# REVIEW ARTICLE



# Biologic characteristics of platelet rich plasma and platelet rich fibrin: A review

Farzaneh Lal Alizade<sup>1</sup>, Mustafa Kazemi<sup>1</sup>, Sahar Irani<sup>1</sup>, Mehdi Sohrabi<sup>2</sup>

<sup>1</sup>Department of Orthodontics, School of Dentistry, Mashhad University of Medical Sciences, Mashhad, Iran, <sup>2</sup>Oral and Maxillofacial Radiologist

#### Correspondence

Dr. Mustafa Kazemi, Department of Orthodontics, Faculty of Dentistry, Mashhad University of Medical Sciences, Mashhad, Iran. Tel/Fax: +9809151734184. E-mail: Mustafaortho94@gmail.com

Received 13 May 2016; Accepted 10 July 2016

doi: 10.15713/ins.ijcdmr.104

### How to cite the article:

Farzaneh Lal Alizade, Mustafa Kazemi, Sahar Irani, Mehdi Sohrabi, "Biologic characteristics of platelet rich plasma and platelet rich fibrin: A review," Int J Contemp Dent Med Rev, vol. 2016, Article ID: 030516, 2016. doi: 10.15713/ins.ijcdmr.104

# Abstract

The recent development of platelet concentrate for surgical use is an evolution of the fibrin glue technologies used since many years. Fibrin is a biologic glue that compact platelet clusters during coagulation process fibrin glues are biologic products that their local application has been used to decrease bleeding and accelerate tissue healing in the past three decades. Fibrin glue prepared based on a natural biologic mechanism (polymerization of fibrin during homeostasis) that reinforced by an artificial way. However, they were ever criticized because were blood derived products and had a high risk of viral infection the production of these products is based on the mixture of two plasma components including fibrinogen and thrombin. Platelet-rich plasma (PRP) is a fibrin matrix in which platelet cytokines, growth factors, and cells are trapped and may be released after a certain time. Choukroun et al. used platelet rich fibrin (PRF) protocol in oral and maxillofacial surgery to improve bone healing. Autologous PRF is considered to be a healing biomaterial, and presently, studies have shown its application in various disciplines of dentistry. PRF is the second generation of platelet concentrates which allows fibrin membranes to get enriched with platelets and growth factors, starting from an anticoagulant free blood harvest. PRF is similar to a fibrin network that allows cell migration and proliferation, and consequently, a more efficient cicatrization. Many growth factors, such as platelet-derived growth factor and transforming growth factor, are released from PRF. One of the major differences between PRF and PRP is the different polymerization that is responsible for the different biologic properties. PRF released autologous growth factors gradually and expressed stronger and more durable effect on proliferation and differentiation, which means that PRF could stimulate the surrounding environment to a more rapid wound healing. This review aims to evaluate the effect of biologic characteristics of fibrin glues, PRP, and PRF.

Keywords: Fibrin, platelet rich fibrin, platelet rich plasma

## Introduction

One of the researchers' great challenges is to obtain bioactive surgical additives to modulate inflammation and increase healing. After each surgical intervention, surgeons confront the complex phenomenon of tissue remodeling and the effects of it on healing. Although the application of fibrin glues was explained more than 30 years ago,<sup>[1,2]</sup> because of complex preparation protocol and also the risk of cross infection and its usage is still controversial. Development of platelet concentrating technique and enhanced efficacy of protocols lead to produce new fibrin glue were known as platelet-rich plasma (PRP) but due to legal limitation for the usage of blood products; a new generation of concentrating platelet

introduced in France. This new product named platelet rich fibrin (PRF) was like an autologous cycatrical matrix.

## Fibrin

The fibrin is a biologic glue that compact platelet clusters during the coagulation process. Fibrinogen is the final production of coagulation reaction and it is a soluble protein that by thrombin is converted to fibrin. Fibrin polymerized gel is the first matrix formed at the wound site.<sup>[3,4]</sup>

#### Fibrin and surgical additives

Fibrin glue prepared based on a natural biologic mechanism (polymerization of fibrin during homeostasis) that reinforced

by an artificial way.<sup>[5]</sup> However, they were ever criticized because were blood-derived products. Pharmacologic products had a high risk of viral infection, but today simple ways are introduced for production of fibrin autologous glues based on the development of platelet concentrating technique<sup>[6]</sup> that these products act similarly to the final stage of coagulation cascade that fibrinogen converts to fibrin.

## **Clinical use**

Despite significant differences between protocols, most of the studies showed the high efficacy of fibrin glues for controlling of slow and widespread bleeding of fibrin parenchyma but these products cannot be effective for sever vascular bleeding.<sup>[2]</sup> These glues successfully used to stop widespread microvascular bleeding and are famous of their ability to seal wound borders.<sup>[7]</sup> Surgeons use the mechanical properties of these products in addition to their biologic benefits.<sup>[8-10]</sup> Sealing with fibrin glues caused reduction of hematoma before the surgery.<sup>[11]</sup> The usage of fibrin products completely explained for plastic and maxillofacial surgery.<sup>[7-10]</sup> However, many surgeons used this glue in different surgeries on animal models and the results were contradictory specially for orthopedic and neurosurgery.<sup>[12]</sup> Sealing of dura mater or nerves for reconstruction in trauma or tumor surgery was not explained in literature. It showed that these products are not more than autologous fibrin glues that their major role is only connection of tissues.<sup>[12,13]</sup>

#### **Concentrated PRP**

Due to the risk of hepatitis infection, production of fibrin products has been prevented in USA from 1978<sup>[14]</sup> Hence, the combination of concentrated platelet products with the idea of cell therapy with growth factors proposed.<sup>[14]</sup> Concentrated platelets were used for prevention and treatment of bleeding caused by thrombopenic condition like maxillary aplasia and acute leukemia and their usage was limited.<sup>[6,15]</sup> The production protocol of PRR are based on two centrifugation for increased platelet aggregation. According to this protocol, the blood sample collected before surgery immediately centrifuged to concentrated platelets.<sup>[16]</sup> Then, thrombin and calcium chloride added to activate platelets and produced gel.<sup>[17-20]</sup>

Platelet concentration of plasma is 3-5 times more than human plasma.<sup>[21]</sup> Platelets have a major role on homeostasis and are a natural source for growth factors. Activation of platelets with different factors like thrombin, calcium chloride or collagen causes the release of growth factors. Growth factors have a key role on wound healing, regeneration, chemotaxis, proliferation, differentiation, and angiogenesis.<sup>[22]</sup> Platelets also release factors, such as fibronectin, vitronectin, and sphynosins-1-phosphate, that are effective for healing process. <sup>[23]</sup> In addition to release of different factors PRR forms a fibrin network that acts as a regeneration matrix.<sup>[24,25]</sup> PRR used for soft and hard tissue augmentation, because increases vascularization of grafts, improves soft tissue healing, decreases the mobility, and increases bone regeneration.<sup>[26]</sup> Improbable cross infection, immunologic reaction is another advantage of PRR.<sup>[27]</sup> PRR makes the handling of grafted material and packing of them more easier and increase the maintenance of space and bone regeneration potential.<sup>[28,29]</sup> Since PRR contains different growth factors, it stimulates angiogenesis and fibroblast differentiation. Hence, it recommended for improvement of soft tissue healing.<sup>[30]</sup> Studies showed that PRR increases the concentration of grafts and decreases its micron scale movement that provides and appropriate environment for a successful graft.<sup>[31]</sup> PRR increases wound maturation and epithelialization and decreases scar formation.<sup>[32]</sup> Epidermal growth factor and platelet-derived growth factor are the major growth factors responsible for fibroblast migration and collagen synthesis. Increased concentration of these growth factors are the probable causes of increased wound healing that it estimated 2-3 times more than normal situation.<sup>[32]</sup> In hard tissues growth factors affect osteoblast. The addition of PRR to stromal cells in animal model approved its angiogenic and osteogenic properties.<sup>[33]</sup> Studies showed the successful use of PRP for bone regeneration in periodontal defects, extracted sockets, implant surgery, and sinus augmentation.<sup>[34]</sup> One of the major challenges of bone augmentation is the prolonged healing period, so one of the important indications of PRP is to reduced healing period. Shorter time needed for grafts healing in sinus augmentation showed in studies.<sup>[32,33]</sup> Increased regeneration of bone in periodontal defects at the distal aspect of 2<sup>nd</sup> molar after surgical removal of 3rd molar had been reported in studies.<sup>[34,35]</sup> But unfortunately, these results were not complete because the biopsy cannot be taken from the site. In human studies, use of PRR increased bone healing and regeneration.<sup>[16,26]</sup> Short period of improvements of bone regeneration (for 2 weeks) was seen in sinus augmentation.<sup>[36]</sup> Only when target cells like osteoblast and osteocyte are present at the site, bone regeneration can occur.<sup>[37]</sup> Because studies used the combination of non-organic grafts with PRR did not show any benefit in different follow-up periods.[36] In animal studies when spongy bone of iliac crest used as the graft material for sinus lift with PRR the clinical results in case and control group was similar.<sup>[28,38]</sup> One of the results put forward the less than expected effects of PRR in studies, may be that concentration of growth factors in PRR is much less than expected effects of PRR in studies, may be that concentration of growth factors in PRR is much less than the value that has clinical effects. It seems that the cytokines potential that release during the platelet activation and fibrin network formation is extremely limited.<sup>[39,40]</sup> It expected that fibrin network acts as a supportive matrix for cytokines, but these soluble molecules release so fast that they cannot place into this structure.

## PRF

PRF was used for the first time by Choukroun *et al.*<sup>[41]</sup> in oral and maxillofacial surgery. This technique did not require to anticoagulants, thrombin and any other jellying factor and it was not anything except a centrifuged blood product without any other additives that was not included the laws about the

prohibition of blood born products. Only a PC-Q2 centrifuge and a kit are needed for PRF production. PRF preparation protocol is very easy. The blood samples collected without any anticoagulants and immediately centrifuged at 3000 rpm for 10 min. Not the usage of anticoagulants causes platelet and coagulation cascade as a result of its contact to the wall surface within minutes and platelet trapped in the fibrin network. Fibrin clot achieved at the middle part tube between red blood cells in the bottom and acellular plasma in upper. The success of this technique is dependent on the fast collection of blood sample and its transfer to the centrifuge. Without use of anticoagulants, the sample should be centrifuged within minutes otherwise the fibrinogen cannot be concentrated in the middle and upper part of tube. It is necessary to concentrate fibrinogen before thrombin convert it to fibrin. Hence, the fast preparation of this protocol is the key tip to achieve a useful PRF clot. The most important property of this technique is not usage of anticoagulants that leads to extensive platelet activation and release of large amounts of cytokines that these soluble molecules trapped into fibrin network.<sup>[42]</sup> One of the major differences between PRF and PRR is the different polymerization that is responsible for different biologic properties. The addition of thrombin and calcium chloride in PRP causes rapid polymerization, but PRF polymerization is slow and natural and thrombin activity is physiologic. The addition of bone thrombin in PRP leads to formation of compacted tetra molecules with bilateral connections and thickened polymerizes fibrin that is not appropriate for cytokine placement and cell migration. But its high strength is suitable for sealing the tissues. Tri molecules mono lateral connection formed in PRF due to physiologic concentration of thrombin that forms a flexible network that is appropriate for cytokine storage and cell migration and because of its flexibility they are used as membranes.

## Conclusion

Platelets contain important growth factors that increase cell proliferation, collagen production, chemotaxis, angiogenesis, and cell differentiation. PRP and PRF are concentrated platelets suspension in a small amount of plasma. Moreover, PRP and PRF contain three adhesive molecules (fibrin, fibronectin, and vitronectin) that reinforced osteoconduction. The major difference between these two products is their polymerization that leads to their different biologic characteristics. Polymerization of PRP is induced by addition of anticoagulants but PRF polymerization is a natural and slow process. Hence, PRF has more suitable fibrin network for storage of cytokines and growth factors and also cell migration.

# References

- 1. Matras H. Effect of various fibrin preparations on reimplantations in the rat skin. Osterr Z Stomatol 1970;67:338-59.
- Gibble JW, Ness PM. Fibrin glue: The perfect operative sealant? Transfusion 1990;30(8):741-7.

- Clark RA. Fibrin and wound healing. Ann N Y Acad Sci 2001;936:355-67.
- van Hinsbergh VW, Collen A, Koolwijk P. Role of fibrin matrix in angiogenesis. Ann N Y Acad Sci 2001;936:426-37.
- 5. Vinazzer H. Fibrin sealing: Physiologic and biochemical background. Facial Plast Surg 1985;2:291-5.
- Whitman DH, Berry RL, Green DM. Platelet gel: An autologous alternative to fibrin glue with applications in oral and maxillofacial surgery. J Oral Maxillofac Surg 1997;55:1294-9.
- Saltz R, Sierra D, Feldman D, Saltz MB, Dimick A, Vasconez LO. Experimental and clinical applications of fibrin glue. Plast Reconstr Surg 1991;88:1005-15.
- Hotz G. Alveolar ridge augmentation with hydroxylapatite using fibrin sealant for fixation. Part I: An experimental study. Int J Oral Maxillofac Surg 1991;20:204-7.
- Hotz G. Alveolar ridge augmentation with hydroxylapatite using fibrin sealant for fixation. Part II: Clinical application. Int J Oral Maxillofac Surg 1991;20:208-13.
- Bonucci E, Marini E, Valdinucci F, Fortunato G. Osteogenic response to hydroxyapatite-fibrin implants in maxillofacial bone defects. Eur J Oral Sci 1997;105:557-61.
- 11. Matras H. Fibrin sealant in maxillofacial surgery. Development and indications. A review of the past 12 years. Facial Plast Surg 1985;2:297-313.
- 12. Matras H. Fibrin seal: The state of the art. J Oral Maxillofac Surg 1985;43:605-11.
- Soffer E, Ouhayoun JP, Anagnostou F. Fibrin sealants and platelet preparations in bone and periodontal healing. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2003;95:521-8.
- 14. Giannobile WV. Periodontal tissue engineering by growth factors. Bone 1996;19 1 Suppl:23S-37.
- Marx RE, Carlson ER, Eichstaedt RM, Schimmele SR, Strauss JE, Georgeff KR. Platelet-rich plasma: Growth factor enhancement for bone grafts. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1998;85:638-46.
- 16. Weibrich G, Kleis WK, Hafner G, Hitzler WE, Wagner W. Comparison of platelet, leukocyte, and growth factor levels in point-of-care platelet-enriched plasma, prepared using a modified Curasan kit, with preparations received from a local blood bank. Clin Oral Implants Res 2003;14:357-62.
- Arpornmaeklong P, Kochel M, Depprich R, Kübler NR, Würzler KK. Influence of platelet-rich plasma (PRP) on osteogenic differentiation of rat bone marrow stromal cells. An *in vitro* study. Int J Oral Maxillofac Surg 2004;33:60-70.
- Carter CA, Jolly DG, Worden CE Sr, Hendren DG, Kane CJ. Platelet-rich plasma gel promotes differentiation and regeneration during equine wound healing. Exp Mol Pathol 2003;74:244-55.
- Yamada Y, Ueda M, Hibi H, Nagasaka T. Translational research for injectable tissue-engineered bone regeneration using mesenchymal stem cells and platelet-rich plasma: From basic research to clinical case study. Cell Transplant 2004;13:343-55.
- 20. Yamada Y, Ueda M, Naiki T, Takahashi M, Hata K, Nagasaka T. Autogenous injectable bone for regeneration with mesenchymal stem cells and platelet-rich plasma: Tissue-engineered bone regeneration. Tissue Eng 2004;10:955-64.
- Wang HL, Avila G. Platelet rich plasma: Myth or reality? Eur J Dent 2007;1:192-4.
- Bennett NT, Schultz GS. Growth factors and wound healing: Biochemical properties of growth factors and their receptors.

Am J Surg 1993;165:728-37.

- 23. Sánchez AR, Sheridan PJ, Kupp LI. Is platelet-rich plasma the perfect enhancement factor? A current review. Int J Oral Maxillofac Implants 2003;18:93-103.
- El-Sharkawy H, Kantarci A, Deady J, Hasturk H, Liu H, Alshahat M, et al. Platelet-rich plasma: Growth factors and pro- and antiinflammatory properties. J Periodontol 2007;78:661-9.
- 25. Fernández-Barbero JE, Galindo-Moreno P, Avila-Ortiz G, Caba O, Sánchez-Fernández E, Wang HL. Flow cytometric and morphological characterization of platelet-rich plasma gel. Clin Oral Implants Res 2006;17:687-93.
- 26. Anitua E. Plasma rich in growth factors: Preliminary results of use in the preparation of future sites for implants. Int J Oral Maxillofac Implants 1999;14:529-35.
- 27. Weibrich G, Kleis WK, Kunz-Kostomanolakis M, Loos AH, Wagner W. Correlation of platelet concentration in platelet-rich plasma to the extraction method, age, sex, and platelet count of the donor. Int J Oral Maxillofac Implants 2001;16:693-9.
- 28. Jakse N, Tangl S, Gilli R, Berghold A, Lorenzoni M, Eskici A, *et al.* Influence of PRP on autogenous sinus grafts. An experimental study on sheep. Clin Oral Implants Res 2003;14:578-83.
- 29. Freymiller EG, Aghaloo TL. Platelet-rich plasma: Ready or not? J Oral Maxillofac Surg 2004;62:484-8.
- 30. Petrungaro PS. Using platelet-rich plasma to accelerate soft tissue maturation in esthetic periodontal surgery. Compend Contin Educ Dent 2001;22:729-32.
- 31. Dodson TT. Is there a role for reconstructive techniques to prevent periodontal defects after third molar surgery? Oral Maxillofac Surg Clin North Am 2007;19:99-104.
- 32. Dohan DM, Choukroun J, Diss A, Dohan SL, Dohan AJ, Mouhgi J, et al. Platelet-rich fibrin (PRI): A second-generation platelet concentrate. Part I: Technological concepts and evolution. Oral Surg Oral Pathol Oral Radiol Endod 2006;101:37-44.
- 33. Ruga E, Gallesio C, Boffano P. Platelet-rich fibrin and piezoelectric surgery: A safe technique for the prevention of periodontal complications in third molar surgery. J Craniofac Surg 2011;22:1951-5.

- 34. Sammartino G, Tia M, Marenzi G, di Lauro AE, D'Agostino E, Claudio PP. Use of autologous platelet-rich plasma (PRP) in periodontal defect treatment after extraction of impacted mandibular third molars. J Oral Maxillofac Surg 2005;63:766-70.
- 35. Karapataki S, Hugoson A, Kugelberg CF. Healing following GTR treatment of bone defects distal to mandibular 2<sup>nd</sup> molars after surgical removal of impacted 3<sup>rd</sup> molars. J Clin Periodontol 2000;27:325-32.
- 36. Wiltfang J, Schlegel KA, Schultze-Mosgau S, Nkenke E, Zimmermann R, Kessler P. Sinus floor augmentation with betatricalciumphosphate (beta-TCP): Does platelet-rich plasma promote its osseous integration and degradation? Clin Oral Implants Res 2003;14:213-8.
- 37. Kassolis JD, Rosen PS, Reynolds MA. Alveolar ridge and sinus augmentation utilizing platelet-rich plasma in combination with freeze-dried bone allograft: Case series. J Periodontol 2000;71:1654-61.
- Butterfield KJ, Bennett J, Gronowicz G, Adams D. Effect of platelet-rich plasma with autogenous bone graft for maxillary sinus augmentation in a rabbit model. J Oral Maxillofac Surg 2005;63:370-6.
- Aghaloo TL, Moy PK, Freymiller EG. Evaluation of plateletrich plasma in combination with anorganic bovine bone in the rabbit cranium: A pilot study. Int J Oral Maxillofac Implants 2004;19:59-65.
- 40. Zechner W, Tangl S, Tepper G, Fürst G, Bernhart T, Haas R, et al. Influence of platelet-rich plasma on osseous healing of dental implants: A histologic and histomorphometric study in minipigs. Int J Oral Maxillofac Implants 2003;18:15-22.
- 41. Choukroun J, Diss A, Simonpieri A, Girard MO, Schoeffler C, Dohan SL, et al. Platelet-rich fibrin (PRF): A second-generation platelet concentrate. Part V: Histologic evaluations of PRF effects on bone allograft maturation in sinus lift. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2006;101:299-303.
- 42. Simonpieri A, Choukroun J, Girard MO, Ouaknine T, Dohan D. Immediate post-extraction implantation: Interest of the PRF. Implantodontie 2004;13:177-89.