

2019

Proteomic analysis of cortical neuronal cultures treated with poly-arginine peptide-18 (R18) and exposed to glutamic acid excitotoxicity

Gabriella MacDougall

Ryan S. Anderton

The University of Notre Dame Australia, ryan.anderton@nd.edu.au

Frank L. Mastaglia

Neville W. Knuckey

Bruno P. Meloni

Follow this and additional works at: https://researchonline.nd.edu.au/health_article



Part of the [Life Sciences Commons](#), and the [Medicine and Health Sciences Commons](#)

This article was originally published as:

MacDougall, G., Anderton, R. S., Mastaglia, F. L., Knuckey, N. W., & Meloni, B. P. (2019). Proteomic analysis of cortical neuronal cultures treated with poly-arginine peptide-18 (R18) and exposed to glutamic acid excitotoxicity. *Molecular Brain*, 12.

Original article available here:

<https://doi.org/10.1186/s13041-019-0486-8>

This article is posted on ResearchOnline@ND at https://researchonline.nd.edu.au/health_article/281. For more information, please contact researchonline@nd.edu.au.



This article has been published in *Molecular Brain*.

Published by BMC.

Available at: <https://doi.org/10.1186/s13041-019-0486-8>

This is an Open Access article distributed in accordance with the Creative Commons Attribution 4.0 International license (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

See: <https://creativecommons.org/licenses/by/4.0/>


MacDougall, G., Anderton, R.S., Mastaglia, F.L., Knuckey, N.W., and Meloni, B.P. (2019) Proteomic analysis of cortical neuronal cultures treated with poly-arginine peptide-18 (R18) and exposed to glutamic acid excitotoxicity. *Molecular Brain*, 12. doi: 10.1186/s13041-019-0486-8

RESEARCH

Open Access



Proteomic analysis of cortical neuronal cultures treated with poly-arginine peptide-18 (R18) and exposed to glutamic acid excitotoxicity

Gabriella MacDougall^{3,4*} , Ryan S. Anderton^{1,3,4}, Frank L. Mastaglia^{1,3}, Neville W. Knuckey^{1,2,3} and Bruno P. Meloni^{1,2,3}

Abstract

Poly-arginine peptide-18 (R18) has recently emerged as a highly effective neuroprotective agent in experimental stroke models, and is particularly efficacious in protecting cortical neurons against glutamic acid excitotoxicity. While we have previously demonstrated that R18 can reduce excitotoxicity-induced neuronal calcium influx, other molecular events associated with R18 neuroprotection are yet to be investigated. Therefore, in this study we were particularly interested in protein expression changes in R18 treated neurons subjected to excitotoxicity. Proteomic analysis was used to compare protein expression patterns in primary cortical neuronal cultures subjected to: (i) R18-treatment alone (R18); (ii) glutamic acid excitotoxic injury (Glut); (iii) R18-treatment and glutamic acid injury (R18 + Glut); (iv) no treatment (Cont). Whole cell lysates were harvested 24 h post-injury and subjected to quantitative proteomic analysis (iTRAQ), coupled with liquid chromatography-tandem mass spectrometry (LC-MS/MS) and subsequent bioinformatic analysis of differentially expressed proteins (DEPs).

Relative to control cultures, R18, Glut, and R18 + Glut treatment resulted in the detection of 5, 95 and 14 DEPs respectively. Compared to Glut alone, R18 + Glut revealed 98 DEPs, including 73 proteins whose expression was also altered by treatment with Glut and/or R18 alone, as well as 25 other uniquely regulated proteins. R18 treatment reversed the up- or down-regulation of all 73 Glut-associated DEPs, which included proteins involved in mitochondrial integrity, ATP generation, mRNA processing and protein translation. Analysis of protein-protein interactions of the 73 DEPs showed they were primarily associated with mitochondrial respiration, proteasome activity and protein synthesis, transmembrane trafficking, axonal growth and neuronal differentiation, and carbohydrate metabolism. Identified protein pathways associated with proteostasis and energy metabolism, and with pathways involved in neurodegeneration.

Collectively, the findings indicate that R18 neuroprotection following excitotoxicity is associated with preservation of neuronal protein profiles, and differential protein expression that assists in maintaining mitochondrial function and energy production, protein homeostasis, and membrane trafficking.

Keywords: Poly-arginine-18 (R18), iTRAQ proteomics, Neuroprotection, Mito-protection, Excitotoxicity, Stroke

* Correspondence: gabriella01@inet.net.au

³Perron Institute for Neurological and Translational Sciences, QEII Medical Centre, Ground Floor, RR Block, 8 Verdun St, Nedlands, Western Australia 6009, Australia

⁴School of Health Sciences and Institute for Health Research, The University of Notre Dame, Fremantle, Western Australia, Australia

Full list of author information is available at the end of the article



Introduction

A major pathophysiological mechanism responsible for ischaemic stroke injury is excitotoxicity, which is triggered by the excessive release of the excitatory neurotransmitter glutamic acid in response to reduced cerebral blood flow and compromised ATP synthesis. Excitotoxicity initiates a range of forward-feeding biochemical events known as the 'ischaemic cascade', which if not inhibited eventually lead to neuronal death and cerebral infarction [1]. Furthermore, as glutamic acid is the most prominent excitatory neurotransmitter in the CNS [2], the detrimental effects of glutamic acid excitotoxicity also play a role in other acute brain disorders such as traumatic brain injury and epilepsy, as well as chronic neurodegenerative disorders, such as Alzheimer's disease (AD) [3, 4], Huntington's disease (HD) [5, 6], Parkinson's disease (PD) [7, 8], and amyotrophic lateral sclerosis (ALS) [9, 10].

Despite ongoing research, neuroprotective therapies for acute brain injuries and other neurodegenerative disorders are either not available or are extremely limited with modest efficacy. Recent studies in our laboratory have identified cationic arginine-rich peptides (CARPs), which include poly-arginine peptides, as a novel class of neuroprotective agents. In particular, we have demonstrated that poly-arginine-18 (R18, 18-mer of arginine) is neuroprotective in *in vitro* neuronal excitotoxicity models and *in vivo* in rodent models of stroke [11–18], hypoxic-ischaemic encephalopathy (HIE) [19], and traumatic brain injury (TBI) [20, 21].

Given the neuroprotective properties of R18, it is imperative that the molecular pathways that underlie its neuroprotective action are fully elucidated in order to gauge its therapeutic potential. While we have previously demonstrated that R18 has the capacity to reduce glutamic acid-induced excitotoxic neuronal death and intracellular calcium influx, and reduce neuronal NMDA receptor levels [22], CARPs also have cell-penetrating properties and can target mitochondria [23]. Therefore, it is likely that R18 and other CARPs have additional intracellular neuroprotective mechanisms of action. In addition, it is also important to examine the ability of R18 to preserve intracellular protein expression and biochemical pathways following a neurodamaging insult. As such, in this study we performed iTRAQ proteomics and bioinformatic analysis (Fig. 1) of protein cell lysates collected from primary cortical neuronal cultures subjected to glutamic acid excitotoxicity with and without treatment with R18.

Methods

Peptides

Poly-arginine-18 (R18; H-RRRRRRRRRRRRRRRRR-OH) was synthesized by Mimotopes (Australia) and purified to 98% by HPLC. Peptides were prepared as

500 μ M stocks in Baxter water (Australia) and stored at -20°C prior to use.

Primary cortical neuronal cultures

Cortical neuronal tissue was extracted from E18 Sprague-Dawley rat embryos, dissociated, resuspended in Neurobasal/2% B27 supplement (B27) and seeded at approximately 55,000 cells/well into 96-well plates (Nunc, Australia), pre-coated with poly-lysine (Sigma-Aldrich Australia) as previously described [24]. Plates were maintained at 37°C in a CO_2 incubator (95% air balance, 98% humidity, 5% CO_2) until use on day *in vitro* 10, when cultures routinely comprise $>97\%$ neurons and 1–3% astrocytes. Approval for the use of E18 Sprague-Dawley rat embryos for isolation of cortical tissue was obtained by the University of Western Australia Animal Ethics Committee (RA/3/100/1432).

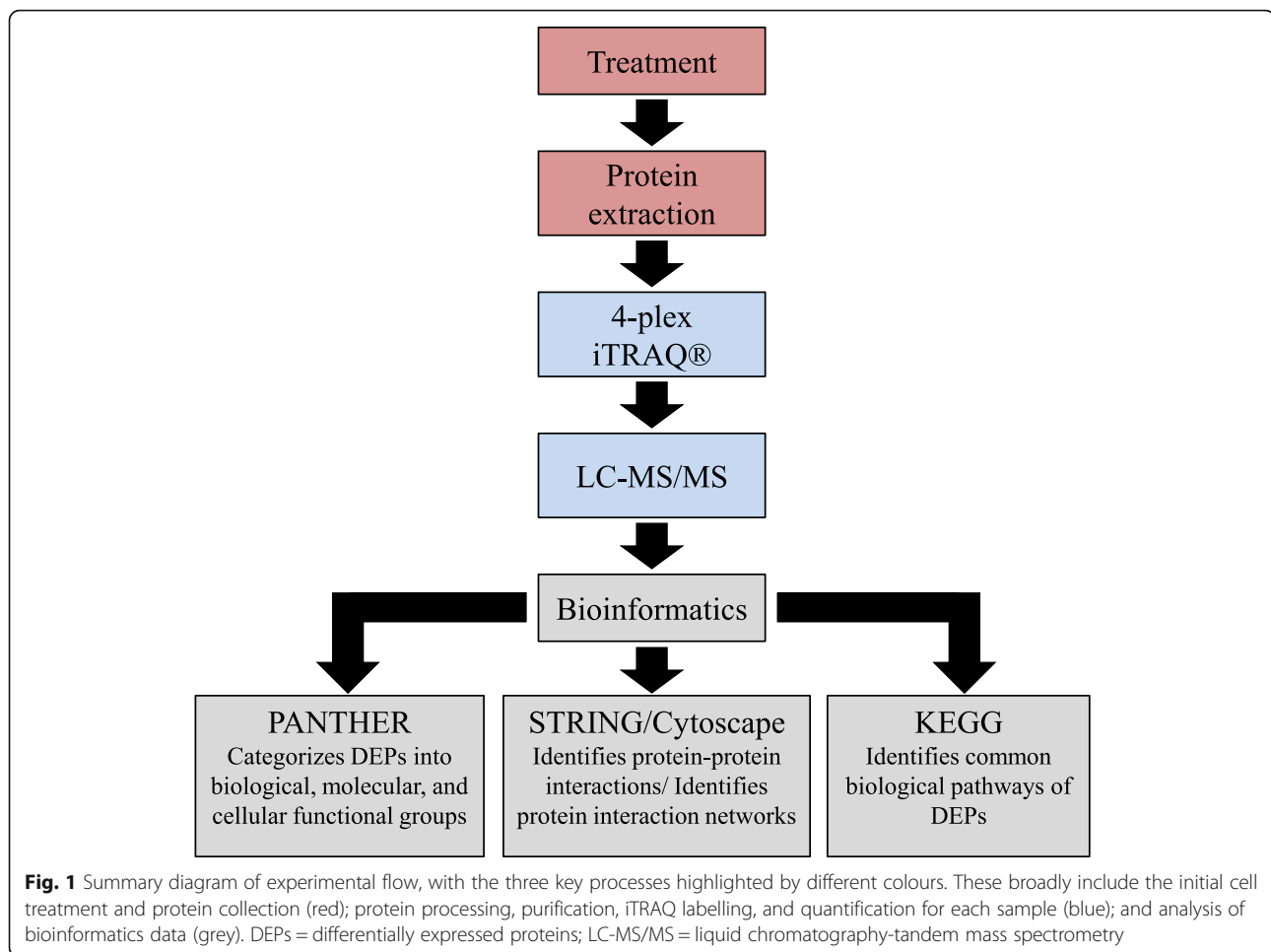
Glutamic acid excitotoxicity model and assessment of cell viability

Cortical neuronal cultures were subjected to glutamic acid excitotoxicity and R18 treatment as previously described [22]. R18 treatment consisted of adding the peptide to culture wells 10 min prior to glutamic acid (L-glutamic acid; Sigma-Aldrich) exposure by removing media and adding 50 μ L of Minimal Essential Media (MEM)/2% B27 containing peptide (2 μ M). To induce excitotoxicity, 50 μ L of MEM/2% B27 containing glutamic acid (200 μ M; final concentration 100 μ M) was added to the culture wells and incubated at 37°C in the CO_2 incubator for 5 min (note: peptide concentration reduced to 1 μ M during this step). Following the 5-min exposure, media was replaced with 100 μ L of MEM/2% B27 and cultures incubated for a further 24 h at 37°C in the CO_2 incubator. Untreated controls with or without glutamic acid treatment underwent the same incubation steps and media additions.

At 24 h post-injury, cell viability was assessed qualitatively by light microscopy, and quantitatively using the CellTiter 96 Aqueous Cell Proliferation MTS assay (Promega, Australia), which determines metabolic capacity of cells through the reduction of the tetrazolium salt (MTS), forming a brown formazan salt that is measured spectrophotometrically at 490 nm.

Protein extraction

At 24 h post-injury, cells were lysed with 20 μ L/well of RIPA buffer (mM: 150 NaCl, 5 EDTA, 50 Tris; %: 1.0 NP-40, 0.5 sodium deoxycholate, 0.1 SDS; pH 8.0) containing protease and phosphatase inhibitor cocktail (Roche Applied Science, Australia). Cell lysates from 8 wells within the same plate were pooled, and this was repeated four times with independent neuronal cultures.



Lysates were clarified by centrifugation at 14,000 g for 5 min at 4 °C, and protein concentration determined via Bradford's assay (Bio-Rad). Aliquots (3.5 mg/mL) of each treatment group were prepared for subsequent iTRAQ analysis and stored at – 20 °C prior to use.

Protein sample preparation and iTRAQ labelling

Quantitative 4-plex iTRAQ proteomics analysis was performed on four independent protein samples for each treatment. Protein sample preparation and iTRAQ labeling was as previously described [25]. Briefly, the protein samples were de-salted, reduced, alkylated, and trypsin-digested according to the iTRAQ protocol [Sciex]. The resulting peptide samples were labeled with iTRAQ reagents as follows: 114: Untreated control (Cont); 115: glutamic acid treated (Glut); 116: R18 treated (R18); 117: R18 and glutamic acid treated (R18 + Glut). All labeled samples were combined to make a pooled sample. Peptides were desalted on a Strata-X 33 µM polymeric reversed phase column (Phenomenex) and dissolved

in a buffer containing 2% acetonitrile 0.1% formic acid before separation by High pH on an Agilent 1100 HPLC system using a Zorbax C18 column (2.1 × 150 mm). Peptides were eluted with a linear gradient of 20 mM ammonium formate, 2% ACN to 20 mM ammonium formate, 90% ACN at 0.2 mL/min. The 95 fractions were concatenated into 12 fractions and dried down. Each fraction was analyzed by electrospray ionization mass spectrometry using the Shimadzu Prominence nano HPLC system [Shimadzu] coupled to a 5600 TripleTOF mass spectrometer [Sciex]. Samples were loaded onto an Agilent Zorbax 300SB-C18, 3.5 µm [Agilent Technologies] and separated with a linear gradient of water/acetonitrile/0.1% formic acid (v/v). Fourteen percent of the labeled sample was loaded on the mass spectrometer.

Proteomic data analysis: qualification and quantitation

Spectral data was qualified using ProteinPilot™ 5.0 software [Sciex] against the SwissProt database, utilizing the

Rattus norvegicus (Rat) taxonomy (Version: April 2017, 7,985 sequences; <https://www.uniprot.org/proteomes/UP000002494>). The False Discovery Rate (FDR) was automatically calculated by the Proteomics System Performance Evaluation Pipeline (PSPEP) feature in the ProteinPilot™ software (AB Sciex, Foster, CA, USA; Version 5.0.1) using the reversed version of the protein sequences contained in the search database. For quantitative protein analysis, a fold change in protein expression $> \pm 1.3$ -fold with a $p < 0.05$ was considered to be a 'differentially expressed protein' (DEP). Protein expression changes with R18, Glut, and R18 + Glut treatment were compared to the control (Cont). In addition, protein changes with Glut treatment were compared with R18 + Glut treatment using Cont protein expression as baseline.

Proteomic data analysis: bioinformatics

Gene ontology analysis with the 'Protein ANalysis THrough Evolutionary Relationships' (PANTHER; Version 14.0, released 2018-12-03; <http://pantherdb.org/>) classification system was utilized to categorize the collective DEPs in the R18, Glut, or R18 + Glut samples, relative to Cont sample, as well as Glut sample, relative to the Glut + R18 sample. These proteins were functionally categorized according to the domains of 'biological processes', 'molecular functions', and 'cellular components' [26].

Protein-protein interaction networks were identified using STRING (Version 11.0, released 2017-05-14; <http://www.string-db.org/>). STRING is a database of known and predicted physical and functional protein-protein interaction, generated through computational prediction from five key databases: 'Genomic Context Predictions', 'High-throughput Lab Experiments', '(Conserved) Co-Expression', 'Automated Textmining', and 'Previous Knowledge in Databases'. Cytoscape (Version 3.7.1) was subsequently utilized to construct and analyze the protein-protein interaction networks, and 'Cluster with overlapping Neighbourhood Expansion' (Cluster ONE; Version 14) was used for network clustering of protein-protein interactions, to identify densely connected and overlapping protein networks.

Identified DEPs were also imported into the Kyoto Encyclopedia of Genes and Genomes (KEGG) Pathway database (<http://www.genome.jp/kegg/pathway.html>) for analysis of common biological pathways and diseases associated with the DEPs.

Statistical analysis

Statistical analysis was conducted with the Prism 8.0 GraphPad statistical software package. Cell viability data were expressed as mean \pm S.E.M. of biological replicates, and multiple comparisons were conducted

with one-way ANOVA and Bonferroni's post hoc test to assess significance, with significance taken as $p < 0.05$. For Cytoscape network analysis, network cut-offs of > 3 proteins were utilized, and a one-sided Mann-Whitney U test was used to identify significant common networks ($p < 0.05$).

Results

R18 improves cell viability in uninjured and glutamic acid-treated neuronal cells

In line with previous studies [12], R18 exhibited potent neuroprotection against glutamic excitotoxic injury in cortical neuronal cultures. In addition, as has been previously reported, cell viability was also significantly increased in neuronal cultures treated with R18 compared to control cultures (Fig. 2).

Quantitative and qualitative proteomic analysis

iTRAQ proteomic analysis detected 7,528 distinct peptide fragments with $> 95\%$ confidence, resulting in the identification of 800 proteins (minimum of ≥ 2 matching peptide hits with $> 95\%$ confidence) consisting of a total of 140 distinct proteins (Table 1 and Additional file 1: Table S1). When compared to Cont, R18, Glut, and Glut + R18 differentially regulated 5, 95 and 14 proteins, respectively (Table 1; see Additional file 2: Table S2 for Glut + R18 DEPs). When compared to Glut, R18 + Glut differentially regulated 98 proteins (Table 1 and Additional file 2: Table S2).

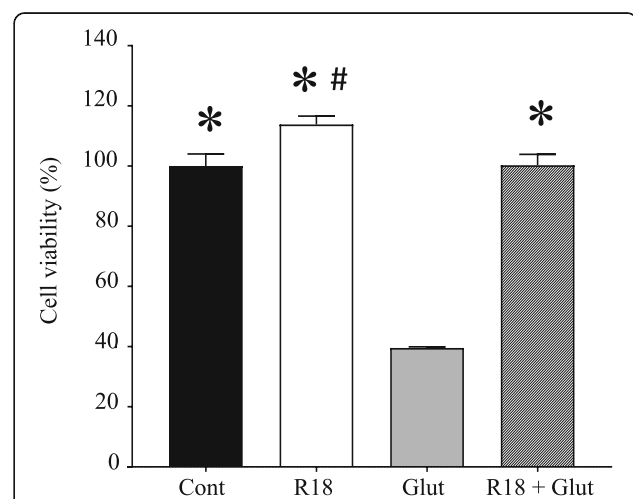


Fig. 2 R18 provides potent neuroprotection against glutamic acid excitotoxicity in primary cortical neurons. Neuronal cultures were subjected to a 10-min R18 pre-treatment (2 μ M) and subsequent 5-min glutamic acid exposure (Glut; 100 μ M). MTS cell viability was assessed at 24 h post-injury. Cell viability was expressed as mean \pm S.E.M. ($p < 0.05$ relative to *Glut or #Cont)

Table 1 Differentially expressed proteins identified in neurons treated with: R18 (R18), glutamic acid (Glut), or R18 and glutamic acid exposure (R18 + Glut)

Gene name	SwissProt Accession Number	Protein	Fold up-/down-regulated ^a		
			R18 vs Cont	Glut vs Cont	R18 + Glut vs Glut
Mitochondrial respiration/function					
Acly	P16638	ATP-citrate synthase	1.076	-2.399^b	1.659
Aco2	Q9ER34	Aconitate hydratase, mitochondrial	-1.028	-3.435	4.285
Atp5a1	P15999	ATP synthase subunit alpha, mitochondrial	-1.472	1.888	-2.421
Atp5b	P10719	ATP synthase subunit beta, mitochondrial	-1.117	2.558	-3.342
Atp5h	P31399	ATP synthase subunit d, mitochondrial	1.076	2.754^b	-1.836
Atp5o	Q06647	ATP synthase subunit O, mitochondrial	1.406	2.032^b	-1.644
Cat	P04762	Catalase	-1.555	1.138	-1.659^b
Cox4i1	P10888	Cytochrome c oxidase subunit 4 isoform 1, mitochondrial	-1.259	2.070	-2.704
Idh1	P41562	Isocitrate dehydrogenase [NADP] cytoplasmic	1.066	1.459	-1.486
Mdh2	P04636	Malate dehydrogenase, mitochondrial	-1.247	2.355	-2.535
Ndufs1	Q66HF1	NADH-ubiquinone oxidoreductase 75 kDa subunit, mitochondrial	1.294	2.443	-5.249
Uqcrc2	P32551	Cytochrome b-c1 complex subunit 2, mitochondrial	-1.247	1.888	-2.704
Proteasome & Protein synthesis					
Asns	P49088	Asparagine synthetase [glutamine-hydrolyzing]	2.089	2.228	2.421^b
Cct2	Q5XIM9	T-complex protein 1 subunit beta	1.106	-3.802	3.631
Cct3	Q6P502	T-complex protein 1 subunit gamma	-1.158	-4.787	4.207
Cct4	Q7TPB1	T-complex protein 1 subunit delta	1.127	-3.944	3.499
Cct5	Q68FQ0	T-complex protein 1 subunit epsilon	1.096	-2.269	2.291
Psmc1	P62193	26S protease regulatory subunit 4	-1.419	-22.67	18.197
Axonal growth/Neuronal differentiation/Cytoskeletal arrangement					
Actr2	Q5M7U6	Actin-related protein 2	-1.355	-1.271	-1.459^b
Ap2m1	P84092	AP-2 complex subunit mu	1.271	-5.394	6.855
Armc10	B1WBW4	Armadillo repeat-containing protein 10	-1.294	-6.667	2.070
Baiap2	Q6GMN2	Brain-specific angiogenesis inhibitor 1-associated protein 2	-1.331	-90.09	99.083
Basp1	Q05175	Brain acid soluble protein 1	1.343	1.614	-1.270
Cntn1	Q63198	Contactin-1	-1.117	1.977	-1.836
Ctnn	Q66HL2	Src substrate cortactin	1.117	-5.345^b	3.251
Dcx	Q9ESI7	Neuronal migration protein doublecortin	1.000	-6.139	6.026
Dnm1	P21575	Dynamamin-1	-2.148	-4.093	3.076
Dpysl2	P47942	Dihydropyrimidinase-related protein 2	-1.225	1.723	-1.690
Dync1h1	P38650	Cytoplasmic dynein 1 heavy chain 1	-1.087	-8.873	8.472
Fabp7	P55051	Fatty acid-binding protein, brain	-1.028	4.406^b	-4.207
Fyn	Q62844	Tyrosine-protein kinase Fyn	1.117	-2.489	2.754^b
Gdi1	P50398	Rab GDP dissociation inhibitor alpha	-1.419	1.820	-2.965
Gfap	P47819	Glial fibrillary acidic protein	1.4322	1.906^b	-1.159
Kif21b	F1M5N7	Kinesin-like protein KIF21B	-1.282	-17.857	16.444
Krt1	Q6IMF3	Keratin, type II cytoskeletal	-4.055^b	-1.236	-6.485
Krt10	Q6IFW6	Keratin, type I cytoskeletal 10	-2.168	-1.722	-16.892^b
Map2	P15146	Microtubule-associated protein 2	1.138	-8.628	11.482
Map4	Q5M7W5	Microtubule-associated protein 4	-2.884	-4.488	4.169
Mapt	P19332	Microtubule-associated protein tau	1.077	-7.179	7.379

Table 1 Differentially expressed proteins identified in neurons treated with: R18 (R18), glutamic acid (Glut), or R18 and glutamic acid exposure (R18 + Glut) (*Continued*)

Gene name	SwissProt Accession Number	Protein	Fold up-/down-regulated ^a		
			R18 vs Cont	Glut vs Cont	R18 + Glut vs Glut
Myh10	Q9JLT0	Myosin-10	-1.459	-2.466	2.228
Ncam1	P13596	Neural cell adhesion molecule 1	-1.180	2.831	-4.093
Pa2g4	Q6AYD3	Proliferation-associated protein 2G4	1.202	-2.188	2.489^b
Pak3	Q62829	Serine/threonine-protein kinase PAK 3	-2.208	-4.365	5.754^b
^b Pebp1	P31044	Phosphatidylethanolamine-binding protein 1	1.5276	2.780^b	-2.128
Rala	P63322	Ras-related protein Ral-A	-1.236	2.704^b	-1.690
Rtn4	Q9JK11	Reticulon-4	1.107	-2.148	2.679
Tpm4	P09495	Tropomyosin alpha-4 chain	-1.097	-2.270	1.738^b
Tuba4a	Q5XIF6	Tubulin alpha-4A chain	1.086	-2.679^b	2.535
Tubb5	P69897	Tubulin beta-5 chain	-1.038	-18.018	16.444
Vesicular/Transmembrane trafficking					
Actn4	Q9QXQ0	Alpha-actinin-4	-1.294	1.906^b	-1.381
Actr1a	P85515	Alpha-centractin	-1.738	-3.565^b	2.148
Ap2a2	P18484	AP-2 complex subunit alpha-2	1.159	-2.582	3.698^b
Atp1a3	P06687	Sodium/potassium-transporting ATPase subunit alpha-3	-2.128	1.660	-2.355
Cadps	Q62717	Calcium-dependent secretion activator 1	-1.486	-11.481	6.194
Camk2a	P11275	Calcium/calmodulin-dependent protein kinase type II subunit alpha	-1.419	-6.667	6.918
Cask	Q62915	Peripheral plasma membrane protein CASK	1.514	1.306	-1.888^b
Dpysl5	Q9JHU0	Dihydropyrimidinase-related protein 5	-1.180	1.213	-1.459^b
Klc1	P37285	Kinesin light chain 1	1.107	-6.024^b	5.058
Nsf	Q9QUL6	Vesicle-fusing ATPase	-1.117	-5.807	4.656
Prkar2b	P12369	cAMP-dependent protein kinase type II-beta regulatory subunit	-1.472	-3.105	1.754
Stx1b	P61265	Syntaxin-1B	1.225	1.600	-1.419
Stxbp1	P61765	Syntaxin-binding protein 1	-1.107	1.600	-2.466
Syn1	P09951	Synapsin-1	-1.786	-3.597^b	3.3113
ER proteostasis/Protein modification					
Calr	P18418	Calreticulin	1.419	1.871	-1.570
Ddost	Q641Y0	Oligosaccharyl transferase 48 kDa subunit	-1.500	2.466	-2.938
Erp29	P52555	Endoplasmic reticulum resident protein 29	-1.067	2.168^b	-3.163
Hsp90aa1	P82995	Heat shock protein HSP 90-alpha	1.038	1.282	-1.446^b
Hspa5	P06761	78 kDa glucose-regulated protein	1.038	2.014	-1.542
Hspa8	P63018	Heat shock cognate 71 kDa protein	1.259	-2.208	2.559^b
Hspa9	P48721	Stress-70 protein, mitochondrial	1.159	-6.083	8.318
Hspd1	P63039	60 kDa heat shock protein, mitochondrial	1.107	1.486	-1.556
Pdia3	P11598	Protein disulfide-isomerase A3	-1.600	1.614	-1.995
Phb	P67779	Prohibitin	-1.213	1.644	-2.535
Phb2	Q5XIH7	Prohibitin-2	-1.047	3.837	-2.992
Por	P00388	NADPH-cytochrome P450 reductase	-1.107	-1.038	1.343^b
Tcp1	P28480	T-complex protein 1 subunit alpha	-1.472	-3.597^b	2.911
Uba1	Q5U300	Ubiquitin-like modifier-activating enzyme 1	-1.107	-5.105	4.406
Glycolysis & Carbohydrate metabolism					
Alb	P02770	Serum albumin	-1.570	1.722	-5.701

Table 1 Differentially expressed proteins identified in neurons treated with: R18 (R18), glutamic acid (Glut), or R18 and glutamic acid exposure (R18 + Glut) (*Continued*)

Gene name	SwissProt Accession Number	Protein	Fold up-/down-regulated ^a		
			R18 vs Cont	Glut vs Cont	R18 + Glut vs Glut
Aldoa	P05065	Fructose-bisphosphate aldolase A	-1.660	1.629	-2.355
Eno1	P04764	Alpha-enolase	1.107	2.911	-1.706
Gapdh	P04797	Glyceraldehyde-3-phosphate dehydrogenase	-1.028	-11.700	10.280
Gpi	Q6P6V0	Glucose-6-phosphate isomerase	-1.318	1.754	-3.435
Hk1	P05708	Hexokinase-1	1.138	2.051^b	-1.888
Ldha	P04642	L-lactate dehydrogenase A chain	-1.786	1.028	-1.675^b
Pkm	P11980	Pyruvate kinase PKM	-1.009	1.787	-1.941
Taldo1	Q9EQS0	Transaldolase	1.259	-1.837	2.109
Mitochondrial fatty acid synthesis					
Acat1	P17764	Acetyl-CoA acetyltransferase, mitochondrial	1.159	2.377	-1.500
Bdh1	P29147	D-beta-hydroxybutyrate dehydrogenase, mitochondrial	-1.213	-5.970	4.246
Fasn	P12785	Fatty acid synthase	-1.117	-3.311	2.704
Got2	P00507	Aspartate aminotransferase, mitochondrial	-1.923	1.213	-2.109^b
Ribosome components/RNA trafficking & processing					
Aars	P50475	Alanine-tRNA ligase, cytoplasmic	-1.556	-5.495	4.699
C1qbp	O35796	ASF/SF2-associated protein p32	1.213	1.600^b	-1.888
Ddx1	Q641Y8	ATP-dependent RNA helicase DDX1	-1.148	-11.173	4.286
Eef1a1	P62630	Elongation factor 1-alpha 1	-1.225	-5.444	4.529
Eef2	P05197	Elongation factor 2	-1.202	-4.131	3.048
Eif4a2	Q5RK11	Eukaryotic initiation factor 4A-II	-1.057	-3.945	3.02
Eif5a	Q3T1J1	Eukaryotic translation initiation factor 5A-1	1.514	-6.983	9.638
Elavl2	Q8CH84	ELAV-like protein 2	-1.107	-4.405	5.598^b
Hnrnpa1	P04256	Heterogeneous nuclear ribonucleoprotein A1	-1.076	-9.911	9.462
Hnrnpa2b1	A7VJC2	Heterogeneous nuclear ribonucleoproteins A2/B1	1.318	-4.325	6.138
Hnrnpa3	Q6URK4	Heterogeneous nuclear ribonucleoprotein A3	1.191	-3.311	4.920
Hnrnpd	Q9JJ54	Heterogeneous nuclear ribonucleoprotein D0	1.202	-5.495	6.252
Hnrnpk	P61980	Heterogeneous nuclear ribonucleoprotein K	-1.057	-8.091	8.872
Hnrnpl	F1LQ48	Heterogeneous nuclear ribonucleoprotein L	-1.028	-2.729	4.207^b
Khsrp	Q99PF5	Far upstream element-binding protein 2	1.556	-3.908	8.017^b
Matr3	P43244	Matrin-3	-1.500	-3.373^b	3.945
Rpl7	P05426	60S ribosomal protein L7	1.225	-2.109	2.188^b
Rpl13	P41123	60S ribosomal protein L13	-1.057	-2.377	2.754^b
Rplp0	P19945	60S acidic ribosomal protein P0	1.486^b	-1.191	-1.076
Rps24	P62850	40S ribosomal protein S24	1.854	-5.754	14.06^b
Rps27	Q71TY3	40S ribosomal protein S27	-1.102	-18.18^b	5.058
Yars	Q4KM49	Tyrosine-tRNA ligase, cytoplasmic	-1.225	-4.656	3.342
Calcium transport and signalling					
Vdac1	Q9Z2L0	Voltage-dependent anion-selective channel protein 1	-1.419	2.466	-2.938
Cacna2d1	P54290	Voltage-dependent calcium channel subunit alpha-2/delta-1	1.306	4.207	-4.018
Gnao1	P59215	Guanine nucleotide-binding protein G(o) subunit alpha	-1.148	-5.152	3.802
Letm1	Q5XIN6	LETM1 and EF-hand domain-containing protein 1, mitochondrial	-3.020	1.459^b	-1.236
Nudt3	Q566C7	Diphosphoinositol polyphosphate phosphohydrolase 1	1.459^b	1.282	1.117

Table 1 Differentially expressed proteins identified in neurons treated with: R18 (R18), glutamic acid (Glut), or R18 and glutamic acid exposure (R18 + Glut) (*Continued*)

Gene name	SwissProt Accession Number	Protein	Fold up-/down-regulated ^a		
			R18 vs Cont	Glut vs Cont	R18 + Glut vs Glut
Ywhaq	P68255	14-3-3 protein theta	-1.087	1.355	-1.615
Miscellaneous					
Ak1	P39069	Adenylate kinase isoenzyme 1 (<i>Cellular energy homeostasis</i>)	1.009	-2.938	4.325
Atic	O35567	Bifunctional purine biosynthesis protein PURH (<i>Purine biosynthesis</i>)	1.038	1.923	-2.630^b
Dnm1l	O35303	Dynamin-1-like protein (<i>Mitochondrial fission</i>)	-1.820	-7.519^b	5.297
Hist1h4b	P62804	Histone H4 (<i>Transcription regulation</i>)	1.754	-3.802	7.516^b

^a Statistically significant values ($p < 0.05$) for fold up-/down-regulation ≥ 1.3 -fold are highlighted in bold. ^b 25 uniquely DEPs in R18 and Glut treatment versus Cont, or R18 + Glut treatment versus Glut

Proteins regulated by R18 treatment alone (R18 vs Cont)

Of the five DEPs identified in the R18 sample, three were uniquely regulated (Table 1). Two of these proteins were upregulated: 60S acidic ribosomal protein P0 (Rplp0; 1.49) and Diphosphoinositol polyphosphate phosphohydrolase 1 (Nudt3; 1.46), and one was down-regulated: keratin, type II cytoskeletal (Krt1; -4.05).

Proteins regulated by glutamic acid injury alone (glut vs Cont)

Of the 95 DEPs identified in the Glut sample, 21 were uniquely regulated (Table 1). The greatest magnitude fold-change in a down-regulated protein was observed for brain-specific angiogenesis inhibitor 1-associated protein 2 (Baiap2; -90.09), while the greatest fold-change in an up-regulated protein was for fatty acid-binding protein, brain (Fabp7; 4.41).

Proteins regulated by R18 plus glutamic acid injury (R18 + Glut vs Cont)

Of the 14 DEPs identified in the R18 + Glut sample none were uniquely regulated (Additional file 2: Table S2). The greatest magnitude fold-change in an up-regulated protein was observed for UV excision repair protein RAD23 homolog B (Rad23b; 3.251), while the greatest fold-change in a down-regulated protein was for Keratin, type I cytoskeletal 10 (Krt10; -33.333).

Proteins regulated by R18 plus glutamic acid injury vs glutamic acid injury alone (R18 + Glut vs glut)

Of the 98 DEPs identified after R18 + Glut treatment (R18 + Glut vs Glut), 73 of the proteins were also regulated by R18 and/or Glut treatments alone, and 25 were uniquely regulated (Table 1; uniquely regulated proteins indicated by *). In addition, R18 treatment reversed the up- or down-regulation of all 73 DEPs (Table 1, Fig. 3). Of the 25 uniquely regulated proteins, the greatest magnitude fold-change in protein expression observed was with Brain-specific angiogenesis inhibitor 1-associated protein 2 (Baiap2; -99.08).

Functional categorization of differentially regulated proteins (DEPs)

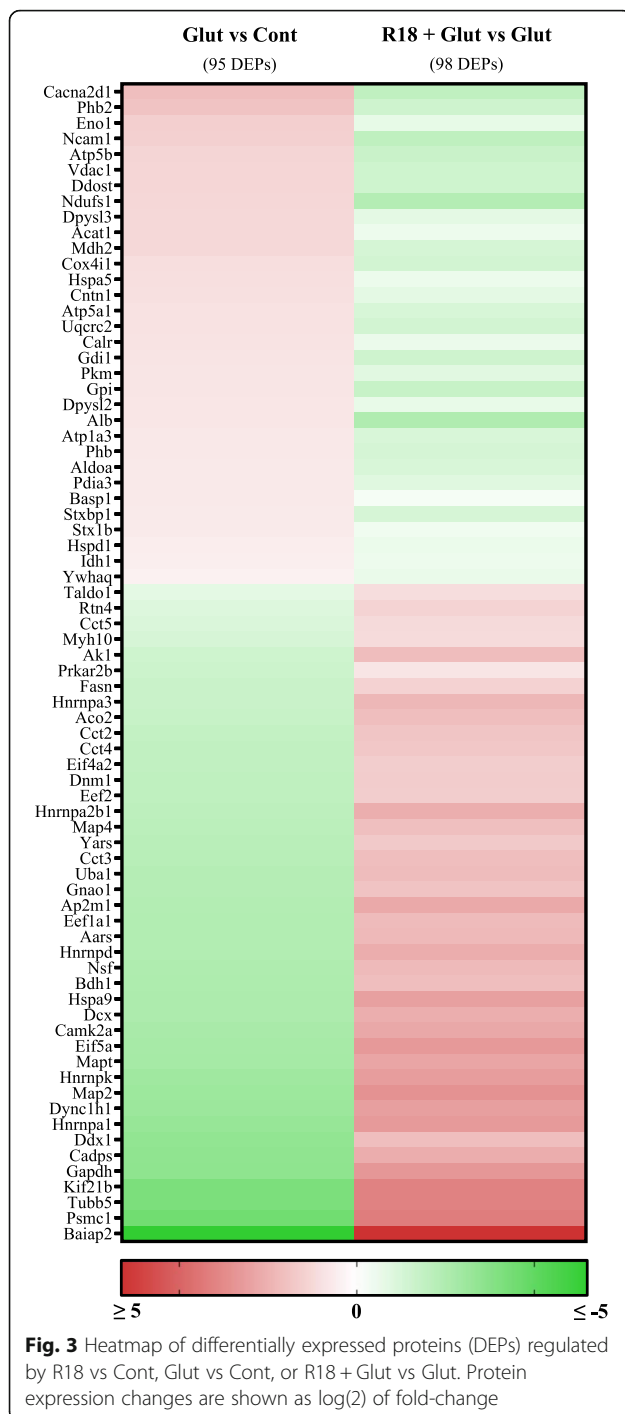
For further functional characterization of DEPs we focused on protein changes in the R18 and R18 + Glut treatments groups as we were most interested in the effects of R18 on proteins regulated in uninjured and injured neurons. PANTHER Gene Ontology analysis was utilized to categorize the DEPs regulated by R18 alone (5 proteins; R18 vs Cont), and the DEPs regulated by R18 + Glut treatment (73 proteins; R18 + Glut vs Glut) according to 'cellular component' (Fig. 4a and d) 'biological process' (Fig. 4b and e), and 'molecular function' (Fig. 4c and f) (Full data provided in Additional file 3: Table S3).

The 5 DEPs regulated by R18 treatment included proteins located in the nucleus and ribosomes which catalyze purine nucleotide catabolic activity (e.g. Nudt3), and modulate rRNA binding (e.g. Rplp0). Other proteins regulated by R18 are involved in intracellular ion trafficking (e.g. Atp1a3) and cytoskeletal structure (e.g. Krt1) (Table 1).

The 73 DEPs regulated by R18 + Glut treatment included proteins involved in mitochondrial respiration and function (e.g. Aco2 and Atp5a1), proteasomal regulation and protein synthesis (e.g. Psmc1 and Cct3), proteostasis/protein modification in the endoplasmic reticulum (e.g. Hspa9 and Uba1), and RNA trafficking/processing (e.g. Eif5a and Hnrnpa1), as well as cytoskeletal rearrangement and axonal growth (e.g. Map2 and Dcx) and vesicular/membrane trafficking (e.g. Atp1a3 and Camk2a). Significant changes of key neuronal proteins are summarized in the schematic detailed in Fig. 5 (full data available in Additional file 2: Table S2 and Additional file 3: Table S3).

Protein-protein interaction network analysis of DEPs regulated by R18 in uninjured and glutamic acid injured neurons

STRING analysis was used to identify potential protein-protein interactions across the 5 DEPs regulated by R18



treatment, and the 73 DEPs regulated by Glut and R18 + Glut. No significant protein-protein interactions were identified for the 5 DEPs regulated by R18 treatment (R18 vs Cont) (Fig. 6a). Two hundred and twenty-two nodes representing direct and indirect protein-protein interactions were identified for the 73 DEPs regulated by R18 + Glut treatment (R18 + Glut vs Glut) (Fig. 6b).

ClusterONE network analysis of the 222 nodes revealed that the protein-protein interactions could be grouped into eight clusters representing distinct biological functional entities (Fig. 7; boxed proteins). The clusters were classified as 'Mitochondrial respiration' (55 nodes), 'Proteasome and Protein synthesis' (43 nodes), 'Axonal growth & neuronal differentiation' (11 nodes), 'Transmembrane trafficking' (10 nodes), 'Endoplasmic reticulum proteostasis' (8 nodes), 'Glycolysis and carbohydrate metabolism' (7 nodes), 'RNA trafficking and processing' (4 nodes), and 'Mitochondrial fatty acid synthesis' (4 nodes) (Full data provided in Additional file 4: Table S4).

KEGG pathway analysis of 73 shared proteins

KEGG pathway analysis to determine the biological pathways and diseases associated with 222 protein-protein interactions identified pathways pertaining to proteostasis ('Proteasome'; 34 of 46 proteins), energy metabolism ('Oxidative phosphorylation'; 52 of 130 proteins), and neurotransmission ('Synaptic vesicle cycle'; 14 of 60 proteins), and 'Retrograde endocannabinoid signaling'; 34 of 144 proteins) (Fig. 8; full data in Additional file 5: Table S5). In addition, KEGG analysis revealed that the protein-protein interactions were associated with the neurodegenerative disorders Parkinson's disease (PD; 52 of 134 proteins), Alzheimer's disease (AD; 47 of 164 proteins), and Huntington's disease (HD; 50 of 181 proteins).

Discussion

In recent years, CARPs have emerged as a novel class of potential neuroprotective therapeutics for a broad range of acute brain injuries and chronic neurodegenerative disorders. These CARPs include short-chained poly-arginine peptides [11, 13, 27], SS-peptides [28, 29], APOE-derived peptides [27, 30], and TAT-fused peptides, including TAT-NR2B9c (NA-1) and JNK1-TAT [11, 31]. Such CARPs have been shown to exert their neuroprotective action through a variety of targets, which include structural and functional preservation of mitochondria [32], reduced ROS generation [33], inhibition of protein aggregation [34], modulation of glutamate or calcium ion receptors (excitotoxicity/calcium influx) [35], and activation of pro-survival signaling [36, 37]. Given the diverse biochemical and cellular effects CARPs can exert on cells, it is likely that other cytoprotective processes are also involved, which have yet to be fully elucidated. To this end, the present study is the first to employ an iTRAQ proteomics approach to gain insight into protein expression changes after poly-arginine-18 (R18) treatment of uninjured neuronal cultures and neuronal cultures subjected to glutamic acid excitotoxic injury.

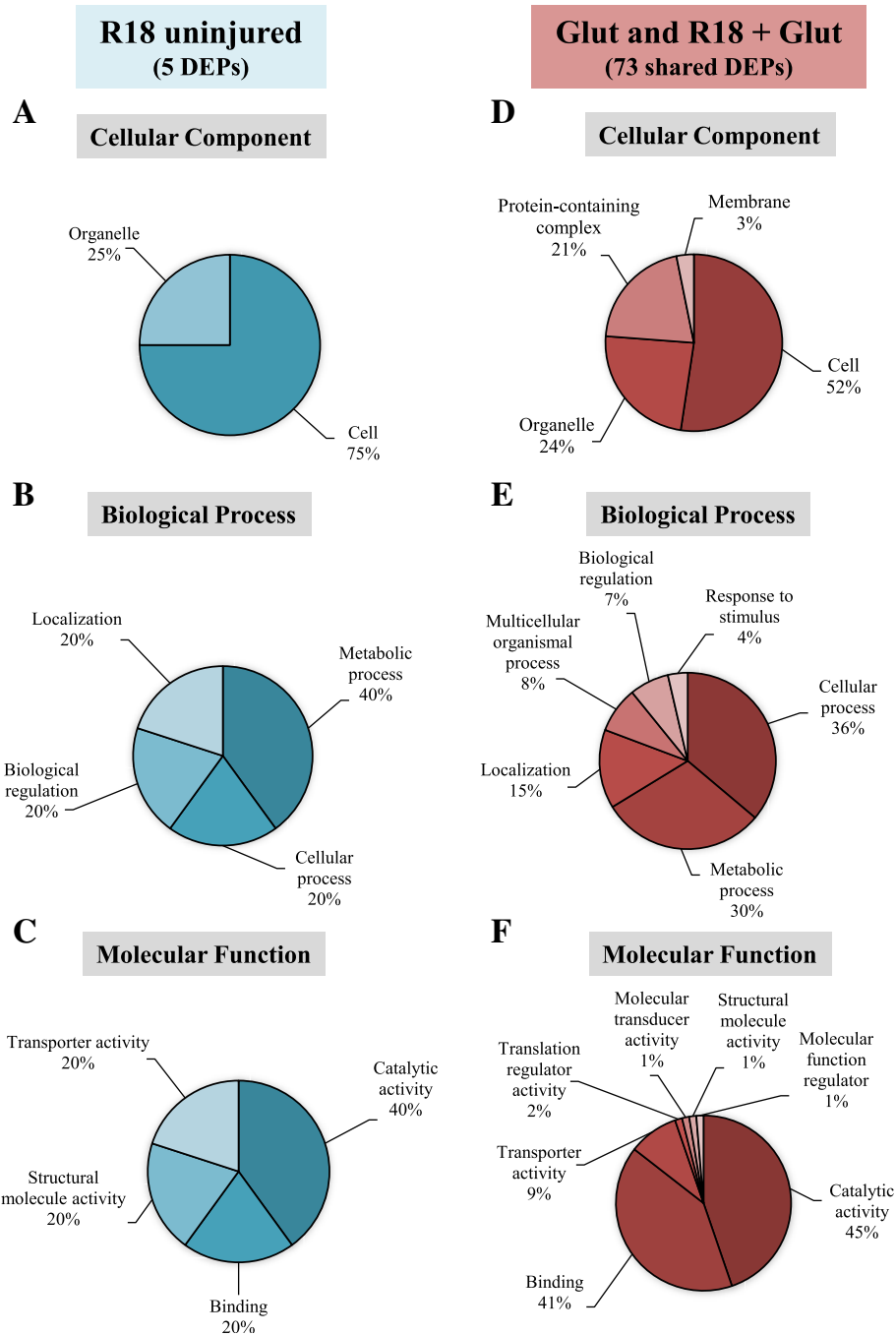
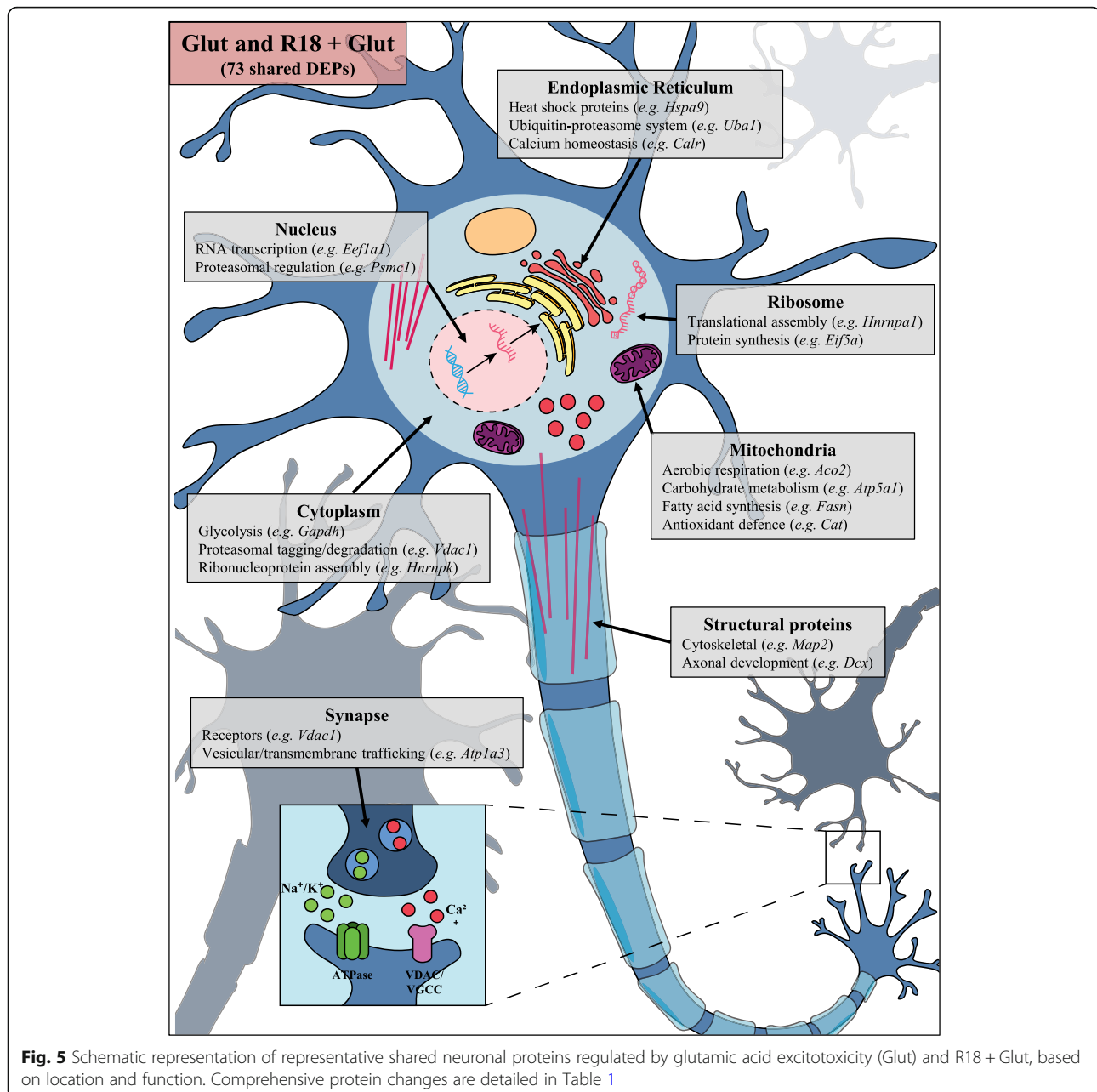


Fig. 4 PANTHER gene-ontology functional categorization of DEPs regulated by R18 in healthy neurons (vs Cont; blue), and R18 + Glut in injured neurons (vs Glut; red). The top five categories are displayed across the functional categories of (a, b) Molecular Function (c, d) Biological Process, and (e, f) Cellular Component

The effects of R18 treatment on uninjured neurons

Proteomics analysis of neuronal cultures 24-h after a 10-min exposure to R18 identified 5 DEPs. The small number of detected protein expression changes was surprising given that CARPs can induce a variety of biological effects on cells [38]. However, it is likely

that the small number of proteins detected was in part due to the 24-h post-R18 treatment time point used to analyze protein expression changes, as majority of the protein changes elicited by R18 treatment may potentially occur within the first few hours, and as such, may no longer have been detectable or did



not fit the requirements for classification of a DEP (e.g. $\pm > 1.3$ fold change) by 24 h.

The proteins that were identified as being affected by R18 were largely associated with protein synthesis and transmembrane protein and cationic ion transport, and did not significantly map onto KEGG pathways, suggesting that the R18 peptide does not exert long-term biological effects in uninjured neurons. This is in line with the proposed notion that neuroprotective agents should preferentially interact with and/or modulate cellular targets activated following

pathological events to minimize the chance of off-target side effects. Such protective agents are deemed 'pathologically-activated' therapeutics, which are thought to have a particularly useful application in neurological disorders, as brain tissue is especially susceptible to drug-induced disruptions and unwanted drug side-effects [38, 39]. However, to provide a more comprehensive assessment of the biological effects of R18 on uninjured neurons, future studies should examine protein expression at earlier time points.

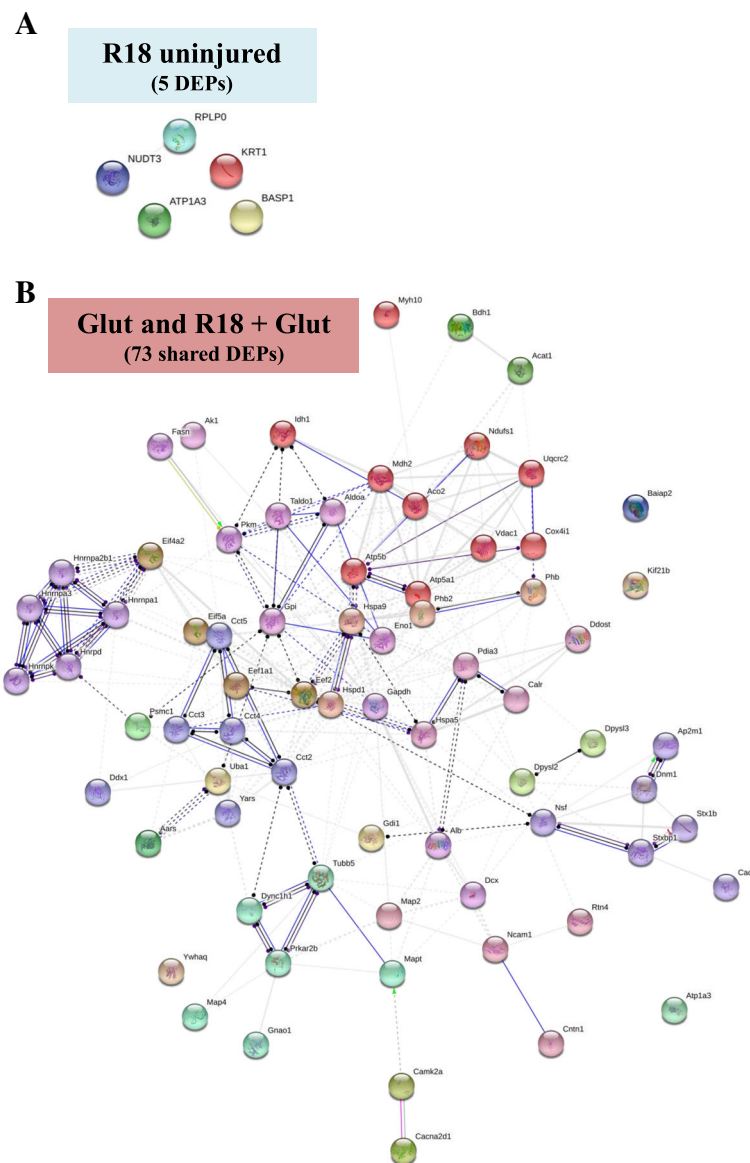


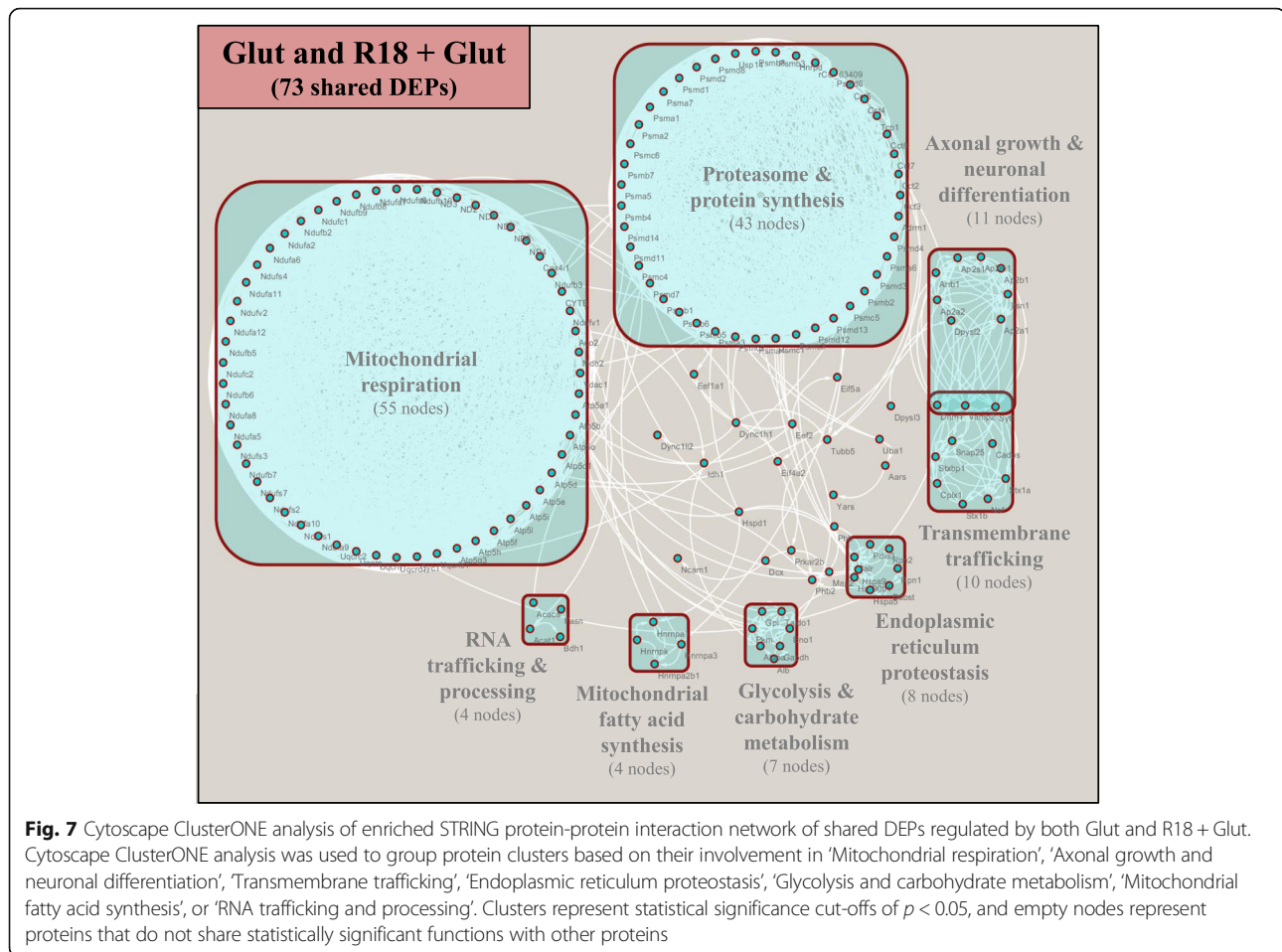
Fig. 6 Enriched STRING PPI network analysis of DEPs regulated by **a.** R18 treatment in healthy neurons (vs Cont), and **b.** DEPs commonly regulated by Glut and R18 + Glut (vs Glut), demonstrating molecular actions of direct and indirect protein-protein interactions between significantly regulated proteins. STRING parameters were set to high confidence (0.700), with only query proteins shown

R18 treatment reduces glutamic acid-induced changes in protein expression

An important finding of this study was the ability of R18 treatment to reverse the majority (74.5%; 73 of 98) of the protein changes induced by glutamic acid excitotoxicity, and thereby preserve the protein expression profiles of cortical neurons post-insult (Table 1; Additional file 4: Table S4). Further analysis revealed that these protein changes underpin key cellular functions, such as mitochondrial respiration and energy production, proteostasis, neuronal transmembrane trafficking, and RNA processing, which are

dysregulated by excitotoxicity. Moreover, KEGG analysis of protein-protein interactions indicated predominant involvement of pathways pertaining to the proteasome and oxidative phosphorylation, which also represent two central biological processes underpinning aspects of neurodegenerative pathophysiology. This likely contributed to the identification of enriched protein-protein interactions pertaining to Parkinson's disease, Alzheimer's disease and Huntington's disease.

Severe and/or prolonged disruptions in the ubiquitin-proteasome system have been implicated in both acute

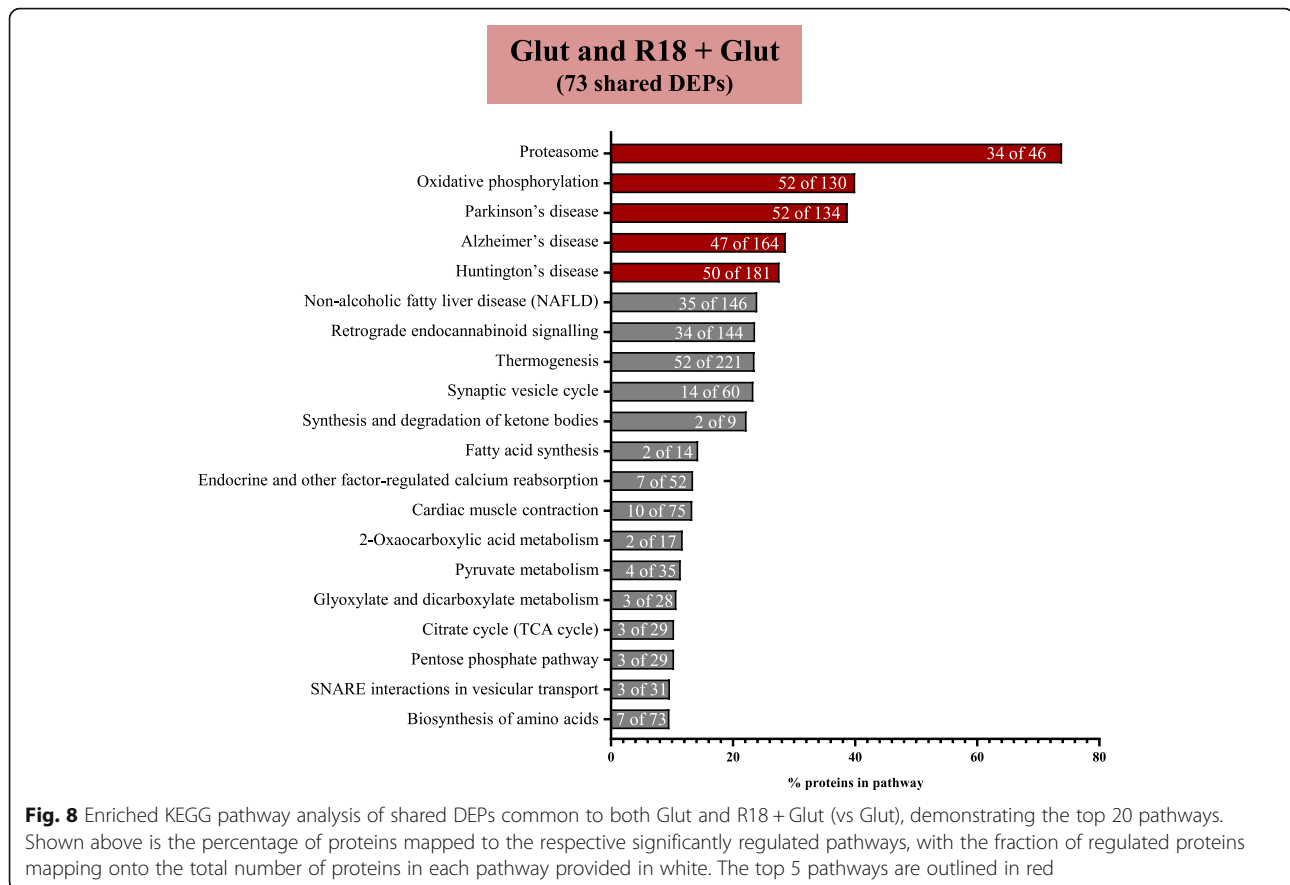


(ischaemic stroke, TBI) and chronic (AD, PD, motor neuron disease) neurological disorders [39, 40]. Previous studies have shown that CARPs exhibit proteasomal modulatory activity and could potentially conserve protein expression profiles through inhibition of injury-induced proteasomal protein degradation. For example, the arginine-rich PR-11 (H-RRRPRPPYLPRRPPPPFFPRLPPRIPPGFPPRFPFRFP-OH; net charge + 11) and PR-39 (H-RRRPRPPYLPR-OH; net charge + 5) peptides attenuate inflammation induced by ischaemia-reperfusion injury through inhibition of proteasomal degradation of I κ B α ; a NF κ B inhibitory protein [41, 42]. Taken together, it appears CARPs can influence the function of the proteasome, and thereby exert neuroprotective benefits during times of cellular stress.

Proteomic analysis also revealed that R18 preserved protein expression profiles pertaining to mitochondrial bioenergetics and structural integrity. Mitochondria are central mediators of intracellular calcium signaling events during excitotoxicity, and as such, are considered the "judge, jury, and executioner" of the cell [31, 43].

During excitotoxic injury, mitochondria act as a buffer for toxic intracellular calcium accumulation, however excessive mitochondrial calcium uptake can disrupt their structural and functional integrity, resulting in the release of pro-death signaling proteins from the mitochondrial inter-membrane space [44, 45]. Therefore, the ability of R18 treatment to attenuate excitotoxicity-induced protein changes underlying loss of mitochondrial integrity, provides evidence that the peptide helps preserve the function of the organelle in times of cellular stress. In line with the ability of CARPs to maintain mitochondrial function and energy generation, in this and previous studies R18 was demonstrated to increase MTS metabolism in uninjured neurons and in neurons after exposure to glutamic acid [14].

Bioinformatic analysis of the DEPs identified in the present study largely focused on the 73 proteins up- or down-regulated by glutamic acid excitotoxic injury. However, it is important to note that 25 other DEPs were also identified to be uniquely regulated by combined R18 + Glut treatment, which may represent additional proteins influenced by R18 and associated with



neuroprotection. Alternatively, these proteins could reflect non-specific changes in protein expression unrelated to neuroprotection.

Limitations and future directions

The proteomics methodology used in this study does not provide insight into other forms of protein modification, such as post-translational changes (e.g. phosphorylation, acetylation, and glycosylation), which may influence protein functions important for neuroprotection. In addition, only a 24-h timepoint was examined and therefore it would also be of interest to examine protein expression changes, as well as post-translational modifications at earlier timepoints after R18 treatment. Further studies are also required to confirm if the DEPs and the biochemical and disease pathways influenced by R18 treatment after glutamic acid excitotoxicity *in vitro* are also affected by the peptide in animal models of acute brain injury (e.g. stroke, TBI) and chronic neurodegenerative disorders (AD, PD).

Conclusion

This exploratory study has demonstrated for the first time that the poly-arginine peptide R18 exerts significant effects in attenuating the protein expression changes

associated with neuronal excitotoxicity *in vitro*, while inducing minimal changes in uninjured neurons. Collectively, our findings indicate that the neuroprotective effects of R18 following excitotoxicity are associated predominantly with preservation of neuronal proteostasis, together with positive effects on mitochondrial and proteasomal function. The findings of this study provide further evidence supporting the role of poly-arginine peptides as a potential neuroprotective therapeutic for both acute and chronic neurodegenerative disorders.

Additional files

Additional file 1: Table S1. Summary of LC-MS/MS spectral data analysis. Summary of LC-MS/MS spectral data analysis with ProteinPilot™ 5.0 Software [Sciex] using the SwissProt database (Version April 2017; 7,985 sequences) against *Rattus norvegicus* (Rat) taxonomy, using the reversed version of the protein sequences contained in the search database. FDR was automatically calculated with the Proteomics System Performance Evaluation Pipeline (PSPEP) feature in the ProteinPilot™ software. (DOCX 17 kb)

Additional file 2: Table S2. Full iTRAQ proteomics data. Full iTRAQ proteomics data showing relative fold changes in protein expression and corresponding *p*-values. (DOCX 54 kb)

Additional file 3: Table S3. PANTHER gene-ontology functional categorization. PANTHER gene-ontology functional categorization of DEPs significantly regulated by R18 treatment alone (R18 vs Cont), glutamic acid exposure (Glut vs Cont), and R18 pre-treatment with glutamic acid

exposure (R18 + Glut vs Glut). Note: proteins may have multiple functions, and as such, the total number of proteins in each category may be greater than the sum of DEPs across each treatment group. (DOCX 49 kb)

Additional file 4: Table S4. Cytoscape quantitative analysis. Cytoscape quantitative analysis of STRING data cluster strength for 73 shared DEPs across Glut and R18 + Glut treatment groups. (DOCX 56 kb)

Additional file 5: Table S5. Quantitative data and full gene list of KEGG pathway analysis. Quantitative data and full gene list of KEGG pathway analysis of 73 shared DEPs across Glut and R18 + Glut treatment groups, with details provided on term ID, overserved gene count vs. background gene count, and FDR. (DOCX 17 kb)

Abbreviations

AD: Alzheimer's disease; ALS: Amyotrophic lateral sclerosis; Cont: 'No treatment' group; DEP: Differentially expressed proteins; Glut: 'Glutamic acid excitotoxic injury' treatment group; HD: Huntington's disease; LC-MS/MS: Liquid chromatography-tandem mass spectrometry; PD: Parkinson's disease; R18 + Glut: 'R18-treatment and glutamic acid injury' treatment group; R18: Poly-arginine-18 peptide

Acknowledgements

iTRAQ proteomic labelling and analysis was conducted with the help of Proteomics International Laboratories LTD, Australia.

Authors' contributions

Sample preparation, testing, and collection conducted by GM. iTRAQ protein labelling and LC-MS/MS analysed by Proteomics International Australia Pty Ltd. Bioinformatics analysed and interpreted by GM. RA, FM, and BM were major contributors to the manuscript. Minor edits were provided by NK. All authors read and approved the final manuscript.

Funding

This study was funded in part by a University Postgraduate Award (UPA) from the University of Notre Dame, Australia. Funding support was also provided by Mr. Torsten Ketelsen and the Perron Institute Fundraising Committee.

Availability of data and materials

All data generated or analysed during this study are included in this published article (and its Suppl. information files).

Ethics approval and consent to participate

Approval for the use of E18 Sprague-Dawley rat embryos for isolation of cortical tissue was obtained by the University of Western Australia Animal Ethics Committee (RA/3/100/1432).

Consent for publication

Not applicable.

Competing interests

B.P. Meloni and N.W. Knuckey are named inventors of several patent applications (Provisional Patents: 2013904197; 30/ 10/2013 and 2014902319; 17/6/2014 and PCT/ AU2014/050326; 30/10/2104) regarding the use of arginine-rich peptides as neuroprotective agents. The other authors declare they have no conflict of interest.

Author details

¹Centre for Neuromuscular and Neurological Disorders, The University of Western Australia, Nedlands, Australia. ²Department of Neurosurgery, Sir Charles Gairdner Hospital, QEII Medical Centre, Nedlands, Western Australia, Australia. ³Perron Institute for Neurological and Translational Sciences, QEII Medical Centre, Ground Floor, RR Block, 8 Verdun St, Nedlands, Western Australia 6009, Australia. ⁴School of Health Sciences and Institute for Health Research, The University Notre Dame, Fremantle, Western Australia, Australia.

Received: 6 June 2019 Accepted: 1 July 2019

Published online: 17 July 2019

References

- Xing C, Arai K, Lo EH, Hommel M. Pathophysiologic cascades in ischemic stroke. *Int J Stroke*. 2012. <https://doi.org/10.1111/j.1747-4949.2012.00839.x>.
- Platt SR. The role of glutamate in central nervous system health and disease - a review. *Vet J*. 2007. <https://doi.org/10.1016/j.tvjl.2005.11.007>.
- Talantova M, Sanz-Blasco S, Zhang X, Xia P, Akhtar MW, Okamoto S -i, et al. A β induces astrocytic glutamate release, extrasynaptic NMDA receptor activation, and synaptic loss. *Proc Natl Acad Sci*. 2013. <https://doi.org/10.1073/pnas.1306832110>.
- De Felice FG, Velasco PT, Lambert MP, Viola K, Fernandez SJ, Ferreira ST, et al. A β oligomers induce neuronal oxidative stress through an N-methyl-D-aspartate receptor-dependent mechanism that is blocked by the Alzheimer drug memantine. *J Biol Chem*. 2007. <https://doi.org/10.1074/jbc.M607483200>.
- Zeron MM, Hansson O, Chen N, Wellington CL, Leavitt BR, Brundin P, et al. Increased sensitivity to N-methyl-D-aspartate receptor-mediated excitotoxicity in a mouse model of Huntington's disease. *Neuron*. 2002. [https://doi.org/10.1016/S0896-6273\(02\)00615-3](https://doi.org/10.1016/S0896-6273(02)00615-3).
- Shehadeh J, Fernandes HB, Zeron Mullins MM, Graham RK, Leavitt BR, Hayden MR, et al. Striatal neuronal apoptosis is preferentially enhanced by NMDA receptor activation in YAC transgenic mouse model of Huntington disease. *Neurobiol Dis*. 2006. <https://doi.org/10.1016/j.nbd.2005.08.001>.
- Helton TD, Otsuka T, Lee M-C, Mu Y, Ehlers MD. Pruning and loss of excitatory synapses by the parkin ubiquitin ligase. *Proc Natl Acad Sci*. 2008. <https://doi.org/10.1073/pnas.0802280105>.
- Zádori D, Klivényi P, Plangár I, Toldi J, Vécsei L. Endogenous neuroprotection in chronic neurodegenerative disorders: with particular regard to the kynurenicines. *J Cell Mol Med*. 2011. <https://doi.org/10.1111/j.1582-4934.2010.01237.x>.
- Carriedo SG, Yin HZ, Weiss JH. Motor neurons are selectively vulnerable to AMPA/kainate receptor-mediated injury in vitro. *J Neurosci*. 1996. <https://doi.org/10.1523/jneurosci.2021-10.2010>.
- Van Damme P, Van den Bosch L, Van Houtte E, Callewaert G, Robberecht W. GluR2-dependent properties of AMPA receptors determine the selective vulnerability of motor neurons to excitotoxicity. *J Neurophysiol*. 2002. <https://doi.org/10.1152/jn.00163.2002>.
- Meloni BP, Craig AJ, Milech N, Hopkins RM, Watt PM, Knuckey NW. The neuroprotective efficacy of cell-penetrating peptides TAT, penetratin, Arg-9, and Pep-1 in glutamic acid, kainic acid, and in vitro ischemia injury models using primary cortical neuronal cultures. *Cell Mol Neurobiol*. 2014. <https://doi.org/10.1007/s10571-013-9999-3>.
- Meloni BP, Brookes LM, Clark VW, Cross JL, Edwards AB, Anderton RS, et al. Poly-arginine and arginine-rich peptides are neuroprotective in stroke models. *J Cereb Blood Flow Metab*. 2015. <https://doi.org/10.1038/jcbfm.2015.11>.
- Milani D, Milani D, Cross JL, Clark VW, Edwards AB, Anderton RS, et al. Assessment of the neuroprotective effects of arginine-rich protamine peptides, poly-arginine peptides (R12-cyclic, R22) and arginine-tryptophan-containing peptides following in vitro excitotoxicity and/or permanent middle cerebral artery occlusion in rats. *NeuroMolecular Med*. 2017. <https://doi.org/10.1007/s12017-017-8441-2>.
- Milani D, Bakeberg MC, Cross JL, Clark VW, Anderton RS, Blacker DJ, et al. Comparison of neuroprotective efficacy of poly-arginine R18 and R18D (D-enantiomer) peptides following permanent middle cerebral artery occlusion in the Wistar rat and in vitro toxicity studies. *PLoS One*. 2018. <https://doi.org/10.1371/journal.pone.0193884>.
- Milani D, Cross JL, Anderton RS, Blacker DJ, Knuckey NW, Meloni BP. Neuroprotective efficacy of poly-arginine R18 and NA-1 (TAT-NR2B9c) peptides following transient middle cerebral artery occlusion in the rat. *Neurosci Res*. 2017. <https://doi.org/10.1016/j.neures.2016.09.002>.
- Milani D, Cross JL, Anderton RS, Blacker DJ, Knuckey NW, Meloni BP. Delayed 2-h post-stroke administration of R18 and NA-1 (TAT-NR2B9c) peptides after permanent and/or transient middle cerebral artery occlusion in the rat. *Brain Res Bull*. 2017. <https://doi.org/10.1016/j.brainresbull.2017.09.012>.
- Milani D, Clark VW, Cross JL, Anderton RS, Knuckey NW, Meloni BP. Poly-arginine peptides reduce infarct volume in a permanent middle cerebral artery rat stroke model. *BMC Neurosci*. 2016. <https://doi.org/10.1186/s12868-016-0253-z>.
- Milani D, Knuckey NW, Anderton RS, Cross JL, Meloni BP. The R18 polyarginine peptide is more effective than the TAT-NR2B9c (NA-1) peptide when

- administered 60 minutes after permanent middle cerebral artery occlusion in the rat. *Stroke Res Treat*. 2016. <https://doi.org/10.1155/2016/2372710>.
19. Edwards AB, Cross JL, Anderton RS, Knuckey NW, Meloni BP. Poly-arginine R18 and R18D (D-enantiomer) peptides reduce infarct volume and improves behavioural outcomes following perinatal hypoxic-ischaemic encephalopathy in the P7 rat. *Mol Brain*. 2018. <https://doi.org/10.1186/s13041-018-0352-0>.
 20. Chiu LS, Anderton RS, Cross JL, Clark VW, Edwards AB, Knuckey NW, et al. Assessment of R18, COG1410, and APP96-110 in excitotoxicity and traumatic brain injury. *Transl Neurosci*. 2017. <https://doi.org/10.1515/tnci-2017-0021>.
 21. Chiu LS, Anderton RS, Cross JL, Clark VW, Knuckey NW, Meloni BP. Poly-arginine peptide R18D reduces neuroinflammation and functional deficits following traumatic brain injury in the long-Evans rat. *Int J Pept Res Ther*. 2019. <https://doi.org/10.1007/s10989-018-09799-8>.
 22. MacDougall G, Anderton RS, Edwards AB, Knuckey NW, Meloni BP. The neuroprotective peptide poly-arginine-12 (R12) reduces cell surface levels of NMDA NR2B receptor subunit in cortical neurons; investigation into the involvement of endocytic mechanisms. *J Mol Neurosci*. 2017. <https://doi.org/10.1007/s12031-016-0861-1>.
 23. Szeto HH. Mitochondria-targeted cytoprotective peptides for ischemia-reperfusion injury. *Antioxid Redox Signal*. 2007. <https://doi.org/10.1089/ars.2007.1892>.
 24. Meloni BP, Majda BT, Knuckey NW. Establishment of neuronal in vitro models of ischemia in 96-well microtiter strip-plates that result in acute, progressive and delayed neuronal death. *Neuroscience*. 2001. [https://doi.org/10.1016/S0306-4522\(01\)00396-7](https://doi.org/10.1016/S0306-4522(01)00396-7).
 25. Casey T, Solomon PS, Bringans S, Tan KC, Oliver RP, Lipscombe R. Quantitative proteomic analysis of G-protein signalling in *Stagonospora nodorum* using isobaric tags for relative and absolute quantification. *Proteomics*. 2010. <https://doi.org/10.1002/pmic.200900474>.
 26. Mi H, Poudel S, Muruganujan A, Casagrande JT, Thomas PD. PANTHER version 10: expanded protein families and functions, and analysis tools. *Nucleic Acids Res*. 2016. <https://doi.org/10.1093/nar/gkv1194>.
 27. Meloni BP, Milani D, Edwards AB, Anderton RS, O'Hare Doig RL, Fitzgerald M, et al. Neuroprotective peptides fused to arginine-rich cell penetrating peptides: neuroprotective mechanism likely mediated by peptide endocytic properties. *Pharmacol Ther*. 2015. <https://doi.org/10.1016/j.pharmthera.2015.06.002>.
 28. Zhao K, Zhao GM, Wu D, Soong Y, Birk AV, Schiller PW, et al. Cell-permeable peptide antioxidants targeted to inner mitochondrial membrane inhibit mitochondrial swelling, oxidative cell death, and reperfusion injury. *J Biol Chem*. 2004. <https://doi.org/10.1074/jbc.M402999200>.
 29. Szeto HH, Liu S, Soong Y, Wu D, Darrah SF, Cheng F-Y, et al. Mitochondria-targeted peptide accelerates ATP recovery and reduces ischemic kidney injury. *J Am Soc Nephrol*. 2011. <https://doi.org/10.1681/ASN.2010080808>.
 30. Aono M, Bennett ER, Kim KS, Lynch JR, Myers J, Pearlstein RD, et al. Protective effect of apolipoprotein E-mimetic peptides on -methyl-D-aspartate excitotoxicity in primary rat neuronal-glial cell cultures. *Neuroscience*. 2003. [https://doi.org/10.1016/S0306-4522\(02\)00709-1](https://doi.org/10.1016/S0306-4522(02)00709-1).
 31. MacDougall G, Anderton RS, Mastaglia FL, Knuckey NW, Meloni BP. Mitochondria and neuroprotection in stroke: cationic arginine-rich peptides (CARPs) as a novel class of mitochondria-targeted neuroprotective therapeutics. *Neurobiol Dis*. 2019. <https://doi.org/10.1016/j.nbd.2018.09.010>.
 32. Birk AV, Liu S, Soong Y, Mills W, Singh P, Warren JD, et al. The mitochondrial-targeted compound SS-31 re-energizes ischemic mitochondria by interacting with cardiolipin. *J Am Soc Nephrol*. 2013. <https://doi.org/10.1681/ASN.2012121216>.
 33. Petri S, Kiaei M, Damiano M, Hiller A, Wille E, Manfredi G, et al. Cell-permeable peptide antioxidants as a novel therapeutic approach in a mouse model of amyotrophic lateral sclerosis. *J Neurochem*. 2006. <https://doi.org/10.1111/j.1471-4159.2006.04018.x>.
 34. Gibson TJ, Murphy RM. Design of peptidyl compounds that affect β -amyloid aggregation: importance of surface tension and context. *Biochemistry*. 2005. <https://doi.org/10.1021/bi050225s>.
 35. Fan J, Cowan CM, Zhang LYJ, Hayden MR, Raymond LA. Interaction of postsynaptic density protein-95 with NMDA receptors influences excitotoxicity in the yeast artificial chromosome mouse model of huntington's disease. *J Neurosci*. 2009. <https://doi.org/10.1523/JNEUROSCI.2491-09.2009>.
 36. Cook DR, Gleichman AJ, Cross SA, Doshi S, Ho W, Jordan-Sciutto KL, et al. NMDA receptor modulation by the neuropeptide apelin: implications for excitotoxic injury. *J Neurochem*. 2011. <https://doi.org/10.1111/j.1471-4159.2011.07383.x>.
 37. O'Donnell LA, Agrawal A, Sabnekar P, Dichter MA, Lynch DR, Kolson DL. Apelin, an endogenous neuronal peptide, protects hippocampal neurons against excitotoxic injury. *J Neurochem*. 2007. <https://doi.org/10.1111/j.1471-4159.2007.04645.x>.
 38. Brugnano J, Ward BC, Panitch A. Cell penetrating peptides can exert biological activity: a review. *Biomol Concepts*. 2010. <https://doi.org/10.1515/bmc.2010.016>.
 39. Ross CA, Poirier MA. Protein aggregation and neurodegenerative disease. *Nat Med*. 2004. <https://doi.org/10.1038/nm1066>.
 40. Ciechanover A, Brundin P. The ubiquitin proteasome system in neurodegenerative diseases: sometimes the chicken, sometimes the egg. *Neuron*. 2003. [https://doi.org/10.1016/S0896-6273\(03\)00606-8](https://doi.org/10.1016/S0896-6273(03)00606-8).
 41. Bao J, Sato K, Li M, Gao Y, Abid R, Aird W, et al. PR-39 and PR-11 peptides inhibit ischemia-reperfusion injury by blocking proteasome-mediated I kappa B alpha degradation. *Am J Physiol Heart Circ Physiol*. 2001. <https://doi.org/10.1152/ajpheart.2001.281.6.H2612>.
 42. Anbanandam A, Albarado DC, Tirziu DC, Simons M, Veeraraghavan S. Molecular basis for proline- and arginine-rich peptide inhibition of proteasome. *J Mol Biol*. 2008. <https://doi.org/10.1016/j.jmb.2008.09.021>.
 43. Yang JL, Mukda S, Der CS. Diverse roles of mitochondria in ischemic stroke. *Redox Biol*. 2018. <https://doi.org/10.1016/j.redox.2018.03.002>.
 44. Stout AK, Raphael HM, Kanterewicz BI, Klann E, Reynolds IJ. Glutamate-induced neuron death requires mitochondrial calcium uptake. *Nat Neurosci*. 1998. <https://doi.org/10.1038/1577>.
 45. Castilho RF, Ward MW, Nicholls DG. Oxidative stress, mitochondrial function, and acute glutamate excitotoxicity in cultured cerebellar granule cells. *J Neurochem*. 1999. <https://doi.org/10.1046/j.1471-4159.1999.721394.x>.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more [biomedcentral.com/submissions](https://www.biomedcentral.com/submissions)

