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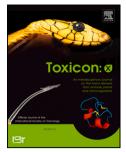
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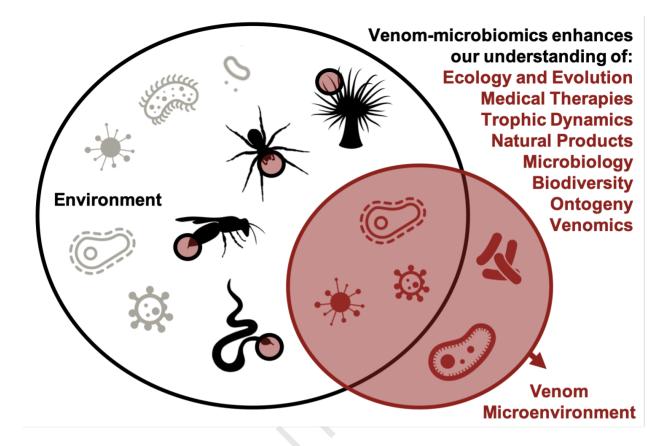
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The emerging field of venom-microbiomics for exploring venom as a microenvironment, and the corresponding Initiative for Venom Associated Microbes and Parasites (iVAMP)

- 5
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38 Abstract

- 39 Venom is a known source of novel antimicrobial natural products. The substantial, increasing
- 40 number of these discoveries have unintentionally culminated in the misconception that venom
- 41 and venom-producing glands are largely sterile environments. Culture-dependent and -
- 42 independent studies on the microbial communities in venom microenvironments reveal the
- 43 presence of archaea, algae, bacteria, fungi, protozoa, and viruses. Venom-centric microbiome
- studies are relatively sparse to date with the adaptive advantages that venom-associated microbes
- 45 might offer to their hosts, or that hosts might provide to venom-associated microbes, remaining
- 46 largely unknown. We highlight the potential for the discovery of venom microbiomes within the
- adaptive landscape of venom systems. The considerable number of convergently evolved
 venomous animals, juxtaposed with the comparatively few known studies to identify microbial
- 49 communities in venom, provides new possibilities for both biodiversity and therapeutic
- 50 discoveries. We present an evidence-based argument for integrating microbiology as part of
- 51 venomics (i.e., venom-microbiomics) and introduce iVAMP, the Initiative for Venom Associated
- 52 Microbes and Parasites (<u>https://ivamp-consortium.github.io/</u>), as a growing collaborative
- 53 consortium. We express commitment to the diversity, inclusion and scientific collaboration
- 54 among researchers interested in this emerging subdiscipline through expansion of the iVAMP
- 55 consortium.
- 56

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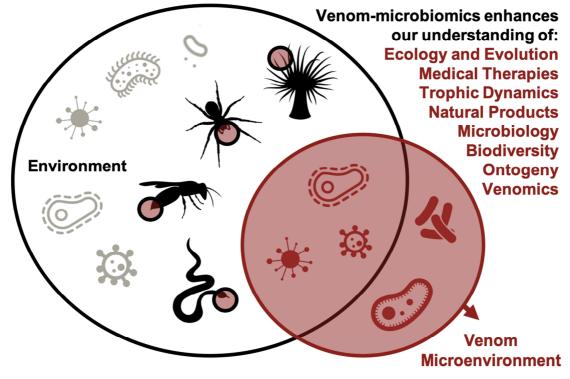
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61 62

57 Keywords: bacteria, coevolution, holobiont, microbiome, symbiont, virus

59 Highlights:

- Venom-microbiome studies as an integrative field of venomics and microbiology
- Argument for multi-omics-based discovery through a microenvironment framework
- Introduction of a venom-microbiome research consortium (iVAMP)



64

65 Text

While scientific research in toxinology and microbiology has persisted for centuries, a cursory 66 search of the literature reveals less than 150 studies overlap between these two fields despite 67 each significantly advancing as a result of next generation sequencing technology (Figure 1, 68 69 Supplemental Table 1, Supplemental Code). The integration of genomics (Moran and Gurevitz, 2006), transcriptomics (Pahari et al., 2007), and proteomics (Fry, 2005) into the study of venom 70 71 has contributed to new toxin discovery and associated biological activity (Oldrati et al., 2016; Calvete, 2017). Over the past 15 years, microbiome research has yielded breakthroughs in our 72 73 knowledge of unculturable microbial "dark matter" (Bernard et al., 2018), the origins of life (Spang et al., 2017), and human health (Arnold et al., 2016; Clavel et al., 2016). Providing 74 ecological and evolutionary context has enhanced both microbiology (Boughner and Singh, 75 2016; Hird, 2017) and venomics (Prashanth et al., 2016; Sunagar et al., 2016; Calvete, 2017). We 76 77 thus propose viewing venom as a microenvironment that occupies a unique niche in which 78 microbes may adapt as a critical perspective for investigating the dynamics of venom-microbe

79 interactions.

80 Researchers in the fields of both venomics and microbiology share common interests in natural products (Katz and Baltz, 2016; Robinson et al., 2017) and adaptive evolution (Phuong et 81 al., 2016; Hird, 2017). With more information on the presence and diversity of venom-associated 82 83 microbiomes (Table 1), future research efforts can focus on how microbes colonize and thrive in venom glands as a starting point for integrating these fields (McFall-Ngai, 2014; Nunes-Alves, 84 2015). For example, examining the biology of the host using microscopy (Schlafer and Meyer, 85 2017) and biomechanics (Yevick and Martin, 2018) could result in translated predictive models 86 (Biggs et al., 2015) for identifying the underlying mechanisms of toxin and metabolite function 87 88 (Sapp, 2016; Adnani et al., 2017). Determining if and which venom microenvironments are truly sterile, and if microbes contribute to shaping the genetic architecture of the venom gland, will 89 90 prove critical in our understanding of venom evolution (Conlin et al., 2014) and antimicrobial 91 resistance (Adnani et al., 2017). Correlating microbial community profiles with functional characteristics of venom could provide yet another layer to the venomics field that would deepen 92 our insight on the mechanisms driving venom variation. Identifying microbial species that have 93 adapted to these seemingly extreme environments (Rampelotto, 2013) will open new avenues of 94 95 research, and emphasizes the need for phylogenetically representative venom host model systems 96 to be bred axenically in vivo to allow researchers to test the functional roles of venom-associated 97 microbes observed in the wild (Figure 2).

98 The host-microbe interactions that naturally occur in the venom microenvironment
99 remain largely unknown, and addressing this knowledge gap through directed microbiome
100 sequencing experiments within a wildtype ecosystem framework will strengthen our
101 understanding of animal associated microbes (McFall-Ngai et al., 2013). A variety of microbial
102 studies have found tetrodotoxin-producing bacteria in venomous and poisonous animals (Hwang

et al., 1989; Cheng et al., 1995; Pratheepa and Vasconcelos, 2013; Stokes et al., 2014) as well as

a number of viruses with RNA genomes residing in venom (Debat, 2017). These studies contrast
 with the notion of the venom microenvironment as largely sterile in that the primary research on

venom-gland derived toxin compounds focuses on antimicrobial properties (Figure 1). However,

107 (1) compounds derived from or contained within venom that demonstrate antimicrobial activity

108 against clinical and/or reference strains (Almeida et al., 2018) may not reflect what occurs

against wild-type strains that co-evolved within venom glands (Reis et al., 2018), and (2)

110 cultured microbes can produce compounds in a lab setting that they may not produce in nature (McCoy and Clapper, 1979; Simmons et al., 2008; Peraud et al., 2009; Catalán et al., 111 112 2010;Quezada et al., 2017b, 2017a, 2017b; Silvestre et al., 2005; Yu et al., 2011). The captive environment, which is already known to affect the host venom profile (Willemse et al., 1979; 113 Freitas-de-Sousa et al., 2015), may also influence microbial composition of the oral and venom 114 115 microbiomes (Hyde et al., 2016), which has led to a call for microbiome studies to utilize wildcollected samples (Colston & Jackson, 2016; Hird, 2017). Studying the venom microbiome, and 116 117 considering the adaptive traits of microbes under selection in an ecological context as it occurs in the wild, clarifies the evolutionary pressures for these antimicrobial compounds found in venom 118 119 (Figure 2). In vitro, in vivo, and natural venom microbiome experiments alongside culture-120 dependent and -independent techniques contribute to our understanding of mutual symbioses, with room for predictive modeling to identify novel niches for microbial adaptation and 121 122 competition (Bull et al., 2010; Zhu et al., 2018). An initial search shows approximately 100 papers per year have consistently been 123 124 published on venom antimicrobial peptides (PubMed search term - antimicrobial AND peptide AND venom 14th Mar 2019) for the past 5 years. The few venom-microbiome studies in the 125 literature to date (Table 1) indicate a clear need for an expansion of the subdiscipline of venom-126 microbiome research, and this has led to the formation of an international, collaborative cohort of 127 128 researchers referred to as the Initiative for Venom Associated Microbes and Parasites (or 129 iVAMP, https://ivamp-consortium.github.io/). A major goal of the iVAMP consortium is to provide a platform for the scientific community to openly discuss areas of interest to the field. 130 Figure 2 outlines some examples of ongoing questions that may be of interest to iVAMP 131 researchers. By emphasizing representation through practice, this consortium supports working 132 with and for communities from which we sample rather than taking from them. Involving 133

scientists across the globe through initiatives like iVAMP extends beyond the requirements of
legislation, such as the Nagoya Protocol (Buck and Hamilton, 2011), to ensure that science is
accessible to the public and inclusive of all parties involved. Overall, the approach taken by this
initiative expands suggested practices (Weber et al., 2001; Cheng et al., 2018) for the benefit of
scientific innovation and discovery.

As an organization, iVAMP has explicit goals and approaches for furthering the fields of 139 microbiome research and venomics (Figure. 2) as well as specific aims for conducting ethical, 140 inclusive, reproducible science. In doing so, our practices seek to prevent counterproductive 141 competition and instead embrace interdisciplinary, collaborative scientific research. The broad 142 scientific disciplines covered by iVAMP members provide a network that allows researchers 143 access to a variety of technical platforms and key resources that otherwise may not be available 144 in individual labs. This is especially important for those researchers who may want to enter the 145 venomics field, but lack accessibility to the necessary resources or instrumentation. Expansion of 146 147 knowledge on microbes living in the many diverse venom host microenvironments additionally contributes to currently absent aspects of holobiont and coevolutionary theory (Faure Denis et 148 al., 2018). Through iVAMP, researchers set an open-access tone for the subdiscipline of venom-149 150 microbiomics that will be useful well into the future.

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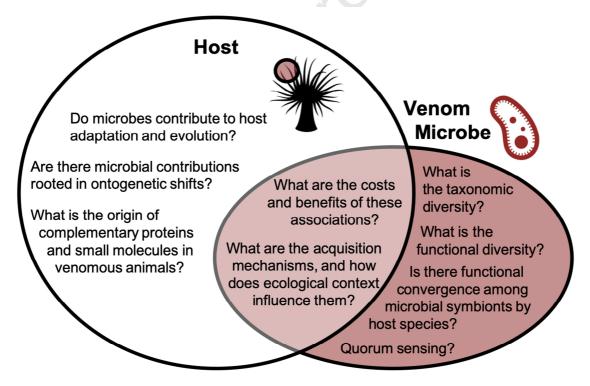


154 Figure 1. Word clouds representative of Supplemental Table 1 content.

155 A breakdown of 140 resultant articles from searching Web of Science for venom-microbe

156 studies. (a) Most articles are either bacteria- or virus- specific, and a subset (16 articles) are not

- related to studies involving microbes. After removing these articles, investigation of the
- remaining 126 show (b) approximately 71% focus on venom toxins exhibiting antimicrobial
- 159 properties with only about 11% focused on venom-microbe interactions. (c) Roughly 57% of the
- 160 surveyed studies focus on snake venom, and the remaining studies are largely from arthropods.
- 161



- 162
- 163 Figure 2. Proposed questions for venom-microbiome exploration of the ecology and
- 164 evolution of venomous hosts and their microbial associates.
- 165 A Venn diagram displaying the intersections of microbiology and venomics through an ecology
- and evolution focus. The questions presented are examples of possible areas of investigation to
- advance the field.

PUBLISHED STUDIES	ORGANISM	TISSUE	WILD / Captive	APPROACH
WEBB B.A., SUMMERS M.D. 1990	Wasp	Venom gland	Captive	Culture, Sanger Sequencing
PERAUD ET AL. 2009	Cone-snail (3 species)	Body, Hepatopancreas, Venom Duct	Wild	Culture, FISH, Sanger Sequencing
GOLDSTEIN ET AL. 2013	Monitor Lizard	Saliva, Gingiva	Captive	Culture, Sanger Sequencing, 16S
SIMMONDS ET AL. 2016	Parasitoid Wasp	Venom Gland	Wild	RNAseq/reverse transcriptomics
D EBAT 2017	Spiders	Transcriptomes of the Body, Brain, Silk Gland Venom Gland	Wild	Data-mining (NGS)
TORRES ET AL. 2017	Cone-snail (8 species)	Venom Duct, Muscle, External Duct	Wild	16S, 454
ESMAEILISHIRAZIFARD ET AL. 2018	Snakes (5 species) Spiders (2 species)	Venom, Oral Cavity	Wild, Captive	Culture, 16S, WGS
IVAMP PROJECTS IN			WILD /	
PROGRESS	ORGANISM	TISSUE	CAPTIVE	APPROACH
Colston	Snakes (multiple)	Venom, Venom Glands, Venom Ducts, Oral Cavity, Muscle, Stomach and GIT	Wild, Captive	16S, RNAseq transcriptomics, Proteomics
HARMS + MACRANDER	Lionfish: Pterois volitans	venom glands, venom	Wild (Invasive)	Transcriptomics, Proteomics
KEISER + COLSTON	Spiders: Stegodyphus	venom glands, venom	Wild, Captive	16S, RNAseq transcriptomics, Proteomics
STIERS, COLSTON	Snake: Crotalus scutulatus	Venom, Venom Glands, Venom Ducts, Oral Cavity, Muscle, Stomach and GIT	Wild, Captive	16S, RNAseq transcriptomics, Proteomics
UL-HASAN, NOBILE, Petras	Cone-snail: Californiconus	Venom, Venom Duct,	Wild, Captive	16S and 18S, Proteomics,

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californicus

Hepatopancreas, Shell, Egg Metabolomics

Table 1. Explicit Sequencing and Next-Generation venom microbiome studies, including published & in progress work within iVAMP ("+" **denotes a collaboration formed because of access to the iVAMP network**). Next-generation venom microbiome studies are comparatively recent, and few in number. Even so, the diversity of these host and microbial community studies highlight the potential benefits of integrating microbiology and venomics (Webb and Summers, 1990; Peraud et al., 2009; Goldstein et al., 2013; Debat, 2017; Torres et al., 2017; Esmaeilishirazifard et al., 2018).

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Conflicts of interest

The authors declare no conflicts of interest.

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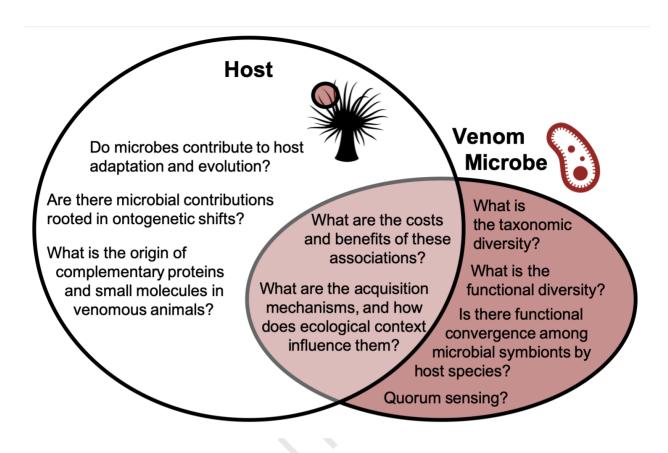
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Declaration of interests

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The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

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