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Scotland's Rural College

# The interplay of dietary nutrient level and varying calcium to phosphorus ratios on efficacy of bacterial phytase: 2. Ileal and total tract nutrient utilization

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1	DIET FACTORS AND PHYTASE EFFECT ON NUTRIENTS
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4	The interplay of dietary nutrient level and varying Ca to phosphorus ratios on efficacy of a
5	bacterial phytase: 2. Ileal and total tract nutrient utilization
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10	ABSTRACT A 14-d broiler experiment was conducted to assess the effects of two
11	dietary variables on efficacy of a bacterial 6-phytase expressed in Aspergillus oryzae on nutrient
12	and phytate phosphorus (PP) utilization. Diets were formulated with or without nutrient matrix
13	values (matrix) for phytase as negative control (NC) or positive control (PC), respectively and
14	with two Ca:tP levels (2:1 or 2.5:1). The diets were supplemented with 0, 1,000 or 2,000 FYT/kg
15	phytase thus producing a $2 \times 2 \times 3$ factorial arrangement. Excreta were collected on d 19 to 21 and
16	ileal digesta on d 21. There was no three-way interaction on digestibility of any nutrient. There
17	was matrix $\times$ phytase (P < 0.01) interaction for Ca and DM digestibility and Ca:tP $\times$ phytase
18	interaction (P < 0.05) for acid hydrolyzed fat, Ca and P digestibility. Pre-cecal flow of Mn, Zn
19	and Na was greater (P < 0.05) in NC diets whereas phytase increased (P < 0.05) pre-cecal flow
20	of Mg, Fe, Mn, and Zn but decreased ( $P < 0.05$ ) pre-cecal Na flow. Total tract PP disappearance
21	and total tract Ca retention increased ( $P < 0.05$ ) with phytase supplementation in diets with 2:1
22	Ca:tP whereas there was no effect of phytase supplementation on PP disappearance or Ca
23	retention in diets with 2.5:1 Ca:tP. Total P and Ca retention were reduced ( $P < 0.05$ ) in PC and
24	NC diets when Ca:tP increased to 2.5:1 but the depression was more pronounced in the NC diet.
25	In addition, PP disappearance decreased (P < $0.05$ ) with increasing Ca:tP in the PC diets but
26	there was no effect of widening Ca:tP on PP disappearance in NC diets. It was concluded from
27	the current study that the effect of phytase supplementation on P utilization is reduced when diets
28	contain adequate P as exemplified in the PC diets and that the negative impact of wide Ca:tP is
29	more pronounced in diets with phytase matrix allowance as exemplified in the NC diets.

31 Key words: broilers, calcium:phosphorus, nutrient utilization, phytase matrix

### **INTRODUCTION**

34 The use of phytase in non-ruminant diets and the effects of different nutritional variables on the efficacy of phytase have been well studied in recent years (Adeola and Cowieson, 2011). 35 However, there is continued interest in understanding the various factors that mitigate the 36 37 efficacy of phytase or that may improve its effect in poultry diets because of the sheer amount of phytate present in typical poultry diets. A typical corn-soybean meal diet for poultry formulated 38 using conventional (i.e. non low-phytate varieties) may contain up to 4 g/kg phytate-P (Selle and 39 40 Ravindran, 2007) which are largely unavailable to birds without the action of phytase (endogenous and exogenous). Liberation of 60% of the P tied up in phytate will be a 41 considerable saving in terms of reducing both the cost for inorganic P supplementation and 42 environmental impact of P excretion. Therefore it is imperative to understand factors that may 43 hinder or enhance the efficacy of phytase. 44

The negative effect of wide Ca:P on phytase efficacy is well known (Tamim et al., 2004; 45 Adeola et al., 2006) and this is related to the formation of recalcitrant calcium-phytate (Taylor, 46 1965, Nelson and Kirby, 1987) or Ca-phosphate complexes (Long et al., 1984). In addition it is 47 common practice to reduce dietary levels of inorganic P, Ca, Na, energy and some digestible 48 49 amino acids (phytase matrix) in phytase-supplemented diets (Shelton et al., 2004). It would seem that supplementation of phytase to diets that already meet birds' requirements for these minerals 50 51 can be both wasteful and counterproductive. However, supplementation of phytase at high levels 52 (Shirley and Edwards, 2003; Cowieson et al., 2006) or to diets that already meet nutrient requirements of broilers has produced improvement in animal performance presumably via 53 mitigation of anti-nutritive effects of phytate rather than supply of limiting nutrients (Walk et al., 54 2013). 55

The current study examines the interplay of the variation in Ca:tP (tp, total P) and dietary 56 nutrient levels on efficacy of phytase added at low and high doses. There have been considerable 57 amount of investigations on the former and much less on the latter. Therefore, the objective of 58 the current experiment was to investigate how the use of a nutrient replacement values for 59 phytase (phytase matrix) affects phytase efficacy on nutrient utilization (with particular focus on 60 utilization of Ca, tP and phytate P), and especially within the context of variable dietary Ca:tP. 61 62 The companion article considers how these dietary factors influence growth performance and bone mineralization in broilers. 63

64

# **MATERIALS AND METHODS**

65 All the animal experimentation procedures used in the current study were approved by 66 the Scotland's Rural College's Animal Experimentation Committee.

## 67 Diets and experimental design

A total of 576 birds were used for the 14-d experiment to study the influence of nutrient 68 specification and Ca:tP on efficacy of phytase on nutrients and minerals utilization in broilers. 69 The birds were brooded together in a floor pen for the first 7 days of age during which they 70 received a standard diet that meets NRC (1994) nutrient requirement for broilers. On day 7, the 71 birds were weighed and allocated to 12 dietary treatments in a randomized complete block 72 design and a  $2 \times 2 \times 3$  factorial arrangement of treatments. Each treatment had 8 replicate cages 73 and 6 birds per replicate cage. The factors were two levels of nutrient specifications (explained 74 below), two levels of Ca:tP (2:1 and 2.5:1) and three levels of phytase supplementation (0, 1,000 75 and 2,000 FYT/kg). Excreta were collected on d 19 to 21 of the birds' age and ileal digesta were 76 77 collected on d 21 after euthanasia of the birds.

The composition of the experimental diets is presented in Table 1. Nutrient specification was used to define the diets that were formulated to meet all the nutrient requirements for broilers (full nutrient specification without phytase matrix or positive control, **PC**) and another

set of diets with reduced nutrient specification formulated to be deficient in P, Ca, crude protein 81 (CP), amino acids, and energy (down specification or negative control, NC). The nutrients and 82 energy levels in the NC diets were reduced relative to the PC diets on the basis of the amount of 83 nutrients and energy that the phytase was expected to release (nutrient matrix values for 84 phytase). The matrix values used per kg feed for 1,000 FYT were approximately, 75 kcal ME, 85 1.5 g for available P, 1.8 g for Ca, 0.26 g for CP, 0.11, 0.07, 0.04, and 0.07 g for digestible 86 lysine, total sulphur amino acids, methionine, and threonine, respectively. One phytase (FYT) 87 unit is defined as the activity that releases 1 µmol inorganic phosphate from 5.0 mM phytate per 88 minute at pH 5.5 and 37°C. 89

# 90 Chemical analysis

Diets, ileal digesta and excreta were analyzed for dry matter, N, gross energy, Ti, and 91 minerals. Dry matter was determined by drying the samples in a drying oven (Uniterm, Russel-92 93 Lindsey Engineering Ltd., Birmingham, England, UK) at 105°C for 24 hours (AOAC Method 934.01; AOAC, 2006). Total N content was determined by the combustion method (Method 94 968.06; AOAC, 2006). Gross energy was determined in an adiabatic bomb calorimeter (Model 95 6200, Parr Instruments, Moline, IL) using benzoic acid as an internal standard. Titanium 96 97 concentration in the samples was determined using the method of Short et al. (1996). Minerals content was determined using Inductively Coupled Plasma - Optical Emission Spectroscopy 98 (AOAC Method 990.08; AOAC, 2006) following digestion, in turn, in concentrated HNO<sub>3</sub> and 99 HCl. Free fat was determined using extraction by petroleum ether in a Soxhlet apparatus for six 100 101 hours whereas acid hydrolyzed fat (AHF) was determined by acid hydrolysis using 30% HCl followed by ether extraction. 102

#### 103 Statistical analysis

104 The data were analyzed by the MIXED procedure of SAS as appropriate for a 105 randomized complete block design and a factorial treatment arrangement. For ease of reference, the two types of control diets (NC and PC) were coded as matrix (i.e. nutrient matrix for phytase) 106 107 in the factorial arrangement with PC (as diets without phytase matrix) and NC (as diets with phytase matrix). The three-way interactions were investigated first in the analysis. Where the 3-108 way interactions were not significant they were dropped from the model and the data re-109 analyzed. Non-significant interactions were dropped for more thorough investigation of the main 110 effects means. Because the two-way interactions were significant for most of the responses even 111 112 though the three-way interactions were not, the simple effects means are presented in the tables. Because of the hierarchical arrangement of main effects and interactions, only the interactions 113 are discussed for responses in which all the two-way interactions are significant, whereas main 114 effects means are also discussed in cases where one or more of the two-way interactions are not 115 significant. 116

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# RESULTS

The analyzed nutrients compositions of the experimental diets are shown in Table 2 and show that expected nutrient compositions were met despite some slightly higher recoveries of the phytase.

The data on ileal nutrient digestibility response to the dietary treatments are presented in Table 3. There were no three-way interaction effects on digestibility of any of the nutrients. There were matrix × Ca:tP (P < 0.05) interaction for DM and AHF digestibility explained by lower (P < 0.01) DM and AHF digestibility in the NC diets with narrow Ca:tP whereas there was no effect of Ca:tP on DM and AHF digestibility in the PC diets. There was also matrix × phytase (P < 0.01) interaction for Ca and DM digestibility with lower (P < 0.01) DM and Ca 127 digestibility (drastic reduction observed for Ca digestibility) in phytase-supplemented NC diets whereas such effect was not observed in the PC diets. Ca:tP  $\times$  phytase interaction was observed 128 (P < 0.05) for AHF, Ca and P digestibility with phytase at 2,000 FYT/kg increasing (P < 0.05)129 130 AHF digestibility in the diets with narrow Ca:tP whereas phytase supplementation had no effect on AHF digestibility in diets with wide Ca:tP. The Ca:tP  $\times$  phytase interaction for Ca 131 digestibility was characterized by drastic and stepwise reduction (P < 0.05) in Ca digestibility 132 with increasing phytase supplemental level in the diets with wide Ca:tP but a less drastic 133 reduction in Ca digestibility at 1,000 FYT/kg in diets with narrow Ca:tP. For P digestibility, the 134 135 Ca:tP  $\times$  phytase interaction was manifested in reduced (P < 0.05) P digestibility at 1.000 FYT/kg and an increase (P < 0.05) at 2,000 FYT/kg in diets with narrow Ca:tP but a reduction (P < 0.05) 136 in P digestibility at both 1,000 and 2,000 FYT/kg in diets with wide Ca:tP. 137

The data on pre-cecal flow of micro-minerals in response to the dietary treatments are 138 139 presented in Table 4. Pre-cecal flow of Na, Mn and Zn was greater (P < 0.05) in NC diets; in addition pre-cecal flow of K and M n was greater (P < 0.05) in diets with wide Ca:tP. On the 140 other hand, phytase supplementation increased (P < 0.05) pre-cecal flow of Mg, Fe, Mn, and Zn, 141 decreased (P < 0.01) flow of Na and had no effect on K flow. There were significant Ca:tP  $\times$ 142 phytase interactions (P < 0.05) for pre-cecal flow of Mg and Mn with a decrease in the pre-cecal 143 flow of the minerals with phytase supplementation in diets with narrow Ca:tP. On the other hand 144 there was an increase in pre-cecal flow of the minerals with phytase supplementation in diets 145 with wide Ca:tP. Ca:tP  $\times$  matrix interaction was significant (P < 0.01) for pre-cecal flow of Na 146 and Mn. Generally pre-cecal flow of Na and Mn was greater (P < 0.01) in PC diets with wide 147 Ca:tP whereas Ca:tP had no effect on flow of the minerals in NC diets. Matrix  $\times$  phytase 148 interaction was observed (P<0.01) for pre-cecal Na and K flow. Phytase supplementation 149

decreased (P < 0.05) pre-cecal Na flow but increased (P < 0.05) K flow in NC diets however phytase supplementation had no effect on pre-cecal Na flow but decreased (P < 0.05) pre-cecal K flow in PC diets.

The effects of the treatments on total tract nutrient retention are shown in Table 5. There 153 154 were Ca:tP  $\times$  phytase interaction (P < 0.05) for total tract retention of DM, fat, AHF, and N, as well as AME. Dry matter and N retention as well as AME increased (P < 0.05) with phytase 155 supplementation in diets with narrow Ca:tP but DM retention decreased (P < 0.05) whereas 156 AME, N and fat retention were unaffected by phytase supplementation in diets with wide Ca:tP. 157 The interaction of Ca:tP  $\times$  matrix was significant (P < 0.05) for retention of DM, AHF, N, and 158 AME. In PC diets, retention of DM, AHF and N decreased (P < 0.05) whereas AME increased (P 159 < 0.05) with widening of Ca:tP. In NC diets, AHF and N retention increased (P < 0.05) whereas 160 there was no change in DM retention and AME with widening of Ca:tP. 161

The effect of the dietary treatments on total tract retention of Ca, P and PP are shown in 162 163 Table 6. Widening Ca:tP to 2.5:1 decreased (P < 0.05) total tract retention of Ca and tP as well as PP disappearance. Ca:tP  $\times$  phytase interaction was significant (P < 0.05) for total tract retention 164 of Ca, tP and PP. In the diets with narrow Ca:tP, only 2,000 FYT phytase increased (P < 0.05) tP 165 retention. In the diets with wide Ca:tP, phytase supplementation at 1,000 FYT/kg improved tP 166 retention. Total tract PP disappearance and Ca retention increased (P < 0.05) with phytase 167 supplementation in diets with narrow Ca:tP but no effect was observed in diets with wide Ca:tP. 168 There was significant Ca:tP  $\times$  matrix (P < 0.05) on total tract retention of tP and Ca as well as PP 169 disappearance. Total P and Ca retention were reduced (P<0.05) in both NC and PC with 170 171 widening of Ca:tP to 2.5:1 but the depression in P and Ca retention due to widening of Ca:tP was more pronounced in the NC diets. In addition, PP disappearance decreased (P<0.05) with 172

widening Ca:tP in the PC diets but there was no effect of widening Ca:tP on PP disappearance in the NC diets. Matrix  $\times$  phytase was significant (P < 0.05) only for total tract P retention with phytase supplementation increasing P retention only at 2,000 FYT/kg in PC diets and only at 1,000 FYT/kg in NC diets.

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# DISCUSSION

There is a preponderance of information on the effects of phytase on nutrient utilization 178 (Selle and Ravindran, 2007; Adeola and Cowieson, 2011) as well as the effect of Ca:P on 179 phytase efficacy (Qian et al, 1997; Selle et al., 2009). In addition, it is a usual practice to reduce 180 nutrient specification in phytase-supplemented diets or provide a nutrient matrix values for 181 phytase (Shelton et al., 2004; Silversides and Hruby, 2009). Therefore the objective of the 182 183 current experiment was to study the interactivity of varying Ca:tP in PC and NC diets on the efficacy of phytase at low and high doses in promoting nutrient utilization in broilers. Although 184 the effects of the treatments in the current experiment were observed on energy and a large 185 186 number of nutrients, the main responses that will subsequently be focused on are phytate and total P as well as Ca in view of their association with phytic acid. 187

# 188 Effects of use of nutrient matrix for phytase on phytase efficacy

The use of a phytase matrix in phytase-supplemented feed enables a reduction in nutrient specification, reduces nutrient excretion and increases the chance of being able to observe phytase effects. Two lines of evidence are presented here to show that the use of a phytase matrix differently affects phytase effects on PP and tP.

At the ileal level, the efficacy of phytase in promoting PP disappearance was the same in both PC and NC diets but at the total tract level, phytase supplementation of PC diet marginally reduced PP disappearance whereas phytase supplementation increased PP disappearance in the NC diets. The PP level was virtually the same across all diets (average of 0.22%) and hence the lower PP disappearance in PC diet without phytase suggests that the higher dietary non-phytate P (**nPP**) level (providing greater quantities of readily available P) in the diet may provide a feedback mechanism inhibiting the degradation of phytic acid. It is not clear if such a mechanism exists, however others have similarly observed reduced PP disappearance in diets with high levels of nPP (Ballam et al. 1982; Olukosi et al., 2013).

Ravindran et al. (2000) observed that hydrolysis of PP increases with an increase in 202 203 dietary PP level. This is intuitive, up to a point, as higher PP provides more substrate for phytase. But it is also of interest to consider how PP hydrolysis is affected by nPP levels in diets with the 204 205 same content of PP. In the current study, ileal PP disappearance was similar in both PC and NC diets supplemented with 2,000 FYT/kg even though ileal PP disappearance in the diets without 206 207 phytase was five percentage units greater in the NC diet. This shows that the effect of phytase on 208 PP disappearance, relative to the control, was greater in the PC diet. The observation that PP disappearance was the same in the diets supplemented with 2,000 FYT/kg in both PC and NC 209 diets indicates that effect of phytase supplementation on PP disappearance did not depend on 210 dietary level of nPP as also observed by Plumstead et al. (2008). 211

Phytate P disappearance at the total tract level in response to phytase supplementation was greater in both PC and NC diets compared with the disappearance at the ileal level. However the difference in PP disappearance at both levels in diets supplemented with 2,000 FYT/kg was greater for NC diet (11 %) compared with PC diet (5%). Phytase did not improve total tract PP disappearance in the PC diet but improved PP disappearance in the NC diet. Increased PP disappearance in the excreta compared with the ileal level is an indication of either that the 218 phytase continued to be effective post-ileal or of the possible effects of microorganisms on 219 phytic acid hydrolysis. Overall the observation on PP disappearance and P utilization indicate 220 that reducing the level of nPP is beneficial in promoting greater PP and total P utilization.

221 Phytase supplementation did not increase ileal P digestibility in both the NC and PC diets 222 but increased total tract P retention by 11% in the NC and had no effect in the PC diets. The numerical increase of 4.3 percentage units for total tract P retention in phytase-supplemented PC 223 diet decreased retained P by 190 mg/kg. On the other hand, phytase supplementation of NC diet 224 increased P retention by 11 percentage units and increased retained P by 830 mg/kg. In spite of 225 226 the greater retained P in phytase-supplemented NC compared with PC diet, the total retained P at 2,000 FYT/kg was 3.74 and 3.62 g/kg for PC and NC diets, respectively. The hydrolyzed PP at 227 228 the total tract level for PC and NC diets supplemented with 2,000 FYT/kg were 1.66 and 1.84 g/kg, respectively. Taken together therefore, the data imply that the high nPP content of the PC 229 230 diets "hinders" phytase from exerting its full effect on phytate. Although the total retained P in 231 PC diet supplemented with 2,000 FYT/kg phytase was greater than retained P in comparable NC diet, this extra retained P could have resulted from the higher dietary P in the PC diet because the 232 amount of PP hydrolyzed was actually lower in the phytase-supplemented PC diet. 233

# 234 The interplay of Ca:tP and dietary nutrient levels (phytase matrix)

The use of a phytase matrix in diet formulation enables a reduction in nutrient content of phytase-supplemented diets. This may be a necessary dietary intervention in phytasesupplemented diets in order to optimize phytase effect and maximize reduction in nutrient excretion (Shelton et al., 2004; Silversides et al., 2009). In the current study, at both the ileal and total tract levels, widening Ca:tP decreased P and Ca digestibility in both PC and NC diets but

the decrease produced by the wider Ca:tP was more pronounced in NC diets and the depression
in Ca utilization due to wide Ca:tP was greater at the total tract level. The decreased digestibility
values were also reflected in decreased digestible and retained Ca and P in the diets in response
to widening the Ca:tP. Two scenarios emerging from these observations are: 1) a decrease in Ca
and P utilization with a widening of Ca:tP irrespective of whether it was in PC or NC diet and, 2)
a more pronounced negative effect of widening Ca:tP in NC diets.

In the first scenario, the decreased Ca utilization with increased Ca:tP can be associated 246 with increased relative dietary concentration of Ca in the diets with wide Ca:tP because the 247 analyzed Ca in these diets was 27% higher than in diets with narrow Ca:tP. This greater dietary 248 249 Ca content produced correspondingly higher Ca intake and hence reduced Ca retained as a 250 percentage of intake. Similar observations have been reported in rats (Hoek et al., 1988), pigs (Qian et al., 1996) and chickens (Qian et al., 1997). Thus it seems that the decreased Ca 251 252 utilization in diets with wide Ca:tP can be largely explained by the presence of an abundance of Ca in the intestine, than can be utilized by the birds, leading to excessive Ca excretion or reduced 253 efficiency of Ca absorption. 254

The decrease in P utilization in the diets with wide Ca:tP ratio is also primarily driven by 255 dietary Ca content because tP content was similar in diets with wide and narrow Ca:tP. Hoek et 256 al. (1988) similarly observed high P excretion in rats receiving diets with high Ca level. This 257 reduced P utilization in diet with wide Ca:tP can be explained by the fact that high concentration 258 of Ca relative to P increases the possibility for negative interaction of Ca and P, leading to 259 greater chances for formation of calcium phosphate (Hurwitz and Bar, 1971). Al-Masri (1995) 260 261 observed that P digestibility, absorption and endogenous excretion in chickens decreased with increasing Ca:P ratio. Similar effect has been reported by Edwards and Veltmann (1983) and 262

Qian et al. (1997). Clearly, an increase in Ca:tP increases the concentration of Ca relative to Pand hence increases the chances of more Ca being chemically bound and becoming indigestible.

It has been suggested that another way by which high Ca:P reduces P and Ca utilization is 265 by the formation of recalcitrant Ca-phytate complex (Wise, 1983; Maenz et al., 1999). The effect 266 267 of Ca:tP on PP disappearance was not consistently observed in the current study. It was only at 268 the total tract level that high Ca:tP decreased PP disappearance in the PC diet. The analyzed PP was the same in all diets in the current experiment and the only differences among diets were Ca 269 and P levels. In addition, the depressed PP disappearance observed in the current study was not 270 dependent on Ca:tP per se but rather on dietary concentration of both Ca and P, i.e. the diet with 271 272 high contents of both Ca and P had depressed PP disappearance.

273 For the second scenario, it is possible that the reason for the decrease in Ca and P utilization in NC diets with wide Ca:tP relative to similar diets in PC diets was due to the lower 274 Ca and P contents of the NC diets compared with PC diets. Phytate P made up a greater 275 276 proportion of total P in NC compared with the PC (additional P in the PC diet was supplied by dicalcium phosphate) and hence the P will be less digestible in NC than the more readily 277 digestible P in the inorganic P sources used in the PC diet. Consequently the current data show 278 that the dietary content of Ca and P, not just the ratio, need to be considered in interpretation of 279 280 the effect of Ca:tP.

In light of the observations in the current experiment, it can be concluded that the effects of wide Ca:tP are more likely to be severe in diets in which nutrient matrix for phytase is used (as exemplified by the NC diets in this experiment) especially as it relates to Ca utilization; and

that the negative effect of high Ca:tP on P and Ca utilization could be mediated via mechanismsindependent of phytic acid degradation.

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# REFERENCES

Adeola, O., and A. J. Cowieson. 2011. Opportunities and challenges in using exogenous
enzymes to improve nonruminant animal production. J. Anim. Sci 89: 3189-3218.

Adeola, O., O. A. Olukosi, J. A. Jendza, R. N. Dilger, and M. R. Bedford. 2006. Response of
 growing pigs to *Peniophora lycii-* and *Escherichia coli-*derived phytases or varying ratios
 of calcium to total phosphorus. Anim. Sci. 82: 637-644.

Al-Masri, M. R. 1995. Absorption and endogenous excretion of phosphorus in growing broiler
chicks, as influenced by calcium and phosphorus ratios in feed. Br. J. Nutr. 74: 407-415.

AOAC. 2006. Official methods of analysis. 18<sup>th</sup> ed. Assoc. Off. Anal. Chem. Washington, DC.

Ballam, G. C., T. S. Nelson and L. K. Kirby. 1982. Effect of fiber and phytate source and of
calcium and phosphorus level on phytate hydrolysis in the chick. Poult. Sci. 63:333-338.

Cowieson, A. J., T. Acamovic, and M. R. Bedford. 2006. Supplementation of corn-soy-based
 diets with an *Eschericia coli*-derived phytase: effects of broiler chick performance and the

- digestiblity of amino acids and metabolizability of minerals and energy. Poult. Sci. 85:
  1389-1397.
- Edwards Jr., H. M., and J. R. Veltmann, 1983. The role of calcium and phosphorus in the
  etiology of tibial dyschondroplasia in young chicks. J. Nutr. 113: 1568-1575.
- Hoek, A. C., A. G. Lemmens, J. W. M. A. Mullink, and A. C. Beynen. 1988. Influence of dietary
  calcium:phosphorus ratio on mineral excrtion and nephrocalcinosis in female rats. J. Nutr.
  118: 1210-1216.
- Hurwitz, S., and A. Bar. 1971. Calcium and phosphorus interrelationships in the intestine of the
  fowl. J. Nutr. 101: 677-686.
- Long, P. H., S. R. Lee, G. N. Rowland, and W. M. Britton. 1984. Experimental rickets in broilers: gross, microscopic, and radiographic lesions. I. phosphorus deficiency and calcium excess. Avian Dis. 28: 460-474
- Maenz, D. D., C. M. Engele-Schaan, R. W. Newkirk, and H. L. Classen. 1999. The effect of
  minerals and mineral chelators on the formation of phytase-resistant and phytasesusceptible forms of phytic acid in solution and in a slurry of canola meal. Anim. Feed Sci.
  Techonol. 81: 177-192.
- National Research Council. 1994. Nutrient Requirements of Poultry. 9th rev. ed. Natl. Acad.
  Press, Washington, DC.
- Nelson, T. S., and L. K. Kirby. 1987. The calcium binding properties of natural phytate in chick
  diets. Nutr. Rep. Int. 35:949–956.

324	Olukosi, O. A., C. Kong, F. Fru-Nji, K. M. Ajuwon, and O, Adeola. 2013. Assessment of a
325	bacterial 6-phytase in the diets of broiler chickens. Poult. Sci. 92: 2101-2108
326	Plumstead P. W., A. B. Leytem, R. O. Maguire, J. W. Spears, P. Kwanyuen, and J. Brake. 2008.
327	Interaction of calcium and phytate in broiler diets. 1. Effects on apparent prececal
328	digestibility and retention of phosphorus. Poult. Sci. 87:449–458.
329	Qian, H., E. T. Kornegay, and D. E. Conner Jr. 1996. Adverse effects of wide
330	calcium:phosphorus ratios on supplemental phytase efficacy for weanling pigs fed two
331	dietary phosphorus levels. J. Anim. Sci. 74:1288-1297.
332	Qian, H., E. T. Kornegay, and D. M. Denbow. 1997. Utilization of phytate phosphorus and
333	calcium as influenced by microbial phytase, cholecalciferol, and the calcium:total
334	phosphorus ratio in broiler diets. Poult. Sci. 76: 37-46.
335	Ravindran, V., S. Cabahug , G. Ravindran, P.H. Selle, W. L. Bryden. 2000. Response of broiler
336	chickens to microbial phytase supplementation as influenced by dietary phytic acid and
337	non-phytate phosphorous levels. II. Effects on apparent metabolisable energy, nutrient
338	digestibility and nutrient retention. Br. Poult. Sci. 41:193-200.
339	Selle, P. H., A. J. Cowieson, and V. Ravindran. 2009. Consequences of calcium interactions with
340	phytate and phytase for poultry and pigs, Livest. Sci. 124: 126-141
341	Selle, P. H., and V. Ravindran. 2007. Microbial phytase in poultry nutrition. Anim. Feed Sci.
342	Technol. 135: 1-41.

Shelton, J. L., L. L. Southern, L. A. Gaston, and A. Foster. 2004. Evaluation of the nutrient
matrix values for phytase in broilers. J. Appl. Poult. Res. 13:213–221.

- Shirley, R. B., and H. M. Edwards Jr. 2003. Graded levels of phytase past industry standards
  improves broiler performance. Poult. Sci. 82: 671-680.
- Short, F. J., P. Gorton, J. Wiseman, and K. N. Boorman. 1996. Determination of titanium dioxide
  added as an inert marker in chicken digestibility studies. Anim. Feed Sci. Technol. 59:
  215-221.
- Silversides, F. G., and M. Hruby. 2009. Feed formulation using phytase in laying hen diets. J
  Appl. Poult. Res.18:15-22
- Tamim, N. M., R. Angel, and M. Christman. 2004. Influence of dietary calcium and phytase on
  phytate phosphorus hydrolysis in broiler chickens. Poult. Sci. 83: 1358-1367.
- Taylor, T. G. 1965. The availability of the calcium and phosphorus of plant materials for
  animals. Proc. Nutr. Soc. 24:105–112.
- Walk, C. L., M. R. Bedford, T. S. Santos, D. Paiva, J. R. Bradley, H. Wladecki, C. Honaker, and
  A. P. McElroy. 2013. Extra-phosphoric effects of superdoses of a novel microbial phytase.
  Poult. Sci. 92: 719-725.
- Wise, A. 1983. Dietary factors determining the biological activities of phytate. Nutr. Abstr. Rev.
  53:791-806.

Basal diet	1	2	3	4
Ca:tP <sup>1</sup>	2	:1	2.5	5:1
Control (phytase matrix)	Positive	Negative	Positive	Negative
Corn	482.6	477.4	466.6	499.4
Wheat	-	50.0	-	-
Soybean meal	397.5	382.5	400.5	394.5
Soybean oil	58.0	40.0	60.0	45.0
Corn Starch	15.0	15.0	15.0	15.0
Dicalcium phosphate	17.5	9.0	17.5	10.0
Limestone	17.0	15.5	28.0	24.0
Titanium dioxide	0.5	0.5	0.5	0.5
L-Lysine HCl	1.0	0.4	1.0	0.7
DL-Methionine	2.8	1.9	2.8	2.8
Threonine	0.6	0.3	0.6	0.6
Vitamin-mineral premix <sup>2</sup>	2.5	2.5	2.5	2.5
Salt	5.0	5.0	5.0	5.0
Phytase premix <sup>3</sup>	To 1,000	To 1,000	To 1,000	To 1,000
Total	1,000	1,000	1,000	1,000
Calculated nutrients and energy,	%			
Metabolizable energy, kcal/kg	3,185	3,125	3,154	3,127
Crude protein	22.9	22.8	22.9	22.9
Total P	0.71	0.56	0.70	0.57
Non-phytate P	0.45	0.30	0.45	0.31
Са	1.06	0.82	1.43	1.13

**Table 1.** Ingredient composition (g/kg) of the experimental basal diets

# <sup>1</sup>Ca:tP based on analyzed chemical composition

<sup>2</sup>Supplied the following per kilogram of diet: vitamin A, 5,484 IU; vitamin D3, 2,643 ICU;
vitamin E, 11 IU; menadione sodium bisulfite, 4.38 mg;riboflavin, 5.49 mg; d-pantothenic
acid, 11 mg; niacin, 44.1 mg; choline chloride, 771 mg; vitamin B12, 13.2 µg; biotin, 55.2
µg; thiamine mononitrate,2.2 mg; folic acid, 990 µg; pyridoxine hydrochloride, 3.3 mg; I,
1.11 mg; Mn, 66.06 mg; Cu, 4.44 mg; Fe, 44.1 mg; Zn, 44.1 mg; Se, 300 µg.

<sup>3</sup>Phytase premix containing 100 phytase units (FYT)/g replaced corn starch to provide
 1,000 or 2,000 FYT/kg.

Diets	1	2	3	4	5	6	7	8	9	10	11	12		
Ca:tP				2:1			2.5:1							
Matrix	P	ositive Cor	ntrol	Ne	gative Con	trol	Po	ositive Cont	rol	Negative Control				
GE, kcal/kg	4,452	4,681	4,652	4,672	4,619	4,603	5,720	5,782	4,610	4,641	4,572	4,580		
Ether extract	10.2	10.3	8.80	7.11	7.83	6.84	11.4	11.3	8.94	7.67	8.26	7.95		
Acid hydrolysed fat	11.1	9.49	9.80	8.03	7.96	8.03	12.1	12.9	9.70	8.30	9.37	8.37		
Ν	3.99	3.86	4.13	3.84	3.90	3.80	5.11	4.70	4.15	4.04	3.73	3.88		
Ca	1.37	1.63	1.40	1.31	1.18	1.17	2.23	2.24	1.87	1.06	1.47	1.38		
Р	0.73	0.81	0.71	0.65	0.61	0.57	0.93	0.91	0.74	0.48	0.60	0.62		
Phytate P	0.26	0.24	0.23	0.25	0.23	0.24	0.28	0.28	0.29	0.24	0.23	0.29		
non-phytate P <sup>1</sup>	0.47	0.57	0.48	0.40	0.38	0.33	0.65	0.63	0.45	0.24	0.37	0.33		
Na	0.23	0.26	0.24	0.26	0.25	0.23	0.24	0.28	0.23	0.17	0.23	0.19		
Mg	0.18	0.18	0.16	0.18	0.18	0.16	0.23	0.22	0.18	0.15	0.18	0.19		
Cu, mg	11.3	15.8	14.7	12.5	10.2	10.2	28.2	18.2	12.4	10.2	13.6	15.8		
Fe, mg	82.4	87.0	79.2	90.7	78.5	71.3	111.3	106.5	83.3	56.6	81.4	79.0		
Mn, mg	81.3	94.9	87.1	100.9	84.2	83.8	107.1	105.1	83.3	61.1	83.7	86.9		
Zn, mg	79.1	96	99.5	92.9	84.2	81.5	101.4	99.5	83.3	58.9	75.7	79.0		
K	1.16	1.14	1.00	1.16	1.19	0.96	1.47	1.43	1.14	0.97	1.15	1.28		
Phytase, FTY/kg <sup>2</sup>	BD	1,121	2,977	BD	1,406	2,400	BD	1,717	2,537	BD	1,011	2,640		

**Table 2.** Analyzed nutrient composition (%, dry matter basis) and phytase activity in the experimental diets

<sup>1</sup>non-phytate phosphorus level was determined by difference (total P – phytate P)

<sup>2</sup>BD – below detection limit

Diet	1	2	3	4	5	6	7	8	9	10	11	12		P-values for interactions <sup>2,3</sup>					
Ca:tP			2	:1					2.5	:1			- SEM						
$M^3$	Pos	itive Cor	ntrol	Neg	ative Co	ntrol	Pos	itive Con	trol	Neg	ative Co	ntrol	SEN	$Ca:tP \times M$	$Ca:tP \times Ph$	$M \times Ph \\$			
$Ph^3$	0	1000	2000	0	1000	2000	0	1000	2000	0	1000	2000							
DM	69.9	67.8	68.6	66.1	62.7	68.5	66.2	65.4	65.4	68.2	66.3	63.1	0.897	0.003	0.635	0.001			
$EE^3$	89.5	88.3	87.2	83.2	83.6	83.0	87.2	88.0	86.7	85.5	86.2	85.0	1.12	0.058	0.794	0.821			
AHF <sup>3</sup>	82.0	79.6	82.4	76.4	74.4	78.0	81.3	81.8	81.0	78.1	81.9	78.4	1.31	0.043	0.003	0.768			
Ν	76.9	73.8	77.1	73.9	69.9	75.0	73.5	71.4	74.8	73.8	70.3	70.0	1.12	0.636	0.156	0.447			
Ca	42.8	32.5	35.9	61.7	45.9	44.9	34.0	31.9	25.8	53.5	34.8	25.9	2.56	0.062	0.029	0.001			
Р	43.8	38.7	47.2	48.5	37.9	46.9	34.0	32.3	33.0	37.6	31.4	30.2	1.97	0.358	0.005	0.071			
PP <sup>3</sup>	48.5	42.7	58.7	57.7	43.3	65.4	36.0	44.1	58.6	39.3	51.1	51.7	6.03	0.493	0.088	0.784			

**Table 3.** Simple effects means for ileal nutrient digestibility response to varying levels of phytase supplementation, dietary Ca:total P broiler diets with or without nutrient matrix values for phytase<sup>1</sup>

<sup>1</sup>Means were obtained from 8 replicate cages of 6 birds per replicate cage

<sup>2</sup>The main effect for Ca:tP was significant (P < 0.05) for all nutrients except EE and PP; The main effect for nutrient matrix was significant for all nutrients except P and PP; The main effect for phytase was significant for all nutrients except EE and AHF; The three-way interaction was not significant for any nutrient

 ${}^{3}M$  = nutrient matrix for phytase; Ph = phytase; EE = crude fat; AHF = acid hydrolysed fat; PP = phytate phosphorus

Diet	1	2	3	4	5	6	7	8	9	10	11	12		P-values	P-values for interactions <sup>2, 3</sup>			
Ca:tP			2	:1					2.5	5:1			SEM					
$M^3$	Pos	itive Co	ntrol	Nega	ative Co	ntrol	Positive Control			Negative Control			SEM	$Ca:tP \times Ph$	$Ca:tP \times M$	$M \times Ph$		
Ph <sup>3</sup>	0	1000	2000	0	1000	2000	0	1000	2000	0	1000	2000						
Na	0.271	0.235	0.232	0.409	0.346	0.292	0.257	0.242	0.260	0.362	0.290	0.294	0.017	0.119	0.032	0.008		
Mg	0.155	0.166	0.158	0.168	0.190	0.157	0.164	0.178	0.171	0.156	0.171	0.186	0.006	0.003	0.096	0.620		
Fe, mg	65.8	73.6	74.5	64.7	70.9	86.9	73.3	75.3	85.5	71.7	78.0	86.6	7.55	0.965	0.804	0.717		
Mn, mg	88.0	92.7	84.5	94.0	99.4	89.4	92.6	96.9	94.2	91.6	91.9	101.0	2.20	< 0.001	0.028	0.309		
Zn, mg	76.7	85.2	77.1	84.2	89.0	83.1	80.6	86.6	81.9	79.8	87.7	88.7	3.45	0.448	0.418	0.676		
Κ	0.158	0.153	0.146	0.141	0.158	0.142	0.191	0.149	0.169	0.161	0.174	0.172	0.023	0.152	0.747	0.006		

**Table 4.** Simple effects means for pre-cecal flow (g/100 g dry matter intake) of micro-minerals in response to varying levels of phytase supplementation, dietary Ca:total P broiler diets with or without nutrient matrix values for phytase<sup>1</sup>

<sup>1</sup>Means were obtained from 8 replicate cages of 6 birds per replicate cage

<sup>2</sup>Phytase matrix effect only significant (P < 0.05) for Na, Mn and Zn; Ca:tP effect only significant for Mn and K; Phytase effect significant (P < 0.05) for all except K; The three-way interaction was not significant for any nutrient

 ${}^{3}M$  = nutrient matrix for phytase; Ph =phytase

Diet	1	2	3	4	5	6	7	8	9	10	11	12		P-value	s for inter	cactions <sup>2, 3</sup>
Ca:tP	2:1 2.5:1													Carth	Cert	
$M^3$	Pos	sitive Con	ve Control Neg			ntrol	Positive Control			Neg	Negative Control			Ca:tP	Ca:tP	$\mathbf{M}\times\mathbf{P}\mathbf{h}$
Ph <sup>3</sup>	0	1000	2000	0	1000	2000	0	1000	2000	0	1000	2000		× Ph	×M	
DM	71.0	68.8	71.4	67.5	66.8	70.2	69.1	67.5	66.2	68.8	70.1	67.2	0.581	0.001	0.020	0.001
AME	3,321	3,557	3,535	3,516	3,469	3,458	3,515	3,540	3,519	3,514	3,453	3,455	5.81	0.001	0.001	0.001
EE	91.7	90.4	90.0	86.7	86.5	86.4	90.6	88.7	89.7	89.0	90.3	88.4	0.636	0.001	0.063	0.876
AHF	89.5	84.4	87.2	83.4	81.8	83.3	86.1	85.0	84.9	85.9	87.8	83.5	0.818	0.001	0.014	0.001
Ν	63.7	57.7	65.2	53.0	54.8	58.3	62.8	58.2	60.0	56.8	59.0	57.3	1.07	0.001	0.001	0.001

**Table 5.** Simple effects means for total tract nutrient retention response to varying levels of phytase supplementation, dietary Ca:total P broiler diets with or without nutrient matrix values for phytase<sup>1</sup>

<sup>1</sup>Means were obtained from 8 replicate cages of 6 birds per replicate cage

<sup>2</sup>Matrix (M) effect was significant for all nutrients except DM; Ca:tP effect was significant for all nutrients except AHF and N;

Phytase (Ph) effect was significant for all nutrients except DM and AME; P-values for three-way interaction was not significant for any of the nutrients

 ${}^{3}M$  = nutrient matrix for phytase; Ph = phytase; EE = crude fat; AHF = acid hydrolysed fat

Diet	1	2	3	4	5	6	7	8	9	10	11	12		P-values for interactions <sup>2, 3</sup>					
Ca:tP			2:	1	-	CartDar	Cert	M											
$M^3$	Pos	itive Cor	ntrol	Neg	ative C	ontrol	Pos	itive Co	ntrol	Neg	Negative Control			Ca:tP ×		M×			
Ph <sup>3</sup>	0	1000	2000	0	1000	2000	0	1000	2000	0	1000	2000		Ph	Μ	Ph			
Р	52.1	49.6	56.8	56.1	61.1	69.1	42.4	46.5	46.5	42.5	54.6	52.5	1.32	< 0.001	0.004	<.0001			
PP <sup>3</sup>	68.4	68.6	68.6	63.6	60.7	71.8	64.7	62.7	58.5	62.9	66.8	67.2	2.41	0.018	0.019	0.096			
Ca	39.9	36.5	42.1	42.3	38.3	46.4	25.2	22.1	22.0	17.8	25.2	16.1	1.59	< 0.001	0.002	0.067			

**Table 6.** Simple effects means for total tract retention response of calcium, total and phytate phosphorus to varying levels of phytase supplementation, dietary Ca:total P broiler diets with or without nutrient matrix values for phytase<sup>1</sup>

<sup>1</sup>Means were obtained from 8 replicate cages of 6 birds per replicate cage

<sup>2</sup>Phytase matrix effect only significant for P; Ca:tP effect significant for all the nutrients; Phytase effect only significant for P only; P-

values for three-way interaction was not significant for any of the nutrients

 ${}^{3}M$  = nutrient matrix for phytase; Ph = phytase; PP = phytate phosphorus