



## Bacterial and Fungal Pathogenesis

**Differential role of *gpaB* and *sidA* gene expressions in relation to virulence in *Aspergillus* species from patients with invasive aspergillosis**Nayereh Ghods<sup>a</sup>, Mehraban Falahati<sup>a,\*</sup>, Maryam Roudbary<sup>a,\*</sup>, Shirin Farahyar<sup>a</sup>, Masoud Shamaei<sup>b</sup>, Mahin Pourabdollah<sup>c</sup>, Farhad Seif<sup>d</sup><sup>a</sup> Iran University of Medical Sciences, School of Medicine, Department of Medical Mycology and Parasitology, Tehran, Iran<sup>b</sup> Clinical Tuberculosis and Epidemiology Research Center, National Research Institute of Tuberculosis and Lung Diseases (NRITLD), Shahid Beheshti University of Medical Sciences, Tehran, Iran<sup>c</sup> Chronic Respiratory Diseases Research Center, National Research Institute of Tuberculosis and Lung Diseases (NRITLD), Shahid Beheshti University of Medical Sciences, Tehran, Iran<sup>d</sup> Iran University of Medical Sciences, School of Medicine, Department of Immunology, Tehran, Iran

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## ABSTRACT

The virulence genes in invasive aspergillosis (IA) have not been analyzed adequately. The present study was designed to evaluate the expression of *gpaB* and *sidA* genes, which are important virulence genes in *Aspergillus* spp. from bronchoalveolar lavage (BAL) samples. Direct examination and culture on Czapek Agar and Sabouraud Dextrose Agar media were performed for 600 BAL specimens isolated from patients with possible aspergillosis. A Galactomannan ELISA assay was also carried out. The expression levels of the *gpaB* and *sidA* genes in isolates were analyzed using quantitative real-time PCR (qRT-PCR). We identified 2 species, including *Aspergillus flavus* (*A. flavus*) and *Aspergillus fumigatus* (*A. fumigatus*) in 25 positive samples for invasive aspergillosis as validated using GM-ELISA. *A. flavus* is the main pathogen threatening transplant recipients and cancer patients worldwide. In this study, *A. flavus* had low levels of the *gpaB* gene expression compared to *A. fumigatus* ( $p=0.006$ ). The highest *sidA* expression was detected in transplant recipients ( $p=0.05$ ). There was no significant correlation between *sidA* expression and underlying disease ( $p=0.15$ ). The *sidA* and *gpaB* gene expression patterns may provide evidence that these virulence genes play important roles in the pathogenicity of *Aspergillus* isolates; however, there are several regulatory genes responsible for the unexpressed *sidA* and *gpaB* genes in the isolates.

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\* Corresponding authors at: Department of Medical Mycology and Parasitology, School of Medicine, Iran University of Medical Sciences, Tehran, Iran.

E-mails: [mehrabanfalahati@yahoo.com](mailto:mehrabanfalahati@yahoo.com) (M. Falahati), [m.roudbary@yahoo.com](mailto:m.roudbary@yahoo.com), [Roudbari.mr@iums.ac.ir](mailto:Roudbari.mr@iums.ac.ir) (M. Roudbary).

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## Introduction

Invasive aspergillosis (IA) is a severe disease and one of the primary causes of mortality among immunocompromised individuals, including cystic fibrosis and cancer patients, as well as transplant recipients.<sup>1</sup> A variety of *Aspergillus* species act as causative agents of IA, but *A. fumigatus* and *A. flavus* are the major causes of the disease. In fact, the small size of their conidia facilitates the entry of fungi into the alveolar cavity.<sup>2–4</sup> If we want to map the gene network, which is associated with the progression of IA in immunocompromised individuals, we will need detailed information related to the virulence gene profiles of *Aspergillus* spp.

Several putative virulence factors contributing to the pathogenesis of aspergillosis,<sup>5</sup> have been identified, including pigment production, adhesion molecules on the cell surface, hydrolytic enzymes and toxin secretion. Additionally, it has been proven that a network of genes is required to support the virulence of the *Aspergillus* spp.<sup>6</sup>

G-proteins, involved in signal transduction pathways, and siderophores are encoded by specific genes in the *Aspergillus* genome<sup>7</sup> and they are essential for *A. fumigatus* to reprogram its physiology in response to the environment and iron uptake.<sup>8,9</sup> G protein signaling and the way by which it is regulated have been studied in a number of models in the past decade. Studies on the pathogenic fungi have opened a new avenue to the concept of the regulatory mechanisms of development, virulence/pathogenicity establishment, mating, and generation of secondary metabolites in different fungi. Certainly, *A. fumigatus* G $\beta$  and G $\gamma$  subunits are inevitable for viability and vegetative growth, because gene deleted mutants are extremely defective in germination and vegetative growth.<sup>10</sup> *gpaB* genes encode G-proteins that have a central role in the integrity, morphogenesis, and virulence of *A. fumigatus*, as well as eliciting the host's immune response. A previous study demonstrated that the *gpaB* gene mutant strain of *Aspergillus* is avirulent in animal-infected IA models.<sup>11</sup>

In a study performed by Gehrke et al. it was shown that deletion of two putative G-protein-coupled receptors, GprC and GprD, caused devastating growth defects. Furthermore, hyphal extension and germination were decreased. They showed that the mutants strain displayed an attenuation of virulence in a murine infection model.<sup>12</sup>

Askew and colleagues showed the gene networks with probable roles in the integrity of the cell wall and environmental change-related signaling pathways.<sup>13</sup> Siderophore proteins are encoded by *sidA* genes and play important roles in several vital processes, including germination, oxidative stress resistance, and iron storage in *Aspergillus* spp.<sup>9,14</sup> Interestingly, it has been shown that *A. fumigatus* exploits *sidA* gene expression for virulence and pathogenicity.<sup>15,16</sup>

Hissen indicated that a rescued strain of *Aspergillus* (Delta *sidA* + *sidA*) exhibited siderophores and had similar growth to the wild type on Iron-limited media. In contrast to the wild-type and other rescued strains, it was reported that the Delta *sidA* strain was virulent in a mouse model of IA. It demonstrates that *sidA* plays a pivotal role in the pathogenesis of *A. fumigatus*.<sup>15</sup> Previous studies have already described the

role of genes in the pathogenesis of *Aspergillus*. Not only it is not well-defined which gene is exactly involved in *Aspergillus* pathogenesis but also the exact function of each gene has remained unknown.<sup>6</sup> The expression of *sidA* and *gpaB* genes in *Aspergillus* spp. was not completely defined in BAL specimens isolated from immunosuppressive subjects. Therefore, regarding the importance of *gpaB* and *sidA* genes in virulence of *Aspergillus* spp. either *in vitro* or *in vivo*. This study focused on *gpaB* and *sidA* gene expressions in *A. fumigatus* and *A. flavus* for the first time, as the primary genes involved (incriminated) in the pathogenesis of *Aspergillus* spp. in BAL samples isolated from immunocompromised and hospitalized subjects predisposed to developing IA.

## Materials and methods

### Patient characteristics

Bronchoalveolar lavage (BAL) samples were collected by a pulmonologist via bronchoscopy from 600 patients with underlying disease and susceptible to IA between 2014 and 2015 at Masih Daneshvari Hospital, Tehran, Iran. The patient populations include cancer patients (leukemia, liver), transplant recipients (bone marrow, lung, and kidney), chronic obstructive pulmonary disease (COPD), hemoptysis (with an immunosuppressive condition), and allergic bronchopulmonary aspergillosis (ABPA) patients taking corticosteroid.<sup>17</sup> All of the participants filled informed consent. The patients' medical records and other data, including age, gender, and type of disease were collected and recorded in a database. These data were kept fully anonymous. The study protocol was approved by the Research Ethics Committee of Iran University of Medical Sciences (IUMS). All BAL samples were collected twice per patient.

### Specimen culture

The BAL samples were immediately transferred to a medical mycology laboratory. The specimens were used in a direct smear with KOH 10% and then cultured on Czapek Agar and Sabouraud Dextrose Agar media (2% glucose, 1% peptone, 0.05% chloramphenicol, 2% agar) (Merck, Germany) for 7 days at 25 °C. The growth rate of *Aspergillus* was checked and the species were identified using conventional phenotyping methods such as colony colors and slide cultures.

### Confirmation of aspergillosis using GM-ELISA

The GM-ELISA (Platelia *Aspergillus*; Bio-Rad, Edmonds, WA) was performed on positive BAL samples, according to the manufacturer's recommendations.<sup>18</sup> Briefly, 100  $\mu$ L of the Platelia treatment solution was added to 300  $\mu$ L of the BAL specimen and heated at 100 °C for 3 min before undergoing centrifugation. The supernatant and the horseradish peroxidase-labeled monoclonal antibody (EBA-2) were incubated at 37 °C in antibody-precoated microplates. The plates were washed and then incubated with a substrate chromogen reaction solution. The reaction was stopped by the use of 100  $\mu$ L of sulfuric acid and after 30 min, the optical density

**Table 1 – Sequence of primers in quantitative real-time PCR.**

| Gene name        | Sequence of primers  |                          | Product size (bp) | T <sub>m</sub> (°C) | Primer efficiency (%) |
|------------------|----------------------|--------------------------|-------------------|---------------------|-----------------------|
|                  | Forward (5'→3')      | Reverse (5'→3')          |                   |                     |                       |
| <i>sidA</i>      | GCGACGATAGCCCATTTGTC | ATTAGCAGGATTAGGATCAAGG   | 155               | 59                  | 98                    |
| <i>gpaB</i>      | CCCGACTTCTATGACGAGC  | CAACGCCTATGCATTGAGCC     | 140               | 60                  | 98                    |
| <i>β-tubulin</i> | CGACAACGAGGCTCTGTACG | CAACTTGCGCAGATCAGAGTTGAG | 200               | 58                  | 99                    |

(OD value) of each well was read using a microplate reader at 450 nm. One well for the negative control, two wells for the cut-off control, and one well for the positive control were used. The results of Galactomannan ELISA assay are expressed as “galactomannan index” (GMI = OD sample/mean OD cut-off control), by comparison to the “cutoff” control. A GMI index of  $\geq 1.0$  in two consecutive samples was considered positive.

#### Culture of *Aspergillus* spp.

For RNA extraction, the *Aspergillus* spp. was cultured in a Yeast Sucrose Broth (YSB) medium (Merck, Germany) at 25 °C for 48–72 h. The optimal growth rate was determined based on the expansion of mycelium on the surface of the medium.

#### RNA extraction, cDNA synthesis, and quantitative real-time PCR

RNA contents within all the samples were extracted using the RNA-X PLUS solution (Cinaclon, cat=RN7713C-s). After disrupting the *Aspergillus* spp. cell wall using liquid nitrogen, the RNA-X PLUS solution was added to the sample. Chloroform was added after centrifugation of liquid at 2000 rpm for 5 min, the aqueous phase was transferred to a new tube. The equal volume of isopropanol added, then the tube was centrifuged at 2000 rpm for 5 min and washed with 75% ethanol, the semidried pellet of RNA was dissolved in DEPC treated water. The RNA was treated with DNase (DNase I) (Invitrogen, Cat No 18068015) to remove genomic DNA contamination. The quality and quantity of the extracted RNA, before and after treatment, were examined using a gel electrophoresis and BioPhotometer plus (Germany), respectively.

First-strand cDNA was synthesized using a high-capacity cDNA reverse transcription kit (Hyper script reverse transcriptase, Gene All, Cat No 601-100), according to the manufacturer's protocol. The cDNA samples were kept at –80 °C until being used. The expression of the *gpaB* and *sidA* genes was quantified by real-time PCR analysis, as previously described.<sup>19</sup> The reactions contained 2  $\mu$ L of template cDNA, 0.6  $\mu$ L of each specific primer (10 pmol) for the *sidA* and *gpaB* genes, 10  $\mu$ L of SYBR Green master mix, and 6.8  $\mu$ L of DEPC water in a final volume of 20  $\mu$ L. The results were normalized using  $\beta$ -tubulin amplifications run on the same plate as the house keeping gene. The real-time PCR system was programmed to an initial denaturation and polymerase activation for 2 min at 95 °C, denaturation for 15 s at 95 °C, and annealing and elongation for 1 min at 62 °C over 35 cycles using a Corbet thermo cycle (RG-6000, Australia). Specific primers for quantitative real-time RT-PCR were designed with online primer design software (Genscript, Piscataway, NJ). For determination of primers efficiency values, we papered the serial dilutions

(25 ng, 5 ng, 1 ng, 0.2 ng, 0.04 ng per microliter) of the positive control for *sidA* and *gpaB* genes. Then, the primers efficiency values were determined using a Corbet thermo cycle (RG-6000, Australia) (Table 1). The specificity of the primers was assessed with genomic DNA from different *Aspergillus* species. The standard species of *A. fumigatus* and *A. flavus* (as control group) were used in relative quantitative method using real-time PCR. The level of *sidA* and *gpaB* expressions was divided into either down-regulation ( $\leq$ mean of gene expression) or up-regulation ( $>$ mean of gene expression), as described previously.<sup>20</sup>

#### Statistical analysis

The mean of gene expression was used as the cut-off for categorization in low and high expression groups. Relative expression was expected using the formula  $2^{-\Delta\Delta Ct}$ . The expression of the *sidA* and *gpaB* genes was normalized to  $\beta$ -tubulin expression. The data were analyzed using SPSS software, version 20 (SPSS, Chicago, IL, USA). The association between the level of *gpaB* and *sidA* expressions and patient characteristics were explored using either Pearson's chi-square or Fisher's exact test. Variations in the expression of *sidA* and *gpaB* between immunosuppressive diseases were assessed using the Mann-Whitney U test. A p-value of  $\leq 0.05$  was considered to be statistically significant.

## Results

#### Population study

Twenty-five positive cases were detected through the screening of 600 BAL samples received from studied patients. The *A. flavus* isolates were found in 17 (68%) of the cases, and *A. fumigatus* isolates were also detected in 8 (32%) of the cases. Twelve (48%) patients were male, and 13 (52%) were female (M/F ratio=0.92); they ranged in age from 40 to 83 years (mean = 59 and median = 60 years). The obtained results from patient's data showed that 9 (36%) of the patients had COPD, while transplant recipients and cancer patients were found in similar frequencies (24%). In addition, hemoptysis and ABPA were detected in only 2 cases (8%) (Table 2). The results of GM-ELISA test indicated that GM antigen was successfully detected in the serum of the patients with positive *Aspergillus* spp. in BAL samples, the result analyzed according to GM index.

#### Analysis of *gpaB* gene expression in patients and its correlation with patient characteristics

For analysis of *gpaB* gene expression, firstly, the expression level of *gpaB* gene in each case was normalized with  $\beta$ -

**Table 2 – Patients' characteristics and correlation between the levels of *sidA* and *gpaB* gene expressions (bold values are significant).**

| Patient data            | Total no. | <i>sidA</i> expression (mean = 2.4) |        | p-Value | <i>gpaB</i> expression (mean = 33) |        | p-Value |
|-------------------------|-----------|-------------------------------------|--------|---------|------------------------------------|--------|---------|
|                         |           | Low                                 | High   |         | Low                                | High   |         |
| <b>Age (mean = 59)</b>  |           |                                     |        |         |                                    |        |         |
| ≤59                     | 14 (56)   | 10 (71)                             | 4 (29) | 0.54    | 13 (93)                            | 1 (7)  | 0.17    |
| >59                     | 11 (44)   | 9 (82)                              | 2 (18) |         | 8 (73)                             | 3 (27) |         |
| <b>Gender</b>           |           |                                     |        |         |                                    |        |         |
| Male                    | 12 (48)   | 9 (75)                              | 3 (25) | 0.9     | 10 (83)                            | 2 (17) | 0.93    |
| Female                  | 13 (52)   | 10 (77)                             | 3 (23) |         | 11 (85)                            | 2 (15) |         |
| <b>Disease</b>          |           |                                     |        |         |                                    |        |         |
| Transplant recipient    | 6 (24)    | 3 (50)                              | 3 (50) | 0.15    | 6 (100)                            | 0 (0)  | 0.07    |
| Cancer                  | 6 (24)    | 6 (100)                             | 0 (0)  |         | 6 (100)                            | 0 (0)  |         |
| COPD <sup>a</sup>       | 9 (36)    | 8 (89)                              | 1 (14) |         | 6 (67)                             | 3 (33) |         |
| Hemoptysis              | 2 (8)     | 1 (50)                              | 1 (50) |         | 1 (50)                             | 1 (50) |         |
| ABPA <sup>b</sup>       | 2 (8)     | 1 (50)                              | 1 (50) |         | 2 (100)                            | 0 (0)  |         |
| <b>Aspergillus spp.</b> |           |                                     |        |         |                                    |        |         |
| <i>flavus</i>           | 17 (68)   | 12 (71)                             | 5 (29) | 0.28    | 17 (100)                           | 0 (0)  | 0.006   |
| <i>fumigatus</i>        | 8 (32)    | 7 (88)                              | 1 (12) |         | 4 (50)                             | 4 (50) |         |

<sup>a</sup> Chronic obstructive pulmonary disease.

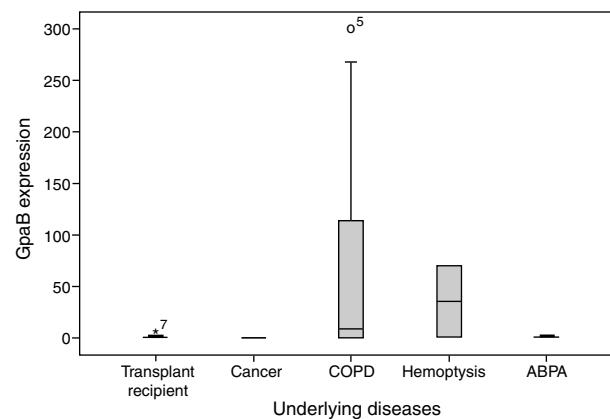
<sup>b</sup> Allergic broncho pulmonary aspergillosis.

tubulin, as housekeeping gene and the final gene expression level has been calculated ( $2^{-\Delta\Delta Ct}$ ). Then, mean value of *gpaB* gene expression was selected as a cut-off value to categorize the level of gene expression. The mean *gpaB* expression (mean = 33) was chosen as a cut-off value to classify the samples into two categories: each case with lower than of mean gene level was considered as the down-regulation expression, and each case with higher than of mean gene level was considered as the up-regulation expression. Down-regulation *gpaB* expression was found mainly in cancer patients and transplant recipients; these same patients showed no instances of up-regulation *gpaB* expression (Table 2).

Correlations between the level of *gpaB* expression and patient characteristics were explored using either Pearson's chi-square or Fisher's exact test. A significant association was observed between *gpaB* expression and *Aspergillus* spp. using fisher exact test, indicating that *A. flavus* expressed lower levels of *gpaB* than *A. fumigatus* ( $p=0.006$ , Table 2). Moreover, a trend became evident between decreased *gpaB* expression and underlying disease using Chi-square test ( $p=0.07$ , Table 2). No correlation was found between *gpaB* expression and other patient characteristics, including age and gender ( $p=0.17$  and  $p=0.93$ , respectively). Among cases, a comparison of the differences between expression using the Mann-Whitney *U* test demonstrated a marginal trend between cancer patients and both hemoptysis patients and transplant recipients ( $p=0.08$  and  $p=0.09$ , respectively). In addition, a marginal trend was found between transplant recipients and hemoptysis patients. ( $p=0.08$ ) (Fig. 1).

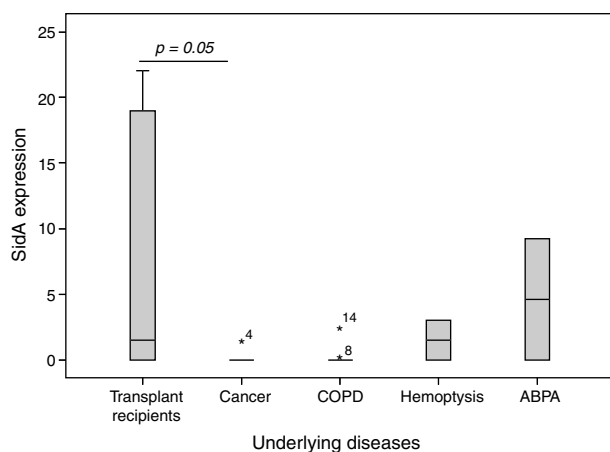
#### Analysis of *sidA* gene expression in patients and its correlation with patient characteristics

For analysis of *sidA* gene expression, firstly, the expression level of *sidA* gene in each case was normalized with  $\beta$ -



**Fig. 1 – Analysis of *gpaB* expression levels in patients using Mann-Whitney *U* test. On the basis of the standard definitions, Box-plot shows median (bold line), interquartile line (box), outliers (circle), and extreme observations (star). COPD, chronic obstructive pulmonary disease; ABPA, allergic broncho pulmonary aspergillosis.**

tubulin, as housekeeping gene and the final gene expression level has been calculated ( $2^{-\Delta\Delta Ct}$ ). Then, mean value of *sidA* gene expression was selected as a cut-off value to categorize the level of gene expression. The mean of *sidA* expression (mean = 2.4) was chosen as a cut-off value to classify the samples into two categories: Each case with lower than of mean gene level was considered as the down-regulation expression, and each case with higher than of mean gene level was considered as the up-regulation expression. Low *sidA* expression was frequently observed in COPD and cancer cases (Table 2). Correlations between the level of *sidA* expression and patient characteristics were explored using either Pearson's chi-square or



**Fig. 2 – Analysis of *sidA* expression levels in patients using Mann–Whitney U test. On the basis of the standard definitions, Box-plot shows median (bold line), interquartile line (box), outliers (circle), and extreme observations (star). COPD, chronic obstructive pulmonary disease; ABPA, allergic broncho pulmonary aspergillosis.**

Fisher's exact test. Down-regulation of *sidA* expression was mainly found in *A. flavus*, but no significant association was found between *sidA* expression and *Aspergillus* spp. ( $p=0.28$ , Table 2). Although down-regulation of *sidA* expression was detected in cancer and COPD patients, no significant correlation was found between *sidA* expression and underlying disease ( $p=0.15$ ). Among the patients, a comparison of the differences between *sidA* expression using the Mann–Whitney U test demonstrated a significant difference between cancer patients and transplant recipients ( $p=0.05$ ), and a marginal trend between cancer patients and both hemoptysis and ABPA patients ( $p=0.08$  and  $p=0.08$ , respectively) (Table 2 and Fig. 2).

#### Correlation between *Aspergillus* spp. and underlying diseases

*A. flavus* was the main pathogen in cancer patients (100%) and transplant recipients (83%). *A. flavus* and *A. fumigatus* play similar roles in other cases, including COPD, hemoptysis, and ABPA.

## Discussion

The incidence of IA has increased in recent years in patients with predisposing factors.<sup>21</sup> *A. fumigatus* and *A. flavus* are known as important agents that can cause IA with enhanced morbidity and mortality in immunocompromised patients, such as chronic obstructive pulmonary disease (COPD) and cancer patients, patients taking corticosteroids and antimicrobial prophylactic drugs, and transplant recipients.<sup>22,23</sup> Due to the suppressed immune systems of such individuals, the *Aspergillus* conidia can pass through the alveolar epithelial cells by conidiation and germination, thereby inducing pathogenicity in patients.<sup>24,25</sup> Furthermore, the expression of

some *Aspergillus* genes has been observed to be changed with extensive prophylaxis with fluconazole, which increases the virulence potency.<sup>26</sup>

The functions and character of the putative virulence factors of *A. fumigatus* are still under investigation. Although several genes have been discovered, there are a lot of other genes which are responsible for the regulation of virulence gene expression. It often makes it impossible to designate with certainty which of them contribute to the mechanism of pathogenesis.<sup>12</sup> Regarding the dominant role of virulence genes in the pathogenesis of IA, there is no comprehensive information concerning gene expression in *Aspergillus*, although a few studies have investigated this issue in recent years.

Our findings demonstrated the up-regulation expression of the *gpaB* gene in *A. fumigatus* compared to *A. flavus*. This may indicate that *gpaB* plays an essential role in the pathogenesis of *A. fumigatus*. G-proteins, as a main component of *gpaB*, have been shown to have a fundamental effect on transduction of the signals into the fungal cells.<sup>27,28</sup> In fact, the activation of G-protein subunits in the cell wall of *A. fumigatus* increases cyclic adenosine monophosphate (cAMP) signaling, mitogen-activated protein kinase (MAPK) and protein kinase A (PKA) pathways in the fungi, and deletion of *A. fumigatus* *GprC* and *GprD* genes lead to growth destruction in all conditions, and an analysis of virulence revealed a significant decrease in virulence and delayed mortality in a murine model of aspergillosis.<sup>12</sup> Down-regulation *gpaB* expression was detected mainly in COPD patients; the patients exhibited no expression of this gene. This finding is supported by the activation of other regulatory mechanisms, such as the MAPK signal pathway, ray proteins, histidine kinases, calcium signaling or signal transduction by other virulence subunits of G-proteins in the pathogenesis of IA.<sup>29</sup>

We observed the highest *sidA* gene expression in transplant recipients and low expression of this gene in cancer and COPD patients. It has been well-established that *sidA* plays a crucial role in virulence due to its high-affinity iron uptake systems in fungi, which triggers pathogenesis in the patients.<sup>30</sup> Additionally, *sidA* genes excite *Aspergillus* growth in lavage fluid, also siderophore deficiency decreases tissue inflammation and invasion.<sup>31</sup>

Consistent with our finding, Gravelat et al. showed *MedA* accomplishes conidiation, because in  $\Delta medA$  strain of *A. fumigatus*, biofilm formation and adherence to pulmonary epithelial, endothelial cells and fibronectin have been impaired *in vitro*. The  $\Delta medA$  strain also had reduced the ability to damage pulmonary epithelial cells. Moreover, *A. fumigatus*  $\Delta medA$  strain displayed low virulence in both an invertebrate and a mammalian model of IA.<sup>32</sup>

The lack of a significant correlation between *sidA* expression and underlying diseases may be related to the activation of other systems for iron uptake by the *Aspergillus* species.<sup>33</sup> The metaloreductases (MR) increased iron uptake upon the initiation of pulmonary IA infections in animal models.<sup>34,35</sup>

It would be beneficial for future studies to investigate *sidA* and *gpaB* gene expressions in other immunocompromised patients, such as HIV and hepatitis patients. In summary, the *sidA* and *gpaB* gene expression patterns in patients indicated

that these genes play important roles in the pathogenicity of *Aspergillus* isolates; nevertheless, several regulatory genes and proteins appeared to be responsible for the unexpressed *sidA* and *gpaB* genes in the isolates, which explain the role of this main mechanism.

## Conclusion

Taken together, our findings indicated that the *A. flavus* was the major pathogen in cancer patients and transplant recipients. A high expression of *gpaB* in COPD samples and an increased expression of *sidA* in transplant recipient may be related to the critical role of predisposing factors in the expression of specific genes in our study population.

## Conflicts of interest

The authors declare no conflicts of interest.

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## REFERENCES

- Pfaller MA, Diekema DJ. Epidemiology of invasive mycoses in North America. *Crit Rev Microbiol.* 2010;36(1):1–53.
- Dagenais TR, Keller NP. Pathogenesis of *Aspergillus fumigatus* in invasive aspergillosis. *Clin Microbiol Rev.* 2009;22(3):447–465.
- Latgé J-P. *Aspergillus fumigatus* and aspergillosis. *Clin Microbiol Rev.* 1999;12(2):310–350.
- Mullins J, Hutcheson PS, Slavin R. *Aspergillus fumigatus* spore concentration in outside air: Cardiff and St Louis compared. *Clin Exp Allergy.* 1984;14(4):351–354.
- Casadevall A. Fungal virulence, vertebrate endothermy, and dinosaur extinction: is there a connection? *Fungal Genet Biol.* 2005;42(2):98–106.
- Abad A. What makes *Aspergillus fumigatus* a successful pathogen? Genes and molecules involved in invasive aspergillosis. *Rev Iberoam Micol.* 2010;27(4):155–182.
- Li H. Glycosylphosphatidylinositol (GPI) anchor is required in *Aspergillus fumigatus* for morphogenesis and virulence. *Mol Microbiol.* 2007;64(4):1014–1027.
- Liebmann B. The cyclic AMP-dependent protein kinase a network regulates development and virulence in *Aspergillus fumigatus*. *Infect Immun.* 2004;72(9):5193–5203.
- Haas H, Eisendle M, Turgeon BG. Siderophores in fungal physiology and virulence. *Annu Rev Phytopathol.* 2008;46:149–187.
- Shin K-S, Kwon N-J, Yu J-H. G $\beta\gamma$ -mediated growth and developmental control in *Aspergillus fumigatus*. *Curr Genet.* 2009;55(6):631.
- Yu J. Heterotrimeric G protein signaling and RGSs in *Aspergillus nidulans*. *J Microbiol Seoul.* 2006;44(2):145.
- Gehrke A. Heptahelical receptors GprC and GprD of *Aspergillus fumigatus* are essential regulators of colony growth, hyphal morphogenesis, and virulence. *Appl Environ Microbiol.* 2010;76(12):3989–3998.
- Askew DS. *Aspergillus fumigatus*: virulence genes in a street-smart mold. *Curr Opin Microbiol.* 2008;11(4):331–337.
- Schrettl M. SreA-mediated iron regulation in *Aspergillus fumigatus*. *Mol Microbiol.* 2008;70(1):27–43.
- Hissen AH. The *Aspergillus fumigatus* siderophore biosynthetic gene *sidA*, encoding L-ornithine N5-oxygenase, is required for virulence. *Infect Immun.* 2005;73(9):5493–5503.
- Schrettl M. Siderophore biosynthesis but not reductive iron assimilation is essential for *Aspergillus fumigatus* virulence. *J Exp Med.* 2004;200(9):1213–1219.
- Ascioglu S, Invasive Fungal Infections Cooperative Group of the European Organization for Research and Treatment of Cancer, Mycoses Study Group of the National Institute of Allergy and Infectious Diseases. Defining opportunistic invasive fungal infections in immunocompromised patients with cancer and hematopoietic stem cell transplants: an international consensus. *Clin Infect Dis.* 2002;34(1):7–14.
- Khodavaysi S. Detection of galactomannan in bronchoalveolar lavage of the intensive care unit patients at risk for invasive aspergillosis. *Curr Med Mycol.* 2015;1(1):12–17.
- Gravelat FN. In vivo analysis of *Aspergillus fumigatus* developmental gene expression determined by real-time reverse transcription-PCR. *Infect Immun.* 2008;76(8):3632–3639.
- Elvidge GP. Concordant regulation of gene expression by hypoxia and 2-oxoglutarate-dependent dioxygenase inhibition the role of HIF-1 $\alpha$ , HIF-2 $\alpha$ , and other pathways. *J Biol Chem.* 2006;281(22):15215–15226.
- Chandrasekar P, Cutright J, Manavathu E. *Aspergillus*: rising frequency of clinical isolation and continued susceptibility to antifungal agents, 1994–1999. *Diagn Microbiol Infect Dis.* 2001;41(4):211–214.
- Rüchel R, Reichard U. Pathogenesis and clinical presentation of aspergillosis. In: *Aspergillus fumigatus*. Karger Publishers; 1999:21–43.
- Samson RA. The genus *Aspergillus* with special regard to the *Aspergillus fumigatus* group. In: *Aspergillus fumigatus*. Karger Publishers; 1999:5–20.
- Dhingra S, Andes D, Calvo AM. VeA regulates conidiation, gliotoxin production, and protease activity in the opportunistic human pathogen *Aspergillus fumigatus*. *Eukaryot Cell.* 2012;11(12):1531–1543.
- Garnacho-Montero J. Isolation of *Aspergillus* spp. from the respiratory tract in critically ill patients: risk factors, clinical presentation and outcome. *Crit Care.* 2005;9(3):R191.
- Singh N. Trends in the epidemiology of opportunistic fungal infections: predisposing factors and the impact of antimicrobial use practices. *Clin Infect Dis.* 2001;33(10):1692–1696.
- Harnett MM, Klaus GG. G protein regulation of receptor signalling. *Immunol Today.* 1988;9(10):315–320.
- Lengeler KB. Signal transduction cascades regulating fungal development and virulence. *Microbiol Mol Biol Rev.* 2000;64(4):746–785.
- Jain R. The MAP kinase MpkA controls cell wall integrity, oxidative stress response, gliotoxin production and iron adaptation in *Aspergillus fumigatus*. *Mol Microbiol.* 2011;82(1):39–53.
- Schrettl M. HapX-mediated adaption to iron starvation is crucial for virulence of *Aspergillus fumigatus*. *PLoS Pathog.* 2010;6(9):e1001124.
- Schrettl M. The crucial role of the *Aspergillus fumigatus* siderophore system in interaction with alveolar macrophages. *Microbes Infect.* 2010;12(12):1035–1041.
- Gravelat FN. *Aspergillus fumigatus* MedA governs adherence, host cell interactions and virulence. *Cell Microbiol.* 2010;12(4):473–488.
- Kosman DJ. Redox cycling in iron uptake, efflux, and trafficking. *J Biol Chem.* 2010;285(35):26729–26735.

34. Blatzer M, Binder U, Haas H. The metalloreductase FreB is involved in adaptation of *Aspergillus fumigatus* to iron starvation. *Fungal Genet Biol.* 2011;48(11):1027–1033.
35. McDonagh A. Sub-telomere directed gene expression during initiation of invasive aspergillosis. *PLoS Pathog.* 2008;4(9):e1000154.