



Original Investigation | Geriatrics

# Association of Altered Liver Enzymes With Alzheimer Disease Diagnosis, Cognition, Neuroimaging Measures, and Cerebrospinal Fluid Biomarkers

Kwangsik Nho, PhD; Alexandra Kueider-Paisley, PhD; Shahzad Ahmad, MSc; Siamak MahmoudianDehkordi, PhD; Matthias Arnold, PhD; Shannon L. Risacher, PhD; Gregory Louie, MS; Colette Blach, MS; Rebecca Baillie, PhD; Xianlin Han, PhD; Gabi Kastenmüller, PhD; John Q. Trojanowski, MD, PhD; Leslie M. Shaw, PhD; Michael W. Weiner, MD; P. Murali Doraiswamy, MBBS; Cornelia van Duijn, PhD; Andrew J. Saykin, PsyD; Rima Kaddurah-Daouk, PhD; for the Alzheimer's Disease Neuroimaging Initiative and the Alzheimer Disease Metabolomics Consortium

## Abstract

**IMPORTANCE** Increasing evidence suggests an important role of liver function in the pathophysiology of Alzheimer disease (AD). The liver is a major metabolic hub; therefore, investigating the association of liver function with AD, cognition, neuroimaging, and CSF biomarkers would improve the understanding of the role of metabolic dysfunction in AD.

**OBJECTIVE** To examine whether liver function markers are associated with cognitive dysfunction and the "A/T/N" (amyloid, tau, and neurodegeneration) biomarkers for AD.

**DESIGN, SETTING, AND PARTICIPANTS** In this cohort study, serum-based liver function markers were measured from September 1, 2005, to August 31, 2013, in 1581 AD Neuroimaging Initiative participants along with cognitive measures, cerebrospinal fluid (CSF) biomarkers, brain atrophy, brain glucose metabolism, and amyloid- $\beta$  accumulation. Associations of liver function markers with AD-associated clinical and A/T/N biomarkers were assessed using generalized linear models adjusted for confounding variables and multiple comparisons. Statistical analysis was performed from November 1, 2017, to February 28, 2019.

**EXPOSURES** Five serum-based liver function markers (total bilirubin, albumin, alkaline phosphatase, alanine aminotransferase, and aspartate aminotransferase) from AD Neuroimaging Initiative participants were used as exposure variables.

**MAIN OUTCOMES AND MEASURES** Primary outcomes included diagnosis of AD, composite scores for executive functioning and memory, CSF biomarkers, atrophy measured by magnetic resonance imaging, brain glucose metabolism measured by fludeoxyglucose F 18 ( $^{18}\text{F}$ ) positron emission tomography, and amyloid- $\beta$  accumulation measured by [ $^{18}\text{F}$ ]florbetapir positron emission tomography.

**RESULTS** Participants in the AD Neuroimaging Initiative ( $n = 1581$ ; 697 women and 884 men; mean [SD] age, 73.4 [7.2] years) included 407 cognitively normal older adults, 20 with significant memory concern, 298 with early mild cognitive impairment, 544 with late mild cognitive impairment, and 312 with AD. An elevated aspartate aminotransferase (AST) to alanine aminotransferase (ALT) ratio and lower levels of ALT were associated with AD diagnosis (AST to ALT ratio: odds ratio, 7.932 [95% CI, 1.673-37.617];  $P = .03$ ; ALT: odds ratio, 0.133 [95% CI, 0.042-0.422];  $P = .004$ ) and poor cognitive performance (AST to ALT ratio:  $\beta$  [SE],  $-0.465$  [0.180];  $P = .02$  for memory composite score;  $\beta$  [SE],  $-0.679$  [0.215];  $P = .006$  for executive function composite score; ALT:  $\beta$  [SE], 0.397 [0.128];

(continued)

## Key Points

**Question** Are liver function markers associated with cognition and the "A/T/N" (amyloid, tau, and neurodegeneration) biomarkers for Alzheimer disease?

**Findings** In this cohort study of 1581 older adults, elevated aspartate aminotransferase to alanine aminotransferase ratios were associated with diagnosis of Alzheimer disease, poor cognition, lower cerebrospinal fluid levels of amyloid- $\beta$  1-42, increased amyloid- $\beta$  deposition, higher cerebrospinal fluid levels of phosphorylated tau and total tau, and reduced brain glucose metabolism. Lower levels of alanine aminotransferase were associated with increased amyloid- $\beta$  deposition, reduced brain glucose metabolism, greater brain atrophy, diagnosis of Alzheimer disease, and poor cognition.

**Meaning** Consistent associations of serum-based liver function markers with Alzheimer disease biomarkers highlight the involvement of metabolic disturbances in the pathophysiology of Alzheimer disease.

## + Supplemental content

Author affiliations and article information are listed at the end of this article.

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Abstract (continued)

$P = .006$  for memory composite score;  $\beta$  [SE], 0.637 [0.152];  $P < .001$  for executive function composite score). Increased AST to ALT ratio values were associated with lower CSF amyloid- $\beta$  1-42 levels ( $\beta$  [SE], -0.170 [0.061];  $P = .04$ ) and increased amyloid- $\beta$  deposition (amyloid biomarkers), higher CSF phosphorylated tau<sub>181</sub> ( $\beta$  [SE], 0.175 [0.055];  $P = .02$ ) (tau biomarkers) and higher CSF total tau levels ( $\beta$  [SE], 0.160 [0.049];  $P = .02$ ) and reduced brain glucose metabolism ( $\beta$  [SE], -0.123 [0.042];  $P = .03$ ) (neurodegeneration biomarkers). Lower levels of ALT were associated with increased amyloid- $\beta$  deposition (amyloid biomarkers), and reduced brain glucose metabolism ( $\beta$  [SE], 0.096 [0.030];  $P = .02$ ) and greater atrophy (neurodegeneration biomarkers).

**CONCLUSIONS AND RELEVANCE** Consistent associations of serum-based liver function markers with cognitive performance and A/T/N biomarkers for AD highlight the involvement of metabolic disturbances in the pathophysiology of AD. Further studies are needed to determine if these associations represent a causative or secondary role. Liver enzyme involvement in AD opens avenues for novel diagnostics and therapeutics.

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## Introduction

Metabolic activities in the liver determine the state of the metabolic readout of peripheral circulation. Mounting evidence suggests that patients with Alzheimer disease (AD) display metabolic dysfunction.<sup>1</sup> Clinical studies suggest that impaired signaling, energy metabolism, inflammation, and insulin resistance play a role in AD.<sup>2,3</sup> This observation is in line with the observation that many metabolic disorders (eg, diabetes, hypertension, obesity, and dyslipidemia) are risk factors for AD.<sup>4</sup> This evidence highlights the importance of the liver in the pathophysiological characteristics of AD. Focused investigation to assess the role of liver function in AD and its endophenotypes is required to bridge the gap between these observations.

Peripheral blood levels of biochemical markers including albumin, alkaline phosphatase, alanine aminotransferase (ALT), aspartate aminotransferase (AST), and total bilirubin are used to assess liver function. Alanine aminotransferase and AST are used in general clinical practice to measure liver injury<sup>5,6</sup> and are factors associated with cardiovascular and metabolic diseases,<sup>7,8</sup> known risk factors of AD and cognitive decline.<sup>9,10</sup> Given this fact, it is conceivable that aminotransferases are surrogate biomarkers of liver metabolic functioning. A systematic search yielded few reports related to research in humans linking peripheral biomarkers of liver functioning to central biomarkers related to AD including amyloid- $\beta$  and tau accumulation, brain glucose metabolism, and structural atrophy.

We investigated the association of peripheral liver function markers with AD diagnosis, cognition, and biomarkers of AD pathophysiological characteristics including neuroimaging (magnetic resonance imaging [MRI] and position emission tomography [PET]) and cerebrospinal fluid (CSF) from older adults in the AD Neuroimaging Initiative (ADNI) cohort. The AD biomarkers were selected and defined consistent with the National Institute on Aging-Alzheimer Association Research Framework (amyloid, tau, and neurodegeneration [A/T/N]) for AD biomarkers that defines 3 general groups of biomarkers based on the nature of pathologic process that each measures.<sup>11</sup>

## Methods

### Study Population

Individuals in this study were participants of ADNI. The initial phase (ADNI-1) was launched in 2003 to test whether serial MRI markers, PET markers, other biological markers, and clinical and neuropsychological assessment could be combined to measure the progression of mild cognitive

impairment (MCI) and early AD. The initial phase was extended to subsequent phases (ADNI-GO, ADNI-2, and ADNI-3) for follow-up of existing participants and additional new enrollments. Inclusion and exclusion criteria, clinical and neuroimaging protocols, and other information are reported elsewhere.<sup>12-14</sup> Demographic and clinical information, raw data from neuroimaging scans, CSF biomarkers, information on *APOE* status, and cognitive scores were downloaded from the ADNI data repository.<sup>12</sup> Baseline data were collected from September 1, 2005, to August 31, 2013. Written informed consent was obtained at enrollment, which included permission for analysis and data sharing. This study was approved by each participating site's institutional review board. This report followed the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) reporting guidelines for cohort studies.

### Liver Function Markers

Five laboratory tests were downloaded from the ADNI data repository and used in the study: total bilirubin, albumin, alkaline phosphatase, ALT, and AST. The liver function markers followed a normal distribution after log transformation. For each marker, participants with values greater or smaller than 4 SDs from its mean value were considered outliers and were removed. To determine if outliers had a significant effect on our findings we performed a sensitivity analysis and observed few differences (or slightly more significant), if any, in results when including outliers (eTable 1 in the Supplement).

### Dementia Diagnosis

Participants in ADNI were classified as cognitively normal controls (CN) or having significant memory concerns (SMC), MCI, or mild clinical AD. Criteria for classification were as follows: Mini-Mental State Examination score range (range, 0 [worst] to 30 [best]) for CN and MCI was 24 to 30, and for AD was 20 to 26; and overall Clinical Dementia Rating score (range for each, 0 [best] to 3 [worst]) for CN was 0, for MCI was 0.5 with a mandatory requirement of memory box score of 0.5 or greater, and for AD was 0.5 or 1.<sup>15</sup> Cognitively normal controls did not have any significant impairment in cognition or activities of daily living. Participants with SMC had normal cognition and no significant impairment in activities of daily living, but had a score of 16 or more on the first 12 items of the self-report version of the Cognitive Change Index (range, 12 [no change] to 60 [severe change]).<sup>16</sup> Participants with MCI had cognitive impairments in memory and/or other domains but were able to perform activities of daily living and did not qualify for a diagnosis of dementia.<sup>15</sup> Participants with AD had to meet the National Institute of Neurological and Communicative Disorders and Stroke-AD and Related Disorders Association criteria for probable AD.<sup>17</sup> Participants from the ADNI-1 cohort with MCI were all classified as late MCI, with a memory impairment approximately 1.5 SD below education-adjusted norms. In the ADNI-GO and ADNI-2 cohort, participants with MCI were classified as either early MCI, with a memory impairment approximately 1 SD below education-adjusted norms, or late MCI (same criteria as in ADNI-1). Both ADNI-1 and ADNI-GO and ADNI-2 participants met the criteria for amnesic MCI, but many in the ADNI-GO and ADNI-2 cohort included the earlier stage MCI designation (ie, early MCI).<sup>18</sup>

### Cognition

Composite scores were used to measure memory and executive functioning. A memory composite score was created from the following: memory tasks from the Alzheimer Disease Assessment Scale-cognitive subscale, the Rey Auditory Verbal Learning Test, memory components of the Mini-Mental State Examination, and the Logical Memory task.<sup>19</sup> An executive function composite score included the following: Wechsler Adult Intelligence Scale-Revised Digit Symbol Substitution task and Digit Span backward task, Trail Making Test Parts A and B, category fluency (animals and vegetables), and 5 clock drawing items. Composite scores have a mean of 0 and an SD of 1.<sup>20</sup>

## Neuroimaging Processing

### MRI Scans

Baseline T1-weighted brain MRI scans were acquired using a sagittal 3-dimensional magnetization prepared rapid gradient echo scans following the ADNI MRI protocol.<sup>21,22</sup> As previously detailed, FreeSurfer, version 5.1, a widely used automated MRI analysis approach, was used to process MRI scans and extract whole-brain and region-of-interest (ROI)-based neuroimaging endophenotypes including volumes and cortical thickness determined by automated segmentation and parcellation.<sup>23-25</sup> The cortical surface was reconstructed to measure thickness at each vertex. The cortical thickness was calculated by taking the Euclidean distance between the gray and white boundary and the gray and CSF boundary at each vertex on the surface.<sup>26-28</sup>

### PET Scans

Preprocessed fludeoxyglucose (FDG) F 18 (<sup>18</sup>F) and [<sup>18</sup>F]florbetapir PET scans (coregistered, averaged, standardized image and voxel size, and uniform resolution) were downloaded from the ADNI Laboratory of Neuro Imaging (LONI) site<sup>12</sup> as described in previously reported methods for acquisition and processing of PET scans.<sup>23,29</sup> For [<sup>18</sup>F]FDG-PET, scans were intensity normalized using a pons ROI to create [<sup>18</sup>F]FDG standardized uptake value ratio (SUVR) images. For [<sup>18</sup>F]florbetapir PET, scans were intensity normalized using a whole cerebellum reference region to create SUVR images.

### CSF Biomarkers

The ADNI generated CSF biomarkers (amyloid- $\beta$  1-42, total tau [t-tau], and phosphorylated tau<sub>181</sub> [p-tau<sub>181</sub>]) in pristine aliquots of 2401 ADNI CSF samples using the validated and highly automated Roche Elecsys electrochemiluminescence immunoassays<sup>30,31</sup> and the same reagent lot for each of these 3 biomarkers. Cerebrospinal fluid biomarker data were downloaded from the ADNI LONI site.<sup>12</sup>

## Statistical Analysis

Statistical analysis was conducted from November 1, 2017, to February 28, 2019. Logistic regression analysis was performed to explore the diagnostic group differences between AD diagnosis and each liver function marker separately. Age, sex, body mass index (BMI), and *APOE*  $\epsilon$ 4 status were used as covariates. We performed a linear regression analysis to assess the association of liver function markers with composite scores for memory and executive functioning using age, sex, years of education, BMI, and *APOE*  $\epsilon$ 4 status as covariates. We also performed a linear regression analysis using age, sex, BMI, and *APOE*  $\epsilon$ 4 status as covariates.

### ROI-Based Analysis of Structural MRI and PET Scans

Mean hippocampal volume was used as an MRI-related phenotype. For FDG-PET, a mean SUVR value was extracted from a global cortical ROI representing regions where patients with AD show decreased glucose metabolism relative to CN participants from the full ADNI-1 cohort, normalized to pons.<sup>29</sup> For [<sup>18</sup>F]florbetapir PET, a mean SUVR value was extracted using MarsBaR from a global cortical region generated from an independent comparison of ADNI-1 [<sup>11</sup>C] Pittsburgh Compound B SUVR scans (regions where AD > CN). We performed a linear regression analysis using age, sex, BMI, and *APOE*  $\epsilon$ 4 status as covariates to evaluate the association of liver function markers with AD-related endophenotypes from MRI and PET scans. For hippocampal volume, years of education, intracranial volume, and magnetic field strength were added as additional covariates.<sup>32</sup>

### Whole-Brain Imaging Analysis

The SurfStat software package<sup>33</sup> was used to perform a multivariable analysis of cortical thickness to examine the association of liver function markers with brain structural changes on a vertex-by-vertex basis using a general linear model approach.<sup>28</sup> General linear models were developed using age, sex, years of education, intracranial volume, BMI, *APOE*  $\epsilon$ 4 status, and magnetic field strength as

covariates. The processed FDG-PET and [<sup>18</sup>F]florbetapir PET images were used to perform a voxelwise statistical analysis of the association of liver function markers with brain glucose metabolism and amyloid- $\beta$  accumulation across the whole brain using SPM8.<sup>34</sup> We performed a multivariable regression analysis using age, sex, BMI, and APOE  $\epsilon$ 4 status as covariates. In the whole-brain surface-based analysis, the adjustment for multiple comparisons was performed using the random field theory correction method with  $P < .05$  adjusted as the level for significance.<sup>35-37</sup> In the voxelwise whole-brain analysis, the significant statistical parameters were selected to correspond to a threshold of  $P < .05$  (false discovery rate [FDR]-corrected).<sup>38</sup>

### Multiple Testing Correction

Results of the analysis of liver function markers with AD diagnosis groups, cognitive composite measures, and A/T/N biomarkers for AD separately were corrected for multiple testing using the FDR with the Benjamini-Hochberg procedure (p.adjust command in R [R Project for Statistical Computing]).

## Results

### Study Sample

Our analyses included 1581 ADNI participants (407 CN, 20 with SMC, 298 with early MCI, 544 with late MCI, and 312 with AD). Demographic information as well as mean and SD of liver function markers stratified by clinical diagnosis are presented in eTable 2 in the [Supplement](#).

### Diagnostic Group Difference of Liver Function Markers With AD Diagnosis

Levels of ALT were significantly decreased in AD compared with CN (odds ratio, 0.133; 95% CI, 0.042-0.422;  $P = .004$ ) (**Table 1**), while AST to ALT ratio values were significantly increased in AD (odds ratio, 7.932; 95% CI, 1.673-37.617;  $P = .03$ ). There was a trend to suggest that ALT levels were increased and AST to ALT ratio values were decreased in MCI compared with CN, but these became nonsignificant after adjustment for multiple comparisons (eTable 3 in the [Supplement](#)).

### Cognition

After adjusting for multiple comparison correction using FDR, we identified significant associations of liver function markers with cognition (**Table 2**). Higher levels of alkaline phosphatase and AST to ALT ratio were associated with lower memory scores (alkaline phosphatase:  $\beta$  [SE], -0.416 [0.162];  $P = .02$ ; AST to ALT ratio:  $\beta$  [SE], -0.465 [0.180];  $P = .02$ ) and executive functioning scores (alkaline phosphatase:  $\beta$  [SE], -0.595 [0.193];  $P = .006$ ; AST to ALT ratio:  $\beta$  [SE], -0.679 [0.215];  $P = .006$ ). Higher ALT levels were associated with higher memory scores ( $\beta$  [SE], 0.397 [0.128];  $P = .006$ ) and executive functioning scores ( $\beta$  [SE], 0.637 [0.152];  $P < .001$ ), whereas higher AST levels were associated with higher executive functioning scores ( $\beta$  [SE], 0.607 [0.215];  $P = .01$ ).

**Table 1. Results of Association of Liver Function Biomarkers With Alzheimer Disease Diagnosis<sup>a</sup>**

Liver Function Marker	Odds Ratio (95% CI)	Corrected P Value
Albumin, g/dL	5.789 (0.040-843.993)	.49
Alkaline phosphatase, U/L	3.620 (0.844-15.529)	.12
ALT, U/L	0.133 (0.042-0.422)	.004
AST, U/L	0.229 (0.045-1.175)	.12
AST to ALT ratio	7.932 (1.673-37.617)	.03
Total bilirubin, mg/dL	1.405 (0.585-3.377)	.49

Abbreviations: ALT, alanine aminotransferase; AST, aspartate aminotransferase.

<sup>a</sup> Cognitively normal vs Alzheimer disease. Analyses were adjusted for age, sex, body mass index, and APOE  $\epsilon$ 4 status.

### Biomarkers of Amyloid-β

We used CSF amyloid-β 1-42 levels and a global cortical amyloid deposition measured from amyloid PET scans as biomarkers of amyloid-β. The regression coefficient of the AST to ALT ratio showed a negative association with CSF amyloid-β 1-42 levels ( $\beta$  [SE], -0.170 [0.061];  $P = .04$ ), indicating that higher AST to ALT ratio values were associated with CSF amyloid-β 1-42 positivity (Figure 1). However, there was no significant correlation between liver function markers and global cortical amyloid deposition.

In the whole-brain analysis using multivariable regression models to determine the association of liver function markers with amyloid-β load measured from amyloid PET scans on a voxelwise level, we identified significant associations for 2 liver function markers. Higher ALT levels were significantly associated with reduced amyloid-β deposition in the bilateral parietal lobes (Figure 2A). Increased AST to ALT ratio values were significantly associated with increased amyloid-β deposition in the bilateral parietal lobes and right temporal lobe (Figure 2C).

### Biomarkers of Fibrillary Tau

We used CSF p-tau levels as a biomarker of fibrillary tau. We investigated the association of liver function markers with CSF p-tau, adjusting for APOE ε4 status as a covariate. Higher AST to ALT ratio values were associated with higher CSF p-tau values ( $\beta$  [SE], 0.175 [0.055];  $P = .02$ ) (Figure 1).

### Biomarkers of Neurodegeneration or Neuronal Injury

We used structural atrophy measured from MRI scans, brain glucose metabolism from FDG-PET scans, and CSF t-tau levels as biomarkers of neurodegeneration or neuronal injury.

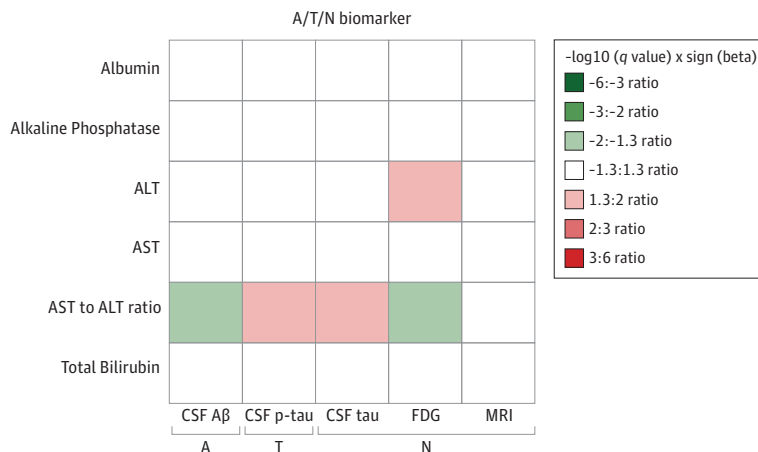
Table 2. Results of Association of Liver Function Biomarkers With Composite Cognitive Performance Measures<sup>a</sup>

Liver Function Marker	Memory Composite Score		Executive Function Composite Score	
	$\beta$ (SE)	Corrected $P$ Value	$\beta$ (SE)	Corrected $P$ Value
Albumin, g/dL	-0.872 (0.576)	.17	-0.203 (0.689)	.77
Alkaline phosphatase, U/L	-0.416 (0.162)	.02	-0.595 (0.193)	.006
ALT, U/L	0.397 (0.128)	.006	0.637 (0.152)	<.001
AST, U/L	0.339 (0.180)	.09	0.607 (0.215)	.01
AST to ALT ratio	-0.465 (0.180)	.02	-0.679 (0.215)	.006
Total bilirubin, mg/dL	-0.068 (0.103)	.61	-0.066 (0.123)	.65

Abbreviations: ALT, alanine aminotransferase; AST, aspartate aminotransferase.

<sup>a</sup> Analyses were adjusted for age, sex, educational level, body mass index, and APOE ε4 status.

Figure 1. Results of Association of Liver Function Biomarkers With Amyloid, Tau, and Neurodegeneration (A/T/N) Biomarkers for Alzheimer Disease



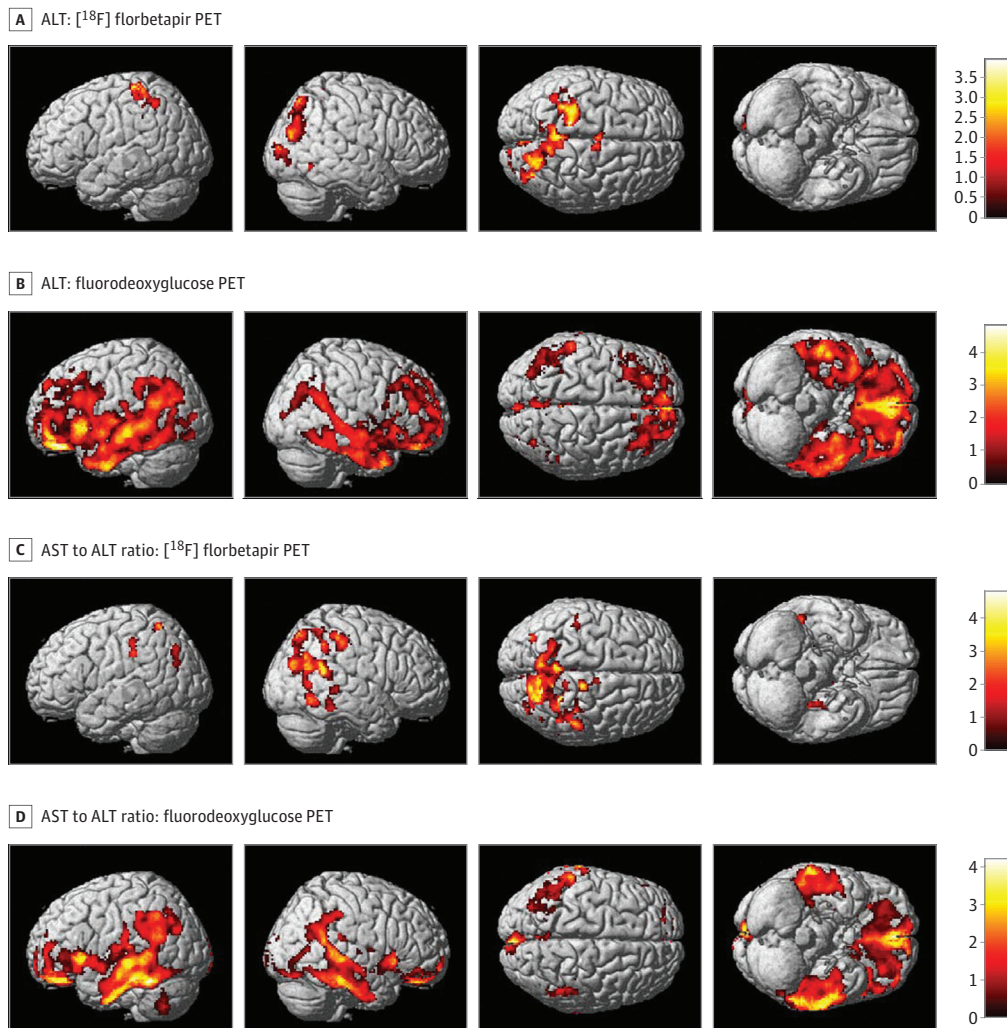
Heat map of q-values of the association between liver function markers and the A/T/N biomarkers for Alzheimer disease.  $P$  values estimated from linear regression analyses were corrected for multiple testing using false discovery rate ( $q$  value). White indicates  $q > 0.05$ , red indicates significant positive association, and green indicates significant negative association. A $\beta$  indicates amyloid- $\beta$ ; ALT, alanine aminotransferase; AST, aspartate aminotransferase; CSF, cerebrospinal fluid; FDG, fludeoxyglucose positron emission tomography; MRI, magnetic resonance imaging; and p-tau, phosphorylated tau.

**Brain Glucose Metabolism**

We performed an ROI-based association analysis of liver function markers with a global cortical glucose metabolism value measured from FDG-PET scans across 1167 ADNI participants with both FDG-PET scans and measurement of liver function markers. The association analysis including *APOE*  $\epsilon 4$  status as a covariate identified 2 markers as significantly associated with brain glucose metabolism after controlling for multiple testing using FDR (Figure 1). For ALT, higher levels were associated with increased glucose metabolism ( $\beta$  [SE], 0.096 [0.030];  $P = .02$ ), while for the AST to ALT ratio, higher ratio values were associated with reduced glucose metabolism ( $\beta$  [SE], -0.123 [0.042];  $P = .03$ ).

In the detailed whole-brain analysis to determine the association of liver function markers with brain glucose metabolism on a voxelwise level, increased ALT levels were associated with increased glucose metabolism in a widespread pattern, especially in the bilateral frontal, parietal, and temporal lobes (Figure 2B). However, higher AST to ALT ratio values were significantly associated with reduced glucose metabolism in the bilateral frontal, parietal, and temporal lobes (Figure 2D).

**Figure 2. Detailed Whole-Brain Voxel-Based Imaging Analysis for Alanine Aminotransferase (ALT) and Aspartate Aminotransferase (AST) to ALT Ratio Levels Using Positron Emission Tomography (PET) Scans**



Whole-brain multivariable analysis was performed to visualize the topography of the association of ALT levels and AST to ALT ratio values with amyloid- $\beta$  load and glucose metabolism on a voxelwise level (false discovery rate-corrected  $P < .05$ ). A, Higher ALT levels were significantly associated with reduced amyloid- $\beta$  deposition in the bilateral parietal lobes. B, Increased ALT levels were significantly associated with increased

glucose metabolism in a widespread manner, especially in the bilateral frontal, parietal, and temporal lobes. C, Increased AST to ALT ratio values were significantly associated with increased amyloid- $\beta$  deposition in the bilateral parietal lobes and the right temporal lobe. D, Increased AST to ALT ratio values were significantly associated with reduced brain glucose metabolism in the bilateral frontal, parietal, and temporal lobes.

### Structural MRI (Atrophy)

In the investigation of the association of liver function markers with mean hippocampal volume with *APOE*  $\epsilon$ 4 status as a covariate, we did not identify any significant association with hippocampal volume after controlling for multiple testing using FDR (Figure 1). Following the detailed whole-brain surface-based analysis of liver function markers using multivariable regression models to assess associations with cortical thickness, higher ALT levels were significantly associated with larger cortical thickness in the bilateral temporal lobes (Figure 3), which showed consistent patterns in the associations of brain glucose metabolism.

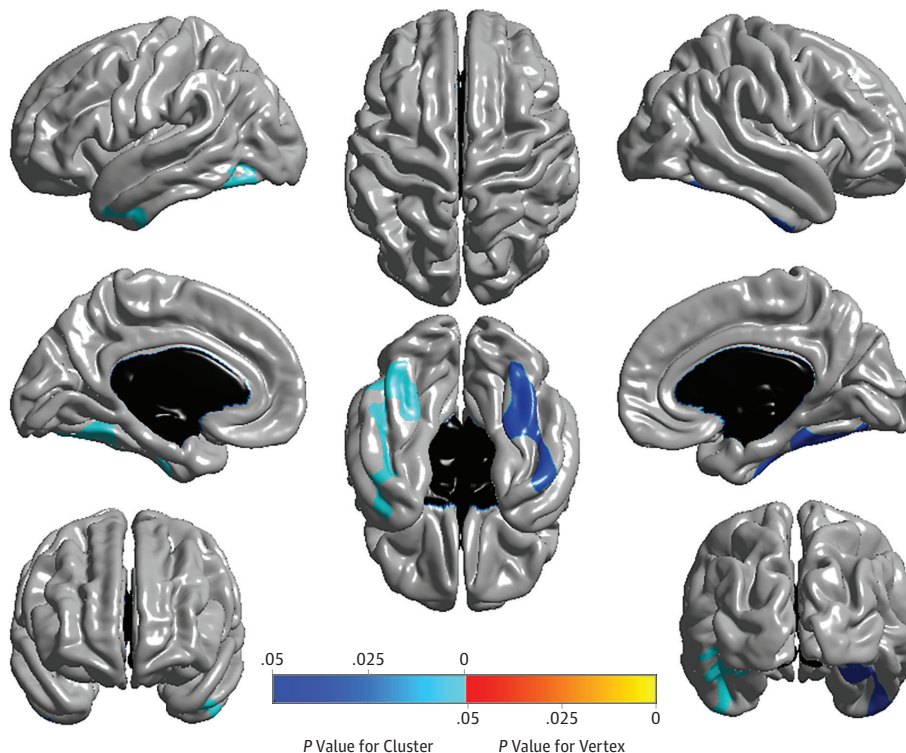
### CSF t-Tau

Higher AST to ALT ratio values were associated with higher CSF t-tau levels ( $\beta$  [SE], 0.160 [0.049];  $P = .02$ ) (Figure 1), which showed consistent patterns in the associations of CSF amyloid- $\beta$  1-42 or p-tau levels and brain glucose metabolism.

## Discussion

We investigated the association between serum-based liver function markers and AD diagnosis, cognition, and AD pathophysiological characteristics based on the A/T/N framework for AD biomarkers in the ADNI cohort.<sup>39</sup> Our findings suggest that the decreased levels of ALT and elevated AST to ALT ratio that were observed in patients with AD were associated with poor cognition and reduced brain glucose metabolism. We also found that an increased AST to ALT ratio was associated with lower CSF amyloid- $\beta$  1-42 levels, greater amyloid- $\beta$  deposition, and higher CSF p-tau and t-tau levels. Furthermore, we observed that decreased levels of ALT were associated with greater amyloid- $\beta$  deposition and structural atrophy.

Figure 3. Detailed Whole-Brain Surface-Based Imaging Analysis for Alanine Aminotransferase (ALT) Levels Using Magnetic Resonance Imaging (MRI) Scans



A whole-brain multivariable analysis of cortical thickness across the brain surface was performed to visualize the topography of the association of ALT levels with brain structure. Statistical maps were thresholded using a random field theory for a multiple testing adjustment to a corrected significance level of  $P < .05$ . The  $P$  value for clusters indicates significant corrected  $P$  values with the lightest blue color. Higher ALT levels were significantly associated with greater cortical thickness, especially in bilateral temporal lobes.



Decreased levels of ALT and increased AST to ALT ratio values were observed in patients with AD and were associated with lower scores on measures of memory and executive function. Our findings are comparable with those of an earlier study that reported increased AST to ALT ratio values and lower levels of ALT in patients with AD compared with controls, although in that study, the association between AD and ALT levels did not reach statistical significance.<sup>40</sup> Altered liver enzymes lead to disturbances in liver-associated metabolites including branched-chain amino acids, ether-phosphatidylcholines, and lipids,<sup>41</sup> which we and others show are altered in AD<sup>1,42-44</sup> and may play a role in disease pathophysiologic characteristics.<sup>45</sup> Disturbed energy metabolism is one of the processes that may explain the observed lower levels of ALT and increased enzyme ratio in individuals with AD and impaired cognition.<sup>3,5</sup> This finding is concordant with our observation that increased AST to ALT ratio values and lower levels of ALT showed a consistent significant association with reduced brain glucose metabolism, particularly in the orbitofrontal cortex and temporal lobes, areas of the brain implicated in memory and executive function. Brain glucose hypometabolism is an early feature of AD and cognitive impairment during the prodromal stage.<sup>46,47</sup> Moreover, ALT and AST are key enzymes in gluconeogenesis in the liver and production of neurotransmitters required in maintaining synapses.<sup>48</sup> Alanine aminotransferase catalyzes a reversible transamination reaction between alanine and  $\alpha$ -ketoglutarate to form pyruvate and glutamate, while AST catalyzes a reversible reaction between aspartate and  $\alpha$ -ketoglutarate to form oxaloacetate and glutamate.<sup>49</sup> Although exact mechanisms remain unclear, 2 possible mechanisms may explain altered levels of enzymes in AD. First, reduced ALT levels lead to reduced pyruvate, which is required for glucose production via gluconeogenesis in the liver and glucose is distributed in various body tissues as an energy source,<sup>50</sup> thus disturbing energy homeostasis. Second, altered levels of ALT and AST may affect levels of glutamate, an excitatory neurotransmitter of the central nervous system involved in synaptic transmission, which also plays an important role in memory.<sup>51</sup>

In the case of low glucose metabolism in the brain, as observed in our current study, less  $\alpha$ -ketoglutarate is available via the tricarboxylic acid cycle that favors glutamate catabolism vs glutamate synthesis in reversible reaction (catalyzed by AST and ALT).<sup>52</sup> Glutamate acts as a neurotransmitter in approximately two-thirds of the synapses in neocortical and hippocampal pyramidal neurons and thus is involved in memory and cognition via long-term potentiation.<sup>53</sup> In a sample of healthy adults, plasma ALT and AST levels were significantly positively correlated with plasma glutamate levels,<sup>5,54</sup> which indicates that lower levels of ALT will decrease glutamate levels in plasma. Based on evidence from earlier studies that peripheral blood levels of glutamate are positively correlated with levels of glutamate in the CSF<sup>55</sup> and studies that reported lower levels of glutamate in patients with AD compared with controls in both blood<sup>56</sup> and brain tissues,<sup>36,57-59</sup> we can infer that lower levels of ALT or AST may affect glutamate levels in AD. In older adults, lower serum ALT levels are associated with mortality<sup>60,61</sup> and are thought to be a biomarker for increased frailty, sarcopenia, and/or reduced levels of pyridoxine (vitamin B<sub>6</sub>).<sup>62</sup> Pyridoxine phosphate is a coenzyme for the synthesis of amino acids, neurotransmitters (eg, serotonin and norepinephrine), and sphingolipids. Alanine aminotransferase decreases with age<sup>63</sup> and may be a sign of hepatic aging. Glutamate levels also decrease with increasing age.<sup>64</sup> Together with the fact that age is the strongest risk factor for AD,<sup>65</sup> decreasing levels of ALT with age may also indicate a possible biological link between aging and AD. Nevertheless, further research is needed to determine the exact cause of reducing ALT levels with age and the pathway through which it can influence neurologic disorders, including AD.

Increased AST to ALT ratios are observed in individuals with nonalcoholic fatty liver disease, which is the hepatic manifestation of metabolic syndrome.<sup>66</sup> In the Framingham Heart Study, nonalcoholic fatty liver disease was associated with smaller total cerebral brain volume even after adjustment for multiple cardiovascular risk factors.<sup>67</sup> Liver dysfunction is also associated with the development of disease including cardiovascular disease and insulin resistance through disruptions in glucose and lipid metabolism, key physiological functions of the liver.<sup>68,69</sup> Thus, using the AST to

ALT ratio as a marker for overall metabolic disturbance,<sup>5</sup> our study provides evidence of an association between altered metabolic status and AD, cognition, and AD endophenotypes.

In addition to ALT levels and the AST to ALT ratio, elevated levels of alkaline phosphatase were significantly associated with poor cognition. This is in line with results from the Oxford Project to Investigate Memory and Aging, which reported increased alkaline phosphatase levels in individuals with AD and an inverse association with cognition.<sup>70</sup> Alkaline phosphatase is an enzyme primarily expressed in the liver and kidneys as well as in endothelial cells in the brain.<sup>71,72</sup> The neuronal form of alkaline phosphatase plays a role in developmental plasticity and activity-dependent cortical functions via contributing in  $\gamma$ -aminobutyric acid metabolism.<sup>73-76</sup> Changes in plasma levels of alkaline phosphatase may occur as a result of central nervous system injury.<sup>77</sup>

### Limitations

This study has several limitations. The observational design of this ADNI cohort study limits our ability to make assumptions about causality. There is need to evaluate the association of liver enzymes with AD in prospective manner. Another limitation of our study is that we did not adjust for alcohol consumption, which was not available in ADNI. Alcohol consumption is associated with altered liver enzymes. Instead, we used a well-established surrogate marker of alcohol consumption,  $\gamma$ -glutamyltransferase. Elevations in  $\gamma$ -glutamyltransferase generally indicate long-term heavy drinking rather than episodic heavy drinking.<sup>78</sup> Our key findings remained significant after adjustment for  $\gamma$ -glutamyltransferase and statin use (eTable 4, eTable 5, and eFigure in the Supplement). However, given the associations with liver function measures and A/T/N biomarkers for AD, it appears that liver function may play a role in the pathogenesis of AD, but limitations should be taken into account before further extrapolating our findings.

### Conclusions

This study's results suggest that altered liver function markers are associated with AD diagnosis and impaired memory and executive function as well as amyloid- $\beta$ , tau, and neurodegenerative biomarkers of AD pathophysiological characteristics. These results are, to our knowledge, the first to show an association of peripheral markers of liver functioning with central biomarkers associated with AD. Although our results suggest an important role of liver functioning in AD pathophysiological characteristics, the causal pathways remain unknown. The liver-brain biochemical axis of communication should be further evaluated in model systems and longitudinal studies to gain deeper knowledge of causal pathways.

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**Corresponding Authors:** Rima Kaddurah-Daouk, PhD, Duke University Medical Center, Room 3552, Duke Blue South, Durham, NC 27710 ([rima.kaddurahdaouk@duke.edu](mailto:rima.kaddurahdaouk@duke.edu)); Andrew J. Saykin, PsyD, Indiana University Neuroscience Center, Department of Radiology and Imaging Sciences, Indiana University School of Medicine, 355 W 16th St, Ste 4100, Indianapolis, IN 46202 ([asaykin@iupui.edu](mailto:asaykin@iupui.edu)).

**Author Affiliations:** Center for Computational Biology and Bioinformatics, Indiana Alzheimer Disease Center, Department of Radiology and Imaging Sciences, Indiana University School of Medicine, Indianapolis (Nho, Risacher, Saykin); Department of Psychiatry and Behavioral Sciences, Duke University, Durham, North Carolina (Kueider-Paisley, MahmoudianDehkordi, Arnold, Louie, Doraiswamy, Kaddurah-Daouk); Department of Epidemiology, Erasmus Medical Centre, Rotterdam, the Netherlands (Ahmad, van Duijn); Institute of Bioinformatics and Systems Biology, Helmholtz Zentrum München, German Research Center for Environmental

Health, Neuherberg, Germany (Arnold, Kastenmüller); Duke Molecular Physiology Institute, Duke University, Durham, North Carolina (Blach); Rosa & Co LLC, San Carlos, California (Baillie); University of Texas Health Science Center at San Antonio, San Antonio (Han); German Center for Diabetes Research, Neuherberg, Germany (Kastenmüller); Department of Pathology and Laboratory Medicine, University of Pennsylvania, Philadelphia (Trojanowski, Shaw); Center for Imaging of Neurodegenerative Diseases, Department of Radiology, San Francisco Veterans Affairs Medical Center and University of California, San Francisco (Weiner); Duke Institute of Brain Sciences, Duke University, Durham, North Carolina (Doraiswamy, Kaddurah-Daouk); Department of Medicine, Duke University, Durham, North Carolina (Doraiswamy, Kaddurah-Daouk); Nuffield Department of Population Health, Oxford University, Oxford, United Kingdom (van Duijn).

**Author Contributions:** Drs Nho and Arnold had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. Drs Nho and Kueider-Paisley and Mr Ahmad contributed equally.

*Concept and design:* Nho, Kueider-Paisley, Ahmad, Trojanowski, Doraiswamy, Kaddurah-Daouk.

*Acquisition, analysis, or interpretation of data:* All authors.

*Drafting of the manuscript:* Nho, Kueider-Paisley, Ahmad, MahmoudianDehkordi, Louie, Trojanowski, Kaddurah-Daouk.

*Critical revision of the manuscript for important intellectual content:* Nho, Kueider-Paisley, Ahmad, Arnold, Risacher, Blach, Baillie, Han, Kastenmüller, Trojanowski, Shaw, Weiner, Doraiswamy, van Duijn, Saykin, Kaddurah-Daouk.

*Statistical analysis:* Nho, MahmoudianDehkordi, Trojanowski, van Duijn.

*Obtained funding:* Nho, Arnold, Weiner, van Duijn, Saykin, Kaddurah-Daouk.

*Administrative, technical, or material support:* Arnold, Louie, Blach, Han, Doraiswamy, Saykin.

*Supervision:* Nho, Arnold, Kastenmüller, Shaw, Kaddurah-Daouk.

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**Group Information:** Alzheimer's Disease Neuroimaging Initiative-I, ADNI-GO, ADNI-II, and ADNI-III investigators include Part A: Leadership and Infrastructure Principal Investigator (PI): Michael W. Weiner, MD, University of California San Francisco; ATRI PI and Director of Coordinating Center Clinical Core: Paul Aisen, MD, University of Southern California; Executive Committee: Michael Weiner, MD, University of California San Francisco; Paul Aisen, MD, University of Southern California; Ronald Petersen, MD, PhD, Mayo Clinic, Rochester, NY; Clifford R. Jack Jr, MD, Mayo Clinic, Rochester, NY; William Jagust, MD, University of California Berkeley; John Q. Trojanowki, MD, PhD, University of Pennsylvania; Arthur W. Toga, PhD, University of Southern California; Laurel Beckett, PhD, University of California Davis; Robert C. Green, MD, MPH, Brigham and Women's Hospital/Harvard Medical School; Andrew J. Saykin, PsyD, Indiana University; John Morris, MD, Washington University St Louis; and Leslie M. Shaw, University of Pennsylvania.

ADNI External Advisory Board members include Zaven Khachaturian, PhD, Prevent Alzheimer's Disease 2020 (Chair); Greg Sorensen, MD, Siemens; Maria Carrillo, PhD, Alzheimer's Association; Lew Kuller, MD, University of Pittsburgh; Marc Raichle, MD, Washington University, St Louis; Steven Paul, MD, Cornell University; Peter Davies, MD, Albert Einstein College of Medicine of Yeshiva University; Howard Fillit, MD, AD Drug Discovery Foundation; Franz Hefti, PhD, Acumen Pharmaceuticals; David Holtzman, MD, Washington University, St Louis; M. Marcel Mesulam, MD, Northwestern University; William Potter, MD, National Institute of Mental Health; and Peter Snyder, PhD, Brown University.

ADNI-3 Private Partner Scientific Board: Veronika Logovinsky, MD, PhD, Eli Lilly (Chair).

Data and Publications Committee members include Robert C. Green, MD, MPH, BWH/HMS (Chair) Resource Allocation Review Committee; Tom Montine, MD, PhD, University of Washington (Chair); Clinical Core Leaders: Ronald Petersen, MD, PhD, Mayo Clinic, Rochester (Core PI); Paul Aisen, MD, University of Southern California Clinical Informatics and Operations; Gustavo Jimenez, MBS, University of Southern California; Michael Donohue, PhD, University of Southern California; Devon Gessert, BS, University of Southern California; Kelly Harless, BA, University of Southern California; Jennifer Salazar, MBS, University of Southern California; Yuliana Cabrera, BS, University of Southern California; Sarah Walter, MSc, University of Southern California; and Lindsey Hergesheimer, BS, University of Southern California. Biostatistics Core Leaders and Key Personnel: Laurel Beckett, PhD, University of California Davis (Core PI); Danielle Harvey, PhD, University of California Davis; and Michael Donohue, PhD, University of California San Diego. MRI Core Leaders and Key Personnel: Clifford R. Jack Jr, MD, Mayo Clinic, Rochester (Core PI); Matthew Bernstein, PhD, Mayo Clinic, Rochester; Nick Fox, MD, University of London; Paul

Thompson, PhD, University of California Los Angeles School of Medicine; Norbert Schuff, PhD, University of California San Francisco MRI; Charles DeCarli, MD, University of California Davis; Bret Borowski, RT, Mayo Clinic; Jeff Gunter, PhD, Mayo Clinic; Matt Senjem, MS, Mayo Clinic; Prashanthi Vemuri, PhD, Mayo Clinic; David Jones, MD, Mayo Clinic; Kejal Kantarci, Mayo Clinic; and Chad Ward, Mayo Clinic. PET Core Leaders and Key Personnel: William Jagust, MD, University of California Berkeley (Core PI); Robert A. Koeppe, PhD, University of Michigan; Norm Foster, MD, University of Utah; Eric M. Reiman, MD, Banner Alzheimer's Institute; Kewei Chen, PhD, Banner Alzheimer's Institute; Chet Mathis, MD, University of Pittsburgh; and Susan Landau, PhD, University of California Berkeley.

Neuropathology Core Leaders include John C. Morris, MD, Washington University, St Louis; Nigel J. Cairns, PhD, FRCPATH, Washington University, St Louis; Erin Franklin, MS, CCRP, Washington University, St Louis; and Lisa Taylor-Reinwald, BA, HTL, Washington University, St Louis.

American Society for Clinical Pathology (ASCP)—Past Investigator: Biomarkers Core Leaders and Key Personnel: Leslie M. Shaw, PhD, University of Pennsylvania School of Medicine; John Q. Trojanowki, MD, PhD, University of Pennsylvania School of Medicine; Virginia Lee, PhD, MBA, University of Pennsylvania School of Medicine; Magdalena Korecka, PhD, University of Pennsylvania School of Medicine; and Michal Figurski, PhD, University of Pennsylvania School of Medicine. Informatics Core Leaders and Key Personnel: Arthur W. Toga, PhD, University of Southern California (Core PI); Karen Crawford, University of Southern California; and Scott Neu, PhD, University of Southern California. Genetics Core Leaders and Key Personnel: Andrew J. Saykin, PsyD, Indiana University; Tatiana M. Foroud, PhD, Indiana University; Steven Potkin, MD, University of California Irvine; Li Shen, PhD, Indiana University; Kelley Faber, MS, CCRC, Indiana University; Sungeun Kim, PhD, Indiana University; and Kwangsik Nho, PhD, Indiana University. Initial Concept Planning & Development: Michael W. Weiner, MD, University of California San Francisco; Lean Thal, MD, University of California San Diego; and Zaven Khachaturian, PhD, Prevent Alzheimer's Disease 2020. Early Project Proposal Development: Leon Thal, MD, University of California San Diego; Neil Buckholtz, National Institute on Aging; Michael W. Weiner, MD, University of California San Francisco; Peter J. Snyder, PhD, Brown University; William Potter, MD, National Institute of Mental Health; Steven Paul, MD, Cornell University; Marilyn Albert, PhD, Johns Hopkins University; Richard Frank, MD, PhD, Richard Frank Consulting; Zaven Khachaturian, PhD, Prevent Alzheimer's Disease 2020; and John Hsiao, MD, National Institute on Aging.

Part B: Investigators by Site: Oregon Health & Science University: Joseph Quinn, MD; Lisa C. Silbert, MD; Betty Lind, BS; Jeffrey A. Kaye, MD, (Past Investigator); Raina Carter, BA (Past Investigator); and Sara Dolen, BS (Past Investigator). University of Southern California: Lon S. Schneider, MD; Sonia Pawluczyk, MD; Mauricio Becerra, BS; Liberty Teodoro, RN; and Bryan M. Spann, DO, PhD (Past Investigator). University of California—San Diego: James Brewer, MD, PhD; Helen Vanderswag, RN; and Adam Fleisher, MD (Past Investigator). University of Michigan: Jaimie Ziolkowski, MA, BS, TLLP; Judith L. Heidebrink, MD, MS; and Joanne L. Lord, LPN, BA, CCRC (Past Investigator). Mayo Clinic, Rochester: Ronald Petersen, MD, PhD; Sara S. Mason, RN; Colleen S. Albers, RN; David Knopman, MD; and Kris Johnson, RN (Past Investigator). Baylor College of Medicine: Javier Villanueva-Meyer, MD; Valory Pavlik, PhD; Nathaniel Pacini, MA; Ashley Lamb, MA; Joseph S. Kass, MD, LD, FAAN; Rachelle S. Doody, MD, PhD (Past Investigator); Victoria Shibley, MS (Past Investigator); Munir Chowdhury, MBBS, MS (Past Investigator); Susan Rountree, MD (Past Investigator); and Mimi Dang, MD (Past Investigator). Columbia University Medical Center: Yaakov Stern, PhD; Lawrence S. Honig, MD, PhD; Karen L. Bell, MD; and Randy Yeh, MD. Washington University in St Louis: Beau Ances, MD, PhD, MSc; John C. Morris, MD; David Winkfield, BS; Maria Carroll, RN, MSN, GCNS-BC; Angela Oliver, RN, BSN, MSG; Mary L. Creech, RN, MSW (Past Investigator); Mark A. Mintun, MD (Past Investigator); and Stacy Schneider, APRN, BC, GNP (Past Investigator). University of Alabama—Birmingham: Daniel Marson, JD, PhD; David Geldmacher, MD; Marissa Natelson Love, MD; Randall Griffith, PhD, ABPP (Past Investigator); David Clark, MD (Past Investigator); and John Brockington, MD (Past Investigator). Mount Sinai School of Medicine: Hillel Grossman, MD; and Effie Mitsis, PhD (Past Investigator). Rush University Medical Center: Raj C. Shah, MD; Melissa Lamar, PhD; and Patricia Samuels. Wien Center: Ranjan Duara, MD; Maria T. Greig-Custo, MD; and Rosemarie Rodriguez, PhD. Johns Hopkins University: Marilyn Albert, PhD; Chiadi Onyike, MD; Daniel D'Agostino II, BS; and Stephanie Kielbaso, BS (Past Investigator). New York University: Martin Sadowski, MD, PhD; Mohammed O. Sheikh, MD; Jamika Singleton-Garvin, CCRP; Anasztasia Ulysse; and Mrunalini Gaikwad. Duke University Medical Center: P. Murali Doraiswamy, MBBS, FRCP; Jeffrey R. Petrella, MD; Olga James, MD; Salvador Borges-Neto, MD; Terence Z. Wong, MD (Past Investigator); and Edward Coleman (Past Investigator). University of Pennsylvania: Jason H. Karlawish, MD; David A. Wolk, MD; Sanjeev Vaishnavi, MD; Christopher M. Clark, MD (Past Investigator); and Steven E. Arnold, MD (Past Investigator). University of Kentucky: Charles D. Smith, MD; Greg Jicha, MD; Peter Hardy, PhD; Riham El Khouli, MD; Elizabeth Oates, MD; and Gary Conrad, MD. University of Pittsburgh: Oscar L. Lopez, MD; MaryAnn Oakley, MA; and Donna M. Simpson, CRNP, MPH. University of Rochester Medical Center: Anton P. Porsteinsson, MD; Kim Martin, RN; Nancy Kowalksi, MS, RNC; Melanie Keltz, RN; Bonnie S. Goldstein, MS, NP (Past Investigator); Kelly M. Makino, BS (Past Investigator); M. Saleem Ismail, MD (Past Investigator); and Connie Brand, RN (Past Investigator). University of California Irvine IMIND: Gaby Thai, MD; Aimee Pierce, MD; Beatriz Yanez, RN; Elizabeth Sosa, PhD; and Megan Witbracht, PhD. University of Texas

Southwestern Medical School: Kyle Womack, MD; Dana Mathews, MD, PhD; and Mary Quiceno, MD. Emory University: Allan I. Levey, MD, PhD; James J. Lah, MD, PhD; and Janet S. Cellar, DNP, PMHCNS-BC. University of Kansas Medical Center: Jeffrey M. Burns, MD; Russell H. Swerdlow, MD; and William M. Brooks, PhD. University of California, Los Angeles: Ellen Woo, PhD; Daniel H.S. Silverman, MD, PhD; Edmond Teng, MD, PhD; Sarah Kremen, MD; Liana Apostolova, MD (Past Investigator); Kathleen Tingus, PhD (Past Investigator); Po H. Lu, PsyD (Past Investigator); and George Bartzokis, MD (Past Investigator). Mayo Clinic, Jacksonville: Neill R Graff-Radford, MBBCH, FRCP (London); Francine Parfitt, MSH, CCRC; and Kim Poki-Walker, BA. Indiana University: Martin R. Farlow, MD; Ann Marie Hake, MD; Brandy R. Matthews, MD (Past Investigator); Jared R. Brosch, MD; and Scott Herring, RN, CCRC. Yale University School of Medicine: Christopher H. van Dyck, MD; Richard E. Carson, PhD; and Pradeep Varma, MD. McGill University, Montreal-Jewish General Hospital: Howard Chertkow, MD; Howard Bergman, MD; and Chris Hosein, Med. Sunnybrook Health Sciences, Ontario: Sandra Black, MD, FRCPC; Bojana Stefanovic, PhD; and Chris (Chinthaka) Heyn, BSc, PhD, MD, FRCPC. U.B.C. Clinic for AD & Related Disorders: Ging-Yuek Robin Hsiung, MD, MHSc, FRCPC; Benita Mudge, BS; Vesna Sossi, PhD; Howard Feldman, MD, FRCPC (Past Investigator); and Michele Assaly, MA (Past Investigator). Cognitive Neurology - St Joseph's, Ontario: Elizabeth Finger, MD; Stephen Pasternack, MD, PhD; William Pavlosky, MD; Irina Rachinsky, MD (Past Investigator); Dick Drost, PhD (Past Investigator); and Andrew Kertesz, MD (Past Investigator). Cleveland Clinic Lou Ruvo Center for Brain Health: Charles Bernick, MD, MPH; and Donna Munic, PhD. Northwestern University: Marek-Marsel Mesulam, MD; Emily Rogalski, PhD; Kristine Lipowski, MA; Sandra Weintraub, PhD; Borna Bonakdarpour, MD; Diana Kerwin, MD (Past Investigator); Chuang-Kuo Wu, MD, PhD (Past Investigator); and Nancy Johnson, PhD (Past Investigator). Premiere Research Inst (Palm Beach Neurology): Carl Sadowsky, MD; and Teresa Villena, MD. Georgetown University Medical Center: Raymond Scott Turner, MD, PhD; Kathleen Johnson, NP; and Brigid Reynolds, NP. Brigham and Women's Hospital: Reisa A. Sperling, MD; Keith A. Johnson, MD; and Gad A. Marshall, MD. Stanford University: Jerome Yesavage, MD; Joy L. Taylor, PhD; Steven Chao, MD, PhD; Barton Lane, MD (Past Investigator); Allyson Rosen, PhD (Past Investigator); and Jared Tinklenberg, MD (Past Investigator). Banner Sun Health Research Institute: Edward Zamrini, MD; Christine M. Belden, PsyD; and Sherye A. Sirrel, CCRC. Boston University: Neil Kowall, MD; Ronald Killiany, PhD; Andrew E. Budson, MD; Alexander Norbash, MD (Past Investigator); and Patricia Lynn Johnson, BA (Past Investigator). Howard University: Thomas O. Obisesan, MD, MPH; Ntekim E. Oyonomo, MD, PhD; Joanne Allard, PhD; and Olu Ogunlana, BPharm. Case Western Reserve University: Alan Lerner, MD; Paula Ogrocki, PhD; Curtis Tatsuoka, PhD; and Parianne Fatica, BA, CCRC. University of California, Davis-Sacramento: Evan Fletcher, PhD; Pauline Maillard, PhD; John Olichney, MD; Charles DeCarli, MD; and Owen Carmichael, PhD (Past Investigator). Neurological Care of CNY: Smita Kittur, MD (Past Investigator). Parkwood Institute: Michael Borrie, MB ChB; T-Y Lee, PhD; and Rob Bartha, PhD. University of Wisconsin: Sterling Johnson, PhD; Sanjay Asthana, MD; and Cynthia M. Carlsson, MD, MS. Banner Alzheimer's Institute: Pierre Tariot, MD; Anna Burke, MD; Joel Hetelle, BS; Kathryn DeMarco, BS; Nadira Trncic, MD, PhD, CCRC (Past Investigator); Adam Fleisher, MD (Past Investigator); and Stephanie Reeder, BA (Past Investigator). Dent Neurologic Institute: Vernice Bates, MD; Horacio Capote, MD; and Michelle Rainka, PharmD, CCRP. The Ohio State University: Douglas W. Scharre, MD; Maria Katakis, MD, PhD; and Rawan Tarawneh, MD. Albany Medical College: Earl A. Zimmerman, MD; Dzintra Celmins, MD; and David Hart, MD. Hartford Hospital, Olin Neuropsychiatry Research Center: Godfrey D. Pearlson, MD; Karen Blank, MD; and Karen Anderson, RN. Dartmouth-Hitchcock Medical Center: Laura A. Flashman, PhD; Marc Seltzer, MD; Mary L. Hynes, RN, MPH; and Robert B. Santulli, MD (Past Investigator). Wake Forest University Health Sciences: Kaycee M. Sink, MD, MAS; Mia Yang, MD; and Akiva Mintz, MD, PhD. Rhode Island Hospital: Brian R. Ott, MD; Geoffrey Tremont, PhD; and Lori A. Daiello, PharmD, ScM. Butler Hospital: Courtney Bodge, PhD; Stephen Salloway, MD, MS; Paul Malloy, PhD; Stephen Correia, PhD; and Athena Lee, PhD. University of California San Francisco: Howard J. Rosen, MD; Bruce L. Miller, MD; and David Perry, MD. Medical University of South Carolina: Jacobo Mintzer, MD, MBA; Kenneth Spicer, MD, PhD; and David Bachman, MD. St Joseph's Health Care: Elizabeth Finger, MD; Stephen Pasternak, MD; Irina Rachinsky, MD; John Rogers, MD; Andrew Kertesz, MD (Past Investigator); and Dick Drost, MD (Past Investigator). Nathan Kline Institute: Nunzio Pomara, MD; Raymundo Hernando, MD; and Antero Sarrael, MD. University of Iowa College of Medicine: Delwyn D. Miller, PharmD, MD; Karen Ekstam Smith, RN; Hristina Koleva, MD; Ki Won Nam, MD; Hyungsub Shim, MD; and Susan K. Schultz, MD (Past Investigator). Cornell University: Norman Relkin, MD, PhD; Gloria Chiang, MD; Michael Lin, MD; and Lisa Ravdin, PhD. University of South Florida Health Byrd Alzheimer's Institute: Amanda Smith, MD; Christi Leach, MD; Balebail Ashok Raj, MD (Past Investigator); and Kristin Fargher, MD (Past Investigator).

DOD ADNI investigators include Part A: Leadership and Infrastructure Principal Investigator: Michael W. Weiner, MD, University of California, San Francisco; ATRI PI and Director of Coordinating Center Clinical Core: Paul Aisen, MD, University of Southern California; Executive Committee: Michael Weiner, MD, University of California San Francisco; Paul Aisen, MD, University of Southern California; Ronald Petersen, MD, PhD, Mayo Clinic, Rochester; Robert C. Green, MD, MPH, Brigham and Women's Hospital/Harvard Medical School; Danielle Harvey, PhD, University of California Davis; Clifford R. Jack Jr, MD, Mayo Clinic, Rochester; William Jagust, MD, University of California Berkeley; John C. Morris, MD, Washington University, St Louis; Andrew J. Saykin, PsyD, Indiana University; Leslie M. Shaw, PhD, Perelman School of Medicine, University of Pennsylvania; Arthur W. Toga, PhD,

University of Southern California; John Q. Trojanowki, MD, PhD, Perelman School of Medicine, University of Pennsylvania; Psychological Evaluation/PTSD Core: Thomas Neylan, MD, University of California San Francisco; Traumatic Brain Injury/TBI Core: Jordan Grafman, PhD, Rehabilitation Institute of Chicago, Feinberg School of Medicine, Northwestern University; Data and Publication Committee: Robert C. Green, MD, MPH BWH/HMS (Chair); Resource Allocation Review Committee: Tom Montine, MD, PhD, University of Washington (Chair); Clinical Core Leaders: Michael Weiner MD, Core PI; Ronald Petersen, MD, PhD, Mayo Clinic, Rochester (Core PI); Paul Aisen, MD, University of Southern California; Clinical Informatics and Operations: Gustavo Jimenez, MBS, University of Southern California; Michael Donohue, PhD, University of Southern California; Devon Gessert, BS, University of Southern California; Kelly Harless, BA, University of Southern California; Jennifer Salazar, MBS, University of Southern California; Yuliana Cabrera, BS, University of Southern California; Sarah Walter, MSc, University of Southern California; Lindsey Hergesheim, BS, University of Southern California; San Francisco Veterans Affairs Medical Center: Thomas Neylan, MD, University of California San Francisco; Jacqueline Hayes, University of California San Francisco; Shannon Finley, University of California San Francisco; Biostatistics Core Leaders and Key Personnel: Danielle Harvey, PhD, University of California Davis (Core PI); Michael Donohue, PhD, University of California San Diego; MRI Core Leaders and Key Personnel: Clifford R. Jack Jr, MD, Mayo Clinic, Rochester (Core PI); Matthew Bernstein, PhD, Mayo Clinic, Rochester; Bret Borowski, RT, Mayo Clinic; Jeff Gunter, PhD, Mayo Clinic; Matt Senjem, MS, Mayo Clinic; Kejal Kantarci, Mayo Clinic; Chad Ward, Mayo Clinic; PET Core Leaders and Key Personnel: William Jagust, MD, University of California Berkeley (Core PI); Robert A. Koeppel, PhD, University of Michigan; Norm Foster, MD, University of Utah; Eric M. Reiman, MD, Banner Alzheimer's Institute; Kewei Chen, PhD, Banner Alzheimer's Institute; Susan Landau, PhD, University of California Berkeley; Neuropathology Core Leaders: John C. Morris, MD, Washington University, St Louis; Nigel J. Cairns, PhD, FRCPATH, Washington University, St Louis; Erin Householder, MS, Washington University, St Louis; Biomarkers Core Leaders and Key Personnel: Leslie M. Shaw, PhD, Perelman School of Medicine, University of Pennsylvania; John Q. Trojanowki, MD, PhD, Perelman School of Medicine, University of Pennsylvania; Virginia Lee, PhD, MBA, Perelman School of Medicine, University of Pennsylvania; Magdalena Korecka, PhD, Perelman School of Medicine, University of Pennsylvania; Michal Figurski, PhD, Perelman School of Medicine, University of Pennsylvania; Informatics Core Leaders and Key Personnel: Arthur W. Toga, PhD, University of Southern California (Core PI); Karen Crawford, University of Southern California; Scott Neu, PhD, University of Southern California; Genetics Core Leaders and Key Personnel: Andrew J. Saykin, PsyD, Indiana University; Tatiana M. Foroud, PhD, Indiana University; Steven Potkin, MD, University of California Irvine; Li Shen, PhD, Indiana University; Kelley Faber, MS, CCRC, Indiana University; Sungeun Kim, PhD, Indiana University; Kwangsik Nho, PhD, Indiana University; Initial Concept Planning & Development: Michael W. Weiner, MD, University of California San Francisco; Karl Friedl, Department of Defense (retired).

Part B: Investigators by Site: University of Southern California: Lon S. Schneider, MD, MS; Sonia Pawluczyk, MD; and Mauricio Becerra. University of California, San Diego: James Brewer, MD, PhD; and Helen Vanderswag, RN. Columbia University Medical Center: Yaakov Stern, PhD; Lawrence S. Honig, MD, PhD; and Karen L. Bell, MD. Rush University Medical Center: Debra Fleischman, PhD; Konstantinos Arfanakis, PhD; and Raj C. Shah, MD. Wien Center: Ranjan Duara, MD (PI); Daniel Varon, MD (Co-PI); and Maria T Greig, HP (Coordinator). Duke University Medical Center: P. Murali Doraiswamy, MBBS; Jeffrey R. Petrella, MD; and Olga James, MD. University of Rochester Medical Center: Anton P. Porsteinsson, MD (director); Bonnie Goldstein, MS, NP (coordinator); and Kimberly S. Martin, RN. University of California, Irvine: Steven G. Potkin, MD; Adrian Preda, MD; and Dana Nguyen, PhD. Medical University of South Carolina: Jacobo Mintzer, MD, MBA; Dino Massoglia, MD, PhD; and Olga Brawman-Mintzer, MD. Premiere Research Inst (Palm Beach Neurology): Carl Sadowsky, MD; Walter Martinez, MD; and Teresa Villena, MD. University of California, San Francisco: William Jagust MD; Susan Landau PhD; Howard Rosen, MD; and David Perry. Georgetown University Medical Center: Raymond Scott Turner, MD, PhD; Kelly Behan; and Brigid Reynolds, NP. Brigham and Women's Hospital: Reisa A. Sperling, MD; Keith A. Johnson, MD; and Gad Marshall, MD. Banner Sun Health Research Institute: Marwan N. Sabbagh, MD; Sandra A. Jacobson, MD; and Sherye A. Sirrel, MS, CCRC. Howard University: Thomas O. Obisesan, MD, MPH; Saba Wolday, MSc; and Joanne Allard, PhD. University of Wisconsin: Sterling C. Johnson, PhD; J. Jay Fruehling, MA; and Sandra Harding, MS. University of Washington: Elaine R. Peskind, MD; Eric C. Petrie, MD, MS; and Gail Li, MD, PhD. Stanford University: Jerome A. Yesavage, MD; Joy L. Taylor, PhD; Ansgar J. Furst, PhD; and Steven Chao, MD. Cornell University: Norman Relkin, MD, PhD; Gloria Chiang, MD; and Lisa Ravdin, PhD.

ADNI Depression: Part A: Leadership and Infrastructure Principal Investigator: Scott Mackin, PhD, University of California San Francisco; ATRI PI and Director of Coordinating Center Clinical Core: Paul Aisen, MD, University of Southern California; Rema Raman, PhD, University of Southern California. Executive Committee: Scott Mackin, PhD, University of California San Francisco; Michael Weiner, MD, University of California San Francisco; Paul Aisen, MD, University of Southern California; Rema Raman, PhD, University of Southern California; Clifford R. Jack Jr, MD, Mayo Clinic, Rochester; Susan Landau, PhD, University of California Berkeley; Andrew J. Saykin, PsyD, Indiana University; Arthur W. Toga, PhD, University of Southern California; Charles DeCarli, MD, University of California Davis; Robert A. Koeppel, PhD, University of Michigan; Data and Publication Committee: Robert C. Green, MD,

MPH, BWH/HMS (Chair); Erin Drake, MA, BWM/HMS (Director); Clinical Core Leaders: Michael Weiner, MD (Core PI); Paul Aisen, MD, University of Southern California; Rema Raman, PhD, University of Southern California; Mike Donohue, PhD, University of Southern California; Clinical Informatics, Operations and Regulatory Affairs: Gustavo Jimenez, MBS, University of Southern California; Devon Gessert, BS, University of Southern California; Kelly Harless, BA, University of Southern California; Jennifer Salazar, MBS, University of Southern California; Yuliana Cabrera, BS, University of Southern California; Sarah Walter, MSc, University of Southern California; Lindsey Hergesheimer, BS, University of Southern California; Elizabeth Shaffer, BS; Psychiatry Site Leaders and Key Personnel: Scott Mackin, PhD, University of California San Francisco; Craig Nelson, MD, University of California San Francisco; David Bickford, BA, University of California San Francisco; Meryl Butters, PhD, University of Pittsburgh; and Michelle Zmuda, MA, University of Pittsburgh.

MRI Core Leaders and Key Personnel: Clifford R. Jack Jr, MD, Mayo Clinic, Rochester (Core PI); Matthew Bernstein, PhD, Mayo Clinic, Rochester; Bret Borowski, RT, Mayo Clinic, Rochester; Jeff Gunter, PhD, Mayo Clinic, Rochester; Matt Senjem, MS, Mayo Clinic, Rochester; Kejal Kantarci, MD, Mayo Clinic, Rochester; Chad Ward, BA, Mayo Clinic, Rochester; Denise Reyes, BS, Mayo Clinic, Rochester; PET Core Leaders and Key Personnel: Robert A. Koeppe, PhD, University of Michigan; Susan Landau, PhD, University of California Berkeley; Informatics Core Leaders and Key Personnel: Arthur W. Toga, PhD, University of Southern California (Core PI); Karen Crawford, University of Southern California; Scott Neu, PhD, University of Southern California.

Genetics Core Leaders and Key Personnel: Andrew J. Saykin, PsyD, Indiana University; Tatiana M. Foroud, PhD, Indiana University; Kelley M. Faber, MS, CCRC, Indiana University; Kwangsik Nho, PhD, Indiana University; Kelly N. Nudelman, Indiana University.

Part B: Investigators by Site: University of California San Francisco: Scott Mackin, PhD; Howard Rosen, MD; Craig Nelson, MD; David Bickford, BA; Yiu Ho Au, BA; Kelly Scherer, BS; Daniel Catalinotto, BA; Samuel Stark, BA; Elise Ong, BA; and Dariella Fernandez, BA. University of Pittsburgh: Meryl Butters, PhD; Michelle Zmuda, MA; Oscar L. Lopez, MD; MaryAnn Oakley, MA; and Donna M. Simpson, CRNP, MPH.

Alzheimer's Disease Metabolomics Consortium Team Members: Indiana University: Andrew Saykin (PI) & Team (ADNI Genomics Core Leader); and Kwangsik Nho. Helmholtz Zentrum Muenchen: Gabi Kastenmüller (PI); and Matthias Arnold (Co-PI). University of Arkansas: Sudeepa Bhattacharyya. University of Texas Health Science Center San Antonio: Xianlin Han (PI). West Coast Metabolomics Center: Oliver Fiehn (PI) & Team, Dinesh Barupal. Baker Heart and Diabetes Institute: Peter Meikle (PI). CalTech: Sarkis Mazmanian (PI). PO Metabolomics: Suzana Petanceska (NIH/NIA: 3 U01AGO24904-09S4; 1R01AGO46171-01). PO ADNI: John Hsiao (NIH/NIA/ERP); Michael Weiner and leadership of ADNI. University of Pennsylvania: Mitchel Kling (PI) & Team; John Toledo; Leslie Shaw (ADNI BiomarkerCore); and John Trojanowski (ADNI Biomarker Core). University of Oxford: Cornelia van Duijin (PI); and Shazad Ahmad (Erasmus). Leiden University Metabolomics Center: Thomas Hankemeier (PI) & Team. National University of Ireland-Galway: Ines Thiele (PI); Almut Heinken (Luxembourg). Institute for Systems Biology: Nathan Price (PI) & Team; Cory Funk; and Priyanka Baloni. University of Hawaii: Wei Jia (PI) & Team. The Metabolomics Innovation Centre Canada tTMIC: David Wishart (PI) & Team.

AMP-AD Collaborations: Rush University (David Bennen); Emory University (Allan Levey); SUNY (Herman Moreno); Columbia (Jose Luchsinger and Phil DeJager); Mt Sinai (Bin Zhang); Mayo-Florida (Nilufer Taner). University of Arizona: Roberta Brinton (PI) and Team; Rui Chang. Boston University: Lindsay Farrer (PI); Rhoda Au and Team. Biocrates Inc Metabolomics: Research Team. Nightingale Health: Peter Wurtz and Research Team. SAGE Networks: Lara Mangravire (PQ) and Team. Cornell University: Jan Krumsiek and Team. USDA: John Newman & Team. Duke University Medical Center, Psychiatry, Metabolomics Core and Statistics (Coordinating Center): Rima Kaddurah-Daouk (Overall PI); Alexandra Kueider-Paisley; P. Murali Doraiswamy (AD clinician); Colette Blach (Database); Art Moseley (Duke Proteomics and Metabolomics Core PI); Will Thompson, (Duke Proteomics and Metabolomics Core, Metabolomics Leader); Siamak Mahmoudiandehkordi (Statistics); Rebecca Baillie (Lipidmetabolism); KathleenWelsh-Bohmer; and Brenda Plassman.

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## REFERENCES

1. Toledo JB, Arnold M, Kastenmüller G, et al; Alzheimer's Disease Neuroimaging Initiative and the Alzheimer Disease Metabolomics Consortium. Metabolic network failures in Alzheimer's disease: a biochemical road map. *Alzheimers Dement*. 2017;13(9):965-984. doi:10.1016/j.jalz.2017.01.020



2. Clarke JR, Ribeiro FC, Frozza RL, De Felice FG, Lourenco MV. Metabolic dysfunction in Alzheimer's disease: from basic neurobiology to clinical approaches. *J Alzheimers Dis*. 2018;64(s1):S405-S426. doi:10.3233/JAD-179911
3. Kapogiannis D, Mattson MP. Disrupted energy metabolism and neuronal circuit dysfunction in cognitive impairment and Alzheimer's disease. *Lancet Neurol*. 2011;10(2):187-198. doi:10.1016/S1474-4422(10)70277-5
4. Craft S. The role of metabolic disorders in Alzheimer disease and vascular dementia: two roads converged. *Arch Neurol*. 2009;66(3):300-305. doi:10.1001/archneurol.2009.27
5. Sookoian S, Castaño GO, Scian R, et al. Serum aminotransferases in nonalcoholic fatty liver disease are a signature of liver metabolic perturbations at the amino acid and Krebs cycle level. *Am J Clin Nutr*. 2016;103(2):422-434. doi:10.3945/ajcn.115.118695
6. Sookoian S, Pirola CJ. Alanine and aspartate aminotransferase and glutamine-cycling pathway: their roles in pathogenesis of metabolic syndrome. *World J Gastroenterol*. 2012;18(29):3775-3781. doi:10.3748/wjg.v18.i29.3775
7. Goessling W, Massaro JM, Vasan RS, D'Agostino RB Sr, Ellison RC, Fox CS. Aminotransferase levels and 20-year risk of metabolic syndrome, diabetes, and cardiovascular disease. *Gastroenterology*. 2008;135(6):1935-1944. doi:10.1053/j.gastro.2008.09.018
8. Sattar N, Scherbakova O, Ford I, et al; West of Scotland Coronary Prevention Study. Elevated alanine aminotransferase predicts new-onset type 2 diabetes independently of classical risk factors, metabolic syndrome, and C-reactive protein in the West of Scotland Coronary Prevention Study. *Diabetes*. 2004;53(11):2855-2860. doi:10.2337/diabetes.53.11.2855
9. Santos CY, Snyder PJ, Wu W-C, Zhang M, Echeverria A, Alber J. Pathophysiologic relationship between Alzheimer's disease, cerebrovascular disease, and cardiovascular risk: a review and synthesis. *Alzheimers Dement (Amst)*. 2017;7:69-87.
10. Fillit H, Nash DT, Rundek T, Zuckerman A. Cardiovascular risk factors and dementia. *Am J Geriatr Pharmacother*. 2008;6(2):100-118. doi:10.1016/j.amjopharm.2008.06.004
11. Jack CR Jr, Bennett DA, Blennow K, et al; Contributors. NIA-AA research framework: toward a biological definition of Alzheimer's disease. *Alzheimers Dement*. 2018;14(4):535-562. doi:10.1016/j.jalz.2018.02.018
12. Alzheimer's Disease Neuroimaging Initiative (ADNI) website. <http://adni.loni.usc.edu/>. Accessed July 8, 2019.
13. Saykin AJ, Shen L, Yao X, et al; Alzheimer's Disease Neuroimaging Initiative. Genetic studies of quantitative MCI and AD phenotypes in ADNI: progress, opportunities, and plans. *Alzheimers Dement*. 2015;11(7):792-814. doi:10.1016/j.jalz.2015.05.009
14. Weiner MW, Veitch DP, Aisen PS, et al; Alzheimer's Disease Neuroimaging Initiative. Recent publications from the Alzheimer's Disease Neuroimaging Initiative: Reviewing progress toward improved AD clinical trials. *Alzheimers Dement*. 2017;13(4):e1-e85. doi:10.1016/j.jalz.2016.11.007
15. Petersen RC, Aisen PS, Beckett LA, et al; Alzheimer's Disease Neuroimaging Initiative (ADNI): clinical characterization. *Neurology*. 2010;74(3):201-209. doi:10.1212/WNL.0b013e3181cb3e25
16. Rattanabannakit C, Risacher SL, Gao S, et al. The Cognitive Change Index as a measure of self and informant perception of cognitive decline: relation to neuropsychological tests. *J Alzheimers Dis*. 2016;51(4):1145-1155. doi:10.3233/JAD-150729
17. McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology*. 1984;34(7):939-944. doi:10.1212/WNL.34.7.939
18. Aisen PS, Petersen RC, Donohue MC, et al; Alzheimer's Disease Neuroimaging Initiative. Clinical core of the Alzheimer's Disease Neuroimaging Initiative: progress and plans. *Alzheimers Dement*. 2010;6(3):239-246. doi:10.1016/j.jalz.2010.03.006
19. Crane PK, Carle A, Gibbons LE, et al; Alzheimer's Disease Neuroimaging Initiative. Development and assessment of a composite score for memory in the Alzheimer's Disease Neuroimaging Initiative (ADNI). *Brain Imaging Behav*. 2012;6(4):502-516. doi:10.1007/s11682-012-9186-z
20. Gibbons LE, Carle AC, Mackin RS, et al; Alzheimer's Disease Neuroimaging Initiative. A composite score for executive functioning, validated in Alzheimer's Disease Neuroimaging Initiative (ADNI) participants with baseline mild cognitive impairment. *Brain Imaging Behav*. 2012;6(4):517-527. doi:10.1007/s11682-012-9176-1
21. Jack CR Jr, Bernstein MA, Borowski BJ, et al; Alzheimer's Disease Neuroimaging Initiative. Update on the magnetic resonance imaging core of the Alzheimer's disease neuroimaging initiative. *Alzheimers Dement*. 2010;6(3):212-220. doi:10.1016/j.jalz.2010.03.004
22. Jack CR Jr, Bernstein MA, Fox NC, et al. The Alzheimer's Disease Neuroimaging Initiative (ADNI): MRI methods. *J Magn Reson Imaging*. 2008;27(4):685-691. doi:10.1002/jmri.21049

23. Kim S, Swaminathan S, Inlow M, et al; Alzheimer's Disease Neuroimaging Initiative (ADNI). Influence of genetic variation on plasma protein levels in older adults using a multi-analyte panel. *PLoS One*. 2013;8(7):e70269. doi:10.1371/journal.pone.0070269
24. Nho K, Corneveaux JJ, Kim S, et al; Multi-Institutional Research on Alzheimer Genetic Epidemiology (MIRAGE) Study; AddNeuroMed Consortium; Indiana Memory and Aging Study; Alzheimer's Disease Neuroimaging Initiative (ADNI). Whole-exome sequencing and imaging genetics identify functional variants for rate of change in hippocampal volume in mild cognitive impairment. *Mol Psychiatry*. 2013;18(7):781-787. doi:10.1038/mp.2013.24
25. Nho K, Kim S, Risacher SL, et al; MIRAGE (Multi-Institutional Research on Alzheimer Genetic Epidemiology) Study; AddNeuroMed Consortium; Indiana Memory and Aging Study; Alzheimer's Disease Neuroimaging Initiative. Protective variant for hippocampal atrophy identified by whole exome sequencing. *Ann Neurol*. 2015;77(3):547-552. doi:10.1002/ana.24349
26. Fischl B, Sereno MI, Dale AM. Cortical surface-based analysis, II: inflation, flattening, and a surface-based coordinate system. *Neuroimage*. 1999;9(2):195-207. doi:10.1006/nimg.1998.0396
27. Dale AM, Fischl B, Sereno MI. Cortical surface-based analysis, I: segmentation and surface reconstruction. *Neuroimage*. 1999;9(2):179-194. doi:10.1006/nimg.1998.0395
28. Chung MK, Worsley KJ, Nacewicz BM, Dalton KM, Davidson RJ. General multivariate linear modeling of surface shapes using SurfStat. *Neuroimage*. 2010;53(2):491-505. doi:10.1016/j.neuroimage.2010.06.032
29. Risacher SL, Kim S, Nho K, et al; Alzheimer's Disease Neuroimaging Initiative (ADNI). APOE effect on Alzheimer's disease biomarkers in older adults with significant memory concern. *Alzheimers Dement*. 2015;11(12):1417-1429. doi:10.1016/j.jalz.2015.03.003
30. Bittner T, Zetterberg H, Teunissen CE, et al. Technical performance of a novel, fully automated electrochemiluminescence immunoassay for the quantitation of  $\beta$ -amyloid (1-42) in human cerebrospinal fluid. *Alzheimers Dement*. 2016;12(5):517-526. doi:10.1016/j.jalz.2015.09.009
31. Hansson O, Seibyl J, Stomrud E, et al; Swedish BioFINDER study group; Alzheimer's Disease Neuroimaging Initiative. CSF biomarkers of Alzheimer's disease concord with amyloid- $\beta$  PET and predict clinical progression: a study of fully automated immunoassays in BioFINDER and ADNI cohorts. *Alzheimers Dement*. 2018;14(11):1470-1481. doi:10.1016/j.jalz.2018.01.010
32. Noble KG, Grieve SM, Korgaonkar MS, et al. Hippocampal volume varies with educational attainment across the life-span. *Front Hum Neurosci*. 2012;6:307. doi:10.3389/fnhum.2012.00307
33. Worsley KJ. SurfStat. <http://www.math.mcgill.ca/keith/surfstat/>. Accessed July 8, 2019.
34. SPM: Statistical Parametric Mapping. <https://www.fil.ion.ucl.ac.uk/spm/>. Accessed July 8, 2019.
35. Hagler DJ Jr, Saygin AP, Sereno MI. Smoothing and cluster thresholding for cortical surface-based group analysis of fMRI data. *Neuroimage*. 2006;33(4):1093-1103. doi:10.1016/j.neuroimage.2006.07.036
36. Hayasaka S, Phan KL, Liberzon I, Worsley KJ, Nichols TE. Nonstationary cluster-size inference with random field and permutation methods. *Neuroimage*. 2004;22(2):676-687. doi:10.1016/j.neuroimage.2004.01.041
37. Worsley KJ, Taylor JE, Tomaiuolo F, Lerch J. Unified univariate and multivariate random field theory. *Neuroimage*. 2004;23(suppl 1):S189-S195. doi:10.1016/j.neuroimage.2004.07.026
38. Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Stat Soc Series B Stat Methodol*. 1995;57(1):289-300. doi:10.2307/2346101
39. Jack CR Jr, Bennett DA, Blennow K, et al. NIA-AA research framework: towards a biological definition of Alzheimer's disease. Paper presented at: Alzheimer's Association International Conference; November 27, 2017; London, England.
40. Giambattistelli F, Bucossi S, Salustri C, et al. Effects of hemochromatosis and transferrin gene mutations on iron dyshomeostasis, liver dysfunction and on the risk of Alzheimer's disease. *Neurobiol Aging*. 2012;33(8):1633-1641. doi:10.1016/j.neurobiolaging.2011.03.005
41. Pietzner M, Budde K, Homuth G, et al. Hepatic steatosis is associated with adverse molecular signatures in subjects without diabetes. *J Clin Endocrinol Metab*. 2018;103(10):3856-3868. doi:10.1210/jc.2018-00999
42. Varma VR, Oommen AM, Varma S, et al. Brain and blood metabolite signatures of pathology and progression in Alzheimer disease: a targeted metabolomics study. *PLoS Med*. 2018;15(1):e1002482. doi:10.1371/journal.pmed.1002482
43. Kaddurah-Daouk R, Doraiswamy PM, Zhu H, et al. Alterations in metabolic pathways and networks in mild cognitive impairment and early Alzheimer's disease. *Alzheimers Dement*. 2013;9(4)(suppl):P571. doi:10.1016/j.jalz.2013.05.1126

44. Kaddurah-Daouk R, Rozen S, Matson W, et al. Metabolomic changes in autopsy-confirmed Alzheimer's disease. *Alzheimers Dement*. 2011;7(3):309-317. doi:10.1016/j.jalz.2010.06.001
45. Botros M, Sikaris KA. The de ritis ratio: the test of time. *Clin Biochem Rev*. 2013;34(3):117-130.
46. Mosconi L, Sorbi S, de Leon MJ, et al. Hypometabolism exceeds atrophy in presymptomatic early-onset familial Alzheimer's disease. *J Nucl Med*. 2006;47(11):1778-1786.
47. Drzezga A, Lautenschlager N, Siebner H, et al. Cerebral metabolic changes accompanying conversion of mild cognitive impairment into Alzheimer's disease: a PET follow-up study. *Eur J Nucl Med Mol Imaging*. 2003;30(8):1104-1113. doi:10.1007/s00259-003-1194-1
48. Rui L. Energy metabolism in the liver. *Compr Physiol*. 2014;4(1):177-197. doi:10.1002/cphy.c130024
49. Ellinger JJ, Lewis IA, Markley JL. Role of aminotransferases in glutamate metabolism of human erythrocytes. *J Biomol NMR*. 2011;49(3-4):221-229. doi:10.1007/s10858-011-9481-9
50. Qian K, Zhong S, Xie K, Yu D, Yang R, Gong D-W. Hepatic ALT isoenzymes are elevated in gluconeogenic conditions including diabetes and suppressed by insulin at the protein level. *Diabetes Metab Res Rev*. 2015;31(6):562-571. doi:10.1002/dmrr.2655
51. Reis HJ, Guatimosim C, Paquet M, et al. Neuro-transmitters in the central nervous system & their implication in learning and memory processes. *Curr Med Chem*. 2009;16(7):796-840. doi:10.2174/092986709787549271
52. Fleck MW, Henze DA, Barrionuevo G, Palmer AM. Aspartate and glutamate mediate excitatory synaptic transmission in area CA1 of the hippocampus. *J Neurosci*. 1993;13(9):3944-3955. doi:10.1523/JNEUROSCI.13-09-03944.1993
53. Francis PT. Glutamatergic systems in Alzheimer's disease. *Int J Geriatr Psychiatry*. 2003;18(suppl 1):S15-S21. doi:10.1002/gps.934
54. Kamada Y, Hashimoto R, Yamamori H, et al. Impact of plasma transaminase levels on the peripheral blood glutamate levels and memory functions in healthy subjects. *BBA Clin*. 2016;5:101-107. doi:10.1016/j.bbaci.2016.02.004
55. Alfredsson G, Wiesel FA, Tylec A. Relationships between glutamate and monoamine metabolites in cerebrospinal fluid and serum in healthy volunteers. *Biol Psychiatry*. 1988;23(7):689-697. doi:10.1016/0006-3223(88)90052-2
56. Wang G, Zhou Y, Huang FJ, et al. Plasma metabolite profiles of Alzheimer's disease and mild cognitive impairment. *J Proteome Res*. 2014;13(5):2649-2658. doi:10.1021/pr5000895
57. Lowe SL, Bowen DM, Francis PT, Neary D. Ante mortem cerebral amino acid concentrations indicate selective degeneration of glutamate-enriched neurons in Alzheimer's disease. *Neuroscience*. 1990;38(3):571-577. doi:10.1016/0306-4522(90)90051-5
58. Fayed N, Modrego PJ, Rojas-Salinas G, Aguilar K. Brain glutamate levels are decreased in Alzheimer's disease: a magnetic resonance spectroscopy study. *Am J Alzheimers Dis Other Dement*. 2011;26(6):450-456. doi:10.1177/1533317511421780
59. Procter AW, Palmer AM, Francis PT, et al. Evidence of glutamatergic denervation and possible abnormal metabolism in Alzheimer's disease. *J Neurochem*. 1988;50(3):790-802. doi:10.1111/j.1471-4159.1988.tb02983.x
60. Liu Z, Ning H, Que S, Wang L, Qin X, Peng T. Complex association between alanine aminotransferase activity and mortality in general population: a systematic review and meta-analysis of prospective studies. *PLoS One*. 2014;9(3):e91410. doi:10.1371/journal.pone.0091410
61. Peltz-Sinvani N, Klempfner R, Ramaty E, Sela BA, Goldenberg I, Segal G. Low ALT levels independently associated with 22-year all-cause mortality among coronary heart disease patients. *J Gen Intern Med*. 2016;31(2):209-214. doi:10.1007/s11606-015-3480-6
62. Vespasiani-Gentilucci U, De Vincentis A, Ferrucci L, Bandinelli S, Antonelli Incalzi R, Picardi A. Low alanine aminotransferase levels in the elderly population: frailty, disability, sarcopenia, and reduced survival. *J Gerontol A Biol Sci Med Sci*. 2018;73(7):925-930. doi:10.1093/gerona/glx126
63. Elinav E, Ben-Dov IZ, Ackerman E, et al. Correlation between serum alanine aminotransferase activity and age: an inverted U curve pattern. *Am J Gastroenterol*. 2005;100(10):2201-2204. doi:10.1111/j.1572-0241.2005.41822.x
64. Kaiser LG, Schuff N, Cashdollar N, Weiner MW. Age-related glutamate and glutamine concentration changes in normal human brain: 1H MR spectroscopy study at 4 T. *Neurobiol Aging*. 2005;26(5):665-672. doi:10.1016/j.neurobiolaging.2004.07.001
65. Guerreiro R, Bras J. The age factor in Alzheimer's disease. *Genome Med*. 2015;7:106. doi:10.1186/s13073-015-0232-5

66. Katsiki N, Perez-Martinez P, Anagnostis P, Mikhailidis DP, Karagiannis A. Is nonalcoholic fatty liver disease indeed the hepatic manifestation of metabolic syndrome? *Curr Vasc Pharmacol*. 2018;16(3):219-227. doi:10.2174/1570161115666170621075619
67. Weinstein G, Zelber-Sagi S, Preis SR, et al. Association of nonalcoholic fatty liver disease with lower brain volume in healthy middle-aged adults in the Framingham Study. *JAMA Neurol*. 2018;75(1):97-104. doi:10.1001/jamaneurol.2017.3229
68. Bedogni G, Gastaldelli A, Tiribelli C, et al. Relationship between glucose metabolism and non-alcoholic fatty liver disease severity in morbidly obese women. *J Endocrinol Invest*. 2014;37(8):739-744. doi:10.1007/s40618-014-0101-x
69. Perla FM, Prelati M, Lavorato M, Visicchio D, Anania C. The role of lipid and lipoprotein metabolism in non-alcoholic fatty liver disease. *Children (Basel)*. 2017;4(6):E46.
70. Kellett KAB, Williams J, Vardy ER, Smith AD, Hooper NM. Plasma alkaline phosphatase is elevated in Alzheimer's disease and inversely correlates with cognitive function. *Int J Mol Epidemiol Genet*. 2011;2(2):114-121.
71. Moss DW. Physicochemical and pathophysiological factors in the release of membrane-bound alkaline phosphatase from cells. *Clin Chim Acta*. 1997;257(1):133-140. doi:10.1016/S0009-8981(96)06438-8
72. Goldstein DJ, Rogers CE, Harris H. Expression of alkaline phosphatase loci in mammalian tissues. *Proc Natl Acad Sci U S A*. 1980;77(5):2857-2860. doi:10.1073/pnas.77.5.2857
73. Fonta C, Négyessy L, Renaud L, Barone P. Areal and subcellular localization of the ubiquitous alkaline phosphatase in the primate cerebral cortex: evidence for a role in neurotransmission. *Cereb Cortex*. 2004;14(6):595-609. doi:10.1093/cercor/bhh021
74. Waymire KG, Mahuren JD, Jaje JM, Guilarte TR, Coburn SP, MacGregor GR. Mice lacking tissue non-specific alkaline phosphatase die from seizures due to defective metabolism of vitamin B-6. *Nat Genet*. 1995;11(1):45-51. doi:10.1038/ng0995-45
75. Narisawa S, Wennberg C, Millán JL. Abnormal vitamin B6 metabolism in alkaline phosphatase knock-out mice causes multiple abnormalities, but not the impaired bone mineralization. *J Pathol*. 2001;193(1):125-133. doi:10.1002/1096-9896(2000)9999:9999::AID-PATH722>3.0.CO;2-Y
76. Langer D, Ikehara Y, Takebayashi H, Hawkes R, Zimmermann H. The ectonucleotidases alkaline phosphatase and nucleoside triphosphate diphosphohydrolase 2 are associated with subsets of progenitor cell populations in the mouse embryonic, postnatal and adult neurogenic zones. *Neuroscience*. 2007;150(4):863-879. doi:10.1016/j.neuroscience.2007.07.064
77. Yamashita M, Sasaki M, Mii K, et al. Measurement of serum alkaline phosphatase isozyme I in brain-damaged patients. *Neurol Med Chir (Tokyo)*. 1989;29(11):995-998. doi:10.2176/nmc.29.995
78. Gjerde H, Amundsen A, Skog O-J, Mørland J, Aasland OG. Serum gamma-glutamyltransferase: an epidemiological indicator of alcohol consumption? *Br J Addict*. 1987;82(9):1027-1031. doi:10.1111/j.1360-0443.1987.tb01564.x

#### SUPPLEMENT.

**eTable 1.** Sensitivity Analysis for Alzheimer Disease Diagnosis (CN vs AD) Group Differences in Liver Function Biomarkers

**eTable 2.** Demographic Information of ADNI Participants

**eTable 3.** Diagnostic Group Differences in Liver Function Biomarkers

**eTable 4.** Diagnostic Group Differences of Liver Function Biomarkers With Alzheimer Disease Diagnosis (CV vs AD) Adjusted for Gamma-Glutamyltransferase (GGT) and Statin Use

**eTable 5.** Association of Liver Function Biomarkers With Cognition Adjusted for Gamma-Glutamyltransferase (GGT) and Statin Use

**eFigure.** Liver Function Biomarkers and Their Association With A/T/N Biomarkers for Alzheimer Disease Adjusted for Gamma-Glutamyltransferase (GGT) and Statin Use