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Scotland's Rural College

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Fox, NJ; Smith, LA; Houdijk, JGM; Athanasiadou, S; Hutchings, MR

Published in: International Journal for Parasitology

DOI: 10.1016/j.ijpara.2018.06.001

First published: 11/08/2018

Document Version Peer reviewed version

Link to publication

Citation for pulished version (APA):

Fox, NJ., Smith, LA., Houdijk, JGM., Athanasiadou, S., & Hutchings, MR. (2018). Ubiquitous parasites drive a 33% increase in methane yield from livestock. *International Journal for Parasitology*, *48*(13), 1017 - 1021. https://doi.org/10.1016/j.ijpara.2018.06.001

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Manuscript reference number: IJPara18_061R1

1 Ubiquitous parasites drive 33% increase in methane yield from livestock

3	N.J. Fox ^a , L.A. Smith ^{a*} , J.G.M. Houdijk ^a , S. Athanasiadou ^a , M.R. Hutchings ^a
4	^a SRUC, Peter Wilson Building, King's Buildings, West Mains Rd, Edinburgh EH9 3JG
5	*Corresponding author: lesley.smith@sruc.ac.uk +44 (0)131 651 9352
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24 Abstract

Of anthropogenic methane emissions 40% can be attributed to agriculture, the majority of which 25 are from enteric fermentation in livestock. With international commitments to tackle drivers of 26 27 climate change, there is a need to lower global methane emissions from livestock production. Gastrointestinal helminths (parasitic worms) are globally ubiquitous and represent one of the 28 most pervasive challenges to the health and productivity of grazing livestock. These parasites 29 influence a number of factors affecting methane emissions including feed efficiency, nutrient 30 use, and production traits. However, their effects on methane emissions are unknown. This is the 31 first study that empirically demonstrates disease-driven increases in methane (CH₄) yield in 32 livestock (grams of CH₄ per kg of dry matter intake). We do this by measuring methane 33 emissions (in respiration chambers), dry matter intake (DMI), and production parameters for 34 35 parasitised and parasite-free lambs. This study shows that parasite infections in lambs can lead to a 33% increase in methane yield (g CH_4/kg DMI). This knowledge will facilitate more accurate 36 calculations of the true environmental costs of parasitism in livestock, and reveals the potential 37 benefits of mitigating emission through controlling parasite burdens. 38

39

40 Key words: methane, greenhouse gas, climate change, parasites, disease; lambs

41 1. Introduction

Strategies for minimising global greenhouse gas (GHG) emissions from livestock systems are 42 vital. Agriculture contributes an estimated 18% of GHG emissions (Steinfeld et al., 2006), with 43 approximately half of these emissions coming from meat and dairy (Garnett, 2009). Emissions 44 45 are a particular concern in small ruminant (sheep and goat) milk and meat production as 56% of the global domestic ruminant population are small ruminants (Marino et al., 2016), and enteric 46 fermentation is responsible for the majority of emissions in these systems (Gerber et al., 2013). 47 48 Minimising GHG emissions from livestock systems will become increasingly important as demand for livestock products grows; by 2050 the global sheep population is expected to 49 increase by 60%, from 1.7 billion in 2000 to about 2.7 billion by 2050 (Foresight, 2011). With 50 51 little chance of decreasing emissions through an overall reduction in the numbers of farmed ruminants, other ways to mitigate ruminant methane emissions are required (Herrero et al., 2016; 52 Dangal et al., 2017). 53

54

The primary factors affecting ruminant enteric methane emissions are thought to be feed intake 55 56 levels, feed composition, and the microflora of the rumen (Lascano & Cárdenas, 2010). Consequently, mitigation options currently fall into three broad categories: 1) animal breeding 57 for improved efficiency; 2) feed supplements and feed management; and 3) rumen control and 58 59 modifiers (Marino et al., 2016). However, a number of these strategies have high costs and/or inconsistent effects (Marino et al., 2016) and reliable, affordable technologies for reducing 60 methane emissions in grazing livestock in a way that improves overall farm productivity and 61 efficiency remain elusive. 62

63

Gastrointestinal helminths are globally ubiquitous, offering the most pervasive challenge to all
grazing livestock worldwide and compromising animal health, welfare and production
efficiency. They have a substantial impact on the majority of the factors affecting methane
production, including feed intake levels, nutrient use, and rumen retention time (Houdijk et al.,
2016). Controlling gastrointestinal parasites could potentially reduce GHG emissions in grazing
livestock. However, their effects on methane emissions are currently unknown.

70

In addition to revealing the mitigation potential of reducing parasitism, understanding the impact 71 72 of parasitism on methane production is also vital in calculating the true extent of GHG emissions from livestock. Many studies have attempted to calculate the GHG emissions from livestock 73 systems, often with the aim of quantifying how changes in efficiency will impact on methane 74 production (Kipling et al., 2016; Özkan et al., 2016). Emissions estimates are generally 75 calculated based on livestock numbers, time in the production system, and basic national 76 multipliers. However, such calculations ignore the potential impacts of common infections. 77 Efforts have been made to explore the impacts of parasitism on production efficiency and time 78 on pasture, and the consequent implications for emissions (Kenyon et al., 2013). However this 79 approach assumes that methane yield remains constant regardless of infection status, and that 80 parasitism has no additional effect beyond the higher overall feed intake due to decreased 81 production efficiency and increased time to slaughter. 82

83

By quantifying how parasitism affects methane emissions per unit of feed intake, we can obtain a
more complete understanding of the environmental costs of parasitism and the potential benefits
of mitigating emission through controlling parasite burden. Here, we address this aim by

87	evaluating methane emissions per unit of feed intake in parasitised and non-parasitised finishing
88	lambs, using respiration chambers.
89	

90

2. Materials and methods 91

The protocol was conducted under Home Office licence (PPL 60/4489) and was approved 92 by SRUC's Animal Experiment Committee (AE ED 24-2015). 93

94

95 2.1 Animals and experimental design

A total of 72 parasite naïve lambs (Suffolk x Mule), 12-15 weeks old were selected from a 96 commercial sheep flock. All animals were expected to be parasite naïve at the start of the trial, 97 as they were reared indoors and only fed commercial pelleted feed. Their parasite free status was 98 confirmed using faecal egg counts. The animals were divided into three treatment groups, 99

balanced for live weight (mean body weight at day $0 = 36.62 \text{kg} \pm 0.35 \text{ S.E.}$) and sex (mixed 100

101 pens). These treatments were: Ad lib fed control; restricted-fed control; and parasitised.

102

There were a total of eight replicates for each treatment, with each replicate comprised of one 103 pen of three lambs. There were three lambs in each pen to ensure adequate eructation for 104 methane detection. The lambs were housed in indoor pens in these groups of three for the 105 duration of the trial. The trial lasted for 39 days and animals were returned to stock at the end of 106 the trial. 107

108

110 2.2 Parasite challenge

111 The animals in the parasitised treatment were trickle challenged with 7,000 infective *Teladorsagia circumcincta* larvae suspended in 10ml of water, three times a week from days 0 to 112 113 35 (five weeks). T. circumcincta is an abomasal nematode which represents a substantial parasitic challenge to sheep, and is often linked with parasitic gastroenteritis in lambs (Coop et 114 al., 1982). The trickle infection was used to represent the challenge encountered by grazing 115 lambs, and was expected to result in subclinical infection consistent with rates of natural 116 infection on moderately parasitised pasture (Coop et al., 1982). The *ad lib* control, and 117 restricted-fed control treatments were sham infected with 10ml of water, following the same 118 protocol as the parasitised treatment. Parasite levels were monitored through weekly faecal 119 sampling for faecal egg counts (FEC), using the modified flotation method with a sensitivity of 1 120 121 egg per gram of faeces (epg)(Christie & Jackson, 1982). To give an indication of gut damage by *T.circumcinta*, pepsinogen levels were measured from blood samples taken at three points in the 122 trial. Blood samples were collected from all animals at day 0 (pre-challenge), day 36 (peak 123 124 challenge, prior to being placed in the respiration chambers), and day 39 (after removal from the respiration chambers). 125

126

127 2.3 Feeding

The *ad-lib* control and parasitised treatments were fed *ad-lib* access to pelleted grass. The restricted fed treatments were fed 80% of the intake of their *ad lib* fed counterparts, relative to body weight. Parasite induced anorexia was expected in the parasitised treatment, hence the restricted-fed control group enabled the assessment of the impact of parasitism *per se* versus that of anorexia associated with parasitism. All lambs were fed their rations once a day.

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134	2.4 Measurements
135	
136	2.4.1 Methane
137	From days 43 to 46 lambs were housed in indirect open-circuit respiration chambers (No
138	Pollution Industrial Systems Ltd., UK). The trial was conducted over four rounds, using six
139	respiration chambers, with treatments balanced across each round so that each treatment was
140	tested in two respiration chambers per round. The area of each chamber is $25.4m^2$ with penning
141	for three lambs. Temperature and humidity were set at $15 \pm 1^{\circ}C$ and $60 \pm 5\%$ respectively, and
142	air was removed from the chambers by exhaust fans set at 50litres/s. Methane concentration was
143	recorded for the air in each chamber once every six minutes, using infrared absorption
144	spectroscopy. Animals remained in the chambers for three full days (days 43 to 46), the first 24
145	hours were the adaptation period, and measurements taken during the final 48 hours (days 44-46)
146	were used to quantify methane production. Total feed intake in the chambers was measured
147	daily, and methane yield (g CH_4/kg DMI) was calculated by dividing daily methane production
148	by the daily DMI.
149	
150	2.4.2 Digestibility
151	The collection of faeces directly from the rectum of all lambs was carried out over three

152 consecutive days (days 30 to 32), pooled per lamb, and stored at -20°C prior to digestibility

- analysis. Acid insoluble ash (AIA) was used as an internal, indigestible marker to assess the
- apparent total tract digestive matter (DM). Faecal and feed AIA were analysed using the 2N HCl

procedure (Van Keulen & Young, 1977). During feeding, feed samples were collected daily andpooled for analysis.

157

- 158 *2.4.3 Feed intake and weight gain*
- 159 Pelleted feed intake was measured three times a week, to calculate the restricted-feeding
- allowances. Feed intake was also measured daily in each respiration chamber for calculation of
- 161 methane yield (gCH_4/kg DMI). All lambs were weighed weekly.

162

163 2.5 Statistical analysis

164 Data were analysed using ANOVA, with round as the block term. For statistics pertaining to

body weight, day 0 body weight was included as a covariate. Daily intake values are presented

166 per animal, by dividing the total pen intake by three. All statistical analyses were performed in

167 GENSTAT 16

168 **3. Results**

169 3.1 Development of the parasite challenge

170 Lamb FEC increased over time for the parasitised treatment, indicating that parasite infections 171 were achieved in the groups dosed with T. circumcincta, whilst the control and restricted-fed control treatments remained parasite free for the duration of the trial (Fig. 1a). Pepsinogen levels 172 were significantly higher in the parasitised group at the two sample points in the final week of 173 the trial (P<0.001) (Fig. 1b). These increased levels of blood pepsinogen confirmed abomasal 174 damage in the parasitised animals compared to the controls. The FEC and pepsinogen results 175 176 indicate that the parasitised treatment group did harbour helminth infection when in the respiration chambers, whilst the ad lib and restricted fed control groups did not. No clinical 177 signs of parasitism were observed in any groups throughout the experiment. 178

179

180 *3.2 Lamb performance*

Table 1 shows the variation in performance, feed intake, and digestibility, across the three 181 182 treatment groups. The DMI of the parasitised group was significantly lower than the ad lib group (P<0.001), indicative of parasite induced anorexia with their feed intake being an average 183 of approximately 80% of the *ad lib* control group over the study period. Maximum anorexia 184 was found in the final week of the study, where average daily DMI per animal in the parasitised 185 group was approximately 70% of the *ad lib* control group. The highest level of inappetance 186 coincided with the time of highest FEC. Average body weight gain was 174 g/day in control ad 187 lib fed individuals, whilst the average body weight gain for ad lib fed parasitised individuals was 188 7 g/day. 189

190

191 3.3 Methane output and yield

Methane output was significantly higher in the *ad lib* fed control group (P<0.001), than in the 192 other two treatments (Fig. 2). Whilst methane emissions remained relatively steady over time in 193 194 the two *ad lib* fed treatments (*ad lib* control and parasitised), in the restricted fed group the 195 emissions rose steadily shortly after feeding time, before reaching a peak and declining again. 196 197 Although total methane emissions were highest in the *ad lib* fed control group, Fig. 3 reveals that methane yield (g CH₄/kg DMI) was significantly higher in the parasitised group. Methane yield 198 was 33% higher in the parasitised lambs compared to the *ad lib* control group. Whilst there was 199 a significant difference in methane yield between the parasitised treatment group and both 200 control treatment groups (P<0.001), there was no significant difference in methane yield 201 202 between the *ad lib* fed control group and the restricted-fed control group despite a significant 203 difference in feed intake (Fig. 3).

204 4. Discussion

205

This study aimed to quantify the impact of parasitism on methane emissions in lambs. Our results show that methane yield was 33% higher in the parasitised lambs relative to the *ad lib* control group. This is the first study to demonstrate that infectious disease can increase methane yield (g CH_4/kg DMI).

210

The total quantity of methane produced per day was highest in the *ad lib* control group (Fig. 2). 211 212 This is because the primary driver of methane production (g CH_4/day) is dry-matter intake (DMI), with a strong positive correlation between methane emissions and DMI (Buddle et al., 213 2011). The *ad lib* control group had a significantly higher level of DMI (P<0.001), providing a 214 215 higher total supply of substrate for methane production in the rumen. Whilst in the respiration chambers the parasitised lambs consumed 70% of the feed quantity consumed by the ad lib 216 control group. This reduced intake was associated with 20% less methane production in the 217 218 parasitised animals. Snap shot measurements of methane output would therefore show parasitism being associated with a positive environmental impact. However, the methane yield (g CH₄/kg 219 220 DMI) was 33% higher in the parasitised animals compared to the ad-lib control group. The parasitised lambs also had significantly lower weight gain compared to the controls, and would 221 require a higher overall feed intake over their lifetime to reach target weight. Whilst worldwide 222 223 there is a mixture of sheep management practices i.e. intensively and extensively reared lambs and a variety of different nutritional environments, parasite induced anorexia is a phenomena 224 which occurs over all systems where livestock are at risk from gastrointestinal parasites 225 226 (Kyriazakis et al. 1998; Sutherland & Scott, 2010). Thus the combination of increased methane

227 yield, and higher feed intake per kg product demonstrated in this study has substantial

implications for the impacts of parasitism on emissions from meat production.

229

230 Low feed intake can be associated with increased methane yield, however, the methane yield from the parasitised animals was higher than would be expected based solely on their lower 231 DMI (Hammond et al., 2013). Additionally, despite a significant difference in DMI between the 232 ad lib and restricted fed control groups, there was no significant difference in methane yield 233 between these groups (Table 1 and Fig. 3). These findings suggest that parasitism has an impact 234 235 on methane yield beyond that expected from changes in DMI alone. The extent of bacterial fermentation is influenced by myriad elements of gastrointestinal physiology and digesta kinetics 236 (Moraes et al., 2014; Stergiadis et al., 2016). Gastrointestinal nematode infection in small 237 238 ruminants can lead to substantial changes in the digestive tract including increased cell turnover, changes in permeability, changes in pH, altered secretory activities (e.g. mucous production), 239 and inhibited gastric acid production (Li et al., 2016; Louie et al., 2007). Some of these parasite 240 241 induced changes in the gastrointestinal tract will disrupt the intricate interactions between hosts and their gut microbiome, as the large array of products secreted by gastrointestinal nematodes 242 impact on growth and metabolism of resident microbial communities (Zaiss & Harris, 2016). 243 However, we are only now beginning to understand the complexity of microbiota, and the effects 244 of parasitism on interactions between hosts and their gastrointestinal bacteria remain largely 245 unexplored (Buddle et al., 2011; Zaiss & Harris, 2016). Thus the effects of parasitism on 246 microbial survival, proliferation, spatial organisation, and ultimately rate of methanogenesis, are 247 248 yet to be understood. Whilst our results identify a novel phenomenon, they do not reveal the 249 mechanism.

In this study, weight gain was significantly lower in the parasitised group compared to that in 251 other groups. This highlights the substantial impact of parasitism on productivity, with 252 253 parasitised hosts needing to stay in the system much longer to reach slaughter weight. Attempts have previously been made to quantify the impacts of parasitism on emissions through exploring 254 the increased time on pasture, and increased DMI required to reach slaughter weight. Without 255 accounting for the effects of parasitism on emissions per kg DMI such studies will likely 256 underestimate the full influence of parasitism on methane production. The parasite driven 257 increase in methane yield demonstrated in this study, combined with the knowledge that 258 parasitism decreases production efficiency and increases time to achieve production targets 259 (Houdijk et al., 2016; Kenyon et al., 2013), demonstrates that parasitism has the potential to have 260 261 substantial impacts on livestock methane emissions. In addition to emissions increasing with parasitism is the concern that parasite intensity is projected to increase under climate change 262 (Fox et al., 2011, 2012, 2015). 263

264

The potential impact that parasitism has on livestock emissions makes it an attractive target for 265 mitigation. Parasite control practices (i.e. rearing indoors, clean grazing and refugia-based 266 control strategies), which break the parasite lifecycle, provide an opportunity to sustainably 267 reduce GHG emissions as it is cost effective, practical, and improves overall production 268 efficiency. As the increase in ovine meat production is expected to be highest in developing 269 270 countries (O'Mara, 2011), with restricted access to improved feeds, feed supplements and efficiency gains through genetic selection, parasite control offers a viable and accessible way of 271 272 reducing emissions.

274	This study shows that parasite infections in lambs can lead to a 33% increase in methane yield.
275	Combined with impacts of parasitism on production efficiency, and the subsequent increased
276	time on pasture, there is potential for parasitism to have an extensive impact on GHG emissions.
277	There are international commitments to reduce GHG emissions, and an informed understanding
278	of how production-limiting diseases affect GHG production is vital in developing public policies
279	and combating climate change. As we improve our understanding of how parasitism affects
280	livestock methane emissions we begin to elucidate the true environmental costs of parasitism,
281	and reveal the potential benefits of mitigating emission through controlling infectious disease.
282	
283	Acknowledgements
284 285 286 287	Thanks to John Rooke for advice on using the respiration chambers, Dave Anderson, Sandra Terry, Kate Hutchings, Laura Nicoll, Scott Grey, Claire Broadbent, Sokratis Ptochos, Emeric Desjeux, Justine Labbe for care and sampling of the animals, Lesley Deans and Shane Troy assistance with the respiration sampling.
288	This research was financially supported by the Scottish Government.
289	
290	Declarations of interest
291	None

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Figure legends

372

373	Figure 1. Indirect measures of parasitism across all treatment groups.
374	A) Mean faecal egg counts (FEC) (eggs/g faeces) by trial week (\pm SE), and B) mean pepsinogen
375	levels (\pm SE) at three time points, for all three treatment groups of lambs - Ad lib control (<i>ad lib</i>
376	fed), restricted fed control (fed 80% of feed intake of ad lib control, to account for parasite
377	induced anorexia), and parasitised lambs (also ad lib fed).
378	
379	
380	Figure 2. Daily mean methane output
381	Mean methane output in A) grams per hour (\pm SE), and B) grams per day (\pm SE), for Ad lib
382	control (ad lib fed), restricted fed control (fed 80% of feed intake of ad lib control, to account for
383	parasite induced anorexia), and parasitised lambs (also ad lib fed), averaged across individuals.
384	
385	
386	Figure 3. Mean Methane yield
387	Mean methane yield (grams of methane per kg of dry matter intake) (\pm SE) for Ad lib control (<i>ad</i>
388	lib fed), restricted fed control (fed 80% of feed intake of ad lib control, to account for parasite
389	induced anorexia), and parasitised lambs (also ad lib fed).
390	

392 **Table 1. Performance, feed intake and digestibility**

393 The mean body weight parameters, levels of feed intake, and digestibility values for *ad lib*

control lambs, restricted fed control lambs, and parasitised lambs, averaged across individuals.

395 Values in rows with different letter superscripts differed significantly (P<0.05).

	Treatments				
	<i>Ad lib</i> control	Restricted fed control	Parasitised	Standard error	P-value
	1 0 - 2	a- . b	ac ab		
Final BW (kg)	42.6 [°]	37.1°	38.2	0.8	< 0.001
BW gain (g/day) per animal	174 ^a	71 ^b	7 ^c	12.8	<0.001
Daily DMI over trial, per animal (g/day)	1783 ^a	1302 ^b	1396 [°]	28.3	<0.001
Daily DMI per kg BW over trial (g/kg BW/day)	44.6 ^a	34.4 ^b	37.0 ^c	0.98	<0.001
Digestibility Dry Matter (DM, %)	55.4	58.2	58.4	0.01	0.09



Figure 1.

Manuscript reference number: UPara18_061R1

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Figure 2.

Manuscript reference number: UPara18_061R1



Figure 3.

Manuscript reference number: UPara18_061R1