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# The problem of scale in the prediction and management of pathogen spillover

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## Abstract

Epidemics, pandemics, and disease emergence events all underscore the need to predict zoonotic pathogen spillover. Because cross-species transmission is inherently hierarchical, involving processes that occur at varying levels of biological organization, such predictive efforts can be complicated by the many scales and vastness of data potentially required for forecasting. A wide range of approaches are currently used to forecast spillover risk (e.g., macroecology, pathogen discovery, surveillance of human populations, among others), each of which is bound within particular phylogenetic, spatial, and temporal scales of prediction. Here, we contextualize these diverse approaches within their forecasting goals and resulting scales of prediction to illustrate critical areas of conceptual and pragmatic overlap. Specifically, we focus on an ecological perspective to envision a research pipeline that connects these different scales of data and predictions from the aims of discovery to intervention. Pathogen discovery and predictions focused at the phylogenetic scale can first provide coarse and pattern-based guidance for which reservoirs, vectors, pathogens are likely to be involved in spillover, thereby narrowing surveillance targets and where such efforts should be conducted. Next, these predictions can be followed with ecologically driven spatiotemporal studies of reservoirs and vectors to quantify spatiotemporal fluctuations in infection and to mechanistically understand how pathogens circulate and are transmitted to humans. This approach can also help identify general regions and periods for which spillover is most likely. We illustrate this point by highlighting several case studies where long-term, ecologically focused studies (e.g., Lyme disease in the northeast United States, Hendra virus in eastern Australia, *Plasmodium knowlesi* in Southeast Asia) have facilitated predicting spillover in space and time and facilitated design of possible intervention strategies. Such studies can in turn help narrow human surveillance efforts and help refine and improve future large-scale, phylogenetic predictions. We conclude by discussing how greater integration and exchange between data and predictions generated across these varying scales could ultimately help generate more actionable forecasts and interventions.

## 1 Introduction

2 Scale is a key challenge for developing actionable forecasts of pathogen spillover. The processes  
3 that connect reservoir and recipient hosts to facilitate cross-species transmission—which include  
4 infection dynamics in reservoirs, release of infectious agents and survival in the environment,  
5 and recipient host exposure and susceptibility to infection—are inherently hierarchical and occur  
6 over multiple scales of biological organization [1]. Efforts to predict pathogen spillover can thus  
7 be complicated by the many scales and vastness of data potentially required for such forecasts  
8 [Washburne et al., this issue]. However, recent epidemics (e.g., Ebola virus in West Africa [2]),  
9 pandemics (e.g., Zika virus across the Americas [3]), and disease emergence events (e.g.,  
10 *Plasmodium knowlesi* in Malaysia [4]) underscore the need to improve such predictive efforts.

11 Any prediction of pathogen spillover will be bound to a specific scale of space and time  
12 [5], and these scales are determined by objectives of the particular forecast. We rarely aim to  
13 predict an individual that will become infected with a zoonotic pathogen; instead, forecasts often  
14 aim to predict the region and period within which interventions are practical. As an analogue,  
15 weather forecasts do not aim to predict the specific time and place of a particular thunderstorm  
16 but instead predict the probability of rainfall over regional spatial scales (e.g., cities) at hourly,  
17 daily, and weekly temporal scales. Forecasts of pathogen spillover are further restricted not only  
18 to spatial and temporal scales but also to the scale of organisms for which we forecast risk (e.g.,  
19 reservoirs, vectors, pathogens). We refer to this as the “phylogenetic scale”, following recent  
20 developments in ecology and evolution, as it can be defined along a tunable DNA sequence  
21 similarity or given phylogenetic depth [6,7]. As the concept of scale is based upon the order of  
22 entities within some hierarchy (e.g., for spatial scale, continents, biomes, ecoregions, etc [8]),  
23 phylogenetic scale can accordingly encompass several measures, including not only taxonomic  
24 ranks of organisms (e.g., family, genus, species) but also clade age, clade size, and node-to-root  
25 distance, among others [9]. Though we use the term “phylogenetic scale” to be inclusive, its  
26 application here is generally synonymous with taxonomic level. Any spillover prediction will  
27 thus correspond to a particular phylogenetic scale or scales (i.e., a particular lineage of pathogens  
28 from a particular taxon of reservoirs or vectors) within a particular region of space and time.

29 Various approaches are currently used to forecast spillover risk within the bounds of  
30 these scales of prediction. Many macroecological studies focus on processes located upstream in  
31 the spillover pathway and have thus examined trait profiles or cladistic patterns in which species  
32 are likely to serve as reservoirs or vectors of zoonotic pathogens. For example, rodents with a  
33 particularly fast pace of life are more likely to host zoonotic pathogens [10]. Related approaches  
34 have been applied to pathogens [11], and recent work has used viral sequences to predict  
35 reservoirs and vectors [12]. By generating predictions at the phylogenetic scale, these approaches  
36 can guide surveillance, and mapping distributions of known or predicted reservoirs, vectors, and  
37 zoonoses can allow spatial predictions that identify regions where spillover is likely [13,14].  
38 Given phylogenetic and spatial biases in such research [15,16], a related approach has used  
39 pathogen discovery to characterize pathogen diversity; examples include but are not limited to  
40 the U.S. Agency for International Development PREDICT project and Global Virome Project  
41 [17,18]. Many pathogen discovery projects also enhance local capacity and conduct human  
42 surveillance (e.g., VIZIONS in Vietnam [19]), which is another (but not mutually exclusive)  
43 approach [Das et al., this issue]. Spatiotemporal human surveillance can identify spillovers by  
44 screening persons with symptoms that are not easily diagnosable [20] and can facilitate early  
45 detection of known pathogens [21]. These approaches focus on upstream (e.g., reservoirs) and

46 downstream (e.g., humans) processes of the spillover pathway and can generate predictions at  
47 different scales, which has prompted discussions of how best to allocate research efforts [22,23].

48 The problem of scale has been well described for ecology and evolution [8,24] and could  
49 inform discussions of how various research approaches and data streams can contribute to  
50 forecasting pathogen spillover risks [22,25,26]; in particular, scales define the interface between  
51 data collection, forecasts, and interventions. Here, we argue that these various methods, data  
52 streams, and predictions fit along a research pipeline that spans from discovery to intervention  
53 (e.g., Plowright et al. 2019), and we accordingly contextualize these approaches within their  
54 forecasting goals and resulting scales of prediction (Table 1). On the one hand, many approaches  
55 aim to predict spillover of *novel* pathogens (e.g., pathogen discovery in wildlife or in human  
56 populations), and these efforts and the data they collect inform the phylogenetic scales of  
57 prediction (e.g., identifying likely reservoir, vector, and pathogen lineages involved in spillover)  
58 and by extrapolation the spatial scales of prediction (e.g., mapping distributions). On the other  
59 hand, a different suite of efforts aims to predict the spillover of *known* pathogens, which often  
60 focus explicitly on prediction at the scales of space and time within a well-defined phylogenetic  
61 scale (e.g., yellow fever virus [Childs et al., this issue] or Hendra virus [27]). A central concept  
62 connecting these different approaches is scale, but no ecological system has a single scale for  
63 which complex phenomena such as spillover should be studied or predicted [24]. For pathogen  
64 spillover, the scales for which we aim to forecast risk (i.e., phylogenetic, spatial, temporal) vary,  
65 and the resulting predictions made across these variable scales can have different applications.

66 In this paper, we (i) highlight what practical information can be gained from data at  
67 different phylogenetic and spatiotemporal scales and (ii) identify areas of complement between  
68 coarse- and fine-scale forecasts through ecological perspective. Specifically, we envision a  
69 research pipeline that spans from discovery to intervention and connects the data and predictions  
70 provided by macroecology, pathogen discovery, and surveillance in wildlife and humans (Fig. 1).  
71 We also highlight statistical tools for defining the appropriate phylogenetic, spatial, and temporal  
72 scales at which such predictions can be made, which are necessary to better connect data with  
73 proposed interventions. The problem of predicting pathogen spillover is thus best framed as a  
74 challenge in identifying over what scales our data permit forecasts, at what benefits, and at what  
75 costs. Each of these can be quantified for improved surveillance and management decisions.

76

## 77 **Phylogenetic scales of spillover prediction**

78 Do lineages of reservoirs, vectors, and pathogens have common patterns in spillover occurrence?  
79 Phylogenetic predictions for which animals are likely to be reservoirs or vectors and which  
80 pathogens are likely to be zoonotic range from the phylogenetic scale of species to order or even  
81 class. Species-specific predictions, often generated from trait-based analyses and machine  
82 learning algorithms, can offer the most operable resolution of spillover forecasting given a  
83 sufficiently high degree of cross-validation (e.g., area under the receiver operating characteristic  
84 curve [AUC] for out-of-sample prediction) and resulting rank order of likely hosts of zoonotic  
85 pathogens. For example, boosted regression trees with high classification accuracy (AUC=87%)  
86 predicted 112 bat species to be within the 90<sup>th</sup> percentile of likely novel filovirus reservoirs [28].  
87 Mapping the geographic distributions of these likely reservoirs further provided spatially explicit  
88 predictions, with Southeast Asia being a notable hotspot of filovirus-positive bat species where  
89 evidence has otherwise been moderate. Recent filovirus surveys of bats listed in this 90<sup>th</sup>  
90 percentile (e.g., *Eonycteris spelaea*) validate these fine-scale phylogenetic predictions [29].

91 Similar species-specific predictions have been generated for rodent zoonoses [10], Zika virus in  
92 mosquitoes and primates [14,30], and human-to-human transmissibility of zoonotic viruses [31].  
93 Other trait-based approaches generate broader predictions focused on the life history profiles of  
94 likely reservoirs or zoonotic pathogens rather than species predictions [13,32,33].

95 Phylogenetic predictions are also commonly generated at coarser scales, such as clades  
96 (Fig. 1a). Such efforts can be especially insightful when trait data are limited. A comprehensive  
97 study across mammal viruses found that host Order had greater predictive power for explaining  
98 variation in viral diversity compared to traits such as sympatry and body mass [13]. Similarly,  
99 trait-based analyses found no host traits to be predictive of how helminth infections in wildlife  
100 respond to environmental change, yet Old World primates showed high phylogenetic signal [34];  
101 recent work on helminths in more urban vervet monkeys supports such cladistic predictions [35].

102 In general, it may not be safe to assume a single phylogenetic scale (e.g., class, order,  
103 genus, species) captures any pattern in an ecological dataset. As traits driving spillover may  
104 evolve along branches in a phylogeny, there may not be any *a priori* reason to expect the most  
105 important driving patterns to be concentrated along branches of a fixed depth (e.g., only genus).  
106 A novel machine learning algorithm, phylogenetic factorization, was recently developed to more  
107 flexibly identify phylogenetic scales at various depths (e.g., genera and families simultaneously)  
108 underlying patterns in ecological data [36]. A recent application of this method to the taxonomy  
109 of mammal viruses found that the propensity of viruses to be zoonotic is best partitioned along  
110 clades of different phylogenetic scales, such as the order *Nidovirales*, family *Papillomaviridae*,  
111 and genera *Alphavirus* and *Deltaretrovirus* [37]. Such consideration of multiple phylogenetic  
112 scales in future studies of cladistic patterns in reservoirs and vectors would be informative.

113 Generating even coarse predictions of reservoir, vector, or pathogen clades involved in  
114 spillover (Fig. 1a) depends on pathogen discovery and its data on pathogen diversity. However,  
115 such data streams and phylogenetic forecasts provide the first steps that are needed to move from  
116 discovery to intervention by narrowing the range of which reservoirs, vectors, and pathogens  
117 should be the focus of surveillance and where such efforts should be conducted (Fig. 1b). This  
118 initial stage of the pipeline can apply to diverse contexts of spillover (Table 1). Predicting broad  
119 clades of reservoirs or vectors could be especially important for guiding surveillance of novel  
120 pathogens for which little *a priori* data exist [26]; for example, viral genome sequences were  
121 used to forecast artiodactyls as the likely origin of the recently emerged Bas-Congo virus [12].  
122 For known pathogens of public health concern, coarse phylogenetic predictions can still guide  
123 field surveillance; for example, Old World fruit bats, primates, and artiodactyls could be targeted  
124 for further Ebola virus survey efforts [Schmidt et al., this issue]. To move toward interventions,  
125 however, these phylogenetic predictions should be next followed by spatiotemporal studies.

### 126 127 **Spatiotemporal scales of spillover prediction**

128 As pathogen circulation in the reservoir or vector over space and time is the first requisite for  
129 spillover [1], the next stage of the pipeline from discovery to intervention requires phylogenetic  
130 predictions to be supplemented by spatial and temporal surveillance (Fig. 1c). Specifically, such  
131 studies in the proposed reservoir(s) or vector(s) are necessary to quantify spatiotemporal  
132 fluctuations in infection (i.e., pathogen pressure) [Plowright et al., this issue] and assess their  
133 ecological drivers [38–42]. Mechanistic models can help explain whether pathogen pressure is  
134 driven by birth seasonality, metapopulation dynamics, environmental synchrony, and within-host  
135 processes, among others [Glennon et al., this issue][43,44]. Models that integrate such drivers

136 can be highly predictive of spillover. For example, an ecologically driven model of yellow fever  
137 virus spillover that integrated various spatiotemporal data streams had the strongest predictive  
138 accuracy (AUC=0.79) when considering cyclical infection dynamics in wild primate reservoir  
139 hosts; critically, models considering the ecology of mosquito vectors and wild primate reservoirs  
140 were more predictive than those that also included human population size and immunity [Childs  
141 et al., this issue]. As with phylogenetic predictions, such ecologically driven efforts may not  
142 generate exact spatial and temporal spillover predictions. However, these coarser predictions at  
143 the scale of months, seasons, geographic regions, and habitat types can be actionable for guiding  
144 spillover prevention, surveillance, and possible interventions (Fig. 1c).

145 As with phylogenetic predictions, one may not wish to *a priori* assume a given spatial  
146 (e.g., 5 km, 50 km) and temporal scale (e.g., weeks, months) for studying infection dynamics in  
147 reservoirs and vectors. Analyses of spatiotemporal autocorrelation of case data in recipient hosts  
148 or prevalence in related sylvatic systems can facilitate sampling decisions by determining the  
149 spatial and temporal scales at which sampling should occur (e.g., by identifying the range of  
150 spatial and temporal dependence with semivariograms) [45]. Where spatially and temporally  
151 explicit data exist across many reservoir or vector species, phylogenetic factorization could be  
152 applied to identify which spatial and temporal scales may be most appropriate for surveillance or  
153 intervention for specific clades [36]. Determining the spatial and temporal scales at which  
154 infection is correlated can also help identify ecological correlates of infection dynamics and  
155 spillover risk. For example, spatial dependence across large scales can suggest effects of major  
156 climatic drivers, while spatial dependence between nearby locations can instead suggest a highly  
157 localized infection process [46,47]. Additionally, other time series analyses can provide further  
158 epidemiological inference and timescales of spillover risk (e.g., early warning signals) [48].

159 Spatiotemporal surveillance of human populations, especially those in regular contact  
160 with wildlife, is also important for predicting places and times of high spillover risk [22]. For  
161 example, focused surveillance for Ebola virus in humans within West and Central Africa could  
162 alert health systems to early virus detection and avert costs of containing large outbreaks [21].  
163 However, focusing on spatiotemporal data and predictions in the context of reservoirs and  
164 vectors can be particularly informative given the hierarchical nature of spillover, as processes  
165 that occur upstream in the pathway to spillover could have a greater influence on human risk  
166 [Childs et al., this issue; Washburne et al., this issue]. Identifying regions and times of high  
167 pathogen pressure in proposed reservoirs or vectors (e.g., Fig. 1c) could also help prioritize  
168 human surveillance. Lastly, interventions focused upstream on reservoirs or vectors could be  
169 more cost effective and have sustained influence on minimizing risk [Sokolow et al, this issue].

170 Below, we highlight three case studies that demonstrate the value of spatiotemporal data  
171 and understanding ecological mechanism to generate actionable predictions of spillover risks and  
172 to guide long-term interventions (Fig. 2). However, we acknowledge that such efforts can  
173 accordingly carry high logistical costs (e.g., years to establish seasonal pulses of infection from  
174 reservoir populations), and thus ecologically driven studies should be viewed as complementary  
175 to the recipient host surveillance efforts involved in a response to spillover.

176

### 177 *Lyme disease*

178 Lyme disease, which is caused by the bacterium *Borrelia burgdorferi* and transmitted to humans  
179 by *Ixodes* ticks, is the most common vector-borne disease in the United States [49]. Ticks require  
180 three bloodmeals to complete their life cycle. As larvae are born naïve, ticks only obtain *Borrelia*

181 infection after feeding on a competent small vertebrate host. Infected larvae then molt to become  
182 nymphs, which are the most likely life stage to transmit infection to humans. Accordingly, the  
183 density of infected nymphal ticks is a useful proxy for human Lyme disease risk [50,51].

184 Two decades of consistent annual monitoring across replicate sites for the density of  
185 infected nymphs and relevant biotic and abiotic covariates (e.g., acorn abundance, small mammal  
186 abundance, deer abundance, temperature, precipitation) have shown that Lyme disease risk can  
187 be predictable [52–54]. A given year’s density of infected nymphs is predicted by the prior  
188 year’s abundance of white-footed mice and eastern chipmunks and by acorn abundance from two  
189 years prior [52]. Years with strong acorn mast generate high rodent abundances in the next year,  
190 which subsequently drive high nymphal abundances the following year. Such analyses drawn  
191 from years of repeated, ecologically driven surveillance thus show that masting indices can  
192 provide relevant spatial and temporal (e.g., two-year lag) predictions for spillover risk (Fig. 2a).

193 In addition to identifying regions and years for which human risk of exposure to infected  
194 nymphs is high, this mechanistic understanding highlights opportunities to interrupt *Borrelia*  
195 transmission by targeting small mammal reservoirs and their contribution to hosting and  
196 infecting larval ticks. Field experiments have found that direct immunization of white-footed  
197 mice can reduce nymphal infection prevalence [55], though effects were strongest when oral bait  
198 vaccines were used to immunize the broader small mammal host community [56]. Such  
199 ecological approaches were more effective across longer study durations, highlighting how such  
200 interventions may be more sustainable [Sokolow et al., this issue]. Ongoing efforts through The  
201 Tick Project (<https://www.tickproject.org/>) are further assessing what combination of acaricide  
202 treatments, deployed at which spatial and temporal scales, are most capable of reducing risk [57].

203

#### 204 *Hendra virus*

205 Hendra virus is a RNA virus that emerged in 1994, causing an outbreak of a lethal respiratory  
206 and neurological disease in horses and subsequently to humans that has been followed by over  
207 60 spillover events through 2018 across eastern Australia [58,59]. Flying foxes of the genus  
208 *Pteropus* are the natural reservoir hosts, and transmission of virus to horses is assumed to occur  
209 through the ingestion of food or water contaminated with urine [39,60]. Surveillance shows  
210 pulses of bat viral shedding in winter [61], but these do not follow uniform seasonality [47].

211 Despite the relatively small number of spillover events compared to Lyme disease cases,  
212 a mechanistic understanding of Hendra virus spillover informed by long-term ecological work  
213 has likewise provided coarse predictions over space and time [58]. The unprecedented cluster of  
214 spillovers in 2011 was preceded by distinct environmental conditions: a rise in the southern  
215 oscillation index in 2010 that shifted eucalypt plants, the preferred nectar source of bats, into a  
216 growth rather than a flowering phase, in turn restricting food availability [62]. Such food  
217 shortages have been exacerbated by agricultural land conversion and cause periods of intense  
218 nutritional stress for bats [63,64]. Nutritional stress not only drives flying foxes into urban  
219 habitats, where they form sedentary camps near abundant but poor-quality food resources, but  
220 also likely amplifies Hendra virus shedding by impairing bat immunity; for example, nutritional  
221 stress was associated with greater seroprevalence in little red flying foxes [65,66] (Fig. 2b).

222 Recently, environmental conditions and weather events in eastern Australia mirrored  
223 those seen prior to the 2011 Hendra virus spillovers; a severe El Niño in late 2015 and early 2016  
224 was followed by a rise in the southern oscillation index in winter 2016 and a subsequent food  
225 shortage in summer 2016 with concomitant nutritional stress observed in flying foxes [58]. A

226 spillover event in horses was coincident in space and time with nutritional stress in flying foxes  
227 [67]. This chain of predictable events enabled researchers to suggest veterinarians increase  
228 winter monitoring of horses and urge precautionary actions such as vaccination [58]. As with  
229 Lyme disease, a mechanistic ecological understanding facilitated generating such predictions.

230

### 231 *Plasmodium knowlesi*

232 A species of zoonotic malaria, *Plasmodium knowlesi* primarily circulates in monkeys across  
233 Southeast Asia. While human infections were first recorded in the 1960s, only since 2004 have  
234 larger epidemiological clusters been recorded [4]; in some regions (e.g., Sabah, Malaysia), *P.*  
235 *knowlesi* accounts for the majority of all malaria cases [68]. Wild macaques and leaf monkeys  
236 are the primary reservoirs, and transmission occurs through an infectious bite of *Anopheles*  
237 *leucosphyrus* mosquitoes [69]. Most if not all human cases are spillover from primates [70].

238 Unlike Lyme disease and Hendra virus, most research on *P. knowlesi* has focused on  
239 humans, including a large-scale case–control study, MONKEYBAR, in the Philippines and  
240 Malaysian Borneo [71]. Human risk factors for infection include recent activities within or at the  
241 edge of forests (e.g., vegetation clearance, agriculture) where wild macaques occur [72,73], and  
242 human cases have been positively associated with high degrees of local forest cover (2 km) and  
243 recent forest loss [74]. In regions where *P. knowlesi* cases are less common, human risk has been  
244 associated with land clearing activities within 500 meters of an exposed person’s home [75].

245 These findings suggest land conversion may alter interactions between humans,  
246 reservoirs, and vectors differently at distinct spatial scales. Recent work thus used machine  
247 learning tools to flexibly consider the effects of multiple spatial scales on malaria cases [76].  
248 Risk was high when the proportion of cleared land within 1 km was low, suggesting that isolated  
249 households could be more prone to high vector densities in forested habitats. The effect of  
250 deforestation was also high at larger spatial scales (4–5 km), suggesting elevated human  
251 exposure to mosquitoes during commutes to agricultural work or a change in macaque behavior  
252 or demography [76]. Such large-scale deforestation could reduce reservoir host densities and  
253 promote a subsequent behavior change in mosquitoes in these cleared habitats [77]. While  
254 identifying the spatial scales of human risk cannot identify the processes by which deforestation  
255 drives *P. knowlesi* spillover, such work can narrow the space of possible ecological mechanisms  
256 to be elucidated by detailed spatiotemporal studies of reservoirs and vectors (Fig. 2c) [78].

257

### 258 **Interplay between phylogenetic and spatiotemporal scales of prediction**

259 The above examples highlight that ecologically minded spatiotemporal studies of reservoirs and  
260 vectors can provide actionable predictions that can be used to mitigate spillover risks. These  
261 predictions are often coarse in scale and cannot identify the precise spatial location and narrow  
262 timepoint of observed spillover events. However, predictions need not be generated on extremely  
263 fine scales to be useful for prevention or intervention. For Lyme disease, human risk is greatest  
264 in regions with high oak abundance and where acorn masting occurred two years prior. People  
265 can become more vigilant in spring and summer when nymphs are questing, and ecological  
266 interventions can work to break the transmission cycle between rodents and ticks [52,54,57]. For  
267 Hendra virus, veterinarians in regions with active flying fox camps can expect to more closely  
268 monitor horse health during the seasons that follow notable shifts in the southern oscillation  
269 index or observed nutritional stress events in flying foxes [47,58,62]. Although the ecological  
270 mechanisms connecting deforestation to *P. knowlesi* spillover remain less resolved than for



271 Lyme disease or Hendra virus, observed associations between human cases and different spatial  
272 scales of land conversion can guide future ecological studies as well as spatially explicit  
273 interventions [74,76,78]. Such work collectively highlights that spillover can be predicted, but at  
274 varying phylogenetic, spatial, and temporal scales, which carry distinct benefits and costs.

275 We here argue that generating phylogenetic predictions, conducting ecologically driven  
276 spatiotemporal studies, and surveying human populations are complementary approaches to  
277 investigating and predicting spillover. Ecological studies at fine spatial and temporal scales can  
278 complement large-scale pathogen discovery projects, macroecology of zoonoses in reservoirs  
279 and vectors, and human surveillance (Fig. 1). Pathogen discovery can generate the data needed to  
280 narrow the wide range of which reservoirs, vectors, and pathogens should be targets of  
281 surveillance. Detailed ecological field studies can next generate a mechanistic understanding of  
282 pathogen circulation and spillover that coarsely predict when and where cross-species  
283 transmission is most likely. This work can also identify ecological interventions that could occur  
284 prior to human exposure and that may have pronounced and long-term impacts on limiting risks.  
285 Further, spatiotemporal studies of reservoirs and vectors can also narrow human surveillance  
286 efforts toward pathogens with the greatest likelihood for being zoonotic and around those regions  
287 and periods where spillover is most likely. Indeed, projects such as PREDICT currently adhere to  
288 a similar model by integrating wildlife and human data streams to guide surveillance efforts [25].  
289 Moreover, these various data streams on spatial and temporal infection in reservoirs, vectors, and  
290 recipient hosts could help refine and improve macroecological analyses and predictions (Fig. 1d).  
291 This pipeline connecting macroecology, pathogen discovery, and surveillance could facilitate  
292 synergistic hypothesis generation (i.e., prediction) and testing (i.e., surveillance) to continually  
293 refine research efforts. This integrated approach also highlights how spillover predictions can  
294 catalyze both discovery and intervention. Greater exchange between macroecology, pathogen  
295 discovery, and surveillance (e.g., through interdisciplinary working groups [Becker et al., this  
296 issue; [79,80]) could ultimately help generate more actionable predictions and public health  
297 interventions to limit pathogen spillover risks.

298

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316

317 **Author contributions**

318 All authors contributed to the development of ideas and to the writing of this manuscript.

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320

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535 **Tables**

536

537 Table 1. Summary of select approaches to predict pathogen spillover, stratified by the goal and  
 538 the scales of predictions.

Prediction goal	Approach	Scale	Example method	Source
<b>Emergence of novel zoonotic pathogens</b>	Reservoir or vector surveillance	Phylogenetic (reservoir, vector, pathogen), space	Quantify pathogen diversity across reservoirs or vectors globally	[17]
	Macroecology	Phylogenetic (reservoir, vector, pathogen), space <sup>1</sup>	Machine learning to identify likely or novel reservoirs or vectors	[10,12]
	Human surveillance	Phylogenetic (pathogen), space	Survey human populations with high wildlife contact	[22]
<b>Where and when known pathogens may spillover</b>	Macroecology	Phylogenetic (host), space <sup>1</sup>	Machine learning to identify which reservoirs to sample	[14]
	Human surveillance	Space, time	Improve diagnostic capacity to detect early zoonotic outbreaks	[21]
	Early warning signals	Time	Detect transition from stuttering chains to sustained transmission	[48]
	Reservoir surveillance	Space, time	Identify ecological predictors of pathogen spillover (Fig. 2)	[52,58]

539 <sup>1</sup>Space is here implicit through mapping the distribution of predicted reservoirs or vectors

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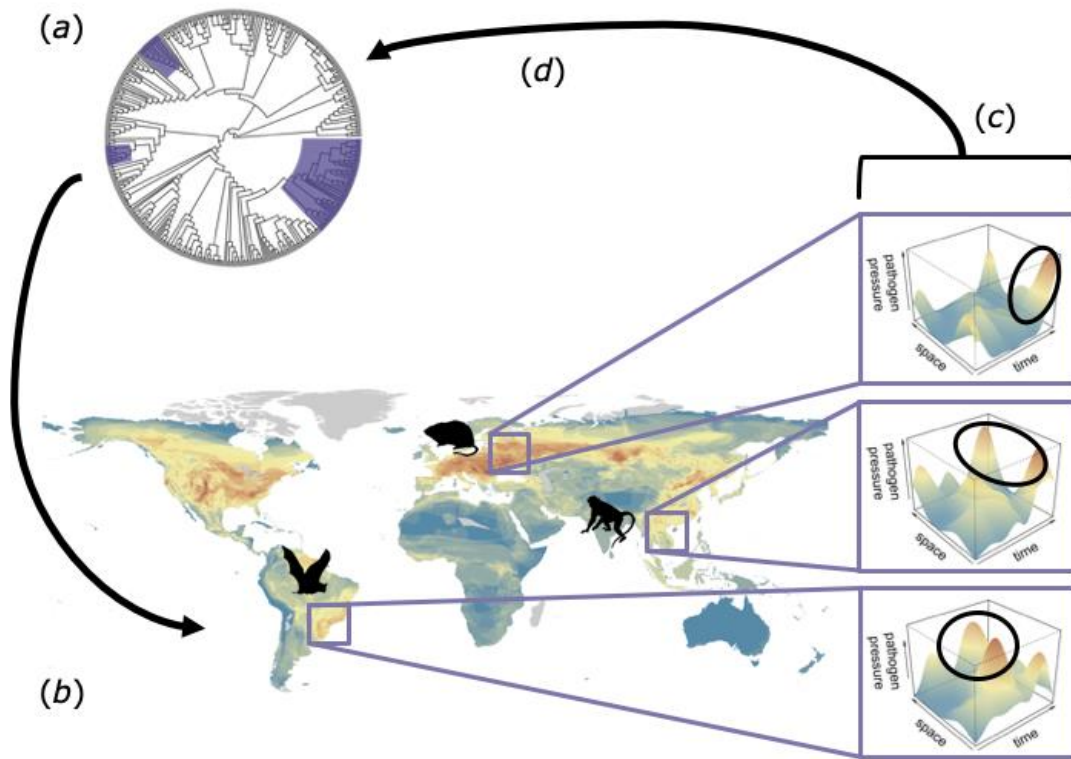
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543 **Figures**

544

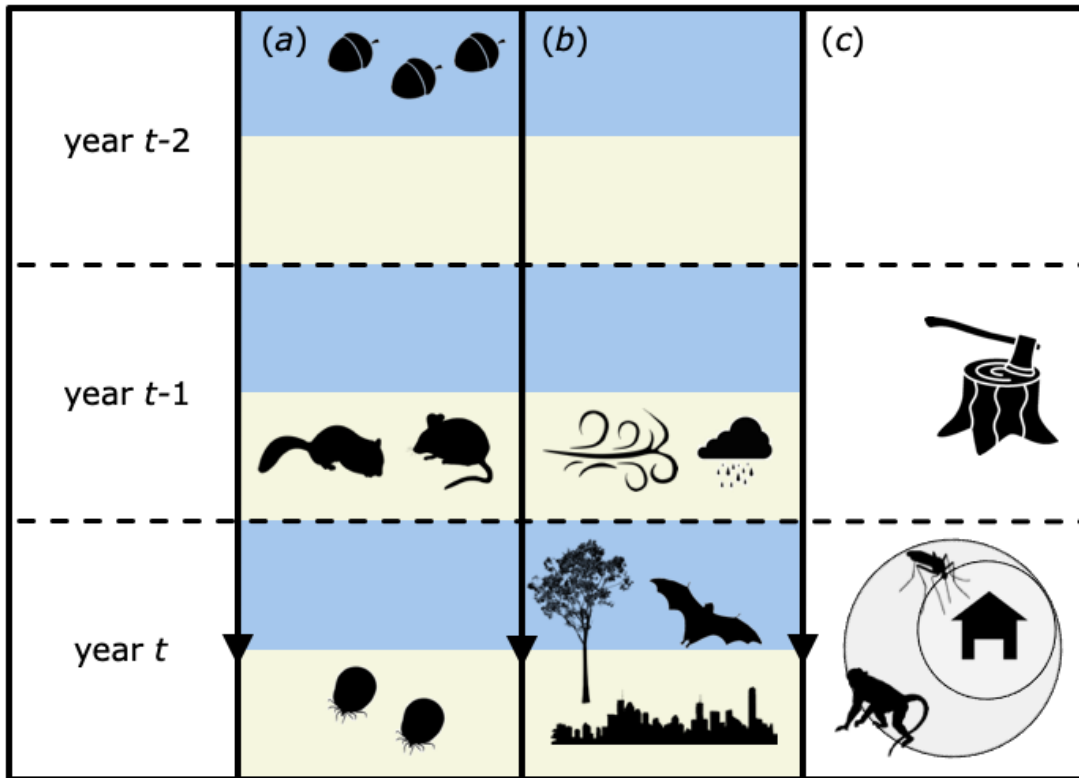
545 Figure 1. Interplay between scales of pathogen spillover prediction and a proposed pipeline for  
546 their integration. Even coarse phylogenetic predictions (a) can narrow the scope of what  
547 reservoirs and vectors for a given set of pathogens, and in which geographic regions, should be  
548 prioritized for surveillance (b); examples based on trait-based or cladistic analyses include  
549 filoviruses in Neotropical bats [28], zoonotic pathogens in European rodents [10], and helminths  
550 in Old world primates [34]. Spatiotemporal studies can next elucidate how zoonotic pathogens  
551 circulate in reservoir or vector populations, identify broad spatial and temporal scales at which  
552 pathogen pressure (e.g., shedding) is greatest, and uncover the ecological mechanisms leading to  
553 spillover (c). Based on these regions and times where risk is greatest (circles), managers can  
554 design preemptive interventions and prioritize human surveillance. Data from spatiotemporal  
555 studies can further address information gaps and refine future macroecological analyses and  
556 predictions in an iterative fashion (d). The map (b) is adapted from Han et al. [10], and  
557 perspective plots (c) were generated using random realizations of a binomial point process with  
558 varying intensities [81]; both are used here simply as heuristic devices.  
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562 Figure 2. Predictive insights into pathogen spillover risk gained from long-term, spatiotemporal  
 563 studies of reservoir hosts and vectors. For Lyme disease in North America (a) and Hendra virus  
 564 in eastern Australia (b), ecological and mechanistic approaches have identified both spatial and  
 565 temporal proxies for spillover risk in recipient hosts. Columns indicate ecological correlates of  
 566 spillover risk at varying time lags, and colors represent differentiate those that occur in autumn  
 567 and winter (blue) or in spring and summer (yellow) for systems with strong seasonality (a–b).  
 568 The ecological mechanisms linking land clearance with *P. knowlesi* spillover in Southeast Asia  
 569 are less well understood (c), but analyses of spatial scale and human cases have generated  
 570 hypotheses for spatiotemporal studies of reservoirs and vectors.  
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