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# OGRDB: a reference database of inferred immune receptor genes

William Lees <sup>1,\*</sup>, Christian E. Busse <sup>2</sup>, Martin Corcoran, Mats Ohlin, Cathrine Scheepers, Frederick A. Matsen, IV, Gur Yaari, Corey T. Watson, The AIRR Community, Andrew Collins and Adrian J. Shepherd

<sup>1</sup>Institute of Structural and Molecular Biology, Birkbeck College, University of London, London WC1E 7HX, UK, <sup>2</sup>Division of B Cell Immunology, German Cancer Research Center, 69120 Heidelberg, Germany, <sup>3</sup>Department of Microbiology, Tumor and Cell Biology, Karolinska Institute, Box 280, 171 77 Stockholm, Sweden, <sup>4</sup>Department of Immunotechnology, Lund University, Medicon Village, S-223 81 Lund, Sweden, <sup>5</sup>Center for HIV and STIs, National Institute for Communicable Diseases of the National Health Laboratory Service, Sandringam, Gauteng 2131, South Africa, <sup>6</sup>Antibody Immunity Research Unit, School of Pathology, University of the Witwatersrand, Johannesburg 2050, South Africa, <sup>7</sup>Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, WA 98109-1024, USA, <sup>8</sup>Faculty of Engineering, Bar Ilan University, Ramat Gan 5290002, Israel, <sup>9</sup>Department of Biochemistry and Molecular Genetics, University of Louisville School of Medicine, Louisville, KY 40202, USA and <sup>10</sup>School of Biotechnology and Biomolecular Sciences, University of New South Wales, Sydney, New South Wales 2052, Australia

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# **ABSTRACT**

High-throughput sequencing of the adaptive immune receptor repertoire (AIRR-seg) is providing unprecedented insights into the immune response to disease and into the development of immune disorders. The accurate interpretation of AIRR-seq data depends on the existence of comprehensive germline gene reference sets. Current sets are known to be incomplete and unrepresentative of the degree of polymorphism and diversity in human and animal populations. A key issue is the complexity of the genomic regions in which they lie, which, because of the presence of multiple repeats, insertions and deletions, have not proved tractable with short-read whole genome sequencing. Recently, tools and methods for inferring such gene sequences from AIRR-seq datasets have become available, and a community approach has been developed for the expert review and publication of such inferences. Here, we present OGRDB, the Open Germline Receptor Database (https://ogrdb. airr-community.org), a public resource for the submission, review and publication of previously unknown receptor germline sequences together with supporting evidence.

# INTRODUCTION

The genes of B-cell and T-cell antigen receptors (IG, TR) lie in some of the most structurally complex and polymorphic regions of vertebrate genomes. Because of their repetitive nature, the presence of many copy number variants, and the variation between individuals, the IG and TR genomic loci are problematic to study via standard high-throughput genomic approaches. For example, short-read surveys of human genetic variation such as the 1000 Genomes Project (1) remain challenging to interpret in these loci, to the extent that it is unclear whether such approaches can reliably deliver information on IG and TR germline variation (2, see also https://www.internationalgenome.org/faq/whyonly-85-genome-assayable). An important consequence is that there are gaps in the current reference sets of IG germline genes and alleles—important gaps in human reference sets, and profound gaps in the sets of all other species, including those of medical and agricultural importance. Many of the sequences underlying the human germline set curated by IMGT (the international ImMunoGeneTics information system (3)), for example, were derived in the 1980s and 1990s from a small number of samples, primarily from either Caucasians or individuals of unknown ethnicity. The full extent of variation among human populations is not well understood and may be substantially underestimated (4–7). In contrast to studies of the human leukocyte antigen (HLA) (8) and the killer-cell immunoglobulinlike receptor (KIR) genes (9), there is little understanding of the common haplotypes of receptor genes. Similar, and

<sup>\*</sup>To whom correspondence should be addressed. Tel: +44 20 7631 6886; Fax: +44 20 7631 6803; Email: william@lees.org.uk

<sup>&</sup>lt;sup>†</sup>The list of endorsing members is provided as Supplementary Information.

possibly deeper, issues are arising in other species. As examples, extensive variation in IG heavy chain (IGH) genes has recently been reported between inbred laboratory mouse strains (10,11), while fish species important for food production exhibit substantial and complex genome and IG region-specific gene duplication (12).

Knowledge of IG gene variation is important. Polymorphism in the human IGHV1-69 gene has been shown to affect the antibody response to influenza A, with implications for vaccine design (13). Similar stereotyped immune responses have been observed in other infectious diseases and in contexts such as cancer and allergy (14–17). The analysis of the high-throughput sequencing of adaptive immune receptor repertoires (AIRR-seq) depends on an accurate germline set in order to identify clonal lineages and to correctly understand the impact of specific germline deletions and polymorphisms on the immune response. Gaps and erroneous sequences in reference sets therefore have a potentially detrimental impact on the development of effective diagnostic and therapeutic strategies (18).

In recent years, methods have been published through which personalized germline repertoires (identifying the set of germline receptor alleles expressed in the repertoire of a specific subject) can be inferred from AIRR-seq datasets (19–23). The personalized germline repertoire (referred to hereafter as a *genotype*) of any given person may be composed of previously unknown alleles as well as those already present in reference sets. Its inference from next-generation sequencing (NGS) provides a means through which highthroughput techniques can be applied to the problems of novel allele identification and population-level genetics. The AIRR Community (www.airr-community.org) - a network of over 300 practitioners in the field of AIRR-seq and the IG, TR and MH Nomenclature Sub-Committee (IMGT-NC) (http://www.imgt.org/IMGTindex/IUIS-NC. php) of the International Union of Immunological Societies (IUIS)—recently reached agreement on a process whereby inferred genes and alleles would first be reviewed by Inferred Allele Review Committees (IARCs) under the auspices of the AIRR Community, and then submitted to IMGT-NC for their consideration (24). The first alleles were submitted for review in late 2018, and the first nine human IGHV genes were affirmed by the human IARC and accepted into IMGT in May 2019. Approximately 50 more are before the review committee, pending final confirmation of supporting data, and formation of IARCs for non-human species is in progress.

Review of inferred alleles is made in the context of individual AIRR-seq based genotypes, together with the accession numbers and details of underlying International Nucleotide Sequence Database Collaboration (INSDC) depositions. Ensuring data quality, tracking the progress of reviews and presenting the outcome to the community transparently was initially daunting, and it soon became apparent that computational support would be necessary. OGRDB (the Open Germline Receptor Database: https://ogrdb.airr-community.org) was developed to meet this need. It provides full supporting evidence for published alleles, recording the repertoires in which they have been observed and preserving references and history as additional information is received. Submitted alleles must be fully sup-

ported by records in public sequence repositories such as NIH Genbank and SRA, and links to these records are provided by OGRDB. For database users, OGRDB provides both the complete set of inferred alleles that have been affirmed through the IARC process, and also the inferred genotypes in which they were found. OGRDB may be viewed on large, medium or even small screen devices. Registration is freely available but only required for making new submissions: all published data is available without the need to register or log in.

# **DATABASE METHODS AND RESULTS**

#### Inferred sequences

On the Sequences tab, OGRDB presents a browsable list of inferred alleles affirmed by the IARC review process, and, where a sequence has been accepted by IMGT, the canonical name allocated by IMGT. The Affirmation Level indicates the number of independent affirmations of the sequence by the IARC, up to a limit of three. The full set of sequences can be downloaded either in FASTA format, or in a provisional AIRR Community-defined format, which contains enriched metadata. Clicking on a sequence name provides a detailed view of the sequence record, including, importantly, the supporting evidence submitted to IARC that underlies the inferred sequence. Sequence entries are versioned: the Notes and History tabs provide details of IARC's review, and any updates made to the record.

# Submissions and inferred genotypes

Supporting evidence for inferred sequences is contained in one or more submissions. Submissions underlying a particular sequence can be accessed by clicking on the Submission IDs in the Evidence section of the sequence record. Alternatively, the Submissions tab of OGRDB provides a browsable view of all published submissions.

A submission reports the analysis of one or more AIRR-seq repertoires. The analysis is provided in the form of an inferred genotype and accompanying information. The genotype lists the set of genes within a particular locus that lead to expressed productive sequences in the repertoire. For each gene, it provides usage statistics, including its observed frequency and statistics on its usage in combination with other genes (Figure 1). OGRDB supports the derivation of a standardized genotype format from a number of analytical tools: conversion scripts and guidance for submitters are available on the website. Alongside the genotype itself, the scripts provide a range of plots that can be used by the submitter and by reviewers to assess the strength and quality of the novel inferences (Figure 2): these can be attached to a submission for additional information.

tThe repertoires underlying a submission are published in an INSDC repository such as the NIH Sequence Read Archive (SRA). Records for novel sequences, and extracted reads from the repertoire(s) that specifically support them, are also deposited. For ease of use and to facilitate review, OGRDB retrieves and displays metadata from NIH repositories. The sequences of reference alleles in each genotype are checked against the current IMGT reference set, and any discrepancies are identified. Human alle-

# **Novel Alleles**

Allele name	Sequences	Closest Reference	Closest in Host	NT Diffs (reference)	NT Diffs (host)	NT Substs (reference)	AA Diffs (reference)	AA Substs (reference)	Sequence			
IGHV1-58*01+G29A	178	IGHV1-58*01	IGHV1-58*01	1		G30A	0		Q	8	•	)
IGHV3-21*01+T52C	9	IGHV3-21*01	IGHV3-21*01	1		T56C	1	L19P	Q			
IGHV3-43*01+A96G.C197T.A261G	146	IGHV3-43*01	IGHV3-9*01	3		A112G,C219T,A286G	2	T38A,T96A	Q	8		
IGHV3-64*05+A188C.G240C	281	IGHV3-64*05	IGHV3-23*04	2		A207C,G265C	1	V89L	Q	0	6	,
IGHV4-38-2*02+G76A.T136G.G169C	1912	IGHV4-38-2*02	IGHV4-38-2*01	3		G80A,T149G,G191C	3	G27D,L50R,S64T	Q			

# Genotype

Allele name	Sequences	Unmutated Seqs	Unmutated % within allele	Unmutated UMIs	Allelic %	Total unmutated population (%)	Unique Ds	Unique Js	Unique CDR3s	Unique Ds with unmutated	Unique Js with unmutated	Unique CDR3s with unmutated	Haplotyping Gene	Haplotyping Ratio	Sequence	
IGHV1-18*01	1946	1314	67.52		100.00	2.00	28	13	952				Haplotyping not possible		Q	<b>✓</b>
IGHV1-2*02	9951	5949	59.78		100.00	9.00	28	13	5049				Haplotyping not possible		Q	•
IGHV1-24*01	5034	3942	78.31		100.00	6.00	28	13	2261				Haplotyping not possible		Q	<b>~</b>
IGHV1-46*01	1007	572	56.80		100.00	0.00	27	13	477				Haplotyping not possible		Q	<b>~</b>

Figure 1. Partial screenshot of a genotype panel showing the statistics provided for inferred alleles, and, beneath, the statistics provided for all alleles (see Table 1 for a description of the information contained in these tables).

les are also checked against IgPdb (http://cgi.cse.unsw.edu. au/~ihmmune/IgPdb/information.php), a repository of suspected polymorphisms in human IG genes. Although Ig-Pdb remains a useful resource for accessing past inferences, the database has been largely inactive in recent years. This resource also differs from OGRDB in that alleles submitted to IgPdb do not require accompanying data that support the inferences. Submissions are not subject to expert review/evaluation and curation, and users of the database are unable to access evidence in support of the inferences. As a consequence of the lack of curation, IgPdb almost certainly includes sequences that have been reported in error. OGRDB also represents a significant improvement over IgPdb because tracking and cross-referencing maintains database correctness over time. For example, a novel sequence that is subsequently accepted by IMGT will still be identified in the underlying genotype as a novel sequence with the name assigned to it by the submitter, but it will be tagged with a note to show that it now matches a sequence in the IMGT reference set.

#### **Submission and review process**

This report focuses on the available published data; however, we encourage researchers who have suitable data sets to submit inferred alleles for review. Submission is online via

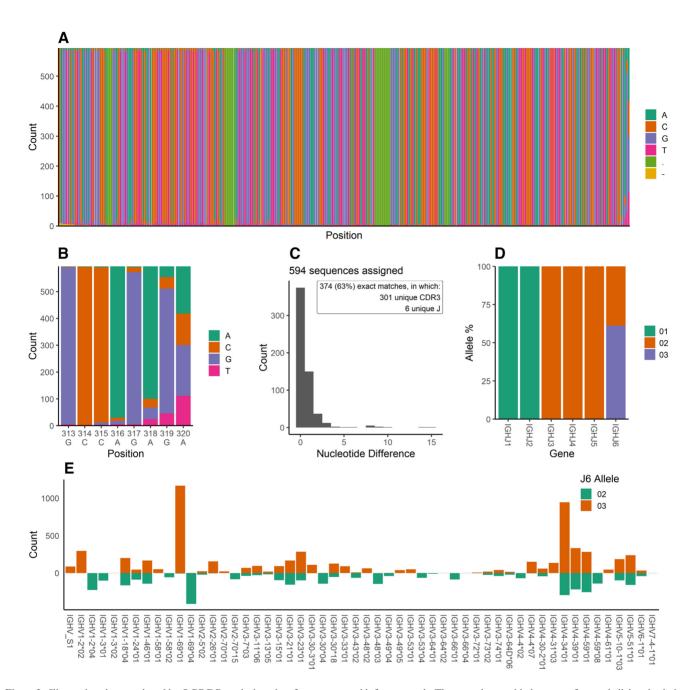


Figure 2. Illustrative plots produced by OGRDB analysis scripts from supported inference tools. These can be provided as part of a novel allele submission, or used independently by researchers interested in exploring the usage characteristics of an AIRR-seq repertoire. (A) IMGT alignment of all sequences assigned to a novel allele (IMGT gaps shown as '.', base calls with low confidence as '-'), showing, in this case, a high-quality underlying dataset. (B) for V-genes, a 'zoom' of the previous plot showing the final 8 nucleotides at the 3' end and illustrating the effect of the recombination process on the difficulty associated with inference of the final nucleotide of the IGHV gene based on the underlying data (27) (for J-genes, a similar plot is shown of the 5' end). (C) a histogram, similar to that produced by IgDiscover (21), showing the reads assigned to the allele, distributed by the number of nucleotide differences to the reference (or inferred reference) sequence. (D) For the analysis of novel V-genes, a plot showing the usage of J-alleles within each J-gene, which can be used to identify heterozygosity (28). In this case, the subject is heterozygous in the IGHJ6 gene. For the analysis of novel J-genes, a similar plot of V-gene allele usage is provided. (E) where heterozygosity may be present, a plot is provided showing the number of reads of each V-gene, split by their usage of the heterozygous allele (in this case alleles \*02 and \*03 of IGHJ6). The novel allele IGHV\_S1 is found exclusively in association with the IGHJ6\*03 and no other alleles of IGHV\_S1 were identified in the genotype: the exclusive association with \*03 therefore provides additional support for the novel inference.

**Table 1.** Information provided in the OGRDB standardized genotype

Field	Description							
sequence_id	Identifier of the allele (either IMGT, or the name assigned by the submitter to an inferred							
	gene)							
sequences	Overall number of sequences assigned to this allele							
closest_reference	For inferred alleles, the closest reference gene and allele, as inferred by the tool							
closest_host	For inferred alleles, the closest reference gene and allele that is in the subject's inferred							
nt_diff	genotype  For inferred alleles, the number of nucleotides that differ between this sequence and the closest reference gene and allele							
nt_diff_host	For inferred alleles, the number of nucleotides that differ between this sequence and the closest reference gene and allele that is in the subject's inferred genotype							
nt_substitutions	For inferred alleles, comma-separated list of nucleotide substitutions (e.g. G112A) between the sequence and the closest reference gene and allele. IMGT numbering is used for V-genes, and number from start of coding sequence for D- or J- genes.							
aa_diff	For inferred alleles, the number of amino acids that differ between this sequence and the closest reference gene and allele							
aa_substitutions	For inferred alleles, the list of amino acid substitutions (e.g. A96N) between the sequence an the closest reference gene and allele. IMGT numbering is used for V-genes, and number from							
unmutated_sequences	start of coding sequence for D- or J- genes.  The number of sequences exactly matching this unmutated sequence							
assigned_unmutated_frequency	The number of sequences exactly matching this allele divided by the number of sequences assigned to this allele, *100							
unmutated_umis	The number of molecules (identified by Unique Molecular Identifiers) exactly matching this							
allelic_percentage	unmutated sequence (if UMIs were used)  The number of sequences exactly matching the sequence of this allele divided by the number of sequences exactly matching any allele of this specific gene, *100							
unmutated_frequency	The number of sequences exactly matching this sequence divided by the number of sequence exactly matching any allele of any gene, *100							
unique_vs	The number of V allele calls (i.e. unique allelic sequences) found associated with this allele							
unique_ds	The number of D allele calls (i.e. unique allelic sequences) found associated with this allele							
unique_js	The number of J allele calls (i.e. unique allelic sequences) found associated with this allele							
unique_cdr3s	The number of unique CDR3s found associated with this allele							
unique_vs_unmutated	The number of V allele calls (i.e. unique allelic sequences) associated with unmutated sequences of this allele							
unique_ds_unmutated	The number of D allele calls (i.e. unique allelic sequences) associated with unmutated sequences of this allele							
unique_js_unmutated	The number of J allele calls (i.e. unique allelic sequences) associated with unmutated sequences of this allele							
unique_cdr3s_unmutated	The number of unique CDR3s associated with unmutated sequences of this allele							
haplotyping_gene	The gene or genes from which haplotyping was inferred, where haplotyping is possible (e.g. IGHJ6)							

Provision of statistics for each allele in the personalized genotype (both reference alleles and novel alleles) allows the novel inferences to be considered in the context of overall gene usage (usage frequency, exact unmutated matches, association with distinct CDR 3 and so on), and also provides useful aggregate information on overall gene usage.

OGRDB, and full details are provided on the site. OGRDB provides a submission and review workflow that supports multiple IARCs covering different species and loci, tracking the progress of all submissions and providing versioning and tracking of published sequences. The software is open source and may be of interest to other groups requiring a system for submission, review and publication.

# DISCUSSION

Our vision for OGRDB is to provide a rich and accessible record of observed receptor gene sequences, including not only the sequences themselves, but also detailed attribution and underlying information on their prevalence. We also aim to support and enhance the productivity of expert review, given the likely explosion of new information as high-throughput methods become increasingly available. While coverage today is limited to sequences inferred from AIRR-seq records, OGRDB contains the necessary fields to support genomic information, including annotation of noncoding regions, and we are interested to explore its extension to records derived from long-read sequencing, which can offer a partial (although to date not a complete) solution to the assembly problems inherent in high-volume genomic sequencing of the receptor loci.

The novel allele submission process is necessarily rigorous, requiring supporting information to be deposited in databases of record. Sequence sets supporting V-gene inferences must be full-length, excluding many data sets available today from consideration. We recognize the success and long-term impact of OGRDB depends greatly on the willingness of AIRR-seq data generators to submit allele inference data and supporting information. We will explore avenues to reduce the submission burden through the use of automated pipelines, and to leverage non-compliant data sets to provide additional support for previously affirmed inferences. VDJbase (https://www.vdjbase.org) (reported separately in this issue) is a database of inferences derived via a computational pipeline from a wider set of repertoires. We will develop integration between the two, and will explore opportunities to identify those datasets analysed within VDJbase which contain good candidate submissions for OGRDB and IARC review. By using results from the VDJbase pipeline, we can encourage submission of candidate sequences and reduce the burden on submitters. We intend to develop the two resources alongside one another to provide a rich and useful resource to the community.

Finally, while we have focussed on IG genes in this description and in the work of the current review committees, at least one analytical tool is available for TR gene inference (20), and OGRDB has been developed to accommodate the review and publication of both. We are keen to extend the scope of review to additional species and loci, and would welcome approaches from any groups interested in participating.

#### CONCLUSION

The study of the adaptive immune system is of high clinical importance. While it has been facilitated by the widespread adoption of high-throughput techniques such as AIRR-seq (25,26), existing genomic resources have been hampered by the complexity of the regions concerned, and do not represent the diversity of human and animal populations. The development of high-quality, publicly available reference sets is a key aim of the AIRR Community. The presentation of inferred alleles through OGRDB is an important part of that vision. The partnerships now formed between OGRDB, IMGT, and VDJbase will further ensure the success of this initiative and increase its long-term impact.

As the application of AIRR-seg becomes even more pervasive in both the research and clinical arenas, we expect that the use of germline databases will also evolve. For example, it is likely that the storage of variant information beyond the coding regions will also offer important value to the community. With the initial foundation of OGRDB now established, we anticipate that OGRDB will be able to take on some of these extended initiatives. In addition, it will be important to begin understanding the prevalence of allelic variants stored in OGRDB and IMGT at the scale of populations and species. The concerted development of OGRDB with VDJbase, which is currently focused on providing rich information on AIRR-seq genotypes and alleles in human populations, will facilitate critical cross-talk between these two databases and ultimately provide a deeper view of IG and TR genetic diversity, and the importance of this diversity to the immune response.

# **DATA AVAILABILITY**

No registration or sign-on is required to access published data on OGRDB. All published information is freely available under a Creative Commons CC0 License. The source code is published at <a href="https://github.com/airr-community/ogre">https://github.com/airr-community/ogre</a> and is available under a European Union Public License v1.2.

# SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online.

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#### REFERENCES

- 1. 1000 Genomes Project Consortium, Auton, A., Brooks, L.D., Durbin, R.M., Garrison, E.P., Kang, H.M., Korbel, J.O., Marchini, J.L., McCarthy, S., McVean, G.A. et al. (2015) A global reference for human genetic variation. *Nature*, 526, 68–74.
- 2. Watson, C.T., Matsen, F.A., Jackson, K.J.L., Bashir, A., Smith, M.L., Glanville, J., Breden, F., Kleinstein, S.H., Collins, A.M. and Busse, C.E. (2017) Comment on 'A database of human immune receptor alleles recovered from population sequencing data'. *J. Immunol.*, 198, 3371–3373
- 3. Lefranc, M.-P., Giudicelli, V., Duroux, P., Jabado-Michaloud, J., Folch, G., Aouinti, S., Carillon, E., Duvergey, H., Houles, A., Paysan-Lafosse, T. *et al.* (2015) IMGT, the international ImMunoGeneTics information system 25 years on. *Nucleic Acids Res.*, 43, D413–D422.
- 4. Watson, C.T., Steinberg, K.M., Huddleston, J., Warren, R.L., Malig, M., Schein, J., Willsey, A.J., Joy, J.B., Scott, J.K., Graves, T.A. *et al.* (2013) Complete haplotype sequence of the human immunoglobulin heavy-chain variable, diversity, and joining genes and characterization of allelic and copy-number variation. *Am. J. Hum. Genet.*, **92**, 530–546.
- Watson, C.T., Glanville, J. and Marasco, W.A. (2017) The individual and population genetics of antibody immunity. *Trends Immunol.*, 38, 459–470.
- Scheepers, C., Shrestha, R.K., Lambson, B.E., Jackson, K.J.L., Wright, I.A., Naicker, D., Goosen, M., Berrie, L., Ismail, A., Garrett, N. et al. (2015) Ability to develop broadly neutralizing HIV-1 antibodies is not restricted by the germline Ig gene repertoire. J. Immunol., 194, 4371–4378.
- 7. Jackson, K.J.L., Wang, Y., Gaeta, B.A., Pomat, W., Siba, P., Rimmer, J., Sewell, W.A. and Collins, A.M. (2011) Divergent human populations show extensive shared IGK rearrangements in peripheral blood B cells. *Immunogenetics*, **64**, 3–14.
- Osoegawa, K., Mallempati, K.C., Gangavarapu, S., Oki, A., Gendzekhadze, K., Marino, S.R., Brown, N.K., Bettinotti, M.P., Weimer, E.T., Montero-Martín, G. et al. (2019) HLA alleles and haplotypes observed in 263 US families. Hum. Immunol., doi:10.1016/j.humimm.2019.05.018.
- Wroblewski, E.E., Parham, P. and Guethlein, L.A. (2019) Two to Tango: Co-evolution of hominid natural killer cell receptors and MHC. Front. Immunol., 10, 177.
- Collins, A.M., Wang, Y., Roskin, K.M., Marquis, C.P. and Jackson, K.J.L. (2015) The mouse antibody heavy chain repertoire is germline-focused and highly variable between inbred strains. *Philos. Trans. R. Soc. B*, 370, 20140236.
- 11. Watson, C.T., Kos, J.T., Gibson, W.S., Newman, L., Deikus, G., Busse, C.E., Smith, M.L., Jackson, K.J.L. and Collins, A.M. (2019) Germline murine immunoglobulin IGHV genes in wild-derived and classical inbred strains: a comparison. *Immunol. Cell Biol.*, doi:10.1111/imcb.12288.
- 12. Magadan, S., Sunyer, O.J. and Boudinot, P. (2015) Unique features of fish immune repertoires: particularities of adaptive immunity within the largest group of vertebrates. *Results Probl. Cell Differ.*, **57**, 235–264.
- 13. Avnir, Y., Watson, C.T., Glanville, J., Peterson, E.C., Tallarico, A.S., Bennett, A.S., Qin, K., Fu, Y., Huang, C.-Y., Beigel, J.H. *et al.* (2016) IGHV1-69 polymorphism modulates anti-influenza antibody repertoires, correlates with IGHV utilization shifts and varies by ethnicity. *Sci. Rep.*, 6, 20842.
- Henry Dunand, C.J. and Wilson, P.C. (2015) Restricted, canonical, stereotyped and convergent immunoglobulin responses. *Philos. Trans. R. Soc. Lond., B, Biol. Sci.*, 370, 20140238.

- 15. Bonsignori, M., Liao, H.-X., Gao, F., Williams, W.B., Alam, S.M., Montefiori, D.C. and Haynes, B.F. (2017) Antibody-virus co-evolution in HIV infection: paths for HIV vaccine development. Immunol. Rev., **275**, 145–160.
- 16. Padyukov, L., Hahn-Zoric, M., Blomqvist, S.R., Ulanova, M., Welch, S.G., Feeney, A.J., Lau, Y.L. and Hanson, L.A. (2001) Distribution of human kappa locus IGKV2-29 and IGKV2D-29 alleles in Swedish Caucasians and Hong Kong Chinese. Immunogenetics, 53, 22-30.
- 17. Levin, M., Levander, F., Palmason, R., Greiff, L. and Ohlin, M. (2017) Antibody-encoding repertoires of bone marrow and peripheral blood-a focus on IgE. J. Allergy Clin. Immunol., 139, 1026–1030.
- 18. Xochelli, A., Agathangelidis, A., Kavakiotis, I., Minga, E., Sutton, L.A., Baliakas, P., Chouvarda, I., Giudicelli, V., Vlahavas, I., Maglaveras, N. et al. (2015) Immunoglobulin heavy variable (IGHV) genes and alleles: new entities, new names and implications for research and prognostication in chronic lymphocytic leukaemia. Immunogenetics, 67, 61-66.
- 19. Gadala-Maria, D., Yaari, G., Uduman, M. and Kleinstein, S.H. (2015) Automated analysis of high-throughput B-cell sequencing data reveals a high frequency of novel immunoglobulin V gene segment alleles. Proc. Natl. Acad. Sci. U.S.A., 112, E862-E870.
- 20. Zhang, W., Wang, I.-M., Wang, C., Lin, L., Chai, X., Wu, J., Bett, A.J., Dhanasekaran, G., Casimiro, D.R. and Liu, X. (2016) IMPre: an accurate and efficient software for prediction of T- and B-cell receptor germline genes and alleles from rearranged repertoire data. Front. Immunol., 7, 457.
- 21. Corcoran, M.M., Phad, G.E., Néstor, V.B., Stahl-Hennig, C., Sumida, N., Persson, M.A.A., Martin, M. and Karlsson Hedestam, G.B. (2016) Production of individualized V gene databases

- reveals high levels of immunoglobulin genetic diversity. Nat. Commun 7 13642
- 22. Ralph, D.K. and Matsen, F.A. (2019) Per-sample immunoglobulin germline inference from B cell receptor deep sequencing data. PLoS Comput. Biol., 15, e1007133.
- 23. Safonova, Y. and Pevzner, P.A. (2019) De novo inference of diversity genes and analysis of Non-canonical V(DD)J recombination in immunoglobulins. Front Immunol., 10, 987.
- 24. Ohlin, M., Scheepers, C., Corcoran, M., Lees, W.D., Busse, C.E., Bagnara, D., Thörnqvist, L., Bürckert, J.-P., Jackson, K.J.L., Ralph, D. et al. (2019) Inferred allelic variants of immunoglobulin receptor genes: a system for their evaluation, documentation, and naming. Front. Immunol., 10, 435.
- 25. Georgiou, G., Ippolito, G.C., Beausang, J., Busse, C.E., Wardemann, H. and Quake, S.R. (2014) The promise and challenge of high-throughput sequencing of the antibody repertoire. Nat. Biotechnol., 32, 158-168.
- 26. Miho, E., Yermanos, A., Weber, C.R., Berger, C.T., Reddy, S.T. and Greiff, V. (2018) Computational strategies for dissecting the high-dimensional complexity of adaptive immune repertoires. Front. Immunol., 9, 224.
- 27. Thörnqvist, L. and Ohlin, M. (2018) Critical steps for computational inference of the 3'-end of novel alleles of immunoglobulin heavy chain variable genes - illustrated by an allele of IGHV3-7. Mol. Immunol., 103, 1-6.
- 28. Kidd, M.J., Chen, Z., Wang, Y., Jackson, K.J., Zhang, L., Boyd, S.D., Fire, A.Z., Tanaka, M.M., Gaëta, B.A. and Collins, A.M. (2012) The inference of phased haplotypes for the immunoglobulin H Chain V region gene loci by analysis of VDJ gene rearrangements. J. Immunol., 188, 1333-1340.