

Journal of Kidney Cancer and VHL 2015; 2(4):163-173.
DOI: <http://dx.doi.org/10.15586/jkcvhl.2015.41>

Review Article

Implications of Von Hippel-Lindau Syndrome and Renal Cell Carcinoma

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Abstract

Von Hippel-Lindau syndrome (VHLS) is a rare hereditary neoplastic disorder caused by mutations in the *vhl* gene leading to the development of tumors in several organs including the central nervous system, pancreas, kidneys, and reproductive organs. Manifestations of VHLS can present at different ages based on the affected organ and subclass of disease. In the subclasses of VHLS that cause renal disease, renal involvement typically begins closer to the end of the second decade of life and can present in different ways ranging from simple cystic lesions to solid tumors. Mutations in *vhl* are most often associated with clear cell renal carcinoma, the most common type of renal cancer, and also play a major role in sporadic cases of clear cell renal carcinoma. The recurrent, multifocal nature of this disease presents difficult challenges in the long-term management of patients with VHLS. Optimization of renal function warrants the use of several different approaches common to the management of renal carcinoma such as nephron sparing surgery, enucleation, ablation, and targeted therapies. In VHLS, renal lesions of 3 cm or bigger are considered to have metastatic potential and even small lesions often harbor malignancy. Many of the aspects of management revolve around optimizing both oncologic outcome and long-term renal function. As new surgical strategies and targeted therapies develop, the management of this complex disease evolves. This review will discuss the key aspects of the current management of VHLS.

Received: 25 August 2015; **Accepted after revision:** 20 September 2015; **Published:** 25 September 2015.

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How to cite: Ashouri K, Mohseni S, Tourtelot J, Sharma P, Spiess PE. Implications of Von Hippel-Lindau Syndrome and Renal Cell Carcinoma. *Journal of Kidney Cancer and VHL* 2015;2(4):163-173. Doi: <http://dx.doi.org/10.15586/jkcvhl.2015.41>

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Introduction

Von Hippel-Lindau syndrome (VHLS) is a rare, but highly penetrant, autosomal dominant hereditary neoplastic disorder caused by a genetic mutation in the *vhl* gene (*vhl*) that occurs in roughly 1 in 36,000

births (1). Mutations in *vhl* can be deletions or missense in the short arm of chromosome 3, resulting in an increased propensity to develop a variety of benign and malignant tumors, such as hemangioblastomas in the central nervous system (CNS), renal cysts and renal cell carcinomas (RCC),

pheochromocytomas, pancreatic islet cell tumors, endolymphatic sac tumors, and both broad ligament and epididymal cystadenomas in women and men, respectively (1, 2). The disease is classified into type 1, the result of a nonsense mutation or deletion, and type 2, the result of a missense mutation, based on adrenal involvement and risk of pheochromocytoma, in which type 2 carries risk of pheochromocytoma and type 1 does not (3, 4). Type 1 typically carries high risk for renal involvement, high risk for CNS hemangioblastomas, and low risk for the development of pheochromocytomas (1). Type 2 VHLS is further divided into A, B, and C based on the predisposition for renal involvement, classified as low risk, high risk, and no risk, respectively (3, 4). Type 2A typically carries low risk for renal involvement and high risk for the development of both hemangioblastomas of the CNS and pheochromocytomas. Type 2B typically carries high risk for renal involvement, CNS hemangioblastomas, and pheochromocytomas. Type 2C typically carries very low risk for renal involvement, low risk for CNS hemangioblastomas and high risk for pheochromocytomas (1). This review will discuss the implications of renal involvement in VHLS and therefore only include VHLS type 1, type 2A, and type 2B.

Materials and Methods

A comprehensive literature review was conducted for articles published between 1990 and 2015 using PubMed with regard to genetic and molecular mechanisms of *vhl* and the clinical and pathological diagnosis, treatments, and prognosis of renal tumors in this disease. Data for this review was acquired using combinations of the following terms: von Hippel-Lindau syndrome, von Hippel-Lindau disease, von Hippel-Lindau, renal cell carcinoma, radiology, screening, treatment, nephrectomy, VHL, VHL gene, HIF, metastatic, nephron sparing surgery, ablation. We excluded articles not written in English and any study that excluded hereditary cancer syndromes from its inclusion criteria, which resulted in the exclusion of approximately 10 peer-reviewed articles each. After assessing the available literature, 81 peer-reviewed articles and

their sources met our inclusion criteria and were included in this review.

Genetics of *vhl* and Molecular Mechanisms of VHL protein

The *vhl* gene is located at chromosome 3p25-p26, and contains 3 exons; over 300 germ-line mutations have been described leading to VHLS (1, 2). The *vhl* gene is also conserved across many taxa of living organisms and thus is considered to be fundamental to life (5). Several studies have implicated the role of *vhl* as a tumor suppressor gene and mutations of the wild-type and loss of heterozygosity are thought to play a major role in tumorigenesis (6).

The protein product of *vhl*, (pVHL), plays a major role in the ubiquitination of multiple proteins involved in many cellular pathways and contains binding domains for elongin C, elongin B, and cullin-2 (7-9). The resulting multimeric protein complex is known as the VHL complex (7). Often mutations in *vhl* affect the expression of pVHL, but some missense mutations can affect pVHL and elongin C binding (10). The role of hypoxia induced factor (HIF), a helix-loop-helix transcription factor that plays a major role in the cellular response mechanism to hypoxia, in the VHL pathway has been well-documented (11-14). The functional, dimeric form of HIF is comprised of 2 subunits, an HIF α subunit (either HIF1 α , HIF2 α , or HIF3 α) and an HIF1 β subunit (15, 16). HIF β is constitutively expressed in contrast to HIF α , whose expression is tightly regulated via ubiquitination by the VHL complex and proteasomal degradation (9). Under normoxic conditions the VHL complex recognizes HIF α , which is polyubiquitinated and degraded, thus reducing cytoplasmic HIF α (17). This recognition of HIF α for ubiquitination by VHL complex requires hydroxylation of proline residues on HIF α subunits by prolyl hydroxylase enzymes (PHD1, PHD2, PHD3), which require oxygen as a cofactor (7, 8, 18, 19). Thus, hydroxylation of HIF α proline residues and subsequent ubiquitination occurs only in the presence of oxygen (20). Under hypoxic conditions, hydroxylation of HIF α prolines does not proceed, preventing or reducing the VHL complex recognition of HIF for ubiquitination, leading to the accumulation of HIF α , making it available to

bind constitutively expressed HIF β (21, 22). This new HIF $\alpha\beta$ complex passes into the nucleus and binds to sequences known as hypoxia response elements that regulate of over 800 hypoxia inducible genes (23). Under normal conditions, expression of these hypoxia inducible genes allows a normal cellular adaptive response to hypoxia; however the aberrant regulation of many of these genes plays a major role in tumorigenesis and angiogenesis. The role in tumorigenesis of these genes varies. Several hypoxia inducible genes such as vascular endothelial growth factor (VEGF), erythropoietin, and platelet-derived growth factor (PDGF) have been found to have roles in angiogenesis (21, 24). Other targets of the pVHL-HIF pathway include glucose uptake via glucose transporter 1 (GLUT1), extracellular matrix remodeling via E-cadherin and matrix metalloproteinases (MMPs), microtubule stabilization via modulation of the primary cilium, regulation of apoptosis via p53, cell proliferation via transforming growth factor- α (TGF α) and epidermal growth factor (EGFR), and cell cycle progression via cyclin D1 (25-31). More recent studies have elucidated the role of mutations in other genes coding for proteins involved in the VHL-HIF pathway, such as TECB1 coding elongin C, resulting in similar aberrancies in the pVHL-mediated regulation of HIF and downstream induction of hypoxia inducible genes (32).

Several studies have also elucidated other roles of pVHL in HIF-independent pathways involved in tumorigenesis including the modulation of extracellular matrix components such as fibronectin, collagen IV, and integrins (33-35). Furthermore, pVHL has been found to be involved in the non-HIF-mediated stabilization of microtubules through a pVHL binding protein (VBP-1) by impacting the assembly of microtubule subunits (11, 18, 36).

Renal Pathology and VHLS

VHLS is the most common cause of hereditary renal cell carcinoma (RCC), specifically clear cell renal carcinoma (ccRCC) (37). Prior to the use of computed tomography (CT) in regular surveillance of VHLS, metastatic ccRCC was thought to be the leading cause of death in VHL patients (38). VHLS-associated RCC comprises about

4% of the total RCC incidence and is equal in both males and females (5, 39). While renal cysts are estimated to occur in up to 63% of patients with VHLS, renal tumors are estimated to occur in 25-45% of patients with VHLS and have a mean age at presentation of 39 years (5, 37). It should be noted that even cases of sporadic, non-familial ccRCC involve mutations or methylation of vhl in up to 91% of cases (40). Renal tumors less than 3 cm tend to be less invasive with lower pathological T stage; however these tumors tend to become more aggressive as they increase in size (41). Growth rates of renal tumors vary from 0.2 cm/year to 2.2 cm/year with a mean of 1.6 cm/year and a mean volume doubling time of approximately 26 months (42, 43). Metastasis is rare in tumors under 3 cm and tumor size on CT has been shown to be a prognostic indicator of overall survival (5, 41).

Renal involvement in VHLS is typically characterized by multiple, bilateral, recurring cysts or tumors (37, 38). Previous studies identified an average of approximately 7 renal cysts and 3 renal tumors per kidney, however it is hypothesized that the number of microscopic foci of malignant and non-malignant neoplasms could be in the hundreds and thousands, respectively (44, 45).

Renal cysts can be classified as simple or complex, depending on the presence and pattern of septation. The Bosniak classification scale is useful in describing radiologic findings in VHLS, but due to the high potential for malignancy of VHL cysts, its prognostic use in management is limited (39). Simple sporadic renal cysts have a very low rate of malignancy under the classic Bosniak classification system, whereas in VHLS simple cysts often already contain malignant characteristics in their walls (45). For this reason, it is recommended to undergo screening and active surveillance every 6 to 12 months depending on the rate of tumor growth and the patient specific history of malignancy (46).

Radiology of renal masses in VHL

Renal masses in VHLS often present as cystic masses on CT with varying degrees of

complexity and septa. Classically, the Bosniak classification scale provides a method of assessing the malignant potential of a renal cyst based on the radiologic characteristics of the cyst and provides diagnostic indications for management (47). It was initially described in the context of CT, but can also be used for MRI and has also been applied to ultrasound (47, 48). As stated above, it is important to note that recommendations for management of renal cysts cannot be made based on the Bosniak classification due to the fact that VHL renal cysts tend to have foci of cellular malignancy in their walls (46). For this reason, even simple cysts are likely to contain some malignancy and require active surveillance. The Bosniak system is useful in stratifying the variation of presentations of renal lesions in VHLS, but management decisions should be based on the consistency of the lesion and size of the largest mass (41). Even larger masses, such as a category III, may necessitate active surveillance instead of treatment based on the consistency of the lesion and number of surgeries in the patient's history (39). The use of ultrasound has proven useful in the surveillance of lesions and identifying their consistency, however, this technique lacks uniformity and is user dependent. CT and MRI are the most useful imaging modalities for the surveillance of renal lesions of VHLS (39).

The feasibility and safety of regular CT scan in patients with VHLS has come into question considering that MRI is a better modality for monitoring and identifying several non-renal tumors common to VHLS and that high levels of radiation exposure can occur with regular CT (46). Several studies suggest the use of regular MRI in surveillance of VHL lesions; however these advantages are not without risk (49, 50). The possibility of nephrogenic systemic fibrosis secondary to the use of gadolinium contrast in patients with compromised renal function and cost should be considered in surveillance strategy (50). For this reason, it is important to consider each patient on a case-by-case basis, but active surveillance has been well established as a key aspect of the management of VHLS (39).

Treatment of renal masses in VHLS

Over the past few decades, nephron-sparing

surgery (NSS), either open partial nephrectomy (PN) or enucleation, has become the standard surgical intervention for managing RCC in VHLS (51-53). Prior to NSS, the major approach was bilateral radical nephrectomy, leaving patients dependent on transplantation or dialysis, with high mortality rate and low quality of life (54). In 1995, Steinbach et al (54) found cancer-specific survival (CSS) rates at 5 and 10 years of 100 and 81 percent for patients undergoing NSS, compared to the CSS rates at 5 and 10 years of 73 and 36 percent for patients undergoing radical nephrectomy. Since then other studies have identified NSS as a superior approach for most cases (55, 56). The goal of NSS is to maximize remaining functional renal parenchyma (56). Since smaller renal tumors are typically considered low stage and low grade with low metastatic potential, PN is usually reserved for tumors greater than 3 cm, at which metastatic potential drastically increases, however some centers prefer a 4 cm cutoff (42, 57). The de novo recurrence rate of RCC in VHL is up to 85% at 10 years and the need for multiple NSSs will arise in many patients (54). It should be noted that in 2007, Ploussard et al (58) found a cancer-specific survival of 93.8% in a 24 VHL patient cohort in which 63.4% of patients required repeat NSS. A more recent study in 2012 by Singer et al (55) found a cancer specific survival at 10 years of 97% in 128 patients who underwent at least bilateral NSS, of which 70% had VHL and 68% of patients required repeat ipsilateral NSS. In 2012, Jilg et al (57) also found repeat NSS to be practical, establishing a limit of up to three repeated NSSs with complication rates of 53.6% for the first NSS, 33.3% for the second NSS, and 67% for the third NSS with a high proportion of more severe complications after the third NSS. Furthermore, NSS has been proven to be superior to bilateral radical nephrectomy and renal replacement therapy in overall survival, cancer-specific survival and morbidity and mortality in the management of VHLS (55). However, this study and other studies highlight the technical skill requirement and difficulty of repeat NSS (55, 59, 60). The number of repeated surgeries is limited by post-operative fibrosis and the increased morbidity and mortality associated with each surgery (53, 56, 61).

The recurrent nature of this disease obviates the need for a less invasive second-line therapy (60). Percutaneous ablation, including radiofrequency ablation and cryoablation, has proven to be a major candidate in both early and later stages of management of smaller renal tumors in VHLS (61-65). Percutaneous ablation is typically reserved for tumors between 2-3 cm and as a secondary procedure in most cases (42, 65). In 2010, Park et al. (66) found that radiofrequency ablation successfully managed 88% of renal masses in 11 patients with recurring tumors secondary to VHLS. In 2013, Yang et al (67) found cryoablation to have a failure rate of 7.7% in patients with VHLS compared to a failure rate of 21.4% for radiofrequency ablation. They also found both radiofrequency ablation and cryoablation to have 100% 5-year CSS (67). Although recurrence rates are higher than NSS, ablation carries lower incidence of morbidity and mortality (66). However, there are several contraindications to ablation including cystic tumors, tumors adjacent to critical structures (e.g. ureter and bowel) and extensively multifocal tumors, all of which are common in VHLS (60).

Several surgical interventions are available for the management of RCC in VHLS and weighing oncologic outcomes against operative morbidity and mortality is crucial in VHL management decisions (60). However, trends in preserving functional renal parenchyma in patients with VHL have improved survival outcomes over the past few decades (56). Pre-operative management and surveillance of other malignancies must also be considered, as these can cause major intraoperative complications, such as hemorrhage of a hemangioblastoma or pheochromocytoma disrupting patient intraoperative stability (53). It is important to keep in mind that these patients are not only subject to RCC but also multifaceted systemic malignant neoplasms, the treatment of which can have high morbidity and severely decrease quality of life (53). Maximizing efforts to use minimally invasive techniques should be a major consideration of future management.

Targeted therapy in RCC and VHL

Advancements in understanding vhl and the activity of pVHL have provided key insight

into potential mechanisms of therapeutic intervention in ccRCC and systemic treatment for VHL using targeted therapies (68). Few studies have been conducted on the efficacy of these drugs specifically in the context of VHSL, however, extensive research has been conducted on metastatic ccRCC. Considering most ccRCC involves aberrancies in vhl, these studies are expected to translate (40).

Older targeted therapies involved immunomodulation through cytokines IFN- α and IL-2, and had low relative risk reduction. IFN- α proved to have little therapeutic value and caused flu-like symptoms, fatigue, and depression; however, it is still seen in some trials as a point of reference (69). IL-2 showed benefit in some patients for long-term use and is still recommended in few select cases, however the adverse events of IL-2 have limited its use in current practice (70). These more classic immunotherapies have given way to newer targeted therapies involving the VEGF and mammalian target of rapamycin (mTOR) pathways that have since been approved by the Food and Drug Administration (FDA).

As noted previously, the loss of functional pVHL results in downstream over-expression of VEGF (21). Several oral tyrosine kinase inhibitors (TKI), sunitinib, sorafenib, pazopanib, and axitinib, and one monoclonal antibody, bevacizumab, decrease VEGF and PDGF signaling thereby decreasing angiogenesis and disrupting tumorigenesis (71). In 2009, Motzer et al (72) found a significantly improved overall survival and progression free survival (PFS) using Sunitinib as a first-line treatment in patients with metastatic ccRCC. Another study by Jonasch et al (73) found a 33% partial regression of RCC in a cohort of 18 patients with VHLS ($p=0.014$). Sorafenib, a TKI similar to sunitinib, as first-line treatment has also been proven to improve PFS in metastatic ccRCC, however, both sunitinib and sorafenib have similar adverse event profiles including diarrhea, rash, fatigue, and hand-foot syndrome (72, 74). Early trials with pazopanib found a significantly increased PFS with a better adverse event profile than other TKI anti-angiogenics, however more recent trials have shown no significant increase in

overall survival over placebo (75, 76). Bevacizumab, a humanized monoclonal antibody for VEGF- α , has shown increased PFS in second-line use with IFN- α (77). Many other TKIs are under investigation for use in ccRCC, but have not yet been approved by the FDA (71).

Furthermore, the FDA has approved two agents, temsirolimus and everolimus, for ccRCC that involve the disruption of the PI3K/AKT/mTOR pathway. Temsirolimus, an inhibitor of mTORC1, is an intravenous agent that has shown significantly improved survival and can be used among high-risk patients (78, 79). Everolimus, an oral mTOR directed therapy, has been approved for second-line use in metastatic ccRCC, showing improved PFS, but no overall survival improvement (80, 81). Both of these agents have a similar adverse event profile of rash, mucositis, interstitial pneumonitis, hyperglycemia, and hyperlipidemia (78, 80).

Conclusion

In conclusion, VHLS is a multifocal, hereditary neoplastic disorder that necessitates a complex multidisciplinary approach in its management (53). The protein product of *vhl*, known as pVHL, forms a multimeric complex with other intracellular proteins to form a molecule capable of ubiquitinating and degrading HIF proteins, preventing the over-expression of hypoxia inducible genes. Aberrant VHL contributes to angiogenesis and tumorigenesis and is implicated in both sporadic and VHLS-associated ccRCC (7-32). Tumors can be cystic or solid in nature and a cutoff of typically 3 cm, but sometimes 4 cm, is used to indicate the necessity for surgical management (42, 57). Radiologic surveillance can be done via several modalities including ultrasound, MRI, and most often CT, however each modality has its own advantages and disadvantages (39). NSS is the preferred surgical approach for renal tumors in VHL, typically using open PN for earlier, larger masses and percutaneous ablation for later, smaller tumors (55, 66). Systemic therapy, involving TKIs of the VEGF pathway and mTOR inhibitors, has been shown to improve CSS in metastatic ccRCC and in the rare studies available on VHL only cohorts (71-81). The multifocal nature of VHL mandates

complex multidisciplinary management strategies that require special considerations (53).

Conflicts of Interest

None to declare.

References

1. Barontini M, Dahia PL. VHL disease. *Best Pract Res Clin Endocrinol Metab.* 2010;24(3):401-13.
<http://dx.doi.org/10.1016/j.beem.2010.01.002>
PMid:20833332
2. Safo AO, Pambuccian SE. Pancreatic manifestations of von Hippel-Lindau disease. *Arch Pathol Lab Med.* 2010;134(7):1080-3.
PMid:20586642
3. Banks RE, Tirukonda P, Taylor C, et al. Genetic and epigenetic analysis of von Hippel-Lindau (VHL) gene alterations and relationship with clinical variables in sporadic renal cancer. *Cancer Res.* 2006;66(4):2000-11.
<http://dx.doi.org/10.1158/0008-5472.CAN-05-3074>
PMid:16488999
4. Ong KR, Woodward ER, Killick P, et al. Genotype-phenotype correlations in von Hippel-Lindau disease. *Hum Mutat.* 2007;28(2):143-9.
<http://dx.doi.org/10.1002/humu.20385>
PMid:17024664
5. Glenn GM. Von Hippel-Lindau Syndrome. *Encyclopedia of endocrine Diseases.* 2008; 4:674-687.
6. Crossey PA, Foster K, Richards FM, et al. Molecular genetic investigations of the mechanism of tumorigenesis in von Hippel-Lindau disease: analysis of allele loss in VHL tumours. *Hum Genet.* 1994;93(1):53-8.
<http://dx.doi.org/10.1007/BF00218913>
PMid:8270255
7. Calzada MJ. Von Hippel-Lindau syndrome: molecular mechanisms of the disease. *Clin Transl Oncol.* 2010;12(3):160-5.
<http://dx.doi.org/10.1007/s12094-010-0485-9>
PMid:20231120
8. Pause A, Lee S, Worrell RA, et al. The von Hippel-Lindau tumor-suppressor gene product forms a stable complex with human CUL-2, a member of the Cdc53 family of proteins. *Proc Natl Acad Sci U S A.* 1997;94(6):2156-61.
<http://dx.doi.org/10.1073/pnas.94.6.2156>
PMid:9122164 PMCID:PMC20057

9. Pause A, Peterson B, Schaffar G, et al. Studying interactions of four proteins in the yeast two-hybrid system: structural resemblance of the pVHL/elongin BC/hCUL-2 complex with the ubiquitin ligase complex SKP1/cullin/F-box protein. *Proc Natl Acad Sci U S A*. 1999;96(17):9533-8.
<http://dx.doi.org/10.1073/pnas.96.17.9533>
PMid:10449727 PMCid:PMC22243
10. Stebbins CE, Kaelin WG, Jr., Pavletich NP. Structure of the VHL-ElonginC-ElonginB complex: implications for VHL tumor suppressor function. *Science*. 1999;284(5413):455-61.
<http://dx.doi.org/10.1126/science.284.5413.455>
PMid:10205047
11. Kim JJ, Rini BI, Hansel DE. Von Hippel Lindau syndrome. *Adv Exp Med Biol*. 2010;685:228-49.
http://dx.doi.org/10.1007/978-1-4419-6448-9_22
PMid:20687511
12. Latif F, Tory K, Gnarra J, et al. Identification of the von Hippel-Lindau disease tumor suppressor gene. *Science*. 1993;260(5112):1317-20.
<http://dx.doi.org/10.1126/science.8493574>
PMid:8493574
13. Woodward ER, Buchberger A, Clifford SC, et al. Comparative sequence analysis of the VHL tumor suppressor gene. *Genomics*. 2000;65(3):253-65.
<http://dx.doi.org/10.1006/geno.2000.6144>
PMid:10857749
14. Stadler W. Chromosomes, hypoxia, angiogenesis, and trial design: A brief history of renal cancer drug development. *Clinical Cancer Research*. 2007;13(6):1630-1633.
<http://dx.doi.org/10.1158/1078-0432.CCR-06-2721>
PMid:17363513
15. Semenza GL. Hypoxia-inducible factors: mediators of cancer progression and targets for cancer therapy. *Trends Pharmacol Sci*. 2012;33(4):207-14.
<http://dx.doi.org/10.1016/j.tips.2012.01.005>
PMid:22398146 PMCid:PMC3437546
16. Wang GL, Semenza GL. General involvement of hypoxia-inducible factor 1 in transcriptional response to hypoxia. *Proc Natl Acad Sci U S A*. 1993;90(9):4304-8.
<http://dx.doi.org/10.1073/pnas.90.9.4304>
17. Ivan M, Kondo K, Yang H, et al. HIFalpha targeted for VHL-mediated destruction by proline hydroxylation: implications for O2 sensing. *Science*. 2001;292(5516):464-8.
<http://dx.doi.org/10.1126/science.1059817>
PMid:11292862
18. Brinke A, Green PM, Giannelli F. Characterization of the gene (VBP1) and transcript for the von Hippel-Lindau binding protein and isolation of the highly conserved murine homologue. *Genomics*. 1997;45(1):105-12.
<http://dx.doi.org/10.1006/geno.1997.4902>
PMid:9339366
19. Yu F, White SB, Zhao Q, et al. HIF-1alpha binding to VHL is regulated by stimulus-sensitive proline hydroxylation. *Proc Natl Acad Sci U S A*. 2001;98(17):9630-5.
<http://dx.doi.org/10.1073/pnas.181341498>
PMid:11504942 PMCid:PMC55503
20. Min JH, Yang H, Ivan M, et al. Structure of an HIF-1alpha-pVHL complex: hydroxyproline recognition in signaling. *Science*. 2002;296(5574):1886-9.
<http://dx.doi.org/10.1126/science.1073440>
PMid:12004076
21. Iliopoulos O, Levy AP, Jiang C, et al. Negative regulation of hypoxia-inducible genes by the von Hippel-Lindau protein. *Proc Natl Acad Sci U S A*. 1996;93(20):10595-9.
<http://dx.doi.org/10.1073/pnas.93.20.10595>
PMid:8855223 PMCid:PMC38198
22. Jaakkola P, Mole DR, Tian YM, et al. Targeting of HIF-alpha to the von Hippel-Lindau ubiquitylation complex by O2-regulated prolyl hydroxylation. *Science*. 2001;292(5516):468-72.
<http://dx.doi.org/10.1126/science.1059796>
PMid:11292861
23. Semenza GL. Oxygen sensing, homeostasis, and disease. *N Engl J Med*. 2011;365(6):537-47.
<http://dx.doi.org/10.1056/NEJMr1011165>
PMid:21830968
24. Gnarra JR, Zhou S, Merrill MJ, et al. Post-transcriptional regulation of vascular endothelial growth factor mRNA by the product of the VHL tumor suppressor gene. *Proc Natl Acad Sci U S A*. 1996;93(20):10589-94.
<http://dx.doi.org/10.1073/pnas.93.20.10589>
PMid:8855222 PMCid:PMC38197
25. Maxwell PH, Wiesener MS, Chang GW, et al. The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. *Nature*. 1999;399(6733):271-5.
<http://dx.doi.org/10.1038/20459>
PMid:10353251

26. Knebelmann B, Ananth S, Cohen HT, et al. Transforming growth factor alpha is a target for the von Hippel-Lindau tumor suppressor. *Cancer Res.* 1998;58(2):226-31. PMID:9443397
27. Gunaratnam L, Morley M, Franovic A, et al. Hypoxia inducible factor activates the transforming growth factor-alpha/epidermal growth factor receptor growth stimulatory pathway in VHL(-/-) renal cell carcinoma cells. *J Biol Chem.* 2003;278(45):44966-74. <http://dx.doi.org/10.1074/jbc.M305502200> PMID:12944410
28. Evans AJ, Russell RC, Roche O, et al. VHL promotes E2 box-dependent E-cadherin transcription by HIF-mediated regulation of SIP1 and snail. *Mol Cell Biol.* 2007;27(1):157-69. <http://dx.doi.org/10.1128/MCB.00892-06> PMID:17060462 PMCID:PMC1800649
29. Petrella BL, Lohi J, Brinckerhoff CE. Identification of membrane type-1 matrix metalloproteinase as a target of hypoxia-inducible factor-2 alpha in von Hippel-Lindau renal cell carcinoma. *Oncogene.* 2005;24(6):1043-52. <http://dx.doi.org/10.1038/sj.onc.1208305> PMID:15592504 PMCID:PMC1847637
30. Sandoel A, Kohler I, Fellmann C, et al. HIF-1 antagonizes p53-mediated apoptosis through a secreted neuronal tyrosinase. *Nature.* 2010;465(7298):577-83. <http://dx.doi.org/10.1038/nature09141> PMID:20520707 PMCID:PMC3328299
31. Baba M, Hirai S, Yamada-Okabe H, et al. Loss of von Hippel-Lindau protein causes cell density dependent deregulation of CyclinD1 expression through hypoxia-inducible factor. *Oncogene.* 2003;22(18):2728-38. <http://dx.doi.org/10.1038/sj.onc.1206373> PMID:12743597
32. Sato Y, Yoshizato T, Shiraishi Y, et al. Integrated molecular analysis of clear-cell renal cell carcinoma. *Nat Genet.* 2013;45(8):860-7. <http://dx.doi.org/10.1038/ng.2699> PMID:2379773
33. Kurban G, Duplan E, Ramlal N, et al. Collagen matrix assembly is driven by the interaction of von Hippel-Lindau tumor suppressor protein with hydroxylated collagen IV alpha 2. *Oncogene.* 2008;27(7):1004-12. <http://dx.doi.org/10.1038/sj.onc.1210709> PMID:17700531
34. Kurban G, Hudon V, Duplan E, et al. Characterization of a von Hippel Lindau pathway involved in extracellular matrix remodeling, cell invasion, and angiogenesis. *Cancer Res.* 2006;66(3):1313-9. <http://dx.doi.org/10.1158/0008-5472.CAN-05-2560> PMID:16452184
35. Ohh M, Yauch RL, Lonergan KM, et al. The von Hippel-Lindau tumor suppressor protein is required for proper assembly of an extracellular fibronectin matrix. *Mol Cell.* 1998;1(7):959-68. [http://dx.doi.org/10.1016/S1097-2765\(00\)80096-9](http://dx.doi.org/10.1016/S1097-2765(00)80096-9)
36. Tsuchiya H, Iseda T, Hino O. Identification of a novel protein (VBP-1) binding to the von Hippel-Lindau (VHL) tumor suppressor gene product. *Cancer Res.* 1996;56(13):2881-5. PMID:8674032
37. Lonser RR, Glenn GM, Walther M, et al. von Hippel-Lindau disease. *Lancet.* 2003;361(9374):2059-67. [http://dx.doi.org/10.1016/S0140-6736\(03\)13643-4](http://dx.doi.org/10.1016/S0140-6736(03)13643-4)
38. Maher ER, Yates JR, Harries R, et al. Clinical features and natural history of von Hippel-Lindau disease. *Q J Med.* 1990;77(283):1151-63. <http://dx.doi.org/10.1093/qjmed/77.2.1151> PMID:2274658
39. Meister M, Choyke P, Anderson C, et al. Radiological evaluation, management, and surveillance of renal masses in Von Hippel-Lindau disease. *Clin Radiol.* 2009;64(6):589-600. <http://dx.doi.org/10.1016/j.crad.2008.10.010> PMID:19414081
40. Nickerson ML, Jaeger E, Shi Y, et al. Improved identification of von Hippel-Lindau gene alterations in clear cell renal tumors. *Clin Cancer Res.* 2008;14(15):4726-34. <http://dx.doi.org/10.1158/1078-0432.CCR-07-4921> PMID:18676741 PMCID:PMC2629664
41. Kwon T, Jeong IG, Pak S, et al. Renal tumor size is an independent prognostic factor for overall survival in von Hippel-Lindau disease. *J Cancer Res Clin Oncol.* 2014;140(7):1171-7. <http://dx.doi.org/10.1007/s00432-014-1654-y> PMID:24671227
42. Walther MM, Choyke PL, Glenn G, et al. Renal cancer in families with hereditary renal cancer: prospective analysis of a tumor size threshold for renal parenchymal sparing surgery. *J Urol.* 1999;161(5):1475-9. [http://dx.doi.org/10.1016/S0022-5347\(05\)68930-6](http://dx.doi.org/10.1016/S0022-5347(05)68930-6)

43. Jilg CA, Neumann HP, Glasker S, et al. Growth kinetics in von Hippel-Lindau-associated renal cell carcinoma. *Urol Int.* 2012;88(1):71-8. <http://dx.doi.org/10.1159/000333348> PMID:22156657
44. Poston CD, Jaffe GS, Lubensky IA, et al. Characterization of the renal pathology of a familial form of renal cell carcinoma associated with von Hippel-Lindau disease: clinical and molecular genetic implications. *J Urol.* 1995;153(1):22-6. <http://dx.doi.org/10.1097/00005392-199501000-00009> PMID:7966777
45. Walther MM, Lubensky IA, Venzon D, et al. Prevalence of microscopic lesions in grossly normal renal parenchyma from patients with von Hippel-Lindau disease, sporadic renal cell carcinoma and no renal disease: clinical implications. *J Urol.* 1995;154(6):2010-4; discussion 2014-5.
46. Choyke PL, Glenn GM, Walther MM, et al. von Hippel-Lindau disease: genetic, clinical, and imaging features. *Radiology.* 1995;194(3):629-42. <http://dx.doi.org/10.1148/radiology.194.3.7862955> PMID:7862955
47. Bosniak MA. The current radiological approach to renal cysts. *Radiology.* 1986;158(1):1-10. <http://dx.doi.org/10.1148/radiology.158.1.3510019> PMID:3510019
48. Karmazyn B, Tawadros A, Delaney LR, et al. Ultrasound classification of solitary renal cysts in children. *J Pediatr Urol.* 2015;11(3):149 e1-6.
49. Hes FJ, Feldberg MA. Von Hippel-Lindau disease: strategies in early detection (renal-, adrenal-, pancreatic masses). *Eur Radiol.* 1999;9(4):598-610. <http://dx.doi.org/10.1007/s003300050717> PMID:10354869
50. Tattersall DJ, Moore NR. von Hippel-Lindau disease: MRI of abdominal manifestations. *Clin Radiol.* 2002;57(2):85-92. <http://dx.doi.org/10.1053/crad.2001.0747> PMID:11977939
51. Novick AC, Zincke H, Neves RJ, et al. Surgical enucleation for renal cell carcinoma. *J Urol.* 1986;135(2):235-8. PMID:3944851
52. Novick AC, Stroom SB. Long-term followup after nephron sparing surgery for renal cell carcinoma in von Hippel-Lindau disease. *J Urol.* 1992;147(6):1488-90. PMID:1593671
53. Herring JC, Enquist EG, Chernoff A, et al. Parenchymal sparing surgery in patients with hereditary renal cell carcinoma: 10-year experience. *J Urol.* 2001;165(3):777-81. [http://dx.doi.org/10.1016/S0022-5347\(05\)66524-X](http://dx.doi.org/10.1016/S0022-5347(05)66524-X)
54. Steinbach F, Novick AC, Zincke H, et al. Treatment of renal cell carcinoma in von Hippel-Lindau disease: a multicenter study. *J Urol.* 1995;153(6):1812-6. [http://dx.doi.org/10.1016/S0022-5347\(01\)67318-X](http://dx.doi.org/10.1016/S0022-5347(01)67318-X)
55. Singer EA, Vourganti S, Lin KY, et al. Outcomes of patients with surgically treated bilateral renal masses and a minimum of 10 years of followup. *J Urol.* 2012;188(6):2084-8. <http://dx.doi.org/10.1016/j.juro.2012.08.038> PMID:23083858 PMCid:PMC3810017
56. Metwalli AR, Linehan WM. Nephron-sparing surgery for multifocal and hereditary renal tumors. *Curr Opin Urol.* 2014;24(5):466-73. <http://dx.doi.org/10.1097/MOU.0000000000000094> PMID:25014245 PMCid:PMC4441729
57. Jilg CA, Neumann HP, Glasker S, et al. Nephron sparing surgery in von Hippel-Lindau associated renal cell carcinoma; clinicopathological long-term follow-up. *Fam Cancer.* 2012;11(3):387-94. <http://dx.doi.org/10.1007/s10689-012-9525-7> PMID:22426863
58. Ploussard G, Droupy S, Ferlicot S, et al. Local recurrence after nephron-sparing surgery in von Hippel-Lindau disease. *Urology.* 2007;70(3):435-9. <http://dx.doi.org/10.1016/j.urology.2007.04.040> PMID:17905091
59. Singer EA, Bratslavsky G. Management of locally recurrent kidney cancer. *Curr Urol Rep.* 2010;11(1):15-21. <http://dx.doi.org/10.1007/s11934-009-0085-9> PMID:20425632
60. Matin SF, Ahrar K, Wood CG, et al. Patterns of intervention for renal lesions in von Hippel-Lindau disease. *BJU Int.* 2008;102(8):940-5. <http://dx.doi.org/10.1111/j.1464-410X.2008.07718.x> PMID:18485044
61. McDougal WS, Gervais DA, McGovern FJ, et al. Long-term followup of patients with renal cell

- carcinoma treated with radio frequency ablation with curative intent. *J Urol*. 2005;174(1):61-3.
<http://dx.doi.org/10.1097/01.ju.0000162046.45024.2b>
 PMid:15947578
62. Gill IS, Remer EM, Hasan WA, et al. Renal cryoablation: outcome at 3 years. *J Urol*. 2005;173(6):1903-7.
<http://dx.doi.org/10.1097/01.ju.0000158154.28845.c9>
 PMid:15879772
63. Pavlovich CP, Walther M, Choyke PL, et al. Percutaneous radio frequency ablation of small renal tumors: initial results. *J Urol*. 2002;167(1):10-5.
[http://dx.doi.org/10.1016/S0022-5347\(05\)65371-2](http://dx.doi.org/10.1016/S0022-5347(05)65371-2)
64. Shingleton WB, Sewell PE. Percutaneous cryoablation of renal cell carcinoma in a transplanted kidney. *BJU Int*. 2002;90(1):137-8.
<http://dx.doi.org/10.1046/j.1464-410X.2002.02761.x>
65. Joly D, Mejean A, Correas JM, et al. Progress in nephron sparing therapy for renal cell carcinoma and von Hippel-Lindau disease. *J Urol*. 2011;185(6):2056-60.
<http://dx.doi.org/10.1016/j.juro.2011.02.007>
 PMid:21496837
66. Park BK, Kim CK. Percutaneous radio frequency ablation of renal tumors in patients with von Hippel-Lindau disease: preliminary results. *J Urol*. 2010;183(5):1703-7.
<http://dx.doi.org/10.1016/j.juro.2010.01.022>
 PMid:20299060
67. Yang B, Autorino R, Remer EM, et al. Probe ablation as salvage therapy for renal tumors in von Hippel-Lindau patients: the Cleveland Clinic experience with 3 years follow-up. *Urol Oncol*. 2013;31(5):686-92.
<http://dx.doi.org/10.1016/j.urolonc.2011.05.008>
 PMid:21723752
68. Rini BI, Small EJ. Biology and clinical development of vascular endothelial growth factor-targeted therapy in renal cell carcinoma. *J Clin Oncol*. 2005;23(5):1028-43.
<http://dx.doi.org/10.1200/JCO.2005.01.186>
 PMid:15534359
69. Coppin C, Porzsolt F, Awa A, et al. Immunotherapy for advanced renal cell cancer. *Cochrane Database Syst Rev* 2005; 10.1002/14651858.CD001425.pub2(1):CD001425.
70. McDermott DF. Update on the application of interleukin-2 in the treatment of renal cell carcinoma. *Clin Cancer Res*. 2007;13(2 Pt 2):716s-720s.
<http://dx.doi.org/10.1158/1078-0432.CCR-06-1872>
 PMid:17255299
71. Randall JM, Millard F, Kurzrock R. Molecular aberrations, targeted therapy, and renal cell carcinoma: current state-of-the-art. *Cancer Metastasis Rev*. 2014;33(4):1109-24.
<http://dx.doi.org/10.1007/s10555-014-9533-1>
 PMid:25365943
72. Motzer RJ, Hutson TE, Tomczak P, et al. Overall survival and updated results for sunitinib compared with interferon alfa in patients with metastatic renal cell carcinoma. *J Clin Oncol*. 2009;27(22):3584-90.
<http://dx.doi.org/10.1200/JCO.2008.20.1293>
 PMid:19487381 PMCID:PMC3646307
73. Jonasch E, McCutcheon IE, Waguespack SG, et al. Pilot trial of sunitinib therapy in patients with von Hippel-Lindau disease. *Ann Oncol*. 2011;22(12):2661-6.
<http://dx.doi.org/10.1093/annonc/mdr011>
 PMid:22105611 PMCID:PMC4542805
74. Escudier B, Eisen T, Stadler WM, et al. Sorafenib in advanced clear-cell renal-cell carcinoma. *N Engl J Med*. 2007;356(2):125-34.
<http://dx.doi.org/10.1056/NEJMoa060655>
 PMid:17215530
75. Sternberg CN, Davis ID, Mardiak J, et al. Pazopanib in locally advanced or metastatic renal cell carcinoma: results of a randomized phase III trial. *J Clin Oncol*. 2010;28(6):1061-8.
<http://dx.doi.org/10.1200/JCO.2009.23.9764>
 PMid:20100962
76. Sternberg CN, Hawkins RE, Wagstaff J, et al. A randomised, double-blind phase III study of pazopanib in patients with advanced and/or metastatic renal cell carcinoma: final overall survival results and safety update. *Eur J Cancer*. 2013;49(6):1287-96.
<http://dx.doi.org/10.1016/j.ejca.2012.12.010>
 PMid:23321547
77. Escudier B, Pluzanska A, Koralewski P, et al. Bevacizumab plus interferon alfa-2a for treatment of metastatic renal cell carcinoma: a randomised, double-blind phase III trial. *Lancet*. 2007;370(9605):2103-11.
[http://dx.doi.org/10.1016/S0140-6736\(07\)61904-7](http://dx.doi.org/10.1016/S0140-6736(07)61904-7)
78. Lockhart AC, Rothenberg ML, Dupont J, et

al. Phase I study of intravenous vascular endothelial growth factor trap, aflibercept, in patients with advanced solid tumors. *J Clin Oncol*. 2010;28(2):207-14.

<http://dx.doi.org/10.1200/JCO.2009.22.9237>
PMid:19949018 PMCID:PMC281571

79. Hudes G, Carducci M, Tomczak P, et al. Temsirolimus, interferon alfa, or both for advanced renal-cell carcinoma. *N Engl J Med*. 2007;356(22):2271-81.

<http://dx.doi.org/10.1056/NEJMoa066838>
PMid:17538086

80. Motzer RJ, Escudier B, Oudard S, et al. Phase 3 trial of everolimus for metastatic renal cell carcinoma : final results and analysis of prognostic factors. *Cancer*. 2010;116(18):4256-65.

<http://dx.doi.org/10.1002/cncr.25219>
PMid:20549832

81. Motzer RJ, Escudier B, Oudard S, et al. Efficacy of everolimus in advanced renal cell carcinoma: a double-blind, randomised, placebo-controlled phase III trial. *Lancet*. 2008;372(9637):449-56.

[http://dx.doi.org/10.1016/S0140-6736\(08\)61039-9](http://dx.doi.org/10.1016/S0140-6736(08)61039-9)