

1 **Process Understanding in Freeze-Drying Cycle Development: Applications for Through Vial**

2 **Impedance Spectroscopy (TVIS) in Mini-Pilot Studies**

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25 Abstract

26 The freeze-drying cycle comprises three stages: (1) Freezing, to form ice and to crystallise out any
27 solutes with a propensity to crystallise, (2) Primary drying to remove the ice phase by sublimation,
28 and (3) Secondary drying to remove the remaining unfrozen water which is bound to the remaining
29 matrix of crystalline and amorphous solids. Given the impact of scale on the process outcomes, any
30 freeze drying cycle developed based on mini-pilot studies, will inevitably require measurement
31 technologies for characterising each stage of the cycle at each scale of the process. However, there
32 are inherent challenges in the development of reliable mini-piloting studies, with the first being the
33 fact that no single PAT technology for freeze-drying may be implemented across all levels of scale,
34 and the second being the inherent changes in process characteristics (process parameters that result
35 from scale up). Here we present a new approach for process understanding in freeze-drying cycle
36 development, which uses a through vial impedance measurement to characterise a broad range of
37 features of the process, including, ice onset times, the completion of ice solidification, the glass
38 transition, and the structural relaxation of the amorphous solid, a surrogate for primary drying rate
39 and the primary drying end point. The on-going development of this technology may see the
40 application with micro-titre plate technologies for formulation screening (micro-scale down) and for
41 scale up into production by using a non-contact probes for monitoring problematic regions within
42 the dryer.

43

44 **Key Words:** Freeze drying, in-line process control, PAT, QbD, critical process parameters

45

46 Introduction

47 The scale up of a lyophilisation cycle is challenging due to multiple differences between
48 small scale and large scale dryers. There is some guidance to facilitate the development of an
49 efficient lyophilisation cycle in the laboratory (1), however, even an optimized cycle from a
50 laboratory freeze-dryer may not transfer smoothly to the manufacturing scale. In addition to the
51 scalability differences, there are several other differences between laboratory and manufacturing
52 scale lyophilisers, which may pose a serious challenge for scale-up. These challenges include: (i) Ice-
53 nucleation differences during freezing, (ii) Heat and mass transfer differences, and (iii) Differences in
54 primary drying time. These differences between laboratory and manufacturing scale lyophilisers can
55 pose a serious challenge and therefore a systematic approach is needed to ensure a smooth scale-
56 up.

57 There are a number of stages in a typical lyophilisation process: Freezing (sometimes
58 including annealing) which transforms the liquid solution into a stable frozen matrix, primary drying
59 to remove the ice, and secondary drying to remove residual water from the unfrozen super-cooled
60 liquid domains of the material [1]. Whilst on the surface these stages appear to be somewhat
61 discrete, they in fact constitute a sequential series of inter-dependent events. The process kinetics
62 (ice formation, sublimation, and moisture desorption) are driven by a number of factors, including
63 heat transfer through the vial base and walls (impacted by any thermal heterogeneities in the shelf
64 temperature and radiant heating on the edge vials), the structure of frozen matrix which evolves
65 from the stochastic ice formation process and the initial water content of the interstitial spaces [2].
66 Any attempt to investigate the freeze drying process is complicated by the fact that the process is
67 undertaken in a closed-system, under extremes of temperature and pressure settings, within a batch
68 of vials in close proximity to each other and many (> 10,000) in number, meaning that direct access
69 by any PAT sensors is limited.

70 It is also widely recognised that any attempt to predict product scale process parameters from the
71 laboratory scale is complicated by the fact that the scale of the operation has a significant impact on
72 the process outcomes. There will be variations in the process parameters, and hence the dependent
73 critical quality attributes, that are considered to be a function of the process scale and care should
74 be exercised to avoid developing a cycle at the mini-pilot scale that cannot be translated to the large
75 scale production [3].

76 It is nevertheless the intention within a scale down/scale up approach to use intermediate or
77 benchtop-scale studies to gain product and process knowledge that helps predict the behaviour
78 (scale-up), assess risks (risk analysis), and diagnose production issues (troubleshooting) at
79 production scales. There will inevitably be limitations to what can be achieved but so long as various
80 factors are considered in the design of a meaningful mini-pilot study then one may be able to
81 maximise the relevance of mini-pilot data to the larger scale production process [4]. The factors to
82 be considered have been discussed elsewhere [5] but include:

83 **1. Formulation composition** - For instance, incomplete crystallization of excipients such as mannitol
84 or glycine [6] with consequential heterogeneity in moisture, crystallinity and appearance.

85 **2. The freezing step** as it defines the ice crystal size, which then defines the pore size for sublimation
86 to occur[7]. Variations in freezing rates associated with differences in temperature at locations
87 around the dryer, coupled to the stochastic nature of super-cooling and nucleation, inevitably
88 introduce heterogeneity [8]. Controlled nucleation is showing promise in delivering shorter primary
89 drying times and greater product homogeneity [9]. Mini-piloting studies might therefore aim to
90 simulate the controlled nucleation that is sometimes practised at the larger scale in an attempt to
91 understand the fast kinetics of the process. However, that may require the development and
92 implementation of new forms of measurement system to track those bulk nucleation and growth
93 phases which result in less efficient secondary drying due to lower surface area.

94 **3. Cycle design & –System Capabilities** – Where a cycle is designed at laboratory or pilot level, the
95 process conditions applied cannot always be achieved in the same way within an industrial scale
96 unit, owing to a number of factors: (i) The rate at which vacuum can be applied may be different
97 between a laboratory system and a production model, especially where process routines require oil
98 free (Rootes) pumps; (ii) The nature of the valve between the chamber and the condenser can also
99 differ between small units (where indeed for some lab models the condenser is inside the chamber)
100 and production units; (iii) The design of the valve and its speed of response, as well as length and
101 diameter of vapour duct, must be taken into account as potential sources of variation on scale up
102 and indeed may make pressure rise testing impractical; (iv) Cooling/heating rates of a mini-pilot
103 dryer and a large process scale machine may differ significantly with the latter only being able to
104 achieve low rates of temperature change owing to the thermal inertia of a large dryer; (v) There may
105 be a significant difference between the temperature achieved on one shelf and another of a large
106 stack of shelves or the time taken to achieve a given temperature across a large stack may differ
107 from the one observed for a small single shelf mini-pilot unit; (vi) The smoothness or ‘roughness’ of
108 the finish on the shelves and the flatness of the shelves are factors which must be considered as the
109 creation of a space between the shelf and the base of the vial will influence heat transfer from the
110 shelf to the product. These factors would be difficult if not impossible to model at small scale.
111 Instead, the freeze drying cycle for a production scale dryer may require adjustment in the shelf
112 temperature and chamber pressure in order to achieve the target product temperature.

113 **4. Impact of scale on sublimation rate** The design of a freeze drying cycle for process scale, if based
114 strictly on a mini-pilot data may be too ambitious for the reasons above. Calculation of the heat
115 transfer properties of the freeze drying system, thermal co-efficient of the vial, resistance to vapour
116 flow posed by dry cake, and the gradient in pressure between sublimation front and condenser
117 allow greater predictability to the scale up (1,2,7,8). In primary drying, it is essential that the product
118 temperature is maintained as high as possible in order to maximise sublimative cooling and reduce

119 the primary drying time, while maintaining the product temperature below certain critical
120 temperatures (the eutectic temperature (crystalline) or glass transition (partially or fully amorphous)
121 in order to avoid melt-back or collapse. Engineering factors to consider in process optimization and
122 scale up include: (i) Minimum achievable pressure as a function of sublimation conditions; (ii)
123 Maximum sublimation rates before losing control of pressure or choke flow; (iii) Condenser
124 temperature; as the driving force for sublimation is the pressure gradient due to the pressure at
125 sublimation surface of the product and the pressure at the surface of the ice on the condenser coil;
126 (iv) The vial heat transfer co-efficient- conduction (heat vial contact between the vial base and the
127 shelf); (v) Convection (heat transfer through gas phase); (iii) Radiation (heat transfer from walls,
128 underside of the upper shelves, etc.); (vi) The amount of radiant heat entering through the
129 transparent Perspex door of the freeze dryer, the scale of the shelves and the heat radiated from the
130 walls are all likely to be very different between a mini-pilot dryer and a process dryer while
131 formulation, vial and stopper format remain constant. The impact of shelf, the effect of nearest
132 neighbour interactions and the impact of radiant heat from the shelves above (and hence the inter-
133 shelf distance) are all factors to bear in mind on scale up [10]. The mapping of sublimation
134 heterogeneity across a shelf has been well demonstrated, with the centre of a tray of vials and
135 indeed the centre tray of a series of vials on a shelf, drying more slowly than those at the extremities
136 where external heating effects are greater. Radiation is the dominant mode of heat transfer during
137 lyophilisation [11] and the edge vial experience most radiation, hence the increased product
138 temperature and therefore rate of sublimation for the edge vs centre vials.

139 **Process monitoring and Process Analytical Technology (PAT)** It is clear that the prudent use of
140 process analytical technology (PAT) is a key factor in being able to achieve an effective scalable
141 freeze drying cycle. There are many different technologies for monitoring and even controlling the
142 freeze-drying process, and these have been reviewed elsewhere in a number of comprehensive texts
143 [12-17, 3]. However, these technologies have not been reviewed within the concept of mini-piloting

144 which includes the use of emerging physical characterization techniques that could significantly
145 improve the measurement of process parameters and product quality attributes during drug
146 development and manufacturing that would potentially replace or supplement traditional
147 approaches in the near future.

148 The inevitable question therefore is “which technologies could be used for mini-piloting
149 studies?” Before addressing that question it is worth stating that mini-piloting studies in freeze-
150 drying may take on a number of forms, in terms of the scale length being investigated. Here we
151 define a number of sample and batch scales which could be considered for the concept of a mini-
152 piloting study: 1. Mini-vials and microtitre plates (< 1 ml)¹ in which the size of the sample and
153 container is reduced from that expected of the final product; 2. Single vials (2- 50 ml)² to be used for
154 a fill volume to be used for the final product; or 3. Clusters of these vials (modelling the impact of
155 radiant heat from the walls of the dryer) where the total volume for each study is defined by the
156 number of vials in the cluster multiplied by the fill volume.

157 Mini-vials and microtitre plates have been used to effect for high throughput formulation
158 screening [18-22]. However, due to the fact that the change in sample size in relation to container
159 geometry (wall and base thickness and materials of construction) has a significant impact on the
160 freezing and drying processes, which is currently difficult to model, then such systems may not be
161 considered presently for a mini-pilot application. Moreover, the commercial sample scale is
162 determined by the product requirements and is not therefore considered as a variable in the
163 development process: Once the formulation has been selected (with or without the use of a high
164 throughput methodology), the process is developed based on a predefined container size and fill
165 volume (and hence fill height). Mini-pilot studies can therefore be considered from a starting point
166 of a single vial.

¹ these are not used in production of any commercial product

² these are the vials that are currently available for most commercial products

167 Here we make a definition of cluster size in terms of micro cluster and meso cluster in order
168 in order to recognise the fact that edge effects extend over the outer 3-4 rows of vials in a large
169 cluster, which means that all vials in a micro-cluster will dry as edge vials, whereas a meso-cluster
170 will have some edge vials and some core vials. It is invariably the case that, for the early stage
171 process development, where there is a requirement to minimise the solution volume (and hence
172 drug consumed) the impact of edge effects is removed by placing the product containing vials at the
173 centre of an array of empty vials.

174 Having clarified the definitions and the impact of scale length in terms of numbers of vials, one
175 then can delineate which PAT might be useable for each scale. Table 1 list a number of commercial
176 PATs and research tools that have been defined as being suitable for each of these scale lengths,
177 whereas others have been excluded. As stated above, it could be argued that microplates and mini
178 vials are best suited for initial formulation screening [18] and their use in predicting scale up is quite
179 limited. These systems are therefore excluded from the scope of a mini-pilot study. Given that mini-
180 piloting must start form a single vial, then one might expect that the model single vial systems that
181 have been described and the 7 vial model lyophilizer might be useful in this regard [23]. However,
182 the use of such systems for a mini-pilot study should be used with caution owing to the fact that a
183 single vial (or a micro-cluster of 7 vials) will behave very differently to a vial in the centre of a meso-
184 cluster or a macro-cluster. The impact of radiant heating on the drying characteristics (shape of the
185 drying front and product temperature) are significantly different for a lone vial to a vial within a large
186 cluster.

187 **Table 1** Comparison of analytical technologies for assessment of critical quality attributes and process
 188 parameters. The table includes both commercially available systems and those which are currently available
 189 only in specific research laboratories.

Sample presentation	Stage	Potential PAT	Excluded PATs
Micro-titre plate	Formulation screening	Uv-vis, Raman, fluorescence, TC	Pressure rise, TDLAS
Single vial	Mini-Pilot Scale to Pilot Scale	OCT, TVIS, NIR, Raman, RTD, TC, Microbalance	Pressure rise, TDLAS
Micro-cluster (7, 19,37 vials)	Mini-Pilot Scale to Pilot Scale	<ul style="list-style-type: none"> • TC, RTD, TVIS, smart sensor, • NIR & Microbalance (“Edge” vial only) 	Pressure rise, TDLAS
Meso-cluster (61,91,127,169+ vials)	Pilot Scale	<ul style="list-style-type: none"> • TC, RTD, TVIS • TDLAS & pressure rise (demand a minimum number of vials) • NIR & Microbalance (“Edge” vial only) 	
Macro-cluster (10,000+ vials)	Production	TC, Pressure rise, TDLAS	Microbalance

190
 191 NIR: Near infrared, TC: thermocouple, ideally wireless; TVIS: Through vial impedance spectroscopy, OCT:
 192 Optical computer tomography. RTD: resistance temperature detector.

193
 194 It is clear that each PAT technology is somewhat limited in its application across all scales
 195 within the development cycle and it remains the case that there is no single PAT technology that can
 196 be applied to assess all quality attributes of the product and process parameters of the cycle, at all
 197 levels of scale. There are a number of reasons for that, with each pertaining to the PAT in question
 198 and the target process parameter for that particular technology. Two process parameters (critical
 199 temperatures and drying rates) and one material attribute (glass formation) are used to illustrate
 200 this point.

201 **Critical temperatures** Single vial systems have been developed for the purpose of evaluating other
 202 more novel process analytics, e.g. OCT (optical coherence tomography) measurements of collapse
 203 [19]. These are beneficial as they demonstrate a minimum requirement that the analysis of some
 204 process parameter or material attribute should be conducted, at least within a container and sample
 205 volume that is consistent with that being freeze-dried at the larger scale. This implies that
 206 techniques such as conventional freeze-drying microscopy (FDM) may not accurately define the
 207 critical parameters (temperatures required to control the process), however they are used extremely
 208 frequently and widely in the formulation characterisation process [24].

209 In attempting to drive process efficiencies one tries to maintain the product temperature as
210 high as possible in order to supply the latent heat of sublimation. However this is complicated by the
211 following issues: The product temperature is always lower than the shelf temperature owing to the
212 heat absorbed from the sublimation of ice and the fact that is difficult to measure the temperature
213 at the sublimation interface. TCs placed in the base of the vial can be used to assess the end point of
214 primary drying (for example) but their position at that point precludes the assessment of the
215 temperature at the sublimation interface, which inevitable moves down the vial contents as the
216 drying process progresses. A technique that can measure the product temperature at the ice
217 sublimation front will inevitably provide greater assurance that the temperature at the sublimation
218 interface does not fall below the critical temperature at which the dry layer (in immediate contact
219 with the sublimation interface) does in fact collapse. The implementation of such a PAT tool within
220 the process control loop will inevitably reduce the risk of product failure through over-aggressive
221 drying profiles. In addition, single vial TCs only provide information on thermal events (and the end
222 point of drying) but not the drying rate.

223 **Primary Drying Rates**

224 Pressure rise and TDLAS techniques are used for the measurement of mass flux and drying rates
225 within dryers that are either partially or fully loaded. However, the determination of drying rates as
226 a function of the location of the vial within a cluster is not accessible with these technologies. A
227 commercially available alternative, i.e. the microbalance, can work at the scale of a single vial but
228 only works on an isolated vial or one which is on the edge of a cluster (which inevitably experiences
229 greater radiant wall effects and does not simply rely on heat transfer through the base). Drying rates
230 and dry layer product resistance calculated by this technique therefore should be used with caution
231 when applied to the behaviour of the same materials when freeze-dried within clusters, owing to the
232 significant impact from radiant heating of the side of the glass vial. It is also the case that single vial
233 spectroscopies cannot be used on clusters of vials; any information on product quality (e.g. protein

234 folding by Raman) and water content during drying cannot be easily translated to populations of
235 vials (because the probes are large and will inevitably cause significant disruption to the thermal
236 heat treatment experiences by vials embedded with a manufacturing scale cluster). It is also
237 apparent that existing PATs are limited in their ability to “look inside” the vial with most optical
238 spectroscopy being limited to a surface measurement of 1-2 mm at best. That said, the application
239 of optical spectroscopy for single vial measurements is the subject of renewed interest, given the
240 potential application in the freeze-drying of vials in a continuous process, where such technologies
241 are expected to excel [25-27].

242

243 **The amorphous state (Mesoscopic properties and Glass Formation)**

244 PATs which enable the measurement of mesoscopic properties (i.e. the material properties at the
245 scale length of molecular clusters) such as the glass transition and the fragility/strength of the glass
246 are desired as these properties have significant impact on the product and process efficiency. The
247 formation of the amorphous phase depends to a large degree on the amount of ice that in turn
248 defines the water content of the unfrozen fraction. In addition, the rate at which the amorphous
249 state forms and the temperatures at which the amorphous phase forms will inevitably influence the
250 enthalpy entrapped within the interstitial phases. The power of molecular vibrational spectroscopies
251 has been well-demonstrated [27] These parameters impact the secondary drying phase [28] and
252 even the stability of the glass matrix that forms [29].

253 **Application of PAT across the scales**

254 One key requirement might be that the prospective analytical technology in question has itself
255 a potential for scale up [30]. The principle drawback of mass-flow based PAT techniques is that
256 these display only one temperature value for the entire batch and do not take into account the
257 inter-vial heterogeneities in different locations of the shelf, which are evidenced through individual
258 temperature sensing devices. Generally, the product temperature values obtained from the MTM

259 technique (used for small scale) are believed to be related with the colder region of the shelf that is
260 non-edge or centre of the array and that temperature as low as -45°C during primary drying may be
261 determined with this technique [31]. End point of primary drying is characterized by a sharp drop in
262 vapour pressure of ice. MTM lead product temperature measurement has been fairly representative
263 with first $2/3^{\text{rd}}$ of the primary drying time however after this time heterogeneities in the rates of ice
264 sublimation amongst vials located at different position in the shelf are predominant [32]. Therefore
265 the PAT measured product temperature after this point of time may be non-representative of the
266 actual product temperature [32, 12]. Furthermore the heat transfer rates were misleading when
267 lyophilisation cycles were performed at very low temperatures and low pressure [33] using low solid
268 contents [31] while a minimum sublimation area of 150 cm^2 is required for an accurate MTM
269 product temperature measurement. Lyophilisation of the formulations with high amorphous solid
270 contents were measured inaccurately with MTM, especially in the early phase of primary drying
271 resulting in a high drying temperatures due to re-adsorption of vapours in the dried layer due to
272 pressure rise [12, 34]. Lastly, the closure of MTM valve hinders the sublimation process owing to
273 slowed self-cooling which may sequence to collapse if the freeze drying cycle is operated at
274 temperatures close enough to collapse temperature [12] or if extended isolator valve closure
275 periods are used.

276 TDLAS, another PAT measures the rate of sublimation from the whole batch by recording the
277 light absorption during the passage of vapour through the duct connecting drying chamber and the
278 condenser [35, 36] provided the freeze drier requires has a conducting duct of an appropriate
279 length. This can be used in-process to feedback and control of the freeze drying cycle as in the
280 LyoStar (Virtis) range of dryers.

281 Both MTM and TDLAS have been used to develop a series of models for the freeze drying
282 process and also to build those algorithms into the first smart freeze drying processes that allow
283 control on-line to be achieved either by intervention or automatically [37]. However, the application

284 of these tools in mini-piloting of freeze drying process is somewhat limited given the requirement for
285 a minimum batch size.

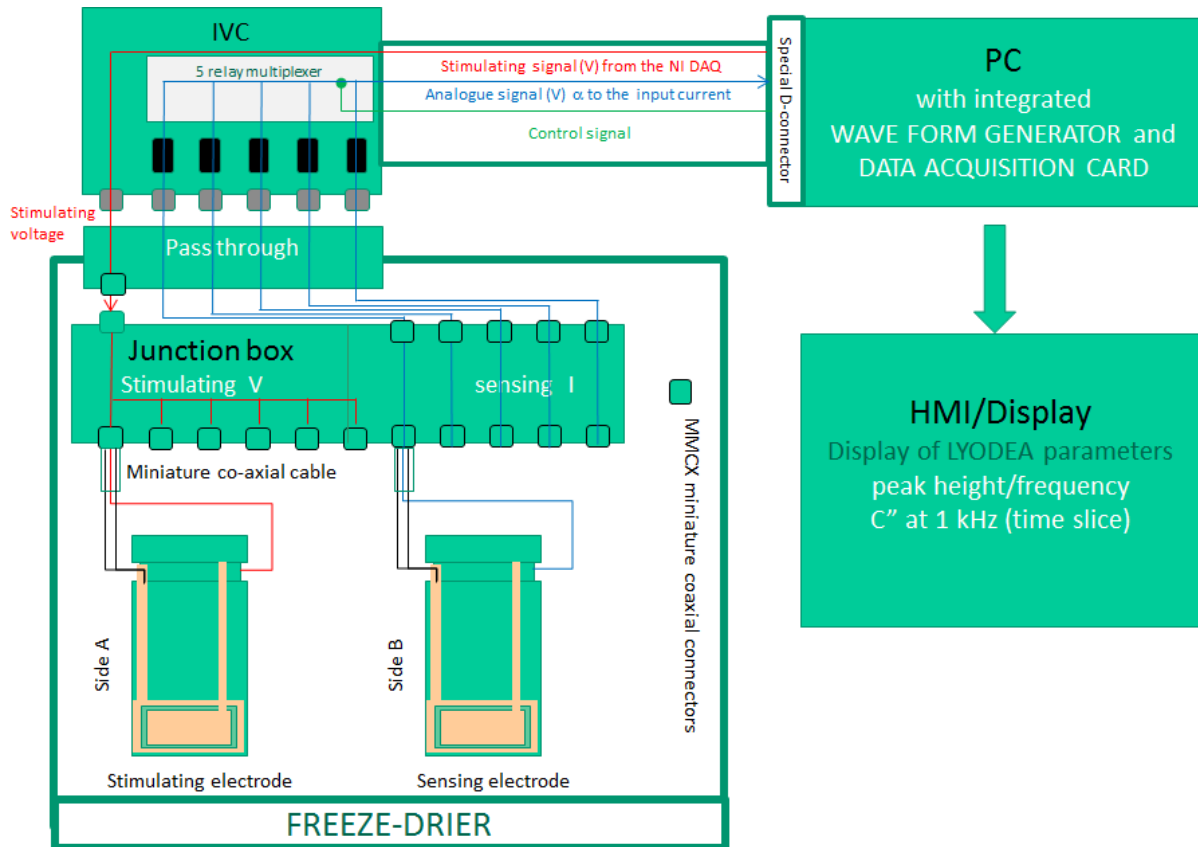
286 As stated above, mini-pilot data may not translate appropriately through scale up unless the
287 PAT technology used to assess the process is also transferrable between scales, and thereby provide
288 an opportunity to unify the PAT signatures at each scale length. For example, in the case of protein
289 formulations (which are sensitive to freeze-concentration stresses) then mini-pilot data from a single
290 vial or micro-cluster of vials may not translate to larger clusters within bigger dryers because the way
291 the sample freezes could impact factors such as aggregation of the active.

292 Techniques which might bridge the gap between the small-scale (e.g. single vial freeze-dryer
293 to the production scale) are limited. One of the few examples are wireless temperature sensors [38]
294 which are an improvement over classical thermocouple or resistance thermometers owing to the
295 lack of wiring, and compatibility with automated loading systems but they are still very much tools
296 used in development, invasive in nature and perturb the ice formation as well as sublimation kinetics
297 . There is an unmet need for a non-invasive technique that can measure both critical events, such as
298 ice formation, glass transition and solidification and collapse, while being able to measure drying
299 rates and end points, and to able to do that across a range of scales from the single vial to multiple
300 vials within clusters, so that the impact of both vial base and radiant wall heating can understood in
301 terms of its impact on process parameters and critical quality attributes. In essence, it is essential
302 that both core and edge vials are assessed for conformity with specification so that the mini-pilot
303 data can be assessed in terms of its direct relevance to production scale and that risks to product
304 quality are understood and mitigated. A more recent PAT technology based on through-vial
305 impedance spectroscopy (TVIS) has been introduced to partly fulfil this need. This technology is non-
306 contacting to the product (unlike an invasive impedance probe in a vial (such as CHRIST's LyoControl
307 technology [39]) and provides some opportunity to characterize material properties across the
308 scales. Albeit in its current form it is a single vial measurement, the opportunity exists to use

309 multiple sensors to track different regions of the dryer and at different scale lengths; and in future
310 the potential development of a non-contact format may allow for such measurements in both scale
311 down application (within mini-vials or micro-wells) and for scale up for multi-vial clusters. The latter
312 is currently the subject of a UK government funded, Innovate UK project called “Biostart”.
313 Theoretical feasibility has been established and a demonstrator unit is currently under development.

314 **Through-Vial Impedance Spectroscopy (TVIS)**

315 Impedance monitoring has a long history as a lyophilisation analysis tool [40, 41]. TVIS
316 measures the electrical impedance of the product, contained within a standard freeze-drying vial
317 that has been modified with electrodes placed on the outside of the glass wall [42]. The impedance
318 measurement vial is connected to a low input-impedance, current to voltage convertor (IVC), via a
319 junction box within the freeze-dryer chamber (mounted close to the shelf on which the vials are
320 located). The signals from the stimulating voltage and that from the resultant current (from the I-to-
321 V convertor) [43] are compared in order to determine the impedance of the measurement vial (and
322 its contents) (Figure 1). The calibration of through vial impedance measurement system is
323 performed by taking in account the impedance contribution at open loop as well as close loop
324 conditions using a reference standard of known capacitance. Details are described in the literature
325 [42].



326

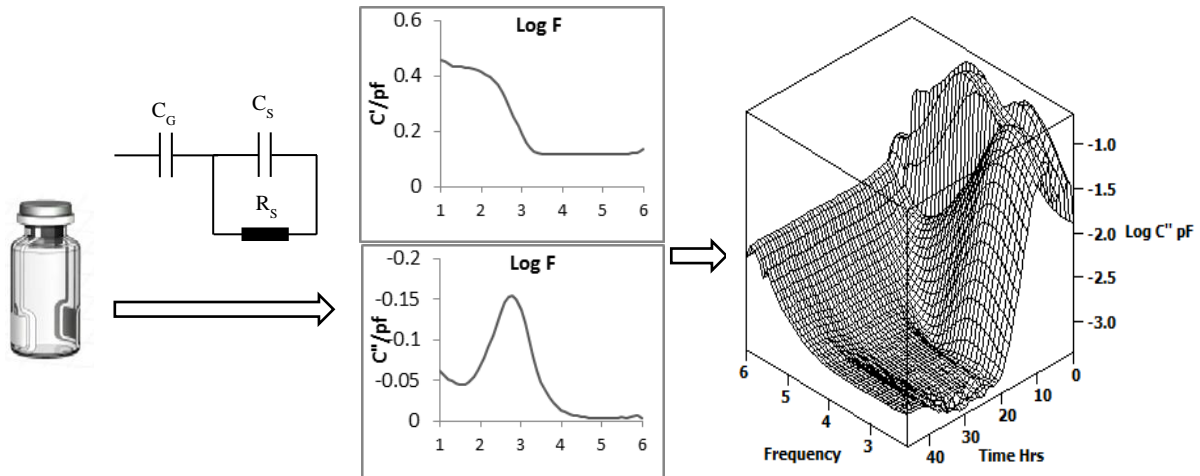
327 **Figure 1** Block diagram of the impedance measurement system. Sides A and B are part of the same vial.

328 The technology combines the function of different single vial PATs (viz. thermocouple and
 329 microbalances) but with a number of advantages: the measurement electrodes are non-intrusive to
 330 the product volume or head space (un-like a thermocouple) which means that the system will not
 331 interfere with the processes of ice nucleation and growth. The measurement hardware has minimal
 332 thermal mass and volume (unlike the microbalance). This minimises the impact on heat transfer
 333 while facilitating measurements on vials which are arranged in the usual hexagonal array (a
 334 requirement for maximising the number of vials loaded into the freeze-dryer). Although the present
 335 design of impedance measurement vials does not support automatic loading, the use of thin foil
 336 electrode makes the tests vials suitable for their placement at any position within the hexagonal
 337 array. This feature, in turn, suggests the potential application of TVIS in spatial mapping of the shelf.
 338 A multichannel (TVIS) instrument design enables the placement of impedance measurement vials at

339 different positions across the shelf which can map the shelf for temperature distributions and
340 variations in the drying rates and to guard against the potential for product collapse.

341 **Electrical Impedance**

342 The electrical impedance of a material determines how easily the material will conduct a current
343 when an alternating voltage is applied to it. Electrical impedance is a function of both the dielectric
344 and conductive properties of the material which are in turn defined by the temperature,
345 composition and physical state of the material contained within the vial (an example TVIS vial is
346 shown in Figure 2i). Changes in these electrical parameters therefore directly mirror the condition of
347 the sample and the progression of the freeze-drying cycle. In order to explain the observed
348 impedance spectrum of the object under test and relate it to the physical properties or changes that
349 may happen during the freeze-drying process, it is necessary to create an appropriate equivalent
350 circuit model. The circuit model (Figure 2 ii) was found to provide an approximate fit to the
351 measured impedance spectrum, where C_G signifies the electrical capacitance of the glass walls of a
352 vial, which is charged through the resistance (R_S) representing the conductivity of the sample, and C_S
353 represents the electrical capacitance of the material within the internal volume of a vial. This imparts
354 a frequency-dependence to the measured dielectric properties, such that the capacitance of the
355 glass wall (C_G) will have sufficient time to charge completely at low frequency, but at high frequency,
356 will not have time to begin to accumulate any of the electrical charge that could otherwise be
357 accommodated.



358

359 **Figure 2** Description of measurement principles; from left to right (i) TVIS measurement vial with external
 360 electrodes attached (in this particular variant there are guard electrodes around each of the measurement
 361 electrodes), (ii) equivalent electrical circuit, with C_G modelling the capacitance of the glass wall of the vial, and
 362 C_S and R_S modelling the capacitance and resistance of the contents of the vial, (iii) individual spectrum (C' vs
 363 log frequency is the real part spectrum, C'' vs log f is the imaginary part spectrum) where the frequency of the
 364 peak in the imaginary capacitance is given by $f = 1/2\pi R_S(C_G + C_S)$ (Note that this particular spectrum is taken
 365 after freezing the sample), and (iv) response surface plot of imaginary capacitance, resulting from measurements
 366 at a range of temperatures. The peak at position A in the early stages of the cycle shows the condition of the
 367 sample in the liquid state. The peak shifts to position B (lower frequencies) when the sample freezes and the
 368 product resistance increases by a factor of 100-1000. The decrease in the peak height over time is a consequence
 369 of the loss of ice on sublimation during the primary drying phase. The wing at low frequency (shaded area D
 370 delineated by the dotted line) is more than likely to be due to the additional distributed element characteristics of
 371 the glass wall.

372 The overall result is that the capacitance spectrum of the material under test (i.e. glass vial, its
 373 contents, and the electrical connections to the vial) will display a step-like decrease in capacitance as
 374 the frequency is increased through that critical frequency which corresponds to the time constant
 375 for the sample ($f = 1/2\pi\tau$, where $\tau = R_S(C_G + C_S)$) (Figure 2 iii top). There is a corresponding peak in
 376 the associated imaginary capacitance spectrum as the material under test starts to conduct
 377 electricity through the phase lag between the response of the sample and the applied electric field
 378 (Figure 2 iii bottom). The step in the real part capacitance and the peak in the imaginary capacitance
 379 are the manifestation of what is known as an interfacial-relaxation process. It is a consequence of
 380 the time dependence of the accumulation of charge at the glass surface as ions migrate through the
 381 liquid (or solid) contained within the glass vial, following the application of an external field [44].

382 It is the characteristics of this process that are used to 'follow' the progression of the freeze-drying
 383 cycle. More specifically, it is the peak frequency and peak value for the imaginary capacitance (which

384 can be considered as the magnitude of the interfacial-relaxation process) that is used to monitor the
 385 freeze-drying cycle. Figure 2iv shows a typical surface plot of the imaginary capacitance as a function
 386 of frequency and time, during the entire freeze-drying cycle. There are characteristic shifts in the
 387 relaxation frequency and change in the peak height as the temperature of the sample changes and
 388 when the material undergoes a phase change (e.g. liquid to ice) [45]. There is then a dramatic
 389 decrease in the magnitude of the interfacial-relaxation peak as ice is removed from the sample.
 390 Factors such as salt content, buffers and tissue culture medium will increase the conductivity and
 391 shift the relaxation peak to the higher frequency end of the experimental frequency window.

392 The impedance of the object under test (namely the glass vial and its contents) can be calculated
 393 from the following equation

$$394 \quad Z^* = \frac{1}{i\omega C^*} = \frac{1}{i\omega C_G} + \frac{1}{\frac{1}{R_S} + i\omega C_S} \quad (1)$$

395 From the complex impedance formula, the expressions for real and imaginary capacitance can be
 396 calculated to explain the origin of interfacial polarization peak

$$397 \quad Z^* = \frac{1}{i\omega C_G} + \frac{R}{1+i\omega R_S C_S} = \frac{1+i\omega R(C_S+C_G)}{i\omega C_G - \omega^2 R_S C_S C_G} \quad (2)$$

$$398 \quad C^* = \frac{1}{i\omega Z^*} = \frac{C_G + i\omega R_S C_S C_G}{1+i\omega R_S(C_S+C_G)} \quad (3)$$

399 By multiplying nominator and denominator by the complex conjugate of denominator

$$400 \quad C^* = \frac{1}{i\omega Z^*} = \frac{(C_G + i\omega R_S C_S C_G)(1 - i\omega R_S(C_S + C_G))}{(1 + i\omega R_S(C_S + C_G))(1 - i\omega R_S(C_S + C_G))} \quad (4)$$

$$401 \quad = \frac{C_G + \omega^2 R_S^2 C_S C_G (C_S + C_G) - i\omega R_S C_G^2}{1 + (\omega R_S(C_S + C_G))^2} \quad (5)$$

402 and grouping the real and imaginary members decomposes C^* into its real C' and imaginary parts

$$403 \quad C' = \frac{C_G + \omega^2 R_S^2 C_S C_G (C_S + C_G)}{1 + (\omega R_S(C_S + C_G))^2} \quad (6)$$

404 and

405
$$C'' = -\frac{\omega R_S C_G^2}{1 + (\omega R_S (C_S + C_G))^2} \quad (7)$$

406 For an example spectrum (Fig. 2) at $\omega \rightarrow 0$, $C'' = 0$.

407 As the frequency is increased, C'' increases to a maximum of

408
$$C''_{max} = \frac{C_G^2}{2(C_S + C_G)} \quad (8)$$

409 at a frequency of

410
$$\omega_{max} = \frac{1}{R(C_S + C_G)}. \quad (9)$$

411 and then decreases to 0 as the frequency $\omega \rightarrow \infty$

412 The value of the real part of capacitance at $\omega \rightarrow 0$ is $C' = C_G$

413 and the value at $\omega \rightarrow \infty$ is

414
$$C' = \frac{C_S C_G}{(C_S + C_G)}. \quad (10)$$

415 It follows that the step change in capacitance is

416
$$\Delta C' = C_G - \frac{C_S C_G}{(C_S + C_G)}, \text{ or } \Delta C' = \frac{C_G^2}{(C_S + C_G)}. \quad (11)$$

417

418 **Measurement of sublimation rate and end of primary drying**

419 The basic assumption is that the capacitance is a function of the amount of ice in the measurement
 420 vial and provides the rationale for the application of TVIS as a determinant of ice sublimation rate
 421 during primary drying. It has been demonstrated that TVIS can be used to measure the onset of
 422 primary drying, rate of ice sublimation and end of primary drying [42].

423 Equation 12 (and the explanation that follows) demonstrates how the magnitude of the peak in the
 424 imaginary part capacitance, during primary drying, provides an assessment of the remaining ice and

425 thereby an opportunity to assess relative drying rates in vials as the process is scaled-up from the
 426 mini-piloting study (e.g. on an individual vial or a small cluster of vials).

$$427 \quad C''_{max} = \frac{C_G^2}{2(C_S + C_G)} \quad (12)$$

428 The magnitude of each lumped circuit element ($C_S + C_G$) is proportional to the cell constant for that
 429 element, i.e.

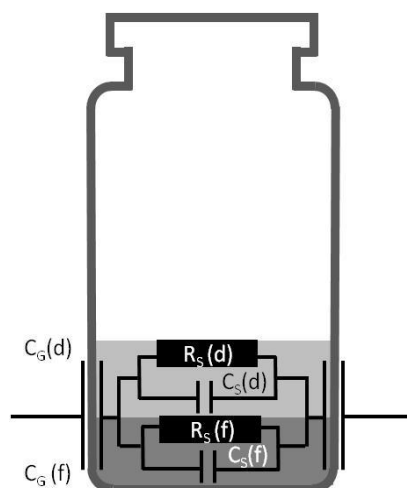
$$430 \quad C_G = \epsilon_G \epsilon_0 \frac{A}{d} \quad (13)$$

431 and

$$432 \quad C_S = \epsilon_S \epsilon_0 \frac{A}{d} \quad (14)$$

433 where A is the area of interface between the frozen mass and the glass adjacent to the electrode.
 434 Provided the sublimation interface is flat then as the ice is removed from the sample and the
 435 sublimation front recedes down the vial, then the interfacial area between the frozen layer and the
 436 juxtaposed glass wall (A) will decrease in proportion to the remaining ice volume. An electrical
 437 model of the drying process is given in Figure 3, in which a dry-layer impedance is incorporated into
 438 the overall impedance of the system. In reality, the impedance of this layer can be ignored given that
 439 the time constant for charging the segment of glass in proximity to the dry layer is very large, owing
 440 to the high resistance of the dried solid, and the only contribution made will be a small contribution
 441 to the real part capacitance.

442



443

444 **Figure 3** Equivalent circuit model of a two layer system, with a dry layer above an ice-rich layer. Here the
 445 model shows an external electrode system that is of a height such that the fill volume extends both above the top
 446 and below the base of the electrode.

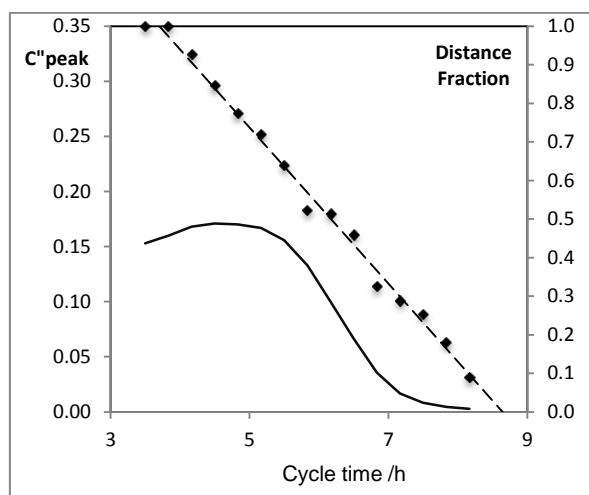
447 The schematic in Figure 3 illustrates how the measured capacitance of the glass and the frozen
 448 solution will change on drying. For a linear drying rate (in time) and for a flat sublimation interface,
 449 both C_S and C_G would be expected to decrease in a linear response. However, given the fact that the
 450 electrodes do not span the full height of the liquid one then needs to consider the impact of the
 451 height of the frozen layer in relation to the electrode height on the measured response. It follows
 452 that one can expect a greater sensitivity to the frozen layer in the middle/centre region of the
 453 electrode than in the regions above and below the electrode. This feature of the measurement
 454 system in its current form (with electrodes on the outside of the vial) partly explains why one
 455 observes the sigmoidal decrease in the magnitude of the peak as the ice sublimates from the sample
 456 (Figure 4).

457 To explore this observation further one needs to consider the fill volume/height in relation to the
 458 vial size and electrode geometry being used (Although we might add that, in the classical scale up
 459 approach in lyophilisation, one would keep the fill height the same but increase surface area in order
 460 to increase drying rates). The theoretical impact of this change in vial geometry in relation to the
 461 electrode geometry was explored in a previous publication, in which it was predicted that the

462 relaxation frequency might increase by up to a factor of two if 2 mL vials are used instead of the 10
463 mL vials we have used in our work [45].

464 In our case, the electrode height in the current presentation of the TVIS measurement system is 5
465 mm, which means that for a fill volume of 3 ml the liquid height is ~9.5 mm. The electrode is placed
466 in such a way that it is 1-2 mm above the vial base (measured externally) and is 2-3mm below the fill
467 height [45]. If there is a 1-2 mm gap (i.e. ice layer) below the electrode then there will be 2.5 to 3.5
468 mm of ice layer above the electrode. It follows that the response of the TVIS spectrum to a reduction
469 in the height of the ice layer won't be registered until $(2.5-3.5)/9.5$ (i.e. 25 to 35%) of the ice is
470 removed. The data generated by the TVIS system supports this observation (Figure 5) which shows
471 that the TVIS system only begins to register the loss of ice when the ice has reduced to
472 approximately 20%. Thereafter, the TVIS system senses a linear decrease in the magnitude of the
473 peak after approximately 40% of the ice has been removed.

474 Extrapolation of the linear portion of the C''_{peak} vs time plot, to the start of the primary drying phase,
475 suggests that a surrogate drying rate may be determined from the imaginary capacitance alone,
476 thereby facilitating comparison between vials placed in different position in the dryer.



477

478 **Figure 4** Capacitance profile of 3% w/v lactose during primary drying; the solid line is C''_{peak} vs primary drying
 479 as measured by the TVIS system. The symbols and the long dashed line is the loss of ice as determined from a
 480 visual assessment of the ice layer from the outside of the vial. The short dotted lines on the linear region of the
 481 C''_{peak} plot is the surrogate drying rate determined by the TVIS system.

482 It remains to be seen whether any general rules may be developed which enables the drying rate of
 483 a range of formulations to be extracted from TVIS measurements. For example, what is unknown at
 484 present is the amount of ice that has formed. This uncertainty may be removed by calculating the
 485 water content of the unfrozen layer from a measurement of the glass transition temperature (see
 486 following section) and the application of the Gordon-Taylor equation. A more straightforward
 487 application of this methodology would be to use it to evaluate the heat transfer coefficient (K_v) at
 488 the base of the vial from a sublimation experiment using water (rather than the product solution).
 489 Other parameters to be recorded, in order to achieve a clear evidence of the sublimation rate,
 490 include the measurement of temperature at the vial bottom and cake resistance to vapour flow. In
 491 that case the amount of ice is known as it is the same as the amount of water that is added to the
 492 vial [46].

493 **Examples of the use of TVIS in mini-piloting (from single to multiple vials) to measure temperature**
 494 **and to characterise critical temperatures and transitions**

495 In the sections that follow a number of applications for TVIS have been described in order to
 496 demonstrate the versatility of the technique in the characterization of first and second order phase

497 transitions (ice formation, eutectic formation and suppression, glass transitions and phase
498 separation), product temperature, and product collapse.

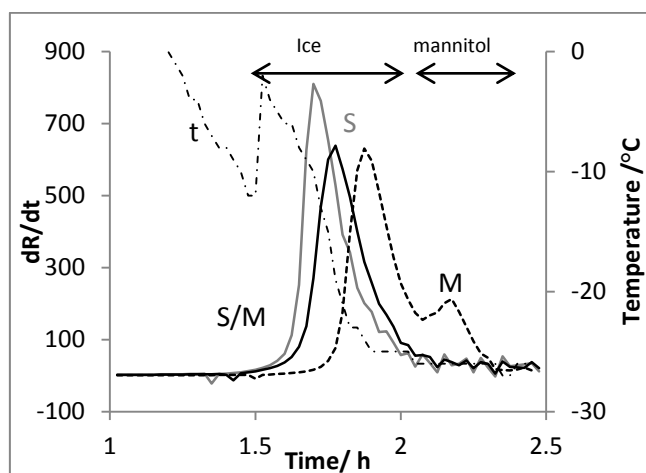
499 In the majority of the following applications, the sensitivity of the TVIS response surface to changes
500 in the resistance of the fill solution have been exploited to determine changes in state (e.g. liquid to
501 solid) and the temperature of the solution (whether liquid or frozen). From equation 9 (re-stated
502 here)

$$503 \quad \omega_{max} = \frac{1}{R(C_S + C_G)} \quad (9)$$

504 one can see the impact of a phase change, which increases the resistance of the sample by a factor
505 of 100-1000 whereas the capacitance will only change by a factor of 25% at best (e.g. 80 to 100). The
506 peak frequency is therefore strongly dependent on the sample resistance.

507 **Measurement of Eutectic crystallization**

508 Classically, eutectic crystallization of an excipient in a formulation is detected using off-line DSC
509 studies, however, until recently there were no techniques capable of recording the manifestation of
510 this crystallization process in-line and therefore it is unclear whether there is a need to include an
511 annealing stage in the drying cycle. The impedance measurements recorded from a surrogate
512 formulation containing mannitol demonstrates a secondary peak in the derivative of the resistance
513 profile at ~-22 °C which was in close agreement with the eutectic crystallization of mannitol as
514 determined by DSC (Figure 5).



515

516 **Figure 5** The electrical resistance (dR/dt) profile during freezing of mannitol solution (M), sucrose (S) and a
 517 50:50 mixture of mannitol and sucrose (S/M)

518 During this study, the impact of a non-crystallizing solid (sucrose) was also shown to suppress the
 519 crystallizing behaviour of mannitol [47].

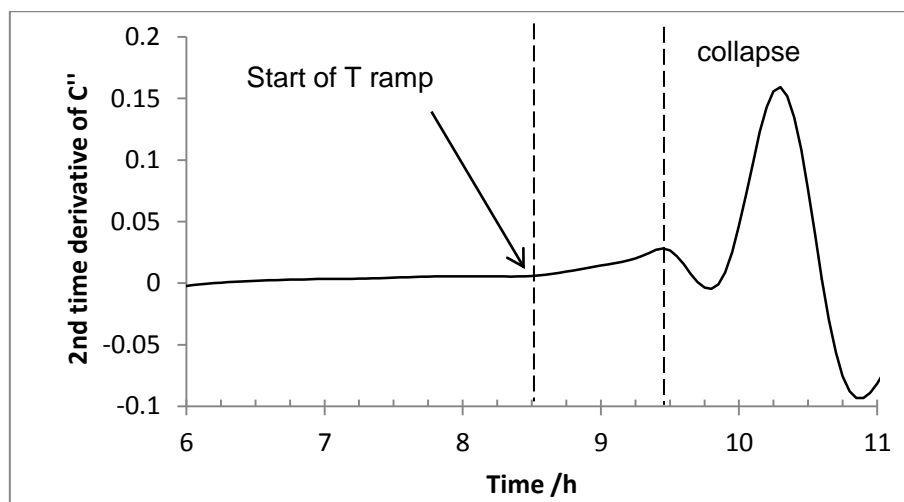
520 **Measurement of product collapse**

521 It is well known fact that the viscosity of a formulation decreases as the temperature increases
 522 above glass transition T_g' . At some temperature exceeding T_g' , the viscosity of the frozen formulation
 523 is insufficient to hold its own weight and the product collapses; the corresponding temperature is
 524 called collapse temperature (T_c) [48]. The measurement of structural collapse in a product during
 525 primary drying remains a challenge for the formulation scientist. In essence, it may provide a realistic
 526 value of temperature that defines the boundary of a design space.

527 Conventionally, T_c is measured by freeze drying microscopy which measures the drying front of small
 528 sample positioned at the temperature controlled freeze drying stage under vacuum [24]. This
 529 temperature is then considered as the upper temperature limit for the primary drying stage.
 530 Recently, Mujat and co-workers used an optical coherence tomography based freeze drying
 531 microscopy (OCT FDM) to record collapse temperature of a surrogate formulation [25, 49]. The
 532 technology is advantageous as it records in-vial measurements of product collapse within a bespoke
 533 design of a single vial freeze dryer. The results from this study indicate that T_c measured by OCT FDM

534 was $\sim 3^{\circ}\text{C}$ higher than the one measured by conventional FD microscopy suggesting a higher primary
 535 drying temperature is permissible that could reduce the primary drying time by up to 30%. Although
 536 the OCT FDM system measures the response of the frozen solution within the glass vial, there are
 537 certain limitations such as: (i) only part of the formulation in direct contact with the probe is
 538 measured, (ii) the presence of the probe may perturb the ice structure and (iii) it is not possible to
 539 use the measurement system on vials placed within the usual configuration of a hexagonal array.

540 In a previous study [44], impedance spectroscopy has been used to record a sudden change in the
 541 capacitance (Fig. 6) which is associated with the macroscopic structural collapse of product (as
 542 measured by photographic images). This observation suggests a useful application of the TVIS in
 543 recording a failure mode.

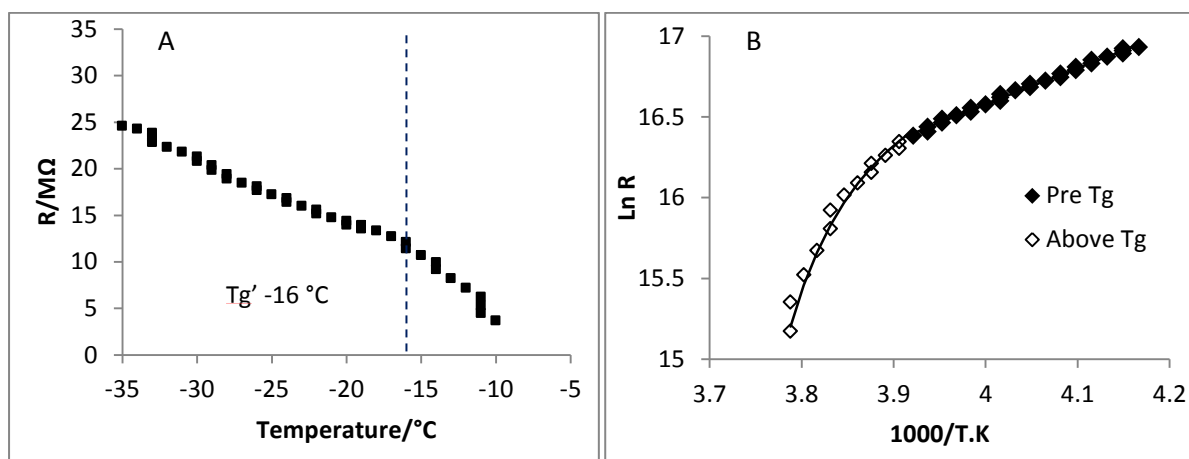


544

545 **Figure 6** Demonstration of collapse in 3% solution of sucrose as the shelf temperature is increased during the
 546 primary drying phase.

547 **Measurement of the glass transition**

548 TVIS has been reported as a direct measurement approach which effectively measures in-vial glass
 549 transition temperatures during re-heating post freezing (Figure 7)[47].



550

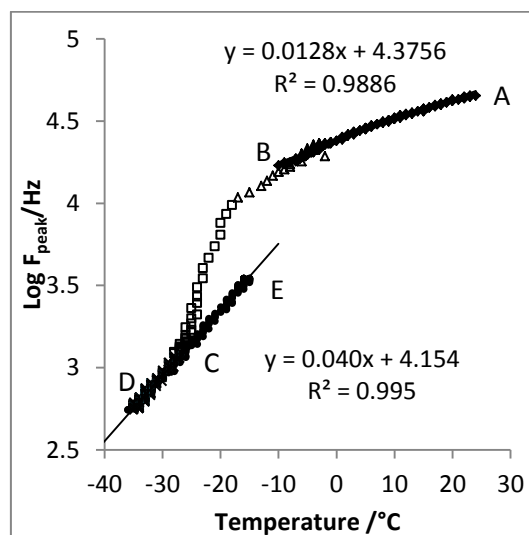
551 **Figure 7** Electrical impedance profile of 10% w/v maltodextrin DE16-19 during re-heating. A) Temperature
 552 profile of equivalent circuit parameter R showing an inflection at the glass transition temperature of $-16\text{ }^{\circ}\text{C}$; B)
 553 Arrhenius plot of $\ln R$ vs $1000/T$ showing Arrhenius behaviour below T_G and non-Arrhenius behaviour above
 554 T_G (the solid line is the VTF model).

555 The additional benefit of this approach is that the impedance data can be modelled with a modified
 556 VTF (Vogel-Tammann-Fulcher) equation which can be employed to calculate the fragility of the
 557 frozen glass. This observation refers to an additional application of TVIS for formulation screening
 558 (especially for micro-plate/min-vial scale) and the potential for validation of formulation behaviour
 559 when transferred to into the process development stage (mini-piloting to scale-up). This parameter
 560 is likely to find application in determining the changes in the strength of the frozen glass following
 561 annealing which might impact the stability of the product. Such information may also provide
 562 additional information on the rationale for the inclusion of annealing step in the freeze drying cycle
 563 [50].

564 Temperature measurements

565 Figure 8 shows that there are correlations between $\log F_{\text{peak}}$ and product temperature during
 566 product cooling in the liquid state (A to B) and the solid state (C to D) and on annealing (D to E). The
 567 temperature coefficient for $\log F_{\text{peak}}$ in the frozen state (D to E) is ~ 0.04 which is approx. x3 of the
 568 temperature coefficient in the solution state.

569 By fitting the equivalent circuit in Figure 2ii to the calibration model data in the region D to E, it is
 570 then possible to estimate the impact that a temperature gradient, within the vial (from the base to
 571 the top of the ice layer) will have on the spectra acquired during primary drying.

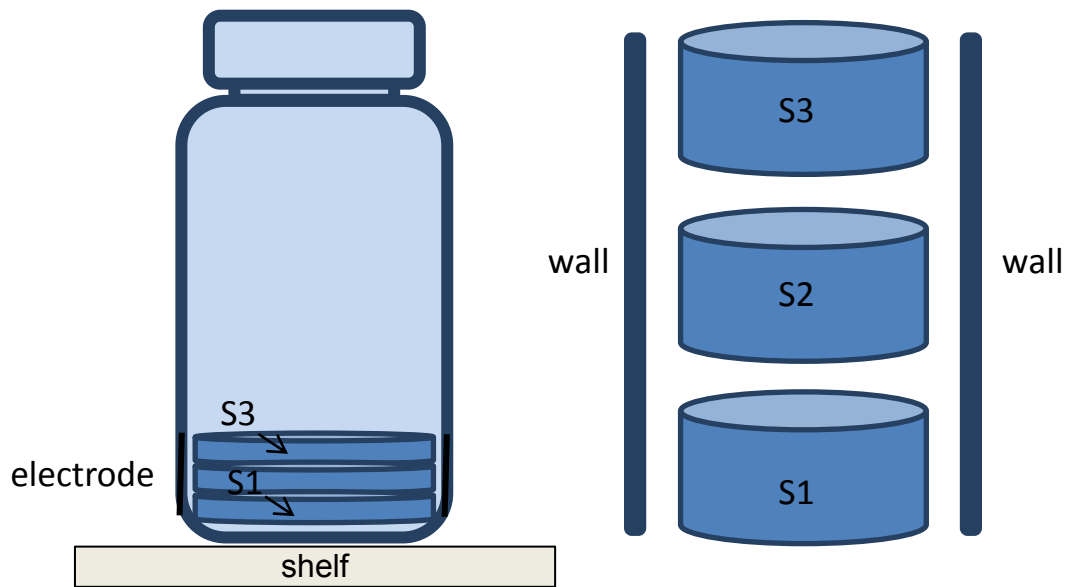


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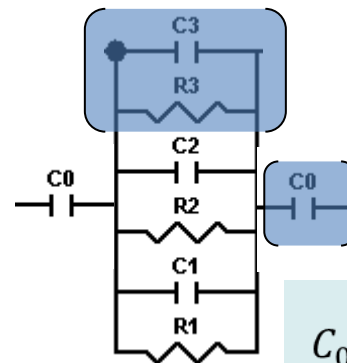
573 **Figure 8** Temperature calibration of the TVIS instrument, the relationship between $\log F_{\text{peak}}$ and temperature is
 574 linear, both in the liquid state (A to B) and the frozen state (C to D to E). Fitting the equivalent circulate model
 575 to the data in the region C to D to E provides an opportunity to create a model for the scenario when there may
 576 be differences in temperature between the top and bottom of the ice layer during primary drying.

577 To this end an equivalent circuit model was built comprising a number of horizontal segments
 578 (Figure 10) with each segment comprising a parallel combination of a capacitor and a resistor.
 579 Estimates for each element were taken from the calibration data represented in Figure 9, having
 580 taken into account the fact that the cell constants for each segment were now a fraction of the cell
 581 constant for the entire volume of the sample (in the case of the capacitive element) and a multiple
 582 of the cell constant for the entire volume of the sample (in the case of the resistive element).

583



$$C_n^*(i\omega) = \frac{1}{R_n} + i\omega C_n \quad (n = 1 \text{ to } 3)$$



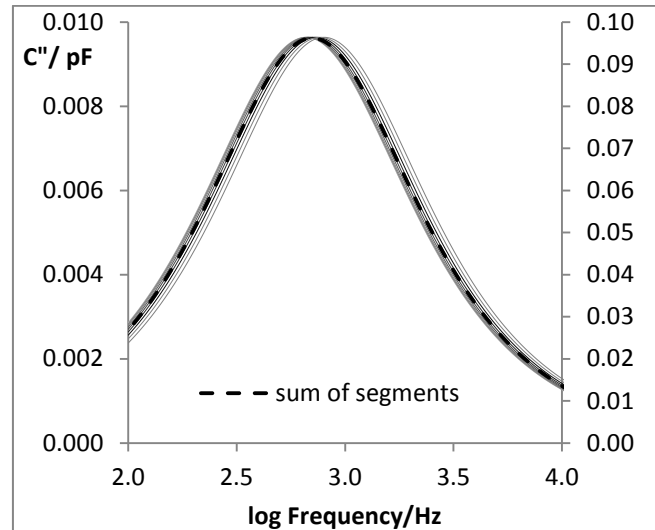
$$C_0^*(i\omega) = \frac{1}{i\omega C_0}$$

584

585

586 **Figure 9** Illustration of how to create a distributed circuit model to account for temperature differences between the top and bottom layers of the frozen solution. Here three
 587 horizontal segments have been used to model a difference in temperature. In each segment the values for the lumped circuit elements have been taken from the calibration
 588 model in the frozen state (Data from region C to E

589



590

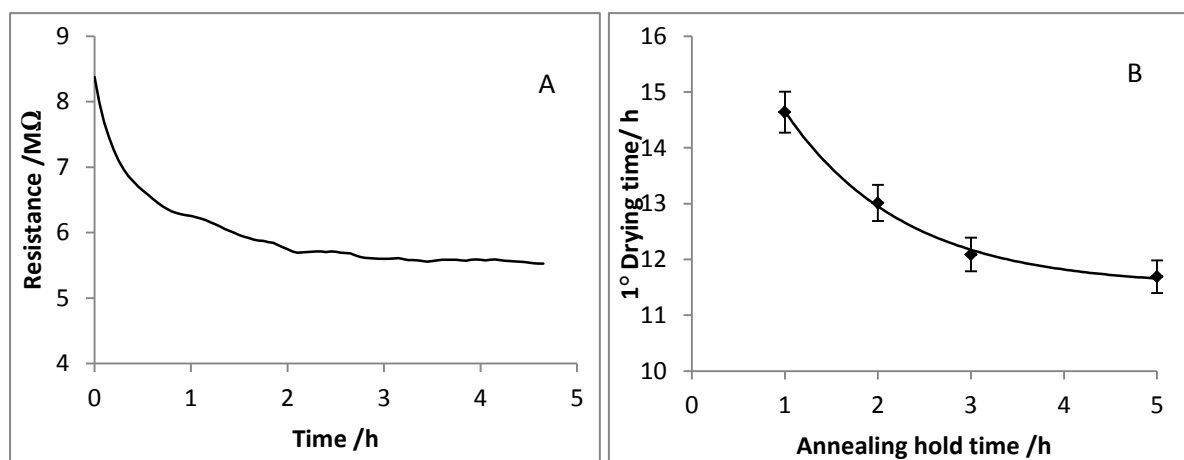
591 **Figure 10** Predicted response from the equivalent circuit model shown in **Figure 9** (here the solution is divided
 592 into 10 horizontal segments rather than the three shown in **Figure 9**). The left hand scale shows the predicted
 593 spectrum of each element, and the right hand scale shows the predicted spectrum when all elements are added
 594 together. The left scale has been magnified by a factor to 10 so that the magnitude of the individual spectra
 595 coincides with the magnitude of the summed spectra.

596 Results from the model (Figure 10) suggests that the shape of the spectrum does not change when
 597 there is a distribution of temperatures across the frozen layer and that the peak frequency provides
 598 an indication of the mean temperature of the frozen mass (dashed line on Figure 10). The mean
 599 temperature may in itself be usefully employed as the driver to set the shelf temperature in a
 600 process control scenario. In the development cycle, one might instead want to include a
 601 thermocouple in the base of the vial to measure the base temperature and then use the TVIS
 602 derived mean temperature to predict the temperature at the sublimation interface. That would
 603 require the assumption that the profile across the frozen layer was linear.

604 **Characterization of Annealing**

605 The inclusion of annealing step increases mass transfer rates during primary drying [51] as it
 606 overcomes ice crystal heterogeneities which arise from uncontrolled freezing a stochastic process,
 607 by promoting the growth of large ice crystals which in turn reduces the dry later resistance. An
 608 annealing step is also included to promote crystallization of bulking agent such as mannitol. TVIS has

609 been used to improve our understanding of how drying rate changes with the annealing hold time
 610 and temperature (Fig. 11) [52].



611

612 **Figure 11** A) Resistance profile of maltodextrin 10% w/v during annealing hold time B) Impact of annealing
 613 hold time on the drying time

614 By recording the changes in electrical impedance of the formulation during annealing as well as the
 615 primary drying stage of the freeze drying cycle, it was possible to demonstrate that Ostwald ripening
 616 was the primary mechanism responsible for faster drying rate. Devitrification, the other mechanism
 617 in question, was ruled out as there was no significant amount of additional ice formation recorded
 618 as glass transition temperature was not increased following annealing.

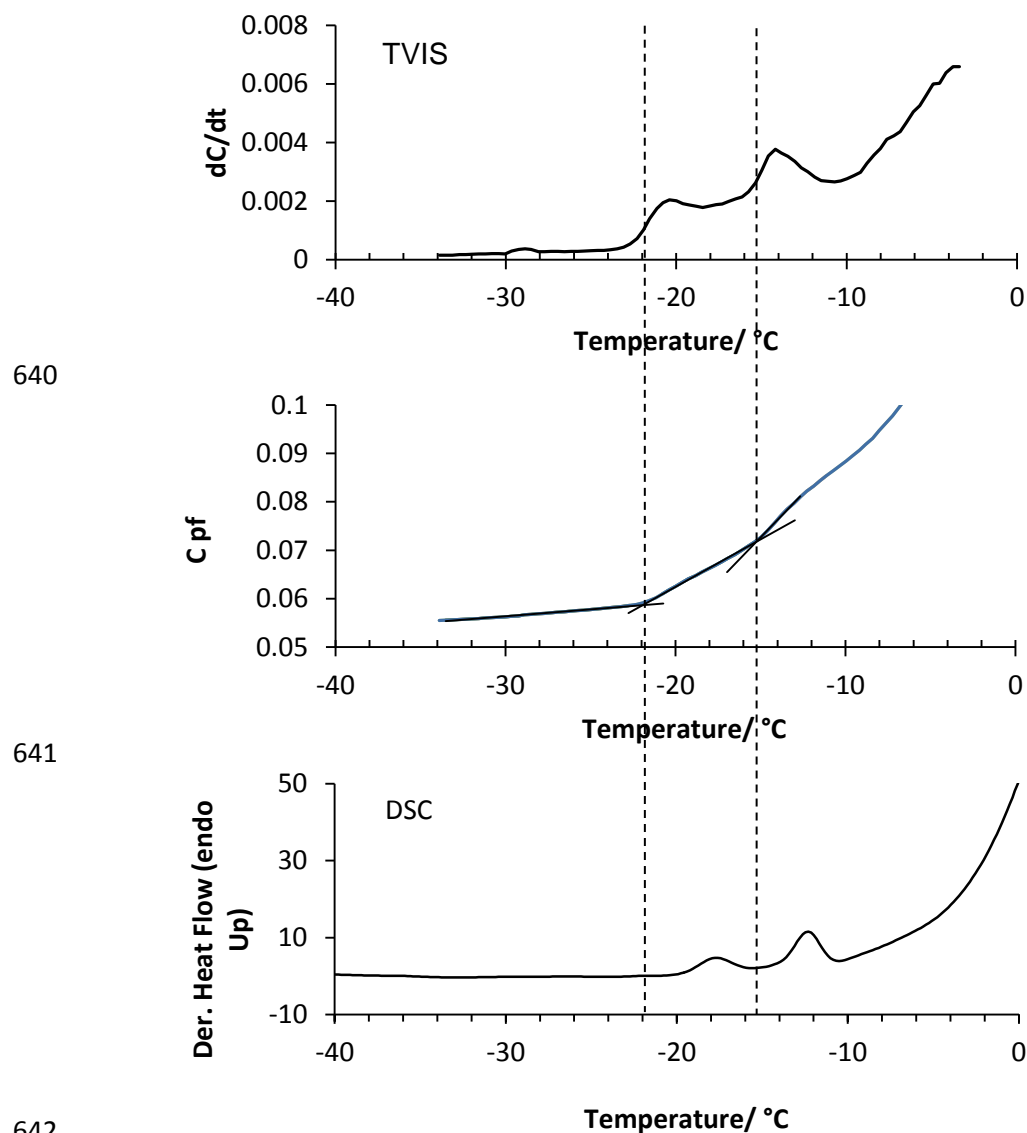
619 The crystal growth occurs at exponential rates and this phenomenon is almost complete after 3h.
 620 Any extension to this hold time in excess of 3 h is largely unjustified as the sublimation rate does not
 621 increase accordingly and these observations are in agreement with the R profile during annealing
 622 which records asymptote after an annealing hold time of 3h.

623 Phase separation

624 In certain cases, the formulation components exhibit physical incompatibility in that different phases
 625 separate out from the solution during freezing and are subsequently dried as separate layers. In
 626 order to demonstrate the potential use for through vial impedance measurement in the
 627 determination of phase separation, a binary solution of 14% w/w dextran (MW 9-11000) (Sigma) and

628 14% w/w polyvinylpyrrolidone PVP K10 (Sigma) was analysed over a frequency range 100Hz-1MHz at
629 scan interval of 0.5 min^{-1} throughout the following freezing cycle: Temperature ramp to $-35 \text{ }^\circ\text{C}$ in 60
630 minutes, hold then at $-35 \text{ }^\circ\text{C}$ for 120 minutes and temperature ramp up from $-35 \text{ }^\circ\text{C}$ to $25 \text{ }^\circ\text{C}$ in 60
631 minutes using a HETO FD 08 freeze dryer. The product temperature was also recorded in a
632 neighbouring vial using a type K thermocouple. Thermal analysis of the formulation was also
633 performed by differential scanning calorimetry, scanning over the same temperature range.

634 After fitting the impedance model in equation 1 to each spectrum, the time derivative of values of
635 the element R_s was seen to undergo a non-linearity with temperature, which was a direct
636 consequence of the sample passing through a glass transition. Time derivatives of the R_s parameter
637 provided estimates for T_g' of $-13 \text{ }^\circ\text{C}$ and $-24 \text{ }^\circ\text{C}$, whereas the T_g' estimates from DSC are $-13 \text{ }^\circ\text{C}$ and -
638 $19.5 \text{ }^\circ\text{C}$ (Figure 12) the latter was in agreement with T_g' of individual components reported the
639 literature [53, 54].



643 **Figure 12** Time derivative and imaginary capacitance of 14% w/w dextran and 14% w/w PVP (b) DSC scan
 644 during re-heating from -35 to 25 °C

645 The close agreement between the two values for the T_g' of the dextran phase (at -13 °C) points to
 646 the reliability of the new method, whereas the disagreement between the two estimates for the
 647 second T_g' may point to real differences in the composition of the PVP phase.

648 **Conclusions**

649 We have reviewed the advantage and limitation of the various available PAT technologies, such as
 650 thermocouples, TDLAS and MTM, commonly used to monitor the freeze drying process. The
 651 translation of process understanding from the mini-pilot to scale up and production demands a new

652 PAT method that can bridge these scales and provide the verification that the process parameters at
653 one scale can be achieved at another. TVIS provides part of that solution on the basis of a two
654 parameter measurement: The first parameter is the measurement of the magnitude of the
655 interfacial relaxation process which has been shown to provide a convenient non-invasive (albeit
656 single vial) measurement of drying rates. The second is the frequency position of the interfacial
657 relaxation which is sensitive to both temperature and phase behaviour. The most important critical
658 product parameter is the product temperature at the ice sublimation interface, T_p , as it defines the
659 efficiency of the process in terms of the rate of drying. Here we demonstrate a possible
660 methodology for determining simultaneously the critical process parameter and the impact it has on
661 drying rates. In future, with TVIS capability embedded in different scales or dryer, it should be
662 possible to track the product temperature and drying rate vs. time profile so that the influence of
663 process- and equipment-related differences may be first understood and then compensated for, and
664 a dynamic cycle developed which adapts the shelf inlet temperature and chamber pressure to
665 maintain this profile. Other capabilities of the TVIS method in the determination of the glass
666 transition and the strength/fragility of the unfrozen phase have been highlighted and suggestions
667 provided as to the relevance these parameters have in assessment of product stability.

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