

This is a repository copy of *Loss of Janus Associated Kinase1 alters urothelial cell function and facilitates the development of bladder cancer*.

White Rose Research Online URL for this paper:
<https://eprints.whiterose.ac.uk/150059/>

Version: Accepted Version

Article:

Daza-Cajigal, Vanessa, Albuquerque, Adriana, Pearson, Joanna Frances et al. (7 more authors) (2019) Loss of Janus Associated Kinase1 alters urothelial cell function and facilitates the development of bladder cancer. *Frontiers in immunology*. ISSN 1664-3224

<https://doi.org/10.3389/fimmu.2019.02065>

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.

Loss of Janus Associated Kinase1 alters urothelial cell function and facilitates the development of bladder cancer

Vanessa Daza-Cajigal^{1,2,3,4,5}, Adriana Albuquerque¹, Joanna Pearson⁶, Jennifer Hinley⁶, Andrew S Mason⁶, Jens Stahlschmidt^{6,7}, Adrian J. Thrasher^{8,9}, Vibhash Mishra¹⁰, Jennifer Southgate⁶, Siobhan O. Burns^{1,2*}.

¹University College London Institute of Immunity and Transplantation, London, UK

²Department of Immunology, Royal Free London NHS Foundation Trust, London, UK

³Universidad Complutense, School of Medicine, Madrid, Spain

⁴Department of Immunology, Hospital Universitario Son Espases, Palma, Spain

⁵Institut d'Investigació Sanitaria de Palma (IdISPa), Palma, Spain

⁶Jack Birch Unit, York Biomedical Research Institute, Department of Biology, University of York, York, UK

⁷Department of Histopathology, St James's University Hospital, Leeds, UK

⁸Great Ormond Street Hospital for Children NHS Foundation Trust, London, UK

⁹University College London Institute of Child Health, London, UK

¹⁰Department of Urology, Royal Free London NHS Foundation Trust, London, UK.

* Correspondence:

Dr. Siobhan O Burns

siobhan.burns@ucl.ac.uk

Keywords: Bladder cancer, IFN γ signalling, immunodeficiency, JAK1, urothelium.

Word count: 3850

Total number of figures and tables: 6

Abstract

Inherited Primary Immunodeficiency (PID) disorders are associated with increased risk of malignancy that may relate to impaired antitumor immune responses or a direct role for PID germline mutations in tumorigenesis. We recently identified germline loss of function mutations in Janus Associated Kinase 1 (*JAK1*) causing primary immunodeficiency characterised by infections and associated with early onset, fatal high-grade bladder carcinoma. Somatic mutations in *JAK1*, required for immune cell signalling in response to interferon gamma ($\text{IFN}\gamma$), have been associated with several non-hematopoietic and hematopoietic cancer cell types but pathogenic mechanisms remain largely unexplored. Here we demonstrate that JAK1 is required for the intrinsic $\text{IFN}\gamma$ response of urothelial cells impacting immunogenicity and cell survival. Specifically, JAK1-deficient urothelial cells showed reduced surface expression of major histocompatibility complex class II (MHC II), intercellular adhesion molecule-1 (ICAM-1) and programmed death-ligand-1 (PD-L1) after $\text{IFN}\gamma$ stimulation and were resistant to $\text{IFN}\gamma$ -induced apoptosis and lymphocyte-mediated killing. In addition, we identify a previously unknown role for $\text{IFN}\gamma$ signalling in modulating urothelial differentiation. Together, our findings support a role for urothelial cell JAK1 in immune surveillance and development of bladder cancer. Our results have implications for patients with rare JAK1 PID and, more broadly, inform development of biomarker and targeted therapies for urothelial carcinoma.

Introduction

Widespread availability of next generation sequencing (NGS) has transformed the field of cancer, identifying candidate genes to enhance our understanding of tumorigenesis and enable stratification of patients for specific therapies. In the field of immunology, NGS has identified new monogenic forms of primary immunodeficiencies (PID): inherited disorders predisposing to infection, autoimmunity and malignancy. Genes identified to cause PID overlap with cancer susceptibility genes, so that clarifying the pathogenesis of PID has impact for oncology. We recently identified a new PID associated with compound homozygous loss of function mutations in the gene coding the signalling protein Janus-Associated Kinase 1 (JAK1) resulting in partial JAK1 deficiency (1). The clinical phenotype was characterised by immunodeficiency plus aggressive urothelial carcinoma that was fatal in the third decade of life, suggesting that impaired JAK1 function may be a predisposing factor for urothelial carcinoma.

JAK1 belongs to a family of widely-expressed tyrosine kinases essential for signal transduction from multiple cytokine receptors through activators of transcription (STAT) proteins(2,3). Individual family members (JAK1, JAK2, JAK3 and Tyrosine kinase 2) associate with selected receptors and have non-redundant roles in cell signalling. Multiple receptors utilise JAK1 for signal transduction including members of the IL-2 receptor family (IL-2R, IL-7R, IL-9R and IL-15R), the IL-4 receptor family (IL-4R, IL-13R), the gp130 receptor family (IL-6R, IL-11R, LIF-R, OSM-R, CT-1R, CNTF-R, NNT-1R/BSF- 3R and Leptin-R) and class II cytokine receptors (type I IFN-R, type II IFN-R, IL-10R (2). Elucidating the non-redundant roles of JAK1 in cell biology has been complicated by the fact that complete JAK1-deficiency results in perinatal lethality as a result of neurological defects in murine models (3)precluding detailed examination of immune competence or malignancy risk.

Activation of the JAK1-dependent interferon gamma (IFN γ) signalling pathway is known to have direct effects on tumour cells, impacting numerous cell programmes including growth, apoptosis, proliferation, differentiation and migration (4–8). In addition, IFN γ induces immunoregulatory functions in cancer cells such as cytokine production, antigen presentation by major histocompatibility complexes (MHC) (9), expression of adhesion molecules such as intercellular adhesion molecule-1 (ICAM-1) (10) and ligands for receptors of immune checkpoints such as the programmed death 1/programmed death-ligand 1 (PD-1/PD-L1) (11). In mouse models, loss of IFN γ signalling accelerated tumour initiation and progression, demonstrating a direct impact of IFN γ signalling on tumorigenesis, the tumour microenvironment and metastatic dissemination (5,12).

To date, bladder urothelial carcinoma has not been described as a feature of other loss of function JAK-STAT or IFN γ -related PID, although different types of tumours have been reported in these conditions, including disseminated cutaneous squamous cell carcinoma (13–16). Somatic mutations in *JAK1* have been identified in multiple tumour types including high-risk bladder cancer, endometrial, colorectal, stomach, and prostate carcinomas (17–19) supporting the idea that alterations in JAK1 signalling, whether through loss or gain of function, could play a role in the pathogenesis of some epithelial cancers. Somatic mutations predicted to cause loss of JAK1 function are associated with reduced expression of IFN-associated genes in different tumour types (19). Although the cellular mechanisms have not been fully elucidated, truncating somatic *JAK1* mutations in gynaecological carcinomas reduced IFN γ -induced MHC class I expression at the tumour cell surface, which could reduce immune recognition and facilitate immune evasion (17).

Here we examine the role of JAK1 in human urothelial cells and demonstrate a requirement of JAK1 for multiple functions including cell survival and interaction with immune cells. We also identify a potential role for JAK1 in modulating urothelial differentiation phenotype through IFN γ -

signalling. Our findings provide new insight into immune and intrinsic signalling regulation of urothelial cells and support a role for JAK1 in the pathogenesis of bladder cancer.

Materials and methods

The Cancer Genome Atlas Analysis

Somatic variants for the muscle invasive bladder cancer (BLCA) cohort from The Cancer Genome Atlas (TCGA) were accessed from the Genome Data Commons(20) as part of dbGaP project 19625. Variants which overlapped the predominant JAK1 transcript (ENST00000342505.5) were extracted and their impact assessed using the Ensembl Variant Effect Predictor (21) including SIFT 4G (22) and Poly-Phen 2 (23), with additional analysis of deleterious effects using HMMvar v1.1.0(24). Mutations and their effects were presented against the JAK1 protein sequence (InterPro P23458).

Ethical approval

Blood samples and biopsies were obtained with ethical approval (National Research ethics numbers 08/H0720/46, 99/095 and 02/208) and informed consent from all subjects in accordance with the Declaration of Helsinki.

Immortalised and normal human urothelial cell cultures

An immortalized normal human urothelial (NHU) subline produced by retroviral transduction with human telomerase reverse transcriptase (hTERT) cells as detailed elsewhere (25), was used in this study. The subline, referred to as Y235hTERT, was previously characterized at passage 40 against the pre-immortalised parental line (passage 7) using comparative genomic hybridization. The Y235hTERT cell lines for this study were cultured using protocols detailed in full elsewhere (26) and used within 20 passages of the CGH analysis. JAK1 knock down (KD) and scrambled control (Sc) hTERT urothelial sub-lines were generated using lentiviral vectors expressing short hairpin RNA (shRNA) sequences and utilised for all experiments unless otherwise indicated. **shRNA**

technology permitted generation of cell lines with sub-total deficiency similar to the effect of loss of function mutations seen in our patient (1).

For differentiation studies only, normal human urothelial (NHU) cells obtained ethically with appropriate informed consent and Research Ethics Committee approvals were maintained in vitro as non-immortalized (finite) cells. For routine culture, NHU cells were grown as adherent monolayers on Primaria™ plasticware (BD Biosciences) in low calcium (0.09 mM) keratinocyte serum-free medium (KSFM) containing bovine pituitary extract and recombinant epidermal growth factor (Life Technologies) supplemented with 30 ng/ml cholera toxin (KSFMc). NHU cells were sub-cultured by trypsinisation at just-confluence and used in experiments between passages 3-5. Differentiation experiments described here were performed on five independent NHU cell cultures from five different donors. Due to the finite nature of these lines, no genotyping of individual cell cultures was performed. Differentiation of NHU cells was induced in just-confluent cell cultures using 1 μ M troglitazone (TZ) as peroxisome proliferator-activated receptor-gamma (PPAR γ) activating ligand with concurrent 1 μ M PD153035 to block epidermal growth factor receptor (EGFR) activation, as previously described (27).

All Y235hTERT cell lines and NHU cell cultures were tested regularly for contamination by *Mycoplasma spp.* using polymerase chain reaction-based kits and DNA-intercalating fluorescent stains for presence of extranuclear DNA. For all stimulation experiments described, concentrations of IFN γ were optimised for each experiment.

Lentivirus preparation and transductions

pGIPZ vectors carrying the short hairpin RNA against JAK1(TAGTACACACATTTCCATG) or scrambled control (TGAACTCATTTTTCTGCTC) sequences as well as puromycin resistance cassette and turbo-GFP marker for selection were supplied by University College London Open Biosystems (London UK). Lentivirus stocks were prepared by transfection of 293T cells (80-90%

confluence) cultured in DMEM medium and 10% heat-inactivated fetal bovine serum, with the envelope plasmid 17.5 µg pMD.G2 (VSV-G/envelop), 32.5 µg p8.74 plasmid (gag-pol) and 25 µg vector construct with the transfection reagent PEI/Opti-MEM™ following the manufacturer's instructions (Promega). Medium was replaced 5h post transfection and medium was harvested after 24 and 48 hours, cleared by centrifugation (4000 rpm, 5 min), filtered through 0.22µm filters and left to spin for 2h 4°C 50,000g. Viruses were titrated on 293T cells by scoring GFP-positive cells (flow cytometry) 3 days post transduction. Virus stocks were stored at -80°C. Transductions of urothelial cells were carried out by infection at a multiplicity of infection of 1:10 for 6h, before replacing virus-containing medium with fresh medium. Cells were selected in puromycin-containing medium and the efficiency of transduction was assessed by percentage of GFP-positive cells. Loss of JAK1 expression in JAK1-deficient hTERT urothelial cells was verified by RT-PCR.

Determination of mRNA levels by real time-quantitative polymerase chain reaction (RT-qPCR) and reverse transcription polymerase chain reaction (RT-PCR)

Cells were left unstimulated or stimulated with 1 ng/ml of IFN γ for different time points. Total RNA from hTERT urothelial cells was extracted using RNeasy kit (Qiagen). RNA from NHU cells was extracted in TRIzol® reagent (Thermo Fisher Scientific, Loughborough, UK); any contaminating DNA was digested using a DNA-free kit (Thermo Fisher Scientific). RNAs were converted to cDNA by reverse-transcription using QuantiTect reverse transcription kit (Qiagen).

Determination of mRNA abundance was performed by RT-PCR using specific primers (Table S1) and QuantiTect SYBR® Green PCR Kit (Qiagen) according to manufacturer's instructions. Fold changes were calculated using the DDCT2 (-Delta Delta C(T)) method and results were normalised with respect to the values obtained for the endogenous GAPDH cDNA.

Flow Cytometry analysis

Sc and KDhTERT urothelial cells, \pm addition of 5 ng/ml of IFN γ for different time points, were detached using Accutase® solution (A6964, Sigma Aldrich), labelled with fluorescent-conjugated antibodies (see Table S2) and washed with phosphate buffered saline (PBS). For STAT1 phosphorylation analysis, cells \pm 1 ng/ml IFN γ for 10 min were fixed and permeabilised using fix buffer I and Perm Buffer III (BD Biosciences) for 30 min at 4°C, washed with PBS and labelled with 5 μ L anti-pSTAT1 antibody (BD Biosciences) for 60min in the dark. For all flow cytometry (BD LSRFortessa) experiments 10,000-30,000 gated events were collected and analysed using FlowJo software.

Cell viability and Apoptosis assays

Sc and KD hTERT urothelial cells were stimulated with IFN γ using different time points and concentrations. Alamar Blue® (AB), diluted 1:10 with KSFMc, was added to urothelial cells grown in 96-well plates (4,000 cells/well). After 3 hours incubation at 37°C, absorbance was measured at 560 and 620 nm. AB reduction was calculated according to manufacturer's instructions (AbD Serotec, Kidlington, UK). Apoptosis was determined by flow cytometry using APC Annexin V apoptosis detection kit with Propidium Iodide (PI) according to manufacturer's instructions (BioLegend 640932).

Lymphocyte Cytotoxicity Assay

Healthy donor peripheral blood mononuclear cells (PBMCs) were isolated by Ficoll™ gradient and frozen in 10% dimethyl sulfoxide (DMSO). To control for differences in the frequency of lymphocyte subsets between donors, PBMCs collected from a single donor buffy coat were stored cryopreserved and used for all killing assays shown. PBMC were thawed and re-suspended in

RPMI (Invitrogen) with 10% heat-inactivated fetal bovine serum and monocytes removed by plastic adherence (1h, 37 °C). Urothelial cells were cultured in 96-well plates \pm IFN γ (5 ng/ml) for 30h and subsequently co-cultured overnight with interleukin-2 (IL-2)25U/ml (Roche) + monocyte-depleted PBMCs (50:1). Cells were detached using Accutase® solution, washed with PBS and resuspended in 200 μ l DNA staining solution which specifically stains nuclei in dead cells and allows identification of necrotic cells (NKTEST, Glycotope Biotechnology). Urothelial cells were gated based on green fluorescent protein (GFP) expression and analysed by flow cytometry.

Immunohistochemistry

5 μ M formalin-fixed tissue sections were dewaxed in xylene, then rehydrated through ethanol into water. Antibodies are detailed in Table S3. Following appropriate antigen retrieval (Table S3), sections were incubated with optimally titrated primary antibodies and immunodetected using secondary antibodies linked to streptavidin-biotin horseradish peroxidase complex (DAKO), ImPRESS™ Excel Polymer system (Vector labs), or BenchMark Ultra automated staining system (Ventana) (Table S3). All slides were counterstained in Mayer's haematoxylin and mounted in DPX (Sigma). Control slides and tissues were included to check specificity.

Statistical analysis

Statistical analysis was performed using Graphpad Prism 5.1 Software. Associations between JAK1-deficient and control cells were tested using one-way ANOVA and appropriate post-test, or a two-tailed Mann Whitney U test. A p value of <0.05 was considered significant.

Results

Partial JAK1 deficiency impairs STAT1 phosphorylation and expression of IFN γ -inducible genes in hTERT urothelial cells

While our patient remains the only reported individual with partial JAK1 deficiency caused by germline loss of function mutations, somatic variants in *JAK1* have been described in other patients with very high-risk bladder cancer(18). Analysis of the muscle invasive bladder cancer (BLCA) cohort from The Cancer Genome Atlas (TCGA) (20) identified 31 different *JAK1* variants in 25/412 TCGA BLCA samples, including single nucleotide variants predicted to alter JAK1 protein function and nonsense/frameshift variants predicted to impair JAK1 signalling (Figure 1a).

To model and test the impact of loss of JAK1 function in urothelial cells, we generated a JAK1 KD immortalised hTERT cell line using lentiviral vectors expressing shRNA sequences. Compared to control Sc shRNA, JAK1 shRNA substantially reduced *JAK1* mRNA expression (Fig. S1a). To confirm functional knock down of JAK1, we studied JAK1-mediated activation of STAT1 proteins in response to IFN γ stimulation, using flow cytometry. We observed a significant decrease in STAT1 phosphorylation following IFN γ stimulation in the KD cell line compared to untransduced and Sc shRNA lines, ($p < 0.05$) (Fig. 1b,c and S1b). The effect was similar to the reduced, but not abolished, STAT1 phosphorylation observed in fibroblasts of the patient with JAK1 deficiency following IFN γ stimulation (Fig. S1c). Expression of mRNA for the interferon inducible transcription factor IRF1 was also significantly lower in KD than Sc cell lines following stimulation with IFN γ , indicating impaired downstream gene regulation in JAK1 deficiency (Fig. 1d). Reduced responses to IFN γ were not due to alteration in expression of the IFN γ receptor (IFN γ R) as Sc and KD hTERT urothelial cells displayed comparable IFN γ R surface expression both at baseline and following IFN γ stimulation (Fig. 1e,f and S1b).

JAK1 deficiency alters MHC, ICAM-1 and PD-L1 expression in hTERT urothelial cells

It was previously shown that IFNs regulate expression of MHC, ICAM-1 and PD-L1 in cancer cells (9–11). We examined the expression of these cell surface molecules in JAK1-deficient and Sc hTERT urothelial cell lines before and after IFN γ treatment (Fig. 2 and S2,3). While basal expression of MHCII was minimal in both KD and Sc cells, IFN γ induced expression of MHCII in control cells that was significantly lower in the KD cells ($p < 0.05$) (Fig. 2a,b, S2b, S3a). As described for gynecological cancer cells bearing somatic *JAK1* mutations (17), we also observed lower surface MHCI expression by JAK1-deficient hTERT urothelial cells compared with control cells at baseline. After IFN γ stimulation, however, MHCI was upregulated by both control and KD cells with no significant difference (Fig. 2c,d, S2c, S3b). Expression of ICAM-1 was reduced in KD cells at baseline and expressed at significantly lower levels after IFN γ stimulation compared with Sc control ($p < 0.05$, Fig. 2e,f, S2d, S3c). In keeping with the effects of other reported somatic mutations associated with IFN γ resistance in cancer cells (28,29), we observed reduced PD-L1 upregulation after IFN γ stimulation in KD cells compared to Sc ($p < 0.05$, Fig. 2g,h, S2e, S3d). Together these data indicate that JAK1 functions to regulate the expression of multiple immunomodulatory cell surface molecules in urothelial cells and that JAK1-deficiency alters urothelial immunomodulatory phenotype.

JAK1 deficiency impairs apoptosis in response to IFN γ in hTERT urothelial cells

JAK1 KD had no effect on basal growth kinetics of immortalised hTERT cell lines assessed using an Alamar Blue (AB) reduction assay. In both Sc and KD cell lines, the culture biomass increased steadily over 4 days, indicating a similar increase in cell number over time. IFN γ stimulation significantly inhibited the rise in population in the Sc cell line (Fig.3a; $p < 0.05$), in keeping with an anti-proliferative effect and/or increased cell death. By contrast, growth kinetics of the JAK1-deficient hTERT urothelial cell population showed no significant change following IFN γ treatment

(Fig. 3a). To further investigate this, we tested whether JAK1-deficient cells are resistant to IFN γ -induced apoptosis using Annexin V/PI staining. In control Sc hTERT urothelial cells, IFN γ induced cell death (Fig. 3b,e) which was mainly due to an increase in cells in early apoptosis identified as an Annexin V+/PI dim population (Fig. 3c,e), with minimal effect on late apoptosis/necrosis (Fig.3d,e). In contrast, there was no significant impact of IFN γ stimulation on any stage of cell death analysed in the KD cell line (Fig. 3 b-e), leading us to conclude that JAK1 is required for IFN γ -mediated apoptosis in urothelial cells. Together, our data support a requirement for JAK1 in regulating urothelial cell survival and homeostasis in response to IFN γ .

Defective lymphocyte-mediated killing of JAK1-deficient hTERT urothelial cells after IFN γ stimulation

We next sought to establish whether the reduced MHCI and ICAM-1 levels observed would impair immune cell recognition of JAK1-deficient hTERT urothelial cells (9,10,30,31). We tested this using a lymphocyte assay where killing can be mediated by NK or CD8+ T cells through multiple mechanisms including shared perforin-granzyme and death receptor/death ligand mechanisms. IFN γ pre-treatment of the Sc hTERT cell line resulted in significant enhancement in urothelial cell lysis by third party, IL-2 expanded primary lymphocytes (measured as necrotic cells, $p < 0.05$. Fig. 4 a,b). Importantly, lymphocyte mediated cell death was significantly reduced in KD compared to Sc cells following IFN γ pre-treatment ($p < 0.05$. Fig. 4c), suggesting that JAK1 deficiency confers resistance to immune cell killing.

Role of JAK1 in urothelial cytodifferentiation

Histological examination of the invasive urothelial carcinoma found in the JAK1-deficient patient revealed a heterogeneous tumour with distinct central and invasive regions (Fig. 5 and S4). The

tumour showed no infiltrating immune cells, with only infrequent cells identified in the stroma or vasculature (Fig. S5).

Muscle-invasive bladder cancers are classified as “luminal” (urothelial differentiated) or “basal-like” (squamous differentiated) subtypes based on signature markers (32). Using antibodies against urothelial differentiation markers, the tumour was negative for uroplakin 3a, whilst cytokeratin (CK) 20 expression was weak and confined to the central region. The squamous marker CK14 was present in a small patch of invasive cells but was otherwise absent. Transcriptional regulators of urothelial differentiation PPAR γ , GATA3, FOXA1 and IRF1 showed nuclear localisation, with weak variable expression of IRF1; pSTAT1 expression was cytoplasmic and weak in both central and invasive regions. Human epidermal growth factor receptor 2 (HER2) expression was variable within the central portion of the tumour, with some areas containing scant membrane and cytoplasmic expression, but negative in other areas, including the invasive front. These findings indicated the tumour to be fundamentally luminal in subtype, albeit with a suppressed urothelial differentiation phenotype. Given the dual role of IRF1 in both IFN γ signalling and PPAR γ -mediated urothelial differentiation, this led us to test experimentally whether JAK1 has a role in urothelial cell differentiation.

As the immortalised hTERT urothelial cell line is not suitable for differentiation studies, non-immortalised NHU cell cultures were generated from five individual donors. NHU cells were induced to differentiate by coactivation of PPAR γ and inhibition of EGFR signalling, using a combination of troglitazone and PD153035 (TZ/PD) (27). These conditions induce gene expression changes associated with urothelial differentiation via PPAR γ -dependent transcription of intermediary transcription factors, including FOXA1 and IRF1 (33). As *IRF1* knockdown limits uroplakin expression and *IRF1* induction was impaired in JAK1-deficient cells (Fig. 1b), we tested by RT-qPCR whether IFN γ modulates expression of genes associated with IFN γ signalling and urothelial cytodifferentiation pathways. As expected, TZ/PD induced up-regulation of *PPARG* and

FOXA1 (Fig. 6a,b). IFN γ alone had a small effect resulting in a weak up-regulation of *IRF1*, *PPARG* and the major histocompatibility class II transactivator (*CIITA*) (Fig. 6a,c,d). Surprisingly however, stimulation with IFN γ +TZ/PD up-regulated all four genes and substantially increased expression of *IRF1*, *FOXA1* and *CIITA* compared with TZ/PD alone (Fig. 6a-d). These data suggest that the combination of both IFN γ and TZ/PD significantly increases expression of transcription factors involved in urothelial cytodifferentiation and supports a previously unknown role for JAK1 in modulating urothelial phenotype.

Discussion

The fundamental importance of JAK1 signalling has been highlighted by perinatal lethality of JAK1 knock-out mice (3) and, in keeping with this, germline mutations causing complete loss of JAK1 function have not been described in humans. We recently reported the first case of germline loss of function mutations causing partial JAK1 deficiency in humans which resulted in immunodeficiency characterised by mycobacterial infection, suggesting a dominant effect on the IFN γ pathway, and high-grade bladder carcinoma (1). The early-onset and aggressive nature of the malignancy, along with the fact that *JAK1* is a hotspot for damaging somatic mutations in bladder carcinoma (28), led us to investigate whether impaired JAK1 function may be a specific predisposing factor for urothelial carcinoma. The data presented here demonstrate that JAK1 is important for multiple aspects of urothelial cell biology and highlight mechanisms by which loss of JAK1 function may promote tumorigenesis in this cell type.

Using a JAK1-deficient urothelial cell line model, we show that loss of JAK1 function impaired induction of apoptosis in response to IFN γ suggesting a role for JAK1 in regulating intrinsic urothelial cell homeostasis. In addition, JAK1-deficient urothelial cell lines demonstrated reduced surface expression levels of ICAM-1 following IFN γ stimulation, which was associated with resistance to lymphocyte-mediated cell lysis that is known to correlate with cell surface expression

of this molecule (10,34). While we did not observe a significant effect of JAK1 deficiency on IFN γ -mediated upregulation of MHC I in our model, JAK1 was shown to be required for expression of MHC II that can mediate tumour and self-antigen presentation in non-professional antigen presenting cells (35). This difference could be explained by the fact that MHC I is highly expressed constitutively on urothelial cells whereas MHC II expression, like ICAM-1, is induced de novo by IFN γ activity (Fig. 2) and hence demonstrates a highly specific directed response.

Together these data suggest that JAK1-deficient urothelial cells are less susceptible to IFN γ -mediated apoptosis, immune cell recognition and immune-mediated cell death. The absence of an immune infiltrate observed in the patient's tumour by immunohistochemistry supports our conclusion that loss of JAK1 in bladder cancer cells results in a poorly immunogenic tumour. IFN γ is of specific relevance in bladder cancer as it is a major cytokine released by tumour infiltrating lymphocytes thought to be important for anti-tumour responses and has potential for use as a biomarker of outcome in this condition (36–39). It has already been reported that damaging mutations in the IFN γ signalling pathway and antigen presentation pathway are associated with metastasis and higher resistance to the checkpoint blocking therapy with anti-PD-L1/PD-1 in a number of tumour types, including bladder cancer (29,30,40). Our data suggest loss of function *JAK1* mutations are a risk factor for lower tumour cell PD-L1 expression which could impair responsiveness to anti-PD-1 therapy used for advanced urothelial carcinoma (41). Further correlation of tumour genetics with clinical response to treatment is required to test this in practice.

In addition to impacting immune-related functions, we observed a potential role for JAK1 and IFN γ signalling in urothelial cell differentiation. It has been shown that NHU cells can be induced to differentiate using PPAR γ ligands and concurrent EGFR inhibition, for example using TZ/PD (27). In this study, we show that IFN γ had a significant effect on the induction of the transcription factors IRF1 and FOXA1, both known to be involved in urothelial cell differentiation induced by PPAR γ activation (33). In particular, IRF1 is a common downstream mediator for PPAR γ and IFN γ

signalling pathways influencing both urothelial differentiated phenotype and immune cell interactions. This suggests that IRF1 may be a potential novel target for modulating immunotherapy outcomes in urothelial cancer. Further research is needed to understand the mechanisms of IFN γ interaction with PPAR γ and EGFR signalling in urothelial cell differentiation and tumorigenesis.

In summary our findings highlight previously unknown roles for JAK1 in urothelial cell immune recognition and differentiation. Our data suggest that loss of JAK1 function through germline or somatic mutation promotes malignant transformation of urothelial cells, which are intrinsically less immunogenic.

Our results add further weight to arguments for sequencing urothelial cell tumours for clinical trials of immunotherapy agents to test whether the mutational burden of JAK1 and other IFN γ -related genes represent a biomarker for responsiveness to treatment in bladder cancer, which can more accurately predict the clinical outcome of these patients.

Acknowledgments:

We thank Louise Griffith of the Haematological Malignancy Diagnostic Service, St James's University Hospital, Leeds for the CD immunohistochemistry.

Conflict of interest disclosure statement

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Author contributions:

VDC, JS and SOB conceived, designed and coordinated the study. VDC, JP, JH performed the experiments. SOB, JS, and AJT provided clinical data and gave critical advice. VDC, JP, JH, JS and SOB analyzed the data and interpreted the results. ASM performed bioinformatic analysis. VDC and SOB wrote the manuscript. VM, JS and AJT gave administrative, technical, or material support. The final version of the manuscript was reviewed by all the coauthors.

Financial support

This work was supported by the Alfonso Martin Escudero Foundation (VD), Rosetrees Trust Foundation (VD, SB), the Wellcome Trust (104807/Z/14/Z) (AT) and a programme grant from York Against Cancer (JP, JH, ASM, JS). The project was sponsored by the University College London (UCL) and the National Institute for Health Research Biomedical Research Centre at Great Ormond Street Hospital for Children NHS Foundation Trust (AT).

References

1. Eletto D, Burns SO, Angulo I, Plagnol V, Gilmour KC, Henriquez F, Curtis J, Gaspar M, Nowak K, Daza-Cajigal V, et al. Biallelic JAK1 mutations in immunodeficient patient with mycobacterial infection. *Nat Commun* (2016) **7**:13992. doi:10.1038/ncomms13992
2. Kisseleva T, Bhattacharya S, Braunstein J, Schindler CW. Signaling through the JAK/STAT pathway, recent advances and future challenges. *Gene* (2002) **285**:1–24. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/12039028>
3. Rodig SJ, Meraz MA, White JM, Lampe PA, Riley JK, Arthur CD, King KL, Sheehan KC, Yin L, Pennica D, et al. Disruption of the Jak1 gene demonstrates obligatory and nonredundant roles of the Jaks in cytokine-induced biologic responses. *Cell* (1998) **93**:373–383. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/9590172>
4. Papageorgiou A, Lashinger L, Millikan R, Grossman HB, Benedict W, Dinney CPN, McConkey DJ. Role of tumor necrosis factor-related apoptosis-inducing ligand in interferon-induced apoptosis in human bladder cancer cells. *Cancer Res* (2004) **64**:8973–8979. doi:10.1158/0008-5472.CAN-04-1909
5. Parker BS, Rautela J, Hertzog PJ. Antitumour actions of interferons: implications for cancer therapy. *Nat Rev Cancer* (2016) **16**:131–144. doi:10.1038/nrc.2016.14
6. Sucker A, Zhao F, Pieper N, Heeke C, Maltaner R, Stadtler N, Real B, Bielefeld N, Howe S, Weide B, et al. Acquired IFN γ resistance impairs anti-tumor immunity and gives rise to T-cell-resistant melanoma lesions. *Nat Commun* (2017) **8**:15440. doi:10.1038/ncomms15440
7. Chawla-Sarkar M, Lindner DJ, Liu Y-F, Williams BR, Sen GC, Silverman RH, Borden EC. Apoptosis and interferons: role of interferon-stimulated genes as mediators of apoptosis. *Apoptosis* (2003) **8**:237–249. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/12766484>

8. Matsushita H, Hosoi A, Ueha S, Abe J, Fujieda N, Tomura M, Maekawa R, Matsushima K, Ohara O, Kakimi K. Cytotoxic T lymphocytes block tumor growth both by lytic activity and IFN γ -dependent cell-cycle arrest. *Cancer Immunol Res* (2015) **3**:26–36. doi:10.1158/2326-6066.CIR-14-0098
9. Propper DJ, Chao D, Braybrooke JP, Bahl P, Thavasu P, Balkwill F, Turley H, Dobbs N, Gatter K, Talbot DC, et al. Low-dose IFN-gamma induces tumor MHC expression in metastatic malignant melanoma. *Clin Cancer Res* (2003) **9**:84–92. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/12538455>
10. Wang R, Jaw JJ, Stutzman NC, Zou Z, Sun PD. Natural killer cell-produced IFN- γ and TNF- α induce target cell cytolysis through up-regulation of ICAM-1. *J Leukoc Biol* (2012) **91**:299–309. doi:10.1189/jlb.0611308
11. Eppihimer MJ, Gunn J, Freeman GJ, Greenfield EA, Chernova T, Erickson J, Leonard JP. Expression and regulation of the PD-L1 immunoinhibitory molecule on microvascular endothelial cells. *Microcirculation* (2002) **9**:133–145. doi:10.1038/sj/mn/7800123
12. Koromilas AE, Sexl V. The tumor suppressor function of STAT1 in breast cancer. *JAK-STAT* (2013) **2**:e23353. doi:10.4161/jkst.23353
13. Bax HI, Freeman AF, Anderson VL, Vesterhus P, Laerum D, Pittaluga S, Wilson WH, Holland SM. B-cell lymphoma in a patient with complete interferon gamma receptor 1 deficiency. *J Clin Immunol* (2013) **33**:1062–1066. doi:10.1007/s10875-013-9907-0
14. Toyoda H, Ido M, Nakanishi K, Nakano T, Kamiya H, Matsumine A, Uchida A, Mizutani H, de Beaucoudrey L, Vogt G, et al. Multiple cutaneous squamous cell carcinomas in a patient with interferon gamma receptor 2 (IFN gamma R2) deficiency. *J Med Genet* (2010) **47**:631–634. doi:10.1136/jmg.2009.072108
15. Taramasso L, Boisson-Dupuis S, Garrè ML, Bondi E, Cama A, Nozza P, Morana G,

- Casanova JL, Marazzi MG. Pineal germinoma in a child with interferon- γ receptor 1 deficiency. case report and literature review. *J Clin Immunol* (2014) **34**:922–927.
doi:10.1007/s10875-014-0098-0
16. Camcioglu Y, Picard C, Lacoste V, Dupuis S, Akçakaya N, Cokura H, Kaner G, Demirkesen C, Plancoulaine S, Emile J-F, et al. HHV-8-associated Kaposi sarcoma in a child with IFN γ R1 deficiency. *J Pediatr* (2004) **144**:519–523. doi:10.1016/j.jpeds.2003.11.012
17. Ren Y, Zhang Y, Liu RZ, Fenstermacher DA, Wright KL, Teer JK, Wu J. JAK1 truncating mutations in gynecologic cancer define new role of cancer-associated protein tyrosine kinase aberrations. *Sci Rep* (2013) **3**:3042. doi:10.1038/srep03042
18. Longo T, McGinley KF, Freedman JA, Etienne W, Wu Y, Sibley A, Owzar K, Gresham J, Moy C, Szabo S, et al. Targeted Exome Sequencing of the Cancer Genome in Patients with Very High-risk Bladder Cancer. *Eur Urol* (2016) **70**:714–717.
doi:10.1016/j.eururo.2016.07.049
19. Albacker LA, Wu J, Smith P, Warmuth M, Stephens PJ, Zhu P, Yu L, Chmielecki J. Loss of function JAK1 mutations occur at high frequency in cancers with microsatellite instability and are suggestive of immune evasion. *PLoS One* (2017) **12**:e0176181.
doi:10.1371/journal.pone.0176181
20. Grossman RL, Heath AP, Ferretti V, Varmus HE, Lowy DR, Kibbe WA, Staudt LM. Toward a Shared Vision for Cancer Genomic Data. *N Engl J Med* (2016) **375**:1109–1112.
doi:10.1056/NEJMp1607591
21. McLaren W, Gil L, Hunt SE, Riat HS, Ritchie GRS, Thormann A, Flicek P, Cunningham F. The Ensembl Variant Effect Predictor. *Genome Biol* (2016) **17**:122. doi:10.1186/s13059-016-0974-4
22. Vaser R, Adusumalli S, Leng SN, Sikic M, Ng PC. SIFT missense predictions for genomes.

Nat Protoc (2016) **11**:1–9. doi:10.1038/nprot.2015.123

23. Adzhubei I, Jordan DM, Sunyaev SR. Predicting Functional Effect of Human Missense Mutations Using PolyPhen-2. *Curr Protoc Hum Genet* (2013) **76**:7.20.1-7.20.41. doi:10.1002/0471142905.hg0720s76
24. Liu M, Watson LT, Zhang L. Quantitative prediction of the effect of genetic variation using hidden Markov models. *BMC Bioinformatics* (2014) **15**:5. doi:10.1186/1471-2105-15-5
25. Georgopoulos NT, Kirkwood LA, Varley CL, MacLaine NJ, Aziz N, Southgate J. Immortalisation of Normal Human Urothelial Cells Compromises Differentiation Capacity. *Eur Urol* (2011) **60**:141–149. doi:10.1016/j.eururo.2011.02.022
26. Southgate J, Hutton KA, Thomas DF, Trejdosiewicz LK. Normal human urothelial cells in vitro: proliferation and induction of stratification. *Lab Invest* (1994) **71**:583–594. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/7967513>
27. Varley CL. Role of PPAR and EGFR signalling in the urothelial terminal differentiation programme. *J Cell Sci* (2004) **117**:2029–2036. doi:10.1242/jcs.01042
28. Budczies J, Bockmayr M, Klauschen F, Endris V, Fröhling S, Schirmacher P, Denkert C, Stenzinger A. Mutation patterns in genes encoding interferon signaling and antigen presentation: A pan-cancer survey with implications for the use of immune checkpoint inhibitors. *Genes, Chromosom {&} cancer* (2017) **56**:651–659. doi:10.1002/gcc.22468
29. Shin DS, Zaretsky JM, Escuin-Ordinas H, Garcia-Diaz A, Hu-Lieskovan S, Kalbasi A, Grasso CS, Hugo W, Sandoval S, Torrejon DY, et al. Primary Resistance to PD-1 Blockade Mediated by JAK1/2 Mutations. *Cancer Discov* (2017) **7**:188–201. doi:10.1158/2159-8290.CD-16-1223
30. Grenga I, Donahue RN, Gargulak ML, Lepone LM, Roselli M, Bilusic M, Schlom J. Anti-

PD-L1/TGF β R2 (M7824) fusion protein induces immunogenic modulation of human urothelial carcinoma cell lines, rendering them more susceptible to immune-mediated recognition and lysis. *Urol Oncol* (2017) doi:10.1016/j.urolonc.2017.09.027

31. López-Soto A, Gonzalez S, Smyth MJ, Galluzzi L. Control of Metastasis by NK Cells. *Cancer Cell* (2017) **32**:135–154. doi:10.1016/j.ccell.2017.06.009
32. Dadhania V, Zhang M, Zhang L, Bondaruk J, Majewski T, Siefker-Radtke A, Guo CC, Dinney C, Cogdell DE, Zhang S, et al. Meta-Analysis of the Luminal and Basal Subtypes of Bladder Cancer and the Identification of Signature Immunohistochemical Markers for Clinical Use. *EBioMedicine* (2016) **12**:105–117. doi:10.1016/j.ebiom.2016.08.036
33. Varley CL, Bacon EJ, Holder JC, Southgate J. FOXA1 and IRF-1 intermediary transcriptional regulators of PPAR γ -induced urothelial cytodifferentiation. *Cell Death Differ* (2009) **16**:103–114. doi:10.1038/cdd.2008.116
34. Jackson AM, Alexandrov AB, Prescott S, James K, Chisholm GD. Role of adhesion molecules in lymphokine-activated killer cell killing of bladder cancer cells: further evidence for a third ligand for leucocyte function-associated antigen-1. *Immunology* (1992) **76**:286–91. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/1353063>
35. Crotzer VL, Blum JS. Autophagy and its role in MHC-mediated antigen presentation. *J Immunol* (2009) **182**:3335–3341. doi:10.4049/jimmunol.0803458
36. Shan G, Tang T, Qian H, Xia Y. Certain BCG-reactive responses are associated with bladder cancer prognosis. *Cancer Immunol Immunother* (2018) **67**:797–803. doi:10.1007/s00262-018-2127-y
37. Efsthathiou JA, Mouw KW, Gibb EA, Liu Y, Wu C-L, Drumm MR, da Costa JB, du Plessis M, Wang NQ, Davicioni E, et al. Impact of Immune and Stromal Infiltration on Outcomes Following Bladder-Sparing Trimodality Therapy for Muscle-Invasive Bladder Cancer. *Eur*

Urol (2019) **76**:59–68. doi:10.1016/j.eururo.2019.01.011

38. Ahlén Bergman E, Hartana CA, Johansson M, Linton LB, Berglund S, Hyllienmark M, Lundgren C, Holmström B, Palmqvist K, Hansson J, et al. Increased CD4+ T cell lineage commitment determined by CpG methylation correlates with better prognosis in urinary bladder cancer patients. *Clin Epigenetics* (2018) **10**:102. doi:10.1186/s13148-018-0536-6
39. Ren R, Tyryshkin K, Graham CH, Koti M, Siemens DR. Comprehensive immune transcriptomic analysis in bladder cancer reveals subtype specific immune gene expression patterns of prognostic relevance. *Oncotarget* (2017) **8**:70982–71001. doi:10.18632/oncotarget.20237
40. Bai J, Gao Z, Li X, Dong L, Han W, Nie J. Regulation of PD-1/PD-L1 pathway and resistance to PD-1/PDL1 blockade. *Oncotarget* (2017) **8**: doi:10.18632/oncotarget.22690
41. Kamat AM, Bellmunt J, Galsky MD, Konety BR, Lamm DL, Langham D, Lee CT, Milowsky MI, O'Donnell MA, O'Donnell PH, et al. Society for Immunotherapy of Cancer consensus statement on immunotherapy for the treatment of bladder carcinoma. *J Immunother cancer* (2017) **5**:68. doi:10.1186/s40425-017-0271-0

Figure legends

Figure 1. STAT1 phosphorylation and expression of *IRF1* mRNA is impaired in JAK1-deficient hTERT urothelial cells. (A) JAK1 protein (InterPro P23458) structure showing core domains, TCGA BLCA variants and their predicted impact. Protein domains: FERM (4.1 protein, ezrin, radizin, moesin domain), PHD (pleckstrin homology-like domain), SH2 (Src homology 2), S-TY-PK (unknown specificity serine-threonine/tyrosine protein kinase), SY-PK (serine-tyrosine protein kinase). Variants: nonsense (red cross), missense (yellow inverted triangle), synonymous (green triangle), frameshift (purple square). Variant effects: S (SIFT score < 0.05), P (Poly-Phen score > 0.908), p (Poly-Phen score > 0.446 & ≤ 0.908), H (HMMvar score > 2). A single 5' UTR modifier mutation not shown. (B,C) Analysis of JAK/STAT signalling by flow cytometry in untransduced (Unt), Sc and KD hTERT immortalised urothelial cell lines after stimulation with IFN γ (1ng/ml) for 24h. Data A is from a representative experiment, data B is from three independent experiments. Two-tailed Mann Whitney test. (D) RTqPCR analysis of *IRF1* mRNA expression from KD and Sc hTERT immortalised urothelial cell lines after stimulation with IFN γ (1ng/ml). Data is from four independent experiments. Two-tailed Mann Whitney test.*P <0.05 Error bars represent the SE.(E,F) Flow cytometry analysis of IFNGR expression in KD and Sc hTERT immortalised urothelial cell lines following stimulation with IFN γ (5 ng/ml) for two days. The graph shows mean values \pm SD. Data is from three independent experiments. One-way ANOVA with Tukey's Multiple Comparison Test.

Figure 2. Analysis of MHC I/II, ICAM-1 and PD-L1 expression in hTERT urothelial cells by Flow Cytometry. Flow cytometry analysis of surface receptor expression in KD and Sc hTERT immortalised urothelial cell lines before and after IFN γ stimulation. Mean fluorescence intensity and percentage of positive cells are shown for (A,B) MHCII (C, D) MHC I (E-F) ICAM-1 and (G-H) PD-L1 expression. Data is from four independent experiments for (A-F) and from three

independent experiments (**G-H**). One-way ANOVA with Tukey's Multiple Comparison post-test. Graphs shows mean values \pm SE and regulation of the expression compared to untreated. *P < 0.05. Error bars represent the SE.

Figure 3. JAK1-deficient hTERT urothelial cells demonstrate preserved population growth and reduced apoptosis in response to IFN γ . (A) KD and Sc hTERT immortalised urothelial cell lines were cultured with addition of Alamar Blue (AB) dye and stimulated with IFN γ (5ng/ml) for the given time points. The capacity of viable cells to reduce AB dye was used as a proxy for cell number. Data A is from five independent experiments. (B,C,D,E) KD and Sc hTERT urothelial cell lines were stimulated with the given concentrations of IFN γ for 5 days. Percentage of early and late apoptosis was quantified with Annexin V/PI apoptosis detection kit by flow cytometry. Data B,C and D are from five independent experiments, data E is from a representative experiment. One-way ANOVA with Tukey's Multiple Comparison post-test. Error bars represent the SE.

Figure 4. JAK1-deficient hTERT urothelial cells showed reduced lymphocyte-mediated killing in response to IFN γ . JAK1-deficient and Sc hTERT immortalised urothelial cell lines were pretreated or not with IFN γ and cultured overnight with 25U/ml IL-2 and monocyte-depleted PBMCs (50:1). (A,B) Necrosis induction in JAK1-deficient and Sc hTERT immortalised urothelial cell lines. One-way ANOVA with Dunn's multiple comparisons post-test. (C) Relative change of necrosis compared to untreated. Two-tailed Mann Whitney U test. Data is from five independent experiments. *P < 0.05. Error bars represent the SE.

Figure 5. Immunohistochemical labelling of patient's tumour using panel of antibodies against differentiation (UPK3a, CK20, CK14), transcription factor (PPAR γ , GATA3, FOXA1,

IRF1), and cell signalling (pSTAT1 and HER2) markers. Images from central region of the tumour, except where marked as invasive. Parallel human tissue controls for antibodies are shown in supplementary Figure S3. Scale bar = 12 μm .

Figure 6. Expression of IFN γ -regulatory genes and intermediary transcription factors in NHU cells by RT-qPCR. RT-qPCR analysis of gene expression in five non-immortalized NHU cell cultures obtained from five different donors, with or without IFN γ (200U/ml) and/or TZ (1 μM) + PD153035 (1 μM) stimulation for 48h compared with the vehicle-only (0.1% DMSO) control. Data are from five independent experiments each using a different NHU cell donor. One-way ANOVA with Tukey's Multiple Comparisons Test. *P<0.05. Error bars represent the SE.