# Expression of DNA Methyltransferase (DNMT) 1, 3a and 3b Proteins in Human Hepatocellular Carcinoma

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Alteration of aberrant DNA methylation is one of the most consistent epigenetic changes found in human cancers. DNA methylation is catalyzed by DNA methyltransferase (DNMT). In this study, we examined DNMT protein expression by immunohistochemistry in surgically resected hepatocellular carcinomas (HCCs). Sections of paraffin-embedded specimens were obtained from 95 patients with HCC between 1989 and 2002. The specimens were stained with anti-DNMTs (DNMT1, DNMT3a and DNMT3b) antibodies. There were statistically significant associations between DNMT protein expression and tumor differentiation (P < 0.05) and intrahepatic metastasis (P < 0.05). DNMT3a protein expression was significantly correlated with portal vein involvement of tumors (P < 0.05). The overall survival rates of patients with DNMT3a-positive HCCs and DNMT3b-positive HCCs were significantly lower than those of patients negative for these proteins (P < 0.005, respectively). To further evaluate the correlation between DNMT protein expression and patient survival, we classified patients into 3 groups: Group 1, DNMT1(+), 3a(-) and 3b(-); Group 2) DNMT1(+), 3a or 3b(+); and Group 3) DNMT1(+), 3a(+) and 3b(+). The overall survival rate of patients in Group 3 was significantly lower than those of patients in Groups 1 and 2 (P = 0.0009). In conclusion, the results of this study suggest that DNMT1. DNMT3a and DNMT3b are cooperatively involved in determining the extent of HCCs, and that DNMT protein overexpression in HCCs may be a predictive factor for poor survival.

Key words: DNA methyltransferase; hepatocellular carcinoma; immunohistochemistry; prognosis

DNA methylation plays an important role in the regulation of gene expression, and gene silencing by aberrant hypermethylation of CpG islands in gene promoter regions frequently occurs in human cancers. DNA methyltransferases are involved in the transfer of methyl groups to promoter CpG islands, and many tumor suppressor genes can be silenced by promoter CpG island methylation. Alteration of aberrant DNA methylation is one of the most consistent epigenetic changes found in hu-

man cancers (Girault et al., 2003; Ding et al., 2008; Nosho et al., 2009).

DNA methyltransferases have been reported to be encoded by 3 distinct families of DNMT genes: DNMT1, DNMT2, and DNMT3. DNMT1 is the best known DNMT involved in the maintenance of methylation, while DNMT3 is involved in de novo methylation and is encoded by 2 distinct genes, DNMT3a and DNMT3b (Łuczak and Jagodzinski, 2006). The function of DNMT2

Abbreviations: CH, chronic hepatitis; DNMT, DNA methyltransferase; HCC, hepatocellular carcinoma; LC, liver cirrhosis; siRNA, small interfering RNA

Patients	п	Immunostaining score					
		DNMT	1	DNMT	3a -	DNMT	3b
Age (year)							
< 60	39	$2.179 \pm 0.721$	NS	$1.897 \pm 0.882$	NS	$1.205 \pm 0.656$	NS
≥ 60	56	$2.357 \pm 0.586$		$1.786 \pm 0.847$		$1.232 \pm 0.831$	
Sex							
Male	76	$2.263 \pm 0.640$	NS	$1.776 \pm 0.873$	NS	$1.145 \pm 0.905$	NS
Female	19	$2.368 \pm 0.684$		$2.053 \pm 0.800$		$1.526 \pm 0.905$	
Background of cancerous liver	tissue						
Chronic hepatitis	27	$2.370 \pm 0.655$	NS	$1.556 \pm 1.013$	NS	$1.000 \pm 0.679$	NS
Liver cirrhosis	68	$2.250 \pm 0.629$		$1.941 \pm 0.770$		$1.309 \pm 0.778$	
Tumor differentiation							
Well differentiated	19	$1.842 \pm 0.765$	P < 0.05	$1.421 \pm 0.769$	P < 0.05	$0.895 \pm 0.657$	P < 0.05
Moderately differentiated	59	$2.339 \pm 0.633$		$1.814 \pm 0.880$		$1.254 \pm 0.779$	
Poorly differentiated	17	$2.471 \pm 0.624$		$2.235 \pm 0.831$		$1.412 \pm 0.870$	
Portal vein involvement of tum	or						
Positive	51	$2.314 \pm 0.678$	NS	$2.000 \pm 0.849$	P < 0.05	$1.294 \pm 0.782$	NS
Negative	44	$2.250 \pm 0.615$		$1.636 \pm 0.838$		$1.136 \pm 0.734$	
Pathological findings Fc-inf							
Positive	68	$2.319 \pm 0.675$	NS	$1.870 \pm 0.873$	NS	$1.188 \pm 0.791$	NS
Negative	27	$2.192 \pm 0.567$		$1.731 \pm 0.827$		$1.308 \pm 0.679$	
Intrahepatic metastasis							
Positive	28	$2.571 \pm 0.504$	P < 0.05	$2.143 \pm 0.803$	P < 0.05	$1.464 \pm 0.693$	P < 0.05
Negative	67	$2.164 \pm 0.665$		$1.701 \pm 0.853$		$1.119 \pm 0.769$	
Tumor size							
≤ 20 mm	22	$2.136 \pm 0.640$	NS	$1.864 \pm 0.774$	NS	$1.273 \pm 0.767$	NS
> 20 mm or < 50 mm	50	$2.400 \pm 0.670$		$1.800 \pm 0.948$		$1.180 \pm 0.800$	
≥ 50 mm	23	$2.087 \pm 0.733$		$1.783 \pm 0.723$		$1.217 \pm 0.736$	

Table 1. Relationship between DNMTs and tissue specimens

n, number of patients.

Fc-inf, infiltration to the capsule; NS, not significant.

remains to be determined. Interactions among DNMTs and methyl CpG-binding domain proteins should also be considered as a mechanism for maintaining DNA methylation patterns using the proofreading or remethylation function (Hattori et al., 2004).

Aberrant hypermethylation of CpG islands has recently been implicated in hepatocarcinogenesis, and it has been reported that tumor suppressor genes in hepatocellular carcinomas (HCCs) are affected by such silencing (Saito et al., 2001; Yang et al., 2003). To study the significance of aberrant DNMT (DNMT1, 3a and 3b) expression in human HCC, we evaluated the correlation between DNMT protein expression and clinicopathologic features and prognosis in patients with HCC.

## **Materials and Methods**

#### Patients and samples

Stored HCC and corresponding non-cancerous tissue specimens from our laboratory were obtained for 95 patients from 1989 to 2002. The protocol was approved by the ethics committee of the School of Medicine, Tottori University (1040). Informed consent was obtained from all patients.

The clinicopathologic features of the 95 patients are shown in Table 1. A mean age was 61  $\pm$  10 years (range, 17–80 years). For comparison, normal liver tissues were also obtained from 3 patients who had undergone surgical resection for hepaticolithiasis.

## Immunohistochemistry

Formalin-fixed, paraffin-embedded tissues were cut into 4-µm-thick continuous sections and mounted on poly-L-lysine-coated glass slides. Slides were deparaffinized in xylene, rehydrated in graded alcohols and washed in tap water. Endogenous peroxidase activity was blocked by incubating sections in 3% H<sub>2</sub>O<sub>2</sub>. Slides were heated in a microwave at 600 W for 15 min with 10 mM sodium citrate buffer (pH 6.0) and treated with polyoxyethylene sorbitan monolaurate (Tween 20) for 30 min for antigen retrieval. Rabbit polyclonal antibodies for DNMT1 (H-300, sc-20701, dilution 1:200), DNMT3a (H-295, sc-20703, dilution 1:200) and DNMT3b (H-230, sc-20704, dilution 1:200) (Santa Cruz Biotechnology, Santa Cruz, CA) were applied overnight, followed by incubation with biotinylated secondary antibodies [anti-rabbit IgG, N-Histofine, Simple Stain MAX PO (MULTI)] at room temperature for 30 min. Antigen-antibody complexes were visualized using a streptavidin-horseradish peroxidase conjugate (LSAB kit; Dako, Carpinteria, CA) with diaminobenzidine as a chromogen. Slides were counterstained with Harris' hematoxylin and methylgreen for 3 to 5 s.

## Scoring of immunoreactivity

Two independent observers (T.M. and K.E.) independently evaluated the immunostaining without knowledge of the clinical outcomes; the concordance ratio was > 90%. When differences in scoring were encountered, a consensus of two was used to obtain a final decision.

Nuclear positivities for the 3 proteins were evaluated using the scoring system previously reported by Choi et al. (2003), according to staining intensity (0, negative; 1, weak; 2, moderate; and 3, severe) and proportion of positive cells (0, negative; 1, positive in  $\leq 10\%$ ; 2, positive in > 10% and  $\leq$ 33%; 3, positive in > 33% and  $\leq 66\%$ ; 4, positive in > 66% of cells). The 2 scores were added in each case, and the expression was graded as - (0), 1+ (1 or 2), 2+ (3 to 5) and 3+ (6 or 7).

## Statistical analysis

Statistical analysis was performed using Prism software (GraphPad Software, La Jolla, CA). Spearman's rank correlation test and Student's t-test were used to study the relationships between DNMT immunoreactivities and various clinicopathologic parameters. Overall survival rates were determined by the Kaplan-Meier method, and the significance of differences between survival rates was studied using the log-rank test. Differences or correlations with P values < 0.05 were considered significant. Positive DNMT expression was defined as moderate (2+) or strong (3+), and negative DNMT expression as absent (-) or weak (1+) expression. To further evaluate the correlation between DNMT protein expression and patient survival, we subdivided the patients into 3 groups: Group 1) DNMT1(+), 3a(-) and 3b(-); Group 2) DNMT1(+), 3a or 3b(+); and Group 3) DNMT1(+), 3a(+) and3b (+).

#### Results

The immunohistochemical staining results are shown in Table 2 and Fig. 1. The non-cancerous liver tissues showed no expression of nuclear DNMT proteins by immunohistochemistry. There were no differences in cytoplasmic DNMT expres-

 
 Table 2. Expression of DNMT 1, 3a and 3b proteins in patients with hepatocellular carcinoma

	DNMT protein expression				
		1+	2+	3+	
	n (%)	n (%)	n (%)	n (%)	
DNMT1	0 (0)	10 (10.5)	50 (52.6)	35 (36.8)	
DNMT3a	8 (8.4)	20 (21.1)	48 (50.5)	19 (20.0)	
DNMT3b	14 (14.7)	51 (53.7)	25 (26.3)	5 (5.2)	

n, number of patients: total number, 95.

Nuclear positivities for the 3 proteins were evaluated using the scoring system, according to staining intensity (0, negative; 1, weak; 2, moderate; and 3, severe) and proportion of positive cells (0, negative; 1, positive in  $\le 10\%$ ; 2, positive in > 10% and  $\le 33\%$ ; 3, positive in > 33% and  $\le 66\%$ ; 4, positive in > 66% of cells). The 2 scores are added in each case, and the expression is graded as – (0), 1+ (1 or 2), 2+ (3 to 5) and 3+ (6 or 7).



**Fig. 1.** Immunohistochemical staining results for (**a**) DNMT1, (**b**) DNMT3a and (**c**) DNMT3b proteins in hepatocellular carcinomas. Strong nuclear expression and faint cytoplasmic expression are seen in these 3 cases. **a** and **b**: counterstained with Harris' hematoxylin. **c**: conterstained with methylgreen. Bar =  $100 \mu m$ .

**Fig. 2.** Kaplan-Meier survival curves for patients with hepatocellular carcinomas according to DNMT protein expression levels. [], number of patients.

a:	DNMT 1.
b:	DNMT 3a.
c:	DNMT 3b.



**Fig. 3.** Kaplan-Meier survival curves were plotted for 94 patients according to the correlation with expression of the DNMT protein complex. [], number of patients.

Group 1: DNMT1(+), 3a(-) and 3b(-). Group 2: DNMT1(+), 3a or 3b(+). Group 3: DNMT1(+), 3a(+) and 3b(+).

hand, no significant correlations were noted between DNMT1 or 3b and portal vein involvement. There were no significant associations between DNMT expression and age, sex, tumor size, infiltration to the capsule or underlying liver disease.

Follow-up information was available for all 95 patients, covering periods ranging from 0 to 183 months (mean, 63.7 months). Kaplan-Meier survival curves were plotted for the 95 patients according to expression of each DNMT.

The overall survival rates of DNMT3apositive and DNMT3b-positive HCC patients were significantly lower than those of patients negative for these proteins (Fig. 2b, P = 0.003, log-rank test; Fig. 2c, P = 0.0034, log-rank test). However, the overall survival rate of DNMT1-positive HCC patients was not significantly different from that of DNMT1-negative HCC cases (Fig. 2a, P = 0.1038, log-rank test).

The overall survival rate of patients in Group 3 [DNMT1(+), 3a(+) and 3b(+)] was significantly lower than those of patients in Groups 1 and 2 (Fig. 3, P = 0.0009, log-rank test).

## Discussion

DNA methylation is well known to be involved in the early developmental stages of HCC with a background of chronic liver disease, including CH and LC. It has recently been suggested that aberrant DNA methylation of CpG islands is an early and frequent event, and that stepwise progression of methylating events contributes to multistep hepatocarcinogenesis (Roncalli et al., 2002; Lee et al., 2003; Yang et al., 2003). The levels of DNMT1, DNMT3a and DNMT3b mRNA in HCCs are significantly higher than in noncancerous liver tissues, including normal, chronic CH and LC tissues (Saito et al., 2001; Oh et al., 2007).

In the present study, we used an immunohistochemical technique to directly examine the protein expression of 3 DNMTs in HCCs. The results demonstrated that DNMT protein expression in HCCs was significantly higher than in noncancerous liver tissues including normal, CH and LC tissues. In addition, there was no significant relationship between DNMT expression, CH and LC.

Immunohistochemical staining showed very strong nuclear expression and faint cytoplasmic expression of DNMT1, and strong nuclear and faint cytoplasmic expression of DNMT3a and DNMT3b. Some immunohistochemical staining of these DNMTs has previously been performed in a variety of cancers (Girault et al., 2003; Lin et al., 2007; Ding et al., 2008) and HCC (Saito et al., 2003; Oh et al., 2007). Although nuclear and cytoplasmic DNMT3b expression has been detected in other cancers, no results of immunohistochemical staining of DNMT3b have previously been reported in HCC.

The present study showed a significant relationship between DNMT3a protein expression and portal vein involvement, and significant relationships between DNMT protein expression and tumor differentiation or intrahepatic metastasis. However, a previous report found no significant relationship between DNMT mRNA levels and portal vein involvement and tumor differentiation in HCCs (Saito et al., 2001), but the same research group reported a significant relationship between DNMT1 protein expression and portal vein involvement or tumor differentiation (Saito et al., 2003). Reverse transcriptase-polymerase chain reaction is sufficiently sensitive to detect small elevations in DNMT mRNA levels, while immunohistochemistry is only able to detect elevations in protein expression when they reach a certain level in more malignant HCCs. We suggest that DNMT protein expression may be more strongly correlated with progression of the cancerous stage of HCCs, rather than the precancerous stage. To the best of our knowledge, there have been no reports of any significant relationship between DNMT protein expression and intrahepatic metastasis, and there have been some reports of a lack of any significant relationship between DNMT protein expression and lymphogenous metastasis in other cancers (Ding et al., 2008; Nosho et al., 2009). However, the relationship between DNMT protein expression and hematogenous metastasis has not been previously reported. E-cadherin, an intercellular adhesion-related protein, is coded for by a tumor suppressor gene. E-cadherin gene methylation has been associated with reduced expression of the protein, resulting in a loss of intercellular adhesion and tissue destruction in HCCs (Kanai et al., 1997). In a recent study, E-cadherin expression was reported to be significantly increased after concomitant siRNA-mediated depletion of DNMT1, 3a and 3b in endometrial epithelial carcinoma cells (Rahnama et al., 2009). DNMTs are thought to be involved in E-cadherin methylation. In addition, reduction of E-cadherin may also be related to the acquisition of cell motility, tumor cell invasion and vascular invasion. E-cadherin gene methylation in HCCs has been shown to be associated with a poor prognosis (Kozyraki et al., 1996; Lee et al., 2003). DNMT

protein expression may therefore contribute to hematogenous metastasis in HCC.

We investigated the relationship between expression of the 3 DNMTs and overall patient survival. This is the 1st report to reveal poor overall survival in HCC patients with positive immunoreactivity for DNMT3a and DNMT3b protein expression. No significant relationship between DNMT1 protein expression and overall patient survival was noted, though some studies on HCC and lung cancers have shown a significant relationship between DNMT1 protein expression and overall survival (Saito et al., 2003; Lin et al., 2007). The reasons for this discrepancy remain to be determined. DNMT1 is referred to as a "maintenance" methyltransferase, and DNMT1 protein expression may thus not contribute to overall survival. However, our study demonstrated a significant relationship between protein expression of the 3 DNMTs and poor overall survival in HCCs. It was recently reported that DNMT1 and DNMT3a/3b may associate directly and function as a complex for DNA methylation of CpG islands (Kim et al., 2002; Rhee et al., 2002; Hattori et al., 2004). The results of the present study suggest that DNMT1, DNMT3a and DNMT3b may be cooperatively involved in determining the extent of HCCs. Thus each DNMT could affect survival in patients with HCC.

DNA methylation is known to be a reversible process. In a previous study, promoter methylation by DNMT was shown to be reversible using the DNMT inhibitor 5-aza-2'-deoxycytidine, which induced re-expression of promoter-methylated genes in a cell line in vitro (Juttermann et al., 1994; Yang et al., 2001; Arai et al., 2006). 5-Aza-2'-deoxycytidine and 5-azacytidine in non-toxic doses have been approved by the United States Food and Drug Administration as DNMT inhibitors for hematological malignancies (Mulero-Navarro and Esteller, 2008). The relationship between DNMT protein expression and patient prognosis demonstrated in the present study suggests a potentially important role for DNMT inhibitor therapy in HCC patients with an increased DNMT protein expression.

In conclusion, the present study suggests that overexpression of DNMT proteins in HCCs may be a predictive factor for poor survival. In addition, DNMTs may represent new prognostic markers and potential therapeutic targets in HCC.

#### References

- 1 Arai M, Yokosuka O, Hirasawa Y, Fukai K, et al. Sequential gene expression changes in cancer cell lines after treatment with the demethylation agent 5-aza-2'deoxycytidine. Cancer 2006;106:2514–2525.
- 2 Choi MS, Shim YH, Hwa JY, Lee SK, et al. Expression of DNA methyltransferases in multistep hepatocarcinogenesis. Hum Pathol 2003;34:11–17.
- 3 Ding WG, Fang JY, Chen XY, Peng YS. The expression and clinical significance of DNA methyltransferase proteins in human gastric cancer. Dig Dis Sci 2008;53:2083–2089.
- 4 Girault I, Tozlu S, Lidereau R, Bièche I. Expression analysis of DNA methyltransferases 1, 3A, and 3B in sporadic breast carcinomas. Clin Cancer Res 2003;9:4415–4422.
- 5 Hattori N, Abe T, Hattori N, Suzuki M, et al. Preference of DNA methyltransferases for CpG islands in mouse embryonic stem cells. Genome Res 2004;14:1733–1740.
- 6 Juttermann R, Li E, Jaenische R. Toxicity of 5-aza-2'deoxycytidine to mammalian cells is mediated primarily by covalent trapping of DNA methyltransferase rather than DNA demethylation. Proc Natl Acad Sci USA 1994;91:11797–11801.
- 7 Kanai Y, Ushijima S, Hui AM, Ochiai A, et al. The Ecadherin gene is silenced by CpG methylation in human hepatocellular carcinomas. Int J Cancer 1997;71:355– 359.
- 8 Kim GD, Ni J, Kelesoglu N, Roberts RJ, et al. Cooperation and communication between the human maintenance and de novo DNA (cytosine-5) methyltransferases. EMBO J 2002;21:4183–4195.
- 9 Kozyraki R, Scoazec JY, Flejou JF, D'Errico A, et al. Expression of cadherins and alpha-catenin in primary epithelial tumors of the liver. Gastroenterology 1996;110:1137–1149.
- 10 Lee S, Lee HJ, Kim JH, Lee HS, et al. Aberrant CpG island hypermethylation along multistep hepatocarcinogenesis. Am J Pathol 2003;163:1371–1378.
- 11 Lin RK, Hsu HS, Chang JW, Chen CY, et al. Alteration of DNA methyltransferases contributes to 5'CpG methy-

lation and poor prognosis in lung cancer. Lung Cancer 2007;55:205–213.

- 12 Łuczak MW, Jagodzinski PP. The role of DNA methylation in cancer development. Folia Histochem Cytobiol 2006;44:143–154.
- 13 Mulero-Navarro S, Esteller M. Epigenetic biomarkers for human cancer: The time is now. Crit Rev Oncol Hematol 2008;68:1–11.
- 14 Nosho K, Shima K, Irahara N, Kure S, et al. DNMT3B expression might contribute to CpG island methylator phenotype in colorectal cancer. Clin Cancer Res 2009;15:3663–3671.
- 15 Oh BK, Kim H, Park HJ, Shim YH, et al. DNA methyltransferase expression and DNA methylation in human hepatocellular carcinoma and their clinicopathological correlation. Int J Mol Med 2007;20:65–73.
- 16 Rahnama F, Thompson B, Steiner M, Shafiei F, et al. Epigenetic regulation of E-cadherin controls endometrial receptivity. Endocrinology 2009;150:1466–1472.
- 17 Rhee I, Bachman KE, Park BH, Jair KW, et al. DNMT1 and DNMT3b cooperate to silence genes in human cancer cells. Nature 2002;416:552–556.
- 18 Roncalli M, Bianchi P, Bruni B, Laghi L, et al. Methylation framework of cell cycle gene inhibitors in cirrhosis and associated hepatocellular carcinoma. Hepatology 2002;36:427–432.
- 19 Saito Y, Kanai Y, Nakagawa T, Sakamoto M, et al. Increased protein expression of DNA methyltransferase (DNMT) 1 is significantly correlated with the malignant potential and poor prognosis of human hepatocellular carcinomas. Int J Cancer 2003;105:527–532.
- 20 Saito Y, Kanai Y, Sakamoto M, Saito H, et al. Expression of mRNA for DNA methyltransferases and methyl-CpG-binding proteins and DNA methylation status on CpG islands and pericentromeric satellite regions during human hepatocarcinogenesis. Hepatology 2001;33: 561–568.
- 21 Yang B, Guo M, Herman JG, Clark DP. Aberrant promoter methylation profiles of tumor suppressor genes in hepatocellular carcinoma. Am J Pathol 2003;163:1101– 1107.
- 22 Yang X, Phillips DL, Ferguson AT, Nelson W, et al. Synergistic activation of functional estrogen receptor (ER)-alpha by DNA methyltransferase and histone deacetylase inhibition in human ER-alpha-negative breast cancer cells. Cancer Res 2001;61:7025–7029.

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