VIETNAM JOURNAL OF CHEMISTRY DOI: 10.15625/0866-7144.2015-2e-033 VOL. 53(2e) 142-145

APRIL 2015

SECONDARY METABOLITES FROM MARINE BACTERIUM STREPTOMYCES SP. G039

Vu Thi Quyen¹, Vu Van Nam¹, Tran Van Hieu¹, Le Cong Vinh¹, Doan Thi Mai Huong¹, Le Thi Hong Minh¹, Brian Murphy², Chau Van Minh¹, Pham Van Cuong^{1*}

¹Institute of Marine Biochemistry, VAST, 18 Hoang Quoc Viet, Cau Giay, Hanoi, Vietnam

²University of Illinois at Chicago, Chicago, Illinois, USA

Received 23 January 2015; Accepted for Publication 18 March 2015

Abstract

Nine secondary metabolites, cylo-(Leu-Pro) (1), cyclo-(Pro-Tyr) (2), indole-3-carboxylic acid (3), 3hydroxyacetylindole (4), phenethyl 2-phenyl acetate (5), *n*-butyl–isobutyl phthalate (6), 4-(2-hydroxyethyl) phenol (7), 2-phenylacetic acid (8), and uracil (9) were isolated and identified from marine *Streptomyces sp.* G039. Their structures were determined by spectroscopic analysis including MS and 2D NMR, as well as by comparison with reported data in the literature.

Keywords. *Streptomyces* sp. G039 strain, marine microorganism, cylo-(Leu-Pro), cyclo-(Pro-Tyr), indole-3-carboxylic acid, 3-hydroxyacetylindole.

1. INTRODUCTION

The studies of the secondary substances produced by marine micro-organisms have obtained many significant achievements in the world [1]. Among the secondary metabolites from marine microorganisms, there are many compounds having interesting biological activities that should be useful to development for their pharmaceutical uses. Meanwhile, the search of bioactive secondary metabolites from marine microorganisms is not widely explored in Vietnam [2, 3]. In the course of our screening program, the EtOAc extract of a *Streptomyces sp.* from marine sediment of Ha Long Bay exhibited an inhibition activity against *Staphylococcus aureus, Echerichia coli* and *Fusarium oxysporum.* In this paper, we reported the isolation and structural elucidation of nine secondary metabolites from the cultures broth of *Streptomyces* sp.

2. EXPRIMENTAL

2.1. General procedures

Optical rotations were determined on a JASCO P-2000 polarimeter (Hachioji, Japan). The ESI-MS was measured on Agilent 6120 series single quadrupole LC/MS systems (USA). NMR spectra were recorded on a Bruker AM500 FT-NMR spectrometer using TMS as an internal standard. Column chromatography (CC) was performed using a silica gel (Kiesel gel 60, 70-230 mesh and 230-400 mesh, Merck, Germany). Thin layer chromatography (TLC) used pre-coated silica gel 60 F_{254} (Merck, Germany).

2.2. Actinomycete material

The marine sediment was collected in Ha Long Bay - Quang Ninh in April 2014. The sediment sample (1 g) was added to the 10 mL of sterile sea water in a conical flask. The flask was agitated for about one hour. The marine sediment was filtered and the filtrate was serially diluted to obtain 10^{-1} to 10^{-7} dilutions using the sterilized sea water. An aliquot of 100 µL of each dilution was spread on the media. Different media like Starch Casein Agar (SCA), Glycerol Asparagine Agar (GA Agar), Humic acid-B vitamin agar (HV Agar) and Glucose yeast malt extract agar (GYM) were used for isolation of actinomycetes. The media containing 50% of sterile sea water were supplemented with rifampicin (5 μ g/mL) and nystatin (25 μ g/mL) (Himedia Mumbai) to inhibit bacterial and fungal contamination, respectively. The petriplates were incubated upto 3 weeks at 28 °C. The isolated discrete colonies were observed and used for identification.

2.3. Fermentation, extraction and isolation

Strain G039 (Streptomyces sp.) was cultured in high-nutrient medium (30 g of instant ocean, 10 g of starch, 4 g of yeast, 2 g of peptone, 1 g of calcium carbonate, 40 mg of iron sulfate, and 100 mg of potassium bromate) for 7 days at 25°C while shaking at 200 rpm. The culture solution (30 L) was extracted with EtOAc (15 L \times 3 times). The EtOAc extract solution was concentrated to dryness. The residue (4.03 g) was separated by CC on silica gel, eluted with CH₂Cl₂/MeOH gradient to afford 6 fractions. Fraction 3 (350 mg) was separated by CC on silica gel, eluted with n-hexane/acetone (5 to 50% acetone in *n*-hexane) to give compound **6** (59 mg). Fraction 4 (250 mg) was subjected to CC on Sephadex LH-20 using MeOH followed by preparative TLC to afford compound 5 (10 mg). Fraction 5 (230 mg) was purified by a Sephadex LH-20 CC eluting with MeOH to afford 5 subfractions. Subfraction 1 (65 mg) was recrystallized in CH_2Cl_2 to provide compound 1 (15 mg). Fraction 6 (1.2 g) was subjected to a CC on silica gel, using CH₂Cl₂/MeOH gradient to give 6 subfractions. Subfraction 2 (150 mg) was separated on silica gel CC, eluted with 5 % acetone in methanol, providing compound 7 (8 mg). Subfraction 3 (160 mg) was purified by Sephadex LH-20 CC (MeOH), followed by the preparative TLC to give compound **3** (7 mg). Subfraction 5 (45 mg) was separated by silica gel CC, eluted with CH_2Cl_2 /acetone gradient affording compound 2 (8) mg). Subfraction 6 (84 mg) was recrystallized in CH_2Cl_2 to yield compound 4 (13 mg).

Cylo-(Leu-Pro) (1): White amorphous solid, m.p 147-148 °C, $[\alpha]_D^{25}$ -85 (*c* 0.20, MeOH); ESI-MS: *m/z* 249 [M+K]⁺. ¹H NMR (500 MHz, CD₃OD): 0.98 (3H, d, *J* = 6.5 Hz, CH₃-13), 0.99 (3H, d, *J* = 6.5 Hz, CH₃-12), 1.54 (1H, m, H-10 α), 1.91 (1H, m, H-11), 1.93 (1H, m, H-4 α), 1.98 (1H, m, H-4 β), 2.07 (1H, m, H-10 β), 2.08 (1H, m, H-5 α), 2.34 (1H, m, H-5 β), 3.55 (2H, m, CH₂-3), 4.14 (1H, m, H-9), 4.28 (1H, t, *J* = 7.5 Hz, H-6). ¹³C NMR (125 MHz, CD₃OD): 21.2 (C-13), 22.7 (C-4), 23.3 (C-12), 24.7 (C-11), 28.1 (C-5), 38.7 (C-10), 45.5 (C-3), 53.4 (C-9), 59.0 (C-6), 166.2 (C-1), 170.2 (C-7).

Cyclo-(**Pro-Tyr**) (2): White amorphous solid, m.p 156-158 °C, ESI-MS: m/z 261 $[M+H]^+$. ¹H NMR (500 MHz, CDCl₃): 1.87 (2H, m, CH₂-10), 1.99 (1H, m, H-5b), 2.31 (1H, m, H-5a), 2.79 (1H, dd, J = 9.5, 14.5 Hz, H-10b), 3.44 (1H, dd, J = 9.5, 14.5 Hz, H-10a), 3.53 (1H, m, H-3b), 3.64 (1H, m, H-3a), 4.06 (1H, dd, J = 1.5; 7.5 Hz, H-6), 4.22 (1H, dd, J = 2.5, 9.5 Hz), 6.07 (1H, s, NH), 6.77 (2H, d, J =8.5 Hz, H-3'), 7.03 (2H, d, J = 8.5 Hz, H-2').

Indole-3-carboxylic acid (3): White powder; mp.231-232°C; ESI-MS: m/z162 [M+H]⁺; ¹H NMR (500 MHz, CD₃OD): 7.17 (1H, dd, J = 1.5, 7.0 Hz,

H-5), 7.22 (1H, dd, J = 1.5, 7.0 Hz, H-6), 7.45 (1H, dd, J = 1.0, 7.0 Hz, H-7), 7.96 (1H, s, H-2), 8.08 (1H, dd, J = 1.5, 7.0 Hz, H-4). ¹³C NMR (125 MHz, CD₃OD): 108.6 (C-3a), 112.9 (C-7), 122.0 (C-4), 122.4 (C-5), 123.6 (C-6), 127.5 (C-3a), 133.4 (C-2),138.2 (C-7a), 169.2 (COOH).

3-Hydroxyacetylindole (**4**): Amorphous solid; ¹H NMR (500 MHz, CDCl₃): 8.74 (1H, br. s, NH), 8.28 (1H, m, H-4), 7.92 (1H, d, J = 4.0 Hz), H-2), 7.45 (1H, m, H-7), 7.35 (1H, m, H-5), 7.33 (1H, m, H-6), 4.78 (2H, s, CH₂-9). ¹³C NMR (125 MHz, CDCl₃): 65.4 (C-9), 111.6 (C-7), 114.3 (C-3), 122.0 (C-4), 123.2 (C-5), 124.2 (C-6), 125.3 (C-3a), 130.5 (C-2), 136.2 (C-7a), 193.2 (C-8).

Phenethyl 2-phenyl acetate (5): Yellow oil; ESI-MS: m/z 241 $[M+H]^+$, ¹H NMR (500 MHz, CDCl₃): 2.94 (2H, t, J = 7.0 Hz, CH₂-7'), 3.62 (2H, s, CH₂-7), 4.33 (2H, t, J = 7.0 Hz, CH₂-8'), 7.16-7.35 (10H, m, 2Ph-H). ¹³C NMR (125 MHz, CDCl₃): 35.1 (C-7'), 41.4 (C-7), 65.3 (C-8'), 126.5-129.3 (Ph-CH), 133.9 (C-1), 137.7 (C-1'), 171.4 (C-8).

n-Butyl–isobutyl phthalate (6): ¹H NMR (500 MHz, CDCl₃): 0.96 (3H, t, J = 7.0 Hz, CH₃-4'), 0.98 (6H, d, J = 6.5 Hz, CH₃-3'' and CH₃-4''), 1.44 (2H, sext, J = 7.0 Hz, CH₂-3'), 1.72 (2H, quint, J = 7.0 Hz, CH₂-2'), 2.04 (1H, m, J = 6.5 Hz, H-2''), 4.09 (2H, d, J = 7.0 Hz, CH₂-1''), 4.30 (2H, t, J = 7.0 Hz, CH₂-1'), 7.52 (2H, m, H-4 and H-5), 7.72 (2H, m, H-3 and H-6). ¹³C NMR (125 MHz, CDCl₃): 13.7 (C-4'), 19.2 (C-3'' and C-4''), 27.7 (C-2''), 30.6 (C-2'), 65.6 (C-1'), 71.8 (C-1''), 128.8 (C-4 and C-5), 130.9 (C-3 and C-6), 132.3 (C-1), 132.4 (C-2), 167.6 (C-7), 167.7 (C-8).

4-(2-Hydroxyethyl) phenol (7): White powder, m.p. 91-92 °C, ESI-MS: m/z 139 $[M+H]^+$; ¹H NMR (500 MHz, CDCl₃): 2.73 (2H, t, J = 7 Hz, H-7), 3.71 (2H, t, J = 7.0 Hz, H-8), 6,72 (2H, d, J = 8.5 Hz; H-2+ H-6), 7.05 (2H, d, J = 8.5 Hz, H-3 and H-5).

2-Phenylacetic acid (8): white needle crystal, m.p. 76-77 °C; ESI-MS: m/z 138 [M+H]⁺; ¹H NMR (500 MHz, CDCl₃): 3.59 (2H, s, CH₂), 7.25-7.33 (5H, m, Ph-H). ¹³C NMR (125 MHz, CDCl₃): 41.4 (CH₂), 127.2 (C-4), 128.6 (C-2 and C-6), 129.4 (C-3 and C-5), 123.6 (C-6), 133.8 (C-1), 175.2 (COOH).

Uracil (9): White powder, mp. 320-321 °C; ESI-MS: m/z 111 [M+H]⁺; ¹H NMR (500 MHz, DMSO d_6): 5.45 (1H, d, J = 7.5 Hz, H-5); 7.38 (1H, d, J =7.5 Hz, H-6). ¹³C NMR (125 MHz, DMSO- d_6): 100.3 (C-5), 142.3 (C-6), 151.62 (C-2), 164.5 (C-4).

3. RESULTS AND DISCUSSION

Compound 1 was isolated as white amorphous

Secondary metabolites from marine...

solid and optically active, $\left[\alpha\right]_{D}^{25}$ -85 (c 0.20, MeOH). The ESI-MS indicated the pseudomolecular ion peak at m/z 249 [M+K]⁺. The ¹H NMR spectrum of 1 displayed signals of 2 methyl groups as doublet of doublet at $\delta_{\rm H}$ 0.98 (d, J = 6.5 Hz, CH₃-13), 0.99 (d, J = 6.5 Hz, CH₃-12) and signals of ten aliphatic protons ranging from 1.54 to 4.28 ppm. Analysis of the ${}^{13}C$ NMR and DEPT spectra of 1 revealed the presence of 11 carbons, including two methyl groups at $\delta_C 21.2$ (C-13) and 23.3 (C-12), three methines at $\delta_{\rm C}$ 24.7 (C-11), 53.4 (C-9) and 59.0 (C-6), four methylenes at $\delta_{\rm C}$ 22.7 (C-4), 28.1 (C-5), 38.7 (C-10), 45.5 (C-3), and two carbonyl at $\delta_{\rm C}$ 166.2 (C-1) and 170.2 (C-7). The chemical shifts of CH₂-3, CH-6 and CH-9 suggested their linkage to nitrogen atoms. This data suggested the presence of two amino acid units, proline and leucine in the structure of 1. Detailed analysis of NMR spectra and comparison with reported values in the literature [4, 5], the structure of **1** was determined to be cylo-(Leu-Pro). This cylodipeptide was previously described from *Michelia champaca* and *Ulva pertusa* [4, 5].

Compound **2** was isolated as a white amorphous solid. The ESI-MS indicated the pseudo-molecular ion peak at m/z 261 [M+H]⁺. The 1D NMR spectrum of **2** displayed signals of the proline unit as **1**. However, in comparison with **1**, the presence of an A₂B₂ aromatic system [$\delta_{\rm H}$ 6.77 (2H, d, J = 8.5 Hz, H-3') and 7.03 (2H, d, J = 8.5 Hz, H-2')] instead of

signals of the 2-propyl group was noted for 2. This observation strongly suggested that the leucine unit of 1 was replaced by the 4-hydroxyphenylanaline moiety in 2. Comparison with the literature [5], compound 2 was identified as cyclo-(Pro-Tyr). This cyclodipeptide was previously isolated from **Streptomyces** and Colletotrichum sp. gloeosporioides [6, 7]. It was found that this compound had antibacterial activity against both Gram-positive and Gram-negative bacteria and antifungal property [7].

Compound **3** was obtained as white powder. The ESI-MS indicated the pseudo-molecular ion peak at m/z 162 [M+H]⁺. Its ¹H-NMR spectrum of **3** showed signals of 5 aromatic protons at $\delta_{\rm H}$ 7.17 (dd, J = 1.5, 7.0 Hz, H-5), 7.22 (dd, J = 1.5, 7.0 Hz, H-6), 7.45 (dd, J = 1.0, 7.0 Hz, H-7), 7.96 (s, H-2) and 8.08(dd, J = 1.5, 7.0 Hz, H-4). In the ¹³C-NMR and DEPT spectra of 3, signals of nine carbons were noted, including five aromatic methine groups, a carboxylic group ($\delta_{\rm C}$ 169.2, C-8) and three sp² quaternary carbons at δ_{C} 108.6 (C-3), 127.5 (C-3a) and 138.2 (C-7a). Thus, the presence of an 1,2disubstituted benzene ring was defined. Carefully analysis of the NMR spectra and comparison with reported data [8], compound 3 was identified as indole-3-carboxylic acid which was reported to have antibacterial activity [8].

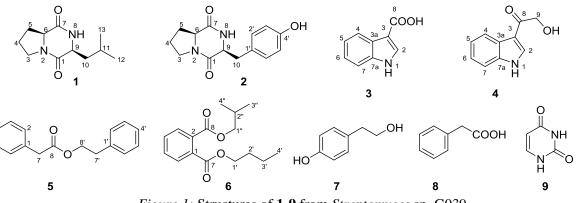


Figure 1: Structures of 1-9 from Streptomyces sp. G039

Compound **4** was isolated as white solids. The 1D NMR spectra of **4** displayed the signals close to those of **3** in which four aromatic protons of 1,2-disubstituted benzene ring [$\delta_{\rm H}$ 8.28 (m, H-4), 7.45 (m, H-7), 7.33 (m, H-5) and 7.35 (m, H-6)] and a proton at $\delta_{\rm H}$ 7.92 (J = 4.0 Hz, H-2) were observed. Comparison of ¹³C NMR and DEPT spectra of **3** and **4** indicated that the carboxylic carbon signal of **3** was replaced by signals of a ketone at $\delta_{\rm C}$ 193.2 and an oxymethylene at $\delta_{\rm C}$ 65.4. Analysis of 2D NMR spectra, especially HMBC spectrum established the

structure of **4** as shown in figure 1. This compound was reported from several marine bacteria and found to be active against parasite *Trypanosoma cruzi* [9].

Compound **5** was obtained as yellow oil. The ESI-MS indicated the pseudo-molecular ion peak at m/z 241 [M+H]⁺. The ¹H NMR spectrum of **5** displayed signals for 10 aromatic protons at $\delta_{\rm H}$ 7.16-7.35 and three methylenes at $\delta_{\rm H}$ 2.94 (t, J = 7.0 Hz, CH₂-7'), 3.62 (s, CH₂-7), 4.33 (t, J = 7.0 Hz, CH₂-8'). Analysis of the ¹³C NMR and DEPT spectra revealed the presence of the groups observed in the

¹H-NMR spectrum (two phenyl rings and three methylenes), with additional signals of a carboxylate carbon at δ_C 171.4 (C-8) and two aromatic quaternary carbons at $\delta_{\rm C}$ 137.7 (C-1') and 133.9 (C-1). Analysis of the HMBC spectrum of 5 allowed determining the structure of **5** as phenethyl-2-phenyl acetate [10].

ESI-MS spectrum of 6 presented the protonated molecular ion at m/z 279 [M+H]⁺. The ¹H-NMR spectrum showed the signals of four aromatic protons and eighteen protons in the aliphatic region. Its ¹³C NMR and DEPT spectra indicated signals of two carboxylate carbons at $\delta_{\rm C}$ 167.6 and 167.7 (C-7 and C-8), six aromatic carbons, a sp³ methine, four methyenes and three methyls. The chemical shifts of the two methylenes at δ_C 65.6 and 71.8 suggested their linkage to oxygens. The NMR data of 6 were consistent with those of *n*-butyl-isobutyl phthalate [11, 12].

By analysis of the NMR spectra, compounds 7, 8 and 9 were determined to be uracil [3, 13], 4-(2hydroxyethyl) phenol [14] and 2-phenylacetic acid [15], respectively. Their NMR data were identical with those reported in the literature.

Acknowledgements. This work was supported by the Vietnam Academy of Science and Technology (VAST) and the Ministry of Science and Technology of Vietnam (MOST), Project: VAST. TĐ. *ÐAB.04/13-15*.

REFERENCES

- 1. A. Debbab, A. H. Aly, W. H. Lin, P. Proksch. Bioactive compounds from marine bacteria and fungi, Microb. Biotechnol., 3(5), 544-563 (2010).
- 2. V. M. Chau, V. K. Phan, V. H. Nguyen, H. N. Nguyen, V. C. Pham. Dược liệu biển Việt Nam: Thực trạng và Cơ hội phát triển, Nxb. Khoa học tự nhiên và Công nghệ, Hà Nội, 2012, tr 3.
- 3. T. T. Do, D. Q. Le, D. T. Quyen, V. C. Pham. Isolation and identification of marine bacteria from marine mud in Vietnam with antimicrobial activity, Journal of Vietnamese Environment, 3(2), 71-75 (2012).
- 4. W. Chung-Yun, H. Lei, K. Kai, S. Chang-Lun, W. Yu-Xi, Z. Cai-Juan, G. Hua-Shi. Secondary metabolites from green algae Ulva pertusa, Chem. Nat. Compd., 46(5), 828-830 (2010).

Corresponding author: Pham Van Cuong

Environ. Microbiol., 3739-3745 (2001).

Institute of Marine Biochemistry, Vietnam Academy of Science and Technology

- M. C. Vanessa, L. Z. Maria, H. L. Loanis, H. S. 5. Geraldo, S. B. Vanderlan, M. Y. Maria, H. P. Ludwig, R. A. Angela. Compounds produced by Colletotrichum gloeosporioides, an Endophytic Fungus from Michelia champaca., Molecules, 19,
- 19243-19252 (2014). 6. S. Smaoui, F. Mathieu, L. Elleuch, Y. Coppe, G. Merlina, I. Karray- Rebai, L. Melloul. Taxonomy purification and chemical characterization of four bioactive compounds from new Streptomyces sp. TN256 strain, World J. Microbiol. Biotechnol., 28, 793-804, (2011).
- 7. P. Wang, F. Kong, J. Wei, Y. Wang, W. Wang, K. Hong, W. Zhu. Alkaloids from the mangrove-derived Actinomycete Jishengella endophytica 161111, Mar. Drugs, 12, 477-490 (2014).
- 8. T. Shouzheng, Y. Yabin; L. Kai, X. Zijun, X. Lihua, Z. Lixing. Antimicrobial metabolites from a novel halophilic actinomycete Nocardiopsis terrae YIM 90022, Nat. Prod. Res., 28(5), 344-346 (2014).
- 9. S. Martínez-Luis, J. F. Gómez, C. Spadafora, H. M. Guzmán, M. Gutiérrez. Antitrypanosomal alkaloids from the marine bacterium Bacillus pumilus, Molecules, 17, 11146-11155 (2012).
- 10. S. I. Murahashi, T. Naota, K. Ito, Y. Maeda, H. Taki. Ruthenium-catalyzed oxidative transformation of alcohols and aldehydes to esters and lactones, J. Org. Chem., 52, 4319-4327 (1987).
- 11. N. S. Radulović, P. D. Blagojević. The most frequently encountered volatile contaminants of essential oils and plant extracts introduced during the isolation procedure: Fast and easy profiling, Phytochem. Anal., 23, 131-142 (2012).
- 12. M. Liu, W. Zhang, L. Qiu, X. Lin. Synthesis of butyl-isobutyl-phthalate and its interaction with α glucosidase in vitro, J. Biochem., 149(1), 27-33 (2011).
- 13. V. N. Vu, V. Trinh Thi Thanh, V. C. Pham, H. Doan Thi Mai, V. M. Chau. Isolation and structural elucidation of compounds from cultures broth of marine bacterium Photobacterium sp., Journal of Science and Technology (Vietnamese), 52(5B), 611-616 (2014).
- 14. V. Somepalli, K. P. Gopala, V. S. Gottumakkala. Synthesis and biological activities of prenylated tyrosine derivatives, the metabolites pithomycesellis, Indian Journal of Chemistry, 45B, 1063-1066 (2006).
- 15. K. H. Byung, W. L. Song, S. K. Beom, Y. L. Jung, S. M. Surk. Isolation and in vivo and in vitro antifungal activity of phenylacetic acid and sodium phenylacetate from Streptomyces humidus, Appl.

18 Hoang Quoc Viet, Cau Giay, Hanoi, Vietnam

E-mail: phamvc@imbc.vast.vn.