- 1 **Full title**: Assessment of interactions between 205 breast cancer susceptibility loci and 13
- 2 established risk factors in relation to breast cancer risk in the Breast Cancer Association
- 3 Consortium
- 4 Short title: Gene-environment interactions and breast cancer risk

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1 Abstract

Background: Previous gene-environment interaction studies of breast cancer risk have provided
sparse evidence of interactions. Using the largest available dataset to date, we performed a
comprehensive assessment of potential effect modification of 205 common susceptibility
variants by 13 established breast cancer risk factors including replication of previously reported
interactions.

Methods: Analyses were performed using 28,176 cases and 32,209 controls genotyped with
iCOGS array and 44,109 cases and 48,145 controls genotyped using OncoArray from the Breast
Cancer Association Consortium (BCAC). Gene-environment interactions were assessed using
unconditional logistic regression and likelihood ratio tests for breast cancer risk overall and by
estrogen-receptor(ER) status. Bayesian False Discovery Probability was used to assess the
noteworthiness of the meta-analyzed array-specific interactions.

13 **Results**: Noteworthy evidence of interaction at $\leq 1\%$ prior probability was observed for three SNP-risk factor pairs. SNP rs4442975 was associated with a greater reduced risk of ER-positive 14 breast cancer ($OR_{int} = 0.85 (0.78 - 0.93), p_{int} = 2.8 \times 10^{-4}$) and overall breast cancer ($OR_{int} = 0.85$) 15 (0.78 - 0.92), $p_{int} = 7.4 \times 10^{-5}$) in current users of estrogen-progesterone therapy compared to 16 non-users. This finding was supported by replication using OncoArray data of the previously 17 reported interaction between rs13387042 ($r^2 = 0.93$ with rs4442975) and current estrogen-18 progesterone therapy for overall disease ($p_{int} = 0.004$). The two other interactions suggested 19 20 stronger associations between SNP rs6596100 and ER-negative breast cancer with increasing parity and younger age at first birth. 21

Conclusion: Overall, our study does not suggest strong effect modification of common breast

2 cancer susceptibility variants by established risk factors.

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4	Key messages
5	• The association between common breast cancer susceptibility loci and breast cancer
6	risk is not strongly modified by established breast cancer risk factors.
	• The combined effect of susceptibility loci and established risk factors is thus well
7	described by a multiplicative model.
8	• We found one noteworthy G x E interaction with overall and ER-positive breast
9	cancer risk, which was replicated, and two novel noteworthy G x E interactions with
5	ER-negative breast cancer risk.
10	• In an independent dataset, we replicated two previously reported G x E interactions.
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1 Introduction

Breast cancer is a complex disease with both environmental and genetic factors contributing to 2 3 risk. Well-established modifiable and non-modifiable environmental factors include age at menarche, parity, age at first birth, breastfeeding, body mass index (BMI), use of menopausal 4 hormonal therapy (MHT), and alcohol consumption (1-6). In addition, high to moderate-risk 5 gene mutations such as BRCA1, BRCA2, TP53, ATM, and CHEK2 increase the risk of breast 6 7 cancer (7-14), as well as multiple common, low-risk single nucleotide polymorphisms (SNPs) discovered through genome-wide association studies (GWAS). Approximately 170 genome-wide 8 9 significant breast cancer susceptibility loci have been identified, including the recently published 65 novel loci associated with overall breast cancer and 10 loci with estrogen receptor (ER)-10 11 negative breast cancer risk, identified through the OncoArray project (15, 16). Estimation of any combined effect of genetic and environmental factors, including gene-12 13 environment (G x E) interactions is considered to possibly improve breast cancer risk prediction, and hence identification of women at high-risk for targeted prevention. However, development 14 of these risk models depends on knowledge of the joint effects of genetic and environmental risk 15 factors, in particular departures from a multiplicative model (that is, G x E interaction on relative 16 risk scale) (17). More importantly, G x E studies of individual susceptibility loci may also 17 provide insight on potential underlying biological mechanisms that could mediate causal effects 18 19 of a factor on risk of breast cancer.

Previous G x E interaction studies of breast cancer have reported nearly 30 potential G x E
interactions with little evidence of departures from multiplicative model (18, 19). Most reported
G x E interactions for breast cancer have not been replicated in independent datasets. Two G x E

interactions were replicated using data from the Breast Cancer Association Consortium (BCAC)
(20), but were not replicated in a smaller study by the Breast and Prostate Cancer Cohort
Consortium (21). In this study, we assess interactions between 205 known common breast
cancer susceptibility loci and 13 established environmental risk factors in relation to risk of
overall and estrogen receptor (ER)-specific breast cancer for women of European ancestry, using
the largest available dataset to date from the Breast Cancer Association Consortium (BCAC).
Additionally, we attempted to replicate previously reported potential G x E interactions (18).

8 Materials and Methods

9 *Study population*

We analyzed data from 46 studies (16 prospective cohorts, 14 population-based case-control 10 studies and 16 non-population based studies) participating in BCAC (Supplementary Table 1). 11 Participants were excluded if they were male, were of non-European descent, had breast tumors 12 13 of unknown invasiveness, or had in-situ disease or prevalent disease at the time of assessment. Women with unknown age at reference date (defined as date of diagnosis for cases and interview 14 15 for controls) were also excluded. For each risk factor, only studies with risk factor information 16 for at least 150 cases and 150 controls were included. All participating studies were approved by the relevant ethics committees and informed consent was obtained from study participants. 17

18 Data harmonization and variable definition

19 Data for risk factors from different studies were harmonized according to a common data

- 20 dictionary and centrally quality controlled. For both case-control and cohort studies,
- 21 epidemiological risk factor data was derived with reference to reference date (described above).
- 22 We used reference age as surrogate to categorize women as probably pre-menopausal (<54

1	years) or post-menopausal (≥54 years) status. The environmental variables available for analysis
2	were: age at menarche (per 2 years), ever parous (yes or no), and for parous women, number of
3	full-term pregnancies (1, 2, 3 and \geq 4), age at first full-term pregnancy (per 5 years), ever
4	breastfed (yes or no), duration of breastfeeding (per 12 months), and for all women, ever use of
5	oral contraceptives (yes or no), adult body mass index (BMI) separately for pre- and
6	postmenopausal women (per 5 kg/m ²), adult height (per 5 cm), lifetime alcohol consumption (per
7	10 g/day), current smoking (yes or no), and current use of combined estrogen-progesterone
8	menopausal hormonal therapy (MHT) (yes or no) as well as current use of estrogen-only MHT
9	for postmenopausal women (yes or no).
10	Genetic data
11	Samples were genotyped using one of the two SNP arrays – iCOGS(22) or OncoArray(15).
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21 interactions (**Supplementary Table 2**). These variants have been associated with breast cancer

risk either through GWAS (24-34) or by fine mapping of associated regions (35-52). Of these,

72 were identified through the OncoArray project and had not been previously evaluated for G x
 E interactions (15, 16).

For replication of the previously reported interactions, we analyzed a subset of 30 544 cases and
37 616 controls genotyped using the OncoArray array, which had not been included in previous
G x E studies. We evaluated 33 potential G x E interactions that had been previously reported
(Supplementary Table 3) (18).

7 Statistical analysis

8 Unconditional logistic regression analysis was employed to assess associations of SNPs and risk 9 factors with breast cancer risk. For SNPs, the estimated number of minor alleles based on imputation was included as a continuous variable. SNP-risk factor interactions were assessed 10 using likelihood ratio tests, based on unconditional logistic regression models with and without 11 an interaction term between the SNP and risk factor of interest. All analyses were adjusted for 12 13 study, reference age, and ten ancestry-informative principal components. To account for differential main effects of risk factors by study design, we included an interaction term between 14 15 the risk factor of interest and an indicator variable for study design (population-based and non-16 population-based), along with the main effect for study design.

Analyses were conducted separately for overall breast cancer risk and for ER-subtype specific breast cancer risk. The analyses were performed separately for women genotyped by iCOGS or OncoArray and the results were meta-analyzed using a fixed-effects inverse-variance weighted model. Between-study heterogeneity in the G x E interaction effect estimates was assessed by Cochrane's Q-test and I^2 index.

MHT was classified into estrogen-progesterone therapy (EPT) and estrogen-only therapy (ET).
Models assessing the association with current MHT use by type were adjusted for former use of
MHT and use of any MHT preparation other than the one of interest. All analyses of MHT use
were restricted to postmenopausal women. Models evaluating the association with current
smoking were adjusted for former smoking.

To assess the noteworthiness of the observed G x E interactions we calculated Bayesian False 6 7 Discovery Probability (BFDP) at five different prior probabilities for a true association (20%, 10%, 1%, 0.1% and 0.01%). G x E interactions with BFDP <80% were considered as 8 9 noteworthy. This was based on the assumption of a four-fold cost of a false non-discovery 10 compared with the cost of a false discovery and that the probability of observing a true 11 interaction odds ratio (OR) inside the range of 0.66-1.50 was 95%, as proposed by Wakefield et 12 al.(53). We also computed a complementary measure to BFDP known as approximate Bayes 13 factor (ABF). It approximates the ratio of the probability of the data given that the null 14 hypothesis is true to the probability of the data when the alternative hypothesis is true, the null 15 hypothesis being absence of any interaction. Therefore, a lower ABF favors the alternative 16 hypothesis over the null hypothesis of absence of an interaction. For noteworthy G x E 17 interactions, we performed stratified analyses by categories of the environmental risk factor using logistic regression. Analyses were carried out using SAS 9.4 or R version 3.4.2. Meta-18 19 analyses and tests of between-study heterogeneity were conducted using the R package "meta" 20 (version 4.9-2).

21 **Results**

The studies included in this analysis are listed in **Supplementary Table 1**. The number of cases and controls with data for each risk factor varied, ranging from 23 755 cases and 30 153 controls with data for parity to 5078 cases and 6867 controls with data for cumulative lifetime intake of alcohol in the iCOGS dataset and from 37 863 cases and 44 533 controls with data for parity to 12 213 cases and 13 232 controls with data for lifetime alcohol intake in the OncoArray dataset **(Supplementary Table 4 & 5)**.

7 The SNP associations with risk of overall as well as ER-subtype breast cancer were consistent with those reported in literature (15, 16) (Supplementary Table 2 & 3). The associations of the 8 environmental risk factors with breast cancer risk were as expected in the population-based 9 studies; in brief, age at menarche, being parous, number of full-term pregnancies, ever 10 11 breastfeeding, cumulative duration of breastfeeding, and premenopausal BMI were negatively 12 associated with breast cancer risk, whereas age at first full-term pregnancy, ever use of oral 13 contraceptives, postmenopausal BMI, current use of EPT, adult height, current smoking and 14 cumulative alcohol consumption were all positively associated with breast cancer risk (Table 1 & Supplementary Figures 1-3). 15

16 We identified three SNP-risk factor interactions as noteworthy (BFDP < 0.8) at ≤ 1 % prior probability (Table 2). The strongest G x E interaction was found for SNP rs4442975 and current 17 use of EPT (OR_{meta-int} = 0.85, 95% CI = 0.78 - 0.92, $p_{\text{meta-int}} = 7.4 \times 10^{-5}$, BFDP = 0.73) with 18 19 overall breast cancer at 0.1% prior probability. The minor allele of SNP rs4442975 was associated with a stronger reduced risk of breast cancer for current users of EPT ($OR_{meta} = 0.74$, 20 95% CI = 0.69 - 0.80) than for never users of MHT (OR_{meta} = 0.87, 95% CI = 0.84 - 0.90) 21 22 (Figure 1A). This interaction was also found to be noteworthy at 1% prior probability for risk of ER-positive breast cancer (OR_{meta-int} = 0.85, 95% CI = 0.78 - 0.93, $p_{meta-int} = 2.8 \times 10^{-4}$, BFDP = 23

0.46). The association of rs4442975 with reduced risk of ER-positive breast cancer was stronger
 for current users of EPT (OR_{meta} = 0.73, 95% CI = 0.68 - 0.79) than for never MHT users
 (OR_{meta} = 0.86, 95% CI = 0.83 - 0.89) (Figure 1B).

The two other noteworthy SNP-risk factor interactions were found for ER-negative breast cancer 4 risk. The interaction between rs6596100 and number of full-term pregnancies was noteworthy at 5 1% prior probability (OR_{meta-int} = 0.91, 95% CI = 0.85 - 0.96, $p_{\text{meta-int}} = 8.2 \times 10^{-4}$, BFDP = 0.74). 6 7 The minor allele of the rs6596100 variant was associated with a reduced risk of overall breast cancer ($OR_{meta} = 0.96, 95\%$ CI = 0.94 – 0.98) and ER-positive breast cancer ($OR_{meta} = 0.94, 95\%$ 8 CI = 0.92 - 0.96), respectively, but not ER-negative breast cancer ($OR_{meta} = 1.01, 95\%$ CI = 0.979 -1.05). The rs6596100 associated risk of ER-negative breast cancer appears to decrease with 10 11 number of full-term pregnancies for parous women, with the estimated per-allele OR_{meta} being 12 1.06 (95% CI = 0.95 - 1.17) for women who had had one full-term pregnancy and 0.92 (95% CI 13 = 0.82 - 1.04) for women who had had four or more full-term pregnancies (Figure 1C).

For parous women, we observed noteworthy evidence that the ER-negative breast cancer risk 14 associated with rs6596100 was also modified by age at first full-term pregnancy (OR_{meta-int} = 15 1.12, 95% CI = 1.05 - 1.19, $p_{\text{meta-int}} = 3.3 \times 10^{-4}$, BFDP = 0.56). The risk conferred by rs6596100 16 on ER-negative breast cancer was decreased for women with age at first full-term pregnancy 17 below 20 years (OR_{meta} of 0.90 (95% CI = 0.79 - 1.03)) but increased for women with age at first 18 full term pregnancy ≥ 30 years (OR_{meta} of 1.10 (95% CI = 0.97 - 1.24)) (Figure 1D). However, 19 we observed between-study heterogeneity for the interaction between rs6596100 and age at first 20 full-term pregnancy (Supplementary Figure 4). Several other interactions were found to be 21 22 noteworthy (BFDP <0.8) at 5% prior probability (Supplementary Table 6). Meta-analyzed

results of all the G x E interactions for overall and ER-subtype risk are shown in Supplementary
 Tables 7-9.

3 In replication analyses, we found evidence for two previously reported associations in the independent subset of OncoArray data (Supplementary Table 10). We estimated an interaction 4 OR for overall breast cancer of 0.80 (95% CI = 0.69-0.93, $p_{int} = 0.004$) for current EPT use and 5 6 rs13387042, a SNP for which we had previously reported an interaction OR of 0.83 (95% CI = 0.74-0.94, $p_{int} = 2.43 \times 10^{-3}$) (20). SNP rs13387042 is in strong linkage disequilibrium with 7 rs4442975; hence this result is consistent with the interaction observed for rs4442975 in the full 8 dataset. In addition, we also observed evidence for a G x E interaction between rs941764 and 9 cumulative lifetime intake of alcohol (<20 g/day vs. ≥ 20 g/day) with ER-negative breast cancer 10 risk (OR_{int} of 0.64, 95% CI = 0.45 - 0.92, p_{int} = 0.01), compared with OR_{int} of 0.53 (95% CI = 11 0.36 - 0.76, $p_{int} = 6.8 \times 10^{-4}$) in Rudolph *et al.* (54). The corresponding meta-analyzed 12 interaction OR (per 10g/day cumulative lifetime alcohol intake) based on OncoArray and iCOGS 13 14 datasets was 0.90 (95% CI = 0.81 - 0.99, $p_{int} = 0.03$). For the G x E interaction between SNP 15 rs3817198 and number of children for parous women, which had the strongest evidence for overall risk of breast cancer in previous analyses (OR_{int} of 1.06 (95% CI = 1.04 - 1.08), p_{int} = 2.416 x 10^{-06}) (20), there was weak evidence of interaction, but in the opposite direction in the 17 replication analyses (OR_{int} of 0.94 (95% CI = 0.94 - 1.00, p_{int} = 0.03). 18

19 Discussion

In this study, we evaluated all known common susceptibility loci for interactions with breast
cancer risk factors, and found little evidence for departures from a multiplicative model. We
refer to G x E interactions as effect modification conferred by epidemiological risk factors on the

association between SNPs and breast cancer risk but, it can very well be SNPs modifying the
association of risk factors with breast cancer risk. We identified three noteworthy (BFDP <0.8) G
x E interactions related to breast cancer risk based on prior probabilities ≤1%. The strongest
evidence was found for effect modification between rs4442975 and current use of EPT with
overall and ER-positive breast cancer risk. Moreover, we found evidence of interactions between
the SNP rs6596100 and number of full-term pregnancies and age at first full-term pregnancy,
respectively, for ER-negative breast cancer risk.

The SNP rs4442975 is located in an intergenic region on the long arm of chromosome 2 (2q35). 8 Another SNP within the same genomic region, rs13387042, was previously reported to show an 9 10 interaction also with current use of EPT (20). We replicated this interaction between rs13387042 11 and current use of EPT using the OncoArray dataset. The two SNPs rs13387042 and rs4442975 are highly correlated ($r^2 = 0.93$) and conditional analysis yielded a significant association only 12 for rs4442975, so that these results reflect the same interaction. Fine-mapping and functional 13 14 analyses have identified rs4442975 to be the most likely causal variant in this region (43). Thus 15 despite the small difference in the risk estimates between never and current EPT, replication of 16 this G x E interaction reinforced what we found previously, implicating the role of the *IGFBP5* 17 gene and estrogen pathway in breast cancer.

Functional analyses indicate that SNP rs4442975 lies near a transcriptional enhancer which
physically interacts with the *IGFBP5* promoter, suggesting that the T allele of rs4442975
decreases susceptibility to breast cancer via increased expression of insulin-like growth factor
binding protein 5 (IGFBP5) (43). IGFBP5 is a key member of the insulin-like growth factor
(IGF) axis which plays an important role in cellular differentiation, proliferation and apoptosis in
breast cancer (55). Activation of the IGF receptors by IGF causes phosphorylation of insulin

1	receptor substrates (IRS-1 & IRS-2). This phosphorylation cascades multiple downstream
2	signaling pathways such as Ras/mitogen-activated protein kinase (MAPK) and phosphoinositide
3	(PI3K) serine-threonine kinase (AkT) which play a role in breast carcinogenesis (56, 57).
4	Estrogen can stimulate the IGF pathway via increased expression of both insulin-like growth
5	factor receptor-1 and IRS-1. Some studies have also reported a positive correlation between
6	overexpression of IGFBP5 and the presence of ER in breast cancer cell lines. Progesterone has
7	been shown to act by increasing levels of IRS-2 and sensitizing breast cancer cells to
8	downstream signaling pathways such as MAPK and Akt (58-60). It is plausible that exogenous
9	hormone exposure due to estrogen and progesterone therapy may affect the regulation of the IGF
10	pathway and thereby modulate germline IGFPB5 variant-related susceptibility to breast cancer.
11	Note however that two other independent breast cancer risk variants in this region (tagged by
12	rs16857609 (13) and a 1.3kb insertion/deletion (49)) are also believed to target IGFBP5 but we
13	did not find evidence for interactions between these variants and current EPT use.
14	Women of young age at first pregnancy are known to have increased circulating sex hormone
15	binding globulin and prolactin but decreased total estrogen levels (61, 62). Likewise, women
16	who have had multiple full-term pregnancies have an overall decreased lifetime exposure to
17	estrogen (61, 63, 64). The association of rs6596100 with ER-negative breast cancer risk was

18 found to be modified by number of full-term pregnancies and age at first full-term pregnancy for

19 parous women. Based on INQUISIT (15), the target genes of rs6596100 and highly correlated

20 SNPs are predicted to be heat shock protein family A member 4 (*HSPA4*) and AF4/FMR2 family

21 member 4 (*AFF4*). INQUISIT predicts *HSPA4* as the most likely target due to overlap of

22 multiple correlated SNPs lying in *HSPA4* promoter region, distal regulatory elements and coding

23 sequence. *HSPA4* gene is responsible for production of heat shock proteins (Hsps), particularly

1	those belonging to the family HSP70. The underlying mechanisms regarding the relationship
2	between rs6596100 and these pregnancy-related risk factors are unknown at present. It is
3	plausible that a lower estrogenic milieu due to reproductive factors may affect the formation of
4	multi-complexes between steroid receptors like ER and heat shock proteins (HSPs), and
5	therefore affecting signaling pathways such as Wnt, ErbB, serine/threonine and tyrosine protein
6	kinase, which are known to be involved in breast carcinogenesis. While there is some biological
7	plausibility regarding the observed interactions with rs6596100, the findings nevertheless could
8	be by chance and thus require independent replication.
9	The SNP rs941764 is located on chromosome 14 in intron of CCDC88C gene (15, 22). The
10	effect modification of rs941764 associated ER-negative breast cancer risk by lifetime intake of
	· ·
11	alcohol was first reported by Rudolph et al.(54). We replicated this G x E interaction in an
12	independent dataset in our study. Mutations in this gene region have been associated with
13	dysregulation of Wnt signaling in neural disorders such as congenital hydrocephalus (65). This
14	gene codes a Hook-related protein (HkRP2) that binds to an important scaffold protein,
15	Dishevelled, in the Wnt signaling pathway, affecting all downstream activity (65).
16	A role of alcohol has been well recognized in initiation and progression of breast cancer
17	presumably via multiple cellular and molecular mechanisms, including the EGFR/ErbB2
18	pathways. Downstream to EGFR/ErbB2 pathways lie multiple pathways such as the MAPK,
19	Wnt/GSK3 β / β -catenin pathways (66). Therefore, alcohol consumption could affect the risk of
20	ER-negative breast cancer through dysregulation of Wnt signaling.

Our study provides the most comprehensive evaluation to date of potential effect modification of
all known common genetic susceptibility variants by environmental risk factors for breast

1 cancer. Our findings are based on the largest available dataset on breast cancer. Despite its large sample size, the study may remain statistically underpowered, considering the rather modest 2 effect sizes of most of the common variants associated with breast cancer risk, and particularly 3 for risk factors for which we have less data (Supplementary table 11) (18). Statistical power was 4 5 further diminished for subtype-specific analyses due to reduced sample sizes, especially for ER-6 negative breast cancer (10,896 ER-negative cases in the combined iCOGS and OncoArray dataset) (18). The lack of strong effect modifications for breast cancer could also be explained 7 by the overall weak to moderate associations of environmental risk factors except for MHT use 8 9 with breast cancer risk along with the modest associations of common genetic variants. A further limitation of our study is that the findings may not be generalizable to other racial/ethnic groups 10 11 since the analyses were restricted to women of European ancestry.

12 In conclusion, our analyses suggest that most of the associated effects of breast cancer 13 susceptibility loci and environmental risk factors are consistent with a multiplicative model. The 14 strongest evidence for an interaction was between the candidate causal variant rs4442975 at 2q35 15 and current use of EPT. The associated effect is supported by a plausible underlying biological 16 mechanism, but further epidemiological and functional validation will be required to determine 17 whether the interaction is genuine. The newly reported results for ER-negative breast cancer risk generate plausible biological hypotheses and may inform future functional studies. Overall, the 18 19 results from our analyses do not suggest strong effect modification of the association between 20 breast cancer susceptibility loci and risk of breast cancer by established epidemiological risk factors. 21

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11	
12	
13	
1.4	
14	
15	
16	
17	
18	
19	

1 References

Breast cancer and hormone replacement therapy: collaborative reanalysis of data from 51
 epidemiological studies of 52,705 women with breast cancer and 108,411 women without breast
 cancer. Collaborative Group on Hormonal Factors in Breast Cancer. Lancet.

5 1997;350(9084):1047-59.

Baer HJ, Rich-Edwards JW, Colditz GA, Hunter DJ, Willett WC, Michels KB. Adult
height, age at attained height, and incidence of breast cancer in premenopausal women. Int J
Cancer. 2006;119(9):2231-5.

9 3. Hunter DJ, Colditz GA, Hankinson SE, Malspeis S, Spiegelman D, Chen W, et al. Oral
10 contraceptive use and breast cancer: a prospective study of young women. Cancer Epidemiol
11 Biomarkers Prev. 2010;19(10):2496-502.

Collaborative Group on Hormonal Factors in Breast C. Menarche, menopause, and breast
 cancer risk: individual participant meta-analysis, including 118 964 women with breast cancer
 from 117 epidemiological studies. Lancet Oncol. 2012;13(11):1141-51.

15 5. Jung S, Wang M, Anderson K, Baglietto L, Bergkvist L, Bernstein L, et al. Alcohol

16 consumption and breast cancer risk by estrogen receptor status: in a pooled analysis of 20

17 studies. Int J Epidemiol. 2016;45(3):916-28.

18 6. World Cancer Research Fund International/American Institute for Cancer Research.

19 Continuous Update Project Report: Diet, Nutrition, Physical Activity and Breast Cancer. 2017.

20 7. Easton DF, Ford D, Bishop DT. Breast and ovarian cancer incidence in BRCA1-mutation

carriers. Breast Cancer Linkage Consortium. Am J Hum Genet. 1995;56(1):265-71.

8. Breast Cancer Linkage C. Cancer risks in BRCA2 mutation carriers. J Natl Cancer Inst.

23 1999;91(15):1310-6.

1	9.	King MC, Marks JH, Mandell JB, New York Breast Cancer Study G. Breast and ovarian
2	cance	r risks due to inherited mutations in BRCA1 and BRCA2. Science. 2003;302(5645):643-6.
3	10.	Garber JE, Goldstein AM, Kantor AF, Dreyfus MG, Fraumeni JF, Jr., Li FP. Follow-up
4	study	of twenty-four families with Li-Fraumeni syndrome. Cancer Res. 1991;51(22):6094-7.
5	11.	Birch JM, Alston RD, McNally RJ, Evans DG, Kelsey AM, Harris M, et al. Relative
6	freque	ency and morphology of cancers in carriers of germline TP53 mutations. Oncogene.
7	2001;	20(34):4621-8.
8	12.	Rapakko K, Allinen M, Syrjakoski K, Vahteristo P, Huusko P, Vahakangas K, et al.
9	Germ	line TP53 alterations in Finnish breast cancer families are rare and occur at conserved
10	mutat	ion-prone sites. Br J Cancer. 2001;84(1):116-9.
11	13.	Meijers-Heijboer H, van den Ouweland A, Klijn J, Wasielewski M, de Snoo A,
12	Olden	burg R, et al. Low-penetrance susceptibility to breast cancer due to CHEK2(*)1100delC in
13	nonca	rriers of BRCA1 or BRCA2 mutations. Nat Genet. 2002;31(1):55-9.
14	14.	Thompson D, Duedal S, Kirner J, McGuffog L, Last J, Reiman A, et al. Cancer risks and
15	morta	lity in heterozygous ATM mutation carriers. J Natl Cancer Inst. 2005;97(11):813-22.
16	15.	Michailidou K, Lindstrom S, Dennis J, Beesley J, Hui S, Kar S, et al. Association
17	analys	sis identifies 65 new breast cancer risk loci. Nature. 2017;551(7678):92-4.
18	16.	Milne RL, Kuchenbaecker KB, Michailidou K, Beesley J, Kar S, Lindstrom S, et al.
19	Identi	fication of ten variants associated with risk of estrogen-receptor-negative breast cancer. Nat
20	Genet	. 2017;49(12):1767-78.
21	17.	Hunter DJ. Gene-environment interactions in human diseases. Nat Rev Genet.

22 2005;6(4):287-98.

1	18.	Rudolph A, Chang-Claude J, Schmidt MK. Gene-environment interaction and risk of
2	breast	cancer. Br J Cancer. 2016;114(2):125-33.

3	19. Barrdahl M, Rudolph A, Hopper JL, Southey MC, Broeks A, Fasching PA, et al. Gene-
4	environment interactions involving functional variants: Results from the Breast Cancer
5	Association Consortium. Int J Cancer. 2017;141(9):1830-40.
6	20. Nickels S, Truong T, Hein R, Stevens K, Buck K, Behrens S, et al. Evidence of gene-
7	environment interactions between common breast cancer susceptibility loci and established
8	environmental risk factors. PLoS Genet. 2013;9(3):e1003284.
9	21. Barrdahl M, Canzian F, Joshi AD, Travis RC, Chang-Claude J, Auer PL, et al. Post-
10	GWAS gene-environment interplay in breast cancer: results from the Breast and Prostate Cancer
11	Cohort Consortium and a meta-analysis on 79,000 women. Hum Mol Genet. 2014;23(19):5260-
12	70.
13	22. Michailidou K, Hall P, Gonzalez-Neira A, Ghoussaini M, Dennis J, Milne RL, et al.
14	Large-scale genotyping identifies 41 new loci associated with breast cancer risk. Nat Genet.
15	2013;45(4):353-61, 61e1-2.
16	23. Amos CI, Dennis J, Wang Z, Byun J, Schumacher FR, Gayther SA, et al. The OncoArray
17	Consortium: A Network for Understanding the Genetic Architecture of Common Cancers.
18	Cancer Epidemiol Biomarkers Prev. 2017;26(1):126-35.
19	24. Easton DF, Pooley KA, Dunning AM, Pharoah PD, Thompson D, Ballinger DG, et al.
20	Genome-wide association study identifies novel breast cancer susceptibility loci. Nature.

21 2007;447(7148):1087-93.

1 25. Thomas G, Jacobs KB, Kraft P, Yeager M, Wacholder S, Cox DG, et al. A multistage genome-wide association study in breast cancer identifies two new risk alleles at 1p11.2 and 2 14q24.1 (RAD51L1). Nat Genet. 2009;41(5):579-84. 3 4 26. Turnbull C, Ahmed S, Morrison J, Pernet D, Renwick A, Maranian M, et al. Genome-5 wide association study identifies five new breast cancer susceptibility loci. Nat Genet. 6 2010;42(6):504-7. 27. 7 Ghoussaini M, Fletcher O, Michailidou K, Turnbull C, Schmidt MK, Dicks E, et al. Genome-wide association analysis identifies three new breast cancer susceptibility loci. Nat 8 9 Genet. 2012;44(3):312-8. 28. Long J, Cai Q, Sung H, Shi J, Zhang B, Choi JY, et al. Genome-wide association study in 10 east Asians identifies novel susceptibility loci for breast cancer. PLoS Genet. 11

12 2012;8(2):e1002532.

13 29. Siddiq A, Couch FJ, Chen GK, Lindstrom S, Eccles D, Millikan RC, et al. A meta-

14 analysis of genome-wide association studies of breast cancer identifies two novel susceptibility

15 loci at 6q14 and 20q11. Hum Mol Genet. 2012;21(24):5373-84.

16 30. Garcia-Closas M, Couch FJ, Lindstrom S, Michailidou K, Schmidt MK, Brook MN, et al.

17 Genome-wide association studies identify four ER negative-specific breast cancer risk loci. Nat

18 Genet. 2013;45(4):392-8, 8e1-2.

19 31. Cai Q, Zhang B, Sung H, Low SK, Kweon SS, Lu W, et al. Genome-wide association

analysis in East Asians identifies breast cancer susceptibility loci at 1q32.1, 5q14.3 and 15q26.1.

21 Nat Genet. 2014;46(8):886-90.

22 32. Milne RL, Burwinkel B, Michailidou K, Arias-Perez JI, Zamora MP, Menendez-

23 Rodriguez P, et al. Common non-synonymous SNPs associated with breast cancer susceptibility:

findings from the Breast Cancer Association Consortium. Hum Mol Genet. 2014;23(22):6096 111.

3 33. Sawyer E, Roylance R, Petridis C, Brook MN, Nowinski S, Papouli E, et al. Genetic
4 predisposition to in situ and invasive lobular carcinoma of the breast. PLoS Genet.

5 2014;10(4):e1004285.

6 34. Couch FJ, Kuchenbaecker KB, Michailidou K, Mendoza-Fandino GA, Nord S, Lilyquist

J, et al. Identification of four novel susceptibility loci for oestrogen receptor negative breast
cancer. Nat Commun. 2016;7:11375.

9 35. Ahmed S, Thomas G, Ghoussaini M, Healey CS, Humphreys MK, Platte R, et al. Newly

discovered breast cancer susceptibility loci on 3p24 and 17q23.2. Nat Genet. 2009;41(5):585-90.

36. Udler MS, Ahmed S, Healey CS, Meyer K, Struewing J, Maranian M, et al. Fine scale
mapping of the breast cancer 16q12 locus. Hum Mol Genet. 2010;19(12):2507-15.

13 37. Haiman CA, Chen GK, Vachon CM, Canzian F, Dunning A, Millikan RC, et al. A

common variant at the TERT-CLPTM1L locus is associated with estrogen receptor-negative

15 breast cancer. Nat Genet. 2011;43(12):1210-4.

16 38. Bojesen SE, Pooley KA, Johnatty SE, Beesley J, Michailidou K, Tyrer JP, et al. Multiple

17 independent variants at the TERT locus are associated with telomere length and risks of breast

18 and ovarian cancer. Nat Genet. 2013;45(4):371-84, 84e1-2.

19 39. French JD, Ghoussaini M, Edwards SL, Meyer KB, Michailidou K, Ahmed S, et al.

20 Functional variants at the 11q13 risk locus for breast cancer regulate cyclin D1 expression

through long-range enhancers. Am J Hum Genet. 2013;92(4):489-503.

1	40. Gaudet MM, Kuchenbaecker KB, Vijai J, Klein RJ, Kirchhoff T, McGuffog L, et al.	
2	Identification of a BRCA2-specific modifier locus at 6p24 related to breast cancer risk. PLoS	
3	Genet. 2013;9(3):e1003173.	
4	41. Long J, Delahanty RJ, Li G, Gao YT, Lu W, Cai Q, et al. A common deletion in the	
5	APOBEC3 genes and breast cancer risk. J Natl Cancer Inst. 2013;105(8):573-9.	
6	42. Meyer KB, O'Reilly M, Michailidou K, Carlebur S, Edwards SL, French JD, et al. Fine-	-
7	scale mapping of the FGFR2 breast cancer risk locus: putative functional variants differentially	
8	bind FOXA1 and E2F1. Am J Hum Genet. 2013;93(6):1046-60.	
9	43. Ghoussaini M, Edwards SL, Michailidou K, Nord S, Cowper-Sal Lari R, Desai K, et al.	
10	Evidence that breast cancer risk at the 2q35 locus is mediated through IGFBP5 regulation. Nat	
11	Commun. 2014;4:4999.	
12	44. Darabi H, McCue K, Beesley J, Michailidou K, Nord S, Kar S, et al. Polymorphisms in	a
13	Putative Enhancer at the 10q21.2 Breast Cancer Risk Locus Regulate NRBF2 Expression. Am	J
14	Hum Genet. 2015;97(1):22-34.	
15	45. Glubb DM, Maranian MJ, Michailidou K, Pooley KA, Meyer KB, Kar S, et al. Fine-sca	le
16	mapping of the 5q11.2 breast cancer locus reveals at least three independent risk variants	
17	regulating MAP3K1. Am J Hum Genet. 2015;96(1):5-20.	
18	46. Lin WY, Camp NJ, Ghoussaini M, Beesley J, Michailidou K, Hopper JL, et al.	
19	Identification and characterization of novel associations in the CASP8/ALS2CR12 region on	
20	chromosome 2 with breast cancer risk. Hum Mol Genet. 2015;24(1):285-98.	
21	47. Orr N, Dudbridge F, Dryden N, Maguire S, Novo D, Perrakis E, et al. Fine-mapping	
22	identifies two additional breast cancer susceptibility loci at 9q31.2. Hum Mol Genet.	
23	2015;24(10):2966-84.	

1	48. Darabi H, Beesley J, Droit A, Kar S, Nord S, Moradi Marjaneh M, et al. Fine scale
2	mapping of the 17q22 breast cancer locus using dense SNPs, genotyped within the Collaborative
3	Oncological Gene-Environment Study (COGs). Sci Rep. 2016;6:32512.
4	49. Dunning AM, Michailidou K, Kuchenbaecker KB, Thompson D, French JD, Beesley J, e
5	al. Breast cancer risk variants at 6q25 display different phenotype associations and regulate
6	ESR1, RMND1 and CCDC170. Nat Genet. 2016;48(4):374-86.
7	50. Ghoussaini M, French JD, Michailidou K, Nord S, Beesley J, Canisus S, et al. Evidence
8	that the 5p12 Variant rs10941679 Confers Susceptibility to Estrogen-Receptor-Positive Breast
9	Cancer through FGF10 and MRPS30 Regulation. Am J Hum Genet. 2016;99(4):903-11.
10	51. Lawrenson K, Kar S, McCue K, Kuchenbaeker K, Michailidou K, Tyrer J, et al.
11	Functional mechanisms underlying pleiotropic risk alleles at the 19p13.1 breast-ovarian cancer
12	susceptibility locus. Nat Commun. 2016;7:12675.
13	52. Wyszynski A, Hong CC, Lam K, Michailidou K, Lytle C, Yao S, et al. An intergenic risk
14	locus containing an enhancer deletion in 2q35 modulates breast cancer risk by deregulating
15	IGFBP5 expression. Hum Mol Genet. 2016;25(17):3863-76.
16	53. Wakefield J. A Bayesian measure of the probability of false discovery in genetic
17	epidemiology studies. Am J Hum Genet. 2007;81(2):208-27.
18	54. Rudolph A, Milne RL, Truong T, Knight JA, Seibold P, Flesch-Janys D, et al.
19	Investigation of gene-environment interactions between 47 newly identified breast cancer
20	susceptibility loci and environmental risk factors. Int J Cancer. 2015;136(6):E685-96.
21	55. Beattie J, Allan GJ, Lochrie JD, Flint DJ. Insulin-like growth factor-binding protein-5
22	(IGFBP-5): a critical member of the IGF axis. Biochem J. 2006;395(1):1-19.

1	56.	Akkiprik M, Feng Y, Wang H, Chen K, Hu L, Sahin A, et al. Multifunctional roles of
2	insulin	-like growth factor binding protein 5 in breast cancer. Breast Cancer Res. 2008;10(4):212.
3	57.	Zhang X, Yee D. Tyrosine kinase signalling in breast cancer: insulin-like growth factors
4	and the	eir receptors in breast cancer. Breast Cancer Res. 2000;2(3):170-5.
5	58.	Cui X, Lazard Z, Zhang P, Hopp TA, Lee AV. Progesterone crosstalks with insulin-like
6	growth	factor signaling in breast cancer cells via induction of insulin receptor substrate-2.
7	Oncog	ene. 2003;22(44):6937-41.
8	59.	Fagan DH, Yee D. Crosstalk between IGF1R and estrogen receptor signaling in breast
9	cancer	. J Mammary Gland Biol Neoplasia. 2008;13(4):423-9.
10	60.	Yee D, Lee AV. Crosstalk between the insulin-like growth factors and estrogens in breast
11	cancer	. J Mammary Gland Biol Neoplasia. 2000;5(1):107-15.
12	61.	Kelsey JL, Gammon MD, John EM. Reproductive factors and breast cancer. Epidemiol
13	Rev. 1	993;15(1):36-47.
14	62.	Bernstein L. Epidemiology of endocrine-related risk factors for breast cancer. J
15	Mamm	nary Gland Biol Neoplasia. 2002;7(1):3-15.
16	63.	Dall GV, Britt KL. Estrogen Effects on the Mammary Gland in Early and Late Life and
17	Breast	Cancer Risk. Front Oncol. 2017;7:110.
18	64.	Fortner RT, Hankinson SE. Reproductive and Hormonal factors and Breast Cancer.
19	Transla	ational Endocrinology & Metabolism: Breast Cancer Update. 32012. p. 95-116.
20	65.	Ekici AB, Hilfinger D, Jatzwauk M, Thiel CT, Wenzel D, Lorenz I, et al. Disturbed Wnt
21	Signal	ling due to a Mutation in CCDC88C Causes an Autosomal Recessive Non-Syndromic
22	Hydro	cephalus with Medial Diverticulum. Mol Syndromol. 2010;1(3):99-112.

1	66. Wang Y, Xu M, Ke ZJ, Luo J. Cellular and molecular mechanisms underlying alcohol-
2	induced aggressiveness of breast cancer. Pharmacol Res. 2017;115:299-308.
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Supplementary Information

Supplementary Table 1 (S1): Studies participating in G x E analysis with number of cases and controls.

Supplementary Table 2 (S2): Associations between 205 common breast cancer susceptibility loci with breast cancer risk in European population, overall and by ER status.

Supplementary Table 3 (S3): Associations between 33 replication SNPs with breast cancer risk in European population, overall and by ER status.

Supplementary Table 4 (S4): Number of cases and controls for each environmental risk factor by study design in iCOGS and OncoArray dataset.

Supplementary Table 5 (S5): Number of cases and controls for each environmental risk factor by overall and ER-status in complete and replication dataset.

Supplementary Table 6 (S6): G x E interactions with BFDP <80% at 5% prior probability (meta-analyzed results).

Supplementary Table 7 (S7): Meta-analyzed G x E interactions between 205 common genetic susceptibility loci and environmental risk factors for overall breast cancer risk.

Supplementary Table 8 (S8): Meta-analyzed G x E interactions between 205 common genetic susceptibility loci and environmental risk factors for ER-positive breast cancer risk.

Supplementary Table 9 (S9): Meta-analyzed G x E interactions between 205 common genetic susceptibility loci and environmental risk factors for ER-negative breast cancer risk.

Supplementary Table 10 (S10): Interaction odds ratio (OR) and 95% confidence intervals (CI) for previously reported G x E interactions in an independent dataset.

Supplementary Table 11 (S11): Power for detecting different gene-environment interaction effect estimates (OR of 0.75 to 1.50) given different minor allele frequencies (0.05 to 0.45) for 1:1 unmatched case-control study. Power calculation is performed by Quanto 1.2.4, assuming a population prevalence of disease of 1%, 15% prevalence of the environmental factor. We assumed a log-additive inheritance model with marginal effect estimate of SNP = 1.10 and marginal effect estimate of environmental factor = 1.20 and two-sided alpha of 5 x 10^{-8} .

Supplementary Figure 1: Forest plot of meta-analyzed study-wise odds ratios and 95% confidence intervals of population-based studies for associations between environmental risk factors and overall breast cancer risk

Supplementary Figure 2: Forest plot of meta-analyzed study-wise odds ratios and 95% confidence intervals of population-based studies for associations between environmental risk factors and ER-positive breast cancer risk

Supplementary Figure 3: Forest plot of meta-analyzed study-wise odds ratios and 95% confidence intervals of population-based studies for associations between environmental risk factors and ER-negative breast cancer risk

Supplementary Figure 4: Forest plot of meta-analyses of study-wise odds ratios and 95% confidence intervals for G x E interactions between SNPs and environmental risk factors of breast cancer (from Table 2) separately for OncoArray and iCOGS datasets.

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Environmental risk factor	Overall brea	st cancer risk	ER-positive b	reast cancer risk	ER-negative breast cancer ri		
	Cases/Controls	OR (95% CI)	Cases/Controls	OR (95% CI)	Cases/Controls	OR (95% CI)	
Age at menarche (per 2 years)	36893/46854	0.91 (0.89-0.92)	26630/46854	0.91 (0.89-0.93)	4255/25233	0.89 (0.85-0.93)	
Ever parous (yes/no)	37242/47173	0.81 (0.77-0.84)	26937/47173	0.78 (0.74-0.81)	4309/25585	0.94 (0.85-1.04)	
Number of full-term pregnancies $(1,2,3,\geq 4)$	31390/41215	0.87 (0.85-0.88)	22720/41215	0.86 (0.84-0.87)	3273/18267	0.90 (0.86-0.94)	
Age at first full-term pregnancy (per 5 years) ¹	30168/39850	1.14 (1.12-1.16)	21869/39850	1.17 (1.14-1.19)	3472/21422	1.02 (0.97-1.06)	
Ever breastfed (yes/no) ¹	27786/30582	0.91 (0.88-0.95)	19691/30582	0.92 (0.88-0.96)	3533/19606	0.96 (0.88-1.03)	
Duration of breastfeeding (per 12 months) ¹	24553/25524	0.96 (0.93-0.98)	17355/25524	0.95 (0.93-0.98)	3315/18012	0.98 (0.94-1.03)	
Adult height (per 5 cm)	35767/46506	1.09 (1.08-1.10)	25763/46506	1.10 (1.09-1.12)	3954/24342	1.03 (1.00-1.05)	
Premenopausal BMI (per 5 kg/m ²)	7994/10066	0.95 (0.92-0.98)	4835/9490	0.92 (0.89-0.95)	913/2030	1.07 (0.98-1.16)	
Postmenopausal BMI (per 5 kg/m ²)	27495/32495	1.07 (1.05-1.09)	20503/32283	1.07 (1.05-1.09)	1758/11859	1.05 (1.00-1.11)	
Ever use of oral contraceptives (yes/no)	35126/44608	1.22 (1.18-1.26)	25271/44608	1.24 (1.20-1.29)	3939/24225	1.14 (1.05-1.23)	
Current use of EPT (yes/no) ^{2,3}	16637/17946	1.75 (1.65-1.87)	12566/17946	1.93 (1.81-2.06)	1190/7353	1.11 (0.92-1.34)	
Current use of ET (yes/no) ^{2,3}	16444/17920	1.10 (1.03-1.17)	11829/16844	1.11 (1.03-1.19)	936/6262	1.35 (1.11-1.64)	
Lifetime intake of alcohol (per 10 g/day)	15827/18723	1.07 (1.05-1.10)	11302/18723	1.09 (1.07-1.11)	1612/11562	1.03 (0.98-1.08)	
Current smoking (yes/no) ⁴	33737/43222	1.18 (1.13-1.24)	24123/43222	1.18 (1.12-1.25)	3707/22573	1.06 (0.96-1.18)	
Pack years smoked (per 10 pack-years) ⁵	7975/11709	1.02 (1.00-1.04)	5944/11709	1.02 (1.00-1.04)	896/6400	1.00 (0.95-1.04)	

Table 1: Main effects for the epidemiologic variables included in the analyses, derived from population-based studies only.

ER: Estrogen receptor, OR: odd ratio, CI: confidence interval, BMI: Body mass index, EPT: Estrogen-Progesterone menopausal hormonal therapy, ET: Estrogen-only menopausal hormonal therapy

All models were adjusted for reference age and study

¹ for parous women

² for postmenopausal women
 ³ Additionally, models were adjusted for former use of menopausal hormonal therapy and use of any other menopausal hormonal therapy preparations

⁴ Additionally, model was adjusted for former smoking

⁵ for ever smokers

Table 2: Gene-environment interactions with Bayesian False Discovery Probability (BFDP) <80% at ≤1% prior probability	1.
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		iCOGS	OncoArray	Meta-an	alysis	Prior probability (BFDP)			FDP)			
Environmental risk factor	SNP (Gene)	OR _{int} (95% CI)	OR _{int} (95% CI)	OR _{int} (95% CI)	p _{int}	0.2	0.1	0.01	0.001	0.0001	ABF	
OVERALL BREAST CANCER RISK												
Current EPT use ¹	rs4442975 (<i>IGFBP5</i>)	0.88 (0.75 – 1.03)	0.83 (0.76 – 0.92)	0.85 (0.78 – 0.92)	7.4E-05	0.011	0.023	0.209	0.727	0.964	0.003	
			ER-POSITIVE	BREAST CAN	ICER RISK	5						
Current EPT use ¹	rs4442975 (<i>IGFBP5</i>)	0.89 (0.75 – 1.06)	0.84 (0.75 - 0.93)	0.85 (0.78 – 0.93)	2.8E-04	0.033	0.072	0.462	0.896	0.989	0.009	
			ER-NEGATIVI	E BREAST CA	NCER RISI	K						
Number of full-term pregnancies ^{2,3}	rs6596100 (HSPA4)	0.84 (0.75 – 0.93)	0.94 (0.87 - 1.01)	0.91 (0.85 – 0.96)	8.2E-04	0.104	0.207	0.742	0.967	0.997	0.029	
Age at FFTP ²	rs6596100 (HSPA4)	1.13 (1.02 – 1.26)	1.11 (1.03 – 1.19)	1.12 (1.05 – 1.19)	3.3E-04	0.048	0.103	0.558	0.927	0.992	0.012	
ER: Estrogen receptor, Estrogen-Progesterone to for postmenopausal we for parous women only categories: 1,2,3, ≥ 4	therapy, FFTP:			val, SNP: Single	e nucleotide	polymor	phism, A	BF: Appr	oximate E	Bayes Factor	:, EPT:	

A. Overall breast cancer, rs4442975 x Current use of Estrogen-Progesterone therapy among postmenopausal women, p-interaction = 7.4E-05 B. ER-positive breast cancer, rs4442975 x Current use of Estrogen-Progesterone therapy among postmenopausal women, p-interaction = 2.8E-04

OR	OR	95%-CI		OR	OR	95%-CI
Never users of Estrogen-Progesterone therapy			Never users of Estrogen-Progesterone thera	py		
ONCO -	0.87	[0.84; 0.90]	ONCO -	20	0.86	[0.82; 0.89]
icogs -	0.87	[0.82; 0.92]	iCOGS -		0.85	[0.80; 0.91]
Fixed effect model	0.87	[0.84; 0.90]	Fixed effect model		0.86	[0.83; 0.89]
Heterogeneity: $I^2 = 0\%$, $\tau^2 = 0$, $p = 0.96$			Heterogeneity: $I^2 = 0\%$, $\tau^2 = 0$, $p = 0.98$			
Current users of Estrogen-Progesterone therapy			Current users of Estrogen-Progesterone			
ONCO —	0.74	[0.67; 0.80]	ONCO		0.72	[0.66; 0.79]
icogs —	0.76	[0.66; 0.89]	icogs —		0.76	[0.64; 0.89]
Fixed effect model	0.74	[0.69; 0.80]	Fixed effect model		0.73	[0.68: 0.79]
Heterogeneity: $I^2 = 0\%$, $\tau^2 = 0$, $p = 0.68$			Heterogeneity: $I^2 = 0\%$, $\tau^2 = 0$, $p = 0.64$			
			· · · · -	1		
0.75 1	1.5		0.75	1	1.5	

Models are adjusted for reference age, study, ten principal components, former use of menopausal hormone therapy (MHT), and use of any other type of MHT preparation than the one of interest

among parous monten, p interaction - ore				parous moment p interaction - sise of		
	OR	OR	95%-CI	OR	OR	95%-CI
No. of full-term pregnancies = 1	ЧĽ			Age at first full-term pregnancy (<20 years)		
ONCO -	-	1.02	[0.89; 1.15]	ONCO	0.89	[0.76; 1.04]
ICOGS		1.13	[0.96; 1.34]	iCOGS	0.92	[0.72; 1.17]
Fixed effect model	-	1.06	[0.95; 1.17]	Fixed effect model	0.90	[0.79; 1.03]
Heterogeneity: $I^2 = 5\%$, $\tau^2 = 0.0003$, $p = 0.31$				Heterogeneity: $I^2 = 0\%$, $\tau^2 = 0$, $p = 0.84$		•
No. of full-term pregnancies = 2				Age at first full-term pregnancy (20-24 years)		
ONCO	-	1.01	[0.94; 1.10]	ONCO —	0.97	[0.89; 1.05]
ICOGS		1.18	[1.06; 1.32]	iCOGS -	- 1.05	[0.92; 1.20]
Fixed effect model	•	1.07	[1.00; 1.14]	Fixed effect model	0.99	[0.92; 1.07]
Heterogeneity: $I^2 = 81\%$, $\tau^2 = 0.0099$, $p = 0.02$				Heterogeneity: $I^2 = 5\%$, $\tau^2 = 0.0002$, $p = 0.30$		
No. of full-term pregnancies = 3				Age at first full-term pregnancy (25-29 years)		
ONCO —	H	0.91	[0.81; 1.02]	ONCO —	0.97	[0.88; 1.07]
iCOGS	•	1.00	[0.85; 1.17]	iCOGS	1.13	[0.97; 1.32]
Fixed effect model		0.94	[0.86; 1.03]	Fixed effect model	1.02	[0.93; 1.10]
Heterogeneity: $I^2 = 0\%$, $\tau^2 = 0$, $p = 0.36$				Heterogeneity: $I^2 = 64\%$, $\tau^2 = 0.0075$, $p = 0.10$		
No. of full-term pregnancies = 4+				Age at first full-term pregnancy (≥30 years)		
ONCO -		0.96	[0.84; 1.09]	ONCO	- 1.06	[0.91; 1.24]
iCOGS		0.81	[0.63; 1.05]	iCOGS	1.16	[0.95; 1.41]
Fixed effect model		0.92	[0.82; 1.04]	Fixed effect model	1.10	[0.97; 1.24]
Heterogeneity: $I^2 = 20\%$, $\tau^2 = 0.0027$, $\rho = 0.26$		l.		Heterogeneity: I ² = 0%, τ ² = 0, ρ = 0.51		
0.75	1 1.	.5		0.8 1	1.25	
Models are adjusted for reference age, st	udy, and ten prin	ncipal co	omponents	Models are adjusted for reference age, study, and te	en principal co	mponents

C. ER-negative breast cancer, rs6596100 x Number of full-term pregnancies among parous women, p-interaction = 8.2E-04 D. ER-negative breast cancer, rs6596100 x Age at first full-term pregnancy among parous women, p-interaction = 3.3E-04