

# n-3 fatty acids as preventive and therapeutic agents in attenuating PCOS complications

Salek, M., Clark, C., Taghizadeh, M. & Jafarnejad, S.

Published PDF deposited in Coventry University's Repository

**Original citation:**

Salek, M, Clark, C, Taghizadeh, M & Jafarnejad, S 2019, 'n-3 fatty acids as preventive and therapeutic agents in attenuating PCOS complications' EXCLI journal, vol. 18, pp. (In-press).

<https://dx.doi.org/10.17179/excli2019-1534>

DOI 10.17179/excli2019-1534

ISSN 1611-2156

ESSN 1611-2156

Publisher: IfADo

**This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>).**

**Copyright © and Moral Rights are retained by the author(s) and/ or other copyright owners. A copy can be downloaded for personal non-commercial research or study, without prior permission or charge. This item cannot be reproduced or quoted extensively from without first obtaining permission in writing from the copyright holder(s). The content must not be changed in any way or sold commercially in any format or medium without the formal permission of the copyright holders.**

## Review article:

# ***N*-3 FATTY ACIDS AS PREVENTIVE AND THERAPEUTIC AGENTS IN ATTENUATING PCOS COMPLICATIONS**

Mina Salek<sup>1</sup>, Cain C. T. Clark<sup>2</sup>, Mohsen Taghizadeh<sup>1</sup>, Sadegh Jafarnejad\*<sup>1</sup>

<sup>1</sup> Research Center for Biochemistry and Nutrition in Metabolic Diseases, Kashan University of Medical Sciences, Kashan, I.R. Iran

<sup>2</sup> Centre for Sport, Exercise and Life Sciences, Coventry University, Coventry, United Kingdom

\* Corresponding author: Research Center for Biochemistry and Nutrition in Metabolic Diseases, Kashan University of Medical Sciences, Kashan, I.R. Iran.

Tel: +98-31-55463378; Fax: +98-31-55463377. E-mail: [sjafarnejad@alumnus.tums.ac.ir](mailto:sjafarnejad@alumnus.tums.ac.ir)

<http://dx.doi.org/10.17179/excli2019-1534>

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>).

## ABSTRACT

To our knowledge, in spite of several trials exploring the beneficial effect of *n*-3 polyunsaturated fatty acids (PUFA) on polycystic ovary syndrome (PCOS), no comprehensive evidence has investigated the effects of *n*-3 PUFA consumption on PCOS complications. Therefore, our aim was to conduct a review to investigate the possible effect and related mechanisms. A comprehensive systematic search was conducted in Embase, MEDLINE/PubMed, Google Scholar, and SCOPUS, to identify studies investigating *n*-3 fatty acids as a preventative or therapeutic agent for the attenuation of PCOS complications. Subsequently, the impact of omega-3 on PCOS, omega-3 and inflammation, omega-3 and insulin resistance, omega-3 and adipokines, omega-3 and lipid metabolism, omega-3 and endothelial function and omega-3 and hormonal factors were discussed. There are multiple mechanisms by which *n*-3 PUFAs may exert their beneficial effects on PCOS, including anti-obesity, glycemic and hormonal hemostasis, anti-inflammatory, regulation of adipokine production and enhancement of endothelial function. *N*-3 PUFAs are a promising agent in relieving complications associated with PCOS. Although most of the studies in patients with PCOS reported an improvement in most complications after administration of omega-3 supplements, there is a distinct dearth of studies investigating the dietary intake of these types of fatty acids. Moreover, favorable effects regarding the improvement of dyslipidemia, regulation of adipokines, regulation of hormonal factors and enhancement of endothelial function are limited. Therefore, more trials are warranted to investigate palatable mechanisms for clarifying the metabolic and hormonal effects of these agents in PCOS.

**Keywords:** *n*-3 PUFA, PCOS, infertility, metabolic disorder, obesity

## INTRODUCTION

Polycystic ovary syndrome (PCOS) is considered one of the most common female endocrine disorders with an approximate prevalence of 5–10 % (Rondanelli et al., 2014; Dunaif, 1997; Asuncion et al., 2000),

with sufferers at an increased risk of reproductive-related disorders (Rondanelli et al., 2014; Goodarzi et al., 2011). There are several characteristics manifest in this syndrome including polycystic ovaries, irregular menstruation, hyperandrogenism and obesity and its associated consequences, such as insulin

resistance, which is reported in about 5 to 20 percent of women, depending upon the applied diagnostic criteria (Faghfoori et al., 2017; Pourghassem Gargari et al., 2015). According to the Rotterdam ESHRE/ASRM sponsored PCOS consensus workshop, a positive diagnosis is considered when any two of the following three features are observed; oligo-ovulation/anovulation, polycystic ovaries on ultrasound examination and/or clinical and/or biochemical signs of hyperandrogenism (Rondanelli et al., 2014; Rotterdam ESHRE/ASRM-sponsored PCOS consensus workshop group, 2004). Additionally, there are several endocrine abnormalities in PCOS which result in metabolic alterations, such as; insulin resistance (IR), hyperinsulinemia, central obesity and metabolic syndrome (MeTS) (Rondanelli et al., 2014), and their associated consequences, including increased risk of hypertension, developing type 2 diabetes (T2DM), dyslipidemia, cardiovascular disorders and atherosclerosis (Rondanelli et al., 2014; Solomon, 1999). The reasons for the manifestation of PCOS is not well established; despite this, a combination of parameters, including genetic and/or epigenetic, environmental factors and exposure to high levels of androgen prenatally are considered to, putatively, play a role in the onset of the disorder. Previous studies have revealed environmental factors may represent fundamental factors in the incidence and treatment of the disorder. Among these environmental factors, dietary patterns are considered as one of the most important and controllable factors (Faghfoori et al., 2017; Nardo et al., 2008; Dumesic et al., 2007; Xita and Tsatsoulis, 2006).

Although optimal treatment for PCOS is not yet fully determined; multifactorial approaches are advisable, including a combination of one or more of the most effective interventional strategies, such as diet and lifestyle changes, or the use of pharmaceuticals (hormonal agents: oral contraceptives/cyclic progestins, gonadotropins, antiestrogens, and metabolic agents such as insulin sensitizers).

It has been shown in a recent survey of 138 endocrinologists and 172 gynecologists, performed by Cussons et al. (2005) and Liepa et al. (2008), that the majority of experts believe diet and exercise is the first step in the treatment of PCOS (Liepa et al., 2008). For instance, dietary fat, such as essential polyunsaturated fatty acids (PUFA) including omega-3 and omega-6 fatty acid, has been studied in relation to PCOS (Liepa et al., 2008). Dietary sources of *n*-3 PUFA are limited in comparison with *n*-6 PUFA sources. Alpha-linolenic acid (ALA) is synthesized by plants and is abundant in sources such as flaxseed and nuts or leaf (Geetha and Chakravarthula, 2018), whilst fish are considered among the richest sources of *n*-3 PUFA eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) (Albracht-Schulte et al., 2018; Ratnayake and Galli, 2009; Jafarnejad and Sadegh, 2011; Li et al., 2018). Omega-3 PUFAs are asserted to exert their favorable effects via the modulation of the amount and type of eicosanoids and by the regulation of both intracellular signaling pathways and transcription factor activities (Gray et al., 2013; Serhan et al., 2004; Babcock et al., 2003). Gene expression is another example of the possible mechanism of *n*-3 PUFA consumption in positively affecting health indices (Gray et al., 2013; Ren et al., 1997). It seems that each of the aforementioned mechanisms, or indeed a combination of them, contributes to the regulation of different aspects of PCOS, including inflammation, blood pressure regulation, platelet adhesion and heart rhythm (Gray et al., 2013; Harris et al., 2008).

It has been shown in previous trials that consumption of unsaturated fat-rich foods may decrease the risk of chronic diseases; which is particularly true regarding risk factors for metabolic disorders seen in PCOS patients, including dyslipidemia, impaired endothelial function, and insulin resistance (Faghfoori et al., 2017; Zivkovic et al., 2007). To our knowledge, in spite of several trials exploring the beneficial effects of *n*-3 PUFA on PCOS (Table 1), no comprehen-

sive review regarding the effects of *n*-3 PUFA consumption on PCOS complications has taken place. Therefore, our aim was to conduct a review to investigate the possible

effect and related mechanisms of *n*-3 PUFA consumption on PCOS complications.

**Table 1:** Characteristics of trials investigating effects of *n*-3 PUFAs on PCOS complications

Study name	Year	Sample size	Follow-up duration (month)	Intervention dosage	Significant outcome
Oner	2013	45	6	1,500 mg of omega-3	BMI, insulin and HOMA levels decreased significantly during treatment, but glucose levels did not change. In the hormonal profile, serum LH and testosterone levels decreased and sex hormone-binding globulin levels increased significantly.
Khani	2017	87	6	2 g/day (two capsules) omega-3 supplements	Waist circumference (WC) was significantly lower in omega-3 as compared to control.
Rafraf	2012	61	2	four 1-g omega-3 fatty acids capsules/d (1,200 mg <i>n</i> -3)	No significant effects on weight, BMI, waist circumference, and waist to hip ratio at the end of the study. Changes in serum visfatin levels were not significant in either of the groups.
McEwen	2017	88	6	2 capsules of omega-3/d (total equivalent to 360 mg EPA and 240 mg DHA/day)	Waist circumference was significantly lower in the omega-3 group as compared to control. The measurements of weight and hip circumference did not change.
Nomura	2018	72	12	pitavastatin with either sarpogrelate (PS) or EPA (PE)	BMI decreased significantly.
Amini	2018	60	3	2×1,000 mg/day fish oil omega-3 fatty acid	The quantitative insulin sensitivity check index increased but serum insulin levels and HOMA-IR significantly decreased.
Mirmasoumi	2018	60	3	1,000 mg flaxseed oil omega-3 fatty acids	Insulin values and homeostasis model of assessment-estimated insulin resistance decreased but quantitative insulin sensitivity check index and hs-CRP significantly decreased compared to the placebo.
Ebrahimi	2017	68	3	1,000 mg omega-3 fatty acids from flaxseed oil containing 400 mg $\alpha$ -linolenic acid plus 400 IU vitamin E supplements	A significant decrease in insulin, homeostasis model of assessment-estimated insulin resistance, a significant increase in quantitative insulin sensitivity check index and any significant effect on fasting plasma glucose and also significant reductions in serum total testosterone and free testosterone was observed.
Jamilian	2018	60	3	50,000 IU vitamin D every 2 weeks plus 2000 mg/day omega-3 fatty acid from fish oil	Serum hs-CRP significantly decreased, gene expression of IL-1 significantly downregulated, a significant effect on gene expression of IL-8, TNF- $\alpha$ and TGF- $\beta$ occurred. Serum total testosterone levels decreased, but other hormonal measures didn't change.

**Table 1 (cont.):** Characteristics of trials investigating effects of *n*-3 PUFAs on PCOS complications

Study name	Year	Sample size	Follow-up duration (month)	Intervention dosage	Significant outcome
Rahmani	2017	68	3	1,000 mg omega-3 fatty acids from flaxseed oil containing 400 mg $\alpha$ -linolenic acid plus 400 IU vitamin E supplements	Gene expression of PPAR- $\gamma$ upregulated, gene expression of IL-1 and IL-8 downregulated, but any significant effect on TNF- $\alpha$ and TGF- $\beta$ wasn't observed.
Talari	2018	60	3	1,000 mg omega-3 from flaxseed oil containing 400 mg $\alpha$ -linolenic acid plus 400 IU vitamin E supplements	Change in hs-CRP was significantly different between the intervention and placebo group. Significant decreases in maximum levels of left CIMT, mean left CIMT levels, maximum levels of right CIMT and mean right CIMT levels and also any significant effect on NO values between the two groups was observed.
Mohammadi	2012	64	2	4 omega-3 fatty acids capsules (each one contained 180 mg EPA and 120 mg DHA)	Levels of adiponectin and HDL-C increased but TC, LDL-C and TG decreased.
Mejia-Montilla	2018	195	3	omega-3 fatty acids	Mean of adiponectin levels showed a statistically significant increase after treatment.
Nadjarzadeh	2015	84	2	3 capsules of omega-3 (each one contained 180 mg EPA and 120 mg DHA) daily	Visfatin concentration did not change in neither groups, but the mean of adiponectin concentration increased in omega-3 group.
Cussions	2009	25	2	4 g/d of omega-3 fatty acids	The mean of LH decreased about 1.74 mIU/ml in omega-3 group. The mean of change of LH/FSH ratio between groups was significant and after the intervention, prolactin and FSH did not meaningfully change in both groups.
Phelan	2011	22	6 (weeks)	LC <i>n</i> -3 PUFA supplement	Fasting and postprandial plasma triacylglycerol, apoB48, total cholesterol, HDL-C, and LDL-C concentrations did not change.
Kalgaonkar	2011	31	6 (weeks)	walnuts or almonds containing 31 g of total fat per day	Walnuts increased sex hormone-binding globulin and almonds reduced free androgen index

### IMPACT OF OMEGA-3 ON PCOS COMPLICATIONS:

#### *Omega-3 and obesity*

Overweight and obesity are observed in 40 %–50 % of patients with PCOS and is considered a major risk factor for PCOS (Liepa et al., 2008; Carmina and Lobo, 1999). The accumulation of excess weight can lead to adipose tissue dysfunction, a phenomenon that is mechanistically connect-

ed to the pathogenesis of MetS and complications such as insulin resistance in liver and skeletal muscle (Albracht-Schulte et al., 2018; Kim and Moustaid-Moussa, 2000). It is assumed that lower serum *n*-3 PUFA concentrations (Albracht-Schulte et al., 2018; Karlsson et al., 2006), particularly DHA (increase of *n*-6:*n*-3 ratio) (Albracht-Schulte et al., 2018; Karlsson et al., 2006), are associated with obesity, elevated waist circumfer-

ence (Albracht-Schulte et al., 2018; Micallef et al., 2009) in adolescents (Karlsson et al., 2006; Albracht-Schulte et al., 2018), and adults with obesity (Albracht-Schulte et al., 2018; Micallef et al., 2009). Adipose tissue is considered an active endocrine organ which has the ability to secrete several adipokines and hormones, such as leptin and adiponectin, and cytokines, such as interleukin-6 (IL-6) (Albracht-Schulte et al., 2018; Kalupahana et al., 2012).

One of the primary treatments in managing obesity and its comorbidities is lifestyle modification with the aim of significant weight loss (Albracht-Schulte et al., 2018). Natural bioactive compounds, such as *n*-3 PUFA, possess few side effects and could be considered a safe approach in comparison to other modalities of treatment (Albracht-Schulte et al., 2018). There are a variety of putative mechanisms by which *n*-3 PUFA, particularly EPA and DHA, could work in improving body composition, modulating energy metabolism and reducing body weight (Albracht-Schulte et al., 2018).

Adipocyte differentiation, which is considered a complex process in making adipose tissue, is mainly regulated by two different families of transcription factors, including peroxisome proliferator-activated receptor  $\gamma$  (PPAR- $\gamma$ ) and CCAAT/enhancer-binding proteins (C/EBPs) (Martinez-Fernandez et al., 2015). It has been shown that DHA, in dosages of  $\geq 50 \mu\text{M}$ , could facilitate the differentiation of adipocytes by up-regulation of mRNA levels of adipocyte protein 2 (aP2) and C/EBP $\alpha$ , PPAR $\gamma$  in 3T3-L1 cell lines (Martinez-Fernandez et al., 2015; Murali et al., 2014). Furthermore, accumulation of lipid droplets results in increased levels of PPAR $\gamma$  following administration of EPA (250  $\mu\text{M}$ ) (Martinez-Fernandez et al., 2015; Hanada et al., 2011). Additionally, previous studies have highlighted that *n*-3 PUFAs can modulate the apoptosis of adipocytes (Martinez-Fernandez et al., 2015; Kim et al., 2006). Collectively, it seems that marine-origin *n*-3 PUFAs may have the potential to regulate the adipocyte number and size via

modulating adipocyte differentiation and apoptosis (Martinez-Fernandez et al., 2015). Indeed, several studies have reported favorable outcomes of omega-3 consumption on obesity. For instance, in Oner and Muderris (2013), BMI decreased significantly with daily oral administration of 1,500 mg omega-3 for 6 months (Oner and Muderris, 2013). In another trial, which was conducted in PCOS patients, the subjects consumed omega-3 in doses of 2 g/day for 6 months, after which, waist circumference (WC) was significantly lower in those that consumed omega-3 vs. the control group (Khani et al., 2017). Similar outcomes were reported in McEwen (2017), where participants received 2 capsules of omega-3 PUFA acids per day, and at the end of the trial, WC was significantly lower in the omega-3 group vs. the control group (McEwen, 2017). The findings of combination therapy have also shown promising effects, for instance, BMI decreased significantly following administration of pitavastatin and, either, EPA 1,800 mg or sarpogrelate 300 mg for 12 months in diabetic patients with arteriosclerosis (Nomura et al., 2018). Conversely, it has been reported that consuming 4 g of omega-3 PUFA capsules, per day, for 8 weeks did not elicit any significant effect on weight, BMI, waist circumference, or waist to hip ratio (Rafraf et al., 2012). However, the contradictory results are limited, and most previous studies have reported favorable effects of *n*-3 PUFA supplementation on obesity.

### ***Omega-3 and insulin resistance***

Insulin resistance is defined as a condition in which the response of peripheral tissue to insulin is reduced (Lepretti et al., 2018). This disorder occurs mostly in the skeletal muscle and the liver, with several negative impacts on glucose metabolism, which is mostly attributed to the defective regulation of glucose transporter isoform 4 (GLUT4). It seems that the skeletal muscle may play a substantial role in insulin resistance (Lepretti et al., 2018; DeFronzo and Tripathy, 2009; Lark et al., 2012). Impaired

suppression of gluconeogenesis, glycogenolysis in the liver, and suppression of glucose output are among the complications associated with insulin resistance (Home and Pacini, 2008; Boden, 2011; Lepretti et al., 2018). The impaired suppression of lipolysis observed in white adipose tissue (WAT) contributes to hyperlipidemia in insulin-resistant subjects (Lepretti et al., 2018; Boden, 2011). Brown adipose tissue (BAT) also plays an essential role in glucose metabolism (Lepretti et al., 2018; Stanford et al., 2013), where it is demonstrated that the whole-body energy homeostasis can be regulated by BAT, with the dissipation of chemical energy as heat via mitochondrial uncoupling protein 1 (UCP1).

Insulin resistance has been reported in 30 % of lean and 75 % of obese patients with PCOS, whilst the severity of insulin resistance is more than would be expected for a given age and body weight (O'Connor et al., 2010; Jonard and Dewailly, 2004). The observed fasting hyperinsulinemia in obese PCOS patients is secondary to increased basal insulin secretion rates (Dunaif, 1997; O'Meara et al., 1993). Obesity results in insulin resistance, which is accountable for the pathogenesis of MetS (Albracht-Schulte et al., 2018; Kim and Moustaid-Moussa, 2000). Previous studies have demonstrated an improvement in insulin resistance after weight loss interventions (Albracht-Schulte et al., 2018; Kim and Moustaid-Moussa, 2000) and several studies recommend that insulin resistance leads to significant disturbances in both metabolic and reproductive homeostasis, and thus may possess an essential role in the pathogenesis of PCOS (Gunalan et al., 2018; Diamanti-Kandarakis and Dunaif, 2012). In brief, insulin may exert its role in hyperandrogenism in PCOS pathophysiology via two distinct pathways: 1) stimulation of androgen production of theca cells with luteinizing hormone (LH) and increased androgen production which results in hirsutism and infertility. 2) Suppression of sex hormone-binding globulin (SHBG) synthesis in the liver (Gunalan et al., 2018; Ehrmann,

2005). SHBG is considered to be a plasma protein for androgen and estrogens; therefore, any decrement in SHBG levels could result in hyperandrogenism in PCOS sufferers. Metabolically, insulin plays an integral role in regulating glucose homeostasis, the activation of amino acid transportation, and suppression of lipolysis (Gunalan et al., 2018; Kahn, 1994). Mechanistically, it has been suggested that omega-3 PUFA may have beneficial effects on insulin sensitivity by attenuating endoplasmic reticulum (ER) stress, and increasing  $\beta$ -oxidation of mitochondrial fatty acid and mitochondrial uncoupling, with a concomitant reduction in lipid deposits and reactive oxygen species (ROS) production. Consequently, inflammatory processes are down-regulated, thereby enhancing insulin sensitivity. Another putative mechanism is related to mitochondrial fusion proteins - mitofusins 2 (Mfn2), which is involved in the maintenance of mitochondrial dynamics homeostasis and integrity of mitochondria-associated endoplasmic reticulum membrane (MAM). In the case of cellular stress, MAM integrity and mitochondrial fusion phenotype may facilitate maintenance of insulin sensitivity (Lepretti et al., 2018).

There are some studies that show the effect of omega-3 supplementation on insulin resistance. In the study of Oner and Muderris (2013) insulin levels significantly decreased following 6 months of omega-3 consumption; whereas there were no reported significant changes in glucose levels and HOMA index. In another trial, 30 participants consumed 2 g/day fish oil omega-3 fatty acid for 12 weeks; following which, a significant increase in the quantitative insulin sensitivity check index, serum insulin levels, and HOMA-IR were reported, respectively (Amini et al., 2018). Mirmasoumi et al. reported a decrease in insulin values and homeostasis model of assessment-estimated insulin resistance, and an increase in quantitative insulin sensitivity check index after administering 1 g/day flaxseed oil omega-3 fatty acids for 12 weeks (Mirmasoumi et al., 2018). Similarly, a significant decrease in in-

sulin, homeostasis model of assessment-estimated insulin resistance and a significant increase in quantitative insulin sensitivity check index were observed after 12 weeks intervention of omega-3 fatty acids and vitamin E co-supplementation (Ebrahimi et al., 2017). However, in a recent meta-analysis which investigated the effect of omega-3 PUFA supplementation on insulin resistance in women with PCOS, no beneficial effect on insulin resistance and HOMA-IR was observed in comparison to the placebo group (Sadeghi et al., 2017).

### ***Omega-3 and inflammation***

Chronic low-grade inflammation is normally involved in the pathogenesis of obesity-related disorders. Leukocytes in the adipose-tissue and circulation can enhance the insulin resistance in type 2 diabetes and obesity (Escobar-Morreale et al., 2011; Fernandez-Real and Ricart, 2003). Polycystic ovary syndrome contributes to a pro-inflammatory state (Escobar-Morreale et al., 2011), and an association between molecular level inflammation and insulin resistance has been well reported (Escobar-Morreale et al., 2011; González et al., 2006a, b). Moreover, there are elevations in several circulating pro-atherogenic inflammatory modulators in PCOS (Escobar-Morreale et al., 2011; Diamanti-Kandarakis et al., 2006a, b; Lewandowski et al., 2006; Hu et al., 2006), which are accompanied by glucose-stimulated up-regulation of pro-atherogenic molecular pathways (Escobar-Morreale et al., 2011; González et al., 2006a). However, it remains unknown whether the pro-inflammatory state and increase of circulating inflammatory mediators in PCOS are related to PCOS itself, or are primary manifestations of the inflamed status of adipose tissue (Escobar-Morreale et al., 2011; Carmina et al., 2007).

*N-3* PUFAs influence the production of several inflammatory markers, such as adhesion molecules and cytokines (Calder, 2017), whilst the most focused markers are the classic pro-inflammatory cytokines, tumor ne-

crisis factor (TNF)- $\alpha$ , IL-6 and interleukin (IL)-1 $\beta$ . Many cell culture and animal reports, respectively, have reported that EPA/DHA can reduce the formation of TNF- $\alpha$ , IL-6, and IL-1 $\beta$  in response to lipopolysaccharide (LPS) exposure (Calder, 2017, 2015). Whilst other studies have shown that EPA and DHA may lead to an increase in the cytokine IL-10 concentration, which is anti-inflammatory (Calder, 2017, 2015). Additionally, it has been reported that *n-3* PUFAs, especially DHA, could decrease the expression of adhesion molecules, such as intercellular adhesion molecule (ICAM)-1 and vascular cell adhesion molecule (VCAM)-1, on the surface of monocytes and endothelial cells (Calder, 2017, 2015). It seems that the alterations in the expression of genes responsible for encoding the cytokines and inflammatory mediators induce the anti-inflammatory effects of *n-3* PUFAs (Calder, 2017, 2015). This suggests that *n-3* PUFAs impact the signaling pathways, and, as a consequence, control gene expression in inflammatory cells.

Another mechanism by which EPA and DHA might have an effect on nuclear factor kappa B (NF- $\kappa$ B) activation, and the ability to induce transcription of pro-inflammatory genes, is related to PPAR- $\gamma$ . This is considered to be a transcription factor which acts in an anti-inflammatory manner, which is likely attributable to its interference with the translocation of NF- $\kappa$ B to the nucleus. It has been reported that *n-3* PUFAs can induce the PPAR- $\gamma$  (Calder, 2017; Forman et al., 1997; Krey et al., 1997) and DHA, specifically, can activate PPAR- $\gamma$  in dendritic cells (Calder, 2017; Kong et al., 2010); which is accountable for inhibition of NF- $\kappa$ B activation and reduced production of the pro-inflammatory cytokines, including TNF- $\alpha$  and IL-6 and LPS stimulation (Calder, 2017; Kong et al., 2010). Furthermore, PPAR- $\gamma$  target genes in dendritic cells are induced by DHA (Calder, 2017; Zapata-Gonzalez et al., 2008), suggesting that it may be a considerable anti-inflammatory mechanism attributable to DHA. Similarly, EPA derivatives, 15-



deoxy-PGD3 and PGD3, can induce PPAR- $\gamma$  in adipocytes, which results in the activation of the anti-inflammatory adipokine adiponectin production (Calder, 2017; Lefils-Lacourtablaise et al., 2013).

There are several reports suggesting that *n*-3 PUFA can improve inflammatory status in PCOS patients (Jamilian et al., 2018; Talari et al., 2018). In one study, co-supplementation of 3 g/day omega-3 from fish oil, with 50000 IU vitamin D, significantly decreased serum high-sensitivity C-reactive protein (hs-CRP), and down-regulated gene expression of IL-1; however, there was no significant effect on gene expression of IL-8, TNF- $\alpha$  or transforming growth factor beta (TGF- $\beta$ ) (Jamilian et al., 2018). In another study, flaxseed oil omega-3 fatty acid supplementation resulted in a significant decrease in hs-CRP (Mirmasoumi et al., 2018), whilst Rahmani et al. (2017) demonstrated that consumption of 1 g/day omega-3 fatty acids from fish oil, for 12-weeks, resulted in up-regulation of PPAR- $\gamma$  and down-regulation in gene expression of IL-1 and interleukin-8 (IL-8) (Rahmani et al., 2017). Evidently, most previous studies emphasize the efficacy of *n*-3 PUFA administration in relieving inflammatory status in PCOS patients.

### ***Omega-3 and adipokines***

The adipokine alteration is considered to be a specific complication of PCOS, yet it is not fully understood whether such dysregulation is secondary to PCOS complications such as obesity, hyperinsulinemia, and hyperandrogenism, or directly related to PCOS (Baldani et al., 2019). Adiponectin and leptin are among the adipokines secreted from adipocytes (Farimani et al., 2018; Das et al., 2013), and it has been shown that different adipokines can promote both inflammatory and anti-inflammatory activities (Farimani et al., 2018; Nomura et al., 2009). It seems that adiponectin expression as an anti-inflammatory adipokine is reduced in obese individuals, whilst expression of leptin as an inflammatory adipokine is elevated, com-

pared to the lean subjects (Farimani et al., 2018; Kang et al., 2016; Ouchi et al., 2011). The higher leptin concentration in obese subjects, which is due to leptin resistance, is related to higher concentrations of inflammatory markers, including TNF- $\alpha$  and CRP. In addition to the inflammatory responses of adipokines, it has been suggested that adiponectin and leptin can regulate appetite and energy expenditure, which results in an alteration of insulin sensitivity. (Farimani et al., 2018; Gray et al., 2013). *N*-3 PUFA's derived from plant and marine sources have been introduced as potential factors affecting the concentration of inflammatory and non-inflammatory adipokines (Farimani et al., 2018; von Frankenberg et al., 2014). Elevation of circulating adiponectin levels and reduced levels of leptin have been reported after *n*-3 PUFAs administration in previous studies (Farimani et al., 2018; von Frankenberg et al., 2014; Hariri et al., 2015), which is associated with improved function of skeletal muscle mitochondria, and, consequently, increased glucose uptake. *N*-3 PUFA normalizes the secretory activity of adipocytes and improves adiponectin-mediated insulin sensitivity in skeletal muscle and the liver (Farimani et al., 2018). Moreover, this can stimulate the activation of 5'AMP-activated protein kinase (AMPK) and PPAR $\gamma$ , which regulate metabolism by increasing glucose consumption and fatty acid oxidation in the liver and skeletal muscle (Farimani et al., 2018; Gray et al., 2013; Perez-Matute et al., 2007; Tomas et al., 2002; Yamauchi et al., 2003). However, it seems that the effect omega-3 elicits on adipokine levels is equivocal, and widely dependent on several parameters, such as diet composition, the physiological and metabolic status of subjects (Farimani et al., 2018; Moreno-Aliaga et al., 2010).

There are several studies related to the effect of omega-3 fatty acids on adipokine secretion, for instance, Mohammadi et al. reported an increase of adiponectin by 19.5 % via inducing omega-3 fatty acids supplementation for 8 weeks (Mohammadi et al., 2012). In another study by Mejia-Montilla et al.

(2018), PCOS patients treated with omega-3 fatty acids for 12 weeks significantly increased mean levels of adiponectin. Similarly, in a study conducted in PCOS sufferers, following omega-3 supplementation (3 caps/day, each containing 180 mg EPA and 120 mg DHA) for 8 weeks, mean adiponectin concentration increased in the omega-3 group, whilst visfatin concentration remained unchanged (Nadjarzadeh et al., 2015). In Rafrat et al. (2012), patients with PCOS took 4 g omega-3 fatty acids capsules per day, providing 1200 mg *n-3* long-chain polyunsaturated fatty acids (*n-3* LC PUFA), for 8 weeks. The incumbent changes in serum visfatin levels were not significant in either the intervention or placebo groups (Rafrat et al., 2012). In summary, the studies investigating the effects of *n-3* PUFA on adipokines, particularly in PCOS, are limited, and further studies are warranted to clarify the possible efficacy of, and mechanisms associated with, improving PCOS complications.

### ***Omega-3 and lipid metabolism***

Significant changes in the production of FFAs, phospholipids, and bioactive lipids have been observed in obese and lean women with PCOS. The reported dyslipidemia in the PCOS population is characterized by reduced high density lipoprotein (HDL) cholesterol concentrations as the main outcome, and possible elevated levels of triacylglycerol (TAG), very low density lipoprotein (VLDL) and/or atherogenic non-A low density lipoprotein (LDL) cholesterol pattern as the secondary outcome (O'Connor et al., 2010; Westerveld et al., 2008).

There are several studies that have reported an inverse association between plasma HDL-C levels and the incidence of CVD disorders (Pizzini et al., 2017; Rader, 2003; Kannel et al., 1964; Assmann et al., 2002), which emphasizes the potential atheroprotective effects of higher concentrations of HDL-C (Pizzini et al., 2017; Siddiqi et al., 2015). The beneficial effect of HDL as a CVD protecting factor is mediated by the Reverse

Cholesterol Transport (RCT) activity of HDL-C. This process explains the cholesterol clearing pathways of peripheral, subendothelial macrophage, and fibroblast-derived cholesterol via direct (via hepatic uptake via scavenger receptor B-I (SR-BI)), and indirect (via shifting cholesterol from HDL particles to apoB-containing lipoproteins) pathways (Mahdy Ali et al., 2012; Siddiqi et al., 2015; Pizzini et al., 2017). Another potential mechanism by which *n-3* PUFAs may exhibit their atheroprotective activities is through enhancement of intracellular catabolism of apolipoprotein B-100 lipoproteins and curbing hepatic apo-B production. This leads to stimulation of lipoprotein lipase (LPL), and increases plasma triglyceride clearance, conversion of the VLDL to LDL, and reducing the synthesis of LDL and reducing postprandial lipemia (Pizzini et al., 2017; Chan et al., 2002, 2003; Park and Harris, 2003; Robinson and Stone, 2006). It seems that *n-3* PUFAs exert their effect by inhibiting sterol regulatory element-binding protein-1 (SREBP-1) mediated pathways, including the activation of Liver X Receptor (LXR), the nuclear transcription factors, Farnesoid X Receptor, Hepatocyte Nuclear Factor-4 Alpha (HNF4A), and Peroxisome Proliferator-activated Receptors (PPARs) (Pizzini et al., 2017; Zuliani et al., 2009). Different types of *n-3* PUFA may elicit different activities regarding lipid metabolism; unlike DPA and DHA, which are deposited in the tissues and spared from catabolism, EPA is mostly directed towards  $\beta$ -oxidation (Albracht-Schulte et al., 2018; Ghasemifard et al., 2015). Additionally, it has been revealed that co-supplementation with EPA/ DHA or single supplementation of DHA leads to a decrease in gene expression of fatty acid synthase, LPL, hormone-sensitive lipase (HSL), and phosphoenolpyruvate carboxykinase (PEPCK) in retroperitoneal fat; however, such effects were not evident in EPA supplementation alone (Albracht-Schulte et al., 2018; Raclot et al., 1997).

Although there are numerous reports of dyslipidemia in PCOS, studies investigating

the effects of *n*-3 PUFA on lipid metabolism in PCOS patients are limited. However, several studies have reported an improvement in lipid regulation after *n*-3 PUFA supplementation in other metabolic disorders. In a study by Cussons et al. (2009) omega-3 fatty acids led to a decrease in hepatic fat in PCOS women with hepatic steatosis. In another study, omega-3 fatty acids supplementation, in PCOS patients, significantly increased HDL-C, and decreased TC, LDL-C and TG (Mohammadi et al., 2012); whilst in Rahmani et al. (2017), 12 weeks co-supplementation of omega-3 fatty acids and vitamin E resulted in a significant decrease in serum triglycerides, VLDL, total, LDL- and total-/HDL-cholesterol in PCOS patients (Rahmani et al., 2017). However, contrary to previous studies, Phelan et al. (2011) reported no significant improvement in fasting and postprandial plasma triacylglycerol, apo-B48, total cholesterol, HDL cholesterol, and LDL cholesterol concentrations in response to LC *n*-3 PUFA supplementation for 6 weeks in PCOS sufferers (Phelan et al., 2011). Notwithstanding, as only a limited number of trials have investigated *n*-3 PUFA supplementation as a conduit to improve PCOS complications, and that different types of *n*-3 PUFA may exert different activities and functions, more studies are warranted to clarify the mechanisms and effects of *n*-3 PUFA on lipid metabolism in PCOS.

### ***Omega-3 and endothelial function***

In addition to dyslipidemia, endothelial dysfunction, which is defined as the reduction of vasodilatory and anti-inflammatory parameters and an increase in vasoconstrictive and pro-inflammatory parameters, is considered to be another significant indicator in women with PCOS which may lead to an increased risk for atherosclerotic cardiovascular (CV) diseases (Wenner et al., 2011; Abbott et al., 2002). Evidence from previous studies has demonstrated that the cardioprotective effects of *n*-3 PUFAs are manifest through improvements in vascular function. Several mechanisms, including decreased ar-

terial plaque build-up (Zehr and Walker, 2018; Renier et al., 1993), increased anti-inflammatory properties (Zehr and Walker, 2018; Calder, 2006), enhanced endothelial-dependent vasodilation as measured by flow-mediated dilation (FMD) (Zehr and Walker, 2018; Wiest et al., 2017; Goodfellow et al., 2000; Siasos et al., 2013), lowering blood pressure (Zehr and Walker, 2018; Ulu et al., 2014; Hoshi et al., 2013; Miller et al., 2014), elevated antioxidant capacity (Zehr and Walker, 2018; Thorlaksdottir et al., 2006) and production of nitric oxide (Albracht-Schulte et al., 2018; Harris et al., 1997) have been suggested. In spite of the apparent dysfunction of endothelial function in PCOS women, there is a dearth of related studies investigating the potential beneficial effects of *n*-3 PUFA, except for Talari et al. (2018), who indicated a significant decrease in maximum levels of left carotid intima-media thickness (CIMT), mean left CIMT levels, maximum levels of right CIMT, and mean right CIMT levels after 12-week co-supplementation with Omega-3 and vitamin E (Talari et al., 2018).

### ***Omega-3 and hormonal factors***

Hyperandrogenism is considered to be a basic characteristic of PCOS (Gunalan et al., 2018; Stein et al., 2016), and is often associated with excessive release of luteinizing hormone (LH) by the pituitary gland and is related with hyperinsulinemia and insulin resistance (Liepa et al., 2008; Ehrmann, 2005). The mechanisms by which insulin resistance impacts the hormonal system are numerous, moreover, it has been shown that insulin induces elevated androgen production and produces a hormonal milieu which results in hirsutism. The improvement in the reproductive system has been attributed to an improvement in LH concentration and LH/Follicle stimulating hormone (FSH) ratio, where the potential mechanistic action can be attributed to arachidonic acids (Nadjarzadeh et al., 2015; Phelan et al., 2011). Arachidonic acid can activate a steroidogenic acute regulatory protein

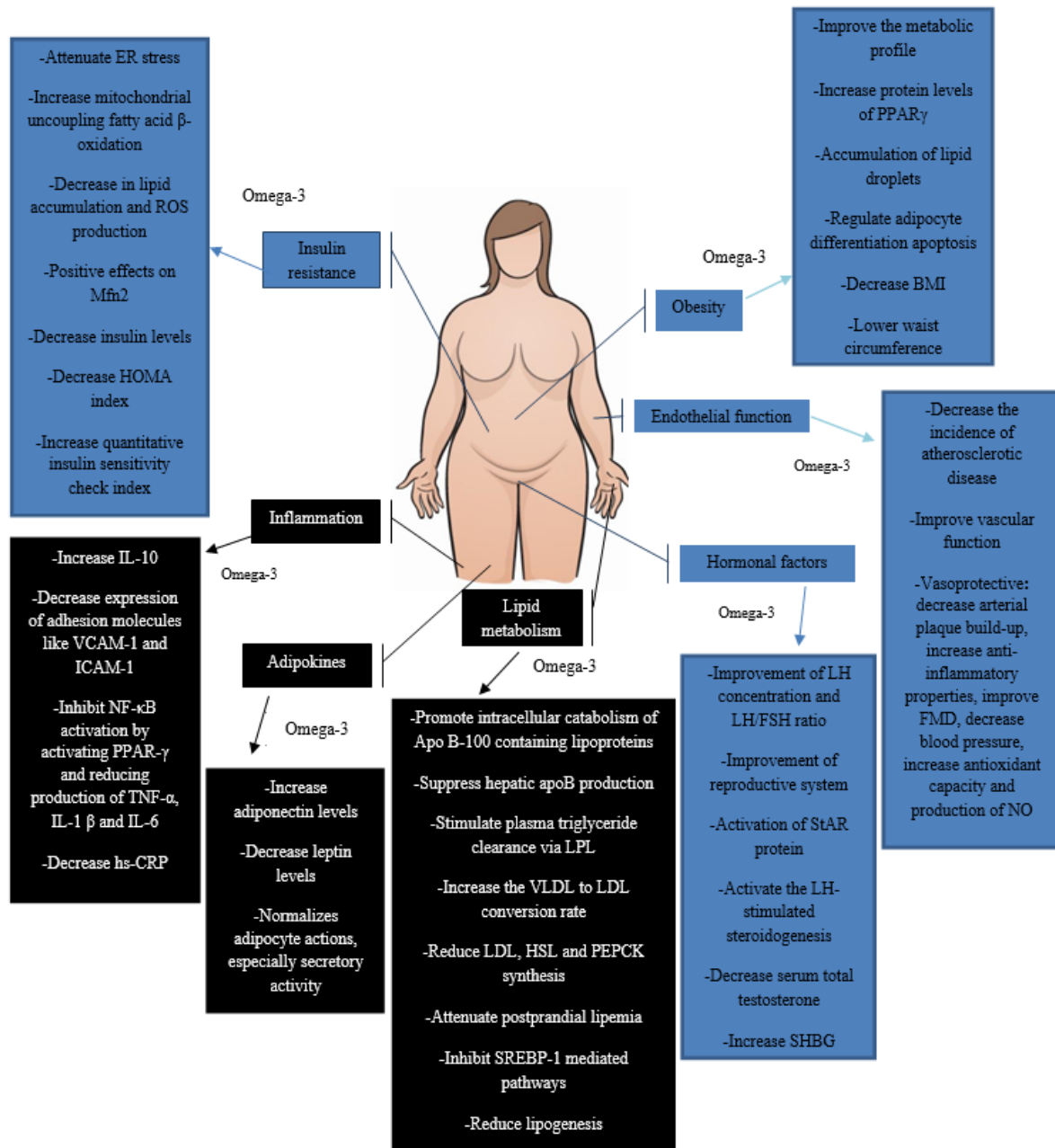
(StAR), which is considered to be the rate-limiting tenet of the steroidogenic pathway. StAR transfers the cholesterol to the inner part of the mitochondrial membrane, which is considered the first and the most important step in the steroidogenic pathway and leads to the production of both androstenedione and testosterone. Therefore, high concentrations of arachidonic acid can result in LH-stimulated steroidogenesis, whilst current evidence suggests that the reduced availability of arachidonic acid is related to levels of omega-3 (Nadjarzadeh et al., 2015; Phelan et al., 2011).

Jamilian et al. (2018) reported that a decreased total serum level of testosterone was evident after co-supplementation of vitamin D and omega-3 fatty acid (50,000 IU vitamin D every 2 weeks plus 2000 mg/day omega-3 fatty acid from fish oil) for 12 weeks, whilst in another study, supplementing with omega-3 fatty acids from flaxseed oil plus vitamin E for 12 weeks in PCOS women, resulted in significant reductions in serum total testosterone and free testosterone (Ebrahimi et al., 2017). A similar effect of omega-3 consumption on hormonal factors in PCOS women was observed in Oner and Muderris (2013); where the participants were treated with 1,500 mg of omega-3, daily, for 6 months. Resultantly, serum LH and testosterone levels decreased and sex hormone-binding globulin levels increased significantly (Oner and Muderris, 2013). Contrastingly, in Nadjarzadeh et al. (2015), PCOS women supplemented with 3 capsules of omega-3 (each one contained 180 mg EPA and 120 mg DHA) daily for 8 weeks, which resulted in no change in FSH. However, mean LH decreased by 1.74 mIU/ml in the omega-3 group, and the mean change in LH/FSH ratio between groups was significant; moreover, the authors reported no meaningful change in prolactin levels (Nadjarzadeh et al., 2015). The aforementioned studies have predominantly explored the effect of *n*-3 PUFA supplements, Kalgaonkar et al., however, focused on dietary intake. In Kalgaonkar et al.

(2011), thirty-one PCOS patients consumed either walnuts or almonds containing 31 g of total fat per day for 6 weeks. Walnut consumption increased sex hormone-binding globulin, whilst almond consumption reduced free androgen index (Kalgaonkar et al., 2011).

## CONCLUSIONS

*N*-3 PUFAs are a promising agent in relieving complications associated with PCOS. There are multiple mechanisms by which *n*-3 PUFAs may exert their beneficial effects on PCOS women, including anti-obesity, glycemic and hormonal homeostasis, anti-inflammatory, regulation of adipokine production and enhancement of endothelial function (Figure 1). Whilst studies investigating the effect of omega-3 supplements in patients with PCOS have reported an improvement in numerous complications and comorbidities, including insulin resistance, dyslipidemia, hyperandrogenism, and regulation of metabolic indicators; there is a distinct dearth of studies investigating the dietary intake of these types of fatty acids. Therefore, more studies are needed to investigate the effect of dietary fatty acids on PCOS complications. Moreover, there are, evidentially, numerous beneficial effects associated with *n*-3 PUFA consumption on PCOS complications, however, favorable effects regarding the improvement of dyslipidemia, regulation of adipokines and regulation of hormonal factors and endothelial function are limited. Therefore, given the importance of these complications in the pathogenesis of PCOS, more trials are warranted to investigate mechanisms to clarify the metabolic and hormonal effects of these agents in PCOS women. Additionally, to our knowledge, the use of *n*-3 PUFA on infertile PCOS patients has not been investigated in previous studies, and more studies are necessary to assess the role of *n*-3 PUFA in the treatment of PCOS induced infertility.



**Figure 1:** Possible mechanisms describing the effects of *n*-3 PUFAs on PCOS complications. PPAR $\gamma$ , peroxisome proliferator-activated receptor  $\gamma$ ; ER, endoplasmic reticulum; ROS, reactive oxygen species; Mfn2, mitochondrial fusion proteins—mitofusins 2; IL-, interleukin; TNF- $\alpha$ , tumour necrosis factor; VCAM-1, vascular cell adhesion molecule; ICAM-1, intercellular adhesion molecule; NF- $\kappa$ B, nuclear factor kappa B; hs-CRP, high-sensitivity C-reactive protein; LPL, lipoprotein lipase; VLDL, very low density lipoprotein; LDL, low density lipoprotein; HSL, hormone-sensitive lipase; PEPCK, phosphoenolpyruvate carboxykinase; SREBP-1, sterol regulatory element-binding protein-1; FMD, flow-mediated dilation; NO, nitric oxide; LH, luteinizing hormone; FSH, follicle stimulating hormone; StAR, steroidogenic acute regulatory protein; SHBG, sex hormone-binding globulin

### Conflict of interest

The authors declare no conflicts of interest associated with the present manuscript.

### REFERENCES

Abbott DH, Dumesic DA, Franks S. Developmental origin of polycystic ovary syndrome - a hypothesis. *J Endocrinol.* 2002;174(1):1-5.

- Albracht-Schulte K, Kalupahana NS, Ramalingam L, Wang S, Rahman SM, Robert-McComb J, et al. Omega-3 fatty acids in obesity and metabolic syndrome: a mechanistic update. *J Nutr Biochem.* 2018; 58:1-16.
- Amini M, Bahmani F, Foroozafard F, Vahedpoor Z, Ghaderi A, Taghizadeh M, et al. The effects of fish oil omega-3 fatty acid supplementation on mental health parameters and metabolic status of patients with polycystic ovary syndrome: a randomized, double-blind, placebo-controlled trial. *J Psychosom Obstet Gynaecol.* 2018; epub ahead of print.
- Assmann G, Cullen P, Schulte H. Simple scoring scheme for calculating the risk of acute coronary events based on the 10-year follow-up of the prospective cardiovascular Munster (PROCAM) study. *Circulation.* 2002;105:310-5.
- Asuncion M, Calvo RM, San Millan JL, Sancho J, Avila S, Escobar-Morreale HF. A prospective study of the prevalence of the polycystic ovary syndrome in unselected Caucasian women from Spain. *J Clin Endocrinol Metab.* 2000;85:2434-8.
- Babcock TA, Kurland A, Helton WS, Rahman A, Anwar KN, Espat NJ. Inhibition of activator protein-1 transcription factor activation by omega-3 fatty acid modulation of mitogen-activated protein kinase signaling kinases. *JPEN J Parenter Enteral Nutr.* 2003;27:176-80; discussion 181.
- Baldani DP, Skrgatic L, Kasum M, Zlopasa G, Kralik Oguic S, Herman M. Altered leptin, adiponectin, resistin and ghrelin secretion may represent an intrinsic polycystic ovary syndrome abnormality. *Gynecol Endocrinol.* 2019;35:401-5.
- Boden G. Obesity, insulin resistance and free fatty acids. *Curr Opin Endocrinol Diabetes Obes.* 2011;18: 139-43.
- Calder PC. n-3 polyunsaturated fatty acids, inflammation, and inflammatory diseases. *Am J Clin Nutr.* 2006;83(6 Suppl):1505s-19s.
- Calder PC. Marine omega-3 fatty acids and inflammatory processes: Effects, mechanisms and clinical relevance. *Biochim Biophys Acta.* 2015;1851:469-84.
- Calder PC. Omega-3 fatty acids and inflammatory processes: from molecules to man. *Biochem Soc Trans.* 2017;45:1105-15.
- Carmina E, Lobo RA. Polycystic ovary syndrome (PCOS): arguably the most common endocrinopathy is associated with significant morbidity in women. *J Clin Endocrinol Metab.* 1999;84:1897-9.
- Carmina E, Bucchieri S, Esposito A, Del Puente A, Mansueto P, Orio F, et al. Abdominal fat quantity and distribution in women with polycystic ovary syndrome and extent of its relation to insulin resistance. *J Clin Endocrinol Metab.* 2007;92:2500-5.
- Chan DC, Watts GF, Barrett PH, Beilin LJ, Redgrave TG, Mori TA. Regulatory effects of HMG CoA reductase inhibitor and fish oils on apolipoprotein B-100 kinetics in insulin-resistant obese male subjects with dyslipidemia. *Diabetes.* 2002;51:2377-86.
- Chan DC, Watts GF, Mori TA, Barrett PH, Redgrave TG, Beilin LJ. Randomized controlled trial of the effect of n-3 fatty acid supplementation on the metabolism of apolipoprotein B-100 and chylomicron remnants in men with visceral obesity. *Am J Clin Nutr.* 2003;77:300-7.
- Cussons AJ, Stuckey BG, Walsh JP, Burke V, Norman RJ. Polycystic ovarian syndrome: marked differences between endocrinologists and gynaecologists in diagnosis and management. *Clin Endocrinol (Oxf).* 2005;62:289-95.
- Cussons AJ, Watts GF, Mori TA, Stuckey BG. Omega-3 fatty acid supplementation decreases liver fat content in polycystic ovary syndrome: a randomized controlled trial employing proton magnetic resonance spectroscopy. *J Clin Endocrinol Metab.* 2009;94: 3842-8.
- Das P, Bhattacharjee D, Bandyopadhyay SK, Bhattacharya G, Singh R. Association of obesity and leptin with insulin resistance in type 2 diabetes mellitus in Indian population. *Indian J Physiol Pharmacol.* 2013;57:45-50.
- DeFronzo RA, Tripathy D. Skeletal muscle insulin resistance is the primary defect in type 2 diabetes. *Diabetes Care.* 2009;32(Suppl 2):S157-63.
- Diamanti-Kandarakis E, Dunaif A. Insulin resistance and the polycystic ovary syndrome revisited: an update on mechanisms and implications. *Endocr Rev.* 2012; 33:981-1030.
- Diamanti-Kandarakis E, Paterakis T, Alexandraki K, Piperi C, Aessopos A, Katsikis I, et al. Indices of low-grade chronic inflammation in polycystic ovary syndrome and the beneficial effect of metformin. *Hum Reprod.* 2006a;21:1426-31.
- Diamanti-Kandarakis E, Alexandraki K, Piperi C, Protogerou A, Katsikis I, Paterakis T, et al. Inflammatory and endothelial markers in women with polycystic ovary syndrome. *Eur J Clin Invest.* 2006b;36:691-7.

- Dumesic DA, Abbott DH, Padmanabhan V. Polycystic ovary syndrome and its developmental origins. *Rev Endocr Metab Disord.* 2007;8:127-41.
- Dunaif A. Insulin resistance and the polycystic ovary syndrome: mechanism and implications for pathogenesis. *Endocr Rev.* 1997;18:774-800.
- Ebrahimi FA, Samimi M, Foroozanfard F, Jamilian M, Akbari H, Rahmani E, et al. The effects of omega-3 fatty acids and vitamin E co-supplementation on indices of insulin resistance and hormonal parameters in patients with polycystic ovary syndrome: a randomized, double-blind, placebo-controlled trial. *Exp Clin Endocrinol Diabetes.* 2017;125:353-9.
- Ehrmann DA. Polycystic ovary syndrome. *N Engl J Med.* 2005;352:1223-36.
- Escobar-Morreale HF, Luque-Ramirez M, Gonzalez F. Circulating inflammatory markers in polycystic ovary syndrome: a systematic review and meta-analysis. *Fertil Steril.* 2011;95:1048-58.e1-2.
- Faghfoori Z, Fazelian S, Shadnough M, Goodarzi R. Nutritional management in women with polycystic ovary syndrome: A review study. *Diabetes Metab Syndr.* 2017;11(Suppl 1):S429-32.
- Farimani AR, Hariri M, Azimi-Nezhad M, Borji A, Zarei S, Hooshmand E. The effect of n-3 PUFAs on circulating adiponectin and leptin in patients with type 2 diabetes mellitus: a systematic review and meta-analysis of randomized controlled trials. *Acta Diabetol.* 2018;55:641-52.
- Fernandez-Real JM, Ricart W. Insulin resistance and chronic cardiovascular inflammatory syndrome. *Endocr Rev.* 2003;24:278-301.
- Forman BM, Chen J, Evans RM. Hypolipidemic drugs, polyunsaturated fatty acids, and eicosanoids are ligands for peroxisome proliferator-activated receptors alpha and delta. *Proc Natl Acad Sci U S A.* 1997;94:4312-7.
- Geetha V, Chakravarthula SN. Chemical composition and anti-inflammatory activity of *Boswellia ovalifoliolata* essential oils from leaf and bark. *J Forestry Res.* 2018;29:373-81.
- Ghasemifard S, Hermon K, Turchini GM, Sinclair AJ. Metabolic fate (absorption, beta-oxidation and deposition) of long-chain n-3 fatty acids is affected by sex and by the oil source (krill oil or fish oil) in the rat. *Br J Nutr.* 2015;114:684-92.
- González F, Rote N, Minium J, Kirwan J. Hyperglycemia stimulates proatherogenic inflammation pathways in polycystic ovary syndrome. *Fertil Steril.* 2006a;86:S450.
- González F, Rote NS, Minium J, Kirwan JP. Increased activation of nuclear factor kappaB triggers inflammation and insulin resistance in polycystic ovary syndrome. *J Clin Endocrinol Metab.* 2006b;91:1508-12.
- Goodarzi MO, Dumesic DA, Chazenbalk G, Azziz R. Polycystic ovary syndrome: etiology, pathogenesis and diagnosis. *Nat Rev Endocrinol.* 2011;7:219-31.
- Goodfellow J, Bellamy MF, Ramsey MW, Jones CJ, Lewis MJ. Dietary supplementation with marine omega-3 fatty acids improve systemic large artery endothelial function in subjects with hypercholesterolemia. *J Am Coll Cardiol.* 2000;35:265-70.
- Gray B, Steyn F, Davies PS, Vitetta L. Omega-3 fatty acids: a review of the effects on adiponectin and leptin and potential implications for obesity management. *Eur J Clin Nutr.* 2013;67:1234-42.
- Gunalan E, Yaba A, Yilmaz B. The effect of nutrient supplementation in the management of polycystic ovary syndrome-associated metabolic dysfunctions: A critical review. *J Turk Ger Gynecol Assoc.* 2018;19:220-32.
- Hanada H, Morikawa K, Hirota K, Nonaka M, Ume-hara Y. Induction of apoptosis and lipogenesis in human preadipocyte cell line by n-3 PUFAs. *Cell Biol Int.* 2011;35:51-9.
- Hariri M, Ghiasvand R, Shiranian A, Askari G, Iraj B, Salehi-Abargouei A. Does omega-3 fatty acids supplementation affect circulating leptin levels? A systematic review and meta-analysis on randomized controlled clinical trials. *Clin Endocrinol (Oxf).* 2015;82:221-8.
- Harris WS, Rambjor GS, Windsor SL, Diederich D. n-3 fatty acids and urinary excretion of nitric oxide metabolites in humans. *Am J Clin Nutr.* 1997;65:459-64.
- Harris WS, Miller M, Tighe AP, Davidson MH, Schaefer EJ. Omega-3 fatty acids and coronary heart disease risk: clinical and mechanistic perspectives. *Atherosclerosis.* 2008;197:12-24.
- Home PD, Pacini G. Hepatic dysfunction and insulin insensitivity in type 2 diabetes mellitus: a critical target for insulin-sensitizing agents. *Diabetes Obes Metab.* 2008;10:699-718.
- Hoshi T, Wissuwa B, Tian Y, Tajima N, Xu R, Bauer M, et al. Omega-3 fatty acids lower blood pressure by directly activating large-conductance Ca(2)(+)-dependent K(+) channels. *Proc Natl Acad Sci U S A.* 2013;110:4816-21.

- Hu WH, Qiao J, Zhao SY, Zhang XW, Li MZ. [Monocyte chemoattractant protein-1 and its correlation with lipoprotein in polycystic ovary syndrome]. *Beijing Da Xue Xue Bao Yi Xue Ban*. 2006;38:487-91.
- Jafarnejad S, Sadegh M. The effects of different levels of dietary protein, energy and using fat on the performance of broiler chicks at the end of the third. *Asian J Poult Sci*. 2011;5:35-40.
- Jamilian M, Samimi M, Mirhosseini N, Afshar Ebrahimi F, Aghadavod E, Talaee R, et al. The influences of vitamin D and omega-3 co-supplementation on clinical, metabolic and genetic parameters in women with polycystic ovary syndrome. *J Affect Disord*. 2018;238:32-8.
- Jonard S, Dewailly D. The follicular excess in polycystic ovaries, due to intra-ovarian hyperandrogenism, may be the main culprit for the follicular arrest. *Hum Reprod Update*. 2004;10:107-17.
- Kahn CR. Banting lecture. Insulin action, diabetogenes, and the cause of type II diabetes. *Diabetes*. 1994;43:1066-84.
- Kalgaonkar S, Almario RU, Gurusinghe D, Garamendi EM, Buchan W, Kim K, et al. Differential effects of walnuts vs almonds on improving metabolic and endocrine parameters in PCOS. *Eur J Clin Nutr*. 2011;65:386-93.
- Kalupahana NS, Moustaid-Moussa N, Claycombe KJ. Immunity as a link between obesity and insulin resistance. *Mol Aspects Med*. 2012;33:26-34.
- Kang YE, Kim JM, Joung KH, Lee JH, You BR, Choi MJ, et al. The roles of adipokines, proinflammatory cytokines, and adipose tissue macrophages in obesity-associated insulin resistance in modest obesity and early metabolic dysfunction. *PLoS One*. 2016;11(4):e0154003.
- Kannel WB, Dawber TR, Friedman GD, Glennon WE, McNamara PM. Risk factors in coronary heart disease. an evaluation of several serum lipids as predictors of coronary heart disease; the framingham study. *Ann Intern Med*. 1964;61:888-99.
- Karlsson M, Marild S, Brandberg J, Lonn L, Friberg P, Strandvik B. Serum phospholipid fatty acids, adipose tissue, and metabolic markers in obese adolescents. *Obesity (Silver Spring)*. 2006;14:1931-9.
- Khani B, Mardanian F, Fesharaki SJ. Omega-3 supplementation effects on polycystic ovary syndrome symptoms and metabolic syndrome. *J Res Med Sci*. 2017;22:64.
- Kim HK, Della-Fera M, Lin J, Baile CA. Docosahexaenoic acid inhibits adipocyte differentiation and induces apoptosis in 3T3-L1 preadipocytes. *J Nutr*. 2006;136:2965-9.
- Kim S, Moustaid-Moussa N. Secretory, endocrine and autocrine/paracrine function of the adipocyte. *J Nutr*. 2000;130:3110s-5.
- Kong W, Yen JH, Vassiliou E, Adhikary S, Toscano MG, Ganea D. Docosahexaenoic acid prevents dendritic cell maturation and in vitro and in vivo expression of the IL-12 cytokine family. *Lipids Health Dis*. 2010;9:12.
- Krey G, Braissant O, L'Horset F, Kalkhoven E, Perroud M, Parker MG, et al. Fatty acids, eicosanoids, and hypolipidemic agents identified as ligands of peroxisome proliferator-activated receptors by coactivator-dependent receptor ligand assay. *Mol Endocrinol*. 1997;11:779-91.
- Lark DS, Fisher-Wellman KH, Neuffer PD. High-fat load: mechanism(s) of insulin resistance in skeletal muscle. *Int J Obes Suppl*. 2012;2(Suppl 2):S31-6.
- Lefils-Lacourtablaise J, Socorro M, Geloën A, Daira P, Debard C, Loizon E, et al. The eicosapentaenoic acid metabolite 15-deoxy-delta(12,14)-prostaglandin J3 increases adiponectin secretion by adipocytes partly via a PPARgamma-dependent mechanism. *PLoS One*. 2013;8(5):e63997.
- Lepretti M, Martucciello S, Burgos Aceves MA, Putti R, Lionetti L. Omega-3 fatty acids and insulin resistance: focus on the regulation of mitochondria and endoplasmic reticulum stress. *Nutrients*. 2018;10(3):E350.
- Lewandowski KC, Komorowski J, O'Callaghan CJ, Tan BK, Chen J, Prelevic GM, et al. Increased circulating levels of matrix metalloproteinase-2 and -9 in women with the polycystic ovary syndrome. *J Clin Endocrinol Metab*. 2006;91:1173-7.
- Li S, Wen W, Gong X, Huang X, Chen N. Variation of lipids and fatty acids composition in the tissues of wild devil stinger (*Inimicus japonicas*) during sexual maturation. *Aquacult Fish*. 2018;3:115-21.
- Liepa GU, Sengupta A, Karsies D. Polycystic ovary syndrome (PCOS) and other androgen excess-related conditions: can changes in dietary intake make a difference? *Nutr Clin Pract*. 2008;23:63-71.
- Mahdy Ali K, Wonnerth A, Huber K, Wojta J. Cardiovascular disease risk reduction by raising HDL cholesterol - current therapies and future opportunities. *Br J Pharmacol*. 2012;167:1177-94.



- Martinez-Fernandez L, Laiglesia LM, Huerta AE, Martinez JA, Moreno-Aliaga MJ. Omega-3 fatty acids and adipose tissue function in obesity and metabolic syndrome. *Prostaglandins Other Lipid Mediat.* 2015; 121(Pt A):24-41.
- McEwen BJ. Can omega-3 polyunsaturated fatty acids improve metabolic profile in polycystic ovary syndrome (PCOS)? *Adv Integr Med.* 2017;4(2):82-3.
- Mejia-Montilla J, Reyna-Villasmil E, Dominguez-Brito L, Naranjo-Rodriguez C, Noriega-Verdugo D, Padilla-Samaniego M, et al. Supplementation with omega-3 fatty acids and plasma adiponectin in women with polycystic ovary syndrome. *Endocrinol Diabetes Nutr.* 2018;65:192-9.
- Micallef M, Munro I, Phang M, Garg M. Plasma n-3 polyunsaturated fatty acids are negatively associated with obesity. *Br J Nutr.* 2009;102:1370-4.
- Miller PE, Van Elswyk M, Alexander DD. Long-chain omega-3 fatty acids eicosapentaenoic acid and docosahexaenoic acid and blood pressure: a meta-analysis of randomized controlled trials. *Am J Hypertens.* 2014;27:885-96.
- Mirmasoumi G, Fazilati M, Foroozafard F, Vahedpoor Z, Mahmoodi S, Taghizadeh M, et al. The effects of flaxseed oil omega-3 fatty acids supplementation on metabolic status of patients with polycystic ovary syndrome: a randomized, double-blind, placebo-controlled trial. *Exp Clin Endocrinol Diabetes.* 2018;126: 222-8.
- Mohammadi E, Rafrat M, Farzadi L, Asghari-Jafarabadi M, Sabour S. Effects of omega-3 fatty acids supplementation on serum adiponectin levels and some metabolic risk factors in women with polycystic ovary syndrome. *Asia Pac J Clin Nutr.* 2012;21:511-8.
- Moreno-Aliaga MJ, Lorente-Cebrian S, Martinez JA. Regulation of adipokine secretion by n-3 fatty acids. *Proc Nutr Soc.* 2010;69:324-32.
- Murali G, Desouza CV, Clevenger ME, Ramalingam R, Saraswathi V. Differential effects of eicosapentaenoic acid and docosahexaenoic acid in promoting the differentiation of 3T3-L1 preadipocytes. *Prostaglandins Leukot Essent Fatty Acids.* 2014;90:13-21.
- Nadjarzadeh A, Dehghani-Firouzabadi R, Daneshbodi H, Lotfi MH, Vaziri N, Mozaffari-Khosravi H. Effect of omega-3 supplementation on visfatin, adiponectin, and anthropometric indices in women with polycystic ovarian syndrome. *J Reprod Infertil.* 2015;16:212-20.
- Nardo LG, Patchava S, Laing I. Polycystic ovary syndrome: pathophysiology, molecular aspects and clinical implications. *Panminerva Med.* 2008;50:267-78.
- Nomura S, Shouzu A, Omoto S, Inami N, Ueba T, Urase F, et al. Effects of eicosapentaenoic acid on endothelial cell-derived microparticles, angiopoietins and adiponectin in patients with type 2 diabetes. *J Atheroscler Thromb.* 2009;16(2):83-90.
- Nomura S, Taniura T, Shouzu A, Omoto S, Suzuki M, Okuda Y, et al. Effects of sarpogrelate, eicosapentaenoic acid and pitavastatin on arteriosclerosis obliterans-related biomarkers in patients with type 2 diabetes (SAREPITASO study). *Vasc Health Risk Manag.* 2018;14:225-32.
- O'Connor A, Gibney J, Roche HM. Metabolic and hormonal aspects of polycystic ovary syndrome: the impact of diet. *Proc Nutr Soc.* 2010;69:628-35.
- O'Meara NM, Blackman JD, Ehrmann DA, Barnes RB, Jaspan JB, Rosenfield RL, et al. Defects in beta-cell function in functional ovarian hyperandrogenism. *J Clin Endocrinol Metab.* 1993;76:1241-7.
- Oner G, Muderris II. Efficacy of omega-3 in the treatment of polycystic ovary syndrome. *J Obstet Gynaecol.* 2013;33:289-91.
- Ouchi N, Parker JL, Lugus JJ, Walsh K. Adipokines in inflammation and metabolic disease. *Nat Rev Immunol.* 2011;11(2):85-97.
- Park Y, Harris WS. Omega-3 fatty acid supplementation accelerates chylomicron triglyceride clearance. *J Lipid Res.* 2003;44:455-63.
- Perez-Matute P, Perez-Echarri N, Martinez JA, Marti A, Moreno-Aliaga MJ. Eicosapentaenoic acid actions on adiposity and insulin resistance in control and high-fat-fed rats: role of apoptosis, adiponectin and tumour necrosis factor-alpha. *Br J Nutr.* 2007;97:389-98.
- Phelan N, O'Connor A, Kyaw Tun T, Correia N, Boran G, Roche H M, et al. Hormonal and metabolic effects of polyunsaturated fatty acids in young women with polycystic ovary syndrome: results from a cross-sectional analysis and a randomized, placebo-controlled, crossover trial. *Am J Clin Nutr.* 2011;93: 652-62.
- Pizzini A, Lunger L, Demetz E, Hilbe R, Weiss G, Ebenbichler C, et al. The role of omega-3 fatty acids in reverse cholesterol transport: A review. *Nutrients.* 2017;9(10):E1099.
- Pourghassem Gargari B, Houjehani S, Farzadi L, Houjehani S, Safaeiyan A. Relationship between serum leptin, ghrelin and dietary macronutrients in women with polycystic ovary syndrome. *Int J Fertil Steril.* 2015;9:313-21.

- Raclot T, Groscolas R, Langin D, Ferre P. Site-specific regulation of gene expression by n-3 polyunsaturated fatty acids in rat white adipose tissues. *J Lipid Res.* 1997;38:1963-72.
- Rader DJ. Regulation of reverse cholesterol transport and clinical implications. *Am J Cardiol.* 2003;92(4a):42j-9.
- Rafraf M, Mohammadi E, Asghari-Jafarabadi M, Farzadi L. Omega-3 fatty acids improve glucose metabolism without effects on obesity values and serum visfatin levels in women with polycystic ovary syndrome. *J Am Coll Nutr.* 2012;31:361-8.
- Rahmani E, Samimi M, Ebrahimi FA, Foroozanfard F, Ahmadi S, Rahimi M, et al. The effects of omega-3 fatty acids and vitamin E co-supplementation on gene expression of lipoprotein(a) and oxidized low-density lipoprotein, lipid profiles and biomarkers of oxidative stress in patients with polycystic ovary syndrome. *Mol Cell Endocrinol.* 2017;439:247-55.
- Ratnayake WM, Galli C. Fat and fatty acid terminology, methods of analysis and fat digestion and metabolism: a background review paper. *Ann Nutr Metab.* 2009;55(1-3):8-43.
- Ren B, Thelen AP, Peters JM, Gonzalez FJ, Jump DB. Polyunsaturated fatty acid suppression of hepatic fatty acid synthase and S14 gene expression does not require peroxisome proliferator-activated receptor alpha. *J Biol Chem.* 1997;272:26827-32.
- Renier G, Skamene E, DeSanctis J, Radzioch D. Dietary n-3 polyunsaturated fatty acids prevent the development of atherosclerotic lesions in mice. Modulation of macrophage secretory activities. *Arterioscler Thromb.* 1993;13:1515-24.
- Robinson JG, Stone NJ. Antiatherosclerotic and antithrombotic effects of omega-3 fatty acids. *Am J Cardiol.* 2006;98(4a):39i-49.
- Rondanelli M, Perna S, Faliva M, Monteferrario F, Repaci E, Allieri F. Focus on metabolic and nutritional correlates of polycystic ovary syndrome and update on nutritional management of these critical phenomena. *Arch Gynecol Obstet.* 2014;290:1079-92.
- Rotterdam ESHRE/ASRM-sponsored PCOS consensus workshop group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome (PCOS). *Hum Reprod.* 2004;19(1):41-7.
- Sadeghi A, Djafarian K, Mohammadi H, Shab-Bidar S. Effect of omega-3 fatty acids supplementation on insulin resistance in women with polycystic ovary syndrome: Meta-analysis of randomized controlled trials. *Diabetes Metab Syndr.* 2017;11:157-62.
- Serhan CN, Arita M, Hong S, Gotlinger K. Resolvins, docosatrienes, and neuroprotectins, novel omega-3-derived mediators, and their endogenous aspirin-triggered epimers. *Lipids.* 2004;39:1125-32.
- Siasos G, Tousoulis D, Oikonomou E, Zaromitidou M, Verveniotis A, Plastiras A, et al. Effects of omega-3 fatty acids on endothelial function, arterial wall properties, inflammatory and fibrinolytic status in smokers: a cross over study. *Int J Cardiol.* 2013;166:340-6.
- Siddiqi HK, Kiss D, Rader D. HDL-cholesterol and cardiovascular disease: rethinking our approach. *Curr Opin Cardiol.* 2015;30:536-42.
- Solomon CG. The epidemiology of polycystic ovary syndrome. Prevalence and associated disease risks. *Endocrinol Metab Clin North Am.* 1999;28:247-63.
- Stanford KI, Middelbeek RJ, Townsend KL, An D, Nygaard EB, Hitchcox KM, et al. Brown adipose tissue regulates glucose homeostasis and insulin sensitivity. *J Clin Invest.* 2013;123:215-23.
- Stein JD, Andrews C, Musch DC, Green C, Lee PP. Sight-Threatening ocular diseases remain underdiagnosed among children of less affluent families. *Health Aff (Millwood).* 2016;35:1359-66.
- Talari HR, Poladchang S, Hamidian Y, Samimi M, Gilasi HR, Ebrahimi FA, et al. The effects of omega-3 and vitamin e co-supplementation on carotid intima-media thickness and inflammatory factors in patients with polycystic ovary syndrome. *Oman Med J.* 2018;33:473-9.
- Thorlaksdottir AY, Skuladottir GV, Petursdottir AL, Tryggvadottir L, Ogmundsdottir HM, Eyfjord JE, et al. Positive association between plasma antioxidant capacity and n-3 PUFA in red blood cells from women. *Lipids.* 2006;41:119-25.
- Tomas E, Tsao TS, Saha AK, Murrey HE, Zhang CcCc, Itani SI, et al. Enhanced muscle fat oxidation and glucose transport by ACRP30 globular domain: acetyl-CoA carboxylase inhibition and AMP-activated protein kinase activation. *Proc Natl Acad Sci U S A.* 2002;99:16309-13.
- Ulu A, Stephen Lee KS, Miyabe C, Yang J, Hammock BG, Dong H, et al. An omega-3 epoxide of docosahexaenoic acid lowers blood pressure in angiotensin-II-dependent hypertension. *J Cardiovasc Pharmacol.* 2014;64:87-99.
- von Frankenberg AD, Silva FM, de Almeida JC, Piccoli V, do Nascimento FV, Sost MM, et al. Effect of dietary lipids on circulating adiponectin: a systematic review with meta-analysis of randomised controlled trials. *Br J Nutr.* 2014;112:1235-50.

- Wenner MM, Taylor HS, Stachenfeld NS. Endothelin B receptor contribution to peripheral microvascular function in women with polycystic ovary syndrome. *J Physiol.* 2011;589:4671-9.
- Westerveld HE, Hoogendoorn M, de Jong AW, Goverde AJ, Fauser BC, Dallinga-Thie GM. Cardiometabolic abnormalities in the polycystic ovary syndrome: pharmacotherapeutic insights. *Pharmacol Ther.* 2008;119:223-41.
- Wiest EF, Walsh-Wilcox MT, Walker MK. Omega-3 polyunsaturated fatty acids protect against cigarette smoke-induced oxidative stress and vascular dysfunction. *Toxicol Sci.* 2017;156:300-10.
- Xita N, Tsatsoulis A. Review: fetal programming of polycystic ovary syndrome by androgen excess: evidence from experimental, clinical, and genetic association studies. *J Clin Endocrinol Metab.* 2006;91:1660-6.
- Yamauchi T, Kamon J, Ito Y, Tsuchida A, Yokomizo T, Kita S, et al. Cloning of adiponectin receptors that mediate antidiabetic metabolic effects. *Nature.* 2003;423(6941):762-9.
- Zapata-Gonzalez F, Rueda F, Petriz J, Domingo P, Villarroya F, Diaz-Delfin J, et al. Human dendritic cell activities are modulated by the omega-3 fatty acid, docosahexaenoic acid, mainly through PPAR (gamma): RXR heterodimers: comparison with other polyunsaturated fatty acids. *J Leukoc Biol.* 2008;84:1172-82.
- Zehr KR, Walker MK. Omega-3 polyunsaturated fatty acids improve endothelial function in humans at risk for atherosclerosis: A review. *Prostaglandins Other Lipid Mediat.* 2018;134:131-40.
- Zivkovic AM, German JB, Sanyal AJ. Comparative review of diets for the metabolic syndrome: implications for nonalcoholic fatty liver disease. *Am J Clin Nutr.* 2007;86:285-300.
- Zuliani G, Galvani M, Leitersdorf E, Volpato S, Cavalieri M, Fellin R. The role of polyunsaturated fatty acids (PUFA) in the treatment of dyslipidemias. *Curr Pharm Des.* 2009;15:4087-93.