

Received: 28 January 2019


Revised: 25 June 2019

Accepted: 4 July 2019

DOI: 10.1111/ddi.12972

BIODIVERSITY RESEARCH**Diversity and Distributions** WILEY

The biodiversity benefit of native forests and mixed-species plantations over monoculture plantations

Xiaoyang Wang^{1,2}  | Fangyuan Hua^{3,4} | Lin Wang¹ | David S. Wilcove^{5,6} | Douglas W. Yu^{1,7,8}

¹State Key Laboratory of Genetic Resources and Evolution, Kunming Institute of Zoology, Chinese Academy of Sciences, Kunming, China

²Kunming College of Life Sciences, University of Chinese Academy of Sciences, Kunming, China

³Conservation Science Group, Department of Zoology, University of Cambridge, Cambridge, UK

⁴Key Laboratory for Plant Diversity and Biogeography of East Asia, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming, China

⁵Department of Ecology and Evolutionary Biology, Princeton University, Princeton, NJ, USA

⁶Program in Science, Technology and Environmental Policy, School of Public and International Affairs, Princeton University, Princeton, NJ, USA

⁷Center for Excellence in Animal Evolution and Genetics, Chinese Academy of Sciences, Kunming, China

⁸School of Biological Sciences, University of East Anglia, Norwich Research Park, Norwich, UK

Correspondence

Douglas W. Yu, State Key Laboratory of Genetic Resources and Evolution, Kunming Institute of Zoology, Chinese Academy of Sciences, Kunming, Yunnan 650223, China. Email: dougwyu@mac.com

Funding information

National Natural Science Foundation of China, Grant/Award Number: 41661144002, 31670536, 31400470, 31500305 and 31872963; Key Research Program of Frontier Sciences, Grant/Award Number: QYZDY-SSW-SMC024; Bureau of International Cooperation, Grant/Award Number: GJHZ1754; Strategic Priority Research Program of the Chinese Academy of Sciences, Grant/Award Number: XDA20050202 and XDB31000000; Ministry of Science and Technology of China, Grant/Award Number: 2012FY110800; University of East Anglia; State Key Laboratory of Genetic Resources and Evolution at the Kunming Institute of Zoology; High Meadows Foundation; Princeton University; Informatization Program of the Chinese Academy of Sciences, Grant/Award Number: XXH13505-03-102-2

Editor: Chris Burridge

Abstract

Aim: China's Grain for Green Program (GFGP) is the largest reforestation programme in the world and has been operating since 1999. The GFGP has promoted the establishment of tree plantations over the restoration of diverse native forests. In a previous study, we showed that native forests support a higher species richness and abundance of birds and bees than do GFGP plantations and that mixed-species GFGP plantations support a higher level of bird (but not bee) diversity than do any individual GFGP monocultures (although still below that of native forests). Here, we use metabarcoding of arthropod diversity to test the generality of these results.

Location: Sichuan, China.

Methods: We sampled arthropod communities using pan traps in the land cover types concerned under the GFGP. These land use types include croplands (the land cover being reforested under the GFGP), native forests (the reference ecosystem as the benchmark for the GFGP's biodiversity effects) and the dominant GFGP reforestation outcomes: monoculture and mixed-species plantations. We used COI-amplicon sequencing ("metabarcoding") of the arthropod samples to quantify and assess the arthropod community profiles associated with each land cover type.

Results: Native forests support the highest overall levels of arthropod species diversity, followed by mixed-species plantations, followed by bamboo and other monocultures. Also, the arthropod community in native forests shares more species with mixed-species plantations than it does with any of the monocultures. Together, these results broadly corroborate our previous conclusions on birds and bees but show

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2019 The Authors. *Diversity and Distributions* published by John Wiley & Sons Ltd.

a higher arthropod biodiversity value of mixed-species plantations than previously indicated by bees alone.

Main conclusion: In our previous study, we recommended that GFGP should prioritize the conservation and restoration of native forests. Also, where plantations are to be used, we recommended that the GFGP should promote mixed-species arrangements over monocultures. Both these recommendations should result in more effective protection of terrestrial biodiversity, which is an important objective of China's land-sustainability spending. The results of this study strengthen these recommendations because our policy prescriptions are now also based on a dataset that includes over 500 species-resolution taxa, ranging across the Arthropoda.

KEYWORDS

Arthropoda, biodiversity, China, forest management, Grain for Green Program, metabarcoding, reforestation

1 | INTRODUCTION

An important challenge for conservation science is to quantify the biodiversity impacts of major policy initiatives, especially in regions undergoing large shifts in land use change. Nowhere is this more true than in China, which combines a high level of native biodiversity (Tao, Huang, Jin, & Guo, 2010) with a large human population that is increasing its ecological footprint (Liu & Diamond, 2005; Pyne, 2013; Sayer & Sun, 2003; Xie et al., 2012). Moreover, for decades, China has had the managerial, political and financial capacity to implement the largest land-sustainability programmes ever seen, from nature-reserve protection to reforestation to de-desertification (Bryan et al., 2018; Liu et al., 2003; Wu et al., 2019; Xu, Wang, & Xue, 1999). These programmes have caused major land use changes and successfully slowed land degradation caused by economic activities (Liu et al., 2008; Ouyang et al., 2016; Ren et al., 2015). For example, China established its first nature reserve in 1956 and reached 2,740 reserves at the end of 2015 (Ma, Shen, Grumbine, & Corlett, 2017). Nearly two-thirds of the area of those nature reserves have national-level status, meaning that they receive the highest level of protection and funding, and analysis of Landsat imagery has shown that national-level reserves successfully deter deforestation (Ren et al., 2015).

Two other major land-sustainability programmes are the Natural Forest Protection Program (NFPP, also known as Natural Forest Conservation Program) and the Grain for Green Program (GFGP, also known as the Sloping Land Conservation Program and the Farm to Forest Program), which were implemented after widespread flooding in 1998 (Liu et al., 2008; Xu, Yin, Li, & Liu, 2006; Yin, Yin, & Li, 2009). The NFPP aims to reduce soil erosion and flooding by protecting native forests in the upstream watersheds of the Yangtze and Yellow rivers (Liu et al., 2008; Ren et al., 2015). The GFGP complements the NFPP by controlling soil erosion on sloping land. The government pays cash and grain to farmers in exchange for tree planting on sloping farmland (Delang & Yuan, 2015; Liu et al., 2008;

Ma et al., 2017; Xu et al., 2006; Zhai, Xu, Dai, Cannon, & Grumbine, 2014). Having reforested 9.06 million ha of cropland over 16 years (~2014) since its inception in 1999, the GFGP is the world's largest reforestation programme.

However, relative to their scale and budgets, little is known about the biodiversity consequences of China's land-sustainability programmes, even though an important and expected co-benefit is biodiversity conservation (Wu et al., 2019). In a recent, massive review, Bryan et al. (2018) were able to cite only one study on the consequences of China's large-scale reforestation programmes for biodiversity, Hua et al. (2016). This paucity of understanding contrasts starkly with the large volume of information on other consequences of these programmes: water and soil maintenance (Deng, Shangguan, & Li, 2012; Long et al., 2006; Wang, Peng, Zhao, Liu, & Chen, 2017; Wang, Jiao, Rayburg, Wang, & Su, 2016), carbon storage (Deng, Liu, & Shangguan, 2014; Wei et al., 2014), vegetation cover (Hua et al., 2018; Zhai et al., 2014; Zhou, Rompaey, & Wang, 2009) and socio-economic outcomes (Liu & Lan, 2015; Yin, Liu, Zhao, Yao, & Liu, 2014; Yin et al., 2009). A better understanding of the biodiversity implications of reforestation programmes is needed to guide these programmes for China and the rest of the world (Turner, Lambin, & Reenberg, 2007; United Nations, 2015).

Guided by the goal of soil erosion control, and operating under the implicit assumption that any type of tree cover should achieve this goal, the GFGP has predominantly established tree plantations ("plantations" hereafter) on retired croplands, rather than restoring native forests (Hua et al., 2018, 2016; Zhai et al., 2014). However, compared with native-forest ecosystems, plantations are known to support lower levels of biodiversity across the world's forest biomes and across taxa (Barlow, Overal, Araujo, Gardner, & Peres, 2007; Bremer & Farley, 2010; Brockerhoff, Jactel, Parrotta, Quine, & Sayer, 2008; Gardner, Hernandez, Barlow, & Peres, 2008; Lindenmayer & Hobbs, 2004), although certain management regimes, such as maintaining understorey structure and mixed cropping, can somewhat increase biodiversity (Hartley, 2002). On the other hand, compared

with croplands, plantations are known to support different species assemblages, with potentially higher levels of biodiversity, although there are indications that croplands in low-intensity agricultural systems—which the croplands retired under GFGP tend to be (Hu, Fu, Chen, & Gulinck, 2006)—may support considerable biodiversity which potentially exceeds that associated with plantations (Allan, Harrison, Navarro, Wilgen, & Thompson, 1997; Buscardo et al., 2008; Elsen, Ramesh, & Wilcove, 2018). Together, these insights suggest that plantations should have been expected to support low levels of biodiversity and that the GFGP could support more biodiversity if it restored native forests.

Indeed, this is what Hua et al. (2016) found. They surveyed bird and bee communities in GFGP-related tree covers in south-central Sichuan, comparing native-forest remnants to GFGP-financed tree-cover types, which include monoculture stands of bamboo, Eucalyptus and Japanese cedar, as well as “mixed plantations”, which are mostly patchworks (checkerboards) of two to five different monocultures and, to a lesser extent, *bona fide* tree-level mixtures (Hua et al., 2018). Most importantly, this study documented that bird and bee species diversities were higher in native forests than in any of the monocultures. In addition, they found that in mixed plantations, bird diversity for non-breeding species was higher than in any of the individual monocultures, albeit lower than in native forests. In contrast, bee diversity was equally low in mixed plantations and monocultures. The lack of a boost to bee diversity in mixed plantations was not surprising, because as with monocultures, the understorey vegetation in mixed plantations was notably lacking in flowering plants (Hua et al., 2016).

The above findings, however, raise the question of why bird diversity was increased just by planting monocultures of different tree species next to each other. One possibility that could not be investigated in Hua et al. (2016) is that general arthropod diversity might also have been boosted in the mixed plantations, as, unlike bees, other arthropods can exploit a range of food resources available even in plantations, via direct consumption of plants and fungi, and via decomposition, parasitism and predation of other animals, including other arthropods (Jactel & Brockerhoff, 2007). Increased arthropod diversity might in turn support more bird diversity. In addition, as a large component of biodiversity, how arthropods themselves (and subgroups thereof) are affected by the GFGP is an important part of understanding the GFGP's biodiversity effects. For instance, Barlow, Gardner, et al. (2007) compared primary forest and Eucalyptus plantations in Brazil and found that birds achieve highest diversity in primary forest, while bees have similar levels of species richness in primary forest and Eucalyptus plantations. They also found that butterflies and dung beetles achieve low diversity but that fruit flies and moths achieve high diversity in Eucalyptus plantations.

The purpose of this study is to test the generality of Hua et al.'s (2016) results by interrogating the “rest of the biodiversity” that was captured in the same sites analysed by Hua et al. (2016). We employ the technique of metabarcoding, which combines traditional DNA barcoding with high-throughput DNA sequencing to characterize

the biodiversity of mixed samples of eukaryotes (Cristescu, 2014; Deiner et al., 2017; Yu et al., 2012), and which has been shown to be a reliable and efficient method for biodiversity characterization (Ji et al., 2013). Through metabarcoding the non-bee arthropods caught in the same pan traps previously used to trap bees in Hua et al. (2016), we hope to answer the following questions: (a) Do native forests support higher levels of arthropod species richness and diversity than all four GFGP plantations? (b) For all GFGP plantations, do mixed plantations support higher levels of arthropod species richness and diversity than do the three individual monocultures? (c) How does community composition compare among these tree covers and what underlies the potential differences?

2 | METHODS

2.1 | Study location

The study region and locations are as in Hua et al. (2016). In short, our study region was a 7,949-km² area in south-central Sichuan Province (Figure 1) spanning 315–1,715 m above sea level, historically forested and then deforested starting in the 1950s. The GFGP established ~54,800 ha of new tree cover between 1999 and 2014, dominated by short-rotation (6–20 years) monocultures of bamboo (BB), Eucalyptus (EC) and Japanese cedar (JC), and short-rotation mixed plantations (MP) of two to five tree species (including the three monoculture species). Monocultures are created by households planting the same tree species in neighbouring landholdings. Correspondingly, mixed plantations are, in most cases, created by planting different species, resulting in a checkerboard, although about a quarter of mixed plantations consist of tree-level mixtures. In Hua et al. (2016), we used the term “mixed forests”, but in Hua et al. (2018), we switched to “mixed plantations”.

The two other surveyed land covers were croplands (CL) and native forests (NF). Croplands mostly consist of low-intensity plantings of rice, corn and vegetables and are the land cover type that has been reforested by GFGP. Native forests are broadleaf, subtropical, evergreen forests that have been subject to decades of selective logging and other forms of extraction. Because this region of China has been inhabited for millennia, there are no undisturbed native forests. Croplands are typically located on flatter land than are the tree covers, as GFGP reforestation targeted sloped land, and the native forests are concentrated towards the more hilly, southern end of the study region. For sampling, we chose larger expanses (>60 ha) of these six land cover types: BB, EC, JC, MP, NF and CL.

2.2 | Sampling design

Each land cover type was represented by at least two locations set ≥ 15 km apart. All tree-cover stands sampled had closed canopy. For each land cover type, we sampled with at least 10 one-ha quadrats, within each of which we operated 40 fluorescent pan traps for 24 hr (Bartholomew & Prowell, 2005; Figure S1). In total, we sampled 74 quadrats (BB: 10, EC: 10, JC: 12, MP: 10, NF: 16, CL: 16). Different

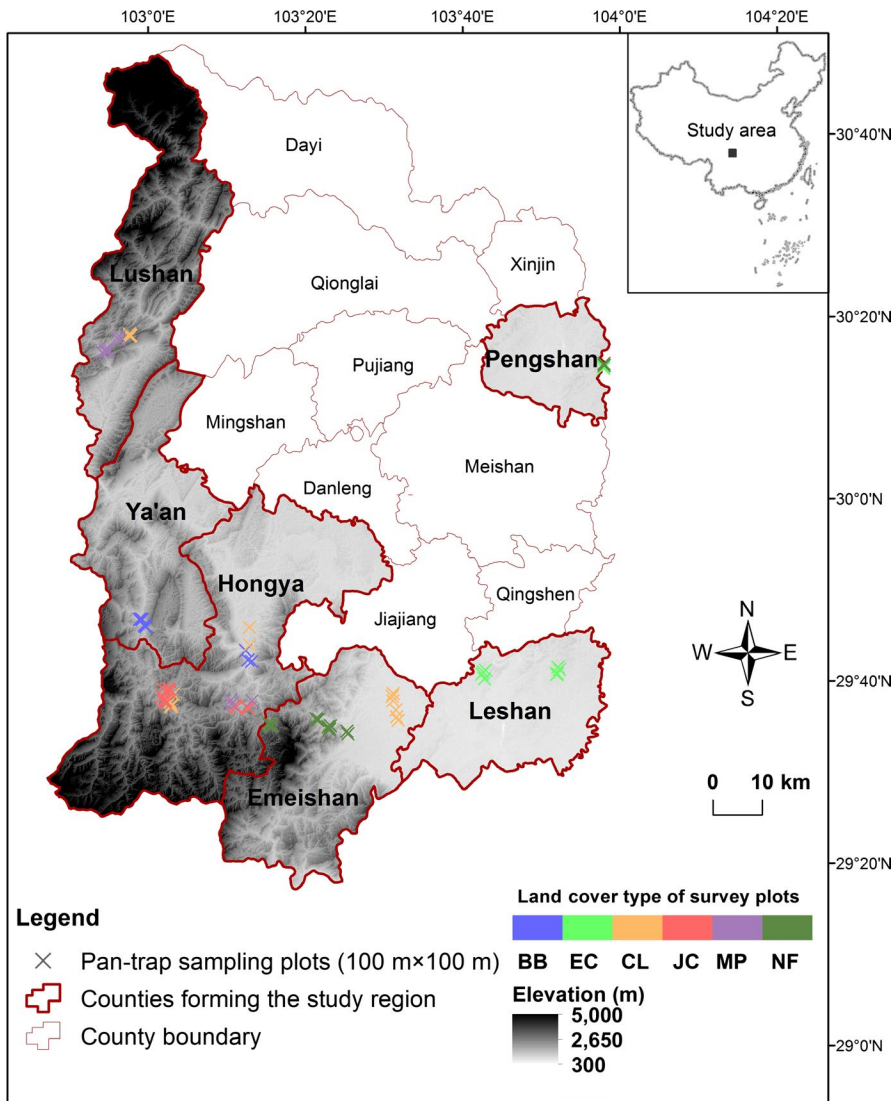


FIGURE 1 Study area in south-central Sichuan Province, subdivided into counties and shaded by elevation. Each cross represents a pan-trap sampling location, colour-coded by land cover type: BB = bamboo monoculture, blue; EC = Eucalyptus monoculture, light green; CL = croplands, orange; JC = Japanese cedar monoculture, red; MP = mixed plantations, purple; NF = native forests, dark green

quadrats were separated by ≥ 300 m if placed in the same tree-cover stand. Samples were stored in 100% ethanol at ambient temperature until shipment to the laboratory, where they were stored at -20°C before DNA extraction. The original reason for using pan traps had been to trap bees, which we individually DNA-barcoded in Hua et al. (2016). Here, we analyse the bycatch.

2.3 | Amplicon preparation

For each of the 74 quadrats, we pooled all 40 pan traps into a single sample. Three quadrats had very few individuals, and we pooled them with their nearest neighbour of the same land cover type (EC01 + EC02 + EC03; NF02 + NF03), leaving us with 71 samples. Storage ethanol was removed by air-drying on single-use filter papers. Our samples were dominated by Diptera and Hymenoptera, as expected. We equalized input DNA across species by using one leg of every individual larger than a mosquito (~ 5 mm long) and the whole body if smaller (e.g. midges). This was to reduce the effect of large-biomass individuals out-competing small-biomass individuals during

PCR, which improves taxon detection (Elbrecht, Peinert, & Leese, 2017). DNA extraction followed the protocols of Qiagen DNeasy Blood & Tissue Kits, followed by quantification via NanoDrop 2000 (Thermo Fisher Scientific).

We amplified a 319-bp fragment of COI using forward primer LCO1490 (5'-GGTCAACAAATCATAAAGATATTGG-3') and reverse primer mICOLintR (5'-GGNGGRTANANNNGTYCANCCNGYNCC-3') (Leray et al., 2013). All samples were carried out with two rounds of PCR. In the first round, both forward and reverse primers were tailed with tags (12–17 bp) for sample identification. In the second round, we added Illumina adapters to the amplicons from the first PCR, thus avoiding the tag jumping that can arise during library preparation of amplicon mixtures (Schnell, Bohmann, & Gilbert, 2015). A table of tags and primers is in Supplementary Information (Table S1). All PCRs were performed on a Mastercycler Pro (Eppendorf) in 20- μl reaction volumes, each containing 2 μl 10 \times buffer (Mg^{2+} plus), 0.2 mM dNTPs, 0.4 μM of each primer, 1 μl DMSO, 0.4 μl BSA (bovine serum albumin) (TaKaRa Biotechnology Co. Ltd), 0.6 U Ex Taq DNA polymerase (TaKaRa Biotechnology) and approximately 60 ng genomic DNA. Both

rounds of PCR started with an initial denaturation at 94°C for 4 min, followed by 35 cycles of 94°C for 45 s, 45°C for 45 s and 72°C for 90 s, and finishing at 72°C for 10 min. PCR products were gel-purified with QIAquick PCR Purification Kit (Qiagen). One sample failed to amplify. We pooled the 70 PCR products into two libraries and sequenced on the Illumina MiSeq (Reagent Kit V3, 300PE) at the Southwest Biodiversity Institute Regional Instrument Center in Kunming. The total number of paired-end reads returned was 13,601,908.

2.4 | Data analyses

The bioinformatic script, including parameters, for the analyses below is in Supplementary Information and will be archived in datadryad.org, along with sequence data and metadata. The R scripts and data tables are on <https://github.com/dougwyu/Sichuan2014>. Below, R packages are indicated with single quotes, and other software is italicized.

2.4.1 | Bioinformatic processing

Initial processing

We removed remnant Illumina adapter sequences with *AdapterRemoval* 2.2.0 (Schubert, Lindgreen, & Orlando, 2016), followed by Schirmer et al.'s (2015) pipeline to filter, trim, denoise and merge read pairs. Specifically, we trimmed low-quality ends using *sickle* 1.33 (Joshi & Fass, 2011), corrected sequence errors using *BayesHammer* in *SPAdes* 3.10.1 (Nikolenko, Korobeynikov, & Alekseyev, 2013) and merged reads using *PandaSeq* 2.11 (Masella, Bartram, Truszkowski, Brown, & Neufeld, 2012), all with default parameters.

Demultiplexing and clustering

We then used *QIIME* 1.9.1's *split_libraries.py* (Caporaso et al., 2010) to demultiplex reads by sample and used *usearch* 9.2.64 (Edgar, 2010) to retain reads between 300 and 330 bp, inclusive, as our amplicon is 319 bp. We used *vsearch* 2.4.3 (Rognes, Flouri, Nichols, Quince, & Mahé, 2016) for de novo chimera removal and used *CROP* 1.33 (Hao, Jiang, & Chen, 2011) to cluster the remaining reads at 97% similarity. This step produced 3,507 OTUs. We also tried *swarm* 2.2.2 (Mahé, Rognes, Quince, Vargas, & Dunthorn, 2015), but it returned huge numbers of OTUs that could not be reduced even after running through "lulu" (see below).

OTU filtration and taxonomic assignment

From the resulting sample X OTU table, we used "lulu" 0.1.0 (Frøslev et al., 2017) to combine OTUs that were likely from the same species but which had failed to be clustered by *CROP*. "lulu" identifies such "parent-child" sets by calculating pairwise similarities of all OTUs (using *vsearch*) to identify sets of high-similarity OTUs and then combining OTUs within such sets that show nested sample distributions. For example, four OTUs might be highly similar, and within this set of four, one OTU contains the most reads and is observed in ten samples. This OTU is the parent, and daughters are inferred if they

are present in a subset of the parent's samples. We ended with 1,506 OTUs.

A common filtering step is to remove OTUs made up of few reads (e.g. 1-read OTUs), as these are more likely to be artefactual (e.g. Yu et al., 2012; Zepeda-Mendoza, Bohmann, Carmona Baez, & Gilbert, 2016). For instance, PCR errors can generate clusters of sequences that are sufficiently different from the parent that they cannot be identified as daughters. Such OTUs are more likely to be small because novel haplotypes typically arise in a later PCR cycle. However, the definition of small is subjective and differs with the size of the sequence dataset. We therefore used "phyloseq" 1.19.1 (McMurdie & Holmes, 2013) to plot the number of OTUs that would be filtered out at different minimum OTU sizes (see <http://evomics.org/wp-content/uploads/2016/01/phyloseq-Lab-01-Answers.html>, accessed 19 July 2018), and we chose a minimum OTU size of 44 reads, which was roughly the graph's inflection point and thus filtered out the most OTUs for the lowest minimum size. We ended with 594 OTUs.

We then used *PyNAST* 1.2.2 to align the 594 OTU sequences to a reference alignment of Arthropoda COI sequences (Yu et al., 2012) at a minimum similarity of 60%; one sequence failed to align and was deleted. The remaining sequences were translated to amino acids using the invertebrate mitochondrial codon table, and we removed 32 OTUs with sequences that contained stop codons. We carried out taxonomic assignment of the OTUs using a Naïve Bayesian Classifier (Wang, Garrity, Tiedje, & Cole, 2007) trained on the Midori UNIQUE COI dataset (Machida, Leray, Ho, & Knowlton, 2017). Sixteen OTUs assigned to non-Arthropoda taxa and two OTUs assigned to Collembola were removed. We ended with 543 OTUs.

Finally, we inspected the OTU table and set to zero those cells that had <5 reads representing that OTU in that sample, as these were more likely to be the result of sequencing error (Yu et al., 2012). In addition, we removed two samples (rows) that contained ≤100 reads total (i.e. samples with little data) and removed seven samples (rows) with <5 OTUs because these samples were potentially overly influential in analyses of species richness. These seven samples included two from native forests and five from monocultures (three BB, one EC and one JC), meaning that we disproportionately removed monocultures, making our species diversity analyses below more conservative. After these sample removals, seven OTUs were removed because they were left with few (<20) reads. Because we do not consider OTU size to be a reliable measure of biomass or abundance (Nichols et al., 2018; Piñol, Mir, Gomez-Polo, & Agustí, 2015; Yu et al., 2012), we converted the OTU table into a presence/absence (0/1) dataset. Throughout, our bias was to remove false-positive detections even at the expense of losing true-positive detections, thereby resulting in a dataset with less, but more reliable (and thus more replicable), data. We ended with 536 OTUs and 61 samples.

2.4.2 | Community analysis

OTU richness and diversities

All community analyses were performed in R 3.3.3 (R Core Team, 2017). We estimated species richness and Shannon and Simpson

diversities using two sample-based estimators: function *specpool* in “vegan” 2.4–5 (Chiu, Wang, Walther, & Chao, 2014) and “iNEXT” 2.0.12 (Hsieh, Ma, & Chao, 2016).

OTU phylogenetic diversities

Because we used a combination of *CROP+“lulu”* and “phyloseq” to combine and remove small OTUs that were likely to be artefactual, the remaining OTUs were more likely to represent true presences. Nonetheless, it remained possible that we had oversplit some biological species into multiple OTUs, as there is no single correct similarity threshold for species delimitation, and this oversplitting might have occurred more often for some taxa in some land cover types, leading to artefactual differences in species richness. However, oversplit OTUs should cluster together in a phylogenetic tree and thus contribute less to estimates of phylogenetic diversity than would OTUs from different biological species. Phylogenetic diversity should thus be a robust estimator of alpha diversity (Yu et al., 2012). To estimate sample phylogenetic diversities, we used “iNextPD” 0.3.2 (Hsieh & Chao, 2017). We built a maximum-likelihood (ML) tree in *RAxML* 8.0.0 (Stamatakis, 2014) with an alignment of the OTU-representative sequences, using a general time-reversible (GTR) model of nucleotide substitution and a gamma model of rate heterogeneity estimating the proportion of invariable sites (-m GTRGAMMAI). The algorithm used a rapid bootstrap analysis and searched for the

best-scoring ML tree (-f a), with -N 1,000 times bootstrap and -p 12,345 as the parsimony random seed. Three OTU sequences produced very long branches in the ML tree, which would skew estimates of phylogenetic diversity, and we removed them. Two of these OTUs were found in all land cover types (and thus would not have been informative), and one was only found in some cropland samples (and thus would not have informed analyses of the tree-cover sites).

Beta diversity

To visualize changes in community composition across land cover types, we ran a Bayesian ordination with “boral” 1.6.1 (Hui, 2016), which is more statistically robust than non-metric multidimensional scaling (NMDS) analysis because “boral” is model-based and thus allows us to apply a suitable error distribution so that fitted-model residuals are properly distributed. We used a binomial error distribution and no row effect as we were using presence/absence data (Figure S5). For the same reasons, we used “mvabund” 3.12.3 (Wang, Naumann, Wright, & Warton, 2012) to test the hypotheses that native forests and mixed plantations differ compositionally from each other and differ from the monocultures and croplands.

We also visualized changes in community composition with an “UpSetR” 1.3.3 intersection diagram, an alternative to Venn diagrams (Conway, Lex, & Gehlenborg, 2017), with a heatmap using the *tabasco* function in “vegan”, and with a “betapart” 1.4-1

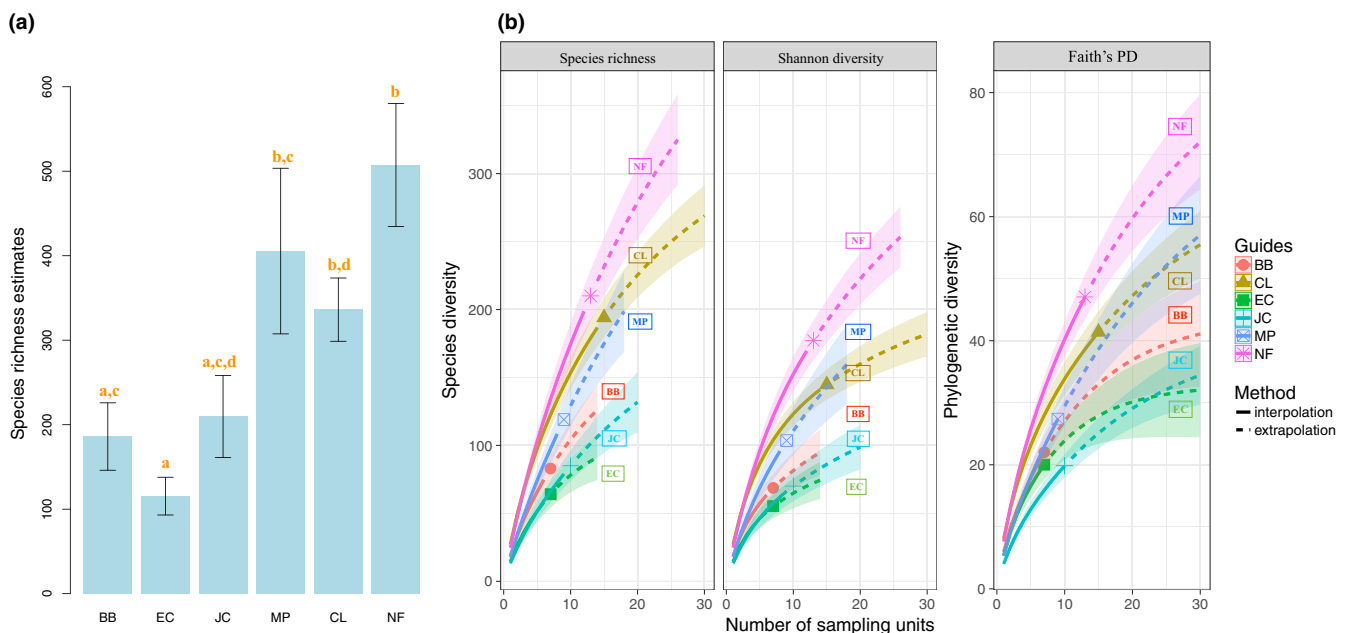


FIGURE 2 Species richness estimates across land cover type. (a) Comparisons of Chao2 species richness estimates. Land cover types sharing the same superscript are not significantly different at the $p = .05$ level (Welch's t test) after table-wide correction for multiple tests (Bonferroni). (b) “iNEXT” estimates of species richness, Shannon diversity and “iNextPD” estimates of phylogenetic diversity by land cover type, using sample-based rarefaction and extrapolation. Native forests (NF) have the highest species richness and diversities, followed by croplands (CL) and mixed plantations (MP), followed by the three monoculture plantations (BB, EC and JC). Codes for land cover types as in Figure 1. Symbols on each curve indicate the number of sampled locations per land cover type, solid lines represent interpolations, and dashed lines represent extrapolations, with 95% confidence intervals. Statistically significant pairwise differences are detected visually by non-overlapping confidence intervals and are considered conservative (MacGregor-Fors & Payton, 2013). Full iNEXT and iNextPD figures are in Figures S3 and S4

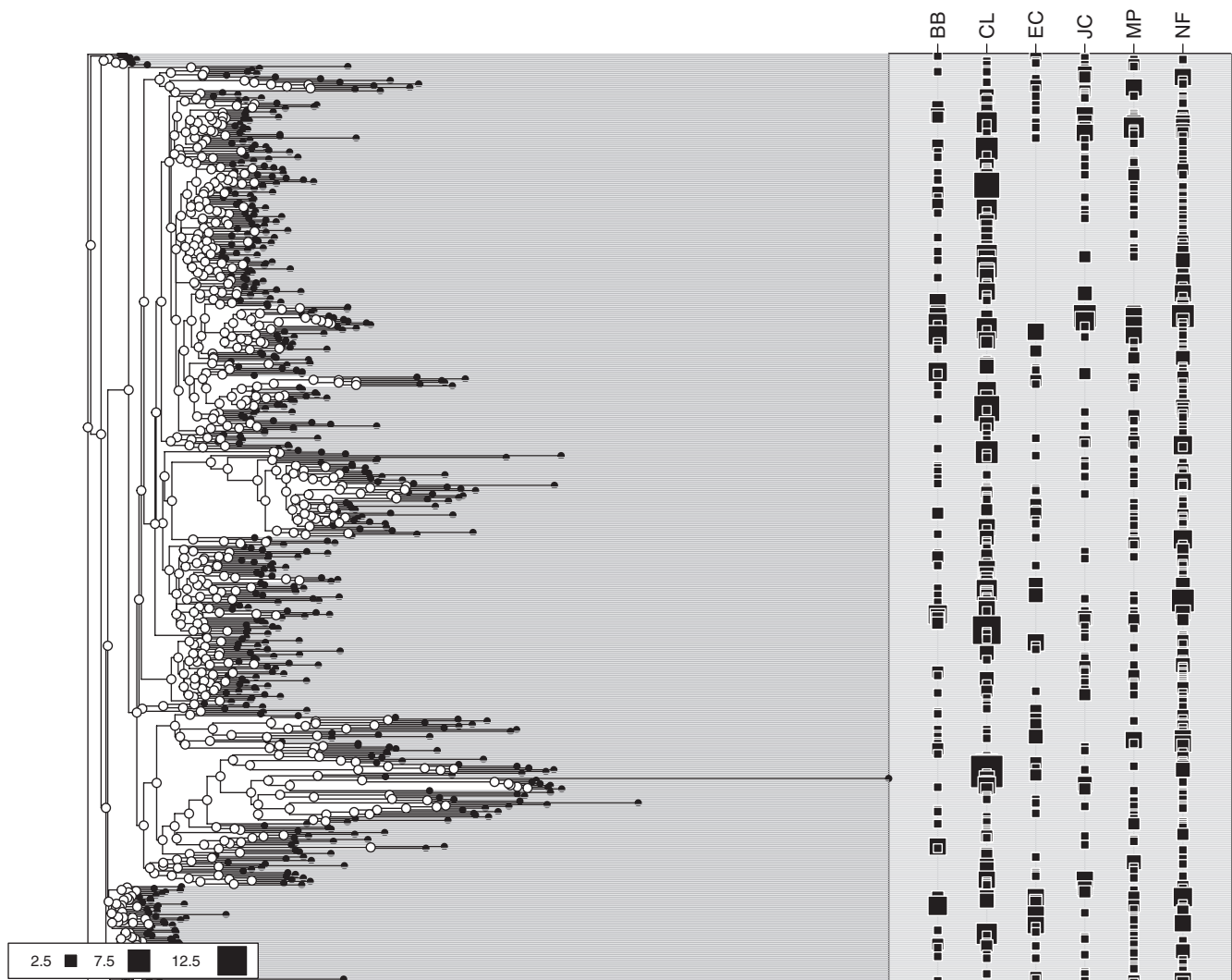


FIGURE 3 Phylogenetic distribution of OTUs by land cover type, created using “iNextPD”. Terminal nodes are black and represent the OTUs. Internal nodes are white. Sizes of the squares on the right indicate each OTU’s incidence frequency (number of samples in which the OTU is observed). Phylogenetic coverage is most complete in native forests (NF) and croplands (CL), followed by mixed plantations (MP), followed by the three monocultures (BB, EC and JC). Codes for land cover types as in Figure 1

(Baselga & Orme, 2012) analysis, which partitions beta diversity into turnover and nestedness components using binary Jaccard dissimilarities, which we visualized with NMDS using the *metaMDS* function in “vegan”. Finally, we used “metacoder” 0.2.0 (Foster, Sharpton, & Grunwald, 2017) to generate taxonomic “heat trees” to pairwise-compare the six land cover types and identify the taxa most strongly driving compositional differences.

3 | RESULTS

3.1 | Alpha diversity

Species richness and diversity are highest in native forests and croplands, followed by mixed plantations, which are in turn richer and more diverse than the monoculture plantations, with the possible exception of bamboo.

3.1.1 | OTU richness and diversities

The Chao2 estimator indicates that native forests, mixed plantations and croplands have the highest estimated species richnesses and do not differ significantly from each other (Figure 2a). Importantly, all three monocultures (bamboo, Eucalyptus and Japanese cedar) exhibit less than half the species richness of native forests and around half the species richness of mixed plantations (Figure 2a). The pairwise differences between native forests and monocultures are all statistically significant (Table S2), and the pairwise differences between mixed plantations and the three monocultures are marginally or significantly different (Figure 2a; Table S2), all after table-wide correction.

The iNEXT analysis reveals even clearer contrasts: native forests have the highest estimated asymptotic species richnesses and Shannon diversities, followed by croplands and mixed plantations,

followed by the three monocultures (Figures 2b and S3). The iNEXT-estimated richness and diversity of mixed plantations are significantly higher than all the monocultures, with the possible exception of bamboo, because the MP and BB confidence intervals touch.

3.1.2 | Phylogenetic diversities

The iNextPD analysis mirrors the iNEXT results (Figures 2b and S4). Using “iNextPD” to visualize phylogenetic coverage by land cover type (Figure 3) reveals that native forests and croplands exhibit almost complete coverage of the OTU tree, whereas mixed plantations and bamboo exhibit some coverage deficits, followed by larger coverage deficits in the other two monocultures.

3.2 | Beta diversity

Native forests are compositionally most similar to mixed plantations and most dissimilar to croplands. The differences in community composition are driven primarily by species turnover.

3.2.1 | Differences in community compositions

Ordination with “boral” (Figure 4a) shows that the primary separation is between the tree-cover types and croplands, with a significantly positive correlation between latent variable 1 and elevation ($r = -.457$, $df = 59$, $p = .0002$). The cropland sites themselves cluster into two groups by elevation. Latent variable 2 largely separates Eucalyptus monoculture from the other tree-cover types, which might reflect its distinct phytochemistry. Importantly, the mixed-plantation and (most of) the native-forest sites overlap and are encircled by the monocultures, indicating that native forests and mixed plantations are compositionally most similar.

The “UpSetR” intersection diagram (Figure 4b) is consistent with the diversity analyses (Figures 2, S3 and S4): native forests (110 OTUs) and croplands (130 OTUs) support more than 2.5 times the number of “unique species” (species detected in only one land cover type) than any of the plantations, and secondly, of the plantations, mixed plantations support the highest number of unique species (44 OTUs). The greater compositional similarity that native forests have with mixed plantations (Figure 4a) is displayed by native forests uniquely sharing more OTUs with mixed plantations (22 OTUs) than with any of the monocultures (13, 9 and 5). However, despite their overlap, “mvabund” analysis shows that the arthropod communities of mixed plantations and native

forests are still significantly distinct from each other, and from the three monocultures and croplands (Table S3).

3.2.2 | Turnover versus nestedness

Consistent with the UpSetR result that the mode in each land cover type is unique species, we found that turnover, not nestedness, dominates compositional differences (Figure 5; see Figure S7 for a heatmap visualization). In other words, the arthropod communities in the monocultures are not simply subsets of native forests or mixed plantations but contain distinct sets of species.

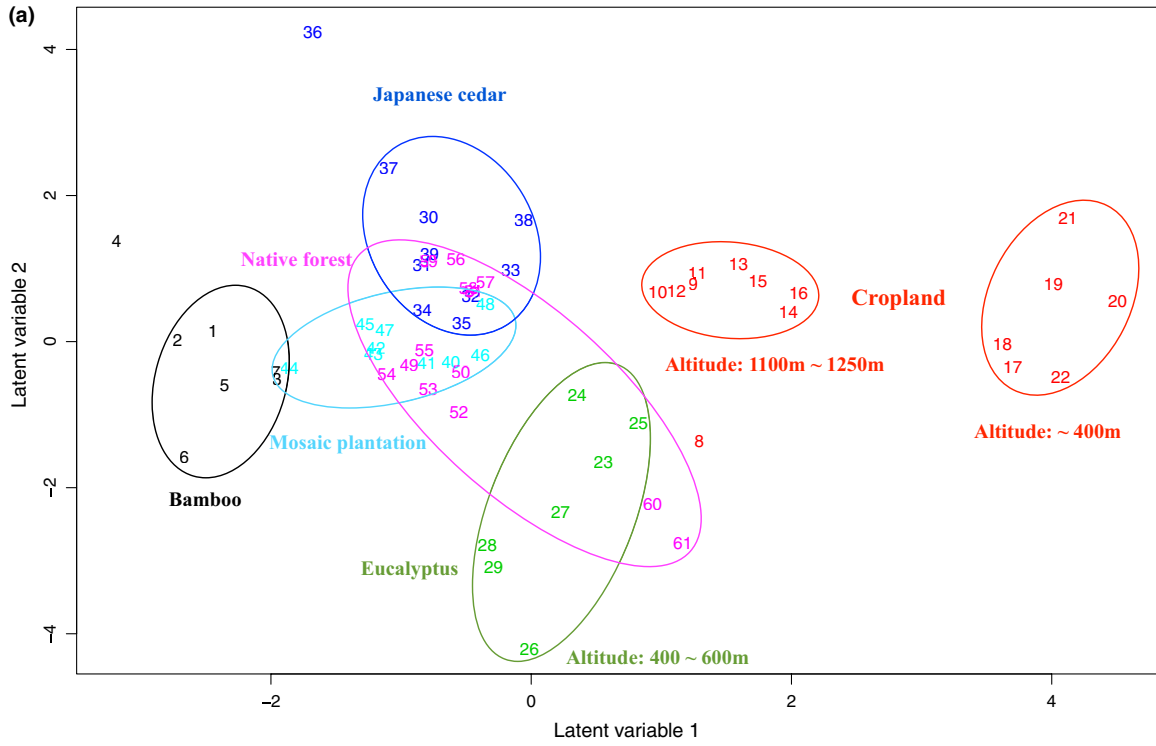
3.2.3 | Taxonomic compositions of and differences between land cover types

The 536 arthropod species in our metabarcoding dataset represent a wide range of arachnid and insect orders and, thus, represent a wide range of ecological functions (Figure 6), including generalist predators (Araneae, Formicidae) and more specialized parasites and parasitoids (Tachinidae, Phoridae, Braconidae) of other arthropods. We also observe taxa that are noted for pollination (Thysanoptera, Syrphidae), xylophagy (Isoptera), and various modes of detritivory, fungivory, frugivory, herbivory and animal parasitism (Lepidoptera, Hemiptera, Diptera, Orthoptera, Formicidae, Thysanoptera).

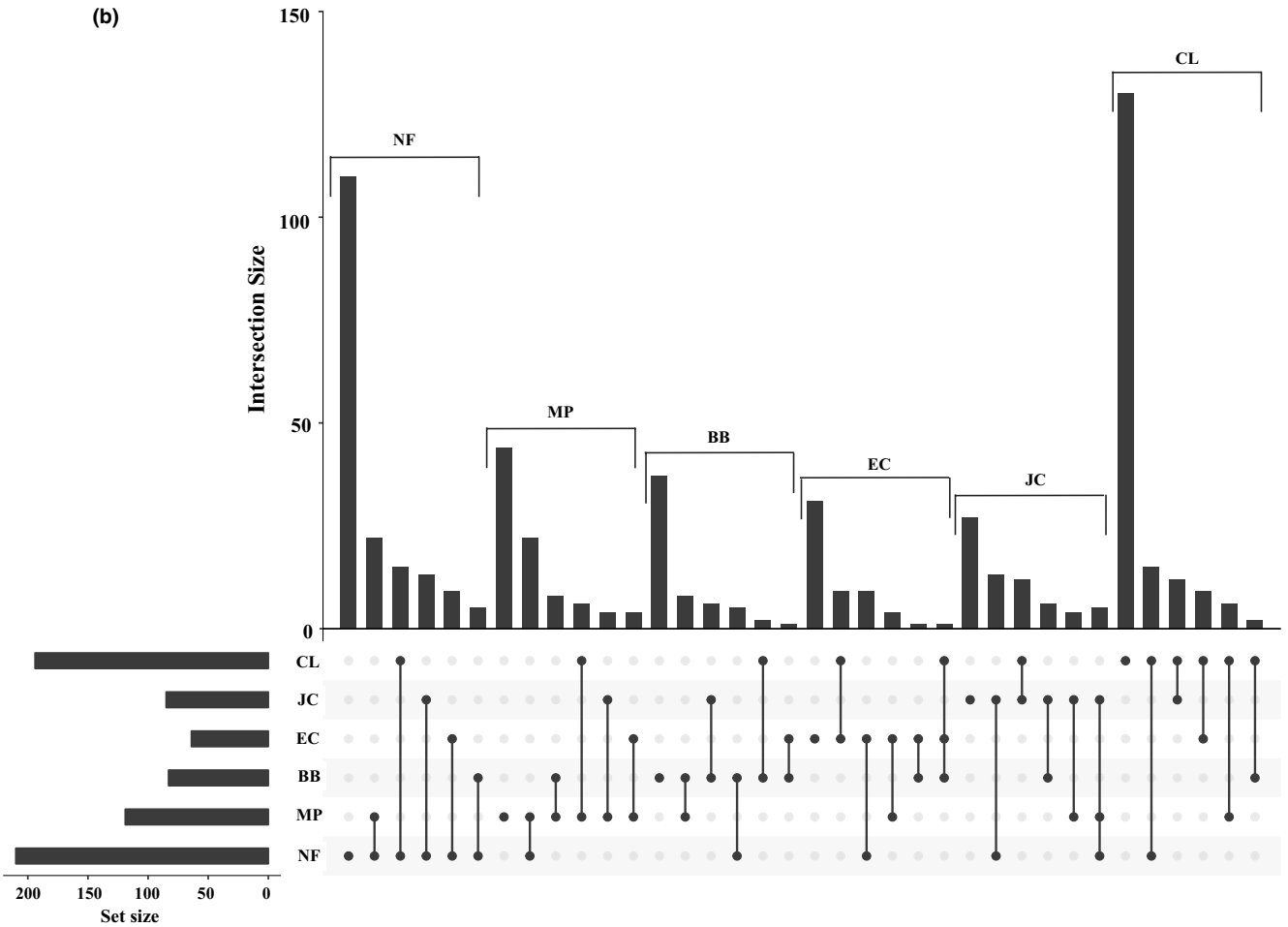
Although the “boral” ordination (Figure 4a) reveals compositional similarity between mixed plantations and native forests, it does not reveal the taxa that are most responsible for this similarity, and for the differences with the other tree-cover types. With “metacoder” heat trees (Figure 6 inset), we can identify the taxa that are driving this similarity and the differences, and what we see is that mixed plantations and native forests “differ in the same ways” from the monocultures. (a) Relative to bamboo, mixed plantations and native forests both have slightly more Lepidoptera OTUs. (b) Relative to Eucalyptus, mixed plantations and native forests both have more Diptera OTUs and fewer of the three OTUs assigned to genera *Mycetophila*, *Sonema* and *Homaloxestis*, which can be taken as Eucalyptus indicator species. (c) Finally, relative to Japanese cedar, mixed plantations and native forests both have more Araneae and Lepidoptera OTUs, fewer Hemiptera OTUs and fewer of the OTU assigned to *Mycetophila*. Heat-tree differences at higher taxonomic ranks (e.g. more Araneae-assigned OTUs) mean that the species which separate the two land cover types differ across samples but nonetheless are in the same higher taxon (e.g. Araneae). Finally, when we include croplands in the heat-tree comparisons (Figure S8),

FIGURE 4 Community composition differences in all land cover types. (a) “Boral” ordination. Colours represent land cover types, and numbers represent individual samples. Cropland (CL) sites separate into two clusters by elevation. Overlap of native forests (NF) and mixed-plantations (MP) points indicates greater compositional similarity between these two land cover types. Ovals manually added to visualize community groupings. Residuals of the “boral” fit in Figure S5. (b) UpSetR intersection map of OTUs unique to and shared between and among land cover types. Croplands and native forests support the highest numbers of unique OTUs (CL = 130, NF = 110), followed by the four plantations (MP = 44, BB = 37, EC = 31, JC = 27). Native forests uniquely share almost as many OTUs with mixed plantations (22 OTUs) as native forests share with the three monocultures combined (27 OTUs, =13 + 9 + 5). Horizontal bars on the left indicate the total number of OTUs in each land cover class. Codes for land cover types as in Figure 1. For clarity, only pairwise comparisons are shown. A non-truncated version is presented in Figure S6

Plot of latent variable posterior medians



(b)



we observe the largest number of heat-tree-tip differences between any two land cover types. In other words, there are multiple species-level indicators of croplands (or in the case of the *Mycetophila* OTU, an indicator of Japanese cedar and Eucalyptus).

4 | DISCUSSION

4.1 | Improving biodiversity conservation under the GFGP

Our study found that native forests support the highest levels of arthropod species richness, Shannon and Simpson diversities, and Faith's and phylogenetic diversity (Figures 2, 3, S3 and S4) and that most of those species are unique to native forests (Figure 4b), consistent with the patterns of bird diversity that were reported in Hua et al. (2016) and other biodiversity studies in plantations (Barlow, Gardner, et al., 2007; Gardner et al., 2008). In addition, our findings pertaining to the higher level of alpha diversity in mixed plantations over monocultures (Figures 2, S3 and S4), and their greater degree of compositional similarity to native forests relative to monocultures (Figures 4, 5, 6, S6, S7 and S8), corroborate those reported for birds

(but not bees) in Hua et al. (2016) and are consistent with other studies of biodiversity in tree plantations. Butterfield and Malvido (1992) showed that mixtures of broadleaves and conifers resulted in a higher species richness of carabid beetles than in conifer monocultures, and Recher, Davis, and Holmes (1987) showed that some bird species are present when in Eucalyptus–pine mixtures but absent from pine monocultures. In short, mixed plantations not only support a higher diversity of non-breeding birds but also provide a small but detectable biodiversity boost for arthropods. Finally, we found that compositional differences among tree-cover types are almost entirely dominated by species turnover, not nestedness, meaning that some species were only detected in the monocultures. This result is consistent with the pattern of moth communities in primary, secondary and plantation forests studied by Hawes et al. (2009). In their findings, all three of their tree-cover types (primary and secondary forest, Eucalyptus plantation) contained large numbers of unique species in three moth families (Arctiidae, Saturniidae and Sphingidae).

Given the balance of evidence, we reaffirm our previous policy recommendations that the GFGP should prioritize the retention and restoration of native forests, and when restoring native forests is

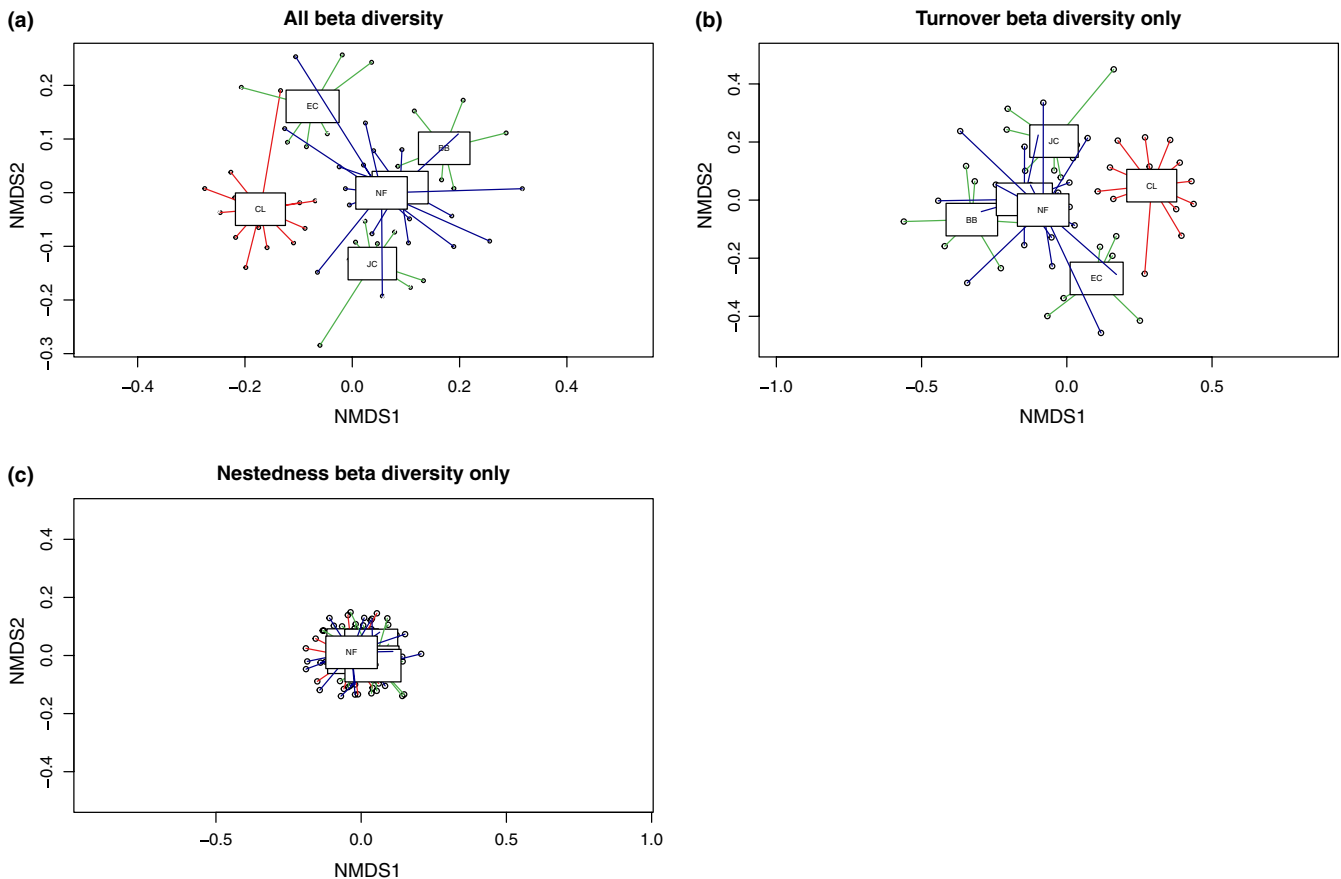


FIGURE 5 NMDS (non-metric multidimensional scaling) ordination of beta diversity by land cover type (binary Jaccard dissimilarities), partitioned with “betapart”. (a) Total beta diversity. (b) Beta diversity based on species turnover only. (c) Beta diversity based on species nestedness only. Turnover accounts for most the observed beta diversity across land cover types, which is visualized as greater distances between points in the turnover figure (b) and almost no distances between points in the nestedness figure (c). Codes for land cover types as in Figure 1

not possible, we secondarily encourage mixed-species plantings over extensive monocultures, at least in western China where we conducted this study. The foundation of these recommendations is now broadened to include 536 species-resolution taxa ranging across the Arthropoda. Given the growing understanding of the biodiversity implications of plantations compared with native forests in different forest biomes across the world (Bremer & Farley, 2010; Fierro, Grez, Vergara, Ramírez-Hernández, & Micó, 2017), these recommendations likely apply to other regions in China where GFGP is relevant, but their applicability will benefit from additional field studies and from anticipated technical advances in DNA-based biodiversity assessment. In the future, it will likely be insightful to carry out time series biodiversity surveys, as our dataset represents only a single time point, but the temporal turnover of forest arthropod communities is high (Barsoum et al., 2019). It is possible that the differences in biodiversity levels that we have detected are even

stronger when integrated over time. Another important variable that we did not measure is sample biomass, given recent evidence that insect biomass has been dropping around the world (e.g. Hallmann et al., 2017). Because we observed high species richness and diversity in our cropland sampling sites (Figures 2, 3, S3 and S4), where agriculture is small-scale in nature, our a priori expectation is that biomass has probably not declined here as rapidly as elsewhere, but this clearly needs testing and should of course now be a standard metric in biodiversity surveys.

Greater levels of arthropod biodiversity in native forest is not a surprise, given their more diverse vegetation structures and species compositions, which are well known to be positively correlated with arthropod diversity (Castagneyrol & Jactel, 2012; Haddad et al., 2009; Stork, Mcbroom, Gely, & Hamilton, 2015; Zhang et al., 2016), but the greater diversity and similarity of mixed plantations to native forests is somewhat surprising, especially as they mostly just

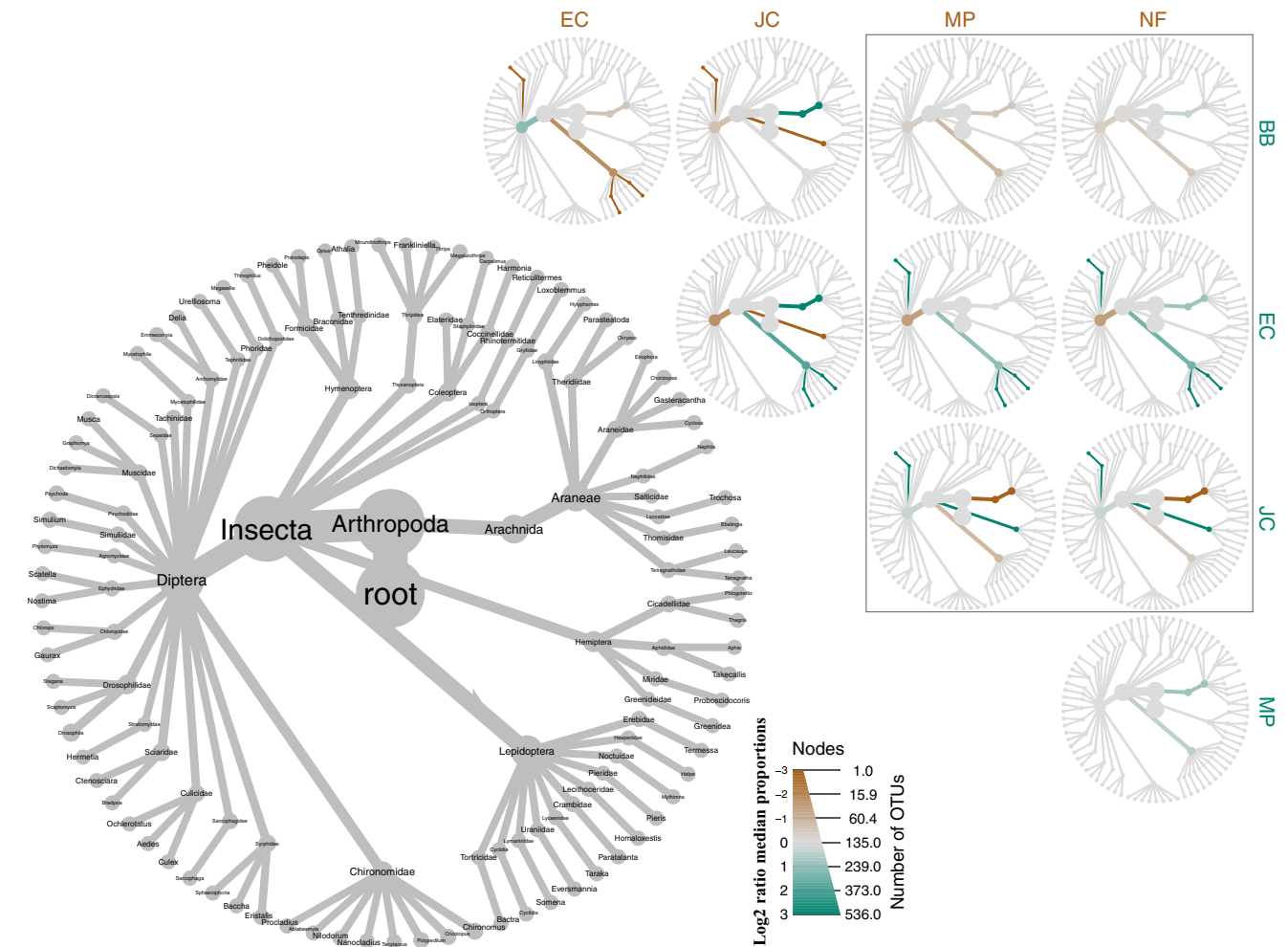


FIGURE 6 Pairwise taxonomic comparisons of all land cover types. Upper right triangle: greener branches indicate taxa that are relatively more abundant (in numbers of OTUs) in the land cover types along the right column, and browner branches indicate taxa that are relatively more abundant in the land cover types along the top row. Lower left: taxonomic identities of the branches. Note that this is a taxonomic tree, not a phylogenetic tree. Legend: width indicates number of OTUs at a given taxonomic rank, and colour indicates relative differences in \log_2 (number of OTUs). Codes for land cover types as in Figure 1. A figure including croplands and a zoomable taxonomic tree is in supplementary information (Figure S8 and S9)

comprise small-scale monocultures, planted in checkerboard pattern. However, planting different tree species near each other not only provides more diverse vegetation per se but also, because the species vary in height and three-dimensional structure, almost certainly allow greater sunlight penetration to the understorey, which in turn should result in greater availability of food and other resources. This mechanism is consistent with our finding that bamboo, which does not create closed canopies, exhibits the highest richness and diversity of the monocultures (Figures 2, S3 and S4). We note that 95% confidence interval overlap is considered an overly conservative test for statistical significance at the $p = .05$ level (MacGregor-Fors & Payton, 2013). A more diverse, and presumably higher-biomass, arthropod community in turn could also support a richer bird community, at least for the insectivorous subset of the community. Our results thus point to a plausible mechanism for why bird diversity is boosted in mixed plantations.

In this study, we report evidence for a biodiversity benefit of native forests over GFPP plantations, which we might think trades off against a greater value of timber sales from plantations. However, even excluding biodiversity, which they did not study, Cao, Zhang, and Su (2019) have recently shown that plantations in China also return a lower net value of other ecosystem services relative to native forests, even after counting income from timber sales. Plantations require a high initial outlay for tree planting, some non-native tree species like Eucalyptus require more water input than do native tree species, and more management effort is required to protect plantations from pest attack. In contrast, timber sale values are low. Cao et al.'s findings complement and strengthen our recommendation (Hua et al., 2016) to prioritize native-forest recovery and expansion over creating plantations.

4.2 | Methodological comments on metabarcoding and studies of biodiversity patterns

Metabarcoding provides an efficient method for interrogating biodiversity samples, but because of its reliance on PCR, metabarcoding datasets tend to contain a non-trivial amount of noise. This noise manifests as a large number of false-positive OTUs, which are filtered out heuristically. Such false OTUs especially complicate efforts to estimate alpha diversity. Here, we applied several filtering steps to remove false OTUs, and we also used "iNextPD" to generate robust comparisons of alpha diversity by estimating phylogenetic diversity instead of species richness. This approach has been previously shown to be reliable (Yu et al., 2012). Another approach, which became available only after we had completed the wet-laboratory portion of our study, is to subject each sample to multiple, independently tagged PCRs (typically three) and to bioinformatically filter out sequences that fail to appear in at least two of the PCRs above some minimum number of reads; such sequences are more likely to be PCR or sequencing errors. This is implemented in the DAME protocol of Zepeda-Mendoza et al. (2016; also see Alberdi, Aizpuru, Gilbert, & Bohmann, 2018).

With regard to studies of biodiversity patterns, we follow Magurran, Dornelas, Moyes, Gotelli, and McGill (2015; Magurran,

2016) in recommending that we should focus less on explaining change in species *richness* and more on explaining change in species *composition* as a function of natural and anthropogenic causes. The argument is that anthropogenically disturbed communities can maintain species richness and even phylogenetic diversity, even as local, or worse still, endemic, species go extinct and are replaced by cosmopolitan species. In our study, croplands support an arthropod community similar in richness and diversity to that of mixed plantations and just below that of native forests (Figures 2, 3, 4b, S3 and S4), but the species composition of croplands is distinct from those in native forests (Figures 4, 5, S6, S7 and S8). Croplands therefore cannot compensate for the loss of the biodiversity dependent on native forests.

ACKNOWLEDGEMENTS

We thank Y. Liu, Q. Wei, S.K. Robinson, S. Francis, R. Zeidler, W. Hua, P. Li, M. Xu and Y. Yao for helpful discussions and logistical support. Special thanks go to our field assistants: S. Huang, M. Tong and K. Du. We thank Ding Zhaoli for sequencing support. We are grateful for the support of Ji Yinqiu, Wu Chunying, Yang Chunyan and Wang Yunyu. D.W. Yu and X. Y. Wang were supported by the National Natural Science Foundation of China (41661144002, 31670536, 31400470, 31500305), the Key Research Program of Frontier Sciences, CAS (QYZDY-SSW-SMC024), the Bureau of International Cooperation project (GJHZ1754), the Strategic Priority Research Program of the Chinese Academy of Sciences (XDA20050202, XDB31000000), the Ministry of Science and Technology of China (2012FY110800), the University of East Anglia, and the State Key Laboratory of Genetic Resources and Evolution at the Kunming Institute of Zoology. F. Hua and D. S. Wilcove were supported by the High Meadows Foundation and Princeton University. L. Wang was supported by the National Nature Science Foundation of China (31872963) and the Informatization Program of the Chinese Academy of Sciences (XXH13505-03-102-2).

DATA AVAILABILITY STATEMENT

Sequence data have been submitted to GenBank under accession number SAMN09981891. Bioinformatic scripts are in Supplementary Information. R scripts and data tables are available at <https://github.com/dougwyu/Sichuan2014>.

ORCID

Xiaoyang Wang  <https://orcid.org/0000-0002-0650-6868>

REFERENCES

- Alberdi, A., Aizpuru, O., Gilbert, M. T. P., & Bohmann, K. (2018). Scrutinizing key steps for reliable metabarcoding of environmental samples. *Methods in Ecology and Evolution*, 9, 134–147. <https://doi.org/10.1111/2041-210X.12849>

- Allan, D. G., Harrison, J. A., Navarro, R. A., Van Wilgen, B. W., & Thompson, M. W. (1997). The impact of commercial afforestation on bird populations in Mpumalanga Province, South Africa—insights from bird-atlas data. *Biological Conservation*, 79, 173–185. [https://doi.org/10.1016/S0006-3207\(96\)00098-5](https://doi.org/10.1016/S0006-3207(96)00098-5)
- Barlow, J., Gardner, T. A., Araujo, I. S., Avila-Pires, T. C., Bonaldo, A. B., Costa, J. E., ... Peres, C. A. (2007). Quantifying the biodiversity value of tropical primary, secondary, and plantation forests. *Proceedings of the National Academy of Sciences*, 104, 18555–18560. <https://doi.org/10.1073/pnas.0703333104>
- Barlow, J., Overall, W. L., Araujo, I. S., Gardner, T. A., & Peres, C. A. (2007). The value of primary, secondary and plantation forests for fruit-feeding butterflies in the Brazilian Amazon. *Journal of Applied Ecology*, 44, 1001–1012. <https://doi.org/10.1111/j.1365-2664.2007.01347.x>
- Barsoum, N., Bruce, C., Forster, J., Ji, Y., Yu, D. W., Lodge, A. H., ... Kingdom, U. (2019). The devil is in the detail: Metabarcoding of arthropods provides a sensitive measure of biodiversity response to forest stand composition compared with surrogate measures of biodiversity. *Ecological Indicators*, 101, 313–323. <https://doi.org/10.1016/j.ecolind.2019.01.023>
- Bartholomew, C. S., & Prowell, D. (2005). Pan compared to malaise trapping for bees (Hymenoptera: Apoidea) in a longleaf pine savanna. *Journal of the Kansas Entomological Society*, 78, 390–392. <https://doi.org/10.2317/0409.24.1>
- Baselga, A., & Orme, C. D. L. (2012). Betapart: An R package for the study of beta diversity. *Methods in Ecology and Evolution*, 3, 808–812. <https://doi.org/10.1111/j.2041-210X.2012.00224.x>
- Bremer, L. L., & Farley, K. A. (2010). Does plantation forestry restore biodiversity or create green deserts? A synthesis of the effects of land-use transitions on plant species richness. *Biodiversity and Conservation*, 19, 3893–3915. <https://doi.org/10.1007/s10531-010-9936-4>
- Brockerhoff, E. G., Jactel, H., Parrotta, J. A., Quine, C. P., & Sayer, J. (2008). Plantation forests and biodiversity: Oxymoron or opportunity? *Biodiversity and Conservation*, 17, 925–951. <https://doi.org/10.1007/s10531-008-9380-x>
- Bryan, B. A., Gao, L., Ye, Y., Sun, X., Connor, J. D., Crossman, N. D., ... Hou, X. (2018). China's response to a national land-system sustainability emergency. *Nature*, 559, 193–204. <https://doi.org/10.1038/s41586-018-0280-2>
- Buscardo, E., Smith, G. F., Kelly, D. L., Freitas, H., Iremonger, S., Mitchell, F. J. G., ... McKee, A.-M. (2008). The early effects of afforestation on biodiversity of grasslands in Ireland. *Biodiversity and Conservation*, 17, 1057–1072. <https://doi.org/10.1007/s10531-007-9275-2>
- Butterfield, J., & Malvido, J. B. (1992). Effect of mixed-species tree planting on the distribution of soil invertebrates. *Special Publications Series of the British Ecological Society*, 11, 255–268.
- Cao, S., Zhang, J., & Su, W. (2019). Difference in the net value of ecological services between natural and artificial forests in China. *Conservation Biology*, 00, 1–8. <https://doi.org/10.1111/cobi.13293>
- Caporaso, J. G., Kuczynski, J., Stombaugh, J., Bittinger, K., Bushman, F. D., Costello, E. K., ... Knight, D. (2010). QIIME allows analysis of high-throughput community sequencing data. *Nature Methods*, 7, 335–336. <https://doi.org/10.1038/nmeth.f.303.QIIME>
- Castagneyrol, B., & Jactel, H. (2012). Unraveling plant – animal diversity relationships: A meta-regression analysis. *Ecology*, 93, 2115–2124. <https://doi.org/10.1890/11-1300.1>
- Chiu, C. H., Wang, Y. T., Walther, B. A., & Chao, A. (2014). An improved nonparametric lower bound of species richness via a modified good-turing frequency formula. *Biometrics*, 70, 671–682. <https://doi.org/10.1111/biom.12200>
- Conway, J. R., Lex, A., & Gehlenborg, N. (2017). UpSetR: An R package for the visualization of intersecting sets and their properties. *Bioinformatics*, 33, 2938–2940. <https://doi.org/10.1093/bioinformatics/btx364>
- Cristescu, M. E. (2014). From barcoding single individuals to metabarcoding biological communities: Towards an integrative approach to the study of global biodiversity. *Trends in Ecology and Evolution*, 29, 566–571. <https://doi.org/10.1016/j.tree.2014.08.001>
- Deiner, K., Bik, H. M., Mächler, E., Seymour, M., Lacoursière-Roussel, A., Altermatt, F., ... Bernatchez, L. (2017). Environmental DNA metabarcoding: Transforming how we survey animal and plant communities. *Molecular Ecology*, 26, 5872–5895. <https://doi.org/10.1111/mec.14350>
- Delang, C. O., & Yuan, Z. (2015). *China's grain for green program*. Switzerland: Springer International Publishing.
- Deng, L., Liu, G., & Shanguan, Z. (2014). Land-use conversion and changing soil carbon stocks in China's "Grain-for-Green" Program: A synthesis. *Global Change Biology*, 20, 3544–3556. <https://doi.org/10.1111/gcb.12508>
- Deng, L., Shanguan, Z., & Li, R. (2012). Effects of the grain-for-green program on soil erosion in China. *International Journal of Sediment Research*, 27, 120–127. [https://doi.org/10.1016/S1001-6279\(12\)60021-3](https://doi.org/10.1016/S1001-6279(12)60021-3)
- Edgar, R. C. (2010). Search and clustering orders of magnitude faster than BLAST. *Bioinformatics*, 26, 2460–2461. <https://doi.org/10.1093/bioinformatics/btq461>
- Elbrecht, V., Peinert, B., & Leese, F. (2017). Sorting things out: Assessing effects of unequal specimen biomass on DNA metabarcoding. *Ecology and Evolution*, 7, 6918–6926. <https://doi.org/10.1002/ece3.3192>
- Elsen, P. R., Ramesh, K., & Wilcove, D. S. (2018). Conserving Himalayan birds in highly seasonal forested and agricultural landscapes. *Conservation Biology*, 32, 1313–1324. <https://doi.org/10.1111/cobi.13145>
- Fierro, A., Grez, A. A., Vergara, P. M., Ramírez-Hernández, A., & Micó, E. (2017). How does the replacement of native forest by exotic forest plantations affect the diversity, abundance and trophic structure of saproxylic beetle assemblages? *Forest Ecology and Management*, 405, 246–256. <https://doi.org/10.1016/j.foreco.2017.09.026>
- Foster, Z. S. L., Sharpton, T., & Grunwald, N. J. (2017). Metacoder: An R package for manipulation and heat tree visualization of community taxonomic data from metabarcoding. *PLoS Computational Biology*, 13, e1005404. <https://doi.org/10.1101/071019>
- Frøslev, T. G., Kjølner, R., Bruun, H. H., Ejrnæs, R., Brunbjerg, A. K., Pietroni, C., & Hansen, A. J. (2017). Algorithm for post-clustering curation of DNA amplicon data yields reliable biodiversity estimates. *Nature Communications*, 8, 1–11. <https://doi.org/10.1038/s41467-017-01312-x>
- Gardner, T. A., Hernandez, M. I. M., Barlow, J., & Peres, C. A. (2008). Understanding the biodiversity consequences of habitat change: The value of secondary and plantation forests for neotropical dung beetles. *Journal of Applied Ecology*, 45, 883–893. <https://doi.org/10.1111/j.1365-2664.2008.01454.x>
- Haddad, N. M., Crutsinger, G. M., Gross, K., Haarstad, J., Knops, J. M. H., & Tilman, D. (2009). Plant species loss decreases arthropod diversity and shifts trophic structure. *Ecology Letters*, 12, 1029–1039. <https://doi.org/10.1111/j.1461-0248.2009.01356.x>
- Hallmann, C. A., Sorg, M., Jongejans, E., Siepel, H., Hofland, N., Schwan, H., ... de Kroon, H. (2017). More than 75 percent decline over 27 years in total flying insect biomass in protected areas. *PLoS ONE*, 12, e0185809. <https://doi.org/10.1371/journal.pone.0185809>
- Hao, X., Jiang, R., & Chen, T. (2011). Clustering 16S rRNA for OTU prediction: A method of unsupervised Bayesian clustering. *Bioinformatics*, 27, 611–618. <https://doi.org/10.1093/bioinformatics/btq725>
- Hartley, M. J. (2002). Rationale and methods for conserving biodiversity in plantation forests. *Forest Ecology and Management*, 155, 81–95. [https://doi.org/10.1016/S0378-1127\(01\)00549-7](https://doi.org/10.1016/S0378-1127(01)00549-7)
- Hawes, J., Motta, S., Overall, W. L., Barlow, J., Gardner, T. A., & Peres, C. A. (2009). Diversity and composition of Amazonian moths in primary, secondary and plantation forests. *Journal of Tropical Ecology*, 25, 281–300. <https://doi.org/10.1017/S0266467409006038>
- Hsieh, T. C., & Chao, A. (2017). Rarefaction and extrapolation: Making fair comparison of abundance-sensitive phylogenetic diversity

- among multiple assemblages. *Systematic Biology*, 66, 100–111. <https://doi.org/10.1093/sysbio/syw073>
- Hsieh, T. C., Ma, K. H., & Chao, A. (2016). iNEXT: An R package for rarefaction and extrapolation of species diversity (Hill numbers). *Methods in Ecology and Evolution*, 7, 1451–1456. <https://doi.org/10.1111/2041-210X.12613>
- Hu, C., Fu, B., Chen, L., & Gulinck, H. (2006). Farmer's attitudes towards the Grain-for-Green programme in the Loess hilly area, China: A case study in two small catchments. *International Journal of Sustainable Development & World Ecology*, 13, 211–220. <https://doi.org/10.1080/13504500609469673>
- Hua, F., Wang, L., Fisher, B., Zheng, X., Wang, X., Yu, D. W., ... Wilcove, D. S. (2018). Tree plantations displacing native forests: The nature and drivers of apparent forest recovery on former croplands in Southwestern China from 2000 to 2015. *Biological Conservation*, 222, 113–124. <https://doi.org/10.1016/j.biocon.2018.03.034>
- Hua, F., Wang, X., Zheng, X., Fisher, B., Wang, L., Zhu, J., ... Wilcove, D. S. (2016). Opportunities for biodiversity gains under the world's largest reforestation programme. *Nature Communications*, 7, 1–11. <https://doi.org/10.1038/ncomms12717>
- Hui, F. K. C. (2016). boral – Bayesian ordination and regression analysis of multivariate abundance data in R. *Methods in Ecology and Evolution*, 7, 744–750. <https://doi.org/10.1111/2041-210X.12514>
- Jactel, H., & Brockerhoff, E. G. (2007). Tree diversity reduces herbivory by forest insects. *Ecology Letters*, 10, 835–848. <https://doi.org/10.1111/j.1461-0248.2007.01073.x>
- Ji, Y., Ashton, L., Pedley, S. M., Edwards, D. P., Tang, Y., Nakamura, A., ... Yu, D. W. (2013). Reliable, verifiable and efficient monitoring of biodiversity via metabarcoding. *Ecology Letters*, 16, 1245–1257. <https://doi.org/10.1111/ele.12162>
- Joshi, N. A., & Fass, J. N. (2011). *Sickle: A sliding-window, adaptive, quality-based trimming tool for FastQ files (Version 1.33) [Software]*. Available at <https://github.com/najoshi/sickle>
- Leray, M., Yang, J. Y., Meyer, C. P., Mills, S. C., Agudelo, N., Ranwez, V., ... Machida, R. J. (2013). A new versatile primer set targeting a short fragment of the mitochondrial COI region for metabarcoding metazoan diversity: Application for characterizing coral reef fish gut contents. *Frontiers in Zoology*, 10, 1–14. <https://doi.org/10.1186/1742-9994-10-34>
- Lindenmayer, D. B., & Hobbs, R. J. (2004). Fauna conservation in Australian plantation forests – a review. *Biological Conservation*, 119, 151–168. <https://doi.org/10.1016/j.biocon.2003.10.028>
- Liu, J., & Diamond, J. (2005). China's environment in a globalizing world. *Nature*, 435, 1179–1186. <https://doi.org/10.1038/4351179a>
- Liu, J., Li, S., Ouyang, Z., Tam, C., Chen, X., Liu, J., ... Chen, X. (2008). Ecological and socioeconomic effects of China's policies for ecosystem services. *Proceedings of the National Academy of Sciences*, 105, 9477–9482. <https://doi.org/10.1073/pnas.0706436105>
- Liu, J., Ouyang, Z., Pimm, S. L., Raven, P. H., Wang, X., Miao, H., & Han, N. (2003). Protecting China's biodiversity. *Science*, 300, 1240–1241. <https://doi.org/10.1126/science.1078868>
- Liu, Z., & Lan, J. (2015). The Sloping Land Conversion Program in China: Effect on the livelihood diversification of rural households. *World Development*, 70, 147–161. <https://doi.org/10.1016/j.worlddev.2015.01.004>
- Long, H. L., Heilig, G. K., Wang, J., Li, X. B., Luo, M., Wu, X. Q., & Zhang, M. (2006). Land use and soil erosion in the upper reaches of the Yangtze River: Some socio-economic considerations on China's Grain-for-Green Programme. *Land Degradation & Development*, 17, 589–603. <https://doi.org/10.1002/ldr.736>
- Ma, K., Shen, X., Grumbine, R. E., & Corlett, R. (2017). China's biodiversity conservation research in progress. *Biological Conservation*, 210, 1–2. <https://doi.org/10.1016/j.biocon.2017.05.029>
- MacGregor-Fors, I., & Payton, M. E. (2013). Contrasting diversity values: Statistical inferences based on overlapping confidence intervals. *PLoS ONE*, 8, e56794. <https://doi.org/10.1371/journal.pone.0056794>
- Machida, R. J., Leray, M., Ho, S. L., & Knowlton, N. (2017). Data descriptor: Metazoan mitochondrial gene sequence reference datasets for taxonomic assignment of environmental samples. *Scientific Data*, 4, 1–7. <https://doi.org/10.1038/sdata.2017.27>
- Magurran, A. E. (2016). How ecosystems change. *Science*, 351, 448–449. <https://doi.org/10.1126/science.aad6758>
- Magurran, A. E., Dornelas, M., Moyes, F., Gotelli, N. J., & McGill, B. (2015). Rapid biotic homogenization of marine fish assemblages. *Nature Communications*, 6, 1–5. <https://doi.org/10.1038/ncomms9405>
- Mahé, F., Rognes, T., Quince, C., de Vargas, C., & Dunthorn, M. (2015). Swarm v2: Highly-scalable and high-resolution amplicon clustering. *PeerJ*, 3, e1420. <https://doi.org/10.7717/peerj.1420>
- Masella, A. P., Bartram, A. K., Truszkowski, J. M., Brown, D. G., & Neufeld, J. D. (2012). PANDAseq: Paired-end assembler for illumina sequences. *BMC Bioinformatics*, 13, 31–37. <https://doi.org/10.1186/1471-2105-13-31>
- McMurdie, P. J., & Holmes, S. (2013). Phyloseq: An R package for reproducible interactive analysis and graphics of microbiome census data. *PLoS ONE*, 8, e61217. <https://doi.org/10.1371/journal.pone.0061217>
- Nichols, R. V., Vollmers, C., Newsom, L. A., Wang, Y., Heintzman, P. D., Leighton, M. K., ... Shapiro, B. (2018). Minimizing polymerase biases in metabarcoding. *Molecular Ecology Resources*, 18, 927–939. <https://doi.org/10.1111/1755-0998.12895>
- Nikolenko, S. I., Korobeynikov, A. I., & Alekseyev, M. A. (2013). BayesHammer: Bayesian clustering for error correction in single-cell sequencing. *BMC Genomics*, 14, 1–11. <https://doi.org/10.1186/1471-2164-14-S1-S7>
- Ouyang, Z., Zheng, H., Xiao, Y., Polasky, S., Liu, J., Xu, W., ... Daily, G. C. (2016). Improvements in ecosystem services from investments in natural capital. *Science*, 352, 1455–1460. <https://doi.org/10.1126/science.aaf2295>
- Piñol, J., Mir, G., Gomez-Polo, P., & Agustí, N. (2015). Universal and blocking primer mismatches limit the use of high-throughput DNA sequencing for the quantitative metabarcoding of arthropods. *Molecular Ecology Resources*, 15, 819–830. <https://doi.org/10.1111/1755-0998.12355>
- Pyne, K. (2013). Conserving China's biodiversity. *Earth Common Journal*, 3, 44–53.
- R Core Team (2017). *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. <https://www.R-project.org/>
- Recher, H. F., Davis, W. E., & Holmes, R. T. (1987). Ecology of brown and striated thornbills in forests of south-eastern New South Wales, with comments on forest management. *Emu - Austral Ornithology*, 87, 1–13. <https://doi.org/10.1071/MU9870001>
- Ren, G., Young, S. S., Wang, L., Wang, W., Long, Y., Wu, R., ... Yu, D. W. (2015). Effectiveness of China's National Forest Protection Program and nature reserves. *Conservation Biology*, 29, 1368–1377. <https://doi.org/10.1111/cobi.12561>
- Rognes, T., Flouri, T., Nichols, B., Quince, C., & Mahé, F. (2016). VSEARCH: A versatile open source tool for metagenomics. *PeerJ*, 4, e2584. <https://doi.org/10.7717/peerj.2584>
- Sayer, J. A., & Sun, C. (2003). The impacts of policy reforms on forest environments and biodiversity. In F. H., William, B., Brian, & X., Jintao (Eds), *China's Forests: Global lessons from market reforms* (pp. 177–194). Beijing: China forestry publishing. doi: <https://doi.org/10.4324/9781936331239>
- Schirmer, M., Ijaz, U. Z., D'Amore, R., Hall, N., Sloan, W. T., & Quince, C. (2015). Insight into biases and sequencing errors for amplicon sequencing with the Illumina MiSeq platform. *Nucleic Acids Research*, 43, e37. <https://doi.org/10.1093/nar/gku1341>
- Schnell, I. B., Bohmann, K., & Gilbert, M. T. P. (2015). Tag jumps illuminated - reducing sequence-to-sample misidentifications in metabarcoding studies. *Molecular Ecology Resources*, 15, 1289–1303. <https://doi.org/10.1111/1755-0998.12402>
- Schubert, M., Lindgreen, S., & Orlando, L. (2016). AdapterRemoval v2: Rapid adapter trimming, identification, and read merging. *BMC Research Notes*, 9, 1–7. <https://doi.org/10.1186/s13104-016-1900-2>

- Stamatakis, A. (2014). RAxML version 8: A tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics*, 30, 1312–1313. <https://doi.org/10.1093/bioinformatics/btu033>
- Stork, N. E., Mcbroom, J., Gely, C., & Hamilton, A. J. (2015). New approaches narrow global species estimates for beetles, insects, and terrestrial arthropods. *Proceedings of the National Academy of Sciences*, 112, 7519–7523. <https://doi.org/10.1073/pnas.1502408112>
- Tao, Y., Huang, D., Jin, X., & Guo, L. (2010). Make an inventory of China's biodiversity. *Bulletin of the Chinese Academy of Sciences*, 24, 212–217.
- Turner, B. L., Lambin, E. F., & Reenberg, A. (2007). The emergence of land change science for global environmental change and sustainability. *Proceedings of the National Academy of Sciences*, 104, 20666–20671. <https://doi.org/10.1073/pnas.0704119104>
- United Nations. (2015). *Transforming our world: The 2030 agenda for sustainable development*. Retrieved from <https://sustainabledevelopment.un.org/post2015>
- Wang, J., Peng, J., Zhao, M., Liu, Y., & Chen, Y. (2017). Significant trade-off for the impact of Grain-for-Green Programme on ecosystem services in North-western Yunnan, China. *Science of the Total Environment*, 574, 57–64. <https://doi.org/10.1016/j.scitotenv.2016.09.026>
- Wang, Q., Garrity, G. M., Tiedje, J. M., & Cole, J. R. (2007). Naïve Bayesian classifier for rapid assignment of rRNA sequences into the new bacterial taxonomy. *Applied and Environmental Microbiology*, 73, 5261–5267. <https://doi.org/10.1128/AEM.00062-07>
- Wang, Y., Naumann, U., Wright, S. T., & Warton, D. I. (2012). Mvabund—an R package for model-based analysis of multivariate abundance data. *Methods in Ecology and Evolution*, 3, 471–474. <https://doi.org/10.1111/j.2041-210X.2012.00190.x>
- Wang, Z. J., Jiao, J. Y., Rayburg, S., Wang, Q. L., & Su, Y. (2016). Soil erosion resistance of "Grain for Green" vegetation types under extreme rainfall conditions on the Loess Plateau, China. *Catena*, 141, 109–116. <https://doi.org/10.1016/j.catena.2016.02.025>
- Wei, Y., Yu, D., Lewis, B. J., Zhou, L. I., Zhou, W., Fang, X., ... Dai, L. (2014). Forest carbon storage and tree carbon pool dynamics under natural forest protection program in Northeastern China. *Chinese Geographical Science*, 24, 397–405. <https://doi.org/10.1007/s11769-014-0703-4>
- Wu, R., Possingham, H. P., Yu, G., Jin, T., Wang, J., Yang, F., ... Zhao, H. (2019). Strengthening China's national biodiversity strategy to attain an ecological civilization. *Conservation Letters*, 1–7. <https://doi.org/10.1111/conl.12660>
- Xie, G., Cao, S., Yang, Q., Xia, L., Fan, Z., Chen, B., & Sarah, H. (2012). *China Ecological Footprint Report 2012*.
- Xu, H., Wang, S., & Xue, D. (1999). Biodiversity conservation in China: Legislation, plans and measures. *Biodiversity and Conservation*, 8, 819–837.
- Xu, J., Yin, R., Li, Z., & Liu, C. (2006). China's ecological rehabilitation: Unprecedented efforts, dramatic impacts, and requisite policies. *Ecological Economics*, 57, 595–607. <https://doi.org/10.1016/j.ecolecon.2005.05.008>
- Yin, R., Liu, C., Zhao, M., Yao, S., & Liu, H. (2014). The implementation and impacts of China's largest payment for ecosystem services program as revealed by longitudinal household data. *Land Use Policy*, 40, 45–55. <https://doi.org/10.1016/j.landusepol.2014.03.002>
- Yin, R., Yin, G., & Li, L. (2009). Assessing China's ecological restoration programs: What's been done and what remains to be done? *Environmental Management*, 45, 442–453. https://doi.org/10.1007/978-90-481-2655-2_2
- Yu, D. W., Ji, Y., Emerson, B. C., Wang, X., Ye, C., Yang, C., & Ding, Z. (2012). Biodiversity soup: Metabarcoding of arthropods for rapid biodiversity assessment and biomonitoring. *Methods in Ecology and Evolution*, 3, 613–623. <https://doi.org/10.1111/j.2041-210X.2012.00198.x>
- Zepeda-Mendoza, M. L., Bohmann, K., Carmona Baez, A., & Gilbert, M. T. P. (2016). DAME: A toolkit for the initial processing of datasets with PCR replicates of double-tagged amplicons for DNA metabarcoding analyses. *BMC Research Notes*, 9, 1–13. <https://doi.org/10.1186/s13104-016-2064-9>
- Zhai, D. L., Xu, J. C., Dai, Z. C., Cannon, C. H., & Grumbine, R. E. (2014). Increasing tree cover while losing diverse natural forests in tropical Hainan, China. *Regional Environmental Change*, 14, 611–621. <https://doi.org/10.1007/s10113-013-0512-9>
- Zhang, K., Lin, S., Ji, Y., Yang, C., Wang, X., Yang, C., ... Yu, D. W. (2016). Plant diversity accurately predicts insect diversity in two tropical landscapes. *Molecular Ecology*, 25, 4407–4419. <https://doi.org/10.1111/mec.13770>
- Zhou, H., Van Rompaey, A., & Wang, J. (2009). Detecting the impact of the "Grain for Green" program on the mean annual vegetation cover in the Shaanxi province, China using SPOT-VGT NDVI data. *Land Use Policy*, 26, 954–960. <https://doi.org/10.1016/j.landusepol.2008.11.006>

BIOSKETCH

Douglas W. Yu is a Principal Investigator at the Kunming Institute of Zoology, China, and Professor of Ecology at the University of East Anglia, UK. His group is a leading developer of methods for rapid assessment of eukaryotic biodiversity using high-throughput DNA sequencing (metabarcoding, mitogenomics, metagenomics) and participates in multiple collaborations to provide biodiversity information for ecosystem-scale management. Yu is also a co-founder of NatureMetrics (www.naturemetrics.co.uk), which supplies DNA-based services to governments, NGOs and businesses so that they can improve policies and processes for protecting and growing biodiversity and natural capital through adaptive management.

Author contributions: X.W., F.H. and D.W.Y. conceived the research and designed the experiments; X.W. and F.H. carried out the fieldwork; L.W. created Figure 1; X.W. carried out the laboratory work; X.W. and D.W.Y. carried out the bioinformatic and statistical analyses; X.W., D.W.Y. and F.H. wrote the manuscript, with comments from all other authors.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

How to cite this article: Wang X, Hua F, Wang L, Wilcove DS, Yu DW. The biodiversity benefit of native forests and mixed-species plantations over monoculture plantations. *Divers Distrib*. 2019;00:1–15. <https://doi.org/10.1111/ddi.12972>