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Merosesquiterpenes from marine sponge Smenospongia cerebriformis

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Abstract

Using various chromatography methods, three merosesquiterpenes belonging to sesquiterpene quinone type, neodactyloquinone (1), dactyloquinone D (2), and dactyloquinone C (3) together with two indole derivatives indole-3-aldehyde (4) and indole-3-cacboxylic methyl ester (5) were isolated from the methanol extract of the Vietnamese marine sponge *Smenospongia cerebriformis*. Their structures were determined by 1D-, 2D-NMR spectra, HR-ESI-MS and in comparison with those reported in the literature.

Keywords. Smenospongia cerebriformis, merosesquiterpene, sesquiterpene quinone, indole derivative.

1. INTRODUCTION

Marine sponges are regarded as a rich source of secondary metabolites with chemically diverse and potential biological benefits. Merosesquiterpenes and indole alkaloid derivatives were found to be the main components of sponges, particularly, the genus Smenospongia. A huge variety of compounds belonging to these two chemical structure classes have been reported from sponges and possessed a broad range of interest bioactivities, such as antimalarial [1, 2], antimicrobial [1-3], anticancer [3, 4] antidepressant [5], as well as inhibition of the neuronal isozyme of nitric oxide synthase (nNOS) [6]. Herein, we report the isolation and structure determination of three merosesquiterpenes and two indole alkaloid derivatives from sponge Smenospongia cerebriformis.

2. MATERIALS AND METHODS

2.1. Sponge materials

The sponge *Smenospongia cerebriformis* (Duchassaing & Michelotti, 1864) was collected in Vinhmoc, Quangtri in August 2015 and identified by Prof. Do Cong Thung, Institute of Marine Environment and Resources, VAST. A voucher specimen (HM08.2015-2) was deposited at the Institute of Marine Biochemistry, Vietnam Academy of Science and Technology.

2.2. General experimental procedures

The ¹H-NMR (500 MHz) and ¹³C-NMR (125 MHz) spectra were recorded on a Bruker AM500 FT-NMR spectrometer and TMS was used as an internal standard. Column chromatography was performed using a silica gel (Kieselgel 60, 70–230 mesh and 230-400 mesh, Merck, Whitehouse Station, NJ) or RP-18 resins (30-50 μm, Fuji Silysia Chemical Ltd.), and thin layer chromatography (TLC) using pre-coated silica-gel 60 F₂₅₄ (0.25 mm, Merck) and RP-18 F_{254S} plates (0.25 mm, Merck).

2.3. Extraction and isolation

Fresh frozen dried samples of Smenospongia cerebriformis (15.0 kg) were well grinded and sonicated with hot MeOH three times and then concentrated under reduced pressure to give MeOH extract (SP, 360 g). This extract was suspended in water and then partitioned with CH₂Cl₂ to give the CH₂Cl₂ (SPD, 102 g) and water (SPW, 250 g) extracts after removal of the solvents in vacuo. Fraction SPD (100 g) was subjected to silica gel column chromatography and eluted with an n-hexane - acetone stepwise gradient to give five fractions SPD1 (39.0 g), SPD2 (5.8 g), SPD3 (12.9 g), SPD4 g), and SPD5 (2.8 g). SPD2 chromatographed on a RP-18 column eluting with acetone - water (1.5:1, v/v) to give four smaller fractions SPD2A-D. Fraction SPD2B was subjected VJC, 55(2), 2017 Phan Van Kiem et al.

to silica gel column chromatography and eluted with a *n*-hexane-ethyl acetate (1.5:1, v/v) to give compound **4** (ASP2, 11.0 mg). Fraction SPD3 was chromatographed on a silica gel column eluting with *n*-hexane-ethyl acetate (3:1, v/v) to give five smaller fractions, SPD3A-E. Fraction SPD3D (2.2 g) was applied to a silica gel column eluting with *n*-hexane-ethyl acetate (2:1, v/v) to give compounds **1** (ASP16A, 10.0 mg) and **2** (ASP15A, 12.0 mg). Fraction SPD3E (1.8 g) was chromatographed on a

RP-18 column eluting with acetone - water (1:1, v/v) to yield compound **5** (16 mg). Fraction SPD5 was subjected to a silica gel column using dichloromethane-ethyl acetate (10:1, v/v) as eluent to give five smaller fractions, SPD5A-D. Furthermore, fraction SPD5B (0.4 g) was firstly chromatographed on a RP-18 column eluting with acetone-water (2:1, v/v) and then further purified on a silica gel column eluting with dichloromethane - acetone (12:1, v/v) to yield compound **3** (ASP27, 11.0 mg).

Figure 1: Chemical structures of compounds 1-5 from S. cerebriformis

Neodactyloquinone (1): White amorphous powder; α_D^{25} : +25.4 (c = 0.1, in CDCl₃); ¹H- and ¹³C-NMR (CDCl₃), see table 1.

Dactyloquinone C (2): White amorphous powder; α_D^{25} : +30.2 (c = 0.1, in CDCl₃); ¹H- and ¹³C-NMR (CDCl₃), see table 2.

Dactyloquinone D (3): White amorphous powder; α_D^{25} : +21.6 (c = 0.1, in CDCl₃); ¹H-NMR (CDCl₃, 500 MHz) δ_H: 5.76 (s, H-19), 4.52 and 4.49 (each 1H, br s, H-11), 3.80 (3H, s, 20-OMe), 1.24 (3H, s, H-13), 1.11 (3H, s, H-12), 1.07 (3H, s, H-14); ¹³C-NMR (CDCl₃, 125 MHz) δ_C: 22.0 (C-1), 28.3 (C-2), 32.6 (C-3), 158.6 (C-4), 39.4 (C-5), 31.2 (C-6), 30.4 (C-7), 84.8 (C-8), 37.1 (C-9), 44.9 (C-10), 103.5 (C-11), 20.9 (C-12), 23.1 (C-13), 21.1 (C-14), 27.8 (C-15), 113.9 (C-16), 152.6 (C-17), 181.7 (C-18), 104.9 (C-19), 159.4 (C-20), 181.0 (C-21), and 56.4 (20-OMe).

Indole-3-aldehyde (**4**): White amorphous powder; 1 H-NMR (CD₃OD, 500 MHz) δ_{H} : 8.04 (s, H-2), 8.13 (d, J=7.5 Hz, H-4), 7.20 (dd, J=7.5, 8.0 Hz, H-5), 7.24 (dd, J=7.5, 8.0 Hz, H-6), 7.45 (d, J=7.5 Hz, H-7), 9.84 (s, H-8); 13 C-NMR (CD₃OD, 125 MHz) δ_{C} : 139.7 (C-2), 120.1 (C-3), 125.7 (C-3a), 122.4 (C-4), 123.6 (C-5), 125.0 (C-6), 113.1 (C-7), 138.9 (C-7a), 187.4 (C-8).

Indole-3-carboxylic methyl ester (5): White amorphous powder, 1 H-NMR (CD₃OD, 500 MHz) δ_{H} : 7.93 (s, H-2), 8.20 (dd, J = 3.0, 9.0 Hz, H-4), 7.28 (overlapped signals, H-5 and H-6), 7.42 (dd, J = 3.0, 9.0 Hz, H-7), 3.93 (s, 8-OMe); 13 C-NMR (CD₃OD, 125 Hz) δ_{C} : 133.1 (C-2), 131.0 (C-3), 136.1 (C-3a), 121.6 (C-4), 122.1 (C-5), 123.2 (C-6), 111.5 (C-7), 125.8 (C-7a), 165.6 (C-8), 51.1 (8-OMe).

3. RESULTS AND DISCUSSION

Compound 1 was isolated as a white amorphous powder. It had a molecular formula C₂₂H₂₈O₄ which was derived from a pseudo-molecular [M+H]⁺ ion peak at m/z 357.2039 (calcd. for $C_{22}H_{29}O_4$, 357.1988) in the HR-ESI-MS and in conjunction with ¹³C NMR data. ¹H NMR and HSQC spectroscopic analysis of 1 showed the presence of three tertiary methyl groups at $\delta_{\rm H}$ 0.93, 1.08 and 1.41 (each 3H, s), exocyclic methylene signal at δ_H 4.54 (2H, br s), methoxy group at δ_H 3.80 (3H, s) and an olefinic proton at $\delta_{\rm H}$ 5.74 (1H, s). The 13 C NMR of 1 revealed signals of 22 carbons which were classified by DEPT as nine non-protonated carbons, two methines, seven methylenes, and four methyl carbons. ¹H NMR and ¹³C NMR disclosed the presence of a dialkoxy-1,4-benzoquinone moiety

Table 1: NMR spectral data for 1-2 and reference compounds

	2 # 2			$\delta_{\rm C}^{~\$,a}$	a l		
С	$\delta_{C}^{\ \#,a}$	- 1-	1		- 1-	2	
		$\delta_{ m C}^{~a,b}$	$\delta_{\rm H}^{\rm a,c}$ (mult., J in Hz)		$\delta_{ m C}^{~a,b}$	$\delta_{\rm H}^{\rm a,c}$ (mult., J in Hz)	
1	21.2	21.2	1.56 (m)	78.5	78.5	4.15 (ddd, 5.5, 10.5, 10.5)	
			1.77 (m)				
2	27.7	27.7	1.27 (m)	35.5	35.5	1.80 (m)	
			1.83 (m)			2.52 (m)	
3	32.7	32.7	2.10 (br d, 14.0)	30.6	30.7	2.26 (m)	
			2.23 (ddd, 5.5, 14.0, 14.0)			2.40 (ddd, 5.0, 14.0, 14.0)	
4	158.3	158.3	-	157.2	157.2	-	
5	41.3	41.4	-	40.2	40.3	-	
6	30.8	30.9	1.41 (m)	37.4	37.4	1.64 (m)	
			2.00 (ddd, 3.5, 3.5, 14.0)				
7	32.3	32.4	1.58 (m)	27.8	27.8	1.53 (m)	
			1.78 (br d, 14.0)			1.60 (m)	
8	34.6	34.6	-	41.8	41.8	1.39 (m)	
9	86.4	86.4	-	37.4	37.4	-	
10	47.8	47.8	1.44 (dd, 2.5, 12.0)	60.2	60.2	1.50 (d, 10.5)	
11	103.7	103.7	4.54 (br s)	104.1	104.1	4.62 (br s)	
12	20.8	20.9	1.08 (s)	21.7	21.8	0.96 (s)	
13	24.3	24.3	0.93 (s)	16.4	16.4	1.03 (d, 6.5)	
14	19.1	19.1	1.41 (s)	15.0	15.0	0.73 (s)	
15	26.8	26.8	2.04 (d, 16.5)	34.8	34.8	1.99 (d, 14.0)	
			2.72 (d, 16.5)			3.13 (d, 14.0)	
16	114.6	114.5	-	130.2	130.3	-	
17	151.2	151.3	-	156.5	156.3	-	
18	181.5	181.5	-	182.6	183.0	1	
19	104.7	104.7	5.74 (s)	105.2	105.2	5.80 (s)	
20	159.5	159.5	-	159.0	159.0	1	
21	181.5	181.5	-	182.6	182.6	1	
20-	56.3	56.4	3.80 (s)	56.4	56.4	3.81 (s)	
OMe							

Measured in ^{a)}CDCl₃, ^{b)}125 MHz, ^{c)}500 MHz. ^{#)} $\delta_{\rm C}$ of neodactyloquinone [8], ^{\$)} $\delta_{\rm C}$ of dactyloquinone C [9].

 $(\delta_{H}: 3.80, 5.74; \delta_{C}: 56.4, 104.7, 114.5, 151.3, 159.5,$ 181.5, 181.5) [7]. The HMBC correlations between H-11 (δ_H 4.54) and C-3 (δ_C 32.7)/C-4 (δ_C 158.3)/C-5 $(\delta_{\rm C} 41.4)$ suggested an exocyclic olefinic methylene forming at C-11/C-4. Methyl protons H-12 ($\delta_{\rm H}$ 1.08) have HMBC correlations with C-4/C-5/C-6 (δ_{C} 30.9)/C-10 (δ_C 47.8), indicating the location of a methyl group at C-5. The HMBC correlations between H-14 (δ_H 1.41) and C-8 (δ_C 34.6)/C-9 (δ_C 86.4)/C-10 indicated a methyl group at C-9. The last tertiary methyl group located at C-8 which was indicated by HMBC correlations between proton H-13 (δ_H 0.93) and carbons C-7 (δ_C 32.4)/C-8/C-9/C-15 ($\delta_{\rm C}$ 26.8). The 1,4-benzoquinone moiety linked to sesquiterpene skeleton at C-15 confirmed by HBMC correlations between methylene protons H-15 ($\delta_{\rm H}$ 2.04, 2.72) and carbons C-7/C-8/C-9/C-13 ($\delta_{\rm C}$ 24.3)/C-16 ($\delta_{\rm C}$ 114.5)/C-17 ($\delta_{\rm C}$ 151.3)/C-21 ($\delta_{\rm C}$ 181.5). The HMBC correlations from protons H-19 $(\delta_{\rm H} 5.74)$ and methoxy $(\delta_{\rm H} 3.80)$ to C-20 $(\delta_{\rm C} 159.5)$ demonstrated for a methoxy group at C-20. Furthermore, carbon chemical shifts of C-9 ($\delta_{\rm C}$ 86.4) and C-17 (δ_C 151.3) suggested an ether bridge between C-9 and C-17 which was agreed with molecular formula of 1 C₂₂H₂₈O₄. Consequently, structure of 1 was established to be neodactyloguinone, a sesquiterpene previously isolated from the sponge Dactylospongia elegans [8]. Its ¹H and ¹³C NMR data were identical with those reported in the literature (table 1) and found to match well [8].

Compound **2** was obtained as a white amorphous powder. The molecular formula of **2** was established as $C_{22}H_{28}O_4$ on the basis of HR-ESI-MS (m/z: 357.2074, [M+H]⁺; calcd. for $C_{22}H_{29}O_4$, 357.1988) and 13 C-NMR analysis. 1 H NMR and HSQC spectroscopic analysis of **2** showed the presence of two tertiary methyl groups at δ_H 0.73, 0.96 (each 3H,

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s) and a secondary methyl group at δ_H 1.03 (3H, d, J = 6.5 Hz), exocyclic methylene signal at $\delta_{\rm H}$ 4.62 (2H, br s), oxygenated methine signal at $\delta_{\rm H}$ 4.15 (1H, ddd, J = 5.5, 10.5, 10.5 Hz), methoxy group at $\delta_{\rm H}$ 3.81 (3H, s) and an olefinic proton at $\delta_{\rm H}$ 5.80 (1H, s). The ¹³C NMR of 2 revealed signals of 22 carbons which were classified by DEPT as eight nonthree protonated carbons, methines, methylenes, and four methyl carbons. ¹H NMR and ¹³C NMR disclosed the presence of a dialkoxy-1,4benzoguinone moiety [δ_H : 3.81, 5.80; δ_C : 56.4, 105.2, 130.3, 156.3, 159.0, 182.6, 183.0] [9]. The HMBC correlations between protons H-11 ($\delta_{\rm H}$ 4.62) and C-3 (δ_C 30.7)/C-4 (δ_C 157.2)/C-5 (δ_C 40.3) suggested an exocyclic olefinic methylene forming at C-11/C-4. Methyl protons H-12 (δ_H 0.96) have HMBC correlations with C-4/C-5/C-6 (δ_C 37.4)/C-10 (δ_C 60.2), indicating the location of a methyl group at C-5. The HMBC correlations between proton H-14 (δ_H 0.73) and C-8 (δ_C 34.6)/C-9 (δ_C 37.4)/C-10/ C-15 (δ_C 34.8) confirmed the methyl group at C-9. The last secondary methyl group located at C-8 which was indicated by HMBC correlations between H-13 (δ_H 1.03) and C-7 (δ_C 27.8)/C-8/C-9. The 1,4-benzoguinone moiety linked to sesquiterpene skeleton at C-15 confirmed by HBMC correlations between methylene protons H-15 ($\delta_{\rm H}$ 1.99, 3.13) and C-9/C-10/C-14 ($\delta_{\rm C}$ 15.0)/C-16 ($\delta_{\rm C}$ 130.3)/C-17 ($\delta_{\rm C}$ 156.3)/C-21 ($\delta_{\rm C}$ 182.6). The HMBC correlations from protons H-19 (δ_H 5.80) and methoxy (δ_H 3.81) to C-20 (δ_C 159.0) demonstrated for a methoxy group at C-20. Carbon chemical shifts of C-1 (δ_C 78.5) and C-17 (δ_C 156.3) suggested an ether bridge between C-1 and C-17 which was agreed with molecular formula of 2 C₂₂H₂₈O₄. In addition, the ¹H- and ¹³C-NMR data of 2 were identical with those of dactyloquinone C, a compound also isolated from sponge Dactylospongia elegans [9] (table 1) and found to match. Consequently, the structure of 2 was established.

Compound 3 was isolated as a white amorphous powder. The molecular formula C₂₂H₂₈O₄ was deduced on the basis of HR-ESI-MS (m/z: 357.2039, $[M+H]^+$; calcd. for $C_{22}H_{29}O_4$, 357.1988) and ¹³C-NMR analysis. ¹H NMR data of 3 also showed the presence of three tertiary methyl groups at $\delta_{\rm H}$ 1.07, 1.11 and 1.23 (each 3H, s), exocyclic methylene signal at δ_H 4.49 and 4.52 (each 1H, br s), methoxy group at δ_H 3.80 (3H, s), and an aromatic proton at $\delta_{\rm H}$ 5.76 (1H, s). The ¹³C NMR and DEPT spectra of 3 revealed signals of 22 carbons including nine nonprotonated carbons, two methines, methylenes, and four methyl carbons. ¹H NMR and ¹³C NMR indicated the presence of a dialkoxy-1,4benzoquinone moiety $[\delta_H$: 3.80, 5.76; δ_C : 56.4, 103.9, 114.5, 152.6, 159.4, 181.0, 181.7]. In comparison with **2**, the 1D-NMR spectra of **3** showed the presence signals of an oxygenated tertiary carbon (84.8), a saturated methylene group, and a tertiary methyl group instead of an oxygenated secondary carbon (δ_C 78.5), a saturated methine group, and a secondary methyl group in the 1D-NMR of **2**, suggesting for the re-arrangement of ether bridge from C-1/C-17 in **2** to C-8/C-17 in **3**. Thus, compound **3** was determined to be dactyloquinone D, a known compound isolated from the sponge *Dactylospongia elegans* [9].

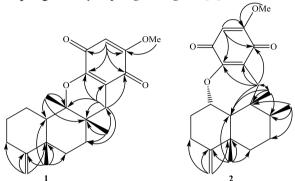


Figure 2: The key HMBC correlations of 1 and 2

The remaining compounds were elucidated to be indole-3-aldehyde (4) and indole-3-cacboxylic methyl ester (5). Their structures were established based on spectral and chemical evidence, which agreed with previous studies [10, 11].

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REFERENCES

- P. Djura, D. B. Stierle, B. Sullivan, D. J. Faulkner, E. V. Arnold, J. Clardy. Some metabolites of the marine sponges Smenospongia aurea and Smenospongia (.ident.Polyfibrospongia) echina, Journal of Organic Chemistry, 45, 1435-1441 (1980).
- 2. Jin-Feng Hu, John A. Schetz, Michelle Kelly, Jiang-Nan Peng, Kenny K. H. Ang, Horst Flotow, Chung Yan Leong, Siew Bee Ng, Antony D. Buss, Scott P. Wilkins, a. M. T. Hamann. New antiinfective and human 5-HT2 receptor binding natural and semisynthetic compounds from the Jamaican sponge Smenospongia aurea, Journal of Natural Products, 65, 476-480 (2002).
- 3. M. L. Kondracki, M. Guyot. *Biologically active quinone and hydroquinone sesquiterpenoids from the sponge Smenospongia* sp., Tetrahedron, **45**, 1995-2004 (1989).

- 4. M. -L. Kondracki, A. M. Guyot. *Smenospongine: a cytotoxic ang antimicrobial aminoquinone isolaed from Smenospongia* sp., Tetrahedron Letters, **28**, 5815-5818 (1987).
- Anna J. Kochanowska, Karumanchi V. Rao, Suzanne Childress, Abir El-Alfy, Rae R. Matsumoto, Michelle Kelly, Gina S. Stewart, Kenneth J. Sufka, a. M. T. Hamann. Secondary metabolites from three Florida sponges with antidepressant activity, Journal of Natural Products, 71, 186-189 (2008).
- 6. E. M. Boyd, J. Sperry. Synthesis of the selective neuronal nitric oxide synthase (nNOS) inhibitor 5,6-dibromo-2'-demethylaplysinopsin, Synlett, 826-830 (2011).
- 7. Hidemichi Mitome, Takahiro Nagasawa, Hiroaki Miyaoka, Yasuji Yamada, a. R. W. M. v. Soest. A new sesquiterpenoid quinone and other related compounds from the Okinawan marine sponge Dactylospongia elegans, Journal of Natural Products,

- **66**, 46-50 (2003).
- 8. C. Shugeng, G. Zhijie, J. T. Shannon, M. H. Sidney, S. L. John, G. I. K. David. *Marine Sesquiterpenoids that Inhibit the Lyase Activity of DNA Polymerase b*, Journal of Natural Products, **67**, 1716-1718 (2004).
- 9. Hidemichi mitome, Takahiro Nagasawa, Hiroaki Miyaoka, Jasuji Yamada, a. R. W. M. v. Soest. Dactyloquinones C, D and E novel sesquiterpenoid quinones, from the Okinawan marine sponge, Dactylospongia elegans, Tetrahedron, 58, 1693-1696 (2002).
- 10. M. A. Ashour, E. S. Elkhayat, R. E. R. Ebel, P. Proksch. *Indole alkaloid from the red sea sponge Hyrtios erectus*, Arkivoc, **xv**, 225-231 (2007).
- 11. Qing-Qing Yang, Marianna Marchini, Wen-Jing Xiao, Paola Ceroni, a. M. Bandini. *Visible-light-induced direct photocatalytic carboxylation of indoles with CBr*₄/MeOH, Chemistry A European Journal, **21**, 18052-18056 (2015).

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