

Fabry Disease: A New Model of Premature Ageing?

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An increase in biological ageing is now commonly recognized in chronic diseases such as chronic kidney disease and chronic obstructive pulmonary disease [1, 2]. In this issue, Vujkovic et al. [3] present data suggesting that Fabry disease may be a disease associated with premature ageing. The authors assessed telomere length (TL) and telomerase activity in a cohort of 33 patients with Fabry disease. Whereas in male patients, the authors observed a reduced TL as compared to age-matched controls, females presented with a higher telomerase activity without a significant difference in TL. The authors did not observe a difference in TL or telomerase between patients with or without enzyme replacement therapy (ERT), nor did they observe a relationship between TL and renal function. Telomere attrition is a feature of ESRD associated with persistent inflammation that predicts poor outcome [4].

Fabry disease is associated with a significantly reduced life expectancy compared to the general population [5], and therefore, the association with a premature ageing process is not unexpected. However, establishing biological age is not straightforward. Both phenotypical criteria and biomarkers are used for this purpose. Whereas phenotypically premature ageing is characterized by frailty, sarcopenia, reduced physical function, and cognitive dysfunction; these are not definite criteria for its diagnosis [6]. Whereas a reduced executive function and informa-

tion processing speed were observed in patients with Fabry disease, and global cognition appeared to be preserved [7]. We could not identify studies from the literature assessing frailty, body composition, or physical function in adults with Fabry disease.

Regarding ageing biomarkers, telomere attrition, which was measured in this study, has been associated with outcomes at a population level in different disease states, but is generally considered to be an insensitive biomarker [8]. The ageing process is typified by significant interindividual variation and differences in physiological function between individuals of the same chronological age. This holds also for TL. The case for TL as a biomarker of ageing has been made repeatedly [9]. Systematic reviews of the evidence relating TL to outcome have found few actual mortality studies, all of which suffered from survivor bias. Few studies have examined the relationship with age-related decline in physiological function. Data from these were equivocal and all lacked statistical power [10, 11]. TL as a biomarker of age-related health is beset by methodological issues and has proven inferior to the use of CDKN2A [12, 13].

An exciting alternative has been the use of methylation-based epigenetic clocks, which show good correlations with chronological age and acceleration in disease states [14]. Caveats to their use, however, include their degree of fit with physiological function in normative

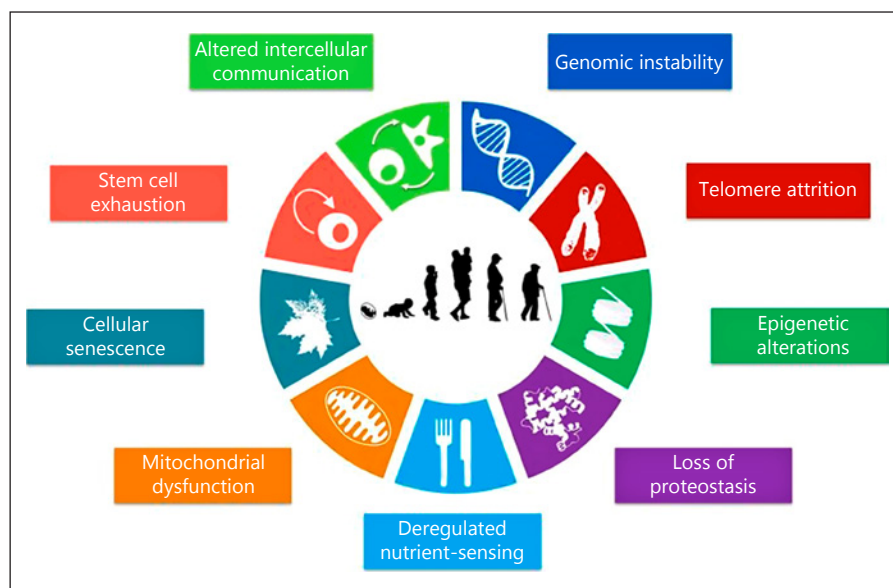


Fig. 1. Hallmarks of aging. From [17], with permission.

ageing, a limited understanding of the significance of their respective methylation sites and the underlying mechanism. Additionally, the associated methylation sites lack conservation between species.

More recently, a ribosomal DNA-based clock has been described [15], with a proposed mechanism based on DNA methylation counteracting increased nucleolar size and rDNA transcription levels. This is also consistent with the MTR (mitochondrial, telomere nucleo-protein complexes, and control of ribosome synthesis) theory of ageing, as described in [16].

At a molecular and cellular level, 9 hallmarks are associated with the ageing process, namely, genomic instability, telomere attrition, altered intracellular communication (such as due to inflammation), epigenetic alterations, loss of proteostasis, deregulated nutrient sensing, mitochondrial dysfunction, cellular senescence, and stem cell exhaustion [17] (Fig. 1).

In patients with Fabry disease or in experimental models, dysregulation of several of these hallmarks has been identified. Whereas telomere attrition was observed in the study of Vujkovic et al. [3] and Squillaro et al. [18] observed DNA damage in a mesenchymal stem cell line despite the activation of DNA damage.

Regarding altered intracellular communication, Fabry disease is characterized by chronic low-grade systemic inflammation [19], resulting from an interaction between Gb3 and toll-like receptors, which likely contributes to tissue damage [20]. In the study of Vujkovic et al. [3], no relation between levels of proinflammatory biomarkers and TL was observed. However, given the small

sample size, this does not necessarily suggest that systemic inflammation is not involved in the ageing process given the cross-sectional nature of the study. With respect to loss of proteostasis, dysregulated autophagy, which serves for the removal of cellular waste products, has been identified in cellular cultures with α -Gal A depletion [21]. Furthermore, nutrient sensing did seem to be in a podocyte cell culture model in Fabry disease, given the lower expression of mechanistic target of rapamycin. Nutrient sensing in the ageing process is a complicated phenomenon. Whereas activation of the anabolic pathways such as mechanistic target of rapamycin leads to accelerated ageing in experimental models, a downregulation of this pathway can be a defense mechanism under cell stress, preserving energy for cellular survival [17, 22].

With respect to mitochondrial dysfunction, a recent study has shown higher plasma markers reflecting oxidative stress in patients with Fabry disease on ERT [23], which is possibly in part related to superoxide dismutase 2 downregulation [24]. Squillaro et al. [18] showed that α -Gal A deficiency induced cell cycle arrest and senescence in cultures of amniotic and bone-marrow derived mesenchymal stem cells. In conclusion, there are several arguments for a disturbance in the “hallmarks” of ageing in patients and models of Fabry disease, although definitely more research is needed.

ERT did not appear to affect telomere attrition. Indeed, it has been shown that ERT can delay disease manifestation, and it is not able to completely prevent

the progression of organ damage in Fabry disease [25]. Inflammatory markers were even higher on patients receiving ERT, which might be a consequence of confounding by indication. An important characteristic of α -Gal A-depleted cell lines is an impairment of autophagy and protein turnover, leading to an increase in oxidative stress, which does not seem to be fully reversible by ERT [21, 23, 26]. The role of defective autophagy in the ageing process is increasingly recognized [27].

In a recent study, ERT cleared Gb3 from podocytes in cell cultures but was not able to normalize dysregulated autophagy, TGF- β expression, or oxidative stress, suggesting that other as yet incompletely identified mechanisms beyond Gb3 accumulation also are involved in the organ damage of FD [21, 23]. Also, ERT did not appear to be able to normalize the decreased energy metabolism in pluripotent stem cell-derived cardiomyocytes that were reprogrammed with a mutated GLA gene of a patient with Fabry disease [28].

Despite existing controversy around this subject [29], it has been suggested that skewing of X chromosome inactivation affects the phenotype of female patients with FD [29]. Whereas patients with random inactivation showed a progressive course with advancing age, Echevarria et al. [30] have observed that those with skewed inactivation displayed either a mild course or a rapidly progressive course of the disease depending on the expression of the mutant GLA allele. Ageing has been shown to induce skewing of lead X chromosome expression, which has been implicated in the pathogenesis of several auto-immune disorders [31]. It might be hypothesized that biological ageing, by inducing skewing of X chromosome inactivation, may also play a role in the progression of FD and its phenotypic expression in females in a mutually reinforcing way. However, this assumption needs to be addressed in future studies.

The finding that females, who usually have a milder form of disease, had telomere levels comparable to controls, but higher telomerase levels are intriguing. It was suggested that a higher telomerase level serves as a temporary compensatory mechanism under stressful conditions in order to preserve TL, whereas data from dialysis patients showing both reduced TL and telomerase activity suggest that this process is exhausted with more progressive disease [32]. This assumption is also supported by the fact that telomerase was decreased in male patients with eGFR levels <60 mL/min/1.73 m². Interestingly, whereas it might be logically argued that a decrease in re-

nal function would be primarily responsible for the telomere attrition in FD patients given the fact that chronic kidney disease is considered a model of premature ageing [2, 4], TL was not related to an impairment in renal function, nor to cardiac biomarkers. In future studies, it would be interesting to study the relation between more specific aging biomarkers and specific organ dysfunction in more detail.

The findings of Vujkovic et al. [3] are intriguing and stimulate larger studies to assess the relation between aging biomarkers, markers of metabolic load such as Gb3, as well as detailed phenotypical alterations. Whether the present results have therapeutic implications is yet uncertain. Generalized strategies to delay premature aging include generic factors, such as exercise [33], whereas at the horizon, senolytic drugs, such as quercetin, fisetin, and dasatinab, are emerging as potential treatments for conditions associated with premature ageing [34]. Another promising target appears to be the transcription factor NRF2-KEAP1 and increased oxidative stress [35], which may be involved in both specific organ damage as well as a generalized aging process [36].

At present, the study of Vujkovic et al. [3] presents preliminary evidence that Fabry disease may represent another model of premature aging. Future research could shed more light on the systemic nature of the disease, which possibly extends even beyond the multiorgan involvement directly related to G3b accumulation. This study should be followed by larger and more detailed studies looking into mechanisms and potential targets to prevent premature ageing in Fabry disease beyond the conventional treatment strategies.

Disclosure Statement

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References

- 1 Barnes PJ. Senescence in COPD and its Comorbidities. *Annu Rev Physiol. Annu Rev Physiol.* 2017 Feb 10;79:517–39.
- 2 Kooman JP, Kotanko P, Schols AM, Shiels PG, Stenvinkel P. Chronic kidney disease and premature ageing. *Nat Rev Nephrol.* 2014 Dec;10(12):732–42.
- 3 Cokan Vujkovic A, Novaković S, Vujkovic B, Števanec M, Škerl P, Šabovič M. Aging in fabry disease: role of telomere length, telomerase activity, and kidney disease. *Nephron.* 2019. DOI: 10.1159/000502909.

- 4 Carrero JJ, Stenvinkel P, Fellström B, Qureshi AR, Lamb K, Heimbürger O, et al. Telomere attrition is associated with inflammation, low fetuin-A levels and high mortality in prevalent haemodialysis patients. *J Intern Med*. 2008 Mar;263(3):302–12.
- 5 Waldek S, Patel MR, Banikazemi M, Lemay R, Lee P. Life expectancy and cause of death in males and females with Fabry disease: findings from the Fabry Registry. *Genet Med*. 2009 Nov;11(11):790–6.
- 6 Eline Slagboom P, van den Berg N, Deelen J. Phenome and genome based studies into human ageing and longevity: an overview. *Biochim Biophys Acta Mol Basis Dis*. 2018 Sep;1864(9 Pt A):2742–51.
- 7 Bolsover FE, Murphy E, Cipolotti L, Werring DJ, Lachmann RH. Cognitive dysfunction and depression in Fabry disease: a systematic review. *J Inherit Metab Dis*. 2014 Mar;37(2):177–87.
- 8 Shiels PG, Stenvinkel P, Kooman JP, McGuinness D. Circulating markers of ageing and allostatic load: A slow train coming. *Pract Lab Med*. 2016 Apr;7:49–54.
- 9 von Zglinicki T, Martin-Ruiz CM. Telomeres as biomarkers for ageing and age-related diseases. *Curr Mol Med*. 2005 Mar;5(2):197–203.
- 10 Shiels PG. CDKN2A might be better than telomere length in determining individual health status. *BMJ*. 2012 Mar;344:e1415.
- 11 Mather KA, Jorm AF, Parslow RA, Christensen H. Is telomere length a biomarker of aging? A review. *J Gerontol A Biol Sci Med Sci*. 2011 Feb;66(2):202–13.
- 12 Martin-Ruiz CM, Baird D, Roger L, Boukamp P, Kronic D, Cawthon R, et al. Reproducibility of telomere length assessment: an international collaborative study. *Int J Epidemiol*. 2015 Oct;44(5):1673–83.
- 13 Gingell-Littlejohn M, McGuinness D, McGlynn LM, Kingsmore D, Stevenson KS, Koppeltaetter C, et al. Pre-transplant CDKN2A expression in kidney biopsies predicts renal function and is a future component of donor scoring criteria. *PLoS One*. 2013 Jul;8(7):e68133.
- 14 Horvath S, Raj K. DNA methylation-based biomarkers and the epigenetic clock theory of ageing. *Nat Rev Genet*. 2018 Jun;19(6):371–84.
- 15 Wang M, Lemos B. Ribosomal DNA harbors an evolutionarily conserved clock of biological aging. *Genome Res*. 2019 Mar;29(3):325–33.
- 16 Shiels PG, McGuinness D, Eriksson M, Kooman JP, Stenvinkel P. The role of epigenetics in renal ageing. *Nat Rev Nephrol*. 2017 Aug;13(8):471–82.
- 17 López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell*. 2013 Jun;153(6):1194–217.
- 18 Squillaro T, Antonucci I, Alessio N, Esposito A, Cipollaro M, Melone MA, et al. Impact of lysosomal storage disorders on biology of mesenchymal stem cells: evidences from in vitro silencing of glucocerebrosidase (GBA) and alpha-galactosidase A (GLA) enzymes. *J Cell Physiol*. 2017 Dec;232(12):3454–67.
- 19 Yogasundaram H, Nikhanj A, Putko BN, Boutin M, Jain-Ghai S, Khan A, et al. Elevated Inflammatory Plasma Biomarkers in Patients With Fabry Disease: A Critical Link to Heart Failure With Preserved Ejection Fraction. *J Am Heart Assoc*. 2018 Nov;7(21):e009098.
- 20 Rozenfeld P, Feriozzi S. Contribution of inflammatory pathways to Fabry disease pathogenesis. *Mol Genet Metab*. 2017 Nov;122(3):19–27.
- 21 Braun F, Blomberg L, Brodessa S, Liebau MC, Schermer B, Benzinger T, et al. Enzyme Replacement Therapy Clears Gb3 Deposits from a Podocyte Cell Culture Model of Fabry Disease but Fails to Restore Altered Cellular Signaling. *Cell Physiol Biochem*. 2019;52(5):1139–50.
- 22 Su KH, Dai C. mTORC1 senses stresses: coupling stress to proteostasis. *BioEssays*. 2017 May;39(5):1600268.
- 23 Ravarotto V, Carraro G, Pagnin E, Bertoldi G, Simioni F, Maiolino G, et al. Oxidative stress and the altered reaction to it in Fabry disease: A possible target for cardiovascular-renal remodeling? *PLoS One*. 2018 Sep;13(9):e0204618.
- 24 Tseng WL, Chou SJ, Chiang HC, Wang ML, Chien CS, Chen KH, et al. Imbalanced Production of Reactive Oxygen Species and Mitochondrial Antioxidant SOD2 in Fabry Disease-Specific Human Induced Pluripotent Stem Cell-Differentiated Vascular Endothelial Cells. *Cell Transplant*. 2017 Mar;26(3):513–27.
- 25 Michaud L. Longitudinal study on ocular manifestations in a cohort of patients with Fabry disease. *PLoS One*. 2019 Jun;14(6):e0213329.
- 26 Song HY, Chien CS, Yarmishyn AA, Chou SJ, Yang YP, Wang ML, et al. Generation of GLA-Knockout Human Embryonic Stem Cell Lines to Model Autophagic Dysfunction and Exosome Secretion in Fabry Disease-Associated Hypertrophic Cardiomyopathy. *Cells*. 2019 Apr;8(4):E327.
- 27 Barbosa MC, Grosso RA, Fader CM. Hallmarks of Aging: An Autophagic Perspective. *Front Endocrinol (Lausanne)*. 2019 Jan;9:790.
- 28 Chou SJ, Yu WC, Chang YL, Chen WY, Chang WC, Chien Y, et al. Energy utilization of induced pluripotent stem cell-derived cardiomyocyte in Fabry disease. *Int J Cardiol*. 2017 Apr;232:255–63.
- 29 Pinto LL, Vieira TA, Giugliani R, Schwartz IV. Expression of the disease on female carriers of X-linked lysosomal disorders: a brief review. *Orphanet J Rare Dis*. 2010 May;5(1):14.
- 30 Echevarria L, Benistan K, Toussaint A, Dubourg O, Hagege AA, Eladari D, et al. X-chromosome inactivation in female patients with Fabry disease. *Clin Genet*. 2016 Jan;89(1):44–54.
- 31 Gentilini D, Castaldi D, Mari D, Monti D, Franceschi C, Di Blasio AM, et al. Age-dependent skewing of X chromosome inactivation appears delayed in centenarians' offspring. Is there a role for allelic imbalance in healthy aging and longevity? *Aging Cell*. 2012 Apr;11(2):277–83.
- 32 Tsirpanlis G, Chatzipanagiotou S, Boufidou F, Kordinas V, Alevyzaki F, Zoga M, et al. Telomerase activity is decreased in peripheral blood mononuclear cells of hemodialysis patients. *Am J Nephrol*. 2006;26(1):91–6.
- 33 Rebelo-Marques A, De Sousa Lages A, Andrade R, Ribeiro CF, Mota-Pinto A, Carrilho F, et al. Aging Hallmarks: The Benefits of Physical Exercise. *Front Endocrinol (Lausanne)*. 2018 May;9:258.
- 34 Hobson S, Arefin S, Kublickiene K, Shiels PG, Stenvinkel P. Senescent Cells in Early Vascular Ageing and Bone Disease of Chronic Kidney Disease-A Novel Target for Treatment. *Toxins (Basel)*. 2019 Feb;11(2):E82.
- 35 Ravarotto V, Simioni F, Carraro G, Bertoldi G, Pagnin E, Calò LA. Oxidative Stress and Cardiovascular-Renal Damage in Fabry Disease: Is There Room for a Pathophysiological Involvement? *J Clin Med*. 2018 Nov;7(11):E409.
- 36 Stenvinkel P, Meyer CJ, Block GA, Chertow GM, Shiels PG. Understanding the role of the cytoprotective transcription factor nuclear factor erythroid 2-related factor 2-lessons from evolution, the animal kingdom and rare progeroid syndromes. *Nephrol Dial Transplant*. 2019 Jul;pii:gfs120.