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1	Effect of the Texel muscling QTL (TM-QTL) on spine characteristics in purebred Texel
2	lambs
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24 Abstract

25

Previous work showed that the Texel muscling QTL (TM-QTL) results in pronounced 26 27 hypertrophy in the loin muscle, with the largest phenotypic effects observed in lambs inheriting a single copy of the allele from the sire. As the loin runs parallel to the spinal 28 vertebrae, and the development of muscle and bone are closely linked, the primary aim of this 29 study was to investigate if there were any subsequent associations between TM-QTL 30 inheritance and underlying spine characteristics (vertebrae number, VN; spine region length, 31 SPL; average length of individual vertebrae, VL) of the thoracic, lumbar, and thoracolumbar 32 spine regions. Spine characteristics were measured from x-ray computed tomography (CT) 33 34 scans for 142 purebred Texel lambs which had been previously genotyped. Least-squares 35 means were significantly different between genotype groups for lumbar and thoracic VN and lumbar SPL. Similarly for these traits, contrasts were shown to be significant for particular 36 modes of gene action but overall were inconclusive. In general, the results showed little 37 38 evidence that spine trait phenotypes were associated with differences in loin muscling associated with the different TM-QTL genotypes. 39

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41 Keywords:

42 Vertebrae, Spine, Texel, Loin, Muscling

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49 **1. Introduction**

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Walling et al. (2004) reported evidence of a quantitative trait locus (QTL) segregating in the 51 52 United Kingdom's Texel sheep population which significantly increased *longissimus dorsi* (loin) muscle depth (up to +1.15 - +2.00 mm, as measured ultrasonically over the third 53 lumbar vertebra). Observing similar results (QTL effect of +2.57 mm) from an analysis 54 including existing and new Texel family data, the QTL, later termed the Texel muscling QTL 55 (TM-QTL), was further verified by Matika et al. (2006). Located on the distal end of the 56 57 ovine chromosome 18 (OAR18) (Walling et al., 2004; Matika et al., 2006), the TM-QTL sits in the same region as the Callipyge (CLPG) and Carwell loci (Cockett et al., 1994; Nicoll et 58 al., 1998) which are also known to affect carcass muscling; the CLPG mutation leads to 59 60 greater muscle mass most pronounced in the hind quarters (loin, pelvis, leg) (Cockett et al., 1994; Koohmaraie et al., 1995; Jackson et al., 1997a, 1997b; Freking et al., 2002), while 61 carriers of Carwell exhibit a larger loin muscle area and weight (Nicoll et al., 1998). 62

Such QTL are of economic interest as there is the potential to utilise their effects through selection programmes to gain greater carcass value (e.g. reducing fat deposition and increasing lean meat production). In the case of the TM-QTL, the proportion of the high value loin cut may be increased e.g. two-dimensional measurements (estimated from crosssectional computed tomography (CT) scans, taken at the fifth lumbar vertebra) describing loin depth, width, and area were found to be ~ 0.5 - 11% greater in TM carrier lambs than non-carrier lambs (Macfarlane et al., 2010).

Moreover, taking the QTL's mode of inheritance into consideration allows the opportunity to exploit the TM-QTL more fully and appropriately in a commercial situation. Similar to *CLPG* (Cockett et al., 1994; Freking et al., 1998a), expression of the Texel muscling phenotype has been suggested to follow the complex parent-of-origin-dependent pattern of 74 inheritance termed polar overdominance (Macfarlane et al., 2010; Matika et al., 2011). This unique type of inheritance is characterised by the instance where heterozygous progeny that 75 inherit a single copy of the allele from the sire exhibit the superior phenotype (Cockett et al., 76 77 1996). Indeed, Macfarlane et al. (2010) observed that the largest phenotypic effects of the TM-QTL were particularly apparent in the TM carrier lambs that had inherited a single copy 78 of the TM allele from the sire and the wild type (+) from the dam (genotype $TM^{S/+D}$; where 79 superscripts S (sire) and D (dam) denote the paternal and maternal origin of the alleles, 80 respectively), with loin depth, width, and area measures ~ 2 - 11% greater in these TM^S/+^D 81 genotype lambs than in the other three genotype groups (homozygote non-carriers, $+^{S}/+^{D}$; 82 heterozygote carriers inheriting TM-QTL from the dam, +^S/TM^D; homozygote carriers, 83 TM^{S}/TM^{D}). 84

85 Essentially, muscle hypertrophy from TM allele segregation appears to be localised to the loin muscle (Macfarlane et al., 2010), which is found along the length of the thoracolumbar 86 (thoracic plus lumbar) spine region. Given that development of muscle and bone are closely 87 linked, it is of further interest to investigate, across genotype groups, if the increased loin 88 muscling is associated with any change to characteristics of the underlying spine section i.e. 89 is there a subsequent effect on spine characteristics in relation to the pattern of TM allele 90 inheritance? Freking et al. (1998b), for instance, found that the spinal column was 91 significantly shorter in CLPG genotype lambs (-2.5 cm; when all animals compared at the 92 same carcass weight) and the carcasses more compact in skeletal structure in comparison to 93 normal genotype lambs. Given its chromosomal position, it may be a similar condition for 94 the TM-QTL. This is a particularly relevant point to assess in terms of a possible 'trade-off' 95 between increasing loin muscle size (e.g. depth) but, in consequence, shortening the spinal 96 column. 97

98 The thoracolumbar spine region, on which the loin is located, encompasses the 'body' (or trunk) vertebrae and the total length of this region (as with any spine region) is a product of 99 the number and length of vertebrae which comprise it. Hence, the difference in body (and 100 101 carcass) lengths observed from individual to individual is contributed to the variation in these vertebral factors. Recent work has demonstrated that the spine characteristics (vertebrae 102 number, vertebrae length), of the thoracolumbar region, can be reliably measured from CT 103 scans. Using such method, it was also identified that these characteristics exhibit significant 104 intra-breed variation in Texel sheep, for example, thoracolumbar vertebrae number was 105 observed to range from 17 to 21 (Donaldson et al., 2013). Therefore, it may be reasonable to 106 use CT measured spine traits to investigate if any association exists between the pattern of 107 TM allele inheritance and spine characteristics. 108

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111 **2. Materials and Methods**

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The present study used a subset of the 209 available purebred Texel lamb records previously 115 used by Macfarlane et al. (2010) and Lambe et al. (2011). Lambs were sired by seven 116 different rams that were previously identified as carriers of at least one copy of TM-QTL; all 117 209 lambs were blood-sampled soon after birth (born 2009) in order to classify their TM-118 QTL genotype (homozygote non-carrier, $+^{S}/+^{D}$; heterozygote carrier inheriting TM-QTL 119 from the sire, $TM^{S/+D}$; heterozygote carrier inheriting TM-OTL from the dam, $+^{S/TM^{D}}$; 120 homozygote carrier, TM^S/TM^D); detailed information on the genotyping of the animals can 121 be found in Macfarlane et al. (2010). However, for a number of animals the genotype could 122

^{113 2.1.} Animals sampled

123 not be fully classified. These unknowns were excluded from this study's analysis, leaving a 124 total of 142 lamb records in the subset, which divided into the TM-QTL genotype groups as 125 follows: $39 + {}^{S}/{}^{+D}$, $52 \text{ TM}^{S}/{}^{+D}$, $17 + {}^{S}/{}^{TM}{}^{D}$, $34 \text{ TM}^{S}/{}^{TM}{}^{D}$. These 142 lamb records included 126 59 entire males and 83 female lambs from the purebred population of Texel sheep kept across 127 two sites, one in Scotland and one in Wales, which were reared as either singles (n = 97), 128 twins (n = 34), or artificially (pet; n = 11) (further details on the management of these 129 animals can be found in Macfarlane et al. (2010) and Lambe et al. (2011)).

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131 2.2. Computed tomography (CT) measurements

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Lambs were CT scanned at ~ 126 days of age (ranging from 93 to 145 days) and their 133 134 topogram images (produced from the CT process) used to quantify spine characteristics for each, details provided in Donaldson et al. (2013). In short, spine traits measured directly 135 from the scans included counts of vertebrae in the thoracic and lumbar regions (VN_{THOR} and 136 VN_{LUM} respectively) and length (mm) of the thoracic and lumbar spine region (SPL_{THOR} and 137 SPL_{LUM} respectively). These measures were used to calculate the average length (mm) of 138 individual vertebrae in the thoracic and lumbar regions (VL_{THOR} (SPL_{THOR}/VN_{THOR}) and 139 VL_{LUM} (SPL_{LUM}/VN_{LUM}) respectively). The results for the thoracic and lumbar spine 140 regions were further used to provide the number of thoracolumbar vertebrae (VN_{T+L}) 141 $(VN_{THOR}+VN_{LUM}))$, and the length (mm) of the thoracolumbar region (SPL_{T+L}) 142 (SPL_{THOR}+SPL_{LUM})). These thoracolumbar spine traits were then used to calculate the 143 average length (mm) of individual vertebrae across the thoracolumbar region (VL_{T+L} 144 145 $(SPL_{T+L}/VN_{T+L})).$

146 For each lamb, the dimensions, width (mm), depth (mm), and area (mm²), of the *longissimus*

147 *lumborum* (CT_MLL_W, CT_MLL_D, and CT_MLL_A respectively), were estimated (from

148 cross-sectional CT scans taken at the fifth lumbar vertebra) by Macfarlane et al. (2010) and 149 included in this study's analysis of the genotypic effect. Essentially, these traits were 150 included to, (i) determine if analysis of the reduced sample of animals shows genotype effects 151 on loin traits similar to that observed for the larger sample, and, (ii) assess, from further 152 analysis of the smaller data set, if the same or similar pattern of TM expression (polar 153 overdominance) can be considered as a source for any genotype differences.

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155 2.3. Statistical Analysis

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Data were analysed using the GLM procedure in SAS (SAS Institute Inc., Cary, NC, USA) to 157 determine the effects of genotype on the collated loin traits and measured spine traits. Fixed 158 effects fitted in the model for loin dimensions (CT_MLL_W, CT_MLL_D, CT_MLL_A), 159 spine length traits (SPL_{THOR}, SPL_{LUM}, SPL_{T+L}, VL_{THOR}, VL_{LUM}, VL_{T+L}) and spine count 160 traits (VN_{THOR}, VN_{LUM}, VN_{T+L}) were site, with two levels (Scotland and Wales), sex, with 161 two levels (male and female), rearing rank, with three levels (single, twin, or pet), and TM-162 QTL-genotype, with four levels $(+^{S}/+^{D}, TM^{S}/+^{D}, +^{S}/TM^{D})$ and TM^{S}/TM^{D}). The model was 163 run with and without covariate adjustments for live weight (LWT); where any of the traits 164 differed significantly between genotype groups, it was of interest to assess if, by testing the 165 groups at a standard live weight, the differences were removed. 166

A set of orthogonal contrasts, as described by Freking et al. (1998a), (additive (-1 0 0 1), dominance (-1 1 1 -1), and reciprocal heterozygote (0 1 -1 0)) was fitted to the $+^{S}/+^{D}$, TM^S/+^D, $+^{S}/TM^{D}$, TM^S/TM^D genotypes, respectively. The contrasts test for any distinct pattern in the differences amongst the genotype group's least-squares means (for loin and spine traits), from which, a particular model for TM gene action might be suggested. Due to the previous evidence supporting the expression of the TM muscling phenotype through a

173	polar overdominant mode of inheritance (Macfarlane et al., 2010), if significant differences
174	were found between the heterozygote groups (reciprocal heterozygote test) a further set of
175	orthogonal contrasts was fitted to the genotypes to include a test for the paternally derived
176	polar overdominant mode of inheritance. Again following that from Freking (1998a), this
177	second set of orthogonal contrasts included additive (-1 0 0 1), maternal dominance (-1 0 2 -
178	1), and polar overdominance (-1 3 -1 -1) models of gene action which were fitted to the
179	$+^{S}/+^{D}$, $TM^{S}/+^{D}$, $+^{S}/TM^{D}$, TM^{S}/TM^{D} genotypes, respectively. Contrasts were performed on
180	the spine count data which was not adjusted for LWT and on the loin muscle and spine length
181	data after the adjustment for LWT.
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184	3. Results
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185 186 187 188 189 190 191	In the context of this work it is useful to note that an earlier study (Macfarlane et al., 2012) found that least-squares means for LWT (measured at birth, 5, 10, 15, and 20 weeks of age) and carcass weight for TM^{S}/TM^{D} animals were consistently larger than that measured for $+^{S}/+^{D}$, $+^{S}/TM^{D}$, and $TM^{S}/+^{D}$ genotype lambs (these differences were significant between TM^{S}/TM^{D} and $+^{S}/+^{D}$ lambs for LWT at birth, 5, and 10 weeks of age, and carcass weight). In the present study, statistical models were first run without an adjustment of LWT but, given
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185 186 187 188 189 190 191 192 193	In the context of this work it is useful to note that an earlier study (Macfarlane et al., 2012) found that least-squares means for LWT (measured at birth, 5, 10, 15, and 20 weeks of age) and carcass weight for TM^S/TM^D animals were consistently larger than that measured for $+^{S}/+^{D}$, $+^{S}/TM^{D}$, and $TM^{S}/+^{D}$ genotype lambs (these differences were significant between TM^S/TM^D and $+^{S}/+^{D}$ lambs for LWT at birth, 5, and 10 weeks of age, and carcass weight). In the present study, statistical models were first run without an adjustment of LWT but, given the above, to remove, as far as possible, any misinterpretation of TM-QTL effects, statistical models were run again with certain traits (loin dimensions, spine length) adjusted for LWT.
185 186 187 188 189 190 191 192 193 194	In the context of this work it is useful to note that an earlier study (Macfarlane et al., 2012) found that least-squares means for LWT (measured at birth, 5, 10, 15, and 20 weeks of age) and carcass weight for TM^{S}/TM^{D} animals were consistently larger than that measured for $+^{S}/+^{D}$, $+^{S}/TM^{D}$, and $TM^{S}/+^{D}$ genotype lambs (these differences were significant between TM^{S}/TM^{D} and $+^{S}/+^{D}$ lambs for LWT at birth, 5, and 10 weeks of age, and carcass weight). In the present study, statistical models were first run without an adjustment of LWT but, given the above, to remove, as far as possible, any misinterpretation of TM-QTL effects, statistical models were run again with certain traits (loin dimensions, spine length) adjusted for LWT. The following sections discussing these traits will therefore focus only on the LWT adjusted
185 186 187 188 189 190 191 192 193 194 195	In the context of this work it is useful to note that an earlier study (Macfarlane et al., 2012) found that least-squares means for LWT (measured at birth, 5, 10, 15, and 20 weeks of age) and carcass weight for TM^S/TM^D animals were consistently larger than that measured for $+^S/+^D$, $+^S/TM^D$, and $TM^S/+^D$ genotype lambs (these differences were significant between TM^S/TM^D and $+^S/+^D$ lambs for LWT at birth, 5, and 10 weeks of age, and carcass weight). In the present study, statistical models were first run without an adjustment of LWT but, given the above, to remove, as far as possible, any misinterpretation of TM-QTL effects, statistical models were run again with certain traits (loin dimensions, spine length) adjusted for LWT. The following sections discussing these traits will therefore focus only on the LWT adjusted results.

Similar to the findings of Macfarlane et al. (2010), TM^S/+^D genotype lambs were observed to 199 generally have the largest loin width, depth, and area, on average (Table 1). The differences 200 in loin dimensions between $TM^{S/+D}$ and $+^{S/+D}$ genotype lambs were consistently significant, 201 however, the larger trait averages observed for the $TM^{S/+D}$ group were not all significantly 202 different from those averages observed for the $+^{S}/TM^{D}$ and TM^{S}/TM^{D} genotype groups. For 203 example, the $TM^{S/+D}$ group was significantly different from the $+^{S}/TM^{D}$ group in regards to 204 loin area (CT MLL A LWT) but the groups were not significantly different for loin width 205 and depth measures (CT_MLL_W_LWT and CT_MLL_D_LWT, respectively). Further to 206 this, and in contrast to Macfarlane et al. (2010), the $TM^{S}/+^{D}$ group in this smaller data set was 207 not significantly different from TM^S/TM^D in regards to all three loin dimension traits. 208 In general, the pattern of results from the analysis of the full data set (Macfarlane et al., 2010)

In general, the pattern of results from the analysis of the full data set (Macfarlane et al., 2010) suggested that the effect of the TM allele on these loin dimensions is expressed through a non-additive mode of inheritance (paternal polar overdominance). Analysis of the subset of records suggests a more general paternal TM-QTL effect on the loin with little evidence of a polar overdominance effect.

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215	3.2.	Spine	length	traits

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Overall, there was no significant effect of the TM-QTL on the thoracic region length traits (SPL_{THOR}, VL_{THOR}). Nor was there an effect of TM-QTL genotype on the average length of individual lumbar vertebrae (VL_{LUM}), however, associations were shown to exist between TM-QTL genotype groups and length of the lumber region (SPL_{LUM}) (Table 1). Leastsquares means showed that, on average, $+^{S}/+^{D}$ and TM^S/+^D genotype lambs had a longer lumbar length compared to $+^{S}/TM^{D}$ and TM^S/TM^D genotype lambs. However, when considering the combined length of the thoracic and lumbar regions (SPL_{T+L}), the genotype effect is negligible (Table 1).

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226 *3.3. Spine count traits*

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The segmentation and anatomical regionalisation of the spinal elements (vertebrae) is established in early development (Wellik, 2007; Iimura et al., 2009), hence, it should not be affected by varying LWT. In running the statistical model with the inclusion and omission of a covariate adjustment of LWT, little difference was found between the least-squares means for each model (Table 1), lending support to the previous statement. Therefore, only the results obtained from the model without LWT adjustment will be discussed (results from the model with LWT covariate adjustment are not shown).

Regarding vertebrae number in the separate thoracic and lumbar spine regions first (VN_{THOR} 235 and VN_{LUM} respectively), there were some significant differences between the genotype 236 groups, however, the magnitude of these differences was relatively small (Table 1). In more 237 detail, it can be seen from the least-squares means that there is much overlap between the 238 genotype classes with regards to VN_{THOR} . The +^S/TM^D and TM^S/TM^D genotype lambs had, 239 on average, a greater number of thoracic vertebrae than $+^{S}/+^{D}$ and $TM^{S}/+^{D}$ genotype lambs, 240 however $+^{S}/TM^{D}$ and $+^{S}/+^{D}$ genotype lambs were not significantly different from each other. 241 With regards to VN_{LUM}, the $+^{S}/+^{D}$ and TM^S/+^D genotype lambs were significantly different 242 from the $+^{S}/TM^{D}$ and TM^{S}/TM^{D} genotype lambs. While observed to possess fewer thoracic 243 vertebrae, $+^{S}/+^{D}$ and $TM^{S}/+^{D}$ genotype lambs had a greater number of lumbar vertebrae, on 244 245 average.

Although significant differences occurred between the genotype groups for the two spine regions when considered separately, when examining the results for the combined thoracic and lumbar vertebrae number (VN_{T+L}), there were no significant differences between the groups (Table 1).

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251 *3.4. Orthogonal contrasts*

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Previous work on loin dimensions had shown strong evidence that the mode of inheritance 253 for the TM-QTL deviates from a simple additive model (Macfarlane et al., 2010). Although 254 the results obtained in this study's subset of data did not fully provide the same results, there 255 256 was certainly an indication for superior loin dimensions in TM-QTL carrier lambs, especially in those with a paternal copy of the TM-QTL. Due to this, sets of orthogonal contrasts were 257 fitted to the genotypes to investigate the situation further. These contrasts allowed testing for 258 259 any particular patterns in the differences among the TM-QTL genotype (least-squares) means, for loin and spine traits, in order to define if certain modes of gene action may be 260 present. 261

The first set of orthogonal contrasts was fitted to the genotypes to test for additive, dominance, and reciprocal heterozygote models of gene action (Table 2; only traits where TM-QTL genotype had a significant effect are shown).

The additive inheritance model was fitted as -1 0 0 1 to the $+^{S}/+^{D}$, $TM^{S}/+^{D}$, $+^{S}/TM^{D}$, and 265 TM^S/TM^D genotypes respectively; testing the difference between the means of the 266 homozygote genotypes. Where the contrast value was positive this showed that TM^S/TM^D 267 had a larger mean than $+^{S}/+^{D}$ for that particular trait and vice versa if the contrast value was 268 negative. The difference between $+^{S}/+^{D}$ and TM^S/TM^D genotype means was significant for 269 all three loin traits CT_MLL_W_LWT, CT_MLL_D_LWT, and CT_MLL_A_LWT, and 270 spine traits, VN_{THOR}, VN_{LUM}, and SPL_{LUM}_LWT. The dominance inheritance model was 271 fitted as -1 1 1 -1 to the +^S/+^D, TM^S/+^D, +^S/TM^D, and TM^S/TM^D genotypes respectively; 272

testing the combined means of the heterozygote genotypes $(TM^{S}/+^{D}, +^{S}/TM^{D})$ with the 273 combined means of the homozygote genotypes $(+^{S}/+^{D}, TM^{S}/TM^{D})$. However, none of the 274 differences between genotype means were significant, providing no evidence of a dominance 275 276 effect on any of the traits. The reciprocal heterozygote model of gene action was fitted as 0.1 -1 0 to the +^S/+^D, TM^S/+^D, +^S/TM^D, and TM^S/TM^D genotypes respectively. This contrast 277 tested the difference between the means of the two heterozygote genotypes $(TM^S/+^D$ and 278 +^S/TM^D), which were significant for traits CT_MLL_A_LWT, VN_{THOR}, VN_{LUM}, and 279 SPL_{LUM}_LWT. 280

Freking et al. (1998a) previously commented that in such a case where the reciprocal 281 heterozygote contrast is shown to be significant, the dominance contrast may be misleading 282 i.e. under and over-estimation of heterozygote genotypes, and further analysis required. 283 284 Therefore, due to this, and with the previous observation of the TM allele's expression through a non-additive mode of inheritance, a further set of orthogonal contrasts, additive, 285 maternal dominance, and polar overdominance, were fitted to the genotypes as (1 0 0 -1) (-1 286 0 2 -1) (-1 3 -1 -1), respectively. Results for the additive model have been discussed above, 287 and with no significant results for a maternal dominance effect only the results for the polar 288 overdominance model from this set of contrasts were shown (Table 2) and discussed further. 289 The polar overdominance inheritance model was fitted as -1 3 -1 -1 to the $+^{S}/+^{D}$, $TM^{S}/+^{D}$, 290 +^S/TM^D, and TM^S/TM^D genotypes respectively and used to test the difference between the 291 mean of the $TM^{S/+D}$ group with each of the means calculated for $+^{S/+D}$, $+^{S/TM^{D}}$, and 292 TM^S/TM^D genotype groups. Contrast values for the paternal polar overdominance model are 293 the combined differences between genotype means (condition as defined above) and were 294 shown to be significant for all traits tested (Table 2); with the exception of VN_{THOR} , $TM^{S}/+^{D}$ 295 genotype lambs had a larger mean compared with each of the other genotype groups. 296

299 4. Discussion

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It should be noted that the data set used in the present report was limited in its size, largely due to the restricted availability of sires (identified as TM-QTL carriers) which could be used to produce a study group of lambs. Nonetheless, to date, it is the only available data set which provides detail of the TM-QTL status for a sufficient number of purebred Texel animals, from which, the effects of TM-QTL on carcass, meat quality, and production traits could be assessed.

The analysis in the present report made use of lamb records, where TM-QTL genotype was unambiguously known, to, (i) determine if similar conclusions for loin dimensions could be formulated using only a subset of data in the analysis, repeating, as close as possible, the model described by Macfarlane et al. (2010), (ii) extend this test to determine if there is an effect of TM-QTL on underlying spine characteristics as the loin muscle is located parallel to spinal vertebrae, and, (iii) fit sets of contrasts to the TM-QTL genotype groups in order to determine the inheritance pattern of the TM-QTL.

It should also be noted that the following discussion will continue to refer only to loin dimension and spine length trait results generated from the model where all lamb records were adjusted for LWT.

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318 *4.1. TM-QTL and loin dimensions*

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Regarding the loin dimensions, CT_MLL_W, CT_MLL_D, and CT_MLL_A, the leastsquares means for these traits reported by Macfarlane et al. (2010) are in strong agreement with an overdominance mode of expression of the TM allele; there is evidence of both heterozygote groups lying outside, in this case above, the phenotypic range of the homozygote groups. The results of Macfarlane et al. (2010) even suggested, more specifically, a paternally expressed polar overdominance effect as $TM^{S/+D}$ genotype lambs consistently exceeded $+^{S}/TM^{D}$ genotype lambs in trait means; the difference between heterozygote groups, however, only appeared to be significant for the loin area (CT_MLL_A).

From the present study, the polar overdominance test (Table 2) did show significance, but the 329 pattern of differences between the TM^S/+^D genotype least-squares means and the least-330 squares means for $+^{S}/+^{D}$, $+^{S}/TM^{D}$, and TM^{S}/TM^{D} genotype groups (Table 1) conflicts with 331 this outcome and could not support a polar overdominance mode of TM gene action i.e. trait 332 means for $TM^{S}/+^{D}$ genotype lambs did not appear to significantly 'out-perform' over all (or 333 the majority) of the other genotype groups (Table 1). Nonetheless, the pattern of least-334 squares means did still infer TM expression which could not be explained by simple additive 335 gene action, and though an overdominacne model could not be supported, there was still 336 indication towards some paternal influence of the TM allele; genotype groups which inherited 337 a copy of the TM allele from the sire $(TM^{S}/+^{D} \text{ and } TM^{S}/TM^{D})$ were observed to have, on 338 average, larger loin width (CT_MLL_W_LWT), depth (CT_MLL_D_LWT), and area 339 (CT_MLL_A_LWT) measures. 340

341

342 *4.2. TM-QTL and spine characteristics*

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Three out of the nine spine traits (VN_{THOR} , VN_{LUM} , and SPL_{LUM}) were observed to be significantly different amongst the TM-QTL genotype groups (Table 1).

346 The least-squares means for vertebrae number traits (VN_{THOR} , VN_{LUM}) followed a curious 347 pattern; means for +^S/+^D and TM^S/+^D genotype lambs were similar and significantly different,

in most instances, to the means for $+^{S}/TM^{D}$ and TM^{S}/TM^{D} genotype lambs, the latter of 348 which were also similar to each other in their mean values. The $+^{S}/+^{D}$ and TM^S/+^D groups 349 had, on average, fewer thoracic vertebrae (VN_{THOR}) but more lumbar vertebrae (VN_{LUM}), 350 with the situation reversed for the $+^{S}/TM^{D}$ and TM^{S}/TM^{D} groups. However, the overall 351 number of thoracolumbar vertebrae (VN_{T+L}) across the four genotype groups was not 352 different. It appeared that TM-QTL inheritance patterns had no substantial effect on the total 353 number of thoracolumbar vertebrae but may have some influence on the thoracic-lumbar 354 vertebral arrangement in the spine, but this remains uncertain as the size of difference 355 356 between genotype groups is small.

Similarly, the total length of the thoracolumbar region (SPL_{T+L}) was not drastically different 357 between the genotype groups. Freking et al. (1998b) observed shorter spinal columns in 358 359 CPLG genotype lambs, and given that TM-QTL falls close to its position on the chromosome, it was an important point to investigate further in connection with TM 360 inheritance. Least-squares means for spine length traits (SPL, VL) from the present study, 361 however, do not suggest any such negative effects of TM-OTL on spine length. The TM^S/+^D 362 genotype lambs, which express the muscle hypertrophy phenotype, in fact, were observed to 363 have, on average, longer thoracolumbar vertebrae (VL_{T+L}_LWT), and subsequently longer 364 thoracolumbar spine regions (SPL_{T+L}_LWT), but these trait values (23.22mm; 442.0mm 365 respectively) were not significantly different from the other three groups. It is interesting that 366 in the animals which were observed to have the largest loin dimension measures (TM^S/+^D), 367 there is not much of a change to the structure on which it lies. 368

Given that differences in spine traits are largely non-significant between the genotype groups, and that the overall pattern of least-squares means is indistinct, interpreting the models of gene action should be done with reservation. For example, the contrast tests showed significance for the polar overdominant model of TM gene action on VN_{THOR}, VN_{LUM}, and 373 SPL_{LUM} spine traits. These results should, again, be carefully considered alongside least-374 squares means (Table 1) as, though slightly larger (for VN_{LUM} and SPL_{LUM}), the means for 375 $TM^{S/+D}$ genotype lambs did not significantly 'out-perform' over all other genotype groups 376 for these spine traits. Hence, there was no strong indication that the observed differences in 377 spine trait phenotypes were associated with increased loin muscling specific to TM gene 378 action.

What is important to take from the present study is that increased loin muscling, particularly associated with $TM^{S/+D}$ genotype lambs, has been shown to have little associated effect on the underlying spine characteristics. Information on spine characteristics, in general, could potentially be used to improve loin production i.e. through increasing the size and/or number of loin chops (Donaldson et al., 2013). Hence, it would be interesting to investigate further the potential size of increase in loin production from those $TM^{S/+D}$ animals which posses a greater number of thoracolumbar vertebrae.

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387

388 **5.** Conclusion

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Given the results from the present study, it was evident that some effect of the TM allele on 390 loin dimension phenotypes was linked to a paternal genetic influence, but, with a weaker data 391 set (67 fewer records) this study could not provide further evidence for a specific polar 392 overdominance inheritance pattern. With regards to spine characteristics, in general terms, 393 the analysis of the subset of data did not reveal any obvious (advantageous or 394 disadvantageous) associations with TM-QTL inheritance. There did not appear to be any 395 effect on spine/vertebrae length and detailing how, or if, the TM allele interacts in the 396 vertebral patterning process (given the thoracic-lumbar vertebral combinations across 397

genotype groups) would require analysing a substantially larger data set than what was available at present.

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Reference List

Cockett, N.E., Jackson, S.P., Shay, T.L., Farnir, F., Berghmans, S., Snowder, G.D., Nielsen, D.M., Georges, M., 1996. Polar overdominance at the Ovine callipyge locus. Science 273, 236-238.

424 Cockett, N.E., Jackson, S.P., Shay, T.L., Nielsen, D., Moore, S.S., Steele, M.R., Barendse, W., Green, R.D., Georges, M., 1994. Chromosomal Localization of the Callipyge 425 Gene in Sheep (Ovis-Aries) Using Bovine Dna Markers. Proceedings of the National 426 Academy of Sciences of the United States of America 91, 3019-3023. 427 Donaldson, C.L., Lambe, N.R., Maltin, C.A., Knott, S., Bunger, L., 2013. Between- and 428 within-breed variations of spine characteristics in sheep. Journal of Animal Science 429 91, 995-1004. 430 Freking, B.A., Keele, J.W., Beattie, C.W., Kappes, S.M., Smith, T.P.L., Sonstegard, T.S., 431 Nielsen, M.K., Leymaster, K.A., 1998a. Evaluation of the ovine Callipyge locus: I. 432 Relative chromosomal position and gene action. Journal of Animal Science 76, 2062-433 2071. 434 Freking, B.A., Keele, J.W., Nielsen, M.K., Leymaster, K.A., 1998b. Evaluation of the ovine 435 Callipyge locus: II. Genotypic effects on growth, slaughter, and carcass traits. Journal 436 of Animal Science 76, 2549-2559. 437 Freking, B.A., Murphy, S.K., Wylie, A.A., Rhodes, S.J., Keele, J.W., Leymaster, K.A., Jirtle, 438 439 R.L., Smith, T.P.L., 2002. Identification of the single base change causing the callipyge muscle hypertrophy phenotype, the only known example of polar 440 overdominance in mammals. Genome Research 12, 1496-1506. 441 Iimura, T., Denans, N., Pourquie, O., 2009. Establishment of Hox Vertebral Identities in the 442 Embryonic Spine Precursors. Current Topics in Developmental Biology 88, 201-234. 443 Jackson, S.P., Miller, M.F., Green, R.D., 1997a. Phenotypic characterization of Rambouillet 444 sheep expressing the Callipyge gene .2. Carcass characteristics and retail yield. 445 Journal of Animal Science 75, 125-132. 446 447 Jackson, S.P., Miller, M.F., Green, R.D., 1997b. Phenotypic characterization of Rambouillet sheep expressing the Callipyge gene .3. Muscle weights and muscle weight 448 distribution. Journal of Animal Science 75, 133-138. 449 Koohmaraie, M., Shackelford, S.D., Wheeler, T.L., Lonergan, S.M., Doumit, M.E., 1995. A 450 451 muscle hypertrophy condition in lamb (callipyge): Characterization of effects on muscle growth and meat quality traits. Journal of Animal Science 73, 3596-3607. 452 Lambe, N.R., Richardson, R.I., Macfarlane, J.M., Nevison, I., Haresign, W., Matika, O., 453 Bunger, L., 2011. Genotypic effects of the Texel Muscling QTL (TM-QTL) on meat 454 455 quality in purebred Texel lambs. Meat Science 89, 125-132. Macfarlane, J.M., Lambe, N.R., Haresign, W., Bunger, L., 2012. The effect of the Texel 456 Muscling QTL on live and carcass weight in Texel lambs. Small Ruminant Research 457 105, 117-121. 458 459 Macfarlane, J.M., Lambe, N.R., Matika, O., Mclean, K.A., Masri, A.Y., Wolf, B.T., Haresign, W., Bishop, S.C., Bunger, L., 2010. Texel loin muscling QTL (TM-QTL) 460 located on ovine chromosome 18 appears to exhibit imprinting and polar 461 overdominance. Proceedings of the 9th World Congress on Genetics Applied to 462 Livestock Production, Leipzig, Germany, August 1-6, 2010 comm. no. 199. 463

- Matika, O., Pong-Wong, R., Woolliams, J.A., Low, J., Nieuwhof, G.J., Boon, S., Bishop,
 S.C., 2006. Verifying quantitative trait loci for muscle depth in commercial terminal
 sire sheep. Proceedings of the 8th World Congress on Genetics Applied to Livestock
 Production, Belo Horizonte, Brazil, August 13-18, 2006, 22-10.
- Matika, O., Sechi, S., Pong-Wong, R., Houston, R.D., Clop, A., Woolliams, J.A., Bishop,
 S.C., 2011. Characterization of OAR1 and OAR18 QTL associated with muscle depth
 in British commercial terminal sire sheep. Animal Genetics 42, 172-180.
- Nicoll, G.B., Burkin, H.R., Broad, T.E., Jopson, N.B., Greer, G.J., Bain, W.E., Wright, C.S.,
 Dodds, K.G., Fennessy, P.F., Mcewan, J.C., 1998. Genetic linkage of microsatellite
 markers to the Carwell locus for rib-eye muscling in sheep. Proceedings of the 6th
 World Congress on Genetics Applied to Livestock Production, Armidale, Australia,
 January 11-16, 1998 26, 529-532.
- Walling, G.A., Visscher, P.M., Wilson, A.D., Mcteir, B.L., Simm, G., Bishop, S.C., 2004.
 Mapping of quantitative trait loci for growth and carcass traits in commercial sheep
 populations. Journal of Animal Science 82, 2234-2245.
- Wellik, D.M., 2007. Hox patterning of the vertebrate axial skeleton. Developmental
 Dynamics 236, 2454-2463.
- 481

482 **Table 1.** Least-squares means (and standard errors) for live weight, loin dimensions¹ and spine traits² for Texel lambs of each TM-QTL

483 genotype³

Genotype														
Trait	+ 8/	/+ ^D	TM	1 ^s /+ ^D	+ ^s /7	^D	TM ^S	/TM ^D			Rearing	TM-	Livo	
	n =	39*	n =	= 52*	n =	17	n =	34	Site	Sex	rank	QTL	weight	R^2
	п	57	"	52		17		51			Tunn	genotype	weight	
Live Weight	30.53	(1.004)	31.78	(0.855)	30.59	(1.402)	31.45	(1.094)	<.001	0.104	<.001	0.672	•	0.253
	66 97 b	(1.02.4)	CO 108	(0.001)	(7.10a.b	(1 4 4 4)	<0.0 0 1	(1.107)	0.004	0.070	0 100	0.024		0.100
CI_MLL_W	66.27°	(1.034)	69.49	(0.881)	67.12 ^{a,a}	(1.444)	68.82	(1.127)	0.084	0.069	0.199	0.034	•	0.126
CT_MLL_D	28.45°	(0.779)	30.96°	(0.663)	29.28 ^{a,b}	(1.087)	30.44 ^ª	(0.848)	0.234	0.411	0.001	0.029	•	0.163
CT_MLL_A	1684 [°]	(64.94)	1883ª	(55.33)	1689 ^{0,c}	(90.68)	1851 ^{a,b}	(70.76)	0.385	0.633	<.001	0.021	•	0.157
CT_MLL_W_LWT	66.27 [°]	(0.591)	68.43 ^a	(0.507)	$67.06^{a,b}$	(0.825)	68.04^{a}	(0.646)	<.001	0.400	0.059	0.001	<.001	0.716
CT_MLL_D_LWT	28.45 ^b	(0.560)	30.28^{a}	(0.481)	$29.25^{a,b}$	(0.782)	29.94 ^a	(0.612)	0.007	0.008	0.148	0.029	<.001	0.570
CT_MLL_A_LWT	1684 ^c	(37.68)	1817^{a}	(32.36)	1685 ^{b,c}	(52.63)	1803 ^{a,b}	(41.17)	<.001	0.002	0.353	0.004	<.001	0.718
VN _{THOR}	12.69 ^{b,c}	(0.064)	12.65 ^c	(0.055)	$12.89^{a,b}$	(0.090)	12.88^{a}	(0.070)	0.026	0.350	0.044	0.006		0.174
VN _{LUM}	6.356 ^a	(0.074)	6.387^{a}	(0.063)	6.111 ^b	(0.104)	6.143 ^b	(0.081)	0.092	0.427	0.061	0.009		0.149
VN _{T+L}	19.05	(0.063)	19.04	(0.054)	19.00	(0.088)	19.02	(0.069)	0.759	0.981	0.645	0.967		0.011
SPL _{THOR}	255.6	(3.941)	260.7	(3.358)	257.9	(5.504)	262.5	(4.295)	0.003	0.429	0.034	0.503		0.122
SPL_{LUM}	$184.4^{a,b}$	(2.411)	186.8 ^a	(2.054)	176.6 ^c	(3.366)	180.3 ^{b,c}	(2.627)	0.208	0.742	0.029	0.018		0.125
SPL_{T+L}	440.0	(5.138)	447.5	(4.378)	434.5	(7.176)	442.8	(5.599)	0.004	0.447	0.015	0.329		0.128
VL _{THOR}	20.13	(0.276)	20.60	(0.235)	20.00	(0.386)	20.38	(0.301)	0.009	0.246	0.007	0.352		0.133
VL _{LUM}	29.03	(0.292)	29.29	(0.249)	28.91	(0.408)	29.36	(0.319)	<.001	0.702	0.045	0.657		0.124
VL _{T+L}	23.10	(0.265)	23.51	(0.226)	22.86	(0.370)	23.28	(0.289)	0.004	0.443	0.007	0.332		0.135
SPL _{THOR} _LWT	255.6	(2.495)	256.9	(2.143)	257.7	(3.485)	259.7	(2.726)	0.379	0.456	0.088	0.610	<.001	0.650
SPL _{LUM} _LWT	184.4^{a}	(2.033)	185.2 ^a	(1.746)	176.5 ^b	(2.839)	179.1 ^b	(2.221)	0.151	0.513	0.333	0.007	<.001	0.382
$SPL_{T+L}LWT$	440.0	(2.720)	442.0	(2.336)	434.2	(3.798)	438.8	(2.971)	0.061	0.241	0.575	0.289	<.001	0.758
VL _{THOR} _LWT	20.13	(0.160)	20.32	(0.138)	19.99	(0.224)	20.18	(0.175)	0.051	0.772	0.595	0.531	<.001	0.710
VL _{LUM} _LWT	29.03	(0.147)	28.97	(0.126)	28.89	(0.205)	29.13	(0.160)	0.350	<.001	0.011	0.721	<.001	0.781
VL _{T+L} LWT	23.10	(0.130)	23.22	(0.112)	22.85	(0.182)	23.07	(0.142)	0.023	0.185	0.488	0.297	<.001	0.793

484 a^{-c} Within a row, means with common letters, or no letters, in their superscript are not significantly different (P > 0.05)

485 LWT = live weight (kg) fitted as a covariate in model

486 ¹ CT_MLL_W = width of *M. longissimus lumborum* (mm); CT_MLL_D = depth of *M. longissimus lumborum* (mm); CT_MLL_A = area of *M. longissimus lumborum* (mm²)

487	2 VN _{THOR} = number of thoracic vertebrae; VN _{LUM} = number of lumbar vertebrae; VN _{T+L} = number of thoracolumbar vertebrae; SPL _{THOR} = length of thoracic spine region
488	(mm); SPL_{LUM} = length of lumbar spine region (mm); SPL_{T+L} = length of thoracolumbar spine region (mm); VL_{THOR} = average length of individual thoracic vertebrae (mm);
489	VL_{LUM} = average length of individual lumbar vertebrae (mm); VL_{T+L} = average length of individual thoracolumbar vertebrae (mm)
490	$^{3} + ^{S} + ^{D} =$ homozygote non-carrier; $TM^{S} + ^{D} =$ heterozygote carrier inheriting TM-QTL from sire; $+ ^{S} / TM^{D} =$ heterozygote carrier inheriting TM-QTL from dam; $TM^{S} / TM^{D} =$
491	homozygote carrier
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	Additive					;	Recip	orocal hetero	zygote	Polar overdominance			
Trait	Cor	ntrast	P- value	Cor	ntrast	P- value	Cor	ntrast	P- value	Con	ıtrast	P- value	
CT_MLL_W_LWT	1.767	(0.736)	0.017	1.184	(1.151)	0.305	1.368	(0.893)	0.128	3.919	(1.698)	0.023	
CT_MLL_D_LWT	1.496	(0.697)	0.034	1.142	(1.090)	0.297	1.032	(0.847)	0.225	3.207	(1.609)	0.048	
CT_MLL_A_LWT	118.9	(46.92)	0.013	15.90	(73.38)	0.829	131.5	(56.98)	0.023	279.0	(108.3)	0.011	
VN _{THOR}	0.186	(0.080)	0.022	-0.032	(0.126)	0.800	-0.243	(0.097)	0.014	-0.518	(0.185)	0.006	
VN_{LUM}	-0.213	(0.092)	0.022	-0.001	(0.144)	0.994	0.275	(0.112)	0.015	0.550	(0.212)	0.012	
SPL _{LUM} _LWT	-5.321	(2.531)	0.037	-1.857	(3.958)	0.640	8.675	(3.073)	0.006	15.49	(5.840)	0.009	
First set of orthogona	al contrasts	was fitted a	s -1 0 0 1(add	litive), -1 1 1	-1(dominan	ce), and 0 1 -	1 0(reciprocal	heterozygot	e) to the $+^{S}/+^{1}$	$^{\rm D}, {\rm TM}^{\rm S}/{\rm +}^{\rm D}, {\rm +}^{\rm S}$	$/TM^{D}$, and	TM ^S /TM ^D	

504 '	Table 2.	Estimates of	TM-Q) TL	genotype contrasts	(and	l standaro	l errors)) and	significance	levels	s (P	-value)) foi	r additive,	dominance,	reciproca
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505	heterozygote, and paternally of	lerived polar overdominant e	effects on loin dimensions ¹	and spine traits ^{2}
505	neterozygote, and paternany (ciived polar overdollillant c		and spine dates

genotypes, respectively. Second set of orthogonal contrasts was fitted as -1 0 0 1(additive), -1 0 2 -1(maternal dominance), and -1 3 -1 -1(polar overdominance) to the +^S/+^D,

 $TM^{S}/+^{D}$, $+^{S}/TM^{D}$, and TM^{S}/TM^{D} genotypes, respectively. From the second set of contrasts, only the results from the polar overdominance test are shown; additive results

previously reported with first set of contrasts and the maternal dominance test was not significant for any of the traits.

LWT = live weight (kg) fitted as a covariate in model

¹ CT_MLL_W = width of *M. longissimus lumborum* (mm); CT_MLL_D = depth of *M. longissimus lumborum* (mm); CT_MLL_A = area of *M. longissimus lumborum* (mm²)

 2 VN_{THOR} = number of thoracic vertebrae; VN_{LUM} = number of lumbar vertebrae; VN_{T+L} = number of thoracolumbar vertebrae; SPL_{THOR} = length of thoracic spine region

(mm); SPL_{LUM} = length of lumbar spine region (mm); SPL_{T+L} = length of thoracolumbar spin region (mm); VL_{THOR} = average length of individual thoracic vertebrae (mm);

 VL_{LUM} = average length of individual lumbar vertebrae (mm); VL_{T+L} = average length of individual thoracolumbar vertebrae (mm)