

# Application of biocomposite edible coatings based on pea starch and guar gum on quality, storability and shelf life of 'Valencia' oranges

Bahareh Saberi, John B. Golding, José R. Marques, Penta Pristijono, Suwimol Chockchaisawasdee, Christopher J. Scarlett and Costas E. Stathopoulos

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1 **Application of biocomposite edible coatings based on pea starch and guar gum on quality,**  
2 **storability and shelf life of ‘Valencia’ oranges**

3 Bahareh Saberi<sup>a\*</sup>, John B. Golding<sup>a,b</sup>, José R. Marques<sup>b</sup>, Penta Pristijono<sup>a</sup>, Suwimol  
4 Chockchaisawasdee<sup>a,c</sup>, Christopher J. Scarlett<sup>a</sup> and Costas E. Stathopoulos<sup>c</sup>

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6 <sup>a</sup> School of Environmental and Life Sciences, University of Newcastle, Ourimbah, NSW 2258,  
7 Australia

8 <sup>b</sup> NSW Department of Primary Industries, Ourimbah, NSW 2258, Australia

9 <sup>c</sup> Division of Food and Drink, School of Science, Engineering and Technology, University of  
10 Abertay, Dundee DD1 1HG, UK

11

12 **\*Correspondence to:**

13 Bahareh Saberi

14 School of Environmental and Life Sciences, Faculty of Science and Information Technology,  
15 University of Newcastle, Brush Road, Ourimbah, NSW 2258, Australia.

16 Tel: +61 449968763; Fax: +61 2 4348 4145; E-mail: bahareh.saberi@uon.edu.au

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19 **ABSTRACT**

20 Novel edible composite coatings based on pea starch and guar gum (PSGG), PSGG blended  
21 with lipid mixture containing the hydrophobic compounds shellac and oleic acid (PSGG-Sh),  
22 and a layer-by-layer (LBL) approach (PSGG as an internal layer and shellac as an external  
23 layer), were investigated and compared with a commercial wax (CW) and uncoated fruit on  
24 postharvest quality of ‘Valencia’ oranges held for up to four weeks at 20 °C and 5 °C with an  
25 additional storage for 7 d at 20 °C. The incorporation of lipid compounds into the PSGG  
26 coatings (PSGG-Sh) generally resulted in the best performance in reducing fruit respiration  
27 rate, ethylene production, weight and firmness loss, peel pitting, and fruit decay rate of the  
28 coated oranges. Fruit coated with PSGG-Sh and a single layer PSGG coatings generally  
29 resulted in higher scores for overall flavor and freshness after four weeks at 5 °C followed by  
30 one week at 20 °C than uncoated fruit, as assessed by a sensory panel. Although the LBL  
31 coating reduced weight loss and respiration rate with improved firmness retention to a greater  
32 extent than the single layer PSGG coating, the bilayer coating also resulted in higher levels of  
33 ethanol causing increased perception of off-flavors. Overall results suggested that PSGG-based  
34 edible coatings could be a beneficial substitute to common commercial waxes for maintaining  
35 quality and storability, as well as extending shelf life of citrus fruit and potentially other fresh  
36 horticultural produce.

37 *Keywords:* Biocomposite edible coating, Citrus, Pea starch, Guar gum, Postharvest quality

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## 40 **1. Introduction**

41 Edible films and coatings are widely used to maintain the quality and shelf life of many  
42 horticultural products, including citrus (Baldwin et al., 2011). Edible films and coatings act as  
43 semi-permeable membranes which restrict the movement of gases and water vapor to reduce  
44 the rate of respiration and water loss from the fruit. Many films/coatings due to their barrier  
45 and mechanical properties can reduce the rate of physiological postharvest degradation  
46 (Baldwin, 1994; Baldwin et al., 1995; Park, 1999).

47 In many countries, harvested citrus fruit are commonly waxed during their processing and  
48 packing. This is to replace the natural wax which is damaged/removed with commercial  
49 harvesting, handling, processing and packing (Valencia- Chamorro et al., 2010). The  
50 commercial application of waxes not only reduces weight loss and shrinkage, but also enhances  
51 shine by increasing gloss (Rojas-Argudo et al., 2009). However, some waxes have been shown  
52 to negatively alter the internal atmosphere of the fruit by inducing anaerobic off-flavor  
53 development with the restriction of respiratory gas exchange (Martínez-Jávega et al., 1989).

54 Many modern citrus waxes are made of shellac (derived from the lac bug, *Kerria lacca*) or  
55 carnauba (derived from the leaves of the carnauba palm, *Copernicia prunifera*). However there  
56 is a need to improve the efficiency and sustainability of waxes applied to citrus.

57 Readily sourced and inexpensive coating materials which are effective at maintaining fruit  
58 quality during storage and shelf life are required. Pea (*Pisum sativum*) is widely grown around  
59 the world and contains 22–45% starch as the most plentiful carbohydrate in the seed (Hoover  
60 and Sosulski, 1991). Pea starch (PS) is comprised of a mixture of two homopolymers; a linear  
61 fraction, amylose, and a highly branched fraction, amylopectin. They are made of units of D-  
62 glucose with only two types of chain linkages, an  $\alpha$ -(1→4) of the main chain and an  $\alpha$ -(1→6)  
63 of the branch chains (Liu, 2005). Pea starch has high content of amylose; therefore it is a

64 potential option for the production of starch-based edible films (Van Soest et al., 2002). Guar  
65 gum (GG), which is derived from the endosperm of the guar bean (*Cyamopsis tetragonoloba*),  
66 is a type of linear galactomannan with ratio of mannose to galactose units of 2:1 (Prajapat and  
67 Gogate, 2015). The molecular structure of guar gum is composed of  $\beta(1\rightarrow4)$ -linked mannopy-  
68 ranose backbone, with several branch points from the C-6 position of mannopyranose, linked  
69 by  $\alpha(1\rightarrow6)$  bond to a single D-galactopyranose sugar (Whistler and BeMiller, 1993). Owing to  
70 the long polymeric chain, high molecular weight and wide availability of pea starch and guar  
71 gum, they can be potential alternatives for production of renewable source based biodegradable  
72 edible coatings or packaging materials. In our previous studies, it has been shown that pea  
73 starch in combination with guar gum can form biocomposite edible films with preferable  
74 physical, optical and mechanical properties (Saber et al., 2016a; Saber et al., 2016b; Saber et  
75 al., 2016c). However, edible coatings based on pea starch and guar gum have not been  
76 comprehensively explored as fruit coatings.

77 Due to the hydrophilic nature of pea starch-guar gum (PSGG) film, it is necessary to add a  
78 hydrophobic substance for decreasing the water sensitivity of the film. In this experiment,  
79 shellac (Sh) was added as a resin-based hydrophobic substance to increase its capability in  
80 increasing gloss and decrease water loss (Arnon et al., 2015). However, an issue with shellac  
81 films is their lack of permeability to gases, which results in the accumulation of ethanol and  
82 carbon dioxide (CO<sub>2</sub>) and the development of off-flavors during storage (Baldwin et al., 1995;  
83 Dhall, 2013; Porat et al., 2005).

84 In this study, we investigated the influence of pea starch-guar gum (PSGG), pea starch-guar  
85 gum-shellac (PSGG-Sh), and PSGG/Sh bilayer composite coating, formed by first applying  
86 PSGG and then shellac (Sh) compared with fruit coated with commercial wax and uncoated  
87 fruit (control) on maintaining the quality of fresh 'Valencia' oranges during four weeks at 20

88 °C and four weeks of storage at 5 °C followed by one week at 20 °C, simulating marketing shelf  
89 life.

## 90 **2. Materials and methods**

### 91 *2.1. Materials*

92 Canadian non-GMO (non-genetically modified organism) yellow pea starch with 13.2 %  
93 moisture, 0.2 % protein, 0.5 % fat, 0.3 % ash, and  $36.25 \pm 0.32$  % amylose was used in all  
94 experiments (supplied by Yantai Shuangta Food Co., Jinling Town, China). Guar gum (E-412)  
95 was purchased from The Melbourne Food Ingredient Depot, Brunswick East, Melbourne,  
96 Australia. Food grade alcohol-based solution of shellac and Citrus Gleam (a shellac-based  
97 commercial wax) were purchased from Castle Chemicals (castlechem.com.au), NSW,  
98 Australia. Oleic acid (OA) and Tween-20 were obtained from Sigma Aldrich, Australia.  
99 Glycerol was from Ajax Finechem Pty. Ltd, Australia and used as a plasticizer. All other  
100 chemicals were purchased from Merck Millipore, Pty., VIC, Melbourne, Australia.

### 101 *2.2. Sample preparation*

102 ‘Valencia’ oranges (*Citrus sinensis* L. Osbeck) were obtained from a local commercial citrus  
103 grower (Griffith, NSW, Australia) at commercial maturity and transported to the NSW  
104 Department of Primary Industries (Ourimbah, NSW, Australia). Oranges were selected based  
105 on homogeneity in shape, color, size, firmness and free of mechanical wounds or fungal decay.  
106 Selected oranges were dipped in a solution of  $1150 \mu\text{L L}^{-1}$  fludioxonil (Scholar<sup>®</sup>, Syngenta  
107 Australia) for one min, then drained and air-dried at 20 °C before coating application.

### 108 *2.3. Coating formulations*

#### 109 *2.3.1. PSGG coatings*

110 Pea starch (2.5 g), guar gum (0.3 g) and 25 % w/w glycerol as plasticizer based on the dry film  
111 matter were dissolved in 100 mL degassed deionized water. The solution was heated at 90 °C  
112 for 20 min upon constant stirring. The suspension was then cooled until room temperature with  
113 mild magnetic stirring (Saber et al., 2016b). The film solution was prepared one day before  
114 use.

### 115 2.3.2. PSGG-Sh coatings

116 The PSGG-Sh composite mixture was prepared by adding oleic acid (1 % of dry weight of pea  
117 starch and guar gum) as emulsifier and Tween-20 (0.3 mL) as surfactant to the PSGG solution  
118 made as described above. Food grade alcohol-based solution of shellac at 40 % (dry weight of  
119 pea starch and guar gum) was added to the PSGG-OA-Tween 20-glycerol mixture. These levels  
120 of film ingredients were optimized using Box–Behnken response surface design (Saber et al.,  
121 2017). The emulsion was gelatinized at 90 °C for 20 min on a hot plate with continuous stirring.  
122 Once the lipids had melted, samples were homogenized for 4 min at 22000 rpm using a T25  
123 Ultra-Turrax (Ika, Staufen, Germany). After homogenization, the film solution was cooled to  
124 room temperature with slow magnetic stirring. The emulsion was prepared one day before use  
125 and was shown to be stable with no phase separation.

### 126 2.4. Experimental design

127 Five series of treatments were applied on oranges: (i) PSGG; (ii) PSGG-Sh; (iii) bilayer  
128 formulation of PSGG as an inner layer with Sh solution as an external layer (PSGG/Sh); (iv)  
129 CW (commercial wax, shellac based ‘Citrus Gleam’) and (v) distilled water acting as a control.  
130 Each treatment for each storage condition included 128 oranges with 8 oranges per plastic  
131 netted bag. There were four replicates per treatment with each bag considered a single replicate.  
132 Data were recorded before treatment (day 0) and at 7 d intervals (four removals) for up to four  
133 weeks storage at 20 °C and relative humidity (RH) of 90–95 %, and logging the temperature

134 and RH with calibrated TinyTag View 2 loggers. Another set of treated oranges was also stored  
135 for 1, 2, 3, and 4 weeks at 5 °C and 90–95 % RH, followed by one additional week at 20 °C to  
136 simulate retail handling and marketing conditions.

### 137 *2.5. Fruit coating*

138 Each coating solution was sprayed uniformly on the whole fruit surface by using a paint sprayer  
139 (High Volume Low Pressure system, 500W Paint Sprayer, 909, Mooroolbark, Vic, Australia).  
140 The bilayer coatings were applied as follows: first the PSGG coating was applied and fruit were  
141 fan dried at room temperature for 2–3 min and then the Sh coating was applied. Then, all coated  
142 oranges were air-dried for 1 h at 20 °C, labelled, weighed, and then randomly packed into  
143 experimental units. Fruits were destructively measured each week for up to four weeks at either  
144 20 °C or 5 °C. Four oranges from each replicate were assessed upon removal (when the fruit  
145 had reached room temperature) and the remaining four fruit were stored for the additional week  
146 at 20 °C.

### 147 *2.6. Fruit quality parameters*

#### 148 *2.6.1. Weight loss*

149 Fruit weight loss was measured by weighing the same marked fruit, at the beginning of the  
150 experiment and at the end of each storage period. The results were presented as the percentage  
151 loss of initial weight (Rojas-Argudo et al., 2009).

#### 152 *2.6.2. Fruit firmness*

153 A texture analyzer (Lloyd Instrument LTD, Fareham, UK) was used to determine firmness of  
154 fruit upon each removal. The maximum force (N) was measured by compressing the fruit in  
155 the equatorial zone between two flat surfaces closing together at the rate of 1 mm min<sup>-1</sup> to a



156 depth of 2 mm. The average of two reading points from each side of the fruit were recorded  
157 (Cháfer et al., 2012).

### 158 2.6.3. *Respiration rate*

159 Respiration rate was measured by the method described by Pristijono et al. (2017a), where 6  
160 oranges from each replicate were allocated into 500 mL hermetic glass jars with a septum in  
161 the lid at 20 °C, and headspace gas sample (1 mL) was collected by a syringe after 1 h, and  
162 transferred to an ICA40 series low-volume gas analysis system (International Controlled  
163 Atmosphere Ltd., Kent, UK). Respiration rate was expressed as  $\mu\text{g CO}_2 \text{ kg}^{-1} \text{ s}^{-1}$ .

### 164 2.6.4. *Ethylene production*

165 Gas sample (1 mL) for analysis was taken 4 h after sealing the container as previous described  
166 for respiration. The concentration of ethylene was calculated by injecting the sample into a  
167 flame ionization gas chromatograph (Gow-Mac 580, Bridgewater NJ) fitted with a stainless  
168 steel column (2 m  $\times$  3.2 mm OD  $\times$  2.2 mm ID) packed with Porapak Q (80-100 mesh) (Altech,  
169 Sydney), with 110, 90 and 70 °C as the operating temperature of the detector, column, and the  
170 injector, respectively. Nitrogen, hydrogen and air were used as carrier and combustion gases at  
171 flow rates of 60, 30 and 300 mL min<sup>-1</sup>, respectively. The ethylene production rate was measured  
172 as ng C<sub>2</sub>H<sub>4</sub> kg<sup>-1</sup> s<sup>-1</sup> (Huque et al., 2013).

### 173 2.6.5. *Skin color*

174 Color was assessed using the CIE L\*, a\*, b\* scale and a Minolta colorimeter (Minolta CR-400,  
175 Osaka). The results were the means of three points on the fruit surface and expressed as Hue  
176 angle (Robles-Sánchez et al., 2013).

$$177 \text{ Hue} = \arctangent \left( \frac{b^*}{a^*} \right) \quad (1)$$

178 2.6.6. *Acetaldehyde and ethanol concentrations in fruit juice*

179 Headspace ethanol (g L<sup>-1</sup>) and acetaldehyde (mg L<sup>-1</sup>) concentration in orange juice was  
180 determined according to Kumar et al. (2014). Ten mL aliquots of orange juice, extracted from  
181 four different fruit in each bag, were transferred into 20 mL vials, sealed with crimp top fitted  
182 with a 2 mm rubber septum, and incubated at 30 °C for 10 min in a water bath. A one mL  
183 sample of the head space was injected in a gas chromatograph (Series 580, GOW MAC,  
184 Bethlehem, PA, USA) equipped with a flame ionization detector and a stainless steel (1.2 m ×  
185 3 mm) filled with Porapak® QS 80/100 column, with nitrogen used as a carrier gas at 30 mL  
186 min<sup>-1</sup>, hydrogen at 19 mL min<sup>-1</sup> and the air flow at 300 mL min<sup>-1</sup>. The column, injector and  
187 detector temperatures were set at 142, 164 and 163 °C, respectively. A 10 mL of solution  
188 containing ethanol and acetaldehyde at 100 µL L<sup>-1</sup> in 20 mL sealed vial was incubated at the  
189 same temperature and used as internal standards for quantity evaluations. The measurements  
190 for standard and samples were made in quadruplicate.

191 2.6.7. *Subjective fruit quality assessments*

192 2.6.7.1. *Peel pitting index (PPI)*

193 Fruit were visually scored to estimate the extent of peel pitting development after each storage  
194 time. Fruit were rated on a scale using the following scores: 0 = no pits, 1 = 1–30 % pitting, 2  
195 = 31–50 % pitting, 3 = severe pitting or > 50 % and the peel pitting index was measured  
196 according to the following formula (Alfárez and Burns, 2004). The results were obtained by  
197 assessing all the fruit (n = 32) per treatment at each storage time.

198

199 
$$PPI = \frac{\sum(\text{rindstaining scale (0-3)} \times \text{number of fruit in each class})}{\text{total number of fruit}} \quad (2)$$

200 2.6.7.2. *Fruit decay rate index (DRI)*

201 The proportion of the decay rate index was evaluated using the following scores: 0 = no area  
202 decay, 1 = 0–10 % area decay, 2 = 11–30 % area decay, 3 = 31–50 % area decay and 4 = 51–  
203 100 % area decay (Wang et al., 2015). The fruit DRI was calculated for the total fruit (n = 32)  
204 per treatment at each storage time as:

$$205 \text{ DRI (\%)} = \frac{\sum(\text{decay grade}(0-4) \times \text{number of fruit at that grade})}{\text{highest level} \times \text{total number of fruit}} \times 100 \quad (3)$$

206 2.6.7.3. *Stem-end rind breakdown (SERB)*

207 The percentage of stem-end rind breakdown development was evaluated visually according to  
208 a four level scale: 0 = no symptoms present; 1 = slight or small symptoms; 2 = moderate or  
209 noticeable symptoms of 30–50 %; and 3 = severe symptoms or > 50 % affected. The SERB  
210 was calculated by assessing all the fruit (n = 32) per treatment at each storage time as follows  
211 (Pristijono et al., 2017b):

$$212 \text{ SERB (\%)} = \frac{\sum(\text{rot score}(0-3) \times \text{number of fruit at that grade})}{\text{highest level} \times \text{total number of fruit}} \times 100 \quad (4)$$

213 2.6.7.4. *Overall visual acceptability (OVA)*

214 Fruit visual acceptability was independently assessed based on a subjective four point scoring  
215 system; 4 = excellent (fresh and high quality fruit with glossy skin and no symptoms of  
216 dehydration, shriveling, and decay); 3 = good (marketable and acceptable fruit quality with  
217 slight shriveling and softness); 2 = not saleable but edible (fruit with moderate signs of  
218 shriveling, dryness, browning, and softness); and 1 = poor quality (fruit with severe signs of  
219 shriveling, significant softness, pitting, and decay) (Golding et al., 2015). The OVA was  
220 calculated as follows:

221 
$$\text{OVA (\%)} = \frac{\sum(\text{rot score}(1-4) \times \text{number of fruit at that grade})}{\text{highest level} \times \text{total number of fruit}} \times 100 \quad (5)$$

222 *2.6.8. Sensory evaluation*

223 Fruit sensory evaluation was performed before treatment (day 0) and after one week at 20 °C  
224 following removal from cold storage. The panel involved twelve staff from NSW Department  
225 of Primary Industries, Ourimbah (6 females and 6 males), aged between 25 and 65 years old  
226 and who are familiar with citrus sensory evaluation. Fruit were brought to room temperature  
227 and hand-peeled, cut in half cross-wise with one half used for sensory analysis and the other  
228 half used for other quality measurements. Fruit were separated into individual segments and  
229 two segments from two different fruit were presented to panelists in coded 60 mL plastic cups.  
230 At each tasting session, panelists were given a rating sheet containing information on the  
231 evaluation procedure, in addition to general verbal instructions and individual clarifications as  
232 required. Panelists were requested to rate their degree of liking for the samples overall flavor  
233 on a 9-point hedonic scale (1 = “dislike extremely”, 9 = “like extremely”). In addition, each  
234 panelist marked an unstructured 10 cm scale, with the anchor points ‘none’ and ‘very strong’  
235 for off-flavor and ‘not fresh at all’ and ‘very fresh’ for freshness, and sensory data were  
236 recorded as distances (mm) from the origin. Five samples at each tasting time were presented  
237 in a random sequence to prevent any positional bias. Panelists were required to cleanse their  
238 palate with a bite of low-salt saltine cracker, a sip of room temperature mineral water, and a  
239 small time lag between samples. The panelists average responses were considered for each  
240 attribute (Tietel et al., 2011).

241 *2.7. Statistical analysis*

242 All analyses were performed in quadruplicate. Sources of variation were storage time and  
243 treatment. The results were statistically assessed by analysis of variance (ANOVA) and  
244 Multiple Ranges Duncan’s test to determine whether differences among treatments and storage

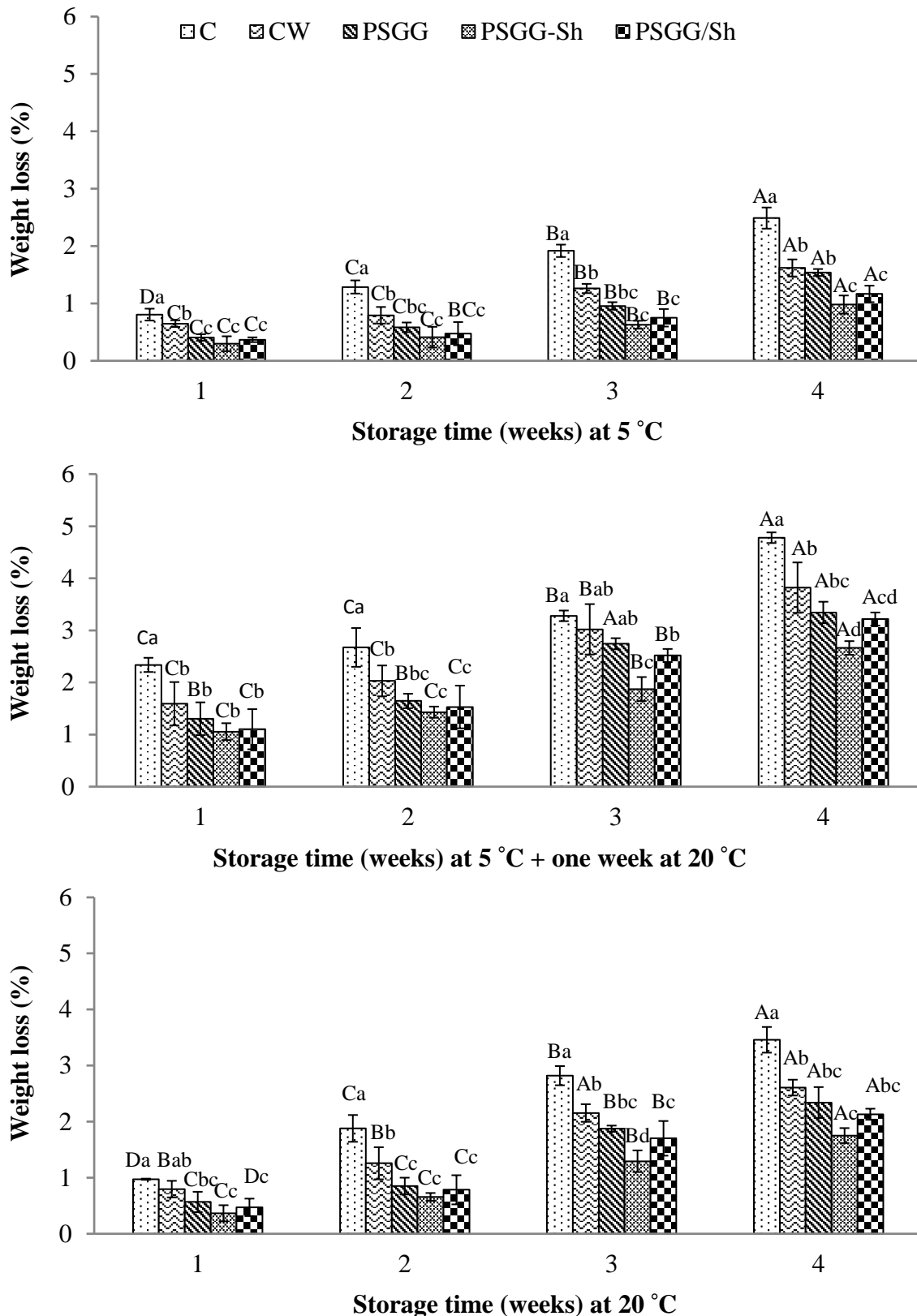
245 times were significant at  $P < 0.05$  using the software SPSS (version 23, SPSS Inc., Chicago,  
246 IL, USA).

### 247 **3. Results and discussion**

#### 248 *3.1. Weight loss*

249 Applied coatings significantly reduced water loss at most temperatures and storage times (Fig.  
250 1), which is an expected and commercially desirable result. The influence of polysaccharide  
251 based coatings on the weight loss is probably associated with the existence of hydroxyl groups  
252 creating hydrogen bonds both inside the coating matrix and with the cuticle on the peel, which  
253 mostly consists of cutin, a polyester polymerized from hydroxylated fatty acids (Arnon et al.,  
254 2015; Koch and Ensikat, 2008).

255 Upon removal from storage at 5 °C, water loss from non-coated fruit was always greater than  
256 the treated fruit, but with the additional week at 20 °C, water loss increased in all treatments.  
257 Similar results were observed in fruit stored constantly at 20 °C. In general, the addition of the  
258 Sh into PSSGG resulted in lower fruit weight loss, suggesting greatest benefit than the bilayer  
259 PSGG/Sh coating due to its likely higher moisture barrier capacity. The apparent synergistic  
260 effect between glycerol, Tween-20, and OA in the blended composite coatings is reported to  
261 result in a more compact and homogenous matrix (Rodríguez et al., 2006), reduce pores and  
262 cracks of films (García et al., 1999), and consequently decrease fruit weight loss in mandarin  
263 fruit (Rojas-Argudo et al., 2009). At constant 20 °C storage, all coatings reduced water loss  
264 after two weeks storage with similar trends noted at the 5 °C followed by one week at 20 °C  
265 storage.



266 **Fig. 1.** Weight loss of 'Valencia' oranges stored at different storage conditions for four weeks as affected by  
 267 various coatings treatments. Each bar represents the means of four replicates of 8 fruit each ( $n = 32$ )  $\pm$  standard  
 268 error. The different lowercase superscript letters in the same storage time indicate significant differences within  
 269 different coating treatments according to Duncan's test ( $P < 0.05$ ). The different uppercase superscript letters in  
 270 the same coating treatment indicate significant differences within different storage time according to Duncan's  
 271 test ( $P < 0.05$ ).

272 3.2. *Fruit firmness*

273 Fruit treated with the blended composite PSGG-Sh coating were significantly firmer than  
274 untreated control at most storage times and temperatures except at one week storage at 5 °C in  
275 which there was little treatment effects (Fig. 2). This related generally to lower weight loss  
276 levels for the same treatment as described above. The loss of fruit firmness for PSGG-Sh coated  
277 fruits after four weeks storage period at constant 5 °C, at 5 °C followed by 7 d at 20 °C, and  
278 constant 20 °C was 0.9 %, 4 %, and 2 %, respectively, with respect to the initial force value of  
279 fruit before treatment ( $46.70 \pm 3.33$  N), whereas the loss in firmness for untreated fruit stored  
280 under the same conditions were 5 %, 15 % and 9 %. The firmness retention of PSGG coating  
281 alone was similar to that of CW in all storage assessments. Moreover, in spite of the good  
282 weight loss inhibition presented by the bilayer PSGG/Sh coating, this coating showed similar  
283 firmness losses compared with single layer PSGG coating during storage. This may be  
284 explained by mechanical tensile strength attributes of PSGG film (Saber et al., 2016a).

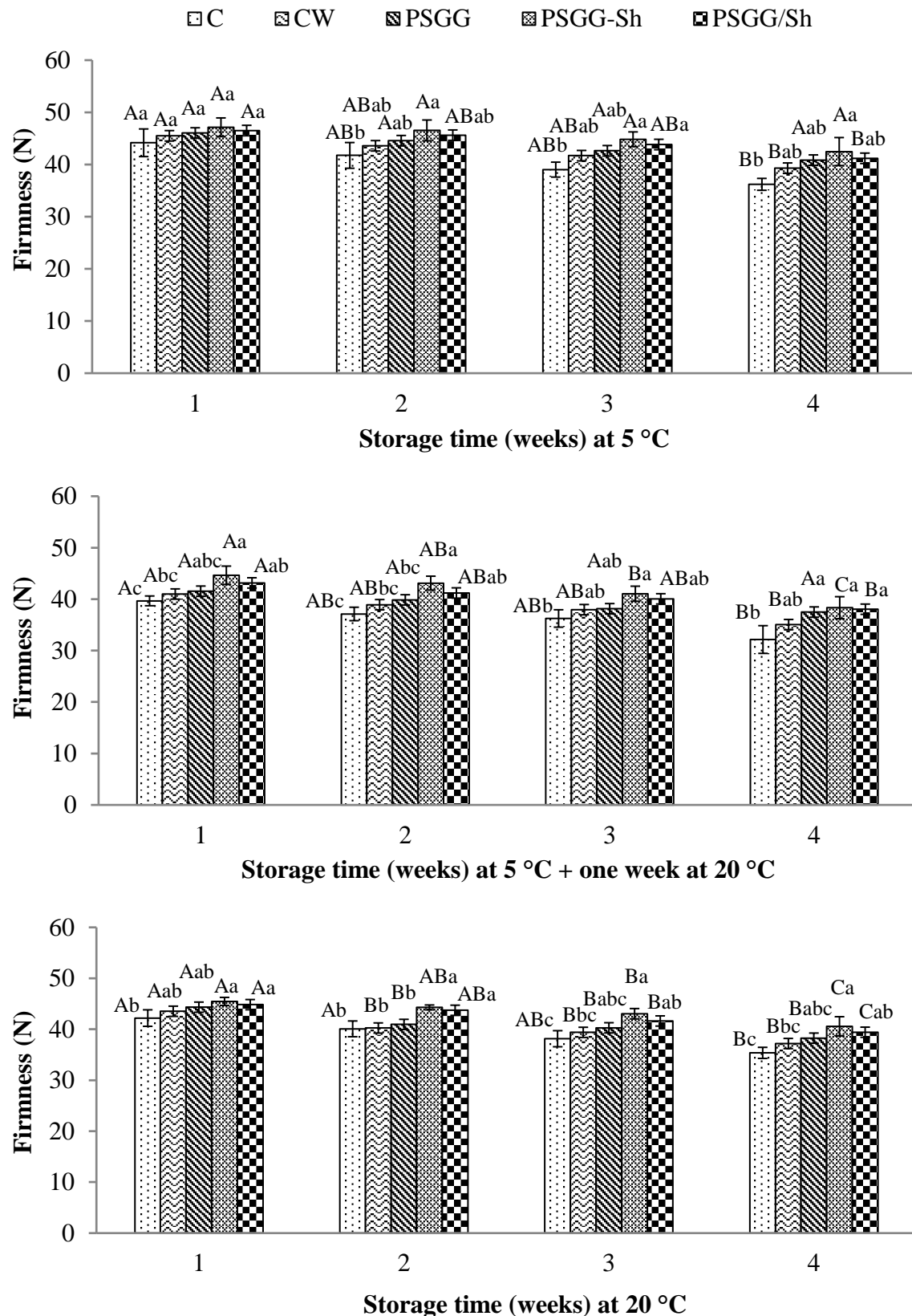
285 The loss of fruit firmness is influenced by the water loss which is considered as main  
286 parameter for texture changes (Del-Valle et al., 2005). In this experiment, the reduction in  
287 firmness losses with the coatings was probably due to the restriction of moisture loss and the  
288 moisture migration from the cells to the surrounding atmosphere through transpiration  
289 (Mahfoudhi and Hamdi, 2015).

290

291

292

293



294 **Fig. 2.** Firmness of 'Valencia' oranges stored at different storage conditions for four weeks as affected by various  
 295 coatings treatments. Each bar represents the means of four replicates of 8 fruit each ( $n = 32$ )  $\pm$  standard error. The  
 296 different lowercase superscript letters in the same storage time indicate significant differences within different  
 297 coating treatments according to Duncan's test ( $P < 0.05$ ). The different uppercase superscript letters in the same  
 298 coating treatment indicate significant differences within different storage time according to Duncan's test ( $P <$   
 299  $0.05$ ).



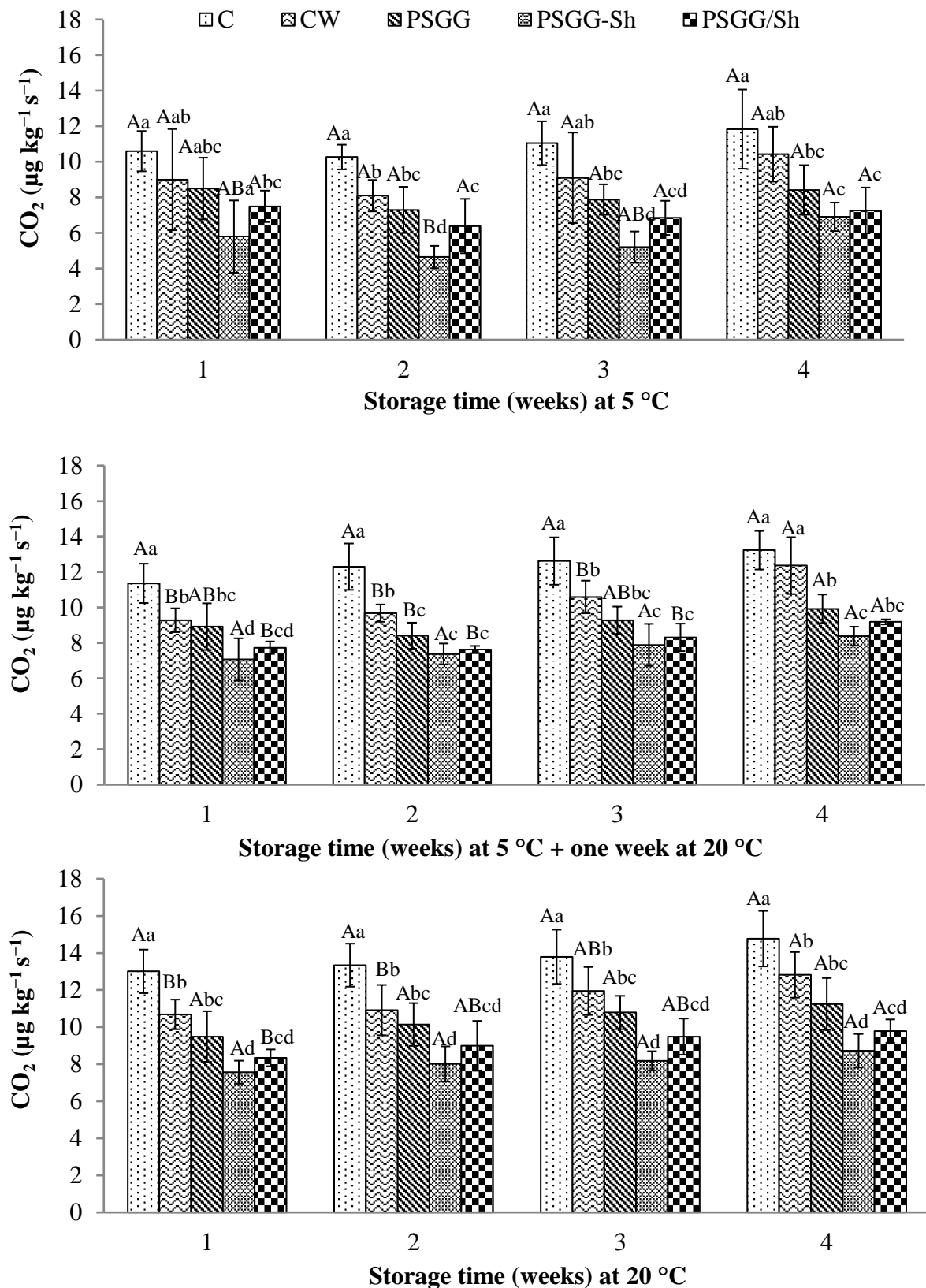
### 300 3.3. *Respiration rate*

301 To estimate the effect of polysaccharide based coatings on respiration rate of oranges during  
302 different storage temperatures, CO<sub>2</sub> concentration in the headspace was calculated (Fig. 3). The  
303 fruit respiration rate was  $13.93 \pm 0.72 \mu\text{g CO}_2 \text{ kg}^{-1} \text{ s}^{-1}$  at the beginning of the experiment and  
304 it decreased in all treatments after storage at 5 °C and 20 °C. Citrus are considered a non-  
305 climacteric fruit, i.e. ethylene production and respiration rates generally do not substantially  
306 increase during ripening and senescence (Wills and Golding, 2016).

307 Coated oranges generally had lower respiration rate than the uncoated control fruit (Fig. 3),  
308 likely due to the modification of internal gas atmosphere by the coatings (Cisneros- Zevallos  
309 and Krochta, 2002). Similar observations have been described by Arnon et al. (2015),  
310 Valencia-Chamorro et al. (2009), and Cháfer et al. (2012) in oranges coated with  
311 polysaccharide based edible coatings. In this experiment, the incorporation of hydrophobic  
312 compounds (oleic acid and shellac) into the coating formulation resulted in decreases in the  
313 respiration rates of treated fruit, such that the PSGG with Sh coated oranges had consistently  
314 lower respiration rates than the commercial wax and the untreated control fruit at all storage  
315 conditions. Similarly, the respiration rate of mandarins and oranges coated by hydroxypropyl  
316 methylcellulose edible coatings containing oleic acid and shellac has been shown to decrease  
317 compared to untreated control fruit (Navarro-Tarazaga et al., 2008; Valencia- Chamorro et al.,  
318 2010).

319 The addition of a shellac layer on PSGG coating generally resulted in lower respiration rates  
320 than those measured for oranges coated by single layer PSGG coatings. A similar trend of  
321 reduced respiration rate upon application of the LBL method was shown in previous studies  
322 with citrus fruit (Arnon et al., 2015; Poverenov et al., 2014).

323



324 **Fig. 3.** Respiration rate of 'Valencia' oranges stored at different storage conditions for four weeks as affected by  
 325 various coatings treatments. Each bar represents the means of four replicates of 8 fruit each ( $n = 32$ )  $\pm$  standard  
 326 error. The different lowercase superscript letters in the same storage time indicate significant differences within  
 327 different coating treatments according to Duncan's test ( $P < 0.05$ ). The different uppercase superscript letters in  
 328 the same coating treatment indicate significant differences within different storage time according to Duncan's  
 329 test ( $P < 0.05$ ).

330 *3.4. Ethylene production*

331 Ethylene production was  $4.92 \pm 0.32$  ng C<sub>2</sub>H<sub>4</sub> kg<sup>-1</sup> s<sup>-1</sup> at the beginning of the experiment and  
332 generally increased during storage (Fig. 4). However, ethylene production in either uncoated  
333 or coated oranges did not exceed 32 ng C<sub>2</sub>H<sub>4</sub> kg<sup>-1</sup> s<sup>-1</sup>. That was expected as citrus are non-  
334 climacteric fruit and the ethylene production rate generally do not considerably increase during  
335 storage (Wills and Golding, 2016). In this experiment, coated fruit generally had lower ethylene  
336 production rates than uncoated ones as expected. In general, fruit coated with PSGG-Sh  
337 produced less ethylene than fruit from most of the other treatments.

338 *3.5. Skin color*

339 There were no significant changes in the peel color as described by the hue angle and measured  
340 with a color meter (data not shown). The coatings treatments did not significantly affect fruit  
341 skin color during storage.

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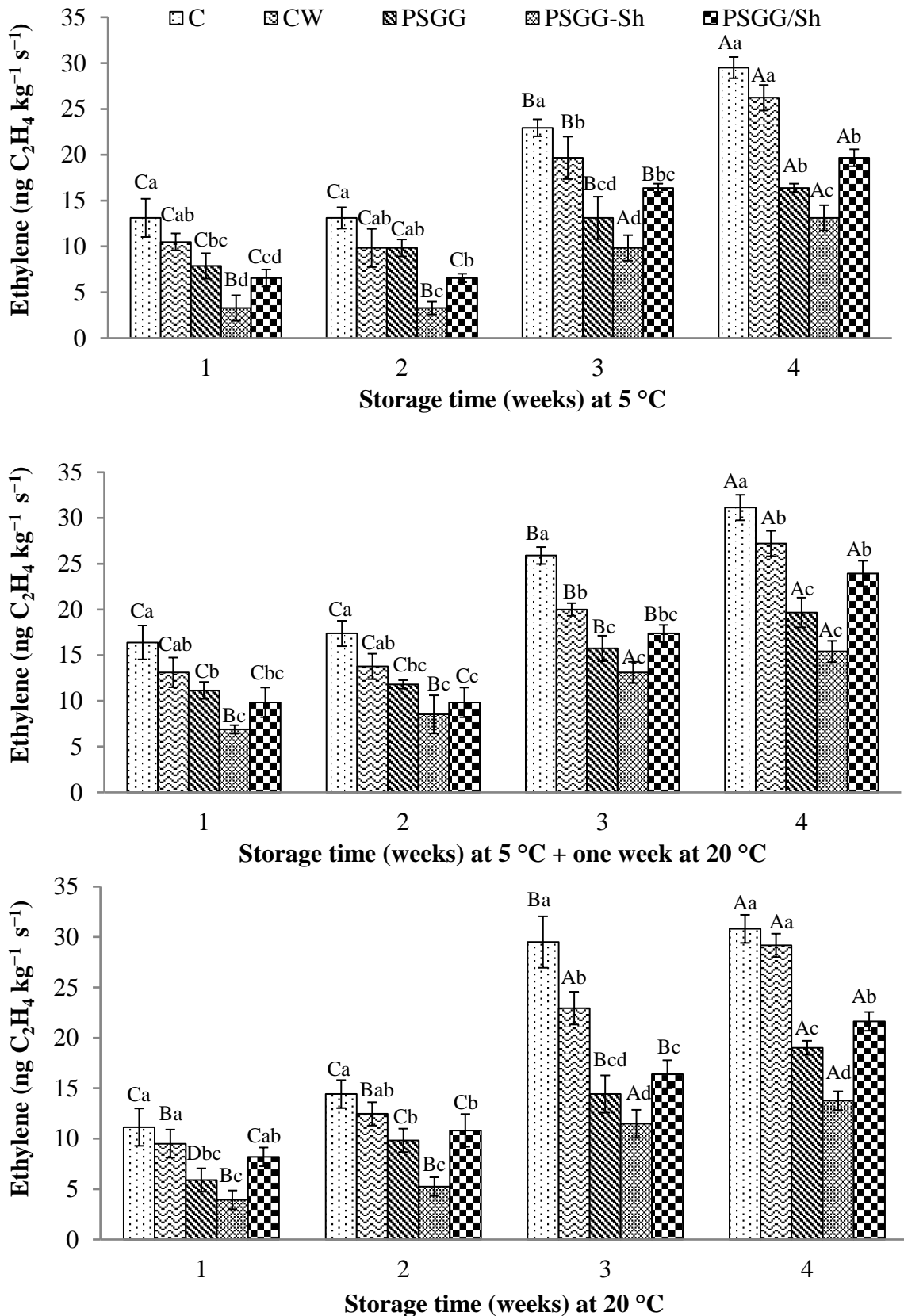
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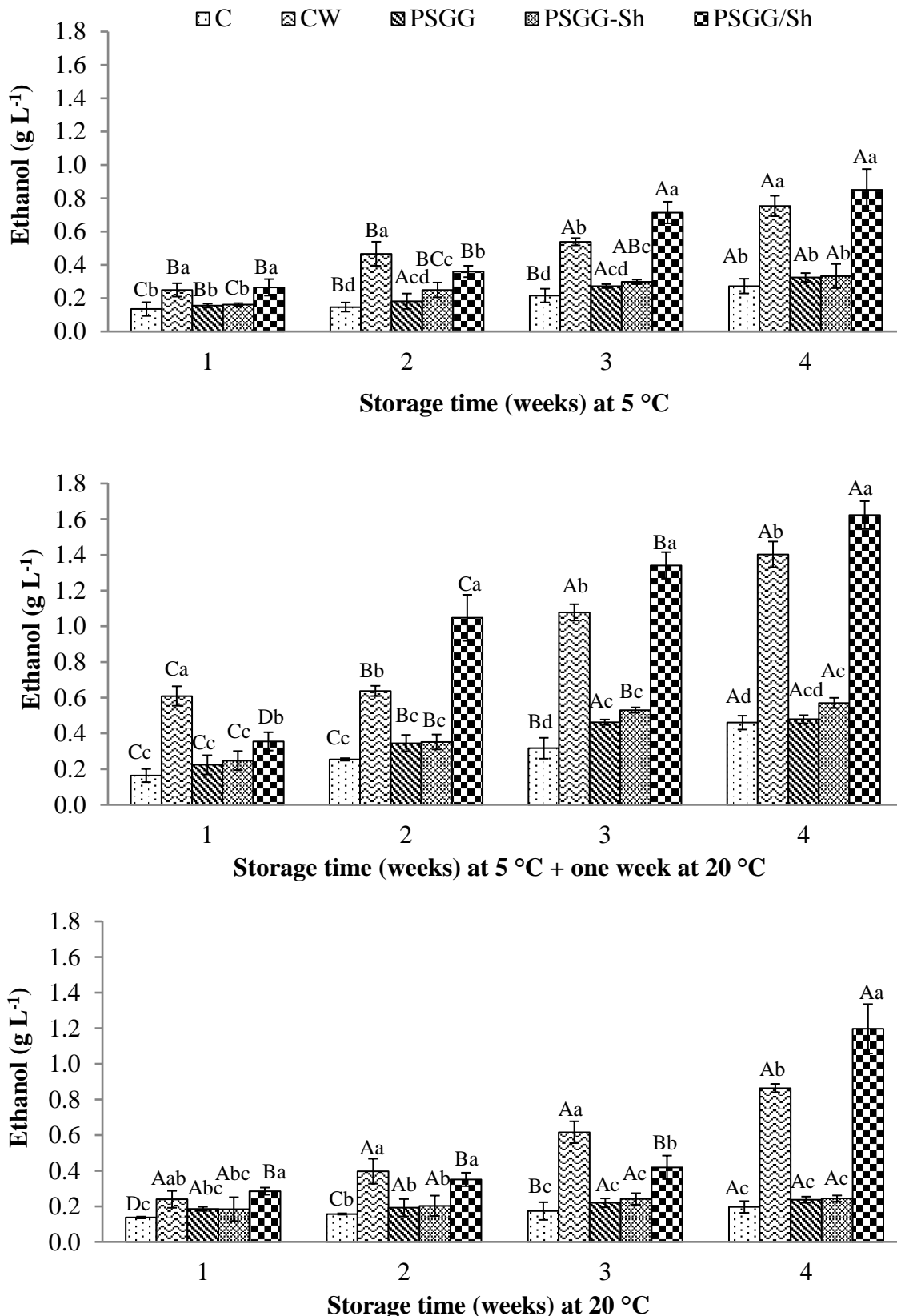
351 **Fig. 4.** Ethylene production of 'Valencia' oranges stored at different storage conditions for four weeks as affected  
 352 by various coatings treatments. Each bar represents the means of four replicates of 8 fruit each ( $n = 32$ )  $\pm$  standard  
 353 error. The different lowercase superscript letters in the same storage time indicate significant differences within  
 354 different coating treatments according to Duncan's test ( $P < 0.05$ ). The different uppercase superscript letters in  
 355 the same coating treatment indicate significant differences within different storage time according to Duncan's  
 356 test ( $P < 0.05$ ).

357 *3.6. Ethanol and acetaldehyde concentrations in fruit juice*

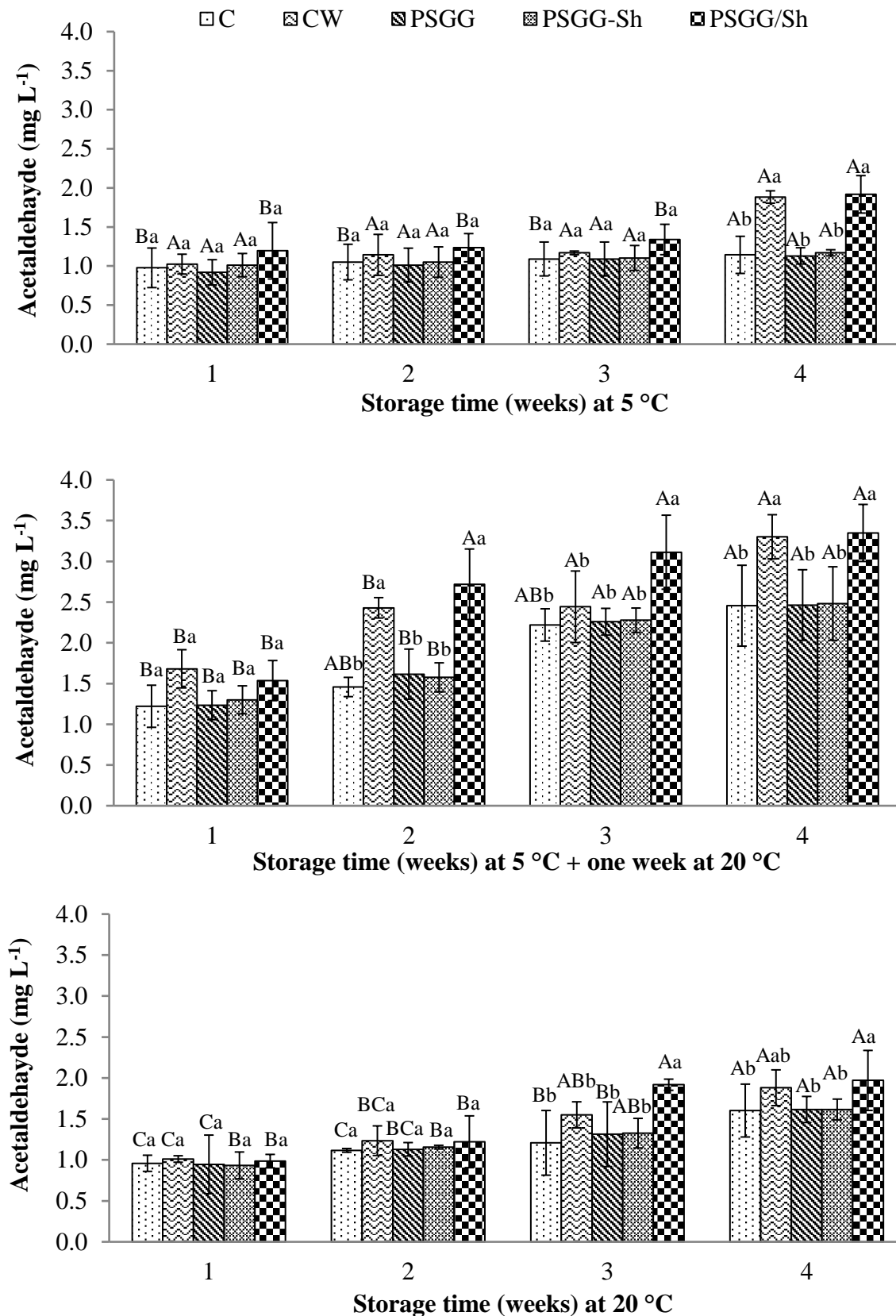
358 The concentrations of ethanol and acetaldehyde in the headspace of the juice of ‘Valencia’  
359 orange fruit at the beginning of the experiment were  $0.13 \pm 0.02 \text{ g L}^{-1}$  and  $0.95 \pm 0.12 \text{ mg L}^{-1}$ ,  
360 respectively (Figs. 5 and 6). In all cases, the ethanol and acetaldehyde concentrations increased  
361 during storage at both 5 °C and 20 °C storage. The production of both volatiles naturally occurs  
362 in fruit and their increased levels have been associated with off-flavors in citrus fruit  
363 (Hagenmaier, 2002).

364 Higher levels of ethanol have been generally reported in coated oranges than in the uncoated  
365 fruit, confirming the development of a modified atmosphere inside the fruit (Valencia-  
366 Chamorro et al., 2009). Though, ethanol concentration in coated oranges did not surpass the  
367 maximum set up at  $2000 \text{ mg L}^{-1}$  as the concentration of off-flavor build-up risk (Rojas-Argudo  
368 et al., 2009). The ethanol concentrations in the juice of bilayer PSGG/Sh-coated fruit were  
369 significantly higher than in the juice of CW-coated fruit after prolonged cold storage followed  
370 by further one week storage at 20 °C. This can be likely attributed to the lower gas permeability  
371 that bilayer coatings created in the fruit. It has been reported that citrus fruit coated with shellac-  
372 based commercial waxes commonly have higher ethanol concentrations than uncoated samples  
373 (Contreras- Oliva et al., 2012). Variations in the ethanol content between the experimental  
374 coatings single layer PSGG and blended composite PSGG-Sh, and the uncoated oranges were  
375 similar, especially after two weeks storage at 20 °C. Moreover, there was generally little  
376 significant differences between coatings treatments on acetaldehyde levels in ‘Valencia’  
377 orange juice especially in weeks one and two; however, bilayer and CW coatings generally  
378 resulted in higher acetaldehyde content in orange juice as the storage period increased.

379



380 **Fig. 5.** Ethanol concentration in the juice of ‘Valencia’ oranges stored at different storage conditions for four  
 381 weeks as affected by various coatings treatments. Each bar represents the means of four replicates of 8 fruit each  
 382 ( $n = 32$ )  $\pm$  standard error. The different lowercase superscript letters in the same storage time indicate significant  
 383 differences within different coating treatments according to Duncan’s test ( $P < 0.05$ ). The different uppercase  
 384 superscript letters in the same coating treatment indicate significant differences within different storage time  
 385 according to Duncan’s test ( $P < 0.05$ ).



386 **Fig. 6.** Acetaldehyde concentration in the juice of ‘Valencia’ oranges stored at different storage conditions for  
 387 four weeks as affected by various coatings treatments. Each bar represents the means of four replicates of 8 fruit  
 388 each ( $n = 32$ )  $\pm$  standard error. The different lowercase superscript letters in the same storage time indicate  
 389 significant differences within different coating treatments according to Duncan’s test ( $P < 0.05$ ). The different  
 390 uppercase superscript letters in the same coating treatment indicate significant differences within different storage  
 391 time according to Duncan’s test ( $P < 0.05$ ).

392 *3.7. Subjective fruit quality assessments*

393 The incidence of peel pitting in coated and uncoated ‘Valencia’ oranges generally increased  
394 with longer storage time (Fig. 7). Higher temperature during storage favored the incidence of  
395 peel pitting in ‘Valencia’ oranges. The uncoated fruit showed the highest PPI value, which  
396 increased from 0.06 to 0.37 and 0.09 to 0.44 during four weeks storage at 5 °C and ambient  
397 temperature, respectively. The rate of increase in the PPI varied among fruit treated with  
398 various coatings stored at different temperatures. No peel pitting was observed in fruit treated  
399 with PSGG-Sh coating after three weeks storage at 5 °C. By four weeks storage at 5 °C the PPI  
400 of fruit coated with PSGG-Sh was 0.09; while that of fruit coated with PSGG and bilayer  
401 PSGG/Sh was 0.13 and 0.28, respectively. Transferring fruit from 5 °C followed by one week  
402 storage at 20 °C increased postharvest peel pitting, possibly because of the increased rate of  
403 dehydration (Alfárez and Burns, 2004). However, it was hard to distinguish the difference  
404 between peel pitting and chilling injury in fruit storage at 5 °C and with the additional storage  
405 for one week at 20 °C, and both considered as physiological disorder and results were reported  
406 together. Peel pitting developed in all fruit after two weeks storage at 5 °C followed by one  
407 week storage at 20 °C.

408 In general, CW and bilayer PSGG/Sh coatings resulted in more pitting on the coated fruit  
409 surface, however the PPI was lower in bilayer coated fruit. Application of more gas-permeable  
410 coatings along with storage at cool temperature appeared to have postponed postharvest peel  
411 pitting in this study. These results indicate that faster water permeability and therefore a faster  
412 water position adjustment in the albedo and flavedo of the fruit peel may alleviate peel pitting  
413 development (Cronjé et al., 2017). Moreover, decreases in internal O<sub>2</sub> and increases in internal  
414 CO<sub>2</sub> in coated fruit can be effective in reducing peel pitting disorder, although the relationship  
415 between the level of pitting and internal CO<sub>2</sub> and O<sub>2</sub> levels in fruit may not be as strong (Alfárez  
416 and Burns, 2004; Petracek et al., 1998). Regardless of higher weight loss value in fruit coated

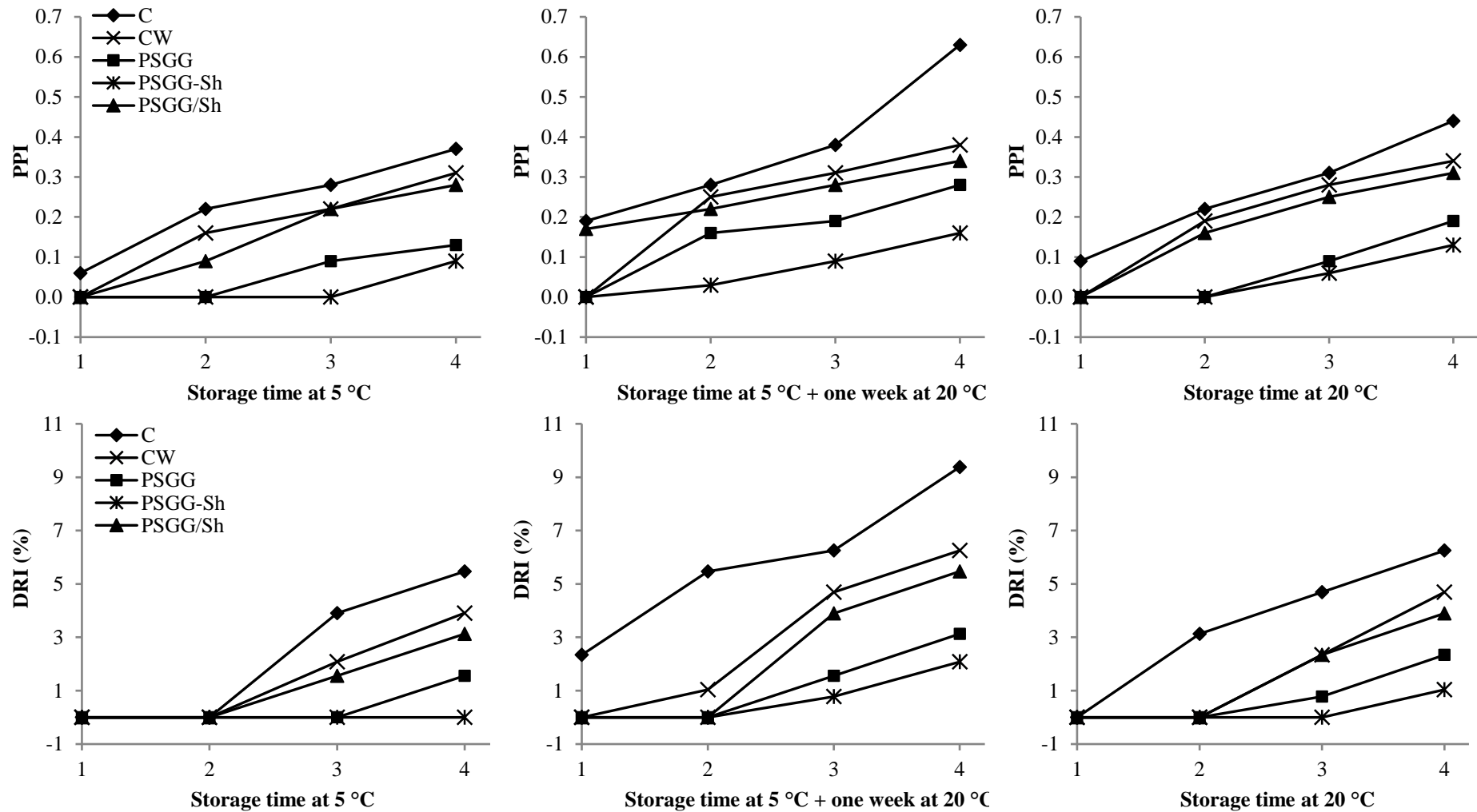


417 with PSGG compared with bilayer coated fruit, subsequent dehydration of orange peel coated  
418 with PSGG did not appear to be enough to stimulate postharvest peel pitting. These results  
419 suggest that a variety of factors including peel maturity and senescence may be involved in  
420 postharvest peel pitting in 'Valencia' oranges.

421 Decay due to natural infection during the whole storage was relatively low as fruit were  
422 sanitized with a fungicide (Fig. 7). The coatings decreased DRI compared with control fruit in  
423 all storage conditions. The films and coatings can suspend decay by reducing senescence,  
424 which causes more susceptibility to pathogenic infection in produce due to damage of cellular  
425 or tissue integrity (Tanada-Palmu and Grosso, 2005). No visible sign of decay in coated or  
426 control fruit was observed until three weeks of the storage period at 5 °C (Fig. 7). Fruit treated  
427 with PSGG-Sh coating remained disease free during four weeks at 5 °C and three weeks at 20  
428 °C. Shelf life of the coated fruit was prolonged up to two weeks without decay at ambient  
429 temperature compared to only one week for uncoated fruit. Removal from the low temperature  
430 and transferring to ambient temperature with further storage of one week generally increased  
431 DRI in fruit from most treatments. At the first week of transferring fruit from 5 °C to 20 °C,  
432 2.3 % of the control fruit decayed and DRI increased to 9.4 % at the end of storage time.

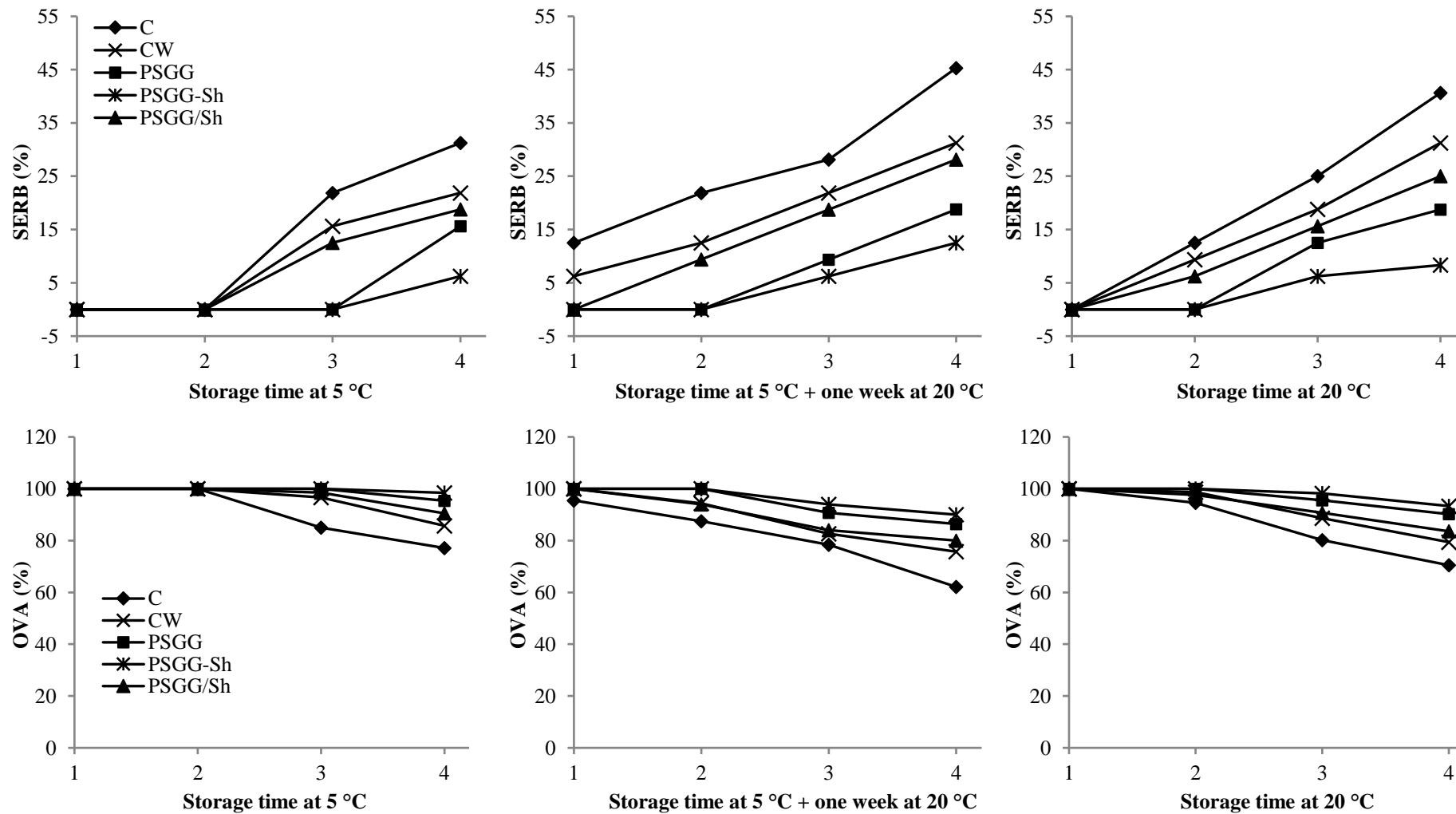
433 The incidence of SERB in fruit maintained at high temperature was higher compared with those  
434 stored at low temperature and control fruit indicated higher SERB value (Fig. 8). Transfer of  
435 fruit to 20 °C after one week storage at 5 °C also caused a noticeable increase in the occurrence  
436 of SERB. Fruit coated by PSGG and PSGG-Sh did not show any sign of SERB after three  
437 weeks storage at 5 °C and two weeks storage at 20 °C (Fig. 8).

438 The OVA of the control and coated fruit decreased throughout storage time. Fruit coated by  
439 PSGG-Sh showed the highest OVA followed by PSGG and bilayer PSGG/Sh-coated fruit at  
440 all storage circumstances (Fig. 8).



441

442 **Fig. 7.** Peel pitting index (PPI) and percentage of fruit decay rate index (DRI) of coated and uncoated 'Valencia' oranges stored at different storage  
 443 conditions for four weeks as affected by various coatings treatments. Each point is total value for n = 32 fruit per treatment at each storage time.



444

445 **Fig. 8.** The percentage of stem-end rind breakdown (SERB) and overall visual acceptability (OVA) of coated and uncoated ‘Valencia’ oranges  
 446 stored at different storage conditions for four weeks as affected by various coatings treatments. Each point is total value for n = 32 fruit per  
 447 treatment at each storage time.

448 3.8. *Sensory evaluation*

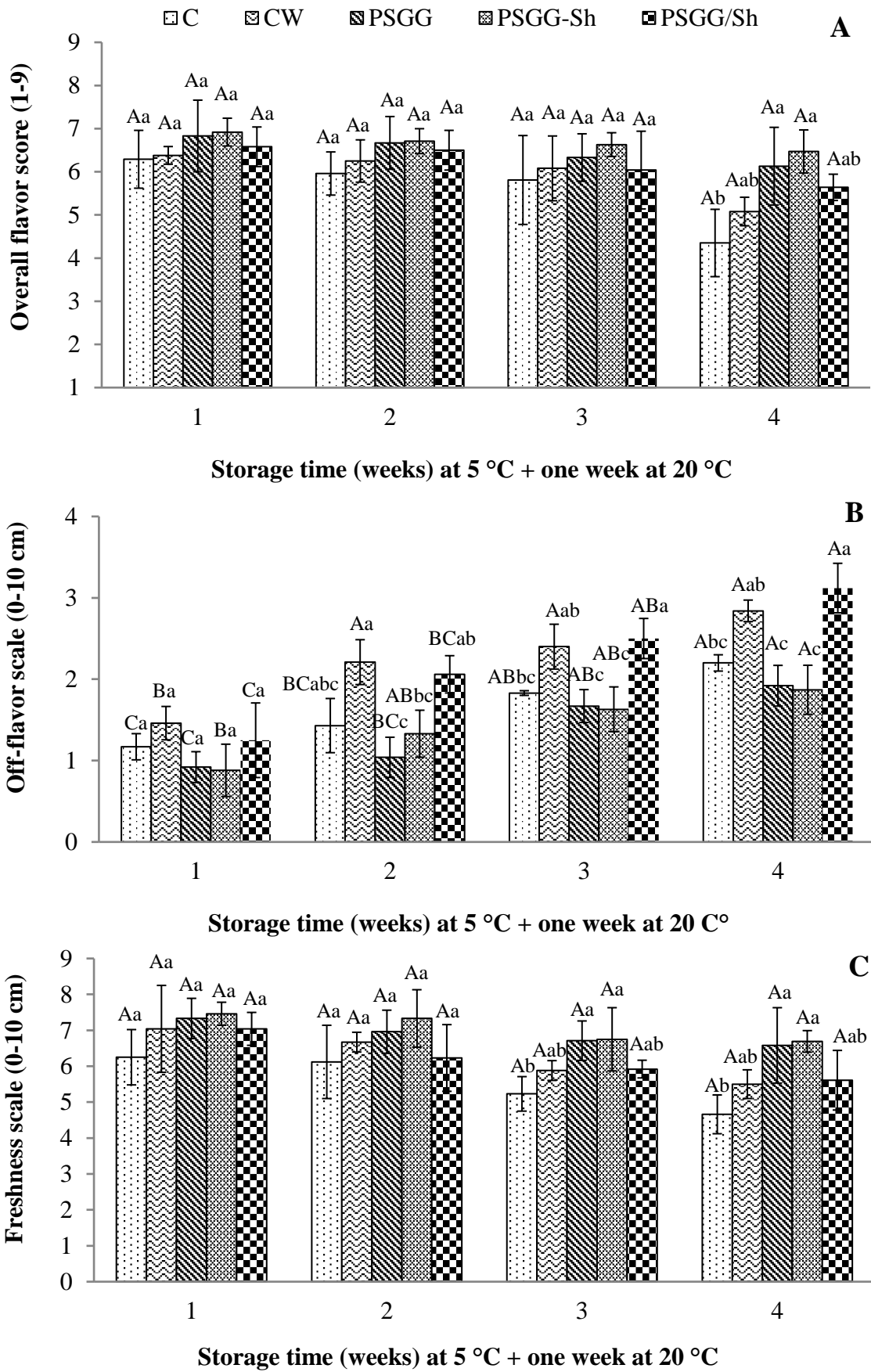
449 Overall flavor scores from fruit stored at 5 °C for four weeks and then held for one week at 20  
450 °C were significantly higher in PSGG and blended composite PSGG-Sh coated compared to  
451 the uncoated control fruit (Fig. 9A). In contrast, there were no differences in overall flavor  
452 between treated and untreated fruit for the first three weeks of storage at 5 °C. Overall flavor  
453 evaluation of uncoated oranges reduced with storage time, from 7.2 before storage to 4.4 at the  
454 end of the storage. Although only a relatively small sensory panel was used, these results  
455 provide an indication of what consumer likeability would be for both treated and untreated  
456 oranges at a likely consumption stage of the fruit.

457 The levels of off-flavor detected in fruit stored for 2, 3 and 4 weeks at 5 °C and then held for 1  
458 week at 20 °C were significantly lower in both the PSGG and PSGG-Sh coated fruit (and  
459 similar to control fruit) compared to CW and to PSGG/Sh (at 3 and 4 weeks) coated fruit (Fig.  
460 9B), the results were correlated with the ethanol and acetaldehyde levels determined in  
461 respective coated fruits. Treatment differences in off-flavor were not significant in fruit stored  
462 for one week at 5 °C and then held for one week at 20 °C. Off-flavor evaluation of uncoated  
463 oranges increased with storage time, from 0 before storage to 2.2 at the end of the storage.  
464 There was a general trend of off-flavor increasing with storage time for most treatments (Fig.  
465 9B).

466 Following 3 and 4 weeks storage at 5 °C plus one week at 20 °C, the fruit coated with single  
467 layer PSGG and blended composite PSGG-Sh were perceived as ‘fresher’ than the untreated  
468 control fruit (Fig. 9C). There were no differences in ‘freshness’ of any treatment for the first  
469 two weeks of storage at 5 °C followed by one week at 20 °C. Freshness evaluation of uncoated  
470 oranges reduced with storage time from 8.0 before storage to 4.7 at the end of the storage.

471 Applied wax or other surface coatings can alter the internal atmosphere in citrus fruit  
472 throughout the supply chain, leading to the accumulation of anaerobic metabolites such as  
473 ethanol and acetaldehyde, which have been associated with poor flavor in a number of studies  
474 (Baldwin et al., 1995; Obenland et al., 2008; Ummarat et al., 2015). Both applied coatings and  
475 cold storage are also reported to change a number of flavor-related aroma volatiles in citrus  
476 fruit, and similar effects could have contributed to the results found in this study. For example,  
477 compared with uncoated fruit, ‘Valencia’ oranges treated with a commercial shellac-based wax  
478 and stored at 16 or 21 °C for up to 56 d had higher concentrations of ethanol, ethyl butanoate,  
479 ethyl acetate, and alpha-pinene as time in storage increased, whereas levels of valencene, alpha-  
480 terpineol, and hexanol were generally lower, especially at the higher storage temperature  
481 (Baldwin et al., 1995). Likewise, flavor quality (i.e. both overall flavor likeability and  
482 freshness) of ‘Navel’ oranges stored at 5 °C for 3 or 6 weeks followed by 4 d at 13 °C and 3 d  
483 at 20 °C was reduced compared to non-stored fruit presumably due to lower levels of limonene  
484 and higher levels of ethyl butanoate, ethyl hexanoate and other four aroma-active compounds  
485 (Obenland et al., 2008).

486



487 **Fig. 9.** Sensory evaluation of ‘Valencia’ oranges stored at 5 °C for four weeks plus one week at 20 °C as affected by various coatings treatments. (A): Overall flavor, (B): Off-flavor, and (C): Freshness. The values represent means of twelve replicates ± standard error. The different lowercase superscript letters in the same storage time

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490 indicate significant differences within different coating treatments according to Duncan's test ( $P < 0.05$ ). The  
491 different uppercase superscript letters in the same coating treatment indicate significant differences within  
492 different storage time according to Duncan's test ( $P < 0.05$ ). Overall flavor was rated on a 9-point hedonic scale  
493 (1="dislike extremely", 9="like extremely"). Off-flavor and freshness were assessed based on an unstructured 10  
494 cm scale, with the anchor points 'none' and 'very strong' for off-flavor, and 'not fresh at all' and 'very fresh' for  
495 freshness.

496

#### 497 **4. Conclusion**

498 The results showed the benefit of applying edible coatings on the maintenance of fruit quality  
499 during storage and shelf life. The lower levels of respiration rates in coated fruit reflected the  
500 capability of the coating to modify the internal atmosphere of fruit as a protective gas barrier.  
501 The incorporation of lipid compounds into the PSGG coatings resulted in the optimum  
502 performance in reducing fruit respiration rate, ethylene production, weight and firmness loss,  
503 peel pitting, and fruit decay index rate of the coated oranges. Although the bilayer PSGG/Sh  
504 coating reduced weight loss and respiration rate with improved firmness retention to a greater  
505 extent than single layer PSGG coating, the bilayer coating also resulted in higher levels of  
506 ethanol causing increased perception of off-flavors. The sensory evaluation of the oranges  
507 showed that the fruit coated with PSGG with the incorporation of Sh and single layer PSGG  
508 coatings maintained overall flavor throughout shelf life with the panellists giving higher  
509 acceptance of freshness and flavor of the coated fruit. These results suggest that PSGG-based  
510 edible coatings could be a beneficial substitute to common commercial waxes for maintaining  
511 quality and extending shelf life of citrus fruit and potentially other fresh horticultural produce.  
512 Further research on the development of new formulations by addition of bioactive compounds  
513 to PSGG-based coating and the application of this coating on microbial growth and on the  
514 physiological processes of various climacteric/non-climacteric fruit and vegetables is of great  
515 interest.

#### 516 **Acknowledgement**

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519 authors acknowledge personnel of the NSW Department of Primary Industries for their  
520 technical assistance on this project.

## 521 **Conflict of Interest**

522 The authors declare no conflict of interest.

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675 **Figure captions**

676 **Fig. 1.** Weight loss of ‘Valencia’ oranges stored at different storage conditions for four weeks  
677 as affected by various coatings treatments. Each bar represents the means of four replicates of  
678 8 fruit each ( $n = 32$ )  $\pm$  standard error. The different lowercase superscript letters in the same  
679 storage time indicate significant differences within different coating treatments according to  
680 Duncan’s test ( $P < 0.05$ ). The different uppercase superscript letters in the same coating  
681 treatment indicate significant differences within different storage time according to Duncan’s  
682 test ( $P < 0.05$ ).

683 **Fig. 2.** Firmness of ‘Valencia’ oranges stored at different storage conditions for four weeks as  
684 affected by various coatings treatments. Each bar represents the means of four replicates of 8  
685 fruit each ( $n = 32$ )  $\pm$  standard error. The different lowercase superscript letters in the same  
686 storage time indicate significant differences within different coating treatments according to  
687 Duncan’s test ( $P < 0.05$ ). The different uppercase superscript letters in the same coating  
688 treatment indicate significant differences within different storage time according to Duncan’s  
689 test ( $P < 0.05$ ).

690 **Fig. 3.** Respiration rate of ‘Valencia’ oranges stored at different storage conditions for four  
691 weeks as affected by various coatings treatments. Each bar represents the means of four  
692 replicates of 8 fruit each ( $n = 32$ )  $\pm$  standard error. The different lowercase superscript letters  
693 in the same storage time indicate significant differences within different coating treatments  
694 according to Duncan’s test ( $P < 0.05$ ). The different uppercase superscript letters in the same  
695 coating treatment indicate significant differences within different storage time according to  
696 Duncan’s test ( $P < 0.05$ ).

697 **Fig. 4.** Ethylene production of ‘Valencia’ oranges stored at different storage conditions for four  
698 weeks as affected by various coatings treatments. Each bar represents the means of four

699 replicates of 8 fruit each ( $n = 32$ )  $\pm$  standard error. The different lowercase superscript letters  
700 in the same storage time indicate significant differences within different coating treatments  
701 according to Duncan's test ( $P < 0.05$ ). The different uppercase superscript letters in the same  
702 coating treatment indicate significant differences within different storage time according to  
703 Duncan's test ( $P < 0.05$ ).

704 **Fig. 5.** Ethanol concentration in the juice of 'Valencia' oranges stored at different storage  
705 conditions for four weeks as affected by various coatings treatments. Each bar represents the  
706 means of four replicates of 8 fruit each ( $n = 32$ )  $\pm$  standard error. The different lowercase  
707 superscript letters in the same storage time indicate significant differences within different  
708 coating treatments according to Duncan's test ( $P < 0.05$ ). The different uppercase superscript  
709 letters in the same coating treatment indicate significant differences within different storage  
710 time according to Duncan's test ( $P < 0.05$ ).

711 **Fig. 6.** Acetaldehyde concentration in the juice of 'Valencia' oranges stored at different storage  
712 conditions for four weeks as affected by various coatings treatments. Each bar represents the  
713 means of four replicates of 8 fruit each ( $n = 32$ )  $\pm$  standard error. The different lowercase  
714 superscript letters in the same storage time indicate significant differences within different  
715 coating treatments according to Duncan's test ( $P < 0.05$ ). The different uppercase superscript  
716 letters in the same coating treatment indicate significant differences within different storage  
717 time according to Duncan's test ( $P < 0.05$ ).

718 **Fig. 7.** Peel pitting index (PPI) and percentage of fruit decay rate index (DRI) of coated and  
719 uncoated 'Valencia' oranges stored at different storage conditions for four weeks as affected  
720 by various coatings treatments. Each point is total value for  $n = 32$  fruit per treatment at each  
721 storage time.

722 **Fig. 8.** The percentage of stem-end rind breakdown (SERB) and overall visual acceptability  
723 (OVA) of coated and uncoated ‘Valencia’ oranges stored at different storage conditions for  
724 four weeks as affected by various coatings treatments. Each point is total value for  $n = 32$  fruit  
725 per treatment at each storage time.

726 **Fig. 9.** Sensory evaluation of ‘Valencia’ oranges stored at 5 °C for four weeks plus one week  
727 at 20 °C as affected by various coatings treatments. (A): Overall flavor, (B): Off-flavor, and  
728 (C): Freshness. The values represent means of twelve replicates  $\pm$  standard error. The different  
729 lowercase superscript letters in the same storage time indicate significant differences within  
730 different coating treatments according to Duncan’s test ( $P < 0.05$ ). The different uppercase  
731 superscript letters in the same coating treatment indicate significant differences within different  
732 storage time according to Duncan’s test ( $P < 0.05$ ). Overall flavor was rated on a 9-point  
733 hedonic scale (1=“dislike extremely”, 9=“like extremely”). Off-flavor and freshness were  
734 assessed based on an unstructured 10 cm scale, with the anchor points ‘none’ and ‘very strong’  
735 for off-flavor, and ‘not fresh at all’ and ‘very fresh’ for freshness.

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