PROCEEDINGS B

Research

THE ROYAL SOCIETY
PUBLISHING

The evolutionary dynamics of the early Palaeozoic marine biodiversity accumulation

Björn Kröger[1], Franziska Franeck[2] and Christian M. Ø. Rasmussen[3,4]

[1]Finnish Museum of Natural History, University of Helsinki, Helsinki, Finland
[2]Natural History Museum, University of Oslo, Oslo, Norway
[3]Natural History Museum of Denmark, University of Copenhagen, Copenhagen, Denmark
[4]Center for Macroecology, Evolution and Climate, University of Copenhagen, Copenhagen, Denmark

BK, 0000-0002-2427-2364; FF, 0000-0002-7909-1800; CMØR, 0000-0003-2982-9931

The early Palaeozoic Era records the initial biodiversification of the Phanerozoic. The increase in biodiversity involved drastic changes in taxon longevity, and in rates of origination and extinction. Here, we calculate these variables in unprecedented temporal resolution. We find that highly volatile origination and extinction rates are associated with short genus longevities during the Cambrian Period. During the Ordovician and Silurian periods, evolutionary rates were less volatile and genera persisted for increasingly longer intervals. The 90%-genus life expectancy doubled from 5 Myr in the late Cambrian to more than 10 Myr in the Ordovician–Silurian periods. Intervals with widespread ecosystem disruption are associated with short genus longevities during the Cambrian and with exceptionally high longevities during the Ordovician and Silurian periods. The post-Cambrian increase in persistence of genera, therefore, indicates an elevated ability of the changing early Palaeozoic marine ecosystems to sustainably maintain existing genera. This is evidence of a new level of ecosystem resilience which evolved during the Ordovician Period.

1. Introduction

The spectacular early Palaeozoic rise in taxonomic richness of marine ecosystems continues to be a focus point of palaeobiological research [1–8]. It featured two distinct events of accelerated biodiversity accumulation, namely the Cambrian explosion (CE) and the Great Ordovician Biodiversification Event (GOBE). In addition, it contained a number of major crises during the Late Ordovician mass extinctions (LOME) [8].

A growing body of evidence suggests that the timing and intensity of the early Palaeozoic biodiversity accumulation was associated with changes in global temperature and oxygen levels [8–13]. However, the mechanisms linking, e.g. change in habitat space [12,14], spread of oxygen minimum zones [15,16], and extent of primary production [17,18] with biodiversity remain elusive [19].

Global biodiversity accumulation results from a combined process of origination and extinction of taxa, or viewed from a different perspective, it builds as a function of longevity of newly originating taxa. Hence, knowledge on taxon longevity and origination/extinction rates is essential to make inferences about the mechanisms of biodiversity accumulation. Many studies on evolutionary rates exist at the Phanerozoic and Palaeozoic scale and at the family and genus level (e.g. [4,20–28]). Longevity and survivorship rates have previously also been the focus of interest (e.g. [29–31]).

Rates of origination and extinction ultimately determine the probability of a taxon (here, a genus) to survive until a time t [29,30]. This relationship should not lead to the conclusion that analyses of longevity and evolutionary rates are redundant. Evolutionary rates inform about the volatility of the evolutionary change at a

given time interval, but they are agnostic about the specific composition of the rates of the individual genera and their life history. Identical evolutionary rates can be produced by originations and extinctions of long-living and short-living genera. Extinctions can preferably affect genera that persisted for a long time or, by contrast, genera that originated shortly before. Conversely, originations may result in long-lasting genera or short-living genera. The ecological mechanisms behind these different scenarios differ drastically and periods of ecosystem disturbance or resilience may remain unnoticed when only described by evolutionary rates.

Here, we present new estimates of rates of origination and extinction at the genus level with an unprecedented temporal resolution, based on a time binning established in Rasmussen *et al.* [8]. Additionally, we present for the first time per time bin estimates of longevity, taxon age, and taxon life expectancy of early Palaeozoic marine genera. Our results allow for a differentiation between taxonomic turnover and genus persistence, that again enables an evaluation of time-specific ecosystem resilience (i.e. the ability of a system to absorb changes and still persist, *sensu* Holling, [32]) as a factor of biodiversity accumulation.

2. Methods

We based our calculations on a sum of 173 293 genus-level Cambrian to Silurian fossil occurrences downloaded from the Paleobiology Database (PBDB, https://paleobiodb.org/#/, download 30 January 2019) and an additional download of 545 449 post-Silurian genus level occurrences from the PBDB (download 02 February 2019). The occurrences were binned into 53 Cambrian–Silurian time intervals with an average duration of 2.3 Myr following [8] and into post-Silurian stage intervals using the binning scheme of the PBDB (https://paleobiodb.org/data1.2/intervals/list.txt?scale=1, accessed 6 July 2019). Details of the data filtering and methodology of time binning and biodiversity calculations have been published in [8]. We estimated genus richness based on the capture-recapture model (CR) approaches [33,34] by fitting the Jolly–Seber model following the POPAN formulation [35]. We calculated relative diversification rates by dividing the richness difference between a time bin and its previous time bin with the richness of the respective time bin ($(n_{gen(t)} - n_{gen(t-1)})/n_{gen(t)}$). With n_{gen} being the number of genera, t being the time bin of interest, and $t-1$ being the previous time bin.

Additionally, we estimated survival and seniority probabilities based on the CR-approach using the Pradel model [36], which were transformed into extinction and origination rates, following the transformation from probabilities into rates described in [37]. The method estimates survival, seniority, and sampling probabilities, which we turn into rates, to account for uneven sampling intervals (see electronic supplementary material, and [34] for details of the method). For comparison of our CR-modelling results with more conventional rate estimations, we calculated origination and extinction rates with the turnover rate metric of Alroy [38] as implemented in the R-package divDyn [39] (see electronic supplementary material).

We estimated genus age, genus life expectancy, and genus longevity indirectly by calculating forward and backward survivorships of cohorts of genera occurring in each time bin. The duration needed to reach the full diversity of genera occurring in each time bin is our measure of backward survivorship (l_{bw}) and can be read as a measure of genus age. The subsequent lifetime of the set of genera occurring in a time bin, is our measure of forward survivorship (l_{fw}) and can be read as a measure of life expectancy. Long-life expectancies of genera indicate a long persistence of the ecological relationships established among these genera. Hence, we interpreted l_{fw} as an indicator of ecosystem

resilience (where resilience determines the persistence of relationships [32]). The backward survivorship can be read as a measure for the age structure of the genera of a time bin and reflects the history of the ecosystems. The sum of l_{bw} and l_{fw} is our overall longevity (l_o), which is a wrapper representing the past and the future of the genera that existed during each time bin.

Our longevity calculations are based on CR-modelled richness curves of the cohorts of genera occurring in each time bin of interest (t_i). In this calculation, a 100% richness always occurs in t_i and the modelled richness always increases in time bins (t_{i-n}) preceding t_i and decreases in the time bins (t_{i+n}) posterior to t_i. We determined the antecedent and posterior time bins containing 50%, 70%, and 90% t_i-richness levels and calculated l_{bw} and l_{fw} as the maximum time ranging from t_i towards these time bins.

The complete algorithm and relevant results are recorded in R-code and can be downloaded at https://doi.org/10.5281/zenodo.3365505.

3. Results

(a) Origination and extinction rates

Our estimated origination and extinction rates reveal basic differences between Cambrian and post-Cambrian evolutionary dynamics (figure 1c). The Cambrian rates are on average much higher than the post-Cambrian rates. Fluctuations of rates between time bins are much greater in the Cambrian Period. The generally decreasing Cambro–Ordovician rates trend was known already from curves with lower stratigraphic resolution [4,25–27]. Additionally, data from trilobites evidenced distinct differences in survivorships between Cambrian and Ordovician cohorts [30]. Our results show that this trilobite survivorship change reflects a more general pattern and that there is a strong change at the Cambro–Ordovician boundary. The significance of the trend change can partially also be demonstrated with a time series changepoint analysis, where a single changepoint of the origination rate time series occurs at the Cambro–Ordovician boundary (electronic supplementary material, figure S1).

Notably, the origination and extinction rates calculated with Alroy's [38] turnover rate metric show a less rapid but more continuous decrease at the Cambro–Ordovician boundary and continued to drop until the beginning of the Middle Ordovician (electronic supplementary material, figure S2), similar to, e.g. in Bambach *et al.* [26]. Bambach's [26] estimations and Alroy's [38] turnover metric result in instantaneous rates that do not account for the length of the time bins. The high estimates in the earliest Ordovician time bins in Alroy's and Bambach's calculations are therefore probably an effect of the poorly constrained timing of these intervals. The relatively long Early Ordovician time bins thus contain a comparatively high number of short-ranging taxa (see below).

Exceptional Cambrian events are the peak origination rates at the late Terreneuvian, early Miaolingian, and early Furongian epochs. Conversely, Cambrian extinction rates peak during the middle Series 2, the late Miaolingian, and early Furongian epochs, reflecting the Botomian [40] and Marjuman extinctions [41]. During the succeeding Ordovician Period, origination rates peaked at the Dapingian–Darriwilian boundary, and extinction rates reached maximum values at the Katian–Hirnantian boundary, reflecting the LOME [42]. Lastly, Silurian extinction rates peaked at the Homerian–Gorstian boundary towards the end of that period. This reflects the Mulde event [43]. The
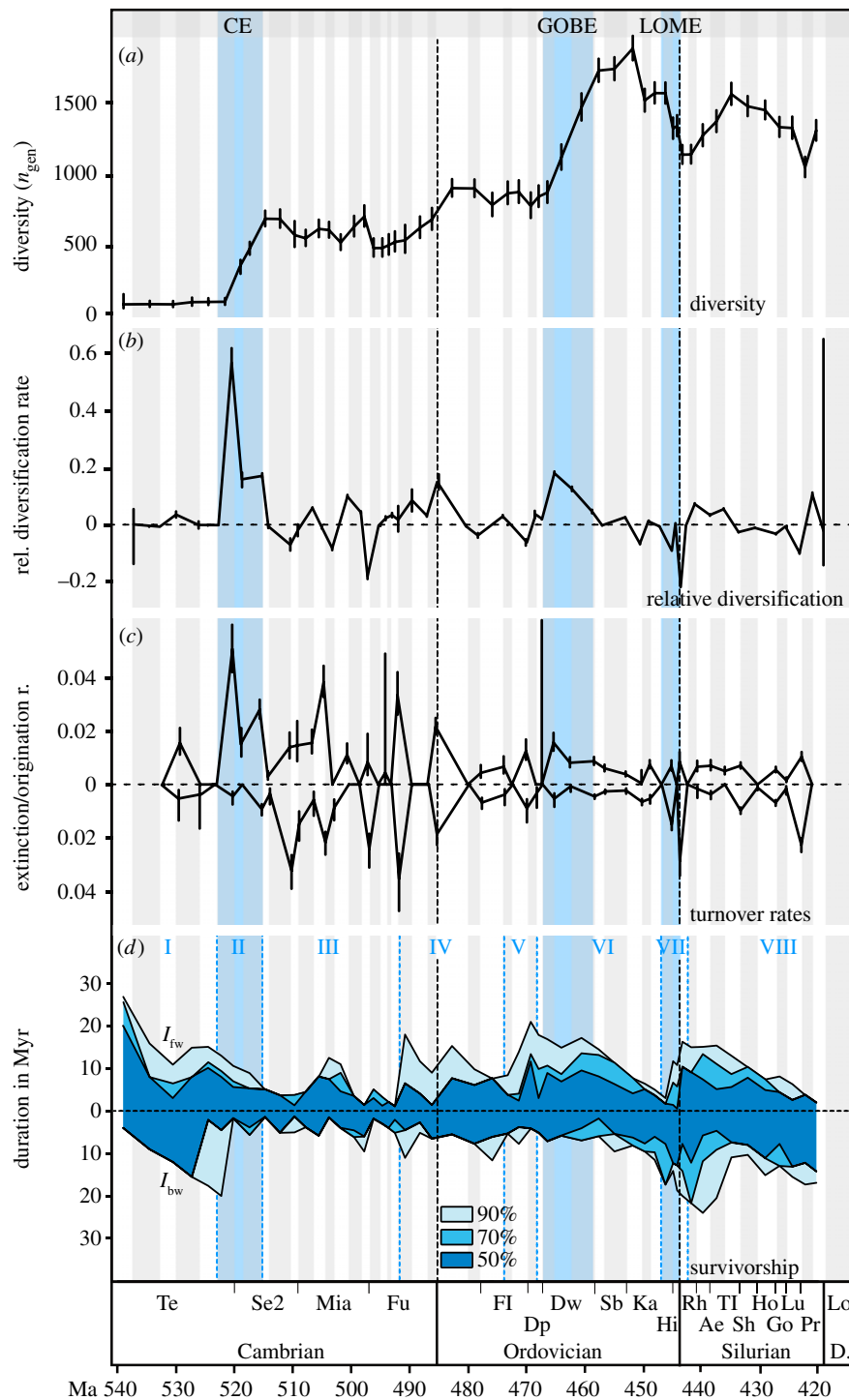
Figure 1. Early Palaeozoic curves of (a) per time bin genus level richness (adapted from [8]), (b) genus level relative diversification rate, (c) genus level extinction and origination rates (r.), and (d) duration of the forward (l_{fw}) and backward (l_{bw}) survivorship of 50%, 70%, and 90% of the cohort of genera of each time bin. (a), (c), and (d) are estimated with CR-modelling. Vertical bars indicate 95% confidence intervals. Note major changes in (a), (c), and (d) during the Furongian–Tremadocian interval. I-IX, designate numbered geo-historical intervals of distinct survivorship trends. Ae, Aeronian; CE, Cambrian Explosion; D., Devonian; Dp, Dapingian; Dw, Darriwilian; Fl, Floian; Fu, Furongian; Go, Gorstian; GOBE; Great Ordovician Biodiversification Event; Hi, Hirnantian; Ho, Homerian; Ka, Katian; Lo, Lochkovian; LOME; Late Ordovician Mass Extinctions; Lu, Ludfordian; Mia, Miaolingian; Pr, Pridolian; Rh, Rhuddanian; Sb, Sandbian; Se2, Cambrian Series 2; Sh, Sheinwoodian; Te, Terreneuvian; Tl, Telychian; Tr, Tremadocian.

observed events here are robust and stand out in the calculations resulting from the CR-modelling and from Alroy's [38] approach (figure 1c; electronic supplement material, figure S2).

(b) Genus survivorships and longevities

The temporal variation of l_{bw} and l_{fw} is expressed in a geo-historical succession of eight distinct intervals (figure 1d), which are best described as follows: the first (I) interval is characterized by increasing l_{bw} and decreasing l_{fw}, reflecting the initial low diversity phase of the Terreneuvian Epoch with the appearance and slow accumulation of more and more new genera. The second (II) interval represents the CE with the rapid appearance of new genera causing l_{bw} to decrease. In the third (III) interval, which lasted until the mid-Furongian Epoch, l_{bw} and l_{fw} remained low at values of, on average, 4–5 Myr. High taxonomic turnover during this time indicates rapid evolutionary change. In the fourth

(IV) interval, which spans the late Furongian Epoch to middle Floian Age, l_{bw} and l_{fw} initially increased and remained at intermediate levels. Hence, during this time more genera persisted for longer and had higher chances to survive for longer times in the future. The overall post-Terreneuvian peak of l_{fw} was reached during the Dapingian Age with more than 21 Myr of 90% life durations, during the fifth (V) interval. The end of the fifth interval marks the beginning of the GOBE. Peak diversification was reached at the beginning of the sixth (VI) interval during the early Darriwilian Age and was paralleled with a decreasing l_{fw}. In the sixth interval, which ranges until the late Katian Age, l_{fw} decreased while l_{bw} increased. Hence, more and more genera occurred with long antecedent life histories, but at the same time the prospect for their future survival decreased. This is clearly an effect of the seventh (VII) interval which lasted from the latest Katian towards the early Rhuddanian Age and which represents the LOME and its direct aftermath. As a consequence of the extinctions, the age structure of the occurring genera was strongly altered and the l_{bw} was at its early Palaeozoic peak in the next interval (VIII) (Rhuddanian–Sheinwoodian ages). A trend of increasing l_{bw} and decreasing l_{fw} during this interval indicates recovery and the appearance of more and more new genera.

4. Discussion

(a) Periods of early Palaeozoic biodiversity accumulation

The synoptic comparison of evolutionary rates, survivorships, and longevity curves allows a periodization of the early Palaeozoic time into a number of intervals characterized by specific evolutionary dynamics. These intervals can be related to the known changes of the biodiversity curve and to the changes in global temperature and oxygen levels. The resulting picture of such a comparison reveals an evolutionary history that began with a relatively stable interval with low evolutionary rates, high genus survivorships, and low diversities (figure 1d, interval I). This relatively stable situation quickly escalated with the rapid appearance of skeletal lophotrochozoans, ecdysozoans, as well as sponge and archaeocyathid reefs and with a climax of the CE during the latest Terreneuvian and Cambrian Epoch 2 [44–46]. The remainder of the Cambrian was characterized by a high volatility of the evolutionary rates, extremely low genus survivorships, and a biodiversity accumulation with a rising trend towards the Ordovician (figure 1d, interval III). Our analysis thus portrays the late middle–late Cambrian as a highly dynamic period with low ecosystem resilience and this is concurrent with a growing body of evidence that the post-CE Cambrian age was a time with recurrent expansions of oxygen minimum zones across the shallow shelf and correspondent habitat disruptions [15,16,47,48].

The terminal Cambrian and the beginning of the Ordovician periods mark another phase in the evolutionary dynamics of the early Palaeozoic that lasted until the end of the Floian Age (figure 1d, intervals IV–V), which was characterized by lowered evolutionary rates, increasing genus survivorships, and a stable level in biodiversity accumulation. This interval coincides with the global expansion and biodiversification of planktic primary producers and with the first appearance of planktic graptolites and cephalopods (the 'plankton revolution' [17]).

The Middle Ordovician time records the main phase of the GOBE with a massive increase in biodiversity accumulation during the Darriwilian Age [8]. Notably, the GOBE peak diversification is preceded by a drastic increase of the forward survivorship rates of genera during the Dapingian Age. This means that Dapingian genera that survived into the Darriwilian had exceptionally high chances to further persist several Myr into the Late Ordovician. Hence, the exceptionally long-life expectancy of Dapingian genera is best explained as an *ex-post* effect of the GOBE. Similarly, the decreasing life expectancy from the Dapingian Age onward until the end of the Ordovician Period is an *ex-post* effect of the LOME. Middle and Late Ordovician genera, successively were doomed to extinction during the LOME. During the LOME genera with short precedent life histories went preferentially extinct. This is consistent with the finding that it was particularly the brachiopod genera with limited geographical ranges that went extinct [49] and that predominantly rare graptolite genera were hit already early on during the LOME [50].

At the same time, genera newly evolving and surviving during the LOME (interval VII, figure 1d) had higher chances to survive for longer. This is the well-known effect of increased life expectancies of genera occurring and originating during and immediately after mass extinctions [51,52]. Previous studies and models show that genera surviving or originating during mass extinctions tend to have a temporal advance to accumulate species [52]. As a consequence, extinctions acted as a filter for long-living genera, causing an early Palaeozoic genus longevity maximum during the Early Silurian Period. Only with the subsequent origination of new short-living Silurian genera during the post-LOME recovery, the genus longevity levels returned to pre-LOME values.

(b) Mechanisms of early Palaeozoic biodiversity accumulation

The existence of an early Palaeozoic maximum in life expectancy (l_{fw}) just before the onset of GOBE is important evidence for the mechanisms behind biodiversity accumulation during this time: the GOBE coincides with an Ordovician peak in origination rates, but not with exceptionally low extinction rates (figure 1c). The exceptionally high life expectancy of Dapingian–early Darriwilian genera, therefore, cannot be explained by lowered extinction rates but as an effect of increasingly long lives of genera that did not go extinct. Dapingian–early Darriwilian genera, which persisted, did so for exceptionally long time intervals. Importantly, the Early Ordovician trend of increasing l_{fw} is succeeded by a Middle–Late Ordovician trend of increasing l_{bw}, resulting in a massive rise in l_o across the entire Ordovician. This means that, despite the environmental perturbations during the LOME, the combined life expectancy and the age structure of genera increased significantly.

The second important conclusion that can be drawn from the pre-GOBE l_{fw} peak is that the maximum life expectancy was not exclusively caused by 'GOBE-specific' novel genera, but also by genera that existed well before the GOBE during the Floian and Dapingian ages. The prolonged life expectancy was not an effect of specific novel genera but an effect of an increased ability of the GOBE ecosystems to sustainably maintain existing genera.

This suggests mechanisms of ecosystem evolution during most of the Ordovician Period, where existing

genera became increasingly successfully integrated under novel ecological conditions such as different temperature and oxygenation regimes while new genera appeared constantly. One example of such an integrative mode of ecosystem evolution is the Ordovician diversification of bryozoan, coral, and stromatoporoid reefs. These clades existed as minor components in tropical shallow-water habitats from the Tremadocian Age, but collectively diversified and became dominant reef builders under cooling climatic conditions during the Middle Ordovician [12,53]. Once established, these reef builders persisted throughout the early Palaeozoic and survived even massive perturbations, such as the LOME [54].

Here, a basic difference between the Cambrian and Ordovician evolutionary dynamics becomes apparent. Cambrian conditions, such as poor oxygenation and high global temperatures are considered to be major factors of ecosystem disruptions that caused origination and extinction rates to fluctuate and genus persistence in ecosystems to decrease, e.g. [47,48]. By contrast, climatically induced global disruptions of the marine ecosystems during the LOME (e.g. [55,56]), had the opposite effect on genus persistence. During the latest Ordovician, genus longevities continued to rise even under drastically reduced biodiversity (figure 1). This basic difference is evidence of a new level of ecosystem resilience that evolved during the Ordovician. It is tempting to suggest that the Early Ordovician revolution in plankton with a first establishment of diverse and stable pelagic food chains that involved common macropredators, such as cephalopods, was an important step towards these new levels. Stable pelagic food webs affected, e.g. larval dispersal and spatial taxon ranges, which in turn potentially affected the taxon longevity.

One general conclusion can be drawn from these geo-historically more specific interpretations: generic life expectancies during the Palaeozoic were highest during time intervals directly preceding diversifications and early during diversification peaks. The diversifications affected novel and established genera likewise by increasing their average life expectancies. Therefore, processes that led to increased levels of ecosystem resilience were major factors of marine biodiversity accumulation of the Palaeozoic.

References

1. Sepkoski JJ. 1979 A kinetic model of phanerozoic taxonomic diversity II. Early phanerozoic families and multiple equilibria. *Paleobiology* **5**, 222–251. (doi:10.1017/S0094837300006539)

2. Miller AI. 1997 Comparative diversification dynamics among Palaeocontinents during the Ordovician radiation. *Geobios* **30**(Supplement 1), 397–406. (doi:10.1016/S0016-6995(97)80044-7)

3. Miller AI. 1997 Dissecting global diversity patterns: examples from the Ordovician radiation. *Annu. Rev. Ecol. Evol. Syst.* **28**, 85–104. (doi:10.1146/annurev.ecolsys.28.1.85)

4. John Sepkoski J. 1998 Rates of speciation in the fossil record. *Phil. Trans. R. Soc. Lond. B* **353**, 315–326. (doi:10.1098/rstb.1998.0212)

5. Webby BD, Paris F, Droser M, Percival I. 2004 *The Great Ordovician Biodiversification Event*. New York, NY: Columbia University Press.

6. Servais T, Owen AW, Harper DAT, Kröger B, Munnecke A. 2010 The Great Ordovician Biodiversification Event (GOBE): the palaeoecological dimension. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **294**, 99–119. (doi:10.1016/j.palaeo.2010.05.031)

7. Miller AI. 2012 The Ordovician Radiation: Macroevolutionary Crossroads of the Phanerozoic Earth and Life: Global Biodiversity, Extinction Intervals and Biogeographic Perturbations Through Time (ed. JA Talent), pp. 381–394. Dordrecht: Springer Netherlands.

8. Rasmussen CMØ, Kröger B, Nielsen ML, Colmenar J. 2019 Cascading trend of Early Paleozoic marine radiations paused by Late Ordovician extinctions. *Proc. Natl Acad. Sci. USA* **116**, 7207–7213. (doi:10.1073/pnas.1821123116)

9. Trotter JA, Williams IS, Barnes CR, Lécuyer C, Nicoll RS. 2008 Did cooling oceans trigger Ordovician biodiversification? Evidence from conodont thermometry. *Science* **321**, 550–554. (doi:10.1126/science.1155814)

10. Sperling EA, Knoll AH, Girguis PR. 2015 The ecological physiology of Earth's second oxygen revolution. *Annu. Rev. Ecol. Evol. Syst.* **46**, 215–235. (doi:10.1146/annurev-ecolsys-110512-135808)

11. Edwards CT, Saltzman MR, Leslie SA, Bergström SM, Sedlacek ARC, Howard A, Bauer JA, Sweet WC, Young SA. 2015 Strontium isotope (87Sr/86Sr) stratigraphy of Ordovician bulk carbonate: implications for preservation of primary seawater values. *Geol. Soc. Am. Bull.* **127**, 1275–1289. (doi:10.1130/B31149.1)

12. Kröger B. 2017 Changes in the latitudinal diversity gradient during the Great Ordovician Biodiversification Event. *Geology* **46**, 127–130. (doi:10.1130/G39587.1)

13. Hearing TW, Harvey THP, Williams M, Leng MJ, Lamb AL, Wilby PR, Gabbott SE, Pohl A, Donnadieu Y. 2018 An early Cambrian greenhouse climate. *Sci. Adv.* **4**, eaar5690. (doi:10.1126/sciadv.aar5690)

14. Sperling EA, Frieder CA, Raman AV, Girguis PR, Levin LA, Knoll AH. 2013 Oxygen, ecology, and the Cambrian radiation of animals. *Proc. Natl Acad. Sci. USA* **110**, 13 446–13 451. (doi:10.1073/pnas.1312778110)

15. Edwards CT, Saltzman MR, Royer DL, Fike DA. 2017 Oxygenation as a driver of the Great Ordovician Biodiversification Event. *Nat. Geosci.* **10**, 925–929. (doi:10.1038/s41561-017-0006-3)

16. Edwards CT. 2019 Links between early paleozoic oxygenation and the Great Ordovician Biodiversification Event (GOBE): a review. *Palaeoworld* **28**, 37–50. (doi:10.1016/j.palwor.2018.08.006)

17. Servais T, Lehnert O, Li J, Mullins GL, Munnecke A, Nützel A, Vecoli M. 2008 The Ordovician Biodiversification: revolution in the oceanic trophic chain. *Lethaia* **41**, 99–109. (doi:10.1111/j.1502-3931.2008.00115.x)

18. Cárdenas AL, Harries PJ. 2010 Effect of nutrient availability on marine origination rates throughout the Phanerozoic eon. *Nat. Geosci.* **3**, 430–434. (doi:10.1038/ngeo869)

19. Stigall AL. 2017 Ordovician oxygen and biodiversity. *Nat. Geosci.* **10**, 887–888. (doi:10.1038/s41561-017-0024-1)

20. Raup DM, Sepkoski JJ. 1982 Mass extinctions in the marine fossil record. *Science* **215**, 1501–1503. (doi:10.1126/science.215.4539.1501)

21. Sepkoski JJ. 1987 Environmental trends in extinction during the Paleozoic. *Science* **235**, 64–66. (doi:10.1126/science.11539724)

22. Foote M. 1994 Temporal variation in extinction risk and temporal scaling of extinction metrics. *Paleobiology* **20**, 424–444. (doi:10.1017/S0094837300012914)

23. Foote M. 2000 Origination and extinction components of taxonomic diversity: general problems. *Paleobiology* **26**, 74–102. (doi:10.1017/S0094837300026890)

24. Connolly SR, Miller AI. 2001 Joint estimation of sampling and turnover rates from fossil databases: capture-mark-recapture methods revisited. *Paleobiology* **27**, 751–767. (doi:10.1666/0094-8373(2001)027<0751:jeosat>2.0.co;2)

25. Foote M. 2003 Origination and extinction through the phanerozoic: a new approach. *J. Geol.* **111**, 125–148. (doi:10.1086/345841)

26. Bambach RK, Knoll AH, Wang SC. 2004 Origination, extinction, and mass depletions of marine diversity. *Paleobiology* **30**, 522–542. (doi:10.1666/0094-8373(2004)030<0522:OEAMDO>2.0.CO;2)

27. Alroy J. 2008 Dynamics of origination and extinction in the marine fossil record. *Proc. Natl Acad. Sci. USA* **105**, 11 536–11 542. (doi:10.1073/pnas.0802597105)

28. Franeck F, Liow LH. 2019 Dissecting the paleocontinental and paleoenvironmental dynamics of the great Ordovician biodiversification. *Paleobiology* **45**, 221–234. (doi:10.1017/pab.2019.4)

29. Raup DM. 1978 Cohort analysis of generic survivorship. *Paleobiology* **4**, 1–15. (doi:10.1017/S0094837300005649)

30. Foote M. 1988 Survivorship analysis of Cambrian and Ordovician trilobites. *Paleobiology* **14**, 258–271. (doi:10.1017/S0094837300011994)

31. Foote M, Miller AI. 2013 Determinants of early survival in marine animal genera. *Paleobiology* **39**, 171–192. (doi:10.1666/12028)

32. Holling CS. 1973 Resilience and stability of ecological systems. *Annu. Rev. Ecol. Syst.* **4**, 1–23. (doi:10.1146/annurev.es.04.110173.000245)

33. Nichols JD, Pollock KH. 1983 Estimating taxonomic diversity, extinction rates, and speciation rates from fossil data using capture-recapture models. *Paleobiology* **9**, 150–163. (doi:10.1017/S0094837300007533)

34. Liow LH, Nichols JD. 2010 Estimating rates and probabilities of origination and extinction using taxonomic occurrence data: capture-mark-recapture (CMR) approaches. In *The paleontological society short course, October 30th 2010* (eds J Alroy, G Hunt), pp. 81–94.

35. Schwarz CJ, Arnason AN. 1996 A general methodology for the analysis of capture-recapture experiments in open populations. *Biometrics* **52**, 860–873. (doi:10.2307/2533048)

36. Pradel R. 1996 Utilization of capture-mark-recapture for the study of recruitment and population growth rate. *Biometrics* **52**, 703–709. (doi:10.2307/2532908)

37. Liow LH, Reitan T, Harnik PG. 2015 Ecological interactions on macroevolutionary time scales: clams and brachiopods are more than ships that pass in the night. *Ecol. Lett.* **18**, 1030–1039. (doi:10.1111/ele.12485)

38. Alroy J. 2015 A more precise speciation and extinction rate estimator. *Paleobiology* **41**, 633–639. (doi:10.1017/pab.2015.26)

39. Kocsis ÁT, Reddin CJ, Alroy J, Kiessling W. 2019 The R package divDyn for quantifying diversity dynamics using fossil sampling data. *Methods Ecol. Evol.* **10**, 735–743. (doi:10.1111/2041-210X.13161)

40. Zhuravlev AY, Wood RA. 1996 Anoxia as the cause of the mid-Early Cambrian (Botomian) extinction event. *Geology* **24**, 311–314. (doi:10.1130/0091-7613(1996)024<0311:AATCOT>2.3.CO;2)

41. Gerhardt AM, Gill BC. 2016 Elucidating the relationship between the later Cambrian end-Marjuman extinctions and SPICE Event. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **461**, 362–373. (doi:10.1016/j.palaeo.2016.08.031)

42. Harper DAT, Hammarlund EU, Rasmussen CMØ. 2014 End Ordovician extinctions: a coincidence of causes. *Gondwana Res.* **25**, 1294–1307. (doi:10.1016/j.gr.2012.12.021)

43. Jeppsson L, Calner M. 2002 The Silurian Mulde Event and a scenario for secundo–secundo events. *Earth Environ. Sci. Trans. R. Soc. Edinburgh* **93**, 135–154. (doi:10.1017/S0263593300000377)

44. Zhuravlev AY, Naimark EB, Wood RA. 2015 Controls on the diversity and structure of earliest metazoan communities: early Cambrian reefs from Siberia. *Earth-Science Rev.* **147**, 18–29. (doi:10.1016/j.earscirev.2015.04.008)

45. Budd GE, Jackson ISC. 2016 Ecological innovations in the Cambrian and the origins of the crown group phyla. *Phil. Trans. R. Soc. B* **371**, 20150287. (doi:10.1098/rstb.2015.0287)

46. Paterson JR, Edgecombe GD, Lee MSY. 2019 Trilobite evolutionary rates constrain the duration of the Cambrian explosion. *Proc. Natl Acad. Sci. USA* **116**, 4394–4399. (doi:10.1073/pnas.1819366116)

47. Saltzman MR, Edwards CT, Adrain JM, Westrop SR. 2015 Persistent oceanic anoxia and elevated extinction rates separate the Cambrian and Ordovician radiations. *Geology* **43**, 807–810. (doi:10.1130/G36814.1)

48. Wood R, Erwin DH. 2018 Innovation not recovery: dynamic redox promotes metazoan radiations. *Biol. Rev.* **93**, 863–873. (doi:10.1111/brv.12375)

49. Finnegan S, Rasmussen CM, Harper DAT. 2016 Biogeographic and bathymetric determinants of brachiopod extinction and survival during the Late Ordovician mass extinction. *Proc. R. Soc. B* **283**, 20160007. (doi:10.1098/rspb.2016.0007)

50. Sheets HD, Mitchell CE, Melchin MJ, Loxton J, Storch P, Carlucci KL, Hawkins AD. 2016 Graptolite community responses to global climate change and the Late Ordovician mass extinction. *Proc. Natl Acad. Sci. USA* **113**, 8380–8385. (doi:10.1073/pnas.1602102113)

51. Kauffman EG, PJ Harries. 1996 The importance of crisis progenitors in recovery from mass extinction. *Geol. Soc. Lond Special Publications* **102**, 15–39. (doi:10.1144/GSL.SP.1996.001.01.02)

52. Miller AI, Foote M. 2003 Increased longevities of post-Paleozoic marine genera after mass extinctions. *Science* **302**, 1030–1032. (doi:10.1126/science.1089719)

53. Webby BD. 2002 Patterns of Ordovician reef development. In *Phanerozoic reef patterns* (eds W Kiessling, EK Flügel, J Golonka), pp. 129–179. SEPM (Society for Sedimentary Geology).

54. Copper P. 2001 Reefs during the multiple crises towards the Ordovician-Silurian boundary: Anticosti Island, eastern Canada, and worldwide. *Can. J. Earth Sci.* **38**, 153–171. (doi:10.1139/e00-071)

55. Crampton JS, Cooper RA, Sadler PM, Foote M. 2016 Greenhouse-icehouse transition in the Late Ordovician marks a step change in extinction regime in the marine plankton. *Proc. Natl Acad. Sci. USA* **113**, 1498–1503. (doi:10.1073/pnas.1519092113)

56. Zou C, Qiu Z, Poulton SW, Dong D, Wang H, Chen D, Lu B, Shi Z, Tao H. 2018 Ocean euxinia and climate change 'double whammy' drove the Late Ordovician mass extinction. *Geology* **46**, 535–538. (doi:10.1130/G40121.1)