



**TOURO COLLEGE &  
UNIVERSITY SYSTEM**

**Touro Scholar**

---

College of Osteopathic Medicine (TUN)  
Publications and Research

College of Osteopathic Medicine

---

3-5-2019

## **Extracellular Vesicles from Thyroid Carcinoma: The New Frontier of Liquid Biopsy**

Germana Rappa

*Touro University Nevada*, [germana.rappa@tun.touro.edu](mailto:germana.rappa@tun.touro.edu)

Caterina Puglisi

Mark F. Santos

*Touro University Nevada*, [mark.santos@tun.touro.edu](mailto:mark.santos@tun.touro.edu)

Stefano Forte

Lorenzo Memeo

*See next page for additional authors*

Follow this and additional works at: [https://touro scholar.touro.edu/tuncom\\_pubs](https://touro scholar.touro.edu/tuncom_pubs)



Part of the [Diagnosis Commons](#), [Investigative Techniques Commons](#), and the [Neoplasms Commons](#)

---

### **Recommended Citation**

Rappa, G., Puglisi, C., Santos, M. F., Forte, S., Memeo, L., & Lorico, A. (2019). Extracellular vesicles from thyroid carcinoma: The new frontier of liquid biopsy. *International Journal of Molecular Sciences*, 20(5), [Article 1114].

This Article is brought to you for free and open access by the College of Osteopathic Medicine at Touro Scholar. It has been accepted for inclusion in College of Osteopathic Medicine (TUN) Publications and Research by an authorized administrator of Touro Scholar. For more information, please contact [touro.scholar@touro.edu](mailto:touro.scholar@touro.edu).

---

**Authors**

Germana Rappa, Caterina Puglisi, Mark F. Santos, Stefano Forte, Lorenzo Memeo, and Aurelio Loricò



Review

# Extracellular Vesicles from Thyroid Carcinoma: The New Frontier of Liquid Biopsy

Germana Rappa <sup>1,†</sup>, Caterina Puglisi <sup>2,†</sup>, Mark F. Santos <sup>1</sup>, Stefano Forte <sup>2</sup>, Lorenzo Memeo <sup>2</sup>  
and Aurelio Lorico <sup>1,\*</sup> 

<sup>1</sup> College of Medicine, Touro University Nevada, 874 American Pacific Drive, Henderson, NV 89014, USA; germana.rappa@tun.touro.edu (G.R.); mark.santos@tun.touro.edu (M.F.S.)

<sup>2</sup> Mediterranean Institute of Oncology Foundation, Via Penninazzo 7, 95029 Viagrande, Italy; caterina.puglisi@grupposamed.com (C.P.); stefano.forte@grupposamed.com (S.F.); lorenzo.memeo@grupposamed.com (L.M.)

\* Correspondence: aurelio.lorico@tun.touro.edu; Tel.: +1-(702)-777-3942; Fax: +1-(702)-777-1758

† These authors were contributed equally to this work.

Received: 8 February 2019; Accepted: 1 March 2019; Published: 5 March 2019



**Abstract:** The diagnostic approach to thyroid cancer is one of the most challenging issues in oncology of the endocrine system because of its high incidence (3.8% of all new cancer cases in the US) and the difficulty to distinguish benign from malignant non-functional thyroid nodules and establish the cervical lymph node involvement during staging. Routine diagnosis of thyroid nodules usually relies on a fine-needle aspirate biopsy, which is invasive and often inaccurate. Therefore, there is an urgent need to identify novel, accurate, and non-invasive diagnostic procedures. Liquid biopsy, as a non-invasive approach for the detection of diagnostic biomarkers for early tumor diagnosis, prognosis, and disease monitoring, may be of particular benefit in this context. Extracellular vesicles (EVs) are a consistent source of tumor-derived RNA due to their prevalence in circulating bodily fluids, the well-established isolation protocols, and the fact that RNA in phospholipid bilayer-enclosed vesicles is protected from blood-borne RNases. Recent results in other types of cancer, including our recent study on plasma EVs from glioblastoma patients suggest that information derived from analysis of EVs from peripheral blood plasma can be integrated in the routine diagnostic tumor approach. In this review, we will examine the diagnostic and prognostic potential of liquid biopsy to detect tumor-derived nucleic acids in circulating EVs from patients with thyroid carcinoma.

**Keywords:** thyroid carcinoma; papillary thyroid carcinoma; liquid biopsy; cancer; extracellular vesicles; exosomes; diagnostic biomarkers; miRNA

## 1. Introduction

Thyroid cancer is the most common malignancy of the endocrine system, representing 3.8% of all new cancer cases in the United States and the ninth most common cancer overall. Its incidence has risen by an average of 5.5% annually over the past ten years [1]. This increase is almost entirely attributed to an increase in papillary thyroid carcinoma (PTC), which comprises 80% of all thyroid cancers [2,3].

Two important diagnostic problems still exist for thyroid cancer that could greatly benefit from the availability of novel blood biomarkers: (i) Distinction between benign and malignant non-functional thyroid nodules. While 3–7% of the world's population has a palpable nodule, only 5% of those nodules are malignant [2,4]. The initial diagnostic work-up includes the measurement of serum thyroid-stimulating hormone levels, which allows differentiation between functional and non-functional nodules [5]. This is an important characteristic because functional nodules

are rarely malignant. If the initial work-up suggests a non-functional nodule with suspicious sonographic features, an ultrasound-guided fine-needle aspirate biopsy should be performed [4]. However, the results of this approach are often inconclusive and may result in over-diagnosis and over-treatment [3]. (ii) Identification of cervical lymph node involvement during staging. Up to 50% of patients with PTC have cervical lymph node involvement. However, preoperative neck ultrasound imaging identifies only half of the lymph nodes that are found during surgery [4].

For the reasons above, liquid biopsy has great potential in the clinical approach to thyroid cancer. It consists of a test done on a sample of blood or, less frequently, other bodily fluids to look for circulating cancer cells, nucleic acids, or proteins from a tumor. Isolation of genetic materials from bodily fluids is a minimally invasive method to diagnose different types of cancer [6]. Thus, the availability of a validated circulating biomarker to distinguish PTCs localized to the thyroid (T+N<sup>-</sup>) from those with lymph node metastasis (T+N<sup>+</sup>) would allow to plan ahead for a localized surgery versus a surgery that includes lymphadenectomy.

Extracellular vesicles (EVs) are nano-biological units released from most cell types into the extracellular environment. They include exosomes and ectosomes, distinguished on the basis of their biogenesis [7]. Exosomes are released when multi-vesicular bodies fuse with the plasma membrane, whereas ectosomes bud directly from the plasma membrane. Afterward, they are taken up by neighboring or distant recipient cells. Given the difficulty to isolate pure populations of exosomes or ectosomes, we will refer to them collectively as EVs, as recommended by the recent guidelines set by the International Society for Extracellular Vesicles [8]. EV cargo includes lipids, proteins, and nucleic acids—DNA and several types of RNA. Thus, both mRNA and many types of non-coding RNA, including micro RNA (miRNA), long non-coding RNA (lncRNA), circular RNA (circRNA), small nucleolar RNA (snoRNA), small nuclear RNAs (snRNA), transfer RNA (tRNA), and piwi-interacting RNAs (piRNA), have been reported in EVs [6,9].

Diverse physiological functions of EVs are ascribed to cell-to-cell communication, such as favoring cellular differentiation and epithelial–mesenchymal transition, promoting angiogenesis, and modulating immune responses [10,11]. EVs play also an important role in pathological conditions such as cancer. Thus, the EV-mediated crosstalk between cancerous and non-cancerous cells, e.g., those found in the bone marrow microenvironment, can modulate the biochemistry and consequently the function of stromal components to stimulate the growth, expansion, and spread of cancer cells [12]. Several reports support the concept that EVs also contribute to distant intercellular communication in cancer [13,14]. Recently, Hoshino et al. [15] showed that EV-associated integrins determine organ-specific metastasis through a selective adhesion of EVs to extracellular matrix-enriched cellular areas, followed by their uptake by resident cells at their predicted metastatic destination [11].

Recent results in PTC [16], as well as in other types of cancer [17], suggest that information derived from analysis of EVs from peripheral blood plasma can be integrated in the routine diagnostic approach for patients with non-functional thyroid nodules. Specifically, analysis of the RNA content of EVs in plasma, in addition to being a less invasive approach than tissue biopsy, may have an important role in early detection of cancer and in the distinction of benign from malignant nodules, as well as from localized metastatic thyroid cancer. It can also be utilized to assess the prognosis and detection of progression or response to treatment.

## 2. Liquid Biopsy: History, Advantages

As precision medicine is emerging as the new paradigm in oncology, tumor sampling for molecular characterization is becoming an almost mandatory procedure. However, tissue biopsy is not always feasible and repeated sampling is often impossible due to invasiveness of the procedure. This restricts the use of tissue sampling and molecular characterization only to the diagnostic approach, while therapy monitoring is still impracticable. Liquid biopsy has the potential to overcome this limit, extending the benefits of molecular characterization to early diagnosis and cancer monitoring.

This may also produce unprecedented advantages in the treatment of cancer by identifying early events of resistance, relapse, and progression.

Much of the early research on liquid biopsies has been on circulating cell-free DNA (cfDNA) from patients with lung, breast, and prostate cancers. cfDNA was first reported in 1948 when Mandel and Métais demonstrated the presence of both DNA and RNA in plasma from 25 individuals with normal and pathological conditions [18]. Tumor DNA fragments are released into the circulation as a result of programmed cell death or necrosis. cfDNA can be sequenced to help diagnosis, predict relapse, and support clinical decisions for changes in the course of treatment.

The first liquid biopsy test approved by the U.S. Food and Drug Administration for a cancer disease is the blood-based companion diagnostic to select patients with non-small cell lung cancer (NSCLC) for treatment with the epidermal growth factor receptor (EGFR) inhibitor, erlotinib [19]. It can detect gene mutations in EGFR, present in approximately 10–20% of NSCLC patients, who must be identified in order to start an anti-EGFR therapy.

In thyroid cancer, several studies have been conducted on the possible use of cfDNA for prognosis and disease monitoring. For example, the detection of RET M918T in cfDNA has been reported in medullary thyroid carcinoma (MTC) as a specific but not very sensitive event during follow-up [20]. In patients with previously assessed somatic RET mutations, the identification of M918T cfDNA constitutes a prognostic factor for overall survival [18]. While in PTC and MTC the concordance between mutations found in cfDNA and those found within the patient's surgical specimen does not surpass 50%, higher concordance has been suggested in anaplastic thyroid cancer patients, where tumor growth is much more sustained [21]. Even with this assumption, the present studies on the cfDNA-based liquid biopsy approach suggest that this may not be the best strategy to pursue in the clinics.

Circulating cell-free RNAs, like miRNAs, have also been explored as candidates for liquid biopsy in cancer patients [22]. It has been suggested that miRNAs are actively released in the extracellular fluid and in the bloodstream by viable cancer cells [23]. This is particularly interesting in the case of naïve patients because these biomarkers are abundantly released by viable cells before treatment.

The potential of circulating tumor cells (CTCs) in liquid biopsy has been widely investigated in an increasing number of clinical studies [24], demonstrating the huge interest of both the medical and scientific communities. Early during pathogenesis of a solid tumor, cells are continuously released from the primary tumor and disseminate through the bloodstream. These cells, that may be responsible for metastatic dissemination of the tumor, can be detected very early during tumor development, even before the manifestation of disease symptoms. Different technologies have been developed for the isolation of such a rare cell population, but the most frequently adopted is based on the epithelial immunophenotype of CTCs. While normal mesenchymal cells are quite common in the blood, the presence of cells presenting epithelial epitopes, like EpCAM, may be considered an unusual event, possibly related to cancer cell dissemination [25]. CTCs can be immunomagnetically isolated based on the presence of this epitope with the help of a negative selection such as the common leukocyte antigen CD45. This technique aims to enrich the population of CTCs, thus providing a sort of sampling of the original tumor. The limits of this approach are mainly related to the low number of CTCs, which affect the sensitivity of the approach, and to the inability to detect cells from non-epithelial tumors like soft tissue sarcomas or some non-epithelial ovarian cancers. It has been also demonstrated that the expression of epithelial markers is strongly modulated during the detachment of cells from the tumor, making some epitopes less effective for the isolation of some CTC populations [26]. Publications describing the use of extracellular vesicles, circulating tumor cells, cell-free DNA, and cell-free RNA for diagnosis and monitoring of thyroid cancer are listed in Table 1.

### 3. Potential Advantages of EVs for Liquid Biopsy

The potential of EVs and their content as cancer biomarkers is increasingly being recognized. Tumor-derived EVs may be investigated for their protein expression or genetic profile as diagnostic or

prognostic markers [27–30]. A potential drawback of plasma/serum EV analysis is that they contain not only cancer-derived EVs, but also EVs released by blood cells, endothelial cells, stromal cells, and others. Moreover, during neoplastic growth, immune response and the often associated inflammation may alter the rate of release of EVs. Intriguingly, in a recent study by our group [17], orthotopically growing glioblastoma cells were responsible for 35–50% of all circulating EVs. Although unrelated to thyroid cancer, this study indicates that the fraction of circulating plasma EVs derived from cancer cells is highly significant, even in a disease confined to the brain and relatively isolated from the rest of the body by the blood–brain barrier.

### 3.1. Protein Expression Profile

Protein profiling of EVs is challenging because of their small particle size, low abundance of proteins, and heterogeneity. However, using a laboratory-built high-sensitivity flow cytometer, Tian et al. [31] recently reported a quantitative multiparameter analysis of single EVs down to 40 nm with high analysis rate. By this technique, the authors found a significantly elevated level of CD147-positive EVs in colorectal cancer patients compared to healthy controls, thus indicating potential for future proteomics-based development of cancer diagnostic and therapeutic strategies. In another study, by proteomic analysis of EVs from patients with pancreatic cancer, 18 or 14 proteins were found to be up-regulated and 11 or 14 proteins down-regulated compared with EVs from healthy volunteers or from pancreatitis patients, respectively [32]. Also, studies on colorectal cancer cells and malignant mesothelioma identified specific EV proteins that are considered a potential specific signature for these diseases [33,34].

### 3.2. Non-Coding RNA Content

Since the EV membrane protects RNA from blood-borne RNases and EV-associated RNA is generally free of endogenous RNA contaminants such as ribosomal RNA [35], EVs provide a more consistent source of RNA for disease biomarker detection compared with cellular or free plasma RNA. Interestingly, EV-associated miRNAs remain stable for years when EVs are stored at  $-20\text{ }^{\circ}\text{C}$  [36]. The presence of functional RNA in EVs was first described in 2006 for murine stem cell-derived EVs [37] and in 2007 for murine mast cell-derived EVs taken up by human mast cells [27]. In ovarian cancer, a specific exosomal signature consisting of 8 miRNAs has been proposed as surrogate diagnostic for cancer screening in asymptomatic subjects [38]. Another study on EVs of patients with melanoma found a correlation between down-regulation of circulating miR-125b and disease progression [39]. The group of miR-1246, miR-3976, miR-4644, and miR-4306 were up-regulated in EVs from 83% of pancreatic adenocarcinoma patients [40]. Dejima et al. discovered that miR-21 and miR-4257 expression in plasma EVs have potential as predictive biomarkers of recurrence in NSCLC patients [41]. Similarly, circulating EV-associated miR-125a-3p in early-stage colon carcinoma [42], as well as miR-320, miR-574-3p, and RNU6-1 in glioblastoma multiforme [43], have been proposed as diagnostic biomarkers for early detection and monitoring of these specific types of cancer. A comprehensive list of EV-associated miRNAs is available in the miRandola database (<http://mirandola.iit.cnr.it>) [44]. Several studies on esophageal cancer, prostate cancer, and meningioma confirmed the power of EV-associated RNA in cancer diagnosis [45–47]. In the FEMX-I melanoma cell line, we recently reported a higher concentration for 49 miRNAs in EVs than in EV-producing cancer cells, including 20 miRNAs with cancer-related function [48]. A correlation of EV-associated integrin  $\alpha 6\beta 4$  and integrin  $\alpha 6\beta 1$  with lung metastasis, and of integrin  $\alpha V\beta 5$  with liver metastasis were found by Hoshino et al. [15]. Moreover, novel studies point to the clinical relevance of other types of non-coding RNA as more represented in EVs than miRNAs. Nabet and coll. [49] reported the prevalence of non-coding RNA species distinct from miRNAs, in particular signal recognition particles (SRP) RNA in stromal cell-derived EVs released during co-culture with breast cancer cells. As we discussed in a recent Commentary [50], this is surprising given the general consensus in the EV field on the prevalence of coding RNA and miRNA in EVs. In fact, the main EV data repository, “Vesiclopedia” (<https://www.microvesicles.org>), contains

~28,000 entries for mRNAs and ~5,000 entries for miRNAs, but no entries for other non-coding RNAs. Undoubtedly, miRNAs have been studied more in depth than other non-coding RNAs in EVs [27,48,51–53], but the presence of a multitude of other non-coding RNA families in EVs suggests that other types of EV-associated non-coding RNAs, such as SRP RNA, snRNA, snoRNA, and piRNA [52,54], may have clinical potential as biomarkers for thyroid cancer [55]. More studies have shown the presence of tumor RNA in the plasma/serum of cancer patients [56]. These include mRNAs that are correlated with different tumor genes [57–61], tyrosine kinase mRNA, telomerase components, and viral mRNA. The potential value for therapy management of the EV mRNA profile in patients with metastatic breast cancer has been recently reported [62].

#### 4. Potential Advantages of EVs for Liquid Biopsy in Patients with Thyroid Cancer

Although only few studies have been performed on EVs released in the blood by thyroid cancer, recent experimental evidence suggests the involvement of EV miRNAs in thyroid neoplasms, supporting the hypothesis that these non-coding RNAs could be used to develop, refine, or strengthen strategies for diagnosis and management of thyroid cancer. For thyroid cancer patients, EV-based liquid biopsy provides an opportunity to compare a novel tool with the traditional markers used to monitor patients, such as thyroglobulin or calcitonin. While fine-needle aspiration cytology is the gold standard for the differential diagnosis of thyroid nodules [63,64], this procedure has limitations in regard to the discrimination of follicular lesions.

Recent results in other types of cancer, described in the paragraph above, suggest that information derived from analysis of EVs from peripheral blood plasma can be integrated in the routine diagnostic approach to the patient with non-functional thyroid nodules. Moreover, specific alterations of cellular miRNA expression profile have been reported in thyroid carcinoma [65], indicating the possibility that some of these miRNAs, contained in EVs, may be employed as circulating biomarkers. miRNAs in the circulation have been analyzed as potential biomarkers of recurrence in PTC [66]. In many cases in which serum thyroglobulin measurements are difficult to interpret, the analysis of changes in circulating levels of miR-146a-5p and miR-221-3p in PTC patients indicate a good correlation with the American Thyroid Association (ATA)-defined response to therapy classes. Thus, Rosignolo et al. [67] suggested that serum levels of miR-146a-5p and miR-221-3p could be used as complementary biomarkers for the early non-invasive detection of persistent PTC. The association between high circulating levels of miR-146b, miR-222, miR-221, and follicular thyroid proliferation has recently been described [68,69]. Two miRNAs (miR-95, miR-190) were differently expressed in serum of PTC patients. In particular, miR-190 was up-regulated whereas miR-95 was down-regulated, which in combination can be used for the differential diagnosis of thyroid nodules [70].

Other studies have shown that the circulating levels of miR-146b-5p, miR-221-3p, miR-222-3p, and miR-146a-5p were reduced upon tumor excision [67,70–72]. The up-regulated expression of miR-146b-5p, miR-221-3p, and miR-222-3p in the circulation of patients with thyroid cancer has also been demonstrated in PTC [73,74], as well as in anaplastic and follicular thyroid carcinoma [75,76]. Also, Samsonov et al. [71] found that plasma exosomal miR-21 and miR-181a differentiate follicular from PTC.

An analytical approach employing a miRNA-based assay on thyroid fine needle aspirate smears from routinely prepared cytology slides has recently been proposed to improve the diagnostic process [77]. In addition, we have recently proposed a new miRNA-based molecular classification of PTC [73].

**Table 1.** Publications describing the use of extracellular vesicles, circulating tumor cells, cell-free DNA, and cell-free RNA for diagnosis and monitoring of thyroid cancer.

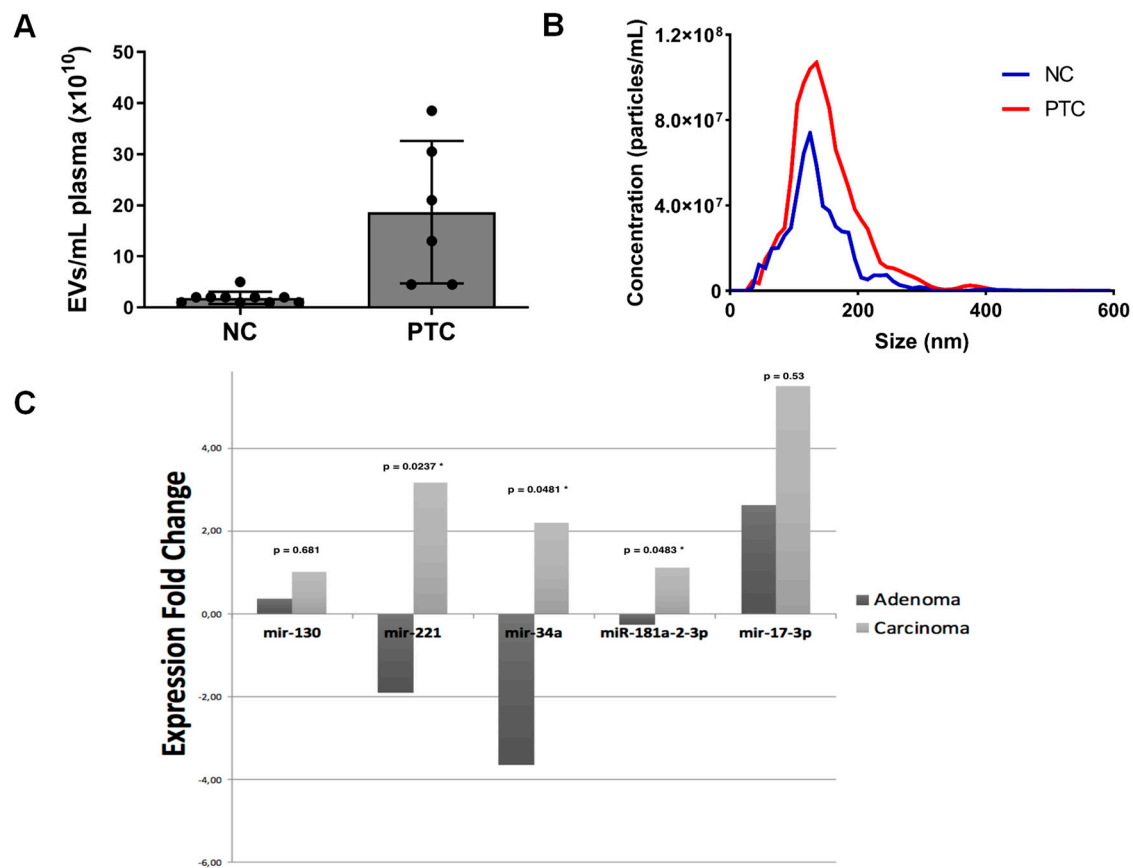
Liquid Biopsy in Thyroid Cancer				
Sample Type	Object	Up-/Down-Regulation	Histotype	References
EV	miR-146b, miR-222	Up	PTC	[66]
	miR-222, miR-142	Up	PTC	[68]
	miR-25-3p, miR-451a, miR-140-3p, let-7	Up	PTC	[70]
	miR-31-5p, miR-126-3p, miR-145-5p, miR-181a miR-21	Up	PTCFTC	[71]
	miR-21, miR-181a-5p	Up	PTC	[73]
	SRC, TLN1, ITGB2, CAPNS1	-	PTC	[78]
	Drug delivery system	-	TC	[79]
	The increase of EPC-EVs and laminins involves folliculogenesis	Up	FTC	[80]
	lncRNAs, linc-ROR	-	PTC	[74]
	lncRNA MALAT1, SLUG, SOX2, and induced EMT	-	PTC	[81]
CTC	Calcitonin-positive CTCs after 12 years	Up	MTC	[82]
	High number of CTCs	Up	DTC DM+	[83]
	CTCs $\geq 5$ is worse OS	-	TC	[84]
	High number of CTCs $\rightarrow$ progressive cancer disease	-	TC	[85]
	PCR detection	-	TC	[86]
cfDNA/ctDNA	BRAF mutation and deregulation miRNA	Up/Down	PTC	[87]
	RET/M91PT mutation	-	MTC	[20]
	cfDNA integrity index	-	TC	[88]
	BRAF mutation	-	TC	[89]
	95% common alteration between cfDNA and tissue DNA	-	FTC	[90]
	BRAF, PIK3CA, NRAS, PTEN, TP53 mutation in cfDNA and tissue DNA	-	ATC	[21]
	BRAF mutation	-	PTC	[91]
	ctDNA panel: 9 cancer gene driver	-	TC	[92]
	BRAFV600	Up	PTC	[93]
	BRAFV600	Up	PTC	[94]
	BRAFV600	Up	DTC	[95]
	cfDNA methylation of $\beta$ -actin, CDH1, DAPK, CALCA, and RAR $\beta$ 2	-	DTC	[96]
	BRAFV600	Up	PTC	[97]
High number	Up	DTC	[98]	
BRAFV600	Up	PTC	[99]	
cfRNA	miR-146a-5p, miR221-3p	Up	PTC	[67]
	let-7e, miR-151-5p, miR-222	Up	PTC	[16]
	miR-579, miR-95, miR-29b, miR-190	Down Up	PTC	[69]
	miR-21, miR-151-5p, miR-222, miR-221	Up	PTC	[72]
	let-7e, miR-151-5p, miR-222	Up	PTC	[16]
miR-146a-5p, miR-150-5p, miR-199b-3p, miR-342-3p	Down	PTC	[100]	

Abbreviations: EV, extracellular vesicles; CTC, circulating tumor cells; cfDNA, cell-free DNA; ctDNA, circulating tumor DNA; cfRNA, cell-free RNA; EPC, endothelial progenitor cells; lncRNA, long non-coding RNA; linc-ROR, long intergenic non-protein coding RNA, regulator of reprogramming; EMT, epithelial-mesenchymal transition; miRNA, micro RNA; PTC, papillary thyroid carcinoma; FTC, follicular thyroid carcinoma; TC, thyroid carcinoma; MTC, medullary thyroid carcinoma; DTC, differentiated thyroid carcinoma; DM+, distant metastasis positive; ATC, anaplastic thyroid carcinoma.

We isolated EVs from blood plasma of patients with PTC before surgery and from age- and sex-matched healthy controls to compare their number and size. Significantly higher numbers of plasma EVs ( $p = 0.025$ ) were found at baseline in thyroid cancer patients ( $n = 6$ ) compared to healthy



controls ( $n = 10$ ), as assessed by nanoparticle tracking analysis (Figure 1A). The average size of EVs was similar (Figure 1B). Thus, EV concentration, not their size, helps distinguish PTC patients from healthy controls. We then performed real-time RT-PCR for five putative thyroid cancer-associated miRNAs on EVs isolated from 0.2 mL of plasma of six patients with PTC, and 4 patients with adenoma of the thyroid. Some of the investigated miRNAs showed an increased expression in PTC patients compared to those with adenoma and healthy controls (Figure 1C). This is particularly evident for miR-34a and miR-17-3p, with the first having a significantly different expression in PTC and normal control and the second showing a significantly augmented expression in carcinoma patients if compared to normal controls ( $p = 0.0476$ ). Both miRNAs have been associated with increased proliferation in PTC [101,102]. However, further studies are needed to confirm that these molecules can be valuable liquid biomarkers in thyroid neoplastic diseases.



**Figure 1.** (A) Scatter plot of extracellular vesicle (EV) plasma concentration in normal controls (NC) and patients with papillary thyroid carcinoma (PTC) by NTA. EVs were isolated by differential centrifugation. (B) Representative size distribution of EVs from plasma of a patient with PTC and a NC. (C) Relative quantitation of miRNAs in patients with thyroid adenoma and carcinoma compared to healthy controls, \*  $p < 0.01$ . Patients' blood samples were collected in disodium EDTA tubes. Plasma samples were prepared by centrifugation and aliquoted into 1.5-mL tubes. EVs were isolated by differential centrifugation as described in Rappa et al. [48]. We measured their plasma concentration and individual size by NTA. Data analysis was performed with NTA 3.0 software. The diffusion coefficient and hydrodynamic radius were determined using the Stokes–Einstein equation, and results displayed as a particle size distribution. Data are presented as the average and standard deviation of six video recordings of 60–90 s per sample. Since NTA is accurate between particle concentrations in the range of  $2 \times 10^7$  to  $2 \times 10^9$  /mL, samples containing higher numbers of particles were diluted before analysis and the relative concentration calculated according to the dilution factor. Silica microspheres of 100 and 200 nm, supplied by Malvern, were used for calibration.

**Author Contributions:** G.R., C.P., M.F.S., S.F., L.M., and A.L. contributed to the text. M.S., G.R., and C.P. performed experiments. A.L. coordinated the review.

**Funding:** This research received no external funding.

**Acknowledgments:** We thank Emmett Findlay and Wolfgang Gilliar for their support and encouragement.

**Conflicts of Interest:** The authors declare no conflicts of interest.

## Abbreviations

EVs	Extracellular vesicles
PTC	Papillary thyroid carcinoma
cfDNA	Cell-free DNA
MTC	Medullary thyroid carcinoma
NSCLC	Non-small cell lung cancer
CTCs	Circulating tumor cell
miRNA	Micro RNA
lncRNA	Long non-coding RNA
circRNA	Circular RNA
snoRNA	Small nucleolar RNA
snRNA	Small nuclear RNA
tRNA	Transfer RNA
piRNA	Piwi-interacting RNA
cfRNA	Cell-free RNA

## References

1. Enewold, L.; Zhu, K.; Ron, E.; Marrogi, A.J.; Stojadinovic, A.; Peoples, G.E.; Devesa, S.S. Rising thyroid cancer incidence in the United States by demographic and tumor characteristics, 1980–2005. *Cancer Epidemiol. Biomark. Prev.* **2009**, *18*, 784–791. [[CrossRef](#)] [[PubMed](#)]
2. Mark, A.; Knox, M.D. Hawaii Island Family Medicine Residency, Hilo, Hawaii. Thyroid nodules. *Am. Fam. Physician* **2013**, *88*, 193–196.
3. Brito, J.P.; Davies, L.; Zeballos-Palacios, C.; Morris, J.C.; Montori, V.M. Papillary lesions of indolent course: Reducing the overdiagnosis of indolent papillary thyroid cancer and unnecessary treatment. *Future Oncol.* **2014**, *10*, 1–4. [[CrossRef](#)] [[PubMed](#)]
4. Cooper, D.S.; Doherty, G.M.; Haugen, B.R.; Kloos, R.T.; Lee, S.L.; Mandel, S.J.; Mazzaferri, E.L.; McIver, B.; Pacini, F.; Scumberger, M.; et al. Revised American Thyroid Association management guidelines for patients with thyroid nodules and differentiated thyroid cancer. *Thyroid* **2009**, *19*, 1167–1214. [[CrossRef](#)] [[PubMed](#)]
5. Nguyen, Q.T.; Lee, E.J.; Huang, M.G.; Park, Y.I.; Khullar, A.; Plodkowski, R.A. Diagnosis and treatment of patients with thyroid cancer. *Am. Health Drug Benefits* **2015**, *8*, 30–40. [[PubMed](#)]
6. Halvaei, S.; Daryani, S.; Eslami-S, Z.; Samadi, T.; Jafarbeik-Iravani, N.; Bakhshayesh, T.O.; Majidzadeh-A, K.; Esmaeili, R. Exosomes in cancer liquid biopsy: A focus on breast cancer. *Mol. Ther. Nucleic Acids* **2018**, *10*, 131–141. [[CrossRef](#)] [[PubMed](#)]
7. Gyorgy, B.; Szabo, T.G.; Pasztoi, M.; Pal, Z.; Misjak, P.; Aradi, B.; Laszlo, V.; Pallinger, E.; Pap, E.; Kittel, A.; et al. Membrane vesicles, current state-of-the-art: Emerging role of extracellular vesicles. *Cell. Mol. Life Sci. CMLS* **2011**, *68*, 2667–2688. [[CrossRef](#)] [[PubMed](#)]
8. Thery, C.; Witwer, K.W.; Aikawa, E.; Alcaraz, M.J.; Anderson, J.D.; Andriantsitohaina, R.; Antoniou, A.; Arab, T.; Archer, F.; Atkin-Smith, G.K.; et al. Minimal information for studies of extracellular vesicles 2018 (MISEV2018): A position statement of the International Society for Extracellular Vesicles and update of the MISEV2014 guidelines. *J. Extracell. Vesicles* **2018**, *7*, 1535750. [[CrossRef](#)] [[PubMed](#)]
9. Kim, K.M.; Abdelmohsen, K.; Mustapic, M.; Kapogiannis, D.; Gorospe, M. RNA in extracellular vesicles. *Wiley Interdiscip. Rev. RNA* **2017**, *8*, 1413. [[CrossRef](#)] [[PubMed](#)]
10. Kowal, J.; Tkach, M.; Thery, C. Biogenesis and secretion of exosomes. *Curr. Opin. Cell Biol.* **2014**, *29*, 116–125. [[CrossRef](#)] [[PubMed](#)]
11. Fujita, Y.; Yoshioka, Y.; Ochiya, T. Extracellular vesicle transfer of cancer pathogenic components. *Cancer Sci.* **2016**, *107*, 385–390. [[CrossRef](#)] [[PubMed](#)]

12. Laurenzana, I.; Lamorte, D.; Trino, S.; De Luca, L.; Ambrosino, C.; Zoppoli, P.; Ruggieri, V.; Del Vecchio, L.; Musto, P.; Caivano, A.; et al. Extracellular Vesicles: A New Prospective in Crosstalk between Microenvironment and Stem Cells in Hematological Malignancies. *Stem Cells Int.* **2018**, *2018*, 9863194. [[CrossRef](#)] [[PubMed](#)]
13. Yáñez-Mó, M.; Siljander, P.R.; Andreu, Z.; Zavec, A.B.; Borràs, F.E.; Buzas, E.I.; Buzas, K.; Casal, E.; Cappello, F.; Carvalho, J.; et al. Biological properties of extracellular vesicles and their physiological functions. *J. Extracell. Vesicles* **2015**, *4*, 27066. [[CrossRef](#)] [[PubMed](#)]
14. Lai, C.P.; Kim, E.Y.; Badr, C.E.; Weissleder, R.; Mempel, T.R.; Tannous, B.A.; Breakefield, X.O. Visualization and tracking of tumour extracellular vesicle delivery and RNA translation using multiplexed reporters. *Nat. Commun.* **2015**, *6*, 7029. [[CrossRef](#)] [[PubMed](#)]
15. Hoshino, A.; Costa-Silva, B.; Shen, T.L. Tumour exosome integrins determine organotropic metastasis. *Nature* **2015**, *527*, 329–335. [[CrossRef](#)] [[PubMed](#)]
16. Yu, S.; Liu, Y.; Wang, J.; Guo, Z.; Zhang, Q.; Yu, F.; Zhang, Y.; Huang, K.; Li, Y.; Song, E.; Zheng, X.L. Circulating microRNA profiles as potential biomarkers for diagnosis of papillary thyroid carcinoma. *J. Clin. Endocrinol. Metab.* **2012**, *97*, 2084–2092. [[CrossRef](#)] [[PubMed](#)]
17. Osti, D.; Del Bene, M.; Rappa, G.; Santos, M.; Matafora, V.; Richichi, C.; Faletti, S.; Beznoussenko, G.V.; Mironov, A.; Bachi, A.; et al. Clinical significance of extracellular vesicles in plasma from glioblastoma patients. *Clin. Cancer Res.* **2019**, *25*, 266–276. [[CrossRef](#)] [[PubMed](#)]
18. Mandel, P.; Metais, P. Les acides nucléiques du plasma sanguin chez l'homme. *C. R. Seances Soc. Biol. Fil.* **1948**, *142*, 241–243. [[PubMed](#)]
19. Kwapisz, D. The first liquid biopsy test approved. Is it a new era of mutation testing for non-small cell lung cancer? *Ann. Transl. Med.* **2017**, *5*, 46. [[CrossRef](#)] [[PubMed](#)]
20. Cote, G.J.; Evers, C.; Hu, M.I.; Grubbs, E.G.; Williams, M.D.; Hai, T.; Duose, D.Y.; Houston, M.R.; Bui, J.H.; Mehrotra, M.; et al. Prognostic significance of circulating *RET* M918T mutated tumor DNA in patients with advanced medullary thyroid carcinoma. *J. Clin. Endocrinol. Metab.* **2017**, *102*, 3591–3599. [[CrossRef](#)] [[PubMed](#)]
21. Sandulache, V.C.; Williams, M.D.; Lai, S.Y.; Lu, C.; William, W.N.; Busaidy, N.L.; Cote, G.J.; Singh, R.R.; Luthra, R.; Cabanillas, M.E. Real-time genomic characterization utilizing circulating cell-free DNA in patients with anaplastic thyroid carcinoma. *Thyroid* **2017**, *27*, 81–87. [[CrossRef](#)] [[PubMed](#)]
22. Trino, S.; Lamorte, D.; Caivano, A.; Laurenzana, I.; Tagliaferri, D.; Falco, G.; Del Vecchio, L.; Musto, P.; De Luca, L. MicroRNAs as New Biomarkers for Diagnosis and Prognosis, and as Potential Therapeutic Targets in Acute Myeloid Leukemia. *Int. J. Mol. Sci.* **2018**, *19*, 460. [[CrossRef](#)] [[PubMed](#)]
23. Fuji, T.; Umeda, Y.; Nyuya, A.; Taniguchi, F.; Kawai, T.; Yasui, K.; Tushima, T.; Yoshida, K.; Fujiwara, T.; Goel, A.; et al. Detection of circulating microRNAs with Ago2 complexes to monitor the tumor dynamics of colorectal cancer patients during chemotherapy. *Int. J. Cancer* **2018**, *142*, 2501–2511. [[CrossRef](#)] [[PubMed](#)]
24. Alix-Panabières, C.; Pantel, K. Circulating tumor cells: Liquid biopsy of cancer. *Clin. Chem.* **2013**, *59*, 110–118. [[CrossRef](#)] [[PubMed](#)]
25. Pantel, K.; Alix-Panabières, C.; Riethdorf, S. Cancer micrometastases. *Nat. Rev. Clin. Oncol.* **2009**, *6*, 339–351. [[CrossRef](#)] [[PubMed](#)]
26. Rao, C.G.; Chianese, D.; Doyle, G.V.; Miller, M.C.; Russell, T.; Sanders, R.A., Jr.; Terstappen, L.W. Expression of epithelial cell adhesion molecule in carcinoma cells present in blood and primary and metastatic tumors. *Int. J. Oncol.* **2005**, *27*, 49–57. [[CrossRef](#)] [[PubMed](#)]
27. Valadi, H.; Ekstrom, K.; Bossios, A.; Sjöstrand, M.; Lee, J.J.; Lötvall, J.O. Exosome-mediated transfer of mRNAs and miRNA is a novel mechanism of genetic exchange between cells. *Nat. Cell Biol.* **2007**, *9*, 654–659. [[CrossRef](#)] [[PubMed](#)]
28. Mitchell, P.S.; Parkin, R.K.; Kroh, E.M.; Fritz, B.R.; Wyman, S.K.; Pogosova-Agadjanyan, E.L.; Peterson, A.; Noteboom, J.; O'Briant, K.C.; Allen, A.; et al. Circulating miRNA as stable blood-based markers for cancer detection. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 10513–10518. [[CrossRef](#)] [[PubMed](#)]
29. Kosaka, N.; Iguchi, H.; Yoshioka, Y.; Takeshita, F.; Matsuki, Y.; Ochiya, T. Secretory mechanisms and intercellular transfer of miRNA in living cells. *J. Biol. Chem.* **2010**, *285*, 17442–17452. [[CrossRef](#)] [[PubMed](#)]
30. Kogure, T.; Lin, W.L.; Yan, I.K.; Braconi, C.; Patel, T. Intercellular nanovesicle-mediated miRNA transfer: A mechanism of environmental modulation of hepatocellular cancer cell growth. *Hepatology* **2011**, *54*, 1237–1248. [[CrossRef](#)] [[PubMed](#)]

31. Tian, Y.; Ma, L.; Gong, M.; Su, G.; Zhu, S.; Zhang, W.; Wang, S.; Li, Z.; Chen, C.; Li, L.; et al. Protein Profiling and Sizing of Extracellular Vesicles from Colorectal Cancer Patients via Flow Cytometry. *ACS Nano* **2018**, *12*, 671–680. [[CrossRef](#)] [[PubMed](#)]
32. Jiao, Y.J.; Jin, D.D.; Jiang, F.; Liu, J.X.; Qu, L.S.; Ni, W.K.; Liu, Z.X.; Lu, C.H.; Ni, R.Z.; Zhu, J.; et al. Characterization and proteomic profiling of pancreatic cancer-derived serum exosomes. *J. Cell Biochem.* **2019**, *120*, 988–999. [[CrossRef](#)] [[PubMed](#)]
33. Mathivanan, S.; Lim, J.W.; Tauro, B.J.; Ji, H.; Moritz, R.L.; Simpson, R.J. Proteomics analysis of A33 immunoaffinity-purified exosomes released from the human colon tumor cell line LIM1215 reveals a tissue-specific protein signature. *Mol. Cell. Proteom.* **2010**, *9*, 197–208. [[CrossRef](#)] [[PubMed](#)]
34. Greening, D.W.; Ji, H.; Chen, M.; Robinson, B.W.; Dick, I.M.; Creaney, J.; Simpson, R.J. Secreted primary human malignant mesothelioma exosome signature reflects oncogenic cargo. *Sci. Rep.* **2016**, *6*, 32643. [[CrossRef](#)] [[PubMed](#)]
35. Cheng, L.; Sharples, R.A.; Scicluna, B.J.; Hill, A.F. Exosomes provide a protective and enriched source of miRNA for biomarker profiling compared to intracellular and cell-free blood. *J. Extracell. Vesicles* **2014**, *3*, 23743. [[CrossRef](#)] [[PubMed](#)]
36. Sanz-Rubio, D.; Martin-Burriel, I.; Gil, A.; Cubero, P.; Forner, M.; Khalyfa, A.; Marin, J.M. Stability of Circulating Exosomal miRNAs in Healthy Subjects. *Sci. Rep.* **2018**, *8*, 10306. [[CrossRef](#)] [[PubMed](#)]
37. Ratajczak, J.; Miekus, K.; Kucia, M.; Zhang, J.; Reza, R.; Dvorak, P.; Ratajczak, M.Z. Embryonic stem cell-derived microvesicles reprogram hematopoietic progenitors: Evidence for horizontal transfer of mRNA and protein delivery. *Leukemia* **2006**, *20*, 847–856. [[CrossRef](#)] [[PubMed](#)]
38. Taylor, D.D.; Gercel-Taylor, C. MicroRNA signatures of tumor-derived exosomes as diagnostic biomarkers of ovarian cancer. *Gynecol. Oncol.* **2008**, *110*, 13–21. [[CrossRef](#)] [[PubMed](#)]
39. Alegre, E.; Sanmamed, M.F.; Rodriguez, C.; Carranza, O.; Martín-Algarra, S.; González, A. Study of circulating microRNA-125b levels in serum exosomes in advanced melanoma. *Arch. Pathol. Lab. Med.* **2014**, *138*, 828–832. [[CrossRef](#)] [[PubMed](#)]
40. Madhavan, B.; Yue, S.; Galli, U.; Rana, S.; Gross, W.; Müller, M.; Giese, N.A.; Kalthoff, H.; Becker, T.; Büchler, M.W.; et al. Combined evaluation of a panel of protein and miRNA serum-exosome biomarkers for pancreatic cancer diagnosis increases sensitivity and specificity. *Int. J. Cancer* **2015**, *136*, 2616–2627. [[CrossRef](#)] [[PubMed](#)]
41. Dejima, H.; Iinuma, H.; Kanaoka, R.; Matsutani, N.; Kawamura, M. Exosomal microRNA in plasma as a non-invasive biomarker for the recurrence of non-small cell lung cancer. *Oncol. Lett.* **2017**, *13*, 1256–1263. [[CrossRef](#)] [[PubMed](#)]
42. Wang, J.; Yan, F.; Zhao, Q.; Zhan, F.; Wang, R.; Wang, L.; Zhang, Y.; Huang, X. Circulating exosomal miR-125a-3p as a novel biomarker for early-stage colon cancer. *Sci. Rep.* **2017**, *7*, 4150. [[CrossRef](#)] [[PubMed](#)]
43. Manterola, L.; Gुरुceaga, E.; Gállego Pérez-Larraya, J.; González-Huarriz, M.; Jauregui, P.; Tejada, S.; Diez-Valle, R.; Segura, V.; Samprón, N.; Barrena, C.; et al. A small noncoding RNA signature found in exosomes of GBM patient serum as a diagnostic tool. *Neuro Oncol.* **2014**, *16*, 520–527. [[CrossRef](#)] [[PubMed](#)]
44. Russo, F.; Di Bella, S.; Nigita, G.; Macca, V.; Laganà, A.; Rosalba, G.; Alfredo, P.; Alfredo, F. miRandola: Extracellular circulating microRNAs database. *PLoS ONE* **2012**, *7*, e47786. [[CrossRef](#)] [[PubMed](#)]
45. Alhasan, A.H.; Scott, A.W.; Wu, J.J.; Feng, G.; Meeks, J.J.; Thaxton, C.S.; Mirkin, C.A. Circulating miRNA signature for the diagnosis of very high-risk prostate cancer. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, 10655–10660. [[CrossRef](#)] [[PubMed](#)]
46. Warnecke-Eberz, U.; Chon, S.H.; Hölscher, A.H.; Drebber, U.; Bollschweiler, E. Exosomal onco-miRs from serum of patients with adenocarcinoma of the esophagus: Comparison of miRNA profiles of exosomes and matching tumor. *Tumour Biol.* **2015**, *36*, 4643–4653. [[CrossRef](#)] [[PubMed](#)]
47. Nilsson, J.; Skog, J.; Nordstrand, A.; Baranov, V.; Mincheva-Nilsson, L.; Breakefield, X.O.; Widmark, A. Prostate cancer-derived urine exosomes: A novel approach to biomarkers for prostate cancer. *Br. J. Cancer* **2009**, *100*, 1603–1607. [[CrossRef](#)] [[PubMed](#)]
48. Rappa, G.; Mercapide, J.; Anzanello, F.; Pope, R.M.; Lorico, A. Biochemical and biological characterization of exosomes containing prominin-1/CD133. *Mol. Cancer* **2013**, *12*, 62. [[CrossRef](#)] [[PubMed](#)]
49. Nabet, B.Y.; Qiu, Y.; Shabason, J.E.; Wu, T.J.; Yoon, T.; Kim, B.C.; Benci, J.L.; DeMichele, A.M.; Tchou, J.; Marcotrigiano, J.; et al. Exosome RNA unshielding couples stromal activation to pattern recognition receptor signaling in cancer. *Cell* **2017**, *170*, 352–366. [[CrossRef](#)] [[PubMed](#)]

50. Rappa, G.; Conigliaro, A.; Santos, M.F.; Alessandro, R.; Lorico, A. Cancer relevance of signal recognition particle and other non-coding RNAs in extracellular vesicles. *Transl. Cancer Res.* **2017**, *6*, S1257–S1260. [[CrossRef](#)]
51. Skog, J.; Wurdinger, T.; van Rijn, S.; Meijer, D.H.; Gainche, L.; Sena-Esteves, M.; Curry, W.T., Jr.; Carter, B.S.; Krichevsky, A.M.; Breakefield, X.O. Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers. *Nat. Cell Biol.* **2008**, *10*, 1470–1476. [[CrossRef](#)] [[PubMed](#)]
52. Nolte-t Hoen, E.N.; Buermans, H.P.; Waasdorp, M.; Stoorvogel, W.; Wauben, M.H.; t Hoen, P.A. Deep sequencing of RNA from immune cell-derived vesicles uncovers the selective incorporation of small non-coding RNA biotypes with potential regulatory functions. *Nucleic Acids Res.* **2012**, *40*, 9272–9285. [[CrossRef](#)] [[PubMed](#)]
53. Raposo, G.; Stoorvogel, W. Extracellular vesicles: Exosomes, microvesicles, and friends. *J. Cell Biol.* **2013**, *200*, 373–383. [[CrossRef](#)] [[PubMed](#)]
54. Liu, Y.; Gu, Y.; Han, Y.; Zhang, Q.; Jiang, Z.; Zhang, X.; Huang, B.; Xu, X.; Zheng, J.; Cao, X. Tumor Exosomal RNAs Promote Lung Pre-metastatic Niche Formation by Activating Alveolar Epithelial TLR3 to Recruit Neutrophils. *Cancer Cell* **2016**, *30*, 243–256. [[CrossRef](#)] [[PubMed](#)]
55. Thind, A.; Wilson, C. Exosomal miRNAs as cancer biomarkers and therapeutic targets. *J. Extracell. Vesicles* **2016**, *5*, 31292. [[CrossRef](#)] [[PubMed](#)]
56. Silva, J.M.; Rodriguez, R.; Garcia, J.M.; Muñoz, C.; Silva, J.; Dominguez, G.; Provencio, M.; España, P.; Bonilla, F. Detection of epithelial tumor RNA in the plasma of colon cancer patients is associated with advanced stages and circulating tumour cells. *Gut* **2002**, *50*, 530–534. [[CrossRef](#)] [[PubMed](#)]
57. Silva, J.M.; Dominguez, G.; Silva, J.; Garcia, J.M.; Sanchez, A.; Rodriguez, O.; Provencio, M.; España, P.; Bonilla, F. Detection of epithelial messenger RNA in the plasma of breast cancer patients is associated with poor prognosis tumor characteristics. *Clin. Cancer Res.* **2001**, *7*, 2821–2825. [[PubMed](#)]
58. Kopreski, M.S.; Benko, F.A.; Gocke, C.D. Circulating RNA as a tumor marker: Detection of 5T4 mRNA in breast and lung cancer patient serum. *Ann. N. Y. Acad. Sci.* **2001**, *945*, 172–178. [[CrossRef](#)] [[PubMed](#)]
59. Gal, S.; Fidler, C.; Lo, Y.M.; Chin, K.; Moore, J.; Harris, A.L.; Wainscoat, J.S. Detection of mammaglobin mRNA in the plasma of breast cancer patients. *Ann. N. Y. Acad. Sci.* **2001**, *945*, 192–194. [[CrossRef](#)] [[PubMed](#)]
60. Fleischhacker, M.; Beinert, T.; Ermitsch, M.; Seferi, D.; Possinger, K.; Engelmann, C.; Jandrig, B. Detection of amplifiable messenger RNA in the serum of patients with lung cancer. *Ann. N. Y. Acad. Sci.* **2001**, *945*, 179–188. [[CrossRef](#)] [[PubMed](#)]
61. Hasselmann, D.O.; Rapp, G.; Rössler, M.; Ugurel, S.; Tilgen, W.; Reinhold, U. Detection of tumor-associated circulating mRNA in serum, plasma and blood cells from patients with disseminated malignant melanoma. *Oncol. Rep.* **2001**, *8*, 115–118. [[CrossRef](#)] [[PubMed](#)]
62. Keup, C.; Mach, P.; Aktas, B.; Tewes, M.; Kolberg, H.C.; Hauch, S.; Sprenger-Haussels, M.; Kimmig, R.; Kasimir-Bauer, S. RNA profiles of circulating tumor cells and extracellular vesicles for therapy stratification of metastatic breast cancer patients. *Clin. Chem.* **2018**, *64*, 1054–1062. [[CrossRef](#)] [[PubMed](#)]
63. Cooper, D.S.; Doherty, G.M.; Haugen, B.R.; Kloos, R.T.; Lee, S.L.; Mandel, S.J.; Mazzaferri, E.L.; McIver, B.; Sherman, S.I.; Tuttle, R.M. Management guidelines for patients with thyroid nodules and differentiated thyroid cancer. *Thyroid* **2006**, *16*, 109–142. [[CrossRef](#)] [[PubMed](#)]
64. Pacini, F.; Schlumberger, M.; Dralle, H.; Elisei, R.; Smit, J.W.; Wiersinga, W.; European Thyroid Cancer Taskforce. European consensus for the management of patients with differentiated thyroid carcinoma of the follicular epithelium. *Eur. J. Endocrinol.* **2006**, *154*, 787–803. [[CrossRef](#)] [[PubMed](#)]
65. Lee, J.C.; Gundara, J.S.; Glover, A.; Serpell, J.; Sidhu, S.B. MicroRNA expression profiles in the management of papillary thyroid cancer. *Oncologist* **2014**, *19*, 1141–1147. [[CrossRef](#)] [[PubMed](#)]
66. Lee, J.C.; Zhao, J.T.; Gundara, J.; Serpell, J.; Bach, L.A.; Sidhu, S. Papillary thyroid cancer derived exosomes contain miRNA-146b and miRNA-222. *J. Surg. Res.* **2015**, *196*, 39–48. [[CrossRef](#)] [[PubMed](#)]
67. Rosignolo, F.; Sponziello, M.; Giacomelli, L. Identification of thyroid-associated serum microRNA profiles and their potential use in thyroid cancer follow-up. *J. Endocr. Soc.* **2017**, *1*, 3–13. [[PubMed](#)]
68. Lee, J.C.; Zhao, J.T.; Clifton-Bligh, R.J.; Gill, A.; Gundara, J.S.; Ip, J.C.; Glover, A.; Sywak, M.S.; Delbridge, L.W.; Robinson, B.G.; et al. MicroRNA-222 and microRNA-146b are tissue and circulating biomarkers of recurrent papillary thyroid cancer. *Cancer* **2013**, *119*, 4358. [[CrossRef](#)] [[PubMed](#)]

69. Cantara, S.; Pilli, T.; Sebastiani, G.; Cevenini, G.; Busonero, G.; Cardinale, S.; Dotta, F.; Pacini, F. Circulating miRNA95 and miRNA190 are sensitive markers for the differential diagnosis of thyroid nodules in a Caucasian population. *J. Clin. Endocrinol. Metab.* **2014**, *99*, 4190–4198. [[CrossRef](#)] [[PubMed](#)]
70. Li, M.; Song, Q.; Li, H.; Lou, Y.; Wang, L. Circulating miR- 25-3p and miR-451a may be potential biomarkers for the diagnosis of papillary thyroid carcinoma. *PLoS ONE* **2015**, *10*, e0135549.
71. Samsonov, R.; Burdakov, V.; Shtam, T.; Radzhabova, Z.; Vasilyev, D.; Tsyrlina, E.; Titov, S.; Ivanov, M.; Berstein, L.; Filatov, M.; et al. Plasma exosomal miR-21 and miR-181a differentiates follicular from papillary thyroid cancer. *Tumour Biol.* **2016**, *37*, 12011–12021. [[CrossRef](#)] [[PubMed](#)]
72. Yoruker, E.E.; Terzioglu, D.; Teksoz, S.; Uslu, F.E.; Gezer, U.; Dalay, N. MicroRNA expression profiles in papillary thyroid carcinoma, benign thyroid nodules and healthy controls. *J. Cancer* **2016**, *7*, 803–809. [[CrossRef](#)] [[PubMed](#)]
73. Rosignolo, F.; Memeo, L.; Monzani, F.; Colarossi, C.; Pecce, V.; Verrienti, A.; Durante, C.; Grani, G.; Lamartina, L.; Forte, S.; et al. MicroRNA-based molecular classification of papillary thyroid carcinoma. *Int. J. Oncol.* **2017**, *50*, 1767–1777. [[CrossRef](#)] [[PubMed](#)]
74. Celano, M.; Rosignolo, F.; Maggisano, V.; Pecce, V.; Iannone, M.; Russo, D.; Bulotta, S. MicroRNAs as biomarkers in thyroid carcinoma. *Int. J. Genom.* **2017**, *2017*, 11. [[CrossRef](#)] [[PubMed](#)]
75. Nikiforova, M.N.; Tseng, G.C.; Steward, D.; Diorio, D.; Nikiforov, Y.E. MicroRNA expression profiling of thyroid tumors: Biological significance and diagnostic utility. *J. Clin. Endocr. Metab.* **2008**, *93*, 1600–1608. [[CrossRef](#)] [[PubMed](#)]
76. Dettmer, M.; Perren, A.; Moch, H.; Komminoth, P.; Nikiforov, Y.E.; Nikiforova, M.N. Comprehensive microRNA expression profiling identifies novel markers in follicular variant of papillary thyroid carcinoma. *Thyroid* **2013**, *23*, 1383–1389. [[CrossRef](#)] [[PubMed](#)]
77. Benjamin, H.; Schnitzer-Perlman, T.; Shtabsky, A.; VandenBussche, C.J.; Ali, S.Z.; Kolar, Z.; Pagni, F.; Rosetta Genomics Group; Bar, D.; Meiri, E. Analytical validity of a microRNA-based assay for diagnosing indeterminate thyroid FNA smears from routinely prepared cytology slides. *Cancer Cytopathol.* **2016**, *124*, 711–721. [[CrossRef](#)] [[PubMed](#)]
78. Luo, D.; Zhan, S.; Xia, W.; Huang, L.; Ge, W.; Wang, T. Proteomics study of serum exosomes from papillary thyroid cancer patients. *Endocr. Relat. Cancer* **2018**, *25*, 879–891. [[CrossRef](#)] [[PubMed](#)]
79. Gangadaran, P.; Li, X.J.; Kalimuthu, S.K.; Min, O.J.; Hong, C.M.; Rajendran, R.L.; Lee, H.W.; Zhu, L.; Baek, S.H.; Jeong, S.Y.; et al. New optical imaging reporter-labeled anaplastic thyroid cancer-derived extracellular vesicles as a platform for *in vivo* tumor targeting in a mouse model. *Sci. Rep.* **2018**, *8*, 13509. [[CrossRef](#)] [[PubMed](#)]
80. Degosserie, J.; Heymans, C.; Spourquet, C.; Halbout, M.; D’Auria, L.; Van Der Smissen, P.; Vertommen, D.; Courtoy, P.J.; Tyteca, D.; Pierreux, C.E. Extracellular vesicles from endothelial progenitor cells promote thyroid follicle formation. *J. Extracell. Vesicles* **2018**, *7*, 1487250. [[CrossRef](#)] [[PubMed](#)]
81. Hardin, H.; Helein, H.; Meyer, K.; Robertson, S.; Zhang, R.; Zhong, W.; Lloyd, R.V. Thyroid cancer stem-like cell exosomes: Regulation of EMT via transfer of lncRNAs. *Lab. Invest.* **2018**, *98*, 1133–1142. [[CrossRef](#)] [[PubMed](#)]
82. Sriramareddy, S.N.; Hamoir, E.; Chavez, M.; Louis, R.; Beckers, A.; Willems, L. Tumor cells may circulate in medullary thyroid cancer patients independently of serum calcitonin. *Endocr. Relat. Cancer* **2018**, *25*, 12. [[CrossRef](#)] [[PubMed](#)]
83. Qiu, Z.L.; Wei, W.J.; Sun, Z.K.; Shen, C.T.; Song, H.J.; Zhang, X.Y.; Zhang, G.Q.; Chen, X.Y.; Luo, Q.Y. Circulating tumor cells correlate with clinicopathological features and outcomes in differentiated thyroid cancer. *Cell. Physiol. Biochem.* **2018**, *48*, 718–730. [[CrossRef](#)] [[PubMed](#)]
84. Xu, J.Y.; Handy, B.; Michaelis, C.L.; Waguespack, S.G.; Hu, M.I.; Busaidy, N.; Jimenez, C.; Cabanillas, M.E.; Fritsche, H.A., Jr.; Cote, G.J.; et al. Detection and prognostic significance of circulating tumor cells in patients with metastatic thyroid cancer. *J. Clin. Endocrinol. Metab.* **2016**, *101*, 4461–4467. [[CrossRef](#)] [[PubMed](#)]
85. Sorg, S.; Pachmann, K.; Brede-Hekimian, K.; Freesmeyer, M.; Winkens, T. Determining tissue origin of circulating epithelial cells (CEC) in patients with differentiated thyroid cancer by real-time PCR using thyroid mRNA probes. *Cancer Lett.* **2015**, *356*, 491–495. [[CrossRef](#)] [[PubMed](#)]
86. Ghossein, R.A.; Carusone, L.; Bhattacharya, S. Polymerase chain reaction detection of micrometastases and circulating tumor cells. *Diagn. Mol. Pathol.* **1999**, *8*, 165–175. [[CrossRef](#)] [[PubMed](#)]

87. Perdas, E.; Stawski, R.; Nowak, D.; Zubrzycka, M. Potential of liquid biopsy in papillary thyroid carcinoma in context of miRNA, BRAF and p53 mutation. *Curr. Drug Targets* **2018**, *19*, 1721–1729. [[CrossRef](#)] [[PubMed](#)]
88. Salvianti, F.; Giuliani, C.; Petrone, L.; Mancini, I.; Vezzosi, V.; Pupilli, C.; Pinzani, P. Integrity and quantity of total cell-free DNA in the diagnosis of thyroid cancer: Correlation with cytological classification. *Int. J. Mol. Sci.* **2017**, *18*, 1350. [[CrossRef](#)] [[PubMed](#)]
89. Fussey, J.M.; Bryant, J.L.; Batis, N.; Spruce, R.J.; Hartley, A.; Good, J.S.; McCabe, C.J.; Boelaert, K.; Sharma, N.; Mehanna, H. The clinical utility of cell-free DNA measurement in differentiated thyroid cancer: A systematic review. *Front. Oncol.* **2018**, *8*, 132. [[CrossRef](#)] [[PubMed](#)]
90. Song, J.; Yang, Z. Whole exome sequencing of circulating cell-free tumor DNA in a follicular thyroid carcinoma patient with lung and bone metastases. *J. Circ. Biomark.* **2018**, *7*. [[CrossRef](#)] [[PubMed](#)]
91. Condello, V.; Macerola, E.; Ugolini, C.; De Napoli, L.; Romei, C.; Materazzi, G.; Elisei, R.; Basolo, F. Analysis of circulating tumor DNA does not improve the clinical management of patients with locally advanced and metastatic papillary thyroid carcinoma. *Head Neck* **2018**, *40*, 1752–1758. [[CrossRef](#)] [[PubMed](#)]
92. Lupo, M.; Guttler, R.; Geck, Z.; Tonozzi, T.R.; Kammesheidt, A.; Braunstein, G.D. Is measurement of circulating tumor DNA of diagnostic use in patients with thyroid nodules? *Endocr. Pract.* **2018**, *24*, 453–459. [[CrossRef](#)] [[PubMed](#)]
93. Pupilli, C.; Pinzani, P.; Salvianti, F.; Fibbi, B.; Rossi, M.; Petrone, L.; Perigli, G.; De Feo, M.L.; Vezzosi, V.; Pazzagli, M.; et al. Circulating BRAFV600E in the diagnosis and follow-up of differentiated papillary thyroid carcinoma. *J. Clin. Endocrinol. Metab.* **2013**, *98*, 3359–3365. [[CrossRef](#)] [[PubMed](#)]
94. Chuang, T.C.; Chuang, A.Y.; Poeta, L.; Koch, W.M.; Califano, J.A.; Tufano, R.P. Detectable BRAF mutation in serum DNA samples from patients with papillary thyroid carcinomas. *Head Neck* **2010**, *32*, 229–234. [[PubMed](#)]
95. Cradic, K.W.; Milosevic, D.; Rosenberg, A.M.; Erickson, L.A.; McIver, B.; Grebe, S.K. Mutant BRAF(T1799A) can be detected in the blood of papillary thyroid carcinoma patients and correlates with disease status. *J. Clin. Endocrinol. Metab.* **2009**, *94*, 5001–5009. [[CrossRef](#)] [[PubMed](#)]
96. Hu, S.; Ewertz, M.; Tufano, R.P.; Brait, M.; Carvalho, A.L.; Liu, D.; Tufano, A.P.; Basaria, S.; Cooper, D.S.; Sidransky, D.; et al. Detection of serum deoxyribonucleic acid methylation markers: A novel diagnostic tool for thyroid cancer. *J. Clin. Endocrinol. Metab.* **2006**, *91*, 98–104. [[CrossRef](#)] [[PubMed](#)]
97. Kim, B.H.; Kim, I.J.; Lee, B.J.; Lee, J.C.; Kim, I.S.; Kim, S.J.; Kim, W.J.; Jeon, Y.K.; Kim, S.S.; Kim, Y.K. Detection of plasma BRAF(V600E) mutation is associated with lung metastasis in papillary thyroid carcinomas. *Yonsei Med. J.* **2015**, *56*, 630–640.
98. Zane, M.; Agostini, M.; Enzo, M.V.; Casal Ide, E.; Del Bianco, P.; Torresan, F.; Merante Boschin, I.; Pennelli, G.; Saccani, A.; Rubello, D.; et al. Circulating cell-free DNA, SLC5A8 and SLC26A4 hypermethylation, BRAF(V600E): A non-invasive tool panel for early detection of thyroid cancer. *Biomed. Pharmacother.* **2013**, *67*, 723–730. [[CrossRef](#)] [[PubMed](#)]
99. Patel, K.B. Detection of Circulating Thyroid Tumor DNA in Patients with Thyroid Nodules. Master's Thesis, The University of Western Ontario, London, ON, Canada, 2015.
100. Graham, M.E.; Hart, R.D.; Douglas, S.; Makki, F.M.; Pinto, D.; Butler, A.L.; Bullock, M.; Rigby, M.H.; Trites, J.R.; Taylor, S.M.; et al. Serum microRNA profiling to distinguish papillary thyroid cancer from benign thyroid masses. *J. Otolaryngol. Head Neck Surg.* **2015**, *44*, 33. [[CrossRef](#)] [[PubMed](#)]
101. Ma, Y.; Qin, H.; Cui, Y. MiR-34a targets GAS1 to promote cell proliferation and inhibit apoptosis in papillary thyroid carcinoma via PI3K/Akt/Bad pathway. *Biochem. Biophys. Res. Commun.* **2013**, *441*, 958–963. [[CrossRef](#)] [[PubMed](#)]
102. Takakura, S.; Mitsutake, N.; Nakashima, M.; Namba, H.; Saenko, V.A.; Rogounovitch, T.I.; Nakazawa, Y.; Hayashi, T.; Ohtsuru, A.; Yamashita, S. Oncogenic role of miR-17-92 cluster in anaplastic thyroid cancer cells. *Cancer Sci.* **2008**, *99*, 1147–1154. [[CrossRef](#)] [[PubMed](#)]

