS and USFWS, 2008; Federal Register, 2011; Richards et al., 2011). Thus NMFS and USFWS recently determined that a loss of this population would mean a loss of genetic diversity to the species as a whole (Federal Register, 2011).

The recovery plan of 2008 highlights two recovery units along the coast of the Northwestern Atlantic Ocean: The Northern (Virginia - North Florida) and the Penninsular Florida Recovery Unit (NRU and PRFU respectively) (NMFS and USFWS, 2008). From 1989-2008, annual nest totals of the NRU averaged 5,125 nests and from 1989-2007, annual nests totals of the PFRU average 64,513. Loggerhead nests have
decreased $1.3 \%$ annually since 1983 in the NRU; overall the unit has experienced a "long-term decline" (NMFS and USFWS, 2011). PFRU nesting has decreased as well by $1.6 \%$ annually in the last 20 years. However, NMFS and USFWS suggest that recent abnormally high nesting years (2008-2010) represent a positive trend for the NRU (Federal Register, 2011). The nesting data suggests that the Northwestern Atlantic population is declining, but there could be signs for recent recruitment.

NMFS and USFWS determined that the Northwestern Atlantic discrete population segment should be considered threatened as opposed to endangered in 2011 (Federal Register, 2011). The factors cited were population abundance and trend: they suggested that the adult population (over 60,000 by conservative estimate) exceeds those of other endangered species in the Atlantic. Furthermore, while acknowledging that the population is experiencing decline, NMFS and USFWS stated that recent nesting events appear to be positive and could stabilize the region. NMFS and USFWS estimate that the nests in Florida comprise approximately $87 \%$ of all nesting effort in the Northwest Atlantic population. Subsequently, the majority of the research concerning this discrete population segment has occurred in Florida regarding nesting counts and female characteristics (Bjorndal et al., 1983; Bollmer et al., 1999; Bowen, 2004; Caldwell, 1962; LeBuff and Beatty, 1971; Dodd et al., 2003; Hanson et al., 1998; Moore and Ball, 2002; Steinitz et al., 1998 etc.).

Our understanding of this discrete population segment is incomplete: there is little information available about the mating males as we cannot obtain a reliable sex ratio because males rarely leave the ocean. Without knowledge of male contribution, the effectiveness with which we manage designated populations will always be limited
(Wibbels et al., 1991;Hanson et al., 1998, Casale et al., 2005, Casale et al., 2011 etc.)
Examining paternity of hatchlings we can evaluate the number of contributing males indirectly (ie. without physically counting each individual). Using genetic techniques we can determine how many males sired a nest, even if we are not able to observe mating behavior. Exclusion paternity analysis can estimate the potential genotypes of contributing fathers, allowing an estimate of how many individuals contribute to nesting sites to be calculated. NMFS and USFWS estimate that there are 30,000 nesting females in the Northwest Atlantic discrete population segment and in 2011 Richards et al. Used models to conservatively estimate that there was just over 38,000 females contributing to the discrete population segment from 2001-2010. The current approximation of adult sex ratios are $1: 1$ suggesting there are $60,000-75,000+$ adult loggerheads in this discrete population segment over the course of this last decade (Federal Register, 2011; Richards et al., 2011 ). The problem with this approximation is that it assumes that all males are reproducing and does not assess the number of males that are actually contributing to this discrete population segment. By looking at the number of genotypes through paternity, we can approach the sex ratio from a different angle and perhaps determine a more robust population estimate.

Previous studies of multiple paternity in sea turtles are summarized in Table 1. These studies indicate that multiple paternity occurs in sea turtle nests (Moore \& Ball, 2002; Zbinden et al., 2007; Fitzsimmons, 1998; Crim et al., 2002; etc.) but varies across populations within a species (Jensen et al., 2006, Zbinden et al. 2007; etc.). It is interesting to note the difference between nesting sites regarding Lepidochelys olivacea: in a regular rookery, only $30 \%$ of nests have multiple paternal contributions, but within
an arribada nesting site (mass mating) the frequency rises significantly to $92 \%$ (Jensen et al., 2006). Jensen et al. attributed the difference in sites due to the abundance of individuals in the mating system, suggesting that as the number of nesting females rise, so does the chance of multiple mating. Following this assessment, larger nesting beaches (more nesting females) should have a higher percentage of nests with multiple paternity. Do smaller nesting beaches (ie. less nesting females) follow this model?

This study examined the occurrence of multiple paternity over the full nesting interval (three years) on Wassaw Island, GA. Loggerheads from the Northwestern Atlantic distinct population segment, return once every three years (Conant et al., 2009), by analyzing 92 non-repeating mothers and 72 nests we are essentially examining a snapshot of the nesting population on Wassaw Island, GA. During the three years, we sampled $19.5 \%$ of the nests laid in that time period.

I will quantify the occurrence of multiple paternity within this nesting population and how it relates to what we know about the whole discrete population segment (specifically with respects to Florida findings). Using genetic data I will estimate how many males are contributing to this nesting beach. Finally I will assess the relationship between the number of fathers per nest and female size, nest hatching success and the timing of the nesting event.

## MATERIALS AND METHODS

## Field Methods

Samples were obtained on the Wassaw National Wildlife Refuge (WASI, $31^{\circ} 54^{\prime} 1.08^{\prime \prime} \mathrm{N} / 80^{\circ} 58^{\prime} 55.92^{\prime \prime}$ ) for three summer nesting seasons (2008-2010). Female loggerheads nest on this 11.3 km beach from May until early August and nests hatch from
late June until early October.
During nightly patrols with the Caretta Research Project adult nesting females were identified from their crawls up the beach (sea turtles all have distinctive crawls, see Figure X). Female sampling was divided into three periods of the nesting season in correspondence to reproductive hormone levels (Drake, 2001): early nests from May 15 June 10; middle nests from June 11 - July 8 and late nests from July 9 - August 4.

Drake (2001) determined that nesting female testosterone levels peaked at the beginning of the nesting season and would drop (step wise) with each subsequent nesting event. Using this step down pattern as a model, we can estimate when nesting events will occur according to our calendar. The first two nesting events roughly from April 30- June 9 (Early Nesting Period), the next two nesting events occur from June 10- July 9 (Middle Nesting Period) and the final nesting events occur from July 10 - August 18 or until the end of the nesting season (Late Nesting Period). There is some overlap between nesting events and this time scale, but if we assume nesting events occur randomly throughout the nesting season, then independent females could be collected in sampling were distributed throughout each nesting period.

Ten individual females from each nesting period (resulting in 90 females) were tagged and measured according to U.S. Fish and Wildlife protocols (Barnard and Keinath, 1999; Williams and Frick, 2000; Drake, 2001). Blood samples were taken from the cervical sinus using a $21 \mathrm{Gx} 1-1 / 2$ " needle and retained in a 5 ml Sodium Heparin vacutainer. No individual was sampled more than once.

Nests of these females were promptly caged to prevent predation as well as the escape of hatchlings. A GPS reading was taken to mark the location of each nest. Stakes
were also added at the dune line to approximate location of nest from the water line, to document the date the nest was laid and to also designate what kind of turtle laid the nest: Caretta caretta (CC); Chelonia mydas (CM); Dermochelys coriacea (DC) were the three typical species. At hatching, 20 random hatchlings were removed from each loggerhead nest, weighed, measured and sacrificed. Residual yolk sacs were removed from these hatchlings, weighed and placed in Whirl-Paks® and stored at $-20^{\circ} \mathrm{C}$.

## Molecular Techniques

Maternal DNA was extracted by adding $2 \mu \mathrm{~L}$ of blood to a lysis buffer (10mM Tris $\mathrm{pH} 8.3,50 \mathrm{mM} \mathrm{KCl}$ and $0.5 \%$ Tween 20) and the mixture was incubated with proteinase $\mathrm{K}(20 \mu \mathrm{~L},>600 \mathrm{mAU} / \mathrm{ml})$ at $65^{\circ} \mathrm{C}$ for one hour followed by $100^{\circ} \mathrm{C}$ for 15 minutes. Hatchling DNA was extracted from residual yolk sacs using the DNeasy blood and tissue kit (QIAGEN) following manufacturer protocol. Polymerase chain reaction (PCR) amplification was carried out using primers for five microsatellite loci (CcP7E05, CcP2F11, CcP7D04, CcP7C06, CcP8D06) designed for Caretta caretta (Shamblin et al., 2009). PCR reactions were carried out as a multiplex reaction in $25 \mu \mathrm{~L}$ volumes, consisting of $2 \mu \mathrm{~L}$ of extracted DNA, $10 \mu \mathrm{~L}$ of Apex Taq Master Mix ( 150 mM TrisHCl pH 8.5, $40 \mathrm{mM}\left(\mathrm{NH}_{4}\right)_{2} \mathrm{SO}_{4}, 3.0 \mathrm{mM} \mathrm{MgCl} 2,0.2 \%$ Tween $20 ; 0.4 \mathrm{mM}$ dNTPs; 0.05 units $/ \mu \mathrm{L}$ Apex Taq DNA polymerase \& stabilizer), $3 \mu \mathrm{~L} \mathrm{H}_{2} \mathrm{O}, 22 \mu \mathrm{~L}$ P7E Forward and Reverse, $1 \mu \mathrm{~L}$ P2F F\&R, $0.5 \mu \mathrm{~L}$ P7D F $\& \mathrm{R}, 1 \mu \mathrm{~L}$ P7C F\&R and $0.5 \mu \mathrm{~L}$ P8D F\&R (10 $\mu \mathrm{M})$. Thermocycling protocol was as follows: $95^{\circ} \mathrm{C}$ for $5 \mathrm{~min} ; 40$ cycles of $95^{\circ} \mathrm{C}$ for 20 s ; $60^{\circ} \mathrm{C}$ for 30 s and $72^{\circ} \mathrm{C}$ for 30 s; and $72^{\circ} \mathrm{C}$ for 10 min .

PCR products were analyzed using an ABI 3500 Genetic Analyzer. Alleles were sized at each locus in relation to an internal size standard using GeneMapper 3.0 software;
also, sizing was checked by eye. Microsatellite loci were checked for null alleles using Micro-Checker 2.2.3 (Van Oosterhout et al., 2004). Probability of Identity (PI) values and probability of exclusion "when only one parent is known" were determined using GenAlEx 6.41 (Peakall and Smouse, 2006). Observed and expected heterozygosity, linkage equilibrium and deviations from Hardy-Weinberg Equilibrium were calculated and assessed using GenAlEx and GDA (Lewis and Zaykin, 2000) (Table 1).

## Paternity Analysis

Analysis of paternity was evaluated with an exclusion paternity analysis. Each hatchling was determined to have its own multilocus genotype. When known maternal alleles were subtracted from each locus, the remaining paternal alleles formed suspected paternal genotypes. This analysis was performed using the programs GERUD 2.0 and COLONY 2.0. GERUD 2.0 assesses the minimum number of fathers per nest. GERUD 2.0 has been employed in a variety of paternity studies (Jones, 2005; Jones et al., 2010, Yue and Chang, 2010, Zbinden et al., 2007) and can be conservative in its estimates of the number of paternal contributions. COLONY 2.0 is a likelihood-based program that determines the maximum number of fathers per clutch. GERUD 2.0 fails if there are more than 6 fathers, so COLONY 2.0 was used to repeat the analysis on nests that GERUD 2.0 could not evaluate because more than 6 fathers were detected. Using both programs we approximated a range, where GERUD 2.0 was the minimum and COLONY 2.0 was the maximum.

COLONY 2.0 was also used to compare sibling relatedness for all the nests (Jones and Wang, 2010). In this analysis, the error rate of genotyping was set to 0.025 as suggested by Wang (Wang, 2004). The determination of multiple paternity within a nest
was established by the occurrence of more than two paternal alleles over at least two loci-this allowed for the possibility of a mutation at one locus (Yue and Chang, 2010). To determine whether paternal contributions were significantly different from equality in each nest, goodness-of-fit $\left.\right|^{2}$-tests were run on all three years of data. When running $\left.\right|^{2}$ tests, the years were separated as we assume that each year is independent of one another.

The "Probability of Identity" (PI) was estimated using the method employed by GenAlEx within our three year data set.

$$
\begin{gathered}
P I=2\left(\sum\left(p_{i}^{2}\right)^{2}\right)-\sum p^{i^{4}} \\
\text { Where } \mathrm{p}_{\mathrm{i}} \text { is the frequency of the } i \text { th allele at a locus }
\end{gathered}
$$

PI provides an estimate of the average probability that two samples will have the exact same genotype given the estimated allele frequencies of the loci used. The Probability of Exclusion (when only one parent is known) was also determined using GenAlEx to estimate the statistical power of our individual loci and our combined loci (Table 1).

$$
P E=1-4 \sum_{\text {Where } \mathrm{p}_{\mathrm{i}} \text { is the frequency of the } i \text { th allele at a locus }} p_{i}{ }^{2}+2\left(\sum p_{i}{ }^{2}\right)^{2}+4 \sum p_{i}{ }^{3}-3 \sum{ }^{4}
$$

Finally each paternal genotype was compared using COLONY 2.0 and GenAlEx to determine if any of the the estimated paternal genotypes were sampled more than once; furthermore using COLONY we could compare all the nests to these genotypes and determine how many nests an individual male contributed to.

## Statistical Analysis

All three years were analyzed together (2008-2010). Every test that was performed using data from GERUD was also performed using data from COLONY. All analyses were carried out using the program SAS 9.3 (SAS Institute Inc).

The original assumption was that female characteristics should vary randomly by
year and nesting events should be independent of one another: no repeat nesters were analyzed. We ran a generalized linear model with a gamma distribution. As our data was skewed to the left, the gamma distribution was the best model to compare variability within number of fathers due to year, season and female size. The model was run twice (once for each paternity program), with the number of fathers per nest as the dependent variable. Female size (Straight Carapace Length measured in centimeters), the period the nest was laid (early, middle or late), the Julian Date the nests were laid and the year the nest was laid (2008-2010) were all defined as independent variables. Initially I ran the model with all the variables and then removed the independent variable with the highest p -value, determining that it had no effect on the number of fathers per nest. Then I reran the model, removing the next highest and repeated until the only independent variable that remained had p -value less than 0.05 .

I ran another generalized linear model (gamma distribution) to determine if the number of fathers per nest affected hatching success of each nest (percent of nest that did not emerge). Again, I ran two separate tests, the hatching success was defined as the dependent variable and the number of fathers per nest was defined as the independent variable.

Separately using the program JMP 9.0 (SAS Institute Inc) I performed a Wilcoxon/Kruskal-Wallis test to determine if there was a difference in the number of fathers between nesting periods. The nesting period (early, middle and late) was defined as the independent variable and the number of fathers as the dependent variable. This separate test was performed to determine the difference within years, whereas the model examined all nests over all years.

## RESULTS

All five loci amplified consistently over the 92 mothers sampled. The program Pedant determined that the combined allelic dropout rate of the samples was 0.0254 and the false allele rate was 0.1070 (Johnson and Haydon, 2007). Allele number ranged from 12 for the locus $C c \mathrm{P} 7 \mathrm{C} 06$ to 27 for the locus $C c \mathrm{P} 8 \mathrm{D} 06$ (Table 2). Deviations from Hardy-Weinberg were not found. Combining all five loci produced an expected exclusion probability of $99.64 \%$, assuming only one parental genotype is known. The combined probability of identity using the five loci (the likelihood that one independent individual will have the exact same genotype as another) was $1.5 \times 10^{-6}$ (Table 2).

Of the original 92 mothers, 72 nests survived: 23 from 2008, 21 from 2009 and 28 from 2010. If we separate by nesting period, there are 23 early nests, 26 middle nests and 23 late nests. Nests were laid from as early as May 25 to as late as July 17. A total of 1282 hatchlings were sampled and analyzed. The average number of hatchlings genotyped per nest was $18.2(\mathrm{SD}=4.3)$. Six nests had less than 10 hatchlings genotyped. Over the three-year period the average nest size was 114.7 ( $\mathrm{SD}=23.9$ ) and the average percentage of a nest sampled from the nesting beach was $16.5 \% ~(\mathrm{SD}=5.7)$. The average percentage of hatchlings that did not emerge from a nest was $21.7 \%$ ( $\mathrm{SD}=13.7$ ). Average female size (straight carapace length) over the three years was 98.6 cm (SD = 6.5).

In 2008, using both GERUD and COLONY multiple paternal contributions were found in 19 of 23 nests ( $82.6 \%$ ), with an average of $3.00( \pm 0.23)$ males per nest. In 2009, using GERUD multiple paternal contributions were found in 15 of 21 nests ( $71.4 \%$ ) with an average of $2.62( \pm 0.38)$ males per nest (COLONY: 18/21, 85.7\%, $2.8( \pm 0.35))$,
averaging $78.6 \%$ with $2.71( \pm 0.25)$ fathers per nest. In 2010, using GERUD multiple paternal contributions were found in 18 of 28 nests (64.3\%) with an average of 2.20 males per nest (COLONY: 19/28, 67.9\%, $2.32( \pm 0.25)$ ). When all three years are combined, using GERUD multiple paternal contributions were found in 52 of 72 nests ( $72.2 \%$ ) with an average of $2.58( \pm 0.17)$ males per nest, $95 \%$ CI: 2.24-2.93 (COLONY: $56 / 72,77.8 \%, 2.72$ ( $\pm 0.18$ ), $95 \%$ CI: $2.36-3.09$ ) (Figure 1).

## Fathers per Nest

Our conservative estimate (GERUD) illustrated that over all three years 20 nests had 1 father, 17 nests had 2 fathers, 21 nests had 3 fathers, 7 nests had 4 fathers, 3 nests had 5 fathers, 2 nests had 6 fathers and finally 2 nests had more than 6 fathers (GERUD could not handle these data sets). Overall, 22 of 52 (42.3\%) of nests with multiple fathers deviated significantly from equality. In 2008, 6 nests deviated significantly from equal contribution among fathers ( $\chi^{2}$ - tests, $p=0.0001-0.0253$ ), 13 nests with multiple fathers did not deviate significantly $(p=0.1651-0.9284)$ (Figure 2a). In 2009, 6 nests deviated significantly from equal contribution among fathers $\left(\chi^{2}\right.$ - tests, $\left.p=0.0006-0.0379\right), 9$ nests with multiple fathers did not deviate significantly $(\mathrm{p}=0.1651-0.9486)$ (Figure 3a). In 2010, 10 nests deviated significantly from equality ( $\chi^{2}$ - tests, $p=0.0001-0.0492$ ), 8 nests with multiple fathers did not deviate significantly ( $p=0.0736-0.7212$ ) (Figure 4a).

The upper range estimate of COLONY indicates 16 nests had 1 father, 23 nests had 2 fathers, 16 nests had 3 fathers, 7 nests had 4 fathers, 4 nests had 5 fathers, 4 nests had 6 fathers and finally 2 nests had 7 fathers. Overall, 23 of the 56 (41.1\%) nests with multiple fathers deviated significantly from equality. In 2008, 9 nests with deviated significantly from equal contribution among fathers ( $\chi^{2}$ - tests, $p=0.0001-0.0253$ ), 10
nests with multiple fathers did not deviate significantly ( $\mathrm{p}=0.1189-0.9735$ ) (Figure 2b). In 2009, 5 nests deviated significantly from equal contribution among fathers $\left(\chi^{2}-\right.$ tests, $p$ $=0.0001-0.0384), 13$ nests with multiple fathers did not deviate significantly $(\mathrm{p}=$ $0.0736-0.9486$ ) (Figure 3b). In 2010, 9 nests deviated significantly from equality ( $\chi^{2}$ tests, $\mathrm{p}=0.0017-0.0423$ ), 10 nests with multiple fathers did not deviate significantly ( p $=0.0736-0.9402)($ Figure 4b).

## Generalized Linear Model

The conservative generalized linear model (Table 4a) shows that when all three variables (year, season and female size) are combined, the variation in the number of fathers per nest was not explained significantly by any of the independent variables ( $\mathrm{p}=$ 0.1303, 0.7410-0.8275, 0.2050 respectively). However when year and female size are run separately, both variables explained significant variation to the number of fathers per nest $\left(\chi^{2}=4.05, p=0.0441\right.$ and $\chi^{2}=5.02, p=0.0251$ respectively). The number of fathers per nest decreases as the years go on (2008-2010, Figure 5), as female size increases, the number of fathers increases (Figure 6). The number of fathers per nest does not explain variation in nest hatching success $\left(\chi^{2}=1.93, \mathrm{p}=0.1645\right)$ (Figure 7).

The less conservative (COLONY) generalized linear model (Table 4b) shows that when all three variables (year, season and female size) are combined, the variation in the number of fathers per nest was not explained significantly by any of the independent variables ( $\mathrm{p}=0.1712,0.6610-0.7981,0.2091$ respectively). However when year and female size are run separately, both variables explained significant variation to the number of fathers per nest $\left(\chi^{2}=3.96, p=0.0465\right.$ and $\chi^{2}=4.63, p=0.0315$ respectively). As year decreases, the number of fathers per nest increases (Figure 5) and as female size
increases, the number of fathers per nest increases (Figure 6). The number of fathers per nest explained no variation in nest hatching success $\left(\chi^{2}=0.97, \mathrm{p}=0.3258\right)$ (Figure 7).

Qualitatively: there was a relationship between female size and nest size $\left(\mathrm{r}^{2}=\right.$ $\left.0.419, \mathrm{~F}_{(1,64)(0.05)}=46.26, \mathrm{p}<0.0001\right)$ (Figure 8).

## Season

When all three years were compared in the model, there was no variation in the number of fathers per nest due to the date the eggs were laid (nesting period: Julian Nest Date). A Wilcoxon/Kruskal-Wallis was performed to determine if the number of fathers per nest varied over the course of the nesting period within each year and between years (not over the course of all three years). Like the model however, there was no difference in the number of fathers due to the nesting period for any of the three years. In 2008: df $=2$, GERUD: $\chi^{2}=1.42, \mathrm{p}=0.4911$, COLONY: $\chi^{2}=0.48, \mathrm{p}=0.7867$ ) (Figure 9a); in 2009: $\left(\mathrm{df}=2\right.$, GERUD: $\chi^{2}=5.33, \mathrm{p}=0.0696$, COLONY: $\left.\chi^{2}=3.43, \mathrm{p}=0.1795\right)$ (Figure 9b); finally in 2010: ( $\mathrm{df}=2$, GERUD: $\chi^{2}=1.49, \mathrm{p}=0.4726$, COLONY: $\chi^{2}=0.72, \mathrm{p}=$ 0.6976 ) (Figure 9c). When all three years were combined, there was no difference in the number of fathers per nest due to the nesting season ( $\mathrm{df}=2$, GERUD: $\chi^{2}=1.75, \mathrm{p}=$ 0.4174 , COLONY: $\chi^{2}=2.12, p=0.3459$ ) (Figure 9 d ). When comparing the number of fathers per nest directly to the Julian Nest Date (the date the nest was laid), there was no significant trend (Figure 10).

## Paternal Genotype Analysis

One hundred and ninety five (195) individual male genotypes were inferred using COLONY over all three years. Figure 12 shows the relationship between nests and fathers, there are no half-sibs, suggesting that every father only contributed to one nest
over all three years.

## DISCUSSION

Multiple paternity has previously been documented in loggerhead sea turtles across the globe. Harry \& Briscoe (1988) determined in Queensland, Australia that 8 of 21 nests samples (38\%) had multiple paternal contributions. In Florida, two studies independently determined that just over $30 \%$ of nests showed multiple paternal contributions (Bollmer et al., 1999 and Moore \& Ball, 2002). Lastly, in Greece, Zbinden et al. (2007) found multiple paternal contributions in 19 of 20 nests ( $95 \%$ ) and found a maximum of 5 fathers per nest. These studies were all conducted at large nesting aggregations (greater than 1000 nests per nesting season) and with the exception of the study in Greece, did not take samples from more than one nesting season.

In my study, over the course of the three year sampling period, the average nest size of C. caretta on Wassaw Island was $114.7(\mathrm{SD}=23.9)$ and emergence success was 0.773 (Caretta Research Project, personal correspondence). The average nest size is comparable to values suggested by NMFS and USFWS of 115, but is higher than those values for emergence of the Northwestern Atlantic Ocean DPS: 0.54 (Conant et al., 2009). It could be suggested that our presence on the small beach of Wassaw is positively influencing the emergence success (less predation due to patrols, caging of nests and people present on the beach). However this could just be a result of the smaller nesting count on a small island in relation to the rest of the DPS, as beaches throughout the more heavily nested areas (such as in Florida) would have more human traffic and lower percentages of nests caged. Also, emergence success of larger nesting aggregations are more likely to approach the mean.

More than one male sired 75\% (72.2-77.8\%) of nests on Wassaw Island during the three year period studied. The average minimum number of fathers per nest was 2.65 (2.58-2.72). Even though Wassaw is a much smaller nesting beach (less nesting females) than either Florida or Greece, this estimate is nearly double the findings in the large Florida rookery of $33 \%$ and 1.4 fathers per nest (Moore \& Ball, 2002). But is less than estimates found in Greece of $95 \%$ and 3.5 fathers per nest (Zbinden et al., 2007) (Figure 11). Previous studies have suggested that larger nesting aggregations should increase the likelihood of multiple paternity (Jensen et al. 2006; Uller and Olsson, 2008; etc.) as there is a positive correlation between multiple paternity and the probability of mate encounters. Regarding the Greece data, this theory makes sense, but there must be something else limiting this theory with regards to Florida (one of the largest nesting aggregations in the world). Perhaps because there are less females nesting in Georgia and Greece, males are able to mate with as many females as possible, but in Florida there are so many nesting females that there is a limit to how many females males can actually mate with (eventually with 30,000 females, a males sperm will run out).

Multiple paternity may be favored if it increases the variability of the offspring (Uller and Olsson, 2008, Olsson et al., 1994), but these are not necessarily related. As I cannot directly study the fitness of all the hatchlings, an easy way to assess variability is by looking at the how many hatchlings actually pip out of the egg. There was no relationship between hatching success and the number of fathers per nest. In other words, the number of fathers per nest did not directly skew the hatchlings to be more or less likely to survive. This could suggest that more fathers do add to the variability of hatchlings, but not to the viability of the hatchlings. We posit that with more fathers there
should be more variability, every additional male will have both "good alleles" and "bad alleles." In this example, "good alleles" favor the hatchling survival (ie. pipping out of the egg) and "bad alleles" could hinder the hatchling survival. By adding variation to a nest it is unlikely every allele passed along will be "good" for the hatchlings. But because female turtles cannot prepare for what their young will face in their future, bethedging against an unpredictable environment could still provide some benefits.

A major goal of this study was to assess the full nesting population of Wassaw Island, GA in comparison to the distinct population segment as a whole. Previous studies within the distinct population segment have focused on the large rookery of Florida (Bollmer et al., 1999; Moore \& Ball, 2002). These studies, as well as others outside of the distinct population segment, have focused on assessing how multiple paternity affects the nesting population directly. (Bollmer et al., 1999; Moore \& Ball, 2002; Zbinden et al., 2007 ; etc.). None of these studies have looked at the trends of multiple paternity over the course of time.

There were three major reasons for us to look at changes in the frequency of multiple paternity in this nesting population over time. First, female turtles return to nest once every three years (Conant et al., 2009); by looking at three years of data, we are essentially looking at a snapshot of the "whole" nesting population on this island. Second, there was a steady decrease of nesting numbers from 1970's until 2008, with a rise of nesting numbers from 2009 to 2011 (Federal Register, 2011), we wanted to see if this sudden increase in nesting affect the number of nests with multiple fathers. Finally, there was an unseasonably cold and wet season in 2009 that invariably affected how many nests I could analyze (only 21 of 30 original nests). As females nest once every three
years on Wassaw, the number of nests per year should be random, allowing our assessment of the number of fathers per nest to be unbiased. But in 2009 many nests were lost due to events outside of the experiment's control. By losing those nests, it is possible that my data has become skewed. There was significant variation in the number of fathers per nest due to the year: from 2008-2010, the number of fathers per nest did increase. However, this finding does not mean that the year caused this increase, the variation is only related to each year, suggesting that each year could affect the results (ie. the difference from 2009 could have been important, but I did not test for that specifically).

We found that the number of fathers per nest did vary with regards to female size (straight carapace length, cm ), such that as female size rose, so did the number of fathers. This is most likely attributed to larger females being older (Casale et al., 2011). Males probably favor older females because they are likely more fecund, they have lived long enough that they have reproduced more than once and if they are larger they can hold more eggs. It is also possible that larger females can swim faster to mating grounds and would have a longer mating period (untested). The females varied significantly in 2010 in comparison to the other two years, the females were smaller, suggesting that these turtles were young. Regarding the difference in number of fathers per nest by year, this could be a difference in sample size: there are 5+ more nests analyzed in 2010 than from 2008 (23) or 2009 (21). Or there might not be any trend at all and 2010 was just an anomaly; the only way to tell for sure would be to continue sampling.

Coupled together, there was no difference in the number of fathers per nest in relation to year and female size. This could suggest that while the weather of 2009 might
have affected the nests on the beach, there was no connection between the climate and the mating offshore. This also suggests that while females in 2010 were smaller and there were less fathers per nest, there was no real difference in the number of fathers per nest because of the smaller females.

There was no difference in the number of fathers per nest due to nesting period. This is consistent with significant sperm mixing at the beginning of the season. (Pearse et al., 2002; Uller and Olsson, 2008; etc.) Females store the sperm in their oviduct after mating and can potentially hold the sperm for at least a year. (Uller and Olsson, 2008) If the sperm from male to male was being packed in one after another, it could be possible to see a skew in the number of fathers per nest over the course of the season. However, because there was no pattern of multiple fathers or single fathers, sperm must mix.

Additionally, $21 \%$ of the nests sired by more than one male showed skewed paternal contributions (ie. proportions of the nest were biased to one of the fathers). When a nest is skewed to one father over another, it is typically hypothesized as evidence for cryptic female choice (allowing females to preferentially choose sperm) or for the promotion of "good alleles," thus supporting sperm competition (Uller and Olsson, 2008). Furthermore, we found that there was no difference in hatching success due to the number of fathers per nest, suggesting that females do not mate repeatedly to guarantee fertilization.

This study estimated 195 unique paternal genotypes. No male sired more than one nest throughout the three years, we posit that this nesting population's mating system is polyandrous and not polygynous. It is probable that males are contributing to other nests, but our data does not support this.

Counting males is difficult as males rarely return to the beach and they return to their feeding grounds immediately after the mating season. By comparing male genotypes we can "count" without having to witness the males or the mating itself. As hatchling sex ratios are becoming increasingly female biased (Hanson et al., 1998; Delgado et al., 2010 etc.), it is important to determine how many males are actually contributing to this nesting population. If we assume that each year in this study is independent of one another, that the females nesting in 2008 are not nesting in 2009 and females in 2009 are not nesting in 2010, then we have to look at the ratio of males to nests individually (3 years, 3 sub-nesting-populations: $3.00,2.71,2.67$ ). But because we didn't find any males that contributed to more than one year's nests we can take the average number of males per nests over all three years ( 2.65 males to nests).

The estimate in Florida, is that there is a 1:1 male to female adult ratio mating and contributing to the nesting beaches, in other words if there are 30,000 females nesting per year in Florida then there are 60,000 active adults (Federal Register, 2011). If the ratio of number of fathers to nests (ie. number of females) is correct and not just a remnant of this experiment's small sample size, Northern nesting beaches could be essential to the continuation of a healthy nesting population of the Northwestern Atlantic distinct population segment.

Table 1
Percentage of nests showing multiple paternity within sea turtles from previous studies using microsatellite markers.

| Species | \# Clutches Analyzed | \% Multiple Paternity | Citation |
| :--- | :---: | :---: | :--- |
| Caretta caretta | 70 | $31 \%(22 / 70)$ | Moore \& Ball, 2002 |
| C. caretta | 20 | $95 \%(19 / 20)$ | Zbinden et al., 2007 |
| Chelonia mydas | 22 | $9 \%(2 / 22)$ | Fitzsimmons, 1998 |
| C. mydas | 18 | $61 \%(11 / 18)$ | Lee \& Hays, 2004 |
| Dermochelys coriacea | 20 | $10 \%(2 / 20)$ | Crim et al., 2002 |
| Lepidochelys olivacea | 13 | $30 \%(4 / 13)$ | Jensen et al., 2006 |
| L. olivacea | 13 | $92 \%(12 / 13)$ | Jensen et al., 2006 |
| Lepidochelys kempi | 26 | $58 \%(15 / 26)$ | Kichler et al., 1999 |

Table 2
Descriptive statistics of the five polymorphic microsatellite markers used Number of alleles $(A)$, expected heterozygosity $\left(H_{E}\right)$ and observed heterozygosity $\left(H_{O}\right)$.

| Locus | Size Range (bp) | Dye | $\mathbf{A}$ | $\mathbf{H}_{\mathbf{E}}$ | $\mathbf{H}_{\mathbf{O}}$ | Expected <br> Exclusion <br> Probability |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| CcP7E05 | $164-236$ | 6FAM | 18 | 0.920 | 0.978 | 0.695 |
| CcP2F11 | $252-308$ | 6FAM | 16 | 0.892 | 0.956 | 0.626 |
| CcP7D04 | $320-376$ | 6FAM | 14 | 0.907 | 0.913 | 0.669 |
| CcP7C06 | $256-296$ | HEX | 12 | 0.864 | 0.858 | 0.541 |
| CcP8D06 | $256-376$ | TAMRA | 27 | 0.941 | 0.956 | 0.792 |

## Table 3

Descriptive table of multiple paternity by year, using GERUD, COLONY and the average between the two programs. The percentage is determined by dividing the number of nests with multiple fathers by the total number of nests and multiplying by 100. The second number is the average number of fathers per nest and SE.

| Year | GERUD | COLONY | Average |
| :---: | :---: | :---: | :---: |
| $\mathbf{2 0 0 8}$ | $19 / 23=82.6 \%$ | $19 / 23=82.6 \%$ | $19 / 23=82.6 \%$ |
|  | $3.00( \pm 0.29)$ | $3.00( \pm 0.35)$ | $3.00( \pm 0.23)$ |
| $\mathbf{2 0 0 9}$ | $15 / 21=71.4 \%$ | $18 / 21=85.7 \%$ | $16.5 / 21=78.6 \%$ |
|  | $2.62( \pm 0.38)$ | $2.80( \pm 0.35)$ | $2.71( \pm 0.25)$ |
| $\mathbf{2 0 1 0}$ | $18 / 28=64.3 \%$ | $19 / 28=67.9 \%$ | $18.5 / 28=66.1 \%$ |
|  | $2.21( \pm 0.23)$ | $2.32( \pm 0.25)$ | $2.67( \pm 0.17)$ |
| Average | $52 / 72=72.2 \%$ | $56 / 72=77.8 \%$ | $54 / 72=75 \%$ |
|  | $2.58( \pm 0.17)$ | $2.72( \pm 0.18)$ | $2.65( \pm 0.13)$ |

Table 4a
Generalized Linear Model for GERUD, distribution gamma. All variables have 1 degree of freedom, $N=72$, Chi-square values are first, followed by the $p$-value.

| Parameter | Initial Model | Step 1 | Step 2 | Run Alone |
| :---: | :---: | :---: | :---: | :---: |
| Year <br> (2008-2010) | $2.29,0.1303$ | $2.34,0.1259$ | $2.18,0.1394$ | $\mathbf{4 . 0 5 , 0 . 0 4 4 1}$ |
| Straight Carapace <br> Length (cm) <br> Season <br> (Early - Middle) | $1.61,0.2050$ | $2.17,0.1408$ | $3.34,0.0676$ | $\mathbf{5 . 0 2 , 0 . 0 2 5 1}$ |
| Season <br> (Middle - Late) <br> Julian Nest Date | $0.05,0.8275$ | $0.21,0.6444$ |  |  |
| Hatching Success | $0.11,0.7410$ |  | $1.93,0.1645$ |  |

Table 4b
Generalized Linear Model for COLONY, distribution gamma. All variables have 1 degree of freedom, $N=72$, Chi-square values are first, followed by the $p$-value.

| Parameter | Initial Model | Step 1 | Step 2 | Run Alone |
| :---: | :---: | :---: | :---: | :---: |
| Year <br> (2008-2010) <br> Straight Carapace <br> Length (cm) <br> Season <br> (Early - Middle) | $1.87,0.1712$ | $1.91,0.1675$ | $1.80,0.1803$ | $\mathbf{3 . 9 6 , 0 . 0 4 6 5}$ |
| Season <br> (Middle - Late) <br> Julian Nest Date | $0.07,0.7880$ | $0.01, .9257$ | $2.91,0.0880$ | $\mathbf{4 . 6 3 , 0 . 3 1 5}$ |
|  | $0.07,0.7981$ | $1.54,0.2152$ |  |  |
| Hatching Success |  | $0.31,0.5801$ |  |  |



Figure 1: Graph showing the relationship between the two programs used and the three years analyzed. $\left(\mathrm{N}_{2008}=23, \mathrm{~N}_{2009}=21, \mathrm{~N}_{2010}=28\right)$. The gray bars are GERUD by year, the white bars are COLONY by year, G F/N represents the number of fathers per nest according to GERUD and C F/N represents the number of fathers per nest according to COLONY
(a)

(b)


Figure 2: Relative contribution of fathers to clutches sired by multiple fathers from 2008. (a): Data estimated using GERUD. (b): Data estimated using COLONY. Stars indicate nests that paternal contributions did deviate significantly from equality (goodness of fit $\chi^{2}$-tests, $P>0.05$ ) in a nest.

(a)

Figure 3: Relative contribution of fathers to clutches sired by multiple fathers from 2009. (a): Data estimated using GERUD. (b): Data estimated using COLONY. Stars indicate nests that paternal contributions did deviate significantly from equality (goodness of fit $\chi^{2}$-tests, $P>0.05$ ) in a nest.


Figure 4: Relative contribution of fathers to clutches sired by multiple fathers from 2010. (a): Data estimated using GERUD. (b): Data estimated using COLONY. Stars indicate nests that paternal contributions did not deviate significantly from equality (goodness of fit $\chi^{2}$-tests, $P>0.05$ ) in a nest.


Figure 5: This graph shows the difference between the average number of fathers by year. There is a visually negative trend as the years increase and using the generalized linear model the trend is significant $\left(\mathrm{G}: \chi^{2}=4.05, \mathrm{p}=0.0441 ; \mathrm{C}:\left.\right|^{2}=3.96, \mathrm{p}=0.0465\right.$ ).


Figure 6: The relationship between female size (Straight Carapace Length, cm ) and the number of fathers per nest. There is a significant difference in number of fathers due to female size: as female size increases, so does the number of fathers $\left(G:\left.\right|^{2}=5.02, p=\right.$ $\left.0.0251 ; C:\left.\right|^{2}=4.63, p=0.0315\right)$.


- GERUD $\times$ COLONY

Figure 7: The relationship between the number of fathers per nest and the hatching success (\%) of each nest, using both programs. The generalized linear model determined there was no significant relationship.


Figure 8: Relationship between Female Straight Carapace Length (cm) and the number of eggs per nest (ie. nest size). There is a positive correlation between the two, as females get larger, the number eggs per nest increases $\left(\mathrm{r}^{2}=0.419, \mathrm{~F}_{(1,64)(0.05)}=46.26, \mathrm{p}<0.001\right)$
(a)

(b)

(c)

(d)


Figure 9: Nesting period (early, middle and late) versus the average number of fathers, (a): 2008, (b): 2009, (c): 2010 and (d): all years combined. The bars represent +1 Standard Deviation. There is no significant difference in the number of fathers due to nesting period across all years.


- GERUD $\times$ COLONY

Figure 10: Julian Nest Date compared to number of fathers per nest. There is a visual negative trend: as the nesting season progresses, the number of fathers decreases (not significant).


Figure 11: Relationship between location, percentage of nests sired by multiple males and the average number of fathers per nest by location. Adapted using data from Moore and Ball, 2002 and Zbinden et al., 2007.


Figure 12: Relatedness of siblings. Estimate of fathers contributing to each nest over all three years. Triangles above the transect represent individual nests (fully related siblings). Triangles below the transect represent individuals who are related to each other (halfsiblings).

## REFERENCES

"Listing and Protecting Loggerhead Sea Turtles as 'Threatened Species’ and Populations of Green and Olive Ridley Sea Turtles as Threatened Species or 'Endangered Species.'" 43 Federal Register 146 (28 July 1978), pg. 32800-32811.
"Endangered and Threatened Species; Determination of Nine Distinct Population Segments of Loggerhead Sea Turtles as Endangered or Threatened." 76 Federal Register 184 (22 September 2011), pg 58868-58952.

Barnard, D.E. and Keinath, J.A. (1999). Sea turtle activities on Blackbeard Island National Wildlife Refuge, Georgia, 1999. Submitted to Savannah Coastal Unit, USFWS.

Beasley, J.C., Beatty, W.S., Olson, Z.H. and Rhodes Jr., O.E. (2010). A Genetic Analysis of the Virginia Opossum Mating System: Evidence of Multiple Paternity in a Highly Fragmented Landscape. Journal of Heredity 101 (3): 368-373.

Bjorndal, K.A., Meylan, A.B., and Turner, B.J. (1983). Sea turtles nesting at Melbourne Beach, Florida, I. Size, growth and reproductive biology. Biological Conservation 26: 65-77.

Bollmer, J.L., Irwin, M.E., Reider, J.P., and Parker, P.G. (1999). Multiple paternity in loggerhead turtle clutches. Copeia 2: 475-478.

Bowen, B.W. (2004). Conservation Implications of Complex Population Structure in the Loggerhead Turtle (Caretta caretta): Final Report to the Seatheast Fisheries Science Center. Hawaii Institute of Marine Biology, University of Hawaii, Kaneohe, HI.

Bowen, B.W., Bass, A.L. , Chow, S., Bostrom, M., Bjorndal, K.A., Bolten, A.B., Okuyama, T., Bolker, B.M., Epperly, S., LaCasella, E., Shaver, D., Dodd, M., Hopkins-Murphy, S.R., Musick, J.A., Swingle, M., Rankin-Baransky, K., Teas, W., Witzell, W.N. and Dutton, P.H. (2004). Natal homing in juvenile loggerhead turtles (Caretta caretta). Molecular Ecology 13: 3797-3808.

Brown, J.L. (1997). A theory of mate choice based on heterozygosity. Behavioral Ecology 8: 60-65.

Caldwell, D.K. (1962). Comments on the nesting behavior of Atlantic loggerhead sea turtles, based primarily on tagging returns. Quarterly Journal of the Florida Academy of Sciences 25(4): 287-302.

Casale, P., Freggi, D. Basso, R. and Argano, R. (2005). Size at Male Maturity, Sexing Methods and Adult Sex Ratio in Loggerhead Turtles (Caretta caretta) from Italian Waters Investigated Through Tail Measurements. The Herpetological Journal 15(3): 145-148.

Casale, P., Mazaris, A.D. and Freggi, D. (2011). Estimation of age at maturity of loggerhead sea turtles Caretta caretta in the Mediterranean using length frequency data. Endangered Species Research 13: 123-129.

Crim, J.L., Spotila, L.D., Spotila, J.R., O’Connor, M., Reina, R., Williams, C.J. and Paladino, F.V. (2002). The leatherback turtle, Dermochelys coriacea, exhibits both polyandry and polygyny. Molecular Ecology 11: 2097-2106.

Conant, T.A., Dutton, P.H., Eguchi, T., Epperly, S.P., Fahy, C.C., Godfrey, M.H., MacPherson, S.L., Possardt, E.E., Schroeder, B.A., Seminoff, J.A., Snover, M.L., Upite, C.M. and Witherington, B.E. (2009). Loggerhead sea turtle (Caretta caretta) 2009 status review under the U.S. Endangered Species Act. Report of the Loggerhead Biological Review Team to the National Marine Fisheries Service, August 2009. 222 pages.

Delgado, C., Canario, A.V.M. and Dellinger, T. (2010). Sex ratios of loggerhead sea turtles Caretta caretta during the juvenile pelagic stage. Marine Biology 157: 979990.

Dodd, C.K. Jr. (1988). Synopsis of the biological data on the loggerhead sea turtle Caretta caretta (Linnaeus 1758). USFWS, Biology Report 88(14). 110 pg.

Dodd, C.K., Jr., and Byles, R.A. (2003). Post-nesting movements and behavior of loggerhead sea turtles (Caretta caretta) departing from east-central Florida nesting beaches. Chelonian Conservation and Biology 4(3): 530-536.

Drake, K. (M.S. 2001). Reproductive parameters of nesting Caretta caretta on Georgia's Barrier Islands. M.S. Thesis, Georgia Southern University (pp. 110).

Encalada, S.E., Bjorndal, K.A., Bolten, A.B., Zurita, J.C., Schroeder, B., Possardt, E., Sears, C.J., Bowen, B.W. (1998). Population structure of loggerhead turtle (Caretta caretta) nesting colonies in the Atlantic and Mediterranean as inferred from mitochondrial DNA control region sequences. Marine Biology 130: 567-575.

FitzSimmons, N.N., Limpus, C.J., Norman, J.A., Goldizen, A.R., Miller, J.D., and Moritz, C. (1997). Philopatry of male marine turtles inferred from mitochondrial DNA markers. Proceedings of the National Academy of Sciences (PNAS) U.S.A. 94: 8912-8917.

Galbraith, D.A. (1993). Multiple paternity and sperm storage in turtles. Herpetological Journal 3: 117-123.

Gist, D.H. and Jones, J.M. (1989). Sperm storage within the oviducts of turtles. Journal of Morphology 199: 379-384.

Hanson, J., Wibbels, T. and Martin, R.E. (1998). Predicted female bias in sex ratios of hatchling loggerhead sea turtles from a Florida nesting beach. Canadian Journal of Zoology 76(10): 1850-1861.

Harry, J.L. and Briscoe, D.A. (1988). Multiple paternity in the loggerhead turtle (Caretta caretta). Journal of Heredity 79(2): 96-99.

Hoekert W.E.J., Neufeglise H., Schouten A.D. and Menken S.B.J. (2002). Multiple paternity and female-biased mutation at a microsatellite locus in the olive ridley sea turtle (Lepidochelys olivacea). Heredity 89: 107-113. Accessed on http://www.nature.com/hdy/journal/v89/n2/full/6800103a.html on 5-27-09.

Ireland, J.S., Broderick, A.C., Glen, F., Godley, B.J., Hays, G.C., Lee, P.L.M. and Skibinski, D.O.F. (2003). Multiple paternity assessed using microsatellite markers, in green turtles Chelonia mydas (Linnaeus, 1758) of Ascension Island, South Atlantic. Journal of Experimental Marine Biology and Ecology 291: 149-160.

Jensen, M.P., Abreu-Grobois, F.A., Frydenberg, J. and Loeshcke, V. (2006). Microsatellites provide insight into contrasting mating patterns in arribada vs. non-arribada olive ridley sea turtle rookeries. Molecular Ecology 15: 2567-2575.

Johnson, P.C.D. and Haydon, D.T. (2007). Maximum-Likelihood Estimation of Allelic Dropout and False Allele Error Rates From Microsatelite Genotypes in the Absence of Reference Data. Genetics 175(2): 827-842.

Jones, A.G. (2005). GERUD 2.0: a computer program for the reconstruction of parental genotypes from half-sib progeny arrays with known or unknown parents. Molecular Ecology Notes 5(3): 708-711.

Jones, O.R. and Wang, J. (2010) COLONY: a program for parentage and sibship inference from multilocus genotype data. Molecular Ecology Resources 10(3): 551-555.

Kalinowski, S.T., Taper, M.L. and Marshall, T.C. (2007). Revising how the computer program CERVUS accommodates genotyping error increases success in paternity assignment. Molecular Ecology 16: 1099-1006.

Kichler, K., Holder, M.T., Davis, S.K., Marquez-M, R. and Owens, D.W. (1999). Detection of multiple paternity in the Kemp's ridley sea turtle with limited sampling. Molecular Ecology 8: 819-830.

LeBuff, C.R. and Beatty, R.W. (1971). Some aspects of nesting of the loggerhead turtle, Caretta caretta (Linne) on the Gulf Coast of Florida. Herpetologica 27: 153-156.

Lee, P.L.M and Hays, G.C. (2004). Polyandry in a marine turtle: females make the best of a bad job. Proceedings of the National Academy of Sciences USA 101:6530-6535.

Lewis, P.O. and Zaykin, D. (2000) Genetic Data Analysis: available at http://hydrodictyon.eeb.uconn.edu/people/plewis/software.php. Last accessed on the $14^{\text {th }}$ of September, 2011.

Limpus, C.J. and Reed, P.C. (1985). The loggerhead turtle, Caretta caretta, in Queensland: breeding males in the southern Great Barrier reef. Australians Wildlife Research 12: 535-540.

McTaggert, S.J. (2000). Good Genes or Sexy Sons? Testing the Benefits of Female Choice in the Painted Turtle, Chrysemys picta. Unpubl. Master's thesis, University of Guelph, Ontario, Canada.

Moore, M.K. and Ball, R.M. (2002). Multiple paternity in loggerhead turtle (Caretta caretta) nests on Melbourne Beach: a microsatellite analysis. Molecular Ecology 11: 281-288.

Moore, J.A., Nelson, N.J., Keall, S.N. and Daugherty, C.H. (2008). Implications of social dominance and multiple paternity for the genetic diversity of a captive bred reptile population (Tuatara). Conservation Genetics 9: 1243-1251.

Moran, S., Turner, P.D. and O'Reilly, C. (2009). Multiple paternity in the European hedgehog. Journal of Zoology 278: 349-353.

National Marine Fisheries Service and U.S. Fish and Wildlife Service (NMFS and USFWS). (2008). Recovery Plan for the Northwest Atlantic Population of the Loggerhead Sea Turtle (Caretta caretta), Second Revision. National Marine Fisheries Service, Silver Spring, MD.

National Oceanic and Atmospheric Administration (NOAA) and U.S. Fish and Wildlife Service (USFWS). (2007). Green Sea Turtle (Chelonia mydas), 5 Year Review: Summary and Evaluation. Accessed and Downloaded from www.fws.gov/northflorida/SeaTurtles/2007-Reviews/2007-green-turtle-5-year-review-final.pdf on 5-30-09.

NOAA Fisheries: Office of Protected Resources. (2009). Loggerhead Turtle (Caretta caretta). Accessed on http://www.nmfs.noaa.gov/pr/species/turtles/loggerhead.htm on 5-26-09.

Olsson, M., Gullberg, A., and Tegelstrom, H. (1994). Sperm competition in the sand lizard, Lacerta agilis. Animal Behavior 48 :193-200.

Palmer, K.S., Rostal, D.C., Grumbles, J.S. and Mulvey, M. (1998). Long-term sperm storage in the desert tortoise (Gopherus agassizii). Copeia 1998: 702-705.

Peakall, R. and Smouse, P.E. (2006). GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and reseach. Molecular Ecology Notes 6: 288-295.

Pearse, D.E. and Avise J.C. (2001). Turtle Mating Systems: Behavior, Sperm Storage and Genetic Paternity. The American Genetic Association 92: 206-211.

Pearse, D.E., Janzen, F.J. and Avise, J.C. (2002) Multiple paternity, sperm storage, and reproductive success of female and male Painted Turtles (Chrysemys picta) in nature. Behavioral Ecology and Sociobiology 51: 164-171.

Pearse, D.E. and Anderson, E.C. (2009). Multiple paternity increases effective population size. Molecular Ecology 18: 3124-3127.

Quinteiro, J., Baibai, T., Oukhattar, L., Soukri, A., Seixas, P and Rey-Mendez, M. (2011). Multiple paternity in the common octopus Octopus vulgaris (Cuvier, 1797), as revealed by microsatellite DNA analysis. Molluscan Research 31 (1): 15-20.

Refsnider, J.M. (2009). High Frequency of Multiple Paternity in Blanding's Turtle (Emys blandingii). Journal of Herpetology 43(1): 74-81.

Richards, P.M., Epperly, S.P., Heppell, S.S., King, R.T., Sasso, C.R., Moncada, F., Nodarse, G., Shaver, D.J., Medina, Y. and Zurita, J. (2011). Sea turtle population estimates incorporating uncertainty: a new approach applied to wester North Atlantic loggerheads Caretta caretta. Endangered Species Research 15: 151-158.

Roques, S., Diaz-Paniagua, C. and Andreu, A.C. (2004). Microsatellite markers reveal multiple paternity and sperm storage in the Mediterranean spur-thighed tortoise, Testudo graeca. Canadian Journal of Zoology 82: 153-159.

Shamblin, B.M., Faircloth, B.C., Dodd, M.G., Bagley, D.A., Ehrhart, L.M., Dutton, P.H., Frey, A., and Nairn, C.J. (2009). Tetranucleotide markers from the loggerhead sea turtle (Caretta caretta) and their cross amplification in other marine turtle species. Conservation Genetics 10: 577-580.

Shamblin, B.M., Dodd, M.G., Bagley, D.A., Ehrhart, L.M., Tucker, A.D., Johnson, C., Carthy, R.R., Scarpino, R.A., McMichael, E., Addison, D.S., Williams, K.L., Frick, M.G., Ouellette, S., Meylan, A.B., Godfrey, M.H., Murphy, S.R. and Nairn, C.J. (2011). Genetic structure of the southeastern United States loggerhead turtle nesting aggregation: evidence of additional structure within the peninsular Florida recovery unit. Marine Biology 158: 571-587.

Schofield, G., Katselidis, K.A., Dimopoulos, P., Pantis, J.D. and Hays, G.C. (2006). Behaviour analysis of the loggerhead sea turtle Caretta caretta from direct inwater observation. Endangered Species Research 2: 71-79.

Shoop, C.R., Ruckdeschel, C.A. and Kenney, R.D. (1998). Female-Biased Sex Ratio of Juvenile Loggerhead Sea Turtles in Georgia. Chelonian Conservation and Biology 3(1): 93-96.

Steinitz, M.J., Salmon, M. and Wyneken, J. (1998). Beach renourishment and loggerhead turtle reproduction: a seven year study at Jupiter Island, Florida. Journal of Coastal Research 14(3): 1000-1013.

Stockley, P., Searle, J.B., MacDonald, D.W., and Jones, C.S. (1993). Female multiple mating behavior in the common shrew as a strategy to reduce inbreeding. Proceedings of the Royal Society of London Series B 254: 173-179.

Uller, T. and Olsson, M. (2008). Multiple paternity in reptiles: patterns and processes. Molecular Ecology 17: 2566-2580.
U.S. Fish and Wildlife Service: North Florida Field Office (NF USFW). (2009). Sea Turtle Information and Species Fact Sheets. Accessed on http://www.fws.gov/northflorida/SeaTurtles/seaturtle-info.htm on 5-29-09.

Van Oosterhout, C., Hutchinson, W.F., Wills, D.P.M. and Shipley, P. (2004). MICROCHECKER: software for identifying and correcting genotyping errors in microsatellite data. Molecular Ecology Notes 4: 535-538.

Walker, D., Power, A.J., Sweeney-Reeves, M. and Avise, J.C. (2007). Multiple paternity and female sperm usage along egg-case strings of the knobbed whelk, Busycon carica (Mollusca; Melongenidae). Marine Biology 151: 53-61.

Wang, J. (2004). Sibship reconstruction from genetic data with typing errors. Genetics 166: 1963.

Wibbels, T., Martin, R.E., Owens, D.W., Amoss, M.S. Jr.. (1991). Female-biased sex ratio of immature loggerhead sea turtles inhabiting the Atlantic coastal waters of Florida. Canadian Journal of Zoology 69(12): 2973-2977.

Williams, K.L. And Frick, M.G. (2000). Annual Report for Caretta Research Project for Wassaw National Wildlife Refuge, Georgia.

Wright, S. (1931). Evolution in Mendelian populations. Genetics 16: 97-159.
Yasui, Y. (2001). Female multiple mating as a genetic bet-hedging strategy when mate choice criteria are unreliable. Ecological Research 16: 605-616.

Yntema, C.L. and Mrosovsky, N. (1980). Sexual Differentiation in Hatchling Loggerheads (Caretta caretta) Incubated at Different Controlled Temperatures. Herpetologica, 36 (1): 33-36.

Yue, G.H., and Chang, A. (2010). Molecular Evidence for High Frequency of Multiple Paternity in a Freshwater Shrimp Species Caridina ensifera. PLos One 5 (9):e12721.doi:10.1371/journal.pone.0012721.

Zbinden, J.A., Largaider, C.R., Leippert, F., Margaritoulis, D. and Arlettaz, R. (2007). High frequency of multiple paternity in the largest rookery of Mediterranean loggerhead sea turtles. Molecular Ecology 16: 3703-3711.

APPENDIX A: FULL NEST DATA

| Season | Date Laid | JND | $\begin{gathered} \hline \text { Nest } \\ \# \\ \text { (CRP) } \\ \hline \end{gathered}$ | Year | Mother SCL | Mother SCW | N | Nest Total | \% Nest Sampled | Hatching Success | HS\% | Gerud | Colony |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 28- |  |  |  |  |  |  |  |  |  |  |  |  |
| Early | May 30- | 149 | 9 | 2008 | 111 | 99.5 | 20 | 147 | 13.60544218 | 91.16 | 0.9116 | 6 | 6 |
| Early | May | 151 | 11 | 2008 | 106 | 97 | 20 | 136 | 14.70588235 | 63.97 | 0.6397 | 1 | 2 |
|  | 30- |  |  |  |  |  |  |  |  |  |  |  |  |
| Early | May | 151 | 12 | 2008 |  |  | 20 | 149 | 13.42281879 | 44.3 | 0.443 | 4 | 4 |
| Early | Jun | 153 | 15 | 2008 | 105 | 97 | 7 | 147 | 4.761904762 | 93.2 | 0.932 | 2 | 2 |
|  | 1- |  |  |  |  |  |  |  |  |  |  |  |  |
| Early | Jun | 153 | 16 | 2008 | 104 | 99 | 9 | 132 | 6.818181818 | 84.85 | 0.8485 | 3 | 3 |
|  | 2- |  |  |  |  |  |  |  |  |  |  |  |  |
| Early | Jun | 154 | 18 | 2008 | 103 | 95.5 | 12 | 127 | 9.448818898 | 81.1 | 0.811 | 3 | 3 |
|  | 15- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 167 | 38 | 2008 | 94 | 83 | 11 | 101 | 10.89108911 | 88.12 | 0.8812 | 3 | 3 |
|  | 15- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 167 | 39 | 2008 | 99 | 92 | 17 | 148 | 11.48648649 | 80.41 | 0.8041 | 5 | 6 |
|  | 16- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 168 | 41 | 2008 | 96 | 95 | 20 | 133 | 15.03759399 | 90.98 | 0.9098 | 5 | 5 |
|  | 17- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 169 | 43 | 2008 | 106 | 101 | 20 | 126 | 15.87301587 | 86.51 | 0.8651 | 3 | 3 |
|  | 17- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 169 | 45 | 2008 | 95 | 91 | 20 | 117 | 17.09401709 | 84.62 | 0.8462 | 4 | 5 |
|  | 18- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 170 | 46 | 2008 | 91 | 85 | 20 | 90 | 22.22222222 | 85.56 | 0.8556 | 3 | 2 |
|  | 19- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 171 | 49 | 2008 | 102 | 94 | 21 | 108 | 19.44444444 | 84.26 | 0.8426 | 2 | 1 |
|  | 19- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 171 | 50 | 2008 |  |  | 20 | 110 | 18.18181818 | 86.36 | 0.8636 | 2 | 1 |
| Late | 7-Jul | 189 | 86 | 2008 | 96 | 82 | 20 | 94 | 21.27659575 | 90.43 | 0.9043 | 1 | 1 |
| Late | 7-Jul | 189 | 88 | 2008 | 102 | 92.5 | 20 | 93 | 21.50537634 | 65.59 | 0.6559 | 5 | 6 |
| Late | 8-Jul | 190 | 91 | 2008 | 97 | 90 | 20 | 119 | 16.80672269 | 78.99 | 0.7899 | 2 | 2 |
|  | 10- |  |  |  |  |  |  |  |  |  |  |  |  |
| Late | Jul | 192 | 94 | 2008 | 99 | 95 | 20 | 106 | 18.86792453 | 86.79 | 0.8679 | 3 | 4 |


|  | 11- |  |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Late | $\begin{gathered} \text { Jul } \\ 14- \end{gathered}$ | 193 | 96 | 2008 | 108 | 97.5 | 21 | 132 | 15.90909091 | 85.61 | 0.8561 | 3 | 2 |
| Late | Jul | 196 | 97 | 2008 | 89.5 | 79.5 | 20 | 106 | 18.86792453 | 91.51 | 0.9151 | 4 | 3 |
|  | 14- |  |  |  |  |  |  |  |  |  |  |  |  |
| Late | Jul | 196 | 99 | 2008 |  |  | 20 | 113 | 17.69911504 | 61.95 | 0.6195 | 3 | 5 |
| Late | Jul | 196 | 100 | 2008 | 105 | 95.5 | 21 | 105 | 20 | 84.62 | 0.8462 | 1 | 2 |
|  | 16- |  |  |  |  |  |  |  |  |  |  |  |  |
| Late | Jul | 198 | 105 | 2008 | 105 | 99 | 20 | 92 | 21.73913044 | 63.04 | 0.6304 | 1 | 1 |
|  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Early | $\begin{aligned} & \text { May } \\ & 26- \end{aligned}$ | 146 | 5 | 2009 | 108.5 | 93 | 20 | 130 | 15.38461539 | 85.38 | 0.8538 | 7 | 7 |
| Early | May | 146 | 6 | 2009 | 103 | 97 | 20 | 153 | 13.07189543 | 80.39 | 0.8039 | 4 | 4 |
|  | 27- |  |  |  |  |  |  |  |  |  |  |  |  |
| Early | $\begin{gathered} \text { May } \\ 29- \end{gathered}$ | 147 | 8 | 2009 | 106 | 94 | 20 | 148 | 13.51351351 | 88.51 | 0.8851 | 2 | 2 |
| Early | May | 149 | 12 | 2009 | 105 | 99 | 19 | 143 | 13.28671329 | 95.1 | 0.951 | 7 | 7 |
|  | 29- |  |  |  |  |  |  |  |  |  |  |  |  |
| Early | May | 149 | 13 | 2009 | 104 | 97 | 20 | 107 | 18.69158879 | 88.79 | 0.8879 | 4 | 4 |
|  | 31- |  |  |  |  |  |  |  |  |  |  |  |  |
| Early | May | 151 | 17 | 2009 | 98.5 | 94.5 | 20 | 145 | 13.79310345 | 88.97 | 0.8897 | 1 | 1 |
|  | 6- |  |  |  |  |  |  |  |  |  |  |  |  |
| Early | Jun | 157 | 22 | 2009 | 108 | 102 | 20 | 168 | 11.90476191 | 57.14 | 0.5714 | 3 | 3 |
|  | 6- |  |  |  |  |  |  |  |  |  |  |  |  |
| Early | Jun | 157 | 23 | 2009 | 100 | 90.5 | 5 | 116 | 4.310344828 | 76.72 | 0.7672 | 3 | 3 |
|  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Early | $\begin{aligned} & \text { Jun } \\ & 17- \end{aligned}$ | 157 | 24 | 2009 | 96 | 90 | 20 | 110 | 18.18181818 | 93.64 | 0.9364 | 2 | 2 |
| Middle | Jun | 168 | 46 | 2009 | 89 | 84 | 19 | 75 | 25.33333333 | 88.67 | 0.8867 | 2 | 3 |
|  | 18- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 169 | 47 | 2009 | 98 | 94 | 5 | 56 | 8.928571429 | 75 | 0.75 | 2 | 2 |
|  | 18- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 169 | 48 | 2009 | 104 | 93.5 | 20 | 132 | 15.15151515 | 56.82 | 0.5682 | 2 | 2 |
|  | 23- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | $\begin{aligned} & \text { Jun } \\ & 23- \end{aligned}$ | 174 | 55 | 2009 | 94 | 86.5 | 20 | 137 | 14.59854015 | 69.34 | 0.6934 | 1 | 1 |
| Middle | Jun | 174 | 56 | 2009 | 101 | 93 | 20 | 137 | 14.59854015 | 90.51 | 0.9051 | 3 | 3 |



|  | 19- |  |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Middle | Jun | 170 | 61 | 2010 | 88 | 77.5 | 20 | 74 | 27.02702703 | 86.49 | 0.8649 | 3 | 2 |
| Middle | 19- | 170 | 62 | 2010 | 92 | 81 | 20 | 110 | 18.18181818 | 86.36 | 0.8636 | 3 | 3 |
|  | 19- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 170 | 64 | 2010 | 87.5 | 79 | 20 | 95 | 21.05263158 | 77.89 | 0.7789 | 1 | 1 |
|  | 20- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 171 | 65 | 2010 | 89 | 84 | 20 | 91 | 21.97802198 | 82.42 | 0.8242 | 2 | 4 |
|  | 20- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 171 | 66 | 2010 | 95.4 | 90.4 | 18 | 121 | 14.87603306 | 75.21 | 0.7521 | 1 | 1 |
|  | 21- |  |  |  |  |  |  |  |  |  |  |  |  |
| Middle | Jun | 172 | 67 | 2010 | 98 | 88 | 20 | 99 | 20.2020202 | 91.92 | 0.9192 | 3 | 4 |
| Late | 7-Jul | 188 | 114 | 2010 | 104 | 92 | 10 | 117 | 8.547008547 | 80 | 0.8 | 3 | 3 |
|  | 10- |  |  |  |  |  |  |  |  |  |  |  |  |
| Late | Jul | 191 | 116 | 2010 | 105 | 97 | 20 | 123 | 16.2601626 | 81.3 | 0.813 | 3 | 3 |
|  | 10- |  |  |  |  |  |  |  |  |  |  |  |  |
| Late | Jul | 191 | 117 | 2010 | 95 | 89 | 15 | 110 | 13.63636364 | 40.91 | 0.4091 | 1 | 1 |
|  | 12- |  |  |  |  |  |  |  |  |  |  |  |  |
| Late | Jul | 193 | 123 | 2010 | 94 | 81 | 20 | 77 | 25.97402597 | 67.53 | 0.6753 | 3 | 3 |
|  | 15- |  |  |  |  |  |  |  |  |  |  |  |  |
| Late | Jul | 196 | 129 | 2010 | 103 | 94 | 13 | 119 | 10.92436975 | 32.77 | 0.3277 | 1 | 1 |
|  | 16- |  |  |  |  |  |  |  |  |  |  |  |  |
| Late | Jul | 197 | 132 | 2010 | 89 | 82 | 20 | 83 | 24.09638554 | 85.54 | 0.8554 | 2 | 2 |
|  | 16- |  |  |  |  |  |  |  |  |  |  |  |  |
| Late | Jul | 197 | 136 | 2010 | 91 | 79 | 20 | 127 | 15.7480315 | 69.29 | 0.6929 | 4 | 5 |
|  | 17- |  |  |  |  |  |  |  |  |  |  |  |  |
| Late | Jul | 198 | 138 | 2010 | 101 | 91 | 20 | 93 | 21.50537634 | 62.07 | 0.6207 | 3 | 2 |

