



**UNIVERSITI PUTRA MALAYSIA**

***INTERACTION BETWEEN AN ENTOMOPATHOGENIC FUNGUS  
Metarhizium anisopliae AND ITS HOST SUBTERRANEAN TERMITES  
Coptotermes curvignathus DURING INFECTION PROCESS***

**AHMAD SYAZWAN BIN SAMSUDDIN**

**FH 2017 5**



**UPM**  
UNIVERSITI PUTRA MALAYSIA  
BERILMU BERBAKTI

**INTERACTION BETWEEN AN ENTOMOPATHOGENIC FUNGUS  
*Metarhizium anisopliae* AND ITS HOST SUBTERRANEAN TERMITES  
*Coptotermes curvignathus* DURING INFECTION PROCESS**

By

**AHMAD SYAZWAN BIN SAMSUDDIN**

**Thesis submitted to School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of Master of Science**

**April 2017**



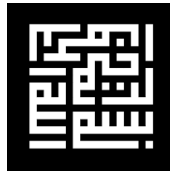
© COPYRIGHT UPM

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia

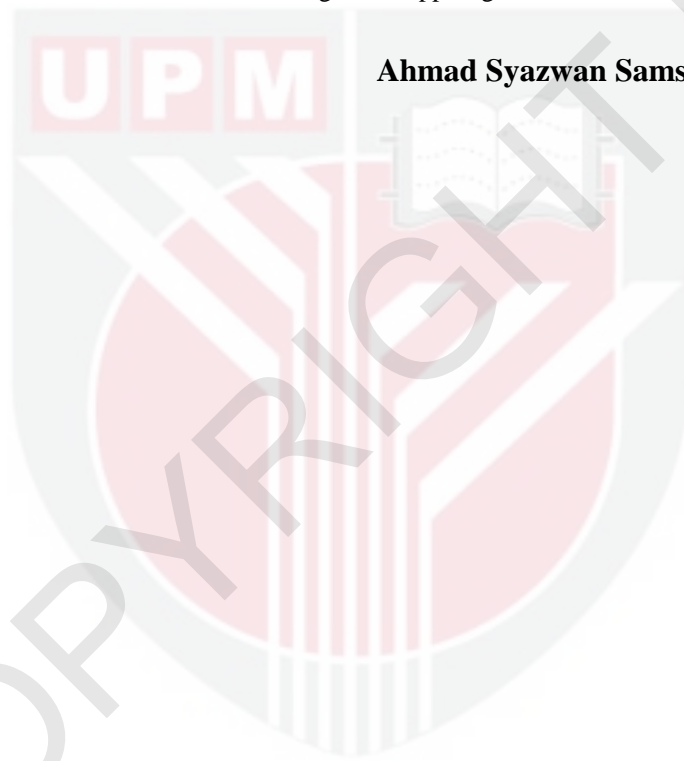


## DEDICATION



This thesis was dedicated to my Ayah and Ibu, Samsuddin Mohamed and Zaharina Sharif for love, courage and support given.

**Ahmad Syazwan Samsuddin**



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Master of Science

**INTERACTION BETWEEN AN ENTOMOPATHOGENIC FUNGUS  
*Metarhizium anisopliae* AND ITS HOST SUBTERRANEAN TERMITES  
*Coptotermes curvignathus* DURING INFECTION PROCESS**

By

**AHMAD SYAZWAN BIN SAMSUDDIN**

**April 2017**

**Chairman : Professor Ahmad Said Bin Sajap, PhD**  
**Faculty : Forestry**

The subterranean termites, *Coptotermes curvignathus* is a destructive pest of tree plantation in Malaysia. Controlling termite population by chemical is not a good option since it may increase the resistance of pest towards the pesticide and cause several implications towards biodiversity and environment. *Metarhizium anisopliae*, an entomopathogenic fungus is a good option by controlling the termite biologically. In this study, ten *M. anisopliae* isolates were screened for their pathogenic effect on termites. The best isolate, PR1 which originated from Pantai Remis, Selangor is the most prudently isolates by yielded the highest mortality (96.97%) and the shortest median lethal time ( $LT_{50}$ = 1.47 days). This isolate was utilized in downstream study by conducting the histopathology and protein expression study; for each stages of pathogenesis up to 144 h of post-inoculation. Results demonstrated that it requires two days to kill infected host starting by germ tube penetration through the termite cuticle observed at 6 h post-inoculation. Injury caused by *M. anisopliae*'s appressorium in a way to penetrate the first layer of host defense mechanism, the cuticle, one of the factors that weakening the infected host. Dissemination and of fungal and hyphal bodies observed on cellular level is a proof that *M. anisopliae* invaded the host starting from the cellular level. Then, inoculated termites were mummified and the sporulation of *M. anisopliae* on host's carcasses observed at 96 h and 144 h of post-inoculation, respectively. Meanwhile in protein expression study, four and five significance expressed proteins have been identified from *C. curvignathus* and *M. anisopliae*, respectively. Expressed proteins in termite shows evidence that being involve in cell regulation and immune response towards infection by this entomopathogenic fungus. Meanwhile, the expressed proteins extracted from *M. anisopliae* were discovered to be involved in the protein transportation and also related with fungus virulency during infection process. By knowing the interaction between this host-pathogen relationship, the achieved knowledge could be utilized in future development of biopesticide by using this fungus to control termite population effectively.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**INTERAKSI DI ANTARA KULAT PEMBUNUH SERANGGA *Metarhizium anisopliae* DAN ANAI-ANAI BAWAH TANAH *Coptotermes curvignathus* SEMASA PROSES JANGKITAN**

Oleh

**AHMAD SYAZWAN BIN SAMSUDDIN**

**April 2017**

**Pengerusi : Profesor Ahmad Said Bin Sajap, PhD**  
**Fakulti : Perhutanan**

*Coptotermes curvignathus* adalah salah satu spesies anai-anai merupakan perosak yang merbahaya terhadap dirian pokok di Malaysia. Pengawalan populasi anai-anai dengan menggunakan bahan kimia adalah salah satu pilihan yang kurang tepat memandangkan ianya mampu meningkatkan kerintangan serangga tersebut terhadap racun kimia tersebut, malahan mampu memberi impak negatif terhadap biodiversiti dan alam sekitar. Penggunaan kulat pembunuh serangga, *Metarhizium anisopliae* adalah salah satu alternatif terbaik dalam mengawal populasi serangga perosak ini secara biologi. Sepuluh pencilan kulat *M. anisopliae* telah disaring bagi menentukan kesan patogeniknya terhadap anai-anai. Kajian mendapati pencilan PR1, pencilan yang dipencil dari Pantai Remis, Selangor adalah pencilan yang terbaik. Ia mampu membunuh 96.97% dari populasi anai anai dan menyebabkan masa membunuh median, LT<sub>50</sub> sebanyak 1.47 hari terhadap anai-anai yang terjangkit. Pencilan ini seterusnya digunakan di dalam kajian berikutnya iaitu kajian histopatologi dan ekspresi protein pada setiap peringkat patogenesis sehingga 144 h pasca-inokulasi. Keputusan menunjukkan ianya memerlukan dua hari untuk membunuh hos terjangkit bermula dengan penembusan tiub germa melalui kutikel pada 6 h pasca-inokulasi. Kecederaan yang disebabkan oleh appressorium kulat untuk menembusi lapisan pertama mekanisme pertahanan, iaitu kutikel serangga itu sendiri adalah merupakan faktor utama yang melemahkan hos terjangkit. Penyebaran kulat dan badan hifa dilihat di peringkat selular membuktikan penaklukan *M. anisopliae* bermula dari peringkat selular. Anai-anai terbalut dengan miselium dan sporulasi oleh kulat ini diperhatikan pada 96 h dan 144 h pasca-inokulasi. Manakala di dalam kajian ekspresi protein, penemuan mendapati empat dan lima protein yang signifikan dari kedua-dua organisma, *C. curvignathus* dan *M. anisopliae* dikenalpasti terlibat di dalam proses jangkitan. Protein yang diekspres oleh anai-anai menunjukkan bukti penglibatan protein di dalam regulasi sel dan tindakbalas imun apabila terjangkit oleh *M. anisopliae*. Selain itu, protein yang diekspres oleh kulat ini mempunyai peranan yang penting didalam pengangkutan protein dan virulensiti kulat ini sendiri ketika proses jangkitan. Berdasarkan interaksi yang dilihat dan dikenalpasti di dalam hubungan perumah-patogen ini, informasi yang didapati mungkin boleh

digunakan di dalam pembangunan racun serangga biologi dengan menggunakan kulat pembunuh serangga ini dengan lebih efektif dalam mengawal populasi anai-anai perosak ini di masa akan datang.





## ACKNOWLEDGEMENT

First and foremost, I would like to thank to Allah for giving me strength, love, courage and wisdom to complete my study.

I would like to express my deepest appreciation to my supervisor, Prof. Ahmad Said Sajap and my co-supervisor, Dr. Rozi Mohamed for the strong and firm provision, sharing their valuable knowledge and intelligence, and encouragement that help me to finish my study. I also would like to thank to my supervisory committee, Dr. Lau Wei Hong and Prof Dzolkifli Omar for the advice and knowledge throughout my study.

I would like to express my sincere gratitude to my parents, Ibu and Ayah for love, trust and endless support which that cannot be replace by others. I also would like to express my special thank to my best friend, Wan Azrul, Fahrul Azmi, Ariff Ramli and Shamil Anwar Ibrahim which always help me, lend a shoulder to cry on and always keep my chin up whenever I needed.

I would also like to thank to my laboratory members, Li Peng, Jessica, Shiou Yih, Mun Theng and Yaqin for all knowledge, guidance and interminable helps throughout my study here in Faculty of Forestry. Not to forget to all my siblings, Angah, Alang, Adik and Amar and also to all my “H’s” friend, Hafifi, Hjis, Hisyam, Hazri and Hairul for always stand behind me and giving me hope and trust in whatever I do in my life.

Not to forget to all staff and friend of Faculty of Forestry, Forest Research Institute Malaysia, Institute of Bioscience, Faculty of Biotechnology and Biomolecular Science, Agro-Biotechnology Institute Malaysia, Faculty of Medicine (Universiti Malaya), Universiti Teknologi Mara, Inbiosis (UKM) and School of Graduate Study and to whom that I forgot to mentioned here; I believe you know who you are that help me whether directly or indirectly throughout my master study here in UPM.

I certify that a Thesis Examination Committee has met on 14 April 2017 to conduct the final examination of Ahmad Syazwan bin Samsuddin on his thesis entitled "Interaction Between an Entomopathogenic Fungus *Metarhizium anisopliae* and its Host Subterranean Termites *Coptotermes curvignathus* during Infection Process" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

**H Ng Paik San, PhD**

Associate Professor  
Faculty of Forestry  
Universiti Putra Malaysia  
(Chairman)

**Puad bin Abdullah, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Idris Abd Ghani, PhD**

Professor  
Universiti Kebangsaan Malaysia  
Malaysia  
(External Examiner)



---

**NOR AINI AB. SHUKOR, PhD**  
Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 8 August 2017

This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Ahmad Said Sajap, PhD**

Professor  
Faculty of Forestry  
Universiti Putra Malaysia  
(Chairman)

**Rozi Mohamed, PhD**

Associate Professor  
Faculty of Forestry  
Universiti Putra Malaysia  
(Member)

**Lau Wei Hong, PhD**

Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Member)

**Dzolkhifli Omar, PhD**

Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia.

Date:

## Declaration by graduate student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations and citations have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office of Deputy Vice Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putera Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Ahmad Syazwan Bin Samsuddin (GS36207)

## Declaration by Members of Supervisory Committee

This is to confirm that:

- The research conducted and the writing of this thesis was under our supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature : \_\_\_\_\_

Name of

Chairman of

Supervisory

Committee: Professor Dr. Ahmad Said Sajap

Signature : \_\_\_\_\_

Name of

Member of

Supervisory

Committee: Associate Professor Dr. Rozi Mohamed

Signature : \_\_\_\_\_

Name of

Member of

Supervisory

Committee: Associate Professor Dr. Lau Wei Hong

Signature : \_\_\_\_\_

Name of

Member of

Supervisory

Committee: Professor Dr. Dzolkhifli Omar

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	ii
<b>ACKNOWLEDGEMENT</b>	iv
<b>APPROVAL</b>	v
<b>DECLARATION</b>	vi
<b>LIST OF TABLES</b>	xii
<b>LIST OF FIGURES</b>	xiii
<b>GLOSSARY OF TERM</b>	xv
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	1
<b>2 LITERATURE REVIEW</b>	
2.1 Termites	3
2.2 Subterranean termites	3
2.3 Overview of <i>Coptotermes curvignathus</i>	4
2.3.1 Biology, morphology and identification	4
2.3.2 Distribution and Infestation	6
2.3.3 Damage and economic importance	8
2.4 Management of <i>Coptotermes curvignathus</i>	8
2.4.1 Chemical control	8
2.4.2 Biological control	9
2.4.2.1 Bacteria	9
2.4.2.2 Entomopathogenic fungi	9
2.5 <i>Metarhizium anisopliae</i> as an entomopathogenic fungus	9
2.6 The host-pathogen interaction between <i>M. anisopliae</i> and termites	10
2.7 Proteomics as a tool in host-pathogen relationship study	11
<b>3 PATHOGENESIS OF DIFFERENT ISOLATES OF <i>Metarhizium anisopliae</i> ON INFECTED <i>Coptotermes curvignathus</i></b>	
3.1 Introduction	12
3.2 Materials and method	
3.2.1 Termite source	12
3.2.2 Fungal isolation	13
3.2.3 Colony morphology	14
3.2.4 DNA isolation	14
3.2.5 PCR for fungal identification	14
3.2.6 Phylogenetic tree construction	15
3.2.7 Preparation of conidial suspension	15
3.2.8 Pathogenicity test	15
3.2.9 Data analysis	16
3.3 Results and discussion	
3.3.1 Colony morphology of isolated fungi	16

3.3.2	Fungal identification through phylogeny tree construction	16
3.3.3	Pathogenicity test and LT <sub>50</sub> values on infected <i>C. curvignathus</i>	19
3.3.4	Growth and development of <i>M. anisopliae</i> on infected <i>C. curvignathus</i>	21
3.4	Conclusion	23
<b>4</b>	<b>HISTOPATHOLOGY OF INFECTED <i>Coptotermes curvignathus</i> WITH <i>Metarhizium anisopliae</i></b>	
4.1	Introduction	24
4.2	Materials and method	
4.2.1	Termite collection	24
4.2.2	Fungal culture	25
4.2.3	Preparation of conidial suspension and inoculation	25
4.2.4	Specimen preparation for electron microscopy	25
4.2.5	Specimen processing for scanning electron microscopy	26
4.2.6	Specimen processing for transmission electron microscopy	26
4.3	Results and discussion	
4.3.1	<i>M. anisopliae</i> pathogenesis on epicuticles of <i>C. curvignathus</i>	26
4.3.2	Dissemination, invasion and colonization of <i>M. anisopliae</i> in termites cellular level	28
4.4	Conclusion	31
<b>5</b>	<b>PROTEIN EXPRESSION AND IDENTIFICATION IN <i>Coptotermes curvignathus</i> AND <i>Metarhizium anisopliae</i> DURING PATHOGENESIS</b>	
5.1	Introduction	32
5.2	Materials and methods	
5.2.1	Fungal inoculation	32
5.2.2	Total protein extraction	33
5.2.3	Bradford Assay	33
5.2.4	Protein clean-up	33
5.2.5	Two-dimensional gel electrophoresis	34
5.2.6	Gel image and protein expression analysis	34
5.2.7	In-gel digestion	35
5.2.8	Mass spectrometry	35
5.2.9	Protein identification	35
5.3	Results and discussion	
5.3.1	Protein expression during pathogenesis	36
5.3.2	Expression of significance identified proteins involved during pathogenesis	37
5.5	Conclusion	44

<b>6</b>	<b>SUMMARY , CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>45</b>
	<b>REFERENCES</b>	<b>47</b>
	<b>APPENDICES</b>	<b>63</b>
	<b>BIODATA OF STUDENT</b>	<b>99</b>
	<b>PUBLICATION</b>	<b>100</b>





## LIST OF TABLES

<b>Table</b>		<b>Page</b>
2.1	List of termites Families and number of genera and species within each Family	3
2.2	List of Subfamilies and number of genera and species within each Subfamily	4
2.3	Origin and distribution of <i>Coptotermes curvignathus</i> in South East Asia region	7
2.4	List of experiment on <i>M. anisopliae</i> in controlling termites	10
3.1	List of local isolates of <i>Metarhizium anisopliae</i> and their localities	13
3.2	Mean percent mortality of <i>C. curvignathus</i> after being infected with different isolates of <i>M. anisopliae</i>	19
3.3	Median lethal time (LT <sub>50</sub> ) of <i>C. curvignathus</i> after being infected with different isolates of <i>M. anisopliae</i>	21
3.4	Mean percent of mycelia formation on <i>C. curvignathus</i> after infected with different isolates of <i>M. anisopliae</i>	22
3.5	Mean percent conidia sporulation on <i>C. curvignathus</i> after infected with different isolates of <i>M. anisopliae</i>	22
C1	Kaplan-Meier analysis log report	68
C2	Case processing summary of analysed sample	69
C3	Means and medians for survival time	70
J1	Summary of mean normalized value of protein expression in each selected protein spots of <i>C. curvignathus</i>	82
J2	Summary of mean normalized value of protein expression in each selected protein spots of <i>M. anisopliae</i>	83
L1	Mass fingerprinting data obtained by MALDI-TOF identification of 14 differentially expressed proteins extracted from infected <i>C. curvignathus</i> with <i>M. anisopliae</i>	95
L2	Mass fingerprinting data obtained by MALDI-TOF identification of 10 differentially expressed proteins extracted from <i>M. anisopliae</i> infecting <i>C. curvignathus</i>	97

## LIST OF FIGURES

Figure		Page
2.1	Life cycle of subterranean termites, <i>C. curvignathus</i> . Q: queen; K: king; L: larvae; W: worker; E: ergatoids; PS: presoldier; S: soldier; N: nymph; A: alate; B: brachypterous neotenic; i: wingless worker pathway; ii: wingpad nymphal pathway	5
2.2	Differences between head of <i>Coptotermes</i> species. A: <i>C. curvignathus</i> ; B: <i>C. kalshoveni</i> ; C: <i>C. sepangensis</i> ; D: <i>C. gestroi</i> ; E: <i>C. havilandi</i>	6
2.3	Morphology of <i>C. curvignathus</i> soldier. A: dorsal view of head; B: lateral view of head; C: pronotum	6
2.4.	Distribution of <i>Coptotermes curvignathus</i> in South East Asia region	7
3.1	Amplified ITS region of all isolated <i>M. anisopliae</i> together with 100bp DNA ladder	17
3.2	Phylogenetic tree of all isolated studied <i>Metarhizium anisopliae</i> together with other same species within same genera <i>Metarhizium</i> . <i>C. sinensis</i> , <i>B. malawiensis</i> and <i>I. fumosoroseus</i> are as an out group	18
3.3	Kaplan-Meier curves showing the survival trend of <i>C. curvignathus</i> over the first 5 days of post infection by the three most pathogenic isolates of <i>M. anisopliae</i>	20
4.1	SEM images of <i>M. anisopliae</i> pathogenesis on <i>C. curvignathus</i> epicuticles. Pathogenesis starts with the attachment of fungal conidia on the thorax (A) and leg (B) leg of <i>C. curvignathus</i> . Conidia successfully attached near hairs (C) 1 to 3 hours after inoculation, after which they germinate at 6h post inoculation (D). Ha: hair; Co: conidia; Gt: germ tube	27
4.2	Germinated conidia start to penetrate the epicuticle at 6h post inoculation (A). Elongation of appressorium tube is assumed along the host's exocuticle (B). Massive attachment of inoculated <i>M. anisopliae</i> 's conidia on the termite's epicuticle is obviously seen at 24h post inoculation (C). Mummification process of termite's body is initiated by elongation of <i>M. anisopliae</i> 's hypha into mycelium at 96h post inoculation (D and E). Production of conidia in abundance at 144h post inoculation (F). Ha: hair; Co: conidia; Gt: germ tube; Ap: Appressorium tube; My: mycelium	28
4.3	Penetrant hypha (Ph) pressing the termite epidermis cell	29
4.4	Dissemination of fungal bodies (arrows) within hemocoel	29

4.5	Mass number of fungal bodies (arrows) of <i>M. anisopliae</i> colonize within termites intercellular	30
5.1	Pool of total protein of <i>C. curvignathus</i> and <i>M. anisopliae</i> during infection process separated by 2-D electrophoresis (green – total protein expressed by <i>M. anisopliae</i> ; blue – total protein expressed by <i>C. curvignathus</i> )	36
5.2	Total protein of <i>C. curvignathus</i> from control group separated by 2-D electrophoresis	38
5.3	Expression of four significance protein extracted from infected <i>C. curvignathus</i> . X-axis: time (hours) of post-inoculation; Y-axis: Y-axis: mean of normalized of protein spot intensities ( $\times 10^3$ )	39
5.4	Total protein of <i>M. anisopliae</i> from control group separated by 2-D electrophoresis	40
5.5	Expression of five significance protein extracted from <i>M. anisopliae</i> . X-axis: time (hours) of post-inoculation; Y-axis: Y-axis: mean of normalized of protein spot intensities ( $\times 10^3$ )	43
A1	Local <i>M. anisopliae</i> isolates culture (A: PR1, B: HSAH5, C: HM, D: GT3, E: GJ4, F: TFFH3, G: PPK, H: TK, I: USSA, J: PKLG)	63
D1	Pathogenesis of <i>M. anisopliae</i> on subteranean termites, <i>C. curvignathus</i> . A: Pre-inoculated termites, B: Termite was mummified with inoculated <i>M. anisopliae</i> , C: Sporulated <i>M. anisopliae</i> conidia on termite carcass	71
H1	Bovine Serum Albumin (BSA) standard curve	78
I1	2D-PAGE gels of separated total protein representative for control groups ( <i>C. curvignathus</i> and <i>M. anisopliae</i> ) and each time point of post-inoculation	80
J1 (a)	Protein expression of significance spot selected from <i>C. curvignathus</i> . X-axis: time (hours) of post-inoculation; Y-axis: mean of normalized of protein spot intensities ( $\times 10^3$ )	84
J1 (b)	Protein expression of significance spot selected from <i>C. curvignathus</i> . X-axis: time (hours) of post-inoculation; Y-axis: mean of normalized of protein spot intensities ( $\times 10^3$ )	85
J2	Protein expression of significance spot selected from <i>M. anisopliae</i> . X-axis: time (hours) of post-inoculation; Y-axis: mean of normalized of protein spot intensities ( $\times 10^3$ )	86
K1	Mass spectrum of digested protein sample from infected <i>C. curvignathus</i> total protein	87
K2	Mass spectrum of digested protein sample from <i>M. anisopliae</i> total protein	92

## GLOSSARY OF TERM

ANOVA	Analysis of Variance
BLAST	Basic Local Alignment Search Tool
bp	Base pair
BSA	Bovine Serum Albumin
cm	centimetre
Da	Dalton
DNA	Deoxyribonucleic Acid
DTT	Dithiothreitol
E	East
g	gram
G	Gamma
h	hour
IPG	Immobilized ph Gradient
ITS	Internal Transcribe Region
L	litre
LT <sub>50</sub>	median lethal time
m	metre
M	molarity
mg	milligram
mg/ml	milligram per millilitre
min	minute
ml	millilitre
ml <sup>-1</sup>	permillilitre
mm	millimetre
MS	Mass Spectra
MS/MS	Tandem Mass Spectrometry
N	North
ng/ml	nanogram per millimetre
OD	Absorbance
PCR	Polymerase Chain Reaction
pH	Potential of hydrogen
pI	Isoelectric point
ppm	Part per million
RH	Relative humidity
RNA	Ribonucleic acid
RPM	Rotation per minute
sec	second
SEM	Scanning electron microscope
TEM	Transmission electron microscope
US\$	United States Dollar
UV	Ultra violet
V	Volt
Vh	Volt x hour
2-D	2-dimension
%	Percentage
°C	Degree Celcius
µl	microlitre
µm	micrometre

## CHAPTER 1

### INTRODUCTION

#### 1.1 *Coptotermes curvignathus* as pest

*Coptotermes curvignathus* is a species of termites commonly found in infested tree orchards in Malaysia. It is a severe pest of tree plantations such as rubber (Sajap et al., 2009) and oil palm (Bakti, 2004). This termite is widespread in subtropical and tropical countries such as Malaysia and other South East Asia countries (Tho, 1992; Klangkaew et al., 2002) as well as countries like Brazil (Costa-Leonardo et al. 2008) and the United States of America (Lenz et al., 2012). The colony performance of this termite is based on the various classes of caste in the colony, like the typical social insects (Deligne et al., 1981), makes it a serious pest of timber-based products and tree plantations.

Chemical-based pesticide generally used in controlling termite population is considered as an inefficient practice that could cause several implications toward the environment and pest resistances (Hokkanen and Lynch, 1995). As alternative application of a biological control such as *Metarhizium anisopliae* would be a potential approach to overcome these problems. *Metarhizium anisopliae* is an entomopathogenic fungus that does not give a negative effect on other organisms and the environment. Furthermore, it would give a long-lasting effect on the pests (Howarth, 1991). When compared to other entomopathogenic fungi such as *Beauveria bassiana*, *M. anisopliae* appeared to sporulated faster than *B. bassiana* (Sun et al., 2002). This criterion is important because it is the first in fungal pathogenicity to infect the host by accumulating its conidia on the host's body.

#### 1.2 Study of *Metarhizium anisopliae* infection on *Coptotermes curvignathus*

The infection of *M. anisopliae* starts with the penetration of the germ tube through the insect's cuticle after germination and through several tissues before the hyphae colonizes the hemocoel and turns the termite into the moribund state (Sajap and Kaur, 1991). The first level of the insect immune system, which is the skin is made up of cuticle. This is the main weakness of the termites when they encounter entomopathogenic fungi.

Several hydrolytic enzymes such as chitinase and protease have been described as proteins that regulate the pathogenicity of *M. anisopliae* (St. Leger et al., 1992). The regulation of a series of chitinase genes such as *chi*, *chi1*, *chi2* and *chi3* shows the different patterns of expression in infected *Spodoptera litura* and *Helicoverpa armigera*. Different structure and composition of the insect's skin may cause different expression of these hydrolytic enzymes when under attack (Prakash et al., 2012). Therefore, hydrolytic enzymes and other pathogen-related protein expressions may be different in termites when attacked by *M. anisopliae*.

The immunity of individual termite plays a major role to encounter pathogen attack since the possibility of subterranean termites to get infected is high because they forage

underground (Chouvenc et al., 2009). Grooming behavior between termites as a social insect when the pathogen infects them is not enough to repel the pathogen due to the slow cannibalism activity by termites in eliminating the infected member in the colony (Yanagawa and Shimizu, 2007).

### 1.3 Problem statement

Previous studies have shown that the application of *M. anisopliae* is commonly successful in the laboratory but not in the field. This problem arises when because there is lack of understanding in the interaction between the termites and *M. anisopliae* during the pathogenesis process (Chouvenc et al., 2011).

The interaction between *C. curvignathus* and several other *Coptotermes* spp. termites with *M. anisopliae* have been studied before (Hoe et al., 2009; Maketon et al., 2007). However, knowledge on the mechanisms of pathogenesis at different stages of infection at cellular and molecular level are still lacking.

### 1.4 Objectives of study

This study was to screen for highly pathogenic isolates from ten isolates of *M. anisopliae* for use in pathogenicity study on infected subterranean termites, *C. curvignathus*. The early and late stages of infection were observed using scanning electron microscopy (SEM) technique. Furthermore, the late stage of pathogenesis was observed by using transmission electron microscopy (TEM). The proteins involved during pathogenesis were separated using 2-dimensional gel electrophoresis. Protein gel images were analyzed and spots of the protein of interest were excised for protein expression profile and determination of protein of interest involved respectively, using software and mass spectrometry. Specific objectives were:

- a) To identify highly virulent *M. anisopliae* isolates against *C. curvignathus*
- b) To monitor the pathogenesis process in *C. curvignathus* using scanning electron microscopy (SEM) and transmission electron microscopy (TEM)
- c) To profile and identify proteins expressed during the interaction between *C. curvignathus* and *M. anisopliae* over a time course using a 2-dimensional gel electrophoresis and mass spectrometry, respectively.

## REFERENCES

- Abdul Hafiz, A.M., Abu Hassan Ahmad, M.Z.A, Abdul Rashid. (2006). Field Efficacy of Premise 200SC Against *Coptotermes gestroi* (Isoptera: Rhinotermitidae) and *Globitermes sulphureus* (Isoptera: Termitidae) in Seberang Perai and Balik Pulau, Penang, Malaysia. Paper presented at The 11th Biological Sciences Graduate Congress, Bangkok, Thailand. December 15-17, 2006.
- Ahmad, M. (1958). Key to the Indo-Malayan termites, *Biologia* 4: 33-198.
- Ahmad, M. (1965). Termites (Isoptera) of Thailand. *Bulletin of the American Museum of Natural History* 131: 1-113.
- Altson, R.A. (1947). A fungus parasitic on *Coptotermes curvignathus*, *Holmgr. Nature* 160: 120.
- Alves, S.B. (1998). Fungos entomopatogênicos, In S.B. Alves (Ed.), *Controle microbiano dos insetos* (2<sup>nd</sup> ed) (pp. 289-381). Piracicaba: FEALQ.
- Bagga, S, Hu, G., Screen, S.E. and St. Leger, R.J. (2004). Reconstructing the diversification of subtilisins in the pathogenic fungus *Metarhizium anisopliae*. *Gene* 324: 159–169.
- Bakti, D. (2004). Strategy of integrated control of termites on oilpalm plantation. *Jurnal Penelitian Bidang Ilmu Pertanian* 2(2): 8-14.
- Barros, B.H.R., Silva, S.H., Marques, E.R. and Rosa, J.C. (2010). A proteomic approach to identifying proteins differentially expressed in conidia and mycelium of the entomopathogenic fungus *Metarhizium acridum*. *Fungal Biology* 14: 572–579.
- Bateman, R.P., Carey, M., Batt, D., Prior, C., Abraham, Y., Moore, D., Jenkins, N. and Fenlon J. (1996). "Screening for virulent isolates of entomopathogenic fungi against the desert locust, *Schistocerca gregaria* (Forskål)". *Biocontrol Science and Technology* 6(4): 549–560.
- Beccaloni, G. and Eggleton, P. (2013). In Z.Q. Zhang (Ed.), *Animal Biodiversity: An Outline of Higher-level Classification and Survey of Taxonomic Richness (Addenda 2013)*. *Zootaxa*, 3703 (pp. 46-48). Auckland: Magnolia Press.
- Berbee, M.L., Yoshimura, A., Sugiyama, J., Taylor, J.W. (1995). Is *Penicillium* monophyletic? An evaluation of phylogeny in the family Trichocomaceae from 18S, 5.8S and ITS ribosomal DNA sequence data. *Mycologia* 87: 210–222.
- Bignell, D.E. (2005). Termites as Soil Engineers and Soil Processors. In E. König and A. Varma (Eds.), *Intestinal Microorganisms of Termites and Other Invertebrates* (pp. 183-185). Heidelberg: Springer.
- Bignell, D.E. and Eggleton, P. (2000). Termites in ecosystems. In T. Abe, D.E. Bignell, M. Higashi (Eds.), *Termites: Evolution, Sociality, Symbioses, Ecology* (pp. 363–387). Massachusetts: Kluwer Academic Publishers.

- Bischoff, J.F., Rehner, S.A. and Humber, R.A. (2009). A multilocus phylogeny of the *Metarhizium anisopliae* lineage. *Mycologia* 101(4), 512-530.
- Borges, J.C. and Ramos, C.H. (2005). Protein folding assisted by chaperones. *Protein and Peptide Letters* 12 (3): 257–261.
- CABI. (n.d.). *Coptotermes curvignathus* (rubber termite). Retrieved March 04, 2016, from <http://www.cabi.org/isc/datasheet/15282>
- Carbone, I. and Kohn, L.M. (1993). Ribosomal DNA sequence divergence within internal transcribed spacer 1 of the Sclerotiniaceae. *Mycologia* 85: 415–427.
- Chan, S.P., Bong, C.F.J. and Lau, W.H. (2011). Damage Pattern and Nesting Characteristic of *Coptotermes curvignathus* (Isoptera: Rhinotermitidae) in Oil Palm on Peat. *American Journal of Applied Sciences* 8(5): 420-427.
- Chey, V.K. (1996). Forest Pest Insects in Sabah. Sabah Forest Records No. 15. Sandakan, Malaysia: Sabah Forest Department
- Choi, Y.W., Hyde, K.D and Ho, W.W.H. (1999). Single spore isolation of fungi. *Fungal Diversity* 3: 29-38.
- Chouvenc, T., Su, N.Y. and Elliot, M.I. (2008). Antifungal activity of the termite alkaloid norharmane against the mycelial growth of *Metarhizium anisopliae* and *Aspergillus nomius*.
- Chouvenc, T., Su, N.Y. and Robert, A. (2009). Cellular encapsulation in the eastern subterranean termite, *Reticulitermes flavipes* (Isoptera), against infection by the entomopathogenic fungus *Metarhizium anisopliae*. *Journal of Invertebrate Pathology* 101: 234-241.
- Chouvenc, T. and Su, N.Y. (2010). Apparent Synergy Among Defense Mechanisms in Subterranean Termites (Rhinotermitidae) Against Epizootic Events: Limits and Potential for Biological Control. *Journal of Economic Entomology* 103(4): 1327-1337.
- Chouvenc, T., Su, N.Y. and Robert, A. (2011). Differences in Cellular Encapsulation of Six Termite (Isoptera) Species Against Infection by the Entomopathogenic Fungus *Metarhizium anisopliae*. *Florida Entomologist* 94(3): 389-397.
- Chouvenc, T., Su, N.Y. and Grace, J.K. (2011). Fifty years of attempted of biological control of termites – analysis of a failure. *Biological Control* 59: 69-82.
- Chouvenc, T. and Su, N.Y. (2012). When subterranean termites challenge the rules of fungal epizootics. *PLoS ONE*, 7: e34484.
- Chouvenc, T., Efstathion, C.A., Elliott, M.L. and Su N.Y. (2013). Extended disease resistance emerging from the faecal nest of a subterranean termite. *Proc R Soc B*, 280: 20131885.



- Colin, M. E., Bonmatin, J. M., Moineau, I., Gaimon, C., Brun, S. and Vermandere, J. P. (2004). A method to quantify and analyze the foraging activity of honey bees: Relevance to the sublethal effects induced by systemic insecticides; *Archives of Environmental Contamination and Toxicology* 47 (3): 387–395.
- Cooper, B., Garret, W.M. and Campbell, K.B. (2006). Shotgun identification of proteins from uredospores of the bean rust *Uromyces appendiculatus*. *Proteomics* 6: 2477-2484.
- Costa-Leonardo, A.M., Casarin, F.E. and Constantini, J.P. (2008). Record of a gregarine (Apicomplexa: Neogregarinida) in the abdomen of the termite *Coptotermes gestroi* (Isoptera, Rhinotermitidae). *Journal of Invertebrate Pathology* 97: 114–118.
- Cranshaw, W. and Redak, R. (2013). Roach Cities and Assassin Cousins. In: W. Cranshaw and R. Redak, (Eds.), *Bugs Rule!: An Introduction to the World of Insects* (pp. 188). Princeton University Press.
- Cruz-Vazquez, C., Carvajal Márquez, J., Lezama-Gutiérrez, R., Vitela-Mendoza, I., Ramos-Parra, M. (2015). Efficacy of the entomopathogenic fungi *Metarhizium anisopliae* in the control of infestation by stable flies *Stomoxys calcitrans* (L.), under natural infestation conditions. *Veterinary Parasitology* 212: 350–355.
- de Castilhos-Fortes, R., Matsumura, A.T.S., Diehl, E. and Fiuza, L.M. (2002). Susceptibility of *Nasutitermes ehrhardti* (Isoptera: Termitidae) to *Bacillus thuringiensis* subspecies. *Brazilian Journal of Microbiology* 33: 219-222.
- Deligne, J., Quennedey, A. and Blum M. S. (1981). The enemies and defense mechanisms of termites, in *Social Insects*, Vol. 2, ed Hermann H. R., editor. (Waltham, MA: Academic Press), 1–76
- Desneux, J. (1904) A propos de la phylogénie des Termitides. *Annales de la Société Entomologique de Belgique* 48: 278–286.
- Drubin, D.G. and Nelson, W.J. (1996). Origins of cell polarity. *Cell* 84: 335–344.
- Duong, N.H., Thanh, H.N., Doan, T., Yen, N.T., Tam, T.T.M., Dung, P.T. and Phuong L.T.T. (1998). Diseases and pests of *Hevea brasiliensis* in Vietnam. Symposium on natural rubber (*Hevea brasiliensis*): Vol. 2 - physiology and exploitation and crop protection and planting methods sessions, Ho Chi Minh City, China, 14-15 October 1997., 80-91.
- Edward, R. and Mill, A.E. (1986). *Termites in Buildings. Their Biology and Control*. East Grinstead: Rentokil Limited.
- Eggleton, P. (2000). Global patterns of termite diversity. In *Termites: Evolution, Sociality, Symbioses, Ecology* (Ed.), T. Abe, D.E. Bignell and M. Higashi, (pp. 25–51). Dordrecht, Netherlands: Kluwer Academic Publishers.

- Engel, M.S., Grimaldi, D.A. and Krishna, K. (2009). Termites (Isoptera): Their phylogeny, classification, and rise to ecological dominance. *American Museum Novitates* 3650: 1-27.
- Evans, J.D., Aronstein, K, Chen, Y.P., Hetru, C, Imler, J.L., Jiang, H., Kanost, M., Thompson, G.J., Zou, Z. and Hultmark, D. (2006). Immune pathways and defense mechanisms in honey bees *Apis mellifera*. *Insect Molecular Biology* 15: 645-656.
- Froggatt, W. (1897). Australian Termitidae part II. *Proceedings of the Linnean Society of New South Wales* 21: 510–552.
- Furukawa, S., Tanaka, H., Nakazawa, H., Ishibashi, J. and Shono, T. (1999). Inducible gene expression of moricin, a unique antibacterial peptide from the silkworm (*Bombyx mori*). *Biochemistry Journal* 340(1): 265–271.
- Gao, Q., Jin, K., Ying, S.H., Zhang, Y., Xiao, G., Shang, Y., Duan, Z., Hu, X., Xie, X.Q., Zhou, G., Peng, G., Luo, Z., Huang, W., Wang, B., Fang, W., Wang, S., Zhong, Y., Ma, L.J. Wang, C. (2011). Genome sequencing and comparative transcriptomics of the model entomopathogenic fungi *Metarhizium anisopliae* and *Metarhizium acridum*. *PLoS Genetics* 7(1), e1001264.
- Gay, F.G. (1963). The synonymy, distribution, and biology of *Coptotermes elisiae* (Desneux). *Pacific Insects* 5(2): 421–423.
- Gasteiger E., Hoogland, C., Gattiker, A., Duvaud, S., Wilkins, M.R., Appel, R.D. and Bairoch, A. (2005). *Protein Identification and Analysis Tools on the ExPASy Server*. In J.M. Walker (Ed.), *The Proteomics Protocols Handbook*. Humana Press.
- Gillooly, J.F., Hou, C. and Kaspari, M. (2010). Eusocial insects as superorganisms. *Communicative and Integrative Biology* 3(4): 360-362.
- Gindin, G., Levski, S., Glazer, I. and Soroker, V. (2006). Evaluation of the Entomopathogenic Fungi *Metarhizium anisopliae* and *Beauveria bassiana* against the Red Palm Weevil *Rhynchophorus ferrugineus*. *Entomolgy* 34: 370-379.
- Giuliano, G.D.B., Krasnoff, S.B., Moon, Y.S., Churchill, A.C.L. and Gibson, D. (2011). Genetic basis of destruxin production in *Metarhizium robertsii* ARSEF 2575. Genetic basis of destruxin production in the entomopathogen *Metarhizium robertsii*. *Current Genetics* 58(2): 105-16.
- Grace, J.K., (2003). Approaches to biological control of termites. *Sociobiology* 41: 115–121.
- Gray, B. (1968). Forest tree and timber insect pests in the territory of Papua and New Guinea. *Pacific Insects* 10(2): 301-323.
- Grindstaff, K.K., Yeaman, C., Anandasabapathy, N., Hsu, S.C., Rodriguez-Boulan, E., Scheller, R.H. and Nelson, W.J. (1998). Sec6/8 Complex Is Recruited to Cell–

Cell Contacts and Specifies Transport Vesicle Delivery to the Basal-Lateral Membrane in Epithelial Cells. *Cell* 93: 731–740.

- Hall, R.A. (1984). Epizootic potential for aphids of different isolates of the fungus *Verticillium lecanii*. *Entomophaga* 29: 311-321.
- Hall, R.A. (1998). Rho GTPases and the actin cytoskeleton. *Science* 279: 509–514.
- Hamid, A.A. (1982). Insect pests of plantation species in Sarawak. In: *Proceedings of the Eighth Malaysian Forestry Conference*, Sabah, 2-8 August 1982. Volume II, 791-796.
- Hänel, H. (1982). Selection of a fungus species, suitable for biological control of the termite *Nasutitermes exitiosus* (Hill). *Zeitschrift für Angewandte Entomologie* 94: 237-245.
- Hänel, H. and Watson, J.A.L. (1983). Preliminary field tests on the use of *Metarhizium anisopliae* for the control of *Nasutitermes exitiosus* (Hill) (Isoptera: Termitidae). *Bull. Journal of the Entomological Research Society* 73: 305-313.
- Hayashi, Y., Lo, N., Miyata, H., Kitade, O. (2007). Sex-linked genetic influence on caste determination in a termite. *Science* 318: 985–987.
- Hernandez, C.E.M., Guerrero, I.E.P., Hernandez, G.A.G., Solis, E.S., Guzman, J.C.T. (2010). Catalase overexpression reduces the germination time and increases the pathogenicity of the fungus *Metarhizium anisopliae*. *Applied Microbiology and Biotechnology* 87:1033 – 1045.
- Herrera, F., Carballo, M. and Shannon, P., (1999). Eficacia de cepas nativas de hongos entomopatogénicos sobre *Bemisia tabaci*, en el laboratorio. *Manejo Integrado de Plagas* 54: 37–43.
- Hoe, P.K., Bong, C.F.J., Jugah, K. and Rajan, A. (2009). Evaluation of *Metarhizium anisopliae* var. *anisopliae* (Deuteromycotina: Hyphomycete) isolates and their effects on subterranean termite *Coptotermes curvignathus* (Isoptera: Rhinotermitidae). *American Journal of Agricultural and Biological Sciences* 4: 289-297.
- Hokkanen, H. and Lynch, J.M. (1995). *Biological Control: Benefits and Risks*. Cambridge, United Kingdom: Cambridge University Press.
- Holmgren, N. (1910). Das System der Termiten. *Zoologischer Anzeiger* 35: 284–286.
- Holmgren, N. (1913). Termitenstudien. 4. Versuch einer systematischen Monographie der Termiten der orientalischen Region. *Kunliga Svenska vetenskapsakademiens handlingar* 50(2): 1-276.
- Holmgren, K. and Holmgren, N. (1917). Report on a collection of termites from India. *Memoirs of the Department of Agriculture in India* 5: 137-171.
- Howarth, F.G. (1991). Environmental impacts of classical biological control. *Annual Review Entomology* 36: 485-509.

- Hsu, S.C., Ting, A.E., Hazuka, C.D., Davanger, S., Kenny, J.W., Kee, Y. and Scheller, R.H. (1996). The mammalian brain rsec6/8 complex. *Neuron* 17: 1209–1219.
- Hu, X.P. 2010. Chapter 12: Biology and Reproductive Strategies in the Subterranean Termites (Isoptera: Rhinotermitidae). In: Liu, T.X. and Kang, L. (Eds): Recent advances in entomological research: from molecular biology to pest management. High Educational Press, Springer, pp 213 - 226.
- Hussain, A., Li, Y.F., Cheng, Y., Liu, Y., Chen, C.C. and Wen, S.Y. (2013). Immune-related transcriptome of *Coptotermes formosanus* Shiraki workers: the defense mechanism. *PLoS ONE* 8(7): e69543.
- Iga, M. & Kataoka, H. (2012). Recent studies on insect hormone metabolic pathways mediated by cytochrome P450 enzymes. *Biol Pharm Bull* 35: 838–843.
- Inglis, D.G., Goettel, M.S., Butt, T.M. and Strasser, H. (2001). Use of Hyphomycetous fungi for managing pests. In T.M. Butt, C. Jackson and I. Magen (Eds.), Fungi as biocontrol agents: progress, problems and potential (pp. 23–39). London, UK: CABI.
- Inoue, M., Chang, L., Hwang, J., Chiang, S.H. and Saltiel, A.R. (2003). The exocyst complex is required for targeting of Glut4 to the plasma membrane by insulin. *Nature*, 422: 629-633.
- Inward, D., Beccaloni, G. and Eggleton, P. (2007). Death of an order: a comprehensive molecular phylogenetic study confirms that termites are eusocial cockroaches. *Biology Letters* 3: 331-335.
- Ishikawa, H. and Marshall, W.F. (2011). Ciliogenesis: building the cell's antenna. *Nature Reviews Molecular Cell Biology* 12: 222-234.
- James, P. (1997). Protein identification in the post-genome era: the rapid rise of proteomics. *Annual Review of biophysics* 30(4): 279-331.
- Javar, S., Mohamed, R., Sajap, A.S. and Lau, W.H. (2015). Expression of pathogenesis-related genes in *Metarhizium anisopliae* when infecting *Spodoptera exigua*. *Biological Control* 85: 30-36.
- Jenkins, T.M., Jones, S.C., Lee, C.Y., Forchler, B.T., Chen, Z., Lopez-Martinez, G., Gallager, N.T., Brown, G., Neal, M., Thirstleton, B. and Kleinschmidt, S. (2007). Phylogeography illuminates maternal origins of exotic *Coptotermes gestroi* (Isoptera: Rhinotermitidae). *Molecular Phylogenetics and Evolution* 42: 612-621.
- Johnson, R.E., Klassen, R., Prakash, L., Prakash, S. (2015). A Major Role of DNA Polymerase  $\delta$  in Replication of Both the Leading Lagging DNA Strands. *Mol. Cell* 59 (2): 163–175.

- Jones, W.E., Grace, J.K. and Tamashiro, M. (1996). Virulence of seven isolates of *Beauveria bassiana* and *Metarhizium anisopliae* to *Coptotermes formosanus* (Isoptera: Rhinotermitidae). *Environmental Entomology* 25: 481-487.
- Kalshoven, L.G.E (1963). *Coptotermes curvignathus* causing the death of trees in Indonesia and Malaya. *Entomologische Berichten* 23: 90-100.
- Kambhampati, S. and Eggleton, P. (2000). Taxonomy and phylogeny of termites. In T. Abe, D.E. Bignell and M. Higsahi (Eds.) *Termites: Evolution, Sociality, Symbioses, and Ecology* (pp. 1-23). Dordrecht, Netherlands: Kluwer Academic Publishers.
- Kaplan, E.L. and Meier, P. (1958). Nonparametric estimation from incomplete observations. *Journal of the American Statistical Association* 53: 457– 481.
- Karkhanis, V., Wang, L., Tae, S., Hu, Y.J., Imbalzano, A.N., Sif, S. (2012). Protein arginine methyltransferase 7 regulates cellular response to DNA damage by methylating promoter histones H2A and H4 of the polymerase  $\delta$  catalytic subunit gene, POLD1. *The Journal of Biological Chemistry* 287 (35): 29801–29814.
- Keller, S., Kessler, P. and Schweizer, C. (2003). Distribution of insect pathogenic soil fungi in Switzerland with special reference to *Beauveria brongniartii* and *Metharhizium anisopliae*. *Bio Control* 48: 307–319.
- Kershaw, M.J., Moorhouse, E.R., Bateman, R.P., Reynolds, S.E. and Charnley, A.K. (1999). The role of destruxins in the pathogenicity of *Metarhizium anisopliae* for three species of insects. *Journal of Invertebrate pathology* 74: 213-223.
- Khoo, K.C., Ooi, P.A.C. and Ho, C.T. (1991). *Crop pests and their management in Malaysia*. Kuala Lumpur, Malaysia: Tropical Press.
- Kim, J.H., Min, J.S., Kang, J.S., Kwon, D.H., Yoon, K.S., Strycharz, J., Koh, YH., Pittendrigh, B.R., Clark, J.M. and Lee, S.H. (2011). Comparison of the humoral and cellular immune responses between body and head lice following bacterial challenge. *Insect Biochemistry and Molecular Biology* 41(5): 332–339.
- Kirton, L.G., Brown, V.K. and Azmi, M. (1999). The pest status of the termite *Coptotermes curvignathus* in *Acacia mangium* plantations: incidence, mode of attack and inherent predisposing factors. *Journal of Tropical Forest Science* 11(4): 822-831.
- Kirton, L.G. and Brown, V.K. (2003). The taxonomic status of pest species of *Coptotermes* in Southeast Asia: resolving the paradox in the pest status of the termites, *Coptotermes gestroi*, *C. havilandi* and *C. travians* (Isoptera: Rhinotermitidae). *Sociobiology* 42: 43–63.
- Kirton, L.G. And Cheng, S. (2007). Ring-Barking And Root Debarking Of Dipterocarp Saplings By Termites In An Enrichment Planting Site In Malaysia *Journal of Tropical Forest Science* 19(2): 67–72.

- Klangkaew, C., Inoue, T., Abe, T., Takematsu, Y., Kudo, T., Noparatnaraporn, N. and Kirtibutr, N. (2002). The diversity and abundance of termites (Isoptera) in the urban area of Bangkok, Thailand. *Sociobiology* 39: 485–493.
- Krutmuang, P. and Mekchay, S. (2005). Pathogenicity of entomopathogenic fungi *Metarhizium anisopliae* against termites. Paper presented at the *Tropentag 2005: Conference on International Agricultural Research for Development*, Stuttgart, Hohenheim.
- Kolykhalov, A.A., Mihalik, K., Feinstone, S.M., Rice, C.M. (2000). Hepatitis C virus-encoded enzymatic activities and conserved RNA elements in the 3' nontranslated region are essential for virus replication in vivo. *J Virol* 74: 2046-2051.
- Korb, J. (2007). Termites. *Current Biology* 17: 995-999.
- Kumar, P.A., Sharma, P.P. and Malik, V.S. (1996). The insecticidal proteins of *Bacillus thuringiensis*. *Advance in Applied Micobiology* 42: 1-43.
- Kusters, J.G., Van Vliet, A.H., Kuipers, E.J. (2006). Pathogenesis of *Helicobacter pylori* infection. *Clin Microbiol Rev* 19: 449-490.
- Latreille, P.A. (1802). *Histoire Naturelle, Général et Particulière des Crustacés et des Insectes*, Vol. 3, Dufart, Paris.
- Lee, C.Y., Vongkaluang, C., Lenz, M. (2007). Challenges to subterranean termite management of multi-genera faunas in Southeast Asia and Australia. *Sociobiology* 50: 213-221.
- Lee, C.Y. (2007). *Perspective in urban insect pest management in Malaysia* (pp. 104). Vector Control Research Unit, Universiti Sains Malaysia.
- Lee, S., Shrestha, S., Prasad, S.V., Kim, Y. (2011). Role of a small G protein Ras in cellular immune response of the beet armyworm, *Spodoptera exigua*. *J Insect Physiol* 57: 356–362.
- Lenz, M., Creffield, J.W., Evans, T.A., Kard, B., Vongkaluang, C., Sornnuwat, Y., Lee, C.Y., Yoshimura, T. and Tsunoda, K. (2012). Resistance of polyamide and polyethylene cable sheathings to termites in Australia, Thailand, USA, Malaysia and Japan: A comparison of four field assessment methods. *International Biodeterioration & Biodegradation* 66: 53-62.
- Liao, X., Lu, H.L., Fang, W., Raymond, J. and St. Leger, R.J. (2014). Overexpression of a *Metarhizium robertsii* HSP25 gene increases thermotolerance and survival in soil. *Applied Microbiology and Biotechnology* 98:777–783.
- Ligoxygakis, P., Pelte, N., Hoffmann, J. and Reichhart, J. (2002). Activation of *Drosophila toll* during fungal infection by a blood serine protease. *Science* 297: 114–116.

- Lim, K.H. and Silek, B. (2001). Termite infestation on oil palms planted on deep peat in Sarawak: Tradewinds experience. Cutting-edge technologies for sustained competitiveness: Proceedings of the 2001 PIPOC International Palm Oil Congress, Agriculture Conference, Kuala Lumpur, Malaysia, 20-22 August 2001, 355-368.
- Linder, P., Lasko, P.F., Ashburner, M., Leroy, P., Nielsen, P.J., Nishi, K., Schnier, J., Slonimski, P.P. (1989). Birth of the D-E-A-D box. *Nature* 337(6203): 121-122.
- Losick, R., Watson, J.D., Baker, T.A., Bell, S., Gann, A. and Levine, M.W. (2008). *Molecular biology of the gene (6th ed.)*. San Francisco: Pearson/Benjamin Cummings.
- Lovett, B. and St. Leger, R.J. (2015). Stress is the rule rather than the exception for *Metarhizium*. *Current Genetics* 61: 253-261.
- Lu, H.L., Wang, J.B., Brown, M.A., Euerle, C. and St Leger, R.J. (2015). Identification of *Drosophila* Mutants Affecting Defense to an Entomopathogenic Fungus. *Scientific Reports* 5: 12350.
- Maketon, M., Sawangwan, P. and Sawatwarakul, W. (2007). Laboratory Study on the Efficacy of *Metarhizium anisopliae* (Deuteromycota: Hyphomycetes) in Controlling *Coptotermes gestroi* (Isoptera: Rhinotermitidae). *Entomol Gener* 30: 203–218.
- Makimura, K., Tamura, Y., Mochizuki, T., Hasegawa, A., Tajiri, Y., Hanazawa, R., Uchida, K., Saito, H. and Yamaguchi, H. (1999). Phylogenetic Classification and Species Identification of Dermatophyte Strains Based on DNA Sequences of Nuclear Ribosomal Internal Transcribed Spacer 1 Regions. *Journal of Clinical Microbiology* 37(64): 920-924.
- Merzendorfer, H., Kim, H.S., Chaudhari, S.S., Kumari, M., Specht, C.A., Butcher, S., Brown, S.J., Robert Manak, J., Beeman, R.W., Kramer, K.J. and Muthukrishnan, S. (2012). Genomic and proteomic studies on the effects of the insect growth regulator diflubenzuron in the model beetle species *Tribolium castaneum*. *Insect Biochemistry and Molecular Biology* 42: 264–276.
- Miller, C.D., Rangel, D., Braga, G.U.L., Flint, S., Kwon, S.I., Messias, C.L., Roberts, D.W. and Anderson, A.J. (2004). Enzyme activities associated with oxidative stress in *Metarhizium anisopliae* during germination, mycelial growth, and conidiation and in response to near-UV irradiation. *Canadian Journal of Microbiology* 50: 41-49.
- Milner, R.J., Staples, J.A. and Lutton, G.G. (1998). The selection of an isolate of the hyphomycete fungus, *Metarhizium anisopliae* for control of termites in Australia. *Biological Control* 11: 240-247.
- Morikawa, T., Yasuno, R. and Wada, H. (2001). Do mammalian cells synthesize lipoic acid? Identification of a mouse cDNA encoding a lipoic acid synthase located in the mitochondria. *FEBS Letters* 498: 16–21.

- Murad, A.M., Noronha, E.F., Miller, R.N.G., Costa, F.T., Pereira, C.D., Mehta, A., Caldas, R.A. and Franco, O.L. (2008). Proteomic analysis of *Metarhizium anisopliae* secretion in the presence of the insect pest *Callosobruchus maculatus*. *Microbiology* 154: 3766–3774.
- Myles, T.G. (2002). Alarm, aggregation, and defense by *Reticulitermes flavipes* in response to a naturally occurring isolate of *Metarhizium anisopliae*. *Sociobiology* 40: 243–255.
- Netz, D.J., Pierik A.J., Stümpfig, M., Mühlenhoff, U. and Lill, R. (2007). The Cfd1-Nbp35 complex acts as a scaffold for iron-sulfur protein assembly in the yeast cytosol. *Nat Chem Biol* 3(5): 278-286.
- Neves, P.M.O.J. and Alves, S.B. (2004). External Events Related to the Infection Process of *Cornitermes cumulans* (Kollar) (Isoptera: Termitidae) by the Entomopathogenic Fungi *Beauveria bassiana* and *Metarhizium anisopliae*. *Neotropical entomology* 33(1): 51-56.
- Ntwasa, M., Goto, A., Kurata, S. (2012). Coleopteran antimicrobial peptides: prospects for clinical applications. *International Journal of Microbiology* 2012: 101989.
- Nyam, V.T. Bong, C.F.J. and King, J.H.P. (2015). Control of subterranean termite *Coptotermes curvignathus* (Isoptera: Rhinotermitidae) by entomopathogen *Metarhizium anisopliae* var. *anisopliae* cultured in liquid state fermentation. *American Journal of Agricultural and Biological Sciences* 10(1): 35-40.
- Osbrink, W.L.A., Williams, K.S., Connick, W.J., Wright, M.S. and Lax, A.R. (2001). Virulence of Bacteria Associated with the Formosan Subterranean Termite (Isoptera: Rhinotermitidae) in New Orleans, LA. *Environmental Entomology* 30(2): 443-448.
- Packer, L., Witt, E.H., Tritschler, H.J. (1995). alpha-Lipoic acid as a biological antioxidant. *Free Radical Biology Medicine* 19(2): 227-250.
- Panepinto, J., Liu, L., Ramos, J., Zhu, X., Valyi-Nagy, T. and Eksi, S. (2005). The DEAD-box RNA helicase Vad1 regulates multiple virulence-associated genes in *Cryptococcus neoformans*. *Journal of Clinical Investigation* 115: 632–641.
- Pedrini, N., Juarez, M.P., Crespo, R., de Alaniz, M.J. (2006). Clues on the role of *Beauveria bassiana* catalases in alkane degradation events. *Mycologia* 98:528-534.
- Petlamul, W. and Prasertsan, P. (2012). Evaluation of Strains of *Metarhizium anisopliae* and *Beauveria bassiana* against *Spodoptera litura* on the Basis of Their Virulence, Germination Rate, Conidia Production, Radial Growth and Enzyme Activity. *Mycobiology* 40: 111-116.
- Prakash, G.V.S.B., Padmaja, V., Jami, S.K. and Kirti, P.B. (2012). Expression of chitinase genes of *Metarhizium anisopliae* isolates in lepidopteran pests and on synthetic media. *Journal of Basic Microbiology* 52: 1-8.



- Quesada-Moraga, E, Santos-Quiróz, R., Valverde-García, P. and Santiago-Álvarez, C. (2004) Virulence, horizontal transmission, and sublethal reproductive effects of *Metarhizium anisopliae* (Anamorphic fungi) on the German cockroach (Blattodea: Blattellidae). *Journal of Invertebrate Pathology* 87: 51-58.
- Rao, B.S. (1974). Diseases and pests of rubber in South and South East Asia. *World Crops* 26(2): 75-81.
- Reddy, G.V.P., Zhao, Z. and Humber, R.A. (2014). Laboratory and field efficacy of entomopathogenic fungi for the management of the sweetpotato weevil, *Cylas formicarius* (Coleoptera: Brentidae). *Journal of Invertebrate Pathology* 122: 10-15.
- Riefler, G.M., Balasingam, G., Lucas, K.G., Wang, S., Hsu, S.C., Firestein, B.L. (2003). Exocyst complex subunit sec8 binds to postsynaptic density protein-95 (PSD-95): a novel interaction regulated by cypin (cytosolic PSD-95 interactor). *Biochemical Journal* 373(1): 49-55.
- Roberts, D.W., and St. Leger, R.J. (2004). *Metarhizium* spp., cosmopolitan insect-pathogenic fungi: mycological aspects. *Advance of Applied Microbiology* 54: 1-70.
- Roh, J.Y., Choi, J.Y. Li, M.S. Jin, B.R. and Je, Y.H. (2007). *Bacillus thuringiensis* as a specific, safe, and effective tool for insect pest control. *Journal of Microbiology and Biotechnology* 17 (4): 547-559.
- Rojas, A.M., Fuentes, G., Rausell, A., Valencia, A. (2012). The Ras protein superfamily: Evolutionary tree and role of conserved amino acids. *Journal of Cel Biology* 196(2): 189-201.
- Roonwal, M.L. and Chhotani O.B. (1962). Indian Species of Termite Genus *Coptotermes*. Indian Council for Agricultural Research Entomological Monograph No. 2.
- Rosengaus, R.B., Jordan, C., Lefebvre, M.L. and Traniello, J.F.A. (1999). Pathogen Alarm Behavior in a Termite: A New Form of Communication in Social Insects. *Naturwissenschaften* 86(11): 544-548.
- Rosengaus, R.B., Moustakes, J.E., Calleri, D.V. and Traniello, J.F.A. (2003). Nesting ecology and cuticular microbial loads in dampwood (*Zootermopsis angusticollis*) and drywood termites (*Incisitermes minor*, *I. schwarzi*, *Cryptotermes cavifrons*). *Journal of Insect Science* 3: 31.
- Rustiguel, C.B., Rosa, J.C., Jorge, J.A., de Oliveira, A.H., Guimarães, L.H. (2015). Secretome Analysis of *Metarhizium anisopliae* Under Submerged Conditions Using *Bombyx mori* Chrysalis to Induce Expression of Virulence-Related Proteins. *Current Microbiology* 72(2): 220-227.
- Sajap, A.S., and Kaur, K. (1990). Histopathology of *Metarhizium anisopliae*, an Entomopathogenic Fungus, Infection in the Termite, *Coptotermes curvignathus*. *Pertanika* 13: 331- 334.

- Sajap, A.S. and Yaacob, A.W. (1997). Termites from selected building premises in Selangor, Peninsular Malaysia. *Malaysian Forester* 93: 203-215.
- Sajap, A.S., Samsudin Amit, Welker, J. (2000). Evaluation of hexaflumuron for controlling the subterranean termite *Coptotermes curvignathus* (Isoptera: Rhinotermitidae) in Malaysia. *Journal of Economic Entomology* 93(2):429-433.
- Sajap, A.S., Sahri, M.H. and Mohd Kasim, M.R. (2009). Feeding response of *Coptotermes curvignathus* and *Coptotermes gestroi* (Isoptera: Rhinotermitidae) to different wood of *Hevea* species. *Sociobiology* 54(3): 873-880.
- Saksamrit, J., Montreesuksirikun, C., Issaranon, T., Atsawat, S., Jitjak, K., Potikhun, P. and Piyaboon, O. (2008). The Efficiency of The Supernatant From *Metarhizium anisopliae* for Eradicating The Termites *Coptotermes curvignathus*. *KMITL Sci. J* 8: 80-85.
- Salman-Dilgimen, A., Hardy, P.O., Dresser, A.R., Chaconas, G. (2011). HrpA, a DEAH-box RNA helicase, is involved in global gene regulation in the Lyme disease spirochete. *PLoS One* 6: e22168.
- Samsuddin, A.S., Sajap, A.S. and Mohamed, R. (2015). *Metarhizium anisopliae* of Peninsular Malaysia origin poses high pathogenicity toward *Coptotermes curvignathus*, a major wood and tree pest. *Malaysian Forester* 78 (1 & 2): 41-48.
- Santi, L., Walter, O.B., Silva, Antonio, F.M., Pinto, Schrank, A., Marilene, H., Vainstein, (2010). *Metarhizium anisopliae* host-pathogen interaction: differential immunoproteomics reveals proteins involved in the infection process of arthropods. *Fungal biology* 114: 312-319.
- Santoro, MG. (2000). Heat shock factors and the control of the stress response. *Biochemical Pharmacology* 59(1): 55-63.
- Schabel H.G. (1978). Percutaneous infection of *Hylobius pales* by *Metarhizium anisopliae*. *Journal of Invertebrate Pathology* 31: 180-187.
- Scheffrahn, H. (2008). Termites (Isoptera). In J.L Capinera (Ed.), *Encyclopedia of Entomology* (pp 3737-3747). Netherlands: Springer.
- Schonauer, M.S., Kastaniotis, A.J., Kursu, V.A., Hiltunen, J.K. and Dieckmann, C.L. (2009). Lipoic Acid Synthesis and Attachment in Yeast Mitochondria. *The Journal of Biological Chemistry* Vol. 284, NO.35, pp.23234-23242.
- Scott, J.G. (1999). Cytochromes P450 and insecticide resistance. *Insect Biochemistry and Molecular Biology* 29: 757-777.
- Sepuri, Naresh Babu, V., Schülke, N., Pain, D. (1998). "GTP Hydrolysis Is Essential for Protein Import into the Mitochondrial Matrix". *Journal of Biological Chemistry* (273): 1420-1424.
- Snyder TE, 1949. Catalog of the Termites (Isoptera) of the World. Smithsonian Miscellaneous Collections No. 112, 490 pp.

- Sornnuwat, Y., Tsunoda, K., Yoshimura, T., Takahashi, M. and Vonckaluanc, C. (1996). Foraging populations of *Coptotermes gestroi* (Isoptera: Rhinotermitidae) in an urban area. *Journal of Economic Entomology* 89: 1485–1490.
- St. Leger, R.J., Butt, T.M., Goettel, M.S., Staples, R.C. and Roberts, D.W. (1989). Production in vitro of appressoria by the entomopathogenic fungus *Metarhizium anisopliae*. *Experimental Mycology* 13: 274-288.
- St. Leger, R.J., Butt, T.M., Stapples, R.C. and Roberts, D.W. (1990). Second messenger involvement in differentiation of the entomopathogenic fungus *Metarhizium anisopliae*. *Journal of General Microbiology* 136: 1779-1789.
- St. Leger, R.J., Frank, D.C., Roberts, D.W. and Staples, R.C. (1992). Molecular cloning and regulatory analysis of the cuticle-degrading protease structural gene from the entomopathogenic fungus *Metarhizium anisopliae*. *European Journal of Biochemistry* 204: 991-1001.
- Stanchi, F., Bertocco, E., Toppo, S., Dioguardi, R., Simionati, B., Cannata, N., Zimbello, R., Lanfranchi, G., Valle, G. (2001). Characterization of 16 novel human genes showing high similarity to yeast sequences. *Yeast* 18(1):69-80.
- Steinhaus, E. A. (1949). *Principles of insect pathology* (757 pp.). New York and London: Hafner Publishing Company.
- Stevenson, B.J., Bibby, J., Pignatelli, P., Muangnoicharoen, S., O'Neill, P.M., Lian, L.Y., Müller, P., Nikou, D., Steven, A., Hemingway, J., Sutcliffe, M.J. and Paine, M.J. (2011). Cytochrome P450 6M2 from the malaria vector *Anopheles gambiae* metabolizes pyrethroids: Sequential metabolism of deltamethrin revealed. *Insect Biochem Mol Biol* 41(7): 492–502.
- Su, N.Y. and Scheffan, R.H. (2000). Termites as pests of buildings. Dordrecht, Netherland: *Kluwer Academic Publishers* 437-453.
- Su, N.Y., Guo, Q., Tu, J, Li, X., Meng, L., Cao, L., Dong, D., Qiu, J. and Guan, X. (2013). Proteins differentially expressed in conidia and mycelium of the entomopathogenic fungus *Metarhizium anisopliae* sensu stricto. *Canadian Journal of Microbiology* 59: 443- 448.
- Sun, J., Fuxa, J.R. and Henderson, G. (2002). Sporulation of *Metarhizium anisopliae* and *Beauveria bassiana* on *Coptotermes formosanus* and in vitro. *Journal of Invertebrate Pathology* 81: 78-85.
- Sun, J., Fuxa, J.R. and Henderson, G. (2003). Effects of virulence, sporulation, and temperature on *Metarhizium anisopliae* and *Beauveria bassiana* laboratory transmission in *Coptotermes formosanus*. *Journal of Invertebrate Pathology* 84: 38-46.
- Tamashiro, M., Fuji, J.K. and Lai, P.Y. (1973). A simple method to observe, trap and prepare large number of subterranean termites for laboratory and field experiments. *Environ. Entomol* 2: 721-722.

- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M. and Kumar, S. (2011). MEGA5: Molecular Evolutionary Genetics Analysis using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. *Molecular Biology and Evolution*.
- TerBush, D.R.1. and Novick, P. (1995). Sec6, Sec8, and Sec15 are components of a multisubunit complex which localizes to small bud tips in *Saccharomyces cerevisiae*. *Journal of Cell Biology* 130(2):299-312.
- Thapa, R.S. and Shim, P.S. (1971). Termite damage in plantation hoop line, *Araucaria cunninghamii* D. Don, in Sabah and its control. *Malaysian Forester* 34: 47-32.
- Tho, Y.P. (1974). The termite problem in plantation forestry in Peninsular Malaysia. *Malaysian Forester* 37: 278-283.
- Tho, Y.P. (1992). Termites of peninsular Malaysia. *Malayan Forest Records* 36: 1-224.
- Tho, Y.P. and Kirton, L.G. (1998). A survey of termite attack in Bahau conifer plantation, Peninsular Malaysia. *Journal of Tropical Forest Science* 10(4): 564-567.
- Timothy, J.K. and Nemat O.K. (2008). Intracellular infection of tick cell lines by the entomopathogenic fungus *Metarhizium anisopliae*. *Microbiology* 154: 1700-1709.
- Toledo, A.V., de Remes Lenicov, A.M.M. and López Lastra, C.C. (2010). Histopathology caused by the entomopathogenic fungi, *Beauveria bassiana* and *Metarhizium anisopliae*, in the adult planthopper, *Peregrinus maidis*, a maize virus vector. *Journal of Insect Science* 10: 35.
- Tsurimoto, T. and Stillman, B. (1991). Replication factors required for SV40 DNA replication in vitro. II. Switching of DNA polymerase alpha and delta during initiation of leading and lagging strand synthesis. *J. Biol. Chem* 266: 1961-1968.
- Walter, S. and Buchner, J. (2002). Molecular chaperones-cellular machines for protein folding. *Angewandte Chemie* 41(7): 1098-1113.
- Walsh, N.P., Alba, B.M., Bose, B., Gross, C.A., Sauer, R.T. (2003). OMP peptide signals initiate the envelope-stress response by activating DegS protease via relief of inhibition mediated by its PDZ domain. *Cell* 113 (1): 61-71.
- Wang, C. and St. Leger, R.J. (2007). The MAD1 adhesin of *Metarhizium anisopliae* links adhesion with blastospore production and virulence to insects: the MAD2 adhesin enables attachment to plants. *Eukaryotic Cell* 6: 808-816.
- Wang, Y., Sumathipala, N., Rayaprolu, S. and Jiang, H. (2011). Recognition of microbial molecular patterns and stimulation of prophenoloxidase activation by a b-1,3-glucanase-related protein in *Manduca sexta* larval plasma. *Insect Biochemistry and Molecular Biology* 41(5): 322-331.

- Wassarman, D.A. and Steitz, J.A. (1991). RNA splicing. Alive with DEAD proteins. *Nature* 349(6309): 463–464.
- Waterhouse DF, 1993. *The Major Arthropod Pests and Weeds of Agriculture in Southeast Asia. ACIAR Monograph No. 21* (141 pp.). Canberra, Australia: Australian Centre for International Agricultural Research.
- Wedde, M., Müller, D., Tintelnot, K., De Hoog, G.S. and Stahl, U. (1998). Phylogenetic Classification and Species Identification of Dermatophyte Strains Based on DNA Sequences of Nuclear Ribosomal Internal Transcribed Spacer 1 Regions. *Medical Mycology* 36: 61-67.
- White, T. J., T. Bruns, S. Lee, and J. W. Taylor. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: M.A. Innis, D. H. Gelfand, J. J. Sninsky, and T. J. White (Eds.), *PCR Protocols: A Guide to Methods and Applications* (pp. 315-322), New York: Academic Press Inc.
- Wilkins, M.R., Pasquali, C., Appel, R.D., Ou, K., Golaz, O., Sanchez, J.C., Yan, J.X., Gooley, A.A., Hughes, G., Humphery-Smith, I., Williams, K.L. and Hochstrasser, D.F. (1996). From proteins to proteomes: large scale protein identification by two-dimensional electrophoresis and amino acid analysis. *Biotechnology (N Y)* 14(1): 61-65.
- Wood, T.G. (1978). In M.V. Brian (ed.) *Production ecology of ants and termites* (pp.55-80). London: Cambridge University Press.
- Wright, M.S. and Cornelius, M.L. (2012). Mortality and repellent effects of microbial pathogens on *Coptotermes formosanus* (Isoptera: Rhinotermitidae). *BMC Microbiology* 12: 291.
- Yanagawa, A. and Shimizu, S. (2007). Resistance of the termite, *Coptotermes formosanus* Shiraki to *Metarhizium anisopliae* due to grooming. *BioControl* 52: 75-85.
- Yanagawa, A., Fujiwara-Tsujii, N., Akino, T., Yoshimura, T., Yanagawa, T. And Shimizu, S. (2011). Behavioral changes in the termite, *Coptotermes formosanus* (Isoptera), inoculated with six fungal isolates. *Journal of Invertebrate Pathology* 107: 100-106.
- Yii, J.E., Bong, C.F.J., King, J.H.P. and Kadir, J. (2016). Synergism of entomopathogenic fungus, *Metarhizium anisopliae* incorporated with fipronil against oil palm pest subterranean termite, *Coptotermes curvignathus*. *Plant Protection Science* 52(1): 35-44.
- Yu, L., Tang, W., He, W., Ma, X., Vasseur, L., Simon, W., Baxter, Yang, G., Huang, S., Song, F. and You, M., (2015). Characterization and expression of the cytochrome P450 gene family in diamondback moth, *Plutella xylostella* (L.). *Scientific Reports* 5: 8952.
- Zettervall, C.J., Anderl, I., Williams, M.J., Palmer, R., Kurucz, E., et al. (2004). A directed screen for genes involved in *Drosophila* blood cell activation. *Proc Natl Acad Sci USA* 101: 14192–14197.

Zhang, W., Chen, J., Keyhani, N.O., Zhang, Z., Li, S., Xia, Y. (2015). Comparative transcriptomic analysis of immune responses of the migratory locust, *Locusta migratoria*, to challenge by the fungal insect pathogen, *Metarhizium acridum*. *BMC Genomics* 16(1):867.

Zimmerman, G. (1998). Suggestions for a standardized method for re-isolation of entomopathogenic fungi from soil using the bait method. *Journal of Applied Entomology* 102: 213-215.

