



UNIVERSITI PUTRA MALAYSIA

***MECHANISM OF
EFFECTS OF ONE AMINO ACID SUBSTITUTIONS AT THE C-TERMINAL
REGION OF THERMOSTABLE L2 LIPASE FROM *Bacillus* sp. L2***

HARTINI AHMAD SANI

FBSB 2017 23



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By

HARTINI AHMAD SANI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

June 2017

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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June 2017

Chairman : Fairolniza Mohd Shariff, PhD
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The substitutions of the amino acid residue at the predetermined critical point of the C-terminal of L2 lipase may increase its thermostability and lipase activity. N and C-terminal regions in most proteins are often disordered and flexible. However, some protein function was closely related to flexibility as well as play a role in the enzyme reaction. The critical point of the stability of L2 lipase structure was predicted at position 385 (wild type residue Serine) of the L2 sequence based on I-Mutant2.0 software. The effects of substitution of the amino acids at the critical point with Glutamic acid, Isoleucine, and Valine were analyzed with Molecular Dynamics (MD) simulation by using Yet Another Scientific Artificial Reality Application (YASARA) software and it showed that the best predicted mutant L2 lipases had lower RMSD value as compared to L2 lipase. It indicated that the three mutants had higher compactness in the structure consequently enhancing the stability. From RMSF analysis, mutations had reduced the flexibility of the enzyme. The best predicted mutants (S385E, S385I, and S385V) were produced in the experimental lab by site-directed mutagenesis. The mutant L2 lipases (60.4 kDa) were purified to homogeneity by a single chromatography step before proceeding with characterization. There were high lipase activities produced by purified mutant L2 lipases at a temperature range of 60-85 °C with the optimum temperature of 80 °C, 75 °C and 70 °C for S385E, S385V, and S385I lipases respectively. The optimum temperature for recombinant L2 lipase was at 70 °C. Mutant L2 lipases (S385E and S385V) had higher optimum temperature compared to recombinant L2 lipase. The optimum pH for mutant L2 lipases (S385E and S385V) was found to be at pH 8 and for S385I was at pH 9, whereas the optimum pH for recombinant L2 lipase was at pH 9. S385I lipase was more thermostable as compared to recombinant L2 lipase and other mutants at temperature 60 °C within 16 hours preincubation. The stability of S385V lipase in various organic solvents was higher as compared to recombinant L2 lipase. S385V lipase had relative activities higher than 100% which 111% in DMSO, 105% in acetone, 123% in diethyl ether and 124% in n-hexane. T_m values for S385V and S385E lipases were at 85.96 °C and 84.85 °C and the values were higher as compared to recombinant L2 lipase which is only 66.73 °C. This

showed the higher thermal stability of S385E and S38V lipases as compared to recombinant L2 lipase. Thus, the substitutions at the predetermined critical point of the C-terminal (Ser385) changed the functionality of the protein structure towards the activity, stability, and flexibility of L2 lipase. The critical point mutation towards the structure of L2 lipase provided a very advantageous strategy for the improvement of enzyme with better function to adapt with harsh environment.

Keywords: L2 lipase, thermostability, site-directed mutagenesis, Molecular Dynamics (MD) simulation.



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**KESAN PENGGANTIAN SATU ASID AMINO DI KAWASAN C-TERMINAL
TERMOSTABIL L2 LIPASE DARI BAKTERIA *Bacillus L2***

Oleh

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Penggantian daripada sisa asid amino pada titik kritikal yang telah ditetapkan pada C-terminal lipase L2 boleh meningkatkan termostabiliti dan aktiviti lipase. Kawasan N dan C-terminal di kebanyakan protein adalah sering tidak tersusun dan fleksibel. Walau bagaimanapun, sesetengah fungsi protein berkaitan rapat dengan fleksibiliti serta memainkan peranan dalam tindak balas enzim. Titik kritikal untuk kestabilan struktur lipase L2 telah dijangkakan pada kedudukan 385 (sisa jenis liar Serine) jujukan L2 yang berdasarkan perisian I-Mutant2.0. Kesan penggantian asid amino di titik kritikal dengan Asid glutamik, Isoleucine dan Valine dianalisis dengan Molekul Dinamik (MD) simulasi dengan menggunakan perisian 'Yet Another Scientific Artificial Reality Application' (YASARA) dan ia telah menunjukkan bahawa mutan lipase L2 yang terbaik diramalkan mempunyai nilai RMSD yang lebih rendah berbanding lipase L2. Ia menunjukkan bahawa tiga mutan tersebut mempunyai kepadatan yang lebih tinggi di dalam struktur yang seterusnya meningkatkan kestabilan. Daripada RMSF analisis, mutasi telah mengurangkan fleksibiliti enzim tersebut. Mutan yang terbaik diramalkan (S385E, S385I dan S385V) telah dihasilkan di makmal eksperimen oleh mutagenesis mengarah lokasi. Mutan lipase L2 tersebut (60.4 kDa) telah dituliskan menjadi kehomogenan oleh satu langkah kromatografi sebelum meneruskan dengan pencirian. Terdapat aktiviti-aktiviti lipase yang tinggi dihasilkan oleh mutan lipase L2 yang dituliskan pada julat suhu 60-85 °C dengan suhu optimum masing-masing adalah 80 °C, 75 °C dan 70 °C untuk lipase S385E, S385V dan S385I. Suhu yang optimum untuk lipase L2 rekombinan adalah pada 70 °C. Lipase L2 mutan (S385E dan S385V) mempunyai suhu optimum yang lebih tinggi berbanding dengan lipase L2 rekombinan. PH yang optimum bagi lipase L2 mutan (S385E dan S385V) didapati pada pH 8 dan untuk S385I adalah pada pH 9, manakala pH yang optimum bagi lipase L2 rekombinan adalah pada pH 9. Lipase S385I adalah lebih termostabil berbanding dengan lipase L2 rekombinan dan mutan yang lain pada suhu 60 °C dalam tempoh 16 jam pra-inkubasi. Kestabilan S385V lipase dalam organik pelarut yang berbeza-beza adalah lebih tinggi berbanding lipase L2 rekombinan. Lipase S385V mempunyai aktiviti-aktiviti relatif yang lebih tinggi daripada 100% dimana 111% dalam

DMSO, 105% dalam aseton, 123% dalam dietil eter dan 124% pada n-heksana. Nilai-nilai T_m untuk lipase S385V dan S385E berada pada 85.96 °C dan 84.85 °C dan nilainya adalah lebih tinggi berbanding lipase L2 rekombinan yang hanya 66.73 °C. Ini menunjukkan kestabilan haba lebih tinggi untuk lipase S385E dan S385V berbanding lipase L2 rekombinan. Oleh yang demikian, penggantian pada titik kritikal yang telah ditetapkan pada C-terminal (Ser385) telah menukar fungsi struktur protein terhadap aktiviti, kestabilan, dan fleksibiliti lipase L2. Mutasi titik kritikal terhadap struktur lipase L2 menyediakan satu strategi yang sangat berfaedah untuk peningkatan enzim dengan fungsi yang lebih baik untuk menyesuaikan diri dengan persekitaran yang sukar.

Kata kunci: lipase L2, termostabiliti, mutagenesis mengarah lokasi, simulasi molekul dinamik (MD).

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I certify that a Thesis Examination Committee has met on 13 June 2017 to conduct the final examination of Hartini binti Ahmad Sani on her thesis entitled "Effects of One Amino Acid Substitutions at the C-Terminal Region of Thermostable L2 Lipase from *Bacillus* sp. L2" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

α	Alpha
Å	Angstrom
β	Beta
β -ME	β -Mercaptoethanol
°C	Degree celsius
%	Percentage
μ g	Microgram
μ L	Microlitre
μ m	Micrometre
μ M	Micromolar
Abs	Absorbance
APS	Ammonium persulfate
bp	Base pair
CaCl ₂	Calcium chloride
CD	Circular Dichroism
CV	Column volume
ddH ₂ O	Double distilled water
DNA	Deoxyribonucleic acid
dNTP	Deoxynucleotide
DDG	Free Energy change
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
dsDNA	double stranded DNA
<i>E. coli</i>	<i>Escherichia coli</i>
ESBRI	Evaluating the Salt BRIdges
g	Gram
GAPDH	Glyceraldehyde-3-phosphate dehydrogenase
h	Hour
HCl	Hydrochloric acid
IPTG	Isopropyl β -D-1-thiogalactopyranoside
kb	Kilobase
kcal	Kilocalories
kDA	Kilodaltons
L	Litre
LB	Luria-Bertani
M	Molar
mA	Milliamps
mM	Millimolar
MD	Molecular Dynamics
mg	Milligram
mL	Millilitre
N	Molar
ng	Nanogram
nm	Nanometre
ns	Nanoseconds
NaOH	Sodium hydroxide

OD	Optical density
PAGE	Polyacrylamide Gel Electrophoresis
PCR	Polymerase Chain Reaction
ps	Picoseconds
RI	Reliability index
RMSD	Root Mean Square Deviation
RMSF	Root Mean Square Fluctuations
RNA	Ribonucleic acid
RPM	Rate per minute
SASA	Solvent Accessible Surface Area
SDS	Sodium Dodecyl Sulphate
SVM	Support vector machines
TEMED	TetramethylEthylenediamine
T opt	Optimal temperature
U/mL	Units per millilitre
v/v	Volume/volume
xg	Times gravity

CHAPTER 1

INTRODUCTION

Protein function may be controlled by protein structure. Protein is composed of a precise sequence of amino acids that allow it to fold up into a particular three-dimensional shape, or conformation (Alberts *et al.*, 2002). Besides that, protein contains regions that are directly involved in protein function, such as active sites or binding sites, as well as regions that are less critical to the protein function and where mutations are likely to have less effect (Betts and Russell 2003). Protein sequences are classically considered as consisting of the whole information for their three-dimensional (3D) structure (Bornot *et al.*, 2007). Each type of protein has a unique sequence of amino acids and has a particular three-dimensional structure, which governed by the order of the amino acids in its chain (Alberts *et al.*, 2002). Changing of critical amino acid can cause changes in protein conformation. However, some studies have shown that changing of less hydrophobic residue (Arg) to a more hydrophobic residue (Ser) at the position 157 of ARM lipase increased the internal hydrophobicity to maintain the structural stability at a high temperature (Salleh *et al.*, 2012).

It is long known that the N and C-terminal regions in most proteins are often disordered and flexible (Kamarudin *et al.*, 2014). Flexibility may be closely related to protein function, as well as play a role in enzyme catalysis (Karshikoff *et al.*, 2015). C-terminal of a protein is known as the residue that has free carboxyl group or at least does not acylate to another residue of amino acid, means that this residue is the end residue of the protein (Hardy *et al.*, 1985). Gudiukaite *et al.*, (2014) reported that 10 and 20 C-terminal amino acids of GD-95 lipase from *Geobacillus* sp. 95 crucially affect other physicochemical characteristics and the stability of this enzyme.

This research explored the roles of the critical amino acid at the C-terminal towards the structure of L2 lipase. The lipase was isolated from bacteria known as *Bacillus* sp. L2, one of the thermophilic bacteria from a hot spring in Perak. Recombinant L2 lipase was successfully overexpressed with a 178-fold increase in activity compared to crude native L2 lipase. The recombinant L2 lipase (43.2 kDa) was purified to homogeneity in a single chromatography step. The purified lipase was found to be reactive at a temperature range of 55–80 °C and at a pH of 6–10 and the optimum activity was found to be at 70 °C and pH 9. The melting temperature (T_m) of L2 lipase was 59.04 °C when analysed by circular dichroism (CD) spectroscopy studies. (Shariff *et al.*, 2011).

The substitution of the amino acid residues at the predetermined critical point at the C-terminal of the L2 lipase may cause an increase in protein stability and lipase activity or may speed up the unfolding of the protein structure. The prediction of the critical point of the L2 lipase was done by using one of the software from internet known as I-Mutant2.0 where 20 amino acids from the C-terminal were analysed in terms of stability. Then, the effects of substitution of the amino acids at the critical point were proceeded

by Molecular Dynamics (MD) simulation by using other software known as Yet Another Scientific Artificial Reality Application (YASARA) and followed by mutagenesis in the experimental lab. The mutant L2 lipases were analysed in terms of lipase activity and other physiochemical properties before can proceed with investigating the secondary structure of the protein. Therefore, the main objective of this research is to investigate the effects of critical amino acid at the C-terminal towards the activity, stability, and flexibility o L2 lipase.

There are two sub-objectives of the research:

- To predict the critical point and analyze the effects of amino acid substitutions at the C-terminal by *in silico* study.
- To validate the effects caused by the substitution of selected amino acids at the target residue of the C-terminal of L2 lipase experimentally.

REFERENCES

- Ahmad, S., Kamal, M.Z., Sankaranarayanan, R. and Rao, N.M. (2008). Thermostable *Bacillus subtilis* lipases: in vitro evolution and structural insight. *Journal of Molecular Biology* 381: 324-40.
- Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K. and Walter, P. (2002). *Molecular Biology of the Cell, 4th edition*. New York: Garland Science.
- Ali, M.S.M., Fuzi, S.F.M., Ganesan, M., Rahman, R.N.Z.R.A., Basri, M. and Salleh, A.B. (2013). Structural adaptation of cold-active RTX lipase from *Pseudomonas* sp. strain AMS8 revealed via homology and molecular dynamics simulation approaches. *Biomedical Research International* 2013: 1-9.
- Arpigny, J.L. and Jaeger, K.E. (1999). Bacterial lipolytic enzymes: classification and properties. *The Biochemical Journal* 343: 177-183.
- Aurora, R. and Rose, G.D. (1998). Helix capping. *Protein Science* 7: 21-38.
- Bava, K.A., Gromiha, M.M., Uedaira, H., Kitajima, K. and Sarai, A. (2004). Protherm, version 4.0: thermodynamic database for proteins and mutants. *Nucleic Acids Research* 32: D120-D121.
- Betts, M.J. and Russell, R.B. 2003. Amino acid properties and consequences of substitutions. In *Bioinformatics for Geneticists*, ed. M. R. Barnes and I. C. Gray, pp. 290-312. United Kingdom: John Wiley and Sons, Ltd.
- Bornot, A., Offmann, B. and Brevern, A.D. (2007). How flexible are protein structures? New questions on the protein structure plasticity. *BioForum Europe* 11: 25.
- Bornhorst, J.A. and Falke, J.J. (2000). Purification of proteins using polyhistidine affinity tags. *Methods in Enzymology* 326: 245-254.
- Bradford, M.N. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry* 72: 248-254.
- Brown, T.L., Lemay, H.E., Bursten, B.E. and Burdge, J.R. (2003). *Chemistry The Central Science*. New Jersey: Pearson Education.
- Capriotti, E., Fariselli, P. and Casadio, R. (2005). I-Mutant2.0: Predicting changes upon mutation from the protein sequence or structure. *Nucleic Acids Research* 33: W306-W310.
- Chan, C.H., Yu, T.H. and Wong, K.B. (2011). Stabilizing salt-bridge enhances protein thermostability by reducing the heat capacity change of unfolding. *PLoS One* 6: 1-8.
- Cherepanov, A.V. and de Vries, S. (2001). Binding of nucleotides by T4 DNA ligase and T4 RNA ligase: optical absorbance and fluorescence studies. *Biophysical Journal* 81: 3545- 3559.
- Costantini, S., Colonna, G. and Facchiano, A.M. (2008). ESBRI : A web server for evaluating salt bridges in proteins. *Bioinformation* 3:137-138.

- Constantini, S. and Facchiano, A. (2002). ESBRI: A web server for evaluating salt bridges in proteins. Retrieved 10 November 2016 from <http://bioinformatica.isa.cnr.it/ESBRI/>.
- Coolbear, T., Daniel, R.M. and Morgan, H.W. (1992). The enzymes from extreme thermophiles: bacterial sources, thermostabilities and industrial relevance. *Advances in Biochemical Engineering/biotechnology* 45: 57-98.
- Dill, K.A. (1990). Dominant forces in protein folding. *Biochemistry* 29: 7135-7155.
- Doukyu, N. and Ogino, H. (2010). Organic solvent-tolerant enzymes. *Biochemical Engineering Journal* 48: 270-282.
- Fields, P.A. (2001). Review: Protein function at thermal extremes: balancing stability and flexibility. *Comparative Biochemistry and Physiology* 129: 417-431.
- Fujiwara, K., Toda, H. and Ikeguchi, M. (2012). Dependence of α -helical and β -sheet amino acid propensities on the overall protein fold type. *BMC Structural Biology* 12: 18.
- Garcia, A.E. and Onuchic, J.N. (2003). Folding a protein in a computer: an atomic description of the folding/unfolding of protein A. *Proceedings of the National Academy of Sciences of the United States of America* 100: 13898-13903.
- Greenfield, N.J. (2009). Using circular dichroism collected as a function of temperature to determine the thermodynamics of protein unfolding and binding interactions. *Nature protocols* 1: 2527-2535.
- Gromiha, M. M., Pathak, M.C., Saraboji, K., Ortlund, E.A. and Gaucher, E.A. (2012). Hydrophobic environment is a key factor for the stability of thermophilic proteins. *Proteins* 81: 715-721.
- Guerois, R. and Paz, M.L.D. 2006. Design of structural elementary motifs. In *Protein design: methods and application*, ed. Kohler and Valentin, pp. 3-26. Totowa: Humana Press.
- Guerois, R., Nielsen, J.E. and Serrano, L. (2002). Predicting changes in the stability of proteins and protein complexes: a study of more than 1000 mutations. *Journal of Molecular Biology* 320: 369-387.
- Gudiukaite, R., Gegeckas, A., Kazlauskas, D. and Citavicius, D. (2014). Influence of N- and/or C-terminal regions on activity, expression, characteristics and structure of lipase from *Geobacillus* sp. 95. *Extremophiles: Life under Extreme Conditions* 18: 131-145.
- Guncheva, M. and Zhiryakova, D. (2011). Catalytic properties and potential applications of *Bacillus* lipases. *Journal of Molecular Catalysis B: Enzymatic* 68: 1-21.
- Gupta, R., Gupta, N. and Rathi, P. (2004). Bacterial lipases: an overview of production, purification and biochemical properties. *Applied Microbiology and Biotechnology* 64: 763-781.
- Hardy, P.M., Jeffrey, P.D., Mathews, A., Mercer, W.D., Nichol, L.W., Pain, R.H., Scawen, M.D., Sherwood, R.F., Smith, J.C., Smith, R.A.G., Stephens, R.M., Thornton, J.M., Winzor, D.J. 1985. Amino acid residues. In *Amino acids*,

- Peptides and Proteins*, ed. M. Ryadnov, E. Farkas, and F. Hudecz, pp. 395. London: The Royal Society of Chemistry.
- Harwood, J. (1989). The versatility of lipases for industrial uses. *Trends in Biochemical Sciences* 14: 125-126.
- Hayward, J.A., Finney, J.L., Daniel, R.M. and Smith, J.C. (2003). Molecular dynamics decomposition of temperature-dependent elastic neutron scattering by a protein solution. *Biophysical Journal* 85:679-685.
- Hasan, F., Shah, A.A. and Hameed, A. (2006). Industrial applications of microbial lipases. *Enzyme and Microbial Technology* 39: 235-251.
- Instruction manual Catalog #210518 (10 reactions) and #210519 (30 reactions) [Revision F.0]. Retrieved 15 March 2015 from <http://www.agilent.com/cs/library/usermanuals/public/210518.pdf>
- Instruction manual Catalog #200521 (10 reactions) and #200522 (30 reactions) [Revision F.0]. Retrieved 20 April 2015 from <http://www.agilent.com/cs/library/usermanuals/public/200521.pdf>.
- Invernizzi, G., Papaleo, E., Grandori, R., De Gioia, L. and Lotti, M. (2009). Relevance of metal ions for lipase stability: structural rearrangements induced in the *Burkholderia glumae* lipase by calcium depletion. *Journal of Structural Biology* 168: 562-570.
- Jaeger, K.E. and Reetz, M.T. (1998). Microbial lipases form versatile tools for biotechnology. *Trends in Biotechnology* 16: 396-403.
- Jeffrey, G.A. (1998). An introduction to hydrogen bonding. *Journal of the American Chemical Society* 120: 5604.
- Jeong, S.T., Kim, H.K., Jun, K.S., Chi, S.W., Pan, J.G., Oh, T.K. and Ryu, E.S. (2002). Novel Zinc-binding center and a temperature switch in the *Bacillus stearothermophilus* L1 lipase. *Journal of Biological Chemistry* 277:17041-17047.
- Kamal, M.Z., Ahmad, S., Yedavalli, P. and Rao, N.M. (2010). Stability curves of laboratory evolved thermostable mutants of a *Bacillus subtilis* lipase. *Biochimica et Biophysica Acta (BBA)-Proteins and Proteomics* 1804: 1850-1856.
- Kamarudin, N.H.A., Rahman, R.N.Z.R.A., Ali, M.S.M., Leow, T.C., Basri, M. and Salleh, A.B. (2014). Unscrambling the effect of C-terminal tail deletion on the stability of a cold-adapted organic solvent stable lipase from *Staphylococcus epidermidis* AT2. *Molecular Biotechnology* 56: 747-757.
- Karshikoff, A. and Jelesarov, I. (2008). Salt bridges and conformational flexibility: effect on protein stability. *Biotechnology and Biotechnological Equipment* 22: 606-611.
- Karshikoff, A., Nilsson, L. and Ladenstein, R. (2015). Rigidity versus flexibility: the dilemma of understanding protein thermal stability. *The FEBS Journal* 282: 3899-3917.

- Kessel, A. and Ben-Tal, N. (2011). Protein Structure. In *Introduction to proteins: Structure, Function and Motion*, ed. N.F. Briton, X. Lin, H.M. Safer, M.V. Schneider, M. Singh, and A. Tramontano, pp. 67-73. United Kingdom: CRC Press.
- Kokkinidis, B.M., Glykos, N.M. and Fadouloglou, V.E. (2012). Protein flexibility and enzymatic catalysis. *Advances in protein chemistry and structural biology* 87: 181-218.
- Krieger, E., Vriend, G. and Spronk, C. (2003). YASARA-Yet Another Scientific Artificial Reality Application. Retrieved 13 Oktober 2014 from <http://www.yasara.org/about.htm>.
- Kufareva, I. and Abagyan, R. (2012). Methods of protein structure comparison. *Methods in Molecular Biology* 857: 231-257.
- Kumar, A., Dhar, K., Kanwar, S. S. and Arora, P. K. (2016). Lipase catalysis in organic solvents : advantages and applications. *Biological Procedures Online* 18: 1-11.
- Kumar, S. and Nussinov, R. (2001). How do thermophilic proteins deal with heat? *Cellular and Molecular Life Sciences* 58: 1216-1233.
- Kumar, S. and Nussinov, R. (1999). Salt Bridge Stability in Monomeric Proteins. *Journal of Molecular Biology* 293: 1241-1255.
- Kumar, S., Tsai, C.J. and Nussinov, R. (2000). Factors enhancing protein thermostability. *Protein Engineering* 13: 179-191.
- Kumwenda, B., Litthauer, D., Bishop, O. T. and Reva, O. (2013). Analysis of protein thermostability enhancing factors in industrially important thermus bacteria species. *Evolutionary Bioinformatics Online* 9: 327-342.
- Kundu, S., Melton, J.S., Sorensen, D.C. and Phillips, G.N. (2002). Dynamics of proteins in crystals: Comparison of experiment with simple models. *Biophysical Journal* 83: 723-732.
- Kunkel, T.A. (1985). Rapid and efficient site-specific mutagenesis without phenotypic selection. *Proceedings of the National Academy of Sciences of the United States of America* 82: 488-492.
- Kwon, D.K. and Rhee, J.S. (1986). A simple and rapid colorimetric method for determination of free fatty acids for lipase assay. *Journal of the American Oil Chemist' Society* 63: 89-92.
- Ladenstein, R. and Ren, B. (2006). Protein disulfides and protein disulfide oxidoreductases in hyperthermophiles. *FEBS Journal* 273:4170-4185.
- Langel, U., Cravatt, B.F., Graslund, A., von Heijne, G., Zorko, M., Land, T. and Niessen, S. (2010). *Introduction to Peptides and Proteins*. London: CRC Press.
- Li, H. and Zhang, X. (2005). Characterization of thermostable lipase from thermophilic *Geobacillus* sp. TW1. *Protein Expression and Purification* 42: 153-159.
- Littlechild, J., Novak, H., James, P. and Sayer, C. 2013. Mechanisms of Thermal Stability Adopted by Thermophilic Proteins and Their Use in White Biotechnology. In *Thermophilic Microbes in Enviromental and Industrial*

- Biotechnology*, ed. T. Satyanarayana, J. Littlechild, Y. Kawarabayasi, pp. 481-507. Dordrecht: Springer Netherlands.
- McCammon, J.A., Gelin, B.R. and Karplus, M. (1977). Dynamics of folded protein. *Nature* 267: 585-590.
- Mendonca, L.M.F. and Marana, S.R. (2011). Single mutations outside the active site affect the substrate specificity in a β -glycosidase. *Biochimica et Biophysica Acta (BBA)-Proteins and Proteomics* 1814: 1616-1623.
- Nagao, T., Shimada, Y., Sugihara, A. and Tominaga, Y. (2000). Amino acid residues contributing to stabilization of *Fusarium heterosporum* lipase. *Journal of Bioscience and Bioengineering* 89: 446-450.
- Nelson, M. and McClelland, M. (1992). Use of DNA methyltransferase/endonuclease enzyme combinations for megabase mapping of chromosomes. *Methods in Enzymology* 216: 279-303.
- Nosoh, Y. and Sekiguchi, T. (1991). *Protein stability and stabilization through protein engineering*. Ellis Horwood Limited.
- Ollis, D.L., Cheah, E., Cygler, M., Dijkstra, B., Frolow, F. and Franken, S.M., Harel, M., Remington, S.J., Silman, I. and Schrag, J. (1992). The α/β hydrolase fold. *Protein Engineering* 5: 197-211.
- Pace, C.N. (1992). Contribution of the hydrophobic effect to globular protein stability. *Journal of Molecular Biology* 226: 29-35.
- Pace, C.N., Scholtz, J.M. and Grimsley, G.R. (2014). Forces stabilizing proteins. *FEBS Letters* 588: 2177-2184.
- Rabbani, M., Sadeghi, H.M., Ani, M. and Chegini, K.G., Etemadifar, Z. and Moazen, F. (2009). Cloning and nucleotide sequence of a lipase gene from a soil isolate. *Research in Pharmaceutical Sciences* 4: 25-32.
- Rahman, R.N.Z.R., Shariff, F.M., Basri, M. and Salleh, A.B. (2012). 3D structure elucidation of thermostable L2 lipase from thermophilic *Bacillus* sp. L2. *International Journal of Molecular Sciences* 13: 9207-9217.
- Ramachandran, G.N., Ramakrishnan, C. and Sasisekharan, V. (1963). Stereochemistry of polypeptide chain configurations. *Journal of Molecular Biology* 7: 95-99.
- Rapaport, D.C. (2004). Basic Molecular Dynamics. In *The Art of Molecular Dynamics Simulation*, ed. N.J. Pienta, R.S. Cole, J.M. Risley, G.T. Rushton, J.L. Stewart, D.M. Cullen and A. Halpen, pp. 11-16. United Kingdom: Cambridge University Press.
- Rathi, P.C., Fulton, A., Jaeger, K.E. and Gohlke, H. (2016). Application of rigidity theory to the thermostabilization of lipase A from *Bacillus subtilis*. *PLOS Computational Biology* 12:e1004754.
- Rees, D.C. and Robertson, A.D. (2001). Some thermodynamics implications for the stability of proteins. *Protein Science* 10: 1187-1194.

- Reetz, M.T., Carballeira, J.D. and Vogel, A. (2006). Iterative saturation mutagenesis on the basis of B factors as a strategy for increasing protein thermostability. *Communication* 45: 7745-7751.
- Salleh, A.B., Rahim, A.S.M.A., Rahman, R.N.Z.R.A., Leow, T.C. and Basri, M. (2012). The role of Arg157Ser in improving the compactness and stability of ARM lipase. *Computer Science and System Biology* 5: 39-46.
- Sangeetha, R., Arulpandi, I. and Geetha, A. (2011). Bacterial lipases as potential industrial biocatalysts: an overview. *Research Journal of Microbiology* 6: 1-24.
- Sharma, P.K., Singh, K., Singh, R., Capalash, N., Ali, A., Mohammad, O. and Kaur, J. (2012). Characterization of a thermostable lipase showing loss of secondary structure at ambient temperature. *Molecular Biology Reports* 39: 2795-2804.
- Shirley, B.A., Stanssens, P., Hahn, U. and Pace, C.N. (1992). Contribution of hydrogen bonding to the conformational stability of Ribonuclease T1, *Biochemistry* 31: 725-732.
- Shariff, F.M., Rahman, R.N.Z.R.A., Basri, M. and Salleh, A.B. (2011). A newly isolated thermostable lipase from *Bacillus* sp.. *International Journal of Molecular Sciences* 12: 2917-2934.
- Shen, Y. and Bax, A. (2012). Identification of helix capping and β -turn motifs from NMR chemical shifts. *Journal of Biomolecular NMR* 52: 211-232.
- Singh, R., Gupta, N., Goswami, V.K. and Gupta, R. (2006). A simple activity staining protocol for lipases and esterases. *Applied Microbiology and Biotechnology* 70: 679-682.
- Speranza, P. and Macedo, G.A. (2013). Biochemical characterization of highly organic solvent-tolerant cutinase from *Fusarium oxysporum*. *Biocatalysis and Agricultural Biotechnology* 2: 372-376.
- Spiwok, V., Lipovova, P., Skalova, T., Duskova, J., Dohnalek, J., Hasek, J. and Kralova, B. (2007). Cold-active enzymes studied by comparative molecular dynamics simulation. *Journal of Molecular Modeling* 13: 485-97.
- Tanner, J.J., Hecht, R.M. and Krause, K.L. (1996). Determinants of enzyme thermostability observed in the molecular structure of *Thermus aquaticus* D-glyceraldehyde-3-phosphate dehydrogenase at 25 Angstroms resolution. *Biochemical Journal* 35: 2597-2609.
- Tompa, D.R., Gromiha, M.M. and Saraboji, K. (2016). Contribution of main chain and side chain atoms and their locations to the stability of thermophilic proteins. *Journal of Molecular Graphics and Modelling* 64: 85-93.
- Vieille, C. and Zeikus, G.J. (2001). Hyperthermophilic enzymes : sources, uses, and molecular mechanisms for thermostability. *Microbiology and molecular biology reviews* 65: 1-43.
- Vihinen, M. (1987). Relationship of protein flexibility to thermostability. *Protein Engineering* 1: 477-480.

- Villeneuve, P., Muderhwa, J.M., Graille, J. and Haas, M.J. (2000). Customizing lipases for biocatalysis: a survey of chemical, physical and molecular biological approaches. *Journal of molecular Catalysis B. Enzymatic* 9:113-148.
- Vogt, G., Woell, S. and Argos, P. (1997). Protein thermal stability, hydrogen bonds, and ion pairs. *Journal of molecular biology* 269:631-643.
- Wang, S., Meng, X., Zhou, H., Liu, Y., Secundo, F. and Liu, Y. (2016). Enzyme stability and activity in non-aqueous reaction systems: a mini review. *Catalysts* 6: 32.
- Xu, X., Xiao, J., Huang, H., Li, J. and Xiao, H. (2010). Molecular dynamics simulations on the structures and properties of epsilon-CL-20(0 0 1)/F 2314 PBX. *Hazardous Material* 175: 423-428.
- Yu, H. and Huang, H. (2014). Engineering proteins for thermostability through rigidifying flexible sites. *Biotechnology Advances* 32: 308-315.
- Zhang, H.M., Li, J.F., Wu, M.C., Shi, H.L. and Tang, C.D. (2013). Determination of amino acids and dipeptides is correlated significantly with optimum temperatures of microbial lipases. *Annals of Microbiology* 63: 307-313.