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Inducing pluripotency in the domestic cat (*Felis Catus*)

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Abstract

Domestic cats suffer from a range of inherited genetic diseases, many of which display similarities with equivalent human conditions. Developing cellular models for these inherited diseases would enable drug discovery, benefiting feline health and welfare as well as enhancing the potential of cats as relevant animal models for translation to human medicine. Advances in our understanding of these diseases at the cellular level have come from the use of induced pluripotent stem cells (iPSCs). iPSCs are capable of differentiating into derivatives of all three germ layers, therefore overcoming the limitations of primary differentiated cells and the ethical concerns of using embryonic stem cells. No studies however report the generation of iPSCs from domestic cats (fiPSCs). Feline adipose derived fibroblasts were infected with amphotropic retrovirus containing the coding sequences for human Oct4, Sox2, Klf4, cMyc and Nanog. Isolated iPSC clones were expanded on mouse inactivated embryonic fibroblasts in the presence of feline leukaemia inhibitory factor (LIF). Retroviral delivery of human pluripotent genes gave rise to putative fiPSC colonies within 5-7 days. These iPS-like cells required foetal bovine serum and feline LIF for maintenance. Colonies were domed with refractile edges, similar to mouse iPSCs. Immunocytochemistry demonstrated positive staining for stem cell markers: alkaline phosphatase, Oct4, Sox2, Nanog and SSEA1. Cells were negative for SSEA4. Expression of endogenous feline Nanog was confirmed by qPCR. The cells were able to differentiate *in vitro* into cells representative of the three germ layers. These results confirm the generation of the first induced pluripotent cells from domestic cats. These cells will provide valuable models to study genetic diseases and explore novel therapeutic strategies.

Introduction

Inducing pluripotency is a process where differentiated cells are reprogrammed into a pluripotent or embryonic-stem cell like state [1]. These induced pluripotent stem cells (iPSCs) are then able to differentiate into derivatives of all three germ layers, generating great promise for their use in regenerative medicine and disease modelling, whilst avoiding the ethical implications of using embryonic stem cells [2,3]. They can also be derived from patients with disease causing genetic mutations, allowing patient-specific iPSC lines to be developed for *in vitro* drug testing. For example, iPSC lines harbouring mutations known to cause hypertrophic cardiomyopathy (HCM) effectively recapitulate the disease at the single cell level, allowing discovery of novel drug targets for clinical therapy [4]. iPSC derived cardiomyocytes have further been shown to highlight potential drug side-effects in specific patients and predict drug response [5]. Additionally, the consistency and ability to control for other factors makes iPSC disease models a rapid drug discovery system [6]. Recent discoveries in gene editing mean that iPSC lines can be mutated, providing a powerful tool for analysing disease-causing mutations whilst controlling for genetic background [7].

iPSCs were first generated from mouse cells and subsequently from human cells [3,8,9]. Since then research focussed on optimising human iPSC generation and culture has allowed the whole process to be xeno-free and footprint-free (without inserted genetic sequences), providing a platform for clinical research [10]. Studies deriving iPSCs from other species have shown the process is applicable across a wide range of animals. For example, multiple groups have derived iPSCs from dogs, horses, sheep, pigs, goats and

other wild species [11–18]. With respect to felids, there are two studies describing the generation of iPSCs from wild species [19,20]. Currently no studies have described the generation of iPSCs from domestic cats, an important veterinary species. Cats also provide a spontaneous large animal model for important human diseases, including diabetes, cardiac disease as well as a wide range of genetic conditions, allowing a one health approach to disease [21–24]. For example, HCM in cats closely resembles human HCM at the genetic, clinical and pathological level [25]. Interestingly a causative mutation identified in cats is also present in a human family [26]. HCM has a high prevalence in the feline population, affecting around 15% of all cats and increasing to almost 30% in those over 9 years of age [27]. Given the importance and similarities of disease in both humans and cats, discovering novel drug targets would be beneficial to the health of both species. Drug discovery utilising feline iPSCs could be translated to the feline clinic, where benefits can be assessed more rapidly due to the high prevalence, rapid disease progression and reduced regulation compared to human clinical trials.

iPSCs derived from species other than mouse or human remain cultured under conditions that are not fully optimised or defined. Many still rely on original methods of retroviral insertion of human genes, use of undefined animal-derived media and maintenance on inactivated mouse embryonic fibroblasts [15,28]. Much of what is known about pluripotent stem cells from diverse species is based on knowledge of embryonic stem cell culture. In cats, there are no studies deriving *bona fide* embryonic stem cells. There are two reports of embryonic stem-like cells, however these could not be maintained under the culture

conditions indefinitely, raising the possibility that either the cells were not true ESCs, or that the culture conditions were not able to sustain a pluripotent state [29,30].

The development of a method to derive iPSCs from feline somatic cells would advance feline genetic and embryological research, which is currently hampered also by the lack of ESCs. These methods would provide researchers with a platform to further optimise iPSC culture from a novel species and realise the same potential as for human iPSCs.

Material and Methods

Fibroblast cell culture

Feline adipose tissue derived fibroblasts, isolated as described elsewhere [31], had been cryopreserved in the liquid phase of nitrogen at passage 0 (P0) to P1 prior to commencement of this project as approved by the Ethics and Welfare Committee (Approval number: URN 2012 1192) at the Royal Veterinary College (RVC). Cells were thawed rapidly in a 37°C water bath, then re-suspended in media consisting of Dulbecco's Modified Eagles Medium (DMEM; Thermo Fisher), 10% v/v foetal bovine serum (FBS; Thermo Fisher), 1% v/v L-glutamine (Thermo Fisher) and 10 ng mL⁻¹ basic fibroblast growth factor (bFGF, Peprotech). All cultures were maintained at 37°C, 5% CO₂ in humidified air.

Feeder cell preparation

Proliferating mouse embryonic fibroblasts (MEFs) were purchased at P3 (EmbryoMax, Sigma-Aldrich). The cells were cultured in the same media as for feline fibroblasts but without bFGF. MEFs were thawed onto 0.2% gelatin (Sigma Aldrich) coated 75 cm² flasks at a density of 1.3 x 10⁴ cells cm⁻² and passaged 1:3 at 70-80% confluence. At passage five, MEFs were mitotically inactivated with 10 µg ml⁻¹ mitomycin C (Stem Cell Technologies) for 2 h and cryopreserved in Cell Banker 2 (AMS Bio) at a concentration of 1 x 10⁶ cells ml⁻¹.

Retroviral production and transfection

Moloney murine leukaemia virus (MMLV) based retroviral plasmids (pMXs) containing the coding sequences for human Oct4 (Addgene 17217), Sox2 (Addgene 17218), Klf4 (Addgene 17219), c-Myc (Addgene 17220) and Nanog (Addgene 18115) were purchased from Addgene. A GFP containing plasmid (Cell Biolabs) was used to assess for transfection efficiency. For production of MMLV virus, Platinum-A packaging cells (Cambridge Bioscience), which express the *gag*, *pol* and *env* genes, were plated 24 h prior to transfection at a density of 1×10^5 cells cm^{-2} in 60 mm cell culture dishes, one dish per retroviral vector plasmid. Next, 6 μg of each of the vectors was transfected to Platinum-A cells separately using Lipofectamine 2000 (Thermo Fisher) according to the manufacturers' instruction. To optimise transfection efficiency, feline fibroblasts were transfected with GFP vector at a range of seeding densities (500 - 1×10^4 cells cm^{-2}), cultured for 24, 48 or 72 h and in media with and without bFGF 24 h. Following this optimisation, fibroblasts were plated 24 h before transfection at a density of 500 cells cm^{-2} in 100 mm cell culture dishes, in media containing bFGF. Supernatants containing virus particles were collected from the packaging cells after 48 h, filtered through a 0.45 μm filter, supplemented with polybrene ($8 \mu\text{g mL}^{-1}$, Sigma Aldrich) and 10 ng mL^{-1} bFGF. The five transcription factor virus supernatants were pooled in equal volumes and added to the fibroblasts. We did not quantify viral titre prior to infection since storage of MMLV leads to reduced viral titre making this impractical. Instead we used a parallel GFP experiment to monitor that viral production and cell infection was successful. The procedure was repeated after removal of the previous viral supernatant 24 h later for a total of two rounds of transfection.

Generation and culture of iPSCs

At 24 h following the second retroviral infection, inactivated MEFs (iMEFs) were added to the plate of feline fibroblasts at a density of $2 \times 10^4 \text{ cm}^{-2}$ in fibroblast media. The following day the media was changed to feline iPSC media, which consisted of DMEM-GlutaMax, 20% v/v ESC-qualified FBS (Thermo Fisher), 1% v/v non-essential amino acids (Thermo Fisher), 0.1 mM 2-mercaptoethanol (Thermo Fisher) and 10 ng mL^{-1} feline leukaemia inhibitory factor (fLIF, Kingfisher Biotech). Colonies were manually picked based on ESC-like morphology at 7-10 days using a dissecting microscope and P-20 pipette tip. Colonies were placed into a 96-well plate containing 20 μL of 0.25% trypsin-EDTA (Thermo Fisher), incubated at room temperature for 5 min and then mixed by pipetting to obtain single cells. This cell suspension was then transferred to one well of a 24-well plate, which had been seeded with iMEFs 24 h previously. Individual colonies were treated separately to obtain clonal cell lines. Hereafter feline iPSCs were passaged as single cells using 0.25% trypsin-EDTA at a 1:6 to 1:10 ratio every 2-3 days. Cells for cryopreservation were re-suspended in CellBanker 2 media, transferred to -80°C in a Mr Frosty freezing container (Nalgene), then transferred to liquid nitrogen storage after 24 h.

Embryoid body formation and differentiation

To form embryoid bodies feline iPSCs were treated with 0.25% trypsin-EDTA for 5 mins and cells were transferred to CELLSTAR cell repellent surface 24-well plates (Greiner Bio-one) using a 1:1 split ratio from a confluent 6 well plate. Media was changed to iPSC media without fLIF. After 4-5 days embryoid bodies were collected and re-distributed onto

0.2% gelatin coated 48-well plates and allowed to differentiate for 3 weeks before antibody staining.

Alkaline phosphatase staining and immunocytochemistry

Alkaline phosphatase activity of feline iPSC at passages 2, 15 and 22 was determined using VECTOR Red Alkaline Phosphatase (Red AP) Substrate Kit (Vector Laboratories). Cells for immunocytochemistry at passages 15-22 were plated in 48-well plates for all staining procedures and allowed to adhere for at least 24 h. Immunostaining procedures were performed at room temperature following removal of culture media. Cells were washed once with Dulbecco's phosphate buffered saline (DPBS) and fixed in 4% paraformaldehyde (Sigma-Aldrich) for 15 min. Fixative was then removed and the cells were permeabilised using 0.4% Triton X-100 (Sigma-Aldrich) for 15 min. Cells were washed with 0.05% Tween 20/DPBS (Sigma Aldrich) and blocked with 10% goat serum (Abcam) for 1 h. Cells were washed again and incubated with the primary antibody for 1.5 h. Following three washes cells were incubated with the secondary antibody for 1 h. Cells were then washed again three times and nuclei counterstained using NucBlue ReadyProbe according to the manufacturers' instructions (Thermo Fisher). The primary and secondary antibodies used for ICC are listed in Table 1. Images were captured using an EVOS FL colour imaging system (Thermo Fisher).

Flow cytometry

To determine the GFP positive percentage, cells were detached from culture vessels using 0.25% trypsin-EDTA 48 h post second transfection. Cells were pelleted (400 g for

5 min) and re-suspended in FACSFlow (BD Bioscience). Samples were acquired in polystyrene FACS tubes on a BD FACS Calibur flow cytometer (BD Bioscience). The instrument was calibrated using CaliBRITE 3 colour FACS Comp beads (BD Bioscience) before acquiring and analysing each set of samples using CellQuest Pro software (BD Bioscience). Non-infected cells were acquired in order to set the forward and side scatter parameters to centre the cell population on the scatter plot. Fluorescence intensity was adjusted to set the non-infected cells within $10^0 - 10^1$ on the log scale axis. Cells were then acquired with an event count set to a total of 1×10^4 events. Data was analysed using FlowJo software (FlowJo, Version 10, LLC).

Reverse transcription and quantitative real-time polymerase chain reaction

RNA was extracted from non-infected fibroblasts, infected fibroblasts and iPSCs at passages 1-22 using the GenElute™ Mammalian Total RNA Miniprep Kit (Sigma-Aldrich). Genomic DNA contamination was removed using the TURBO DNA-free kit (Thermo Fisher). Then 1 µg of RNA was reverse transcribed using the SensiFAST cDNA synthesis kit (Bioline). Quantitative real-time PCR (qPCR) was performed using 2 µL of cDNA template, non-reverse transcribed control or water using the SensiMix SYBR green No-ROX kit (Bioline) and run on a CFX96 Real Time PCR Detection System (Bio-rad) with the following conditions: 95°C for 10 min, then 44 cycles of 95°C for 15 s, 60°C for 15s, 72°C for 15s. A final ramp of 60-90°C at 0.5°C increments was used for melt curve analysis. Five housekeeping genes were first analysed to ascertain stability across different cell types (non-infected fibroblasts, pooled infected fibroblasts and three iPSC lines). RPS7 was scored as the most stable using RefFinder, which compares analyses

from 4 computational programs (Delta CT, BestKeeper, Normfinder and Genorm; Supplementary Figure 1). This was then used to normalise qPCR data for other genes analysed. The primer sequences are given in Table 2.

Karyotyping

Feline iPSCs were incubated for 18 h at 37°C in standard cell culture conditions in media containing 0.13 $\mu\text{g mL}^{-1}$ colcemid (KaryoMax; Thermo Fisher). Next cells were dissociated using 0.25% trypsin-EDTA, pelleted (300g for 3 min) and resuspended by adding 4 mL of 0.075 M KCL drop-wise to the cell pellet and then incubated at 37°C for 25 min. Then 10 drops of fixative (3:1 methanol/acetic acid) were added and cells mixed by inversion and incubated for 10 min at room temperature. Cells were then pelleted as before, re-suspended in 4 mL fixative and incubated for 30 min at room temperature. Following a final centrifugation cells were resuspended in 2 mL of fixative and chromosomes visualised using DAPI staining. At least 20 metaphase spreads were counted, with a haploid number of 38 considered normal.

Statistical analysis

The SPSS software package (IBM, version 23 for Mac) was used for statistical analysis. Data was assessed for normality using histogram analysis. All data are presented as the mean \pm SEM of three individual clonal iPSC lines derived from the same reprogramming experiment, with samples run at least in triplicate. Fibroblast data is obtained from 3 independent analyses of the same cell line or transfected pool that generated the iPSC lines. Comparisons between two independent samples were performed using Student's

two-tailed T-test and between three or more groups using one-way ANOVA with post-hoc Tukey analysis. A p value of < 0.05 was considered significant.

Results

Amphotropic retrovirus infects feline cells with high efficiency

Transduction efficiency of feline fibroblasts was assessed by transfection with a GFP containing amphotropic MMLV. After a number of optimisations, the transfection efficiency could be increased from an initial 10-20% to >99% (Figure 1a and b). Seeding the cells at low density (500 cells cm⁻²) in media containing bFGF 24 h prior to transfection led to the highest GFP expression.

Infected feline fibroblasts express human pluripotent genes

Once we confirmed efficient transduction, feline fibroblasts were transduced with pooled viral supernatant containing coding sequences for five human pluripotency associated genes (OSKMN). Immunocytochemistry demonstrated expression of the pluripotent factors at the protein level with a nuclear localisation as expected (Figure 2a). RT-qPCR confirmed that the feline cells had high gene expression levels of integrated human pluripotent factors (Figure 2b).

Feline iPSCs require feline leukaemia inhibitory factor for maintenance

Following a number of different derivation conditions we ascertained a protocol for feline iPSC induction. A timeline for this is shown in Figure 3a. Feline fibroblasts were seeded 24 h prior to transduction at low density in media containing bFGF. Following 2 rounds of retroviral infection 24 h apart, iMEFs are added directly to the plate of infected feline fibroblasts. Colonies are then observed from day 3 to day 7 (Figure 3b-e). The feline iPSC (fiPSC) colonies were observed to be initially relatively two dimensional, peripherally well

demarcated and with cells exhibiting a high nuclear to cytoplasm ratio, more typical of human embryonic stem cell morphology (Figure 3c). Following the first passage colonies formed a domed morphology with a phase bright surface and retractile edges more typical of mouse ESC morphology (Figure 3d and e). iPSCs were able to be propagated using a 1:6 - 1:10 split ratio every 2-3 days for over 20 passages.

fiPSC were further observed to be dependent on FBS and feline leukaemia inhibitory factor (fLIF) for maintenance. Culture of fiPSC in media containing knock-out serum replacer (KSR) or removing fLIF led to loss of colony morphology within 3-5 days (data not shown). Media containing bFGF with or without LIF allowed small colonies to form but they did not progress beyond 10-20 cells and could not be expanded (data not shown). Additionally, murine LIF was unable to maintain alkaline phosphatase expression of fiPSCs (Figure 4a and b).

Feline iPSCs express key pluripotent markers and reactivate Nanog expression

Feline iPSCs were examined for markers of pluripotent stem cells. Colonies were positive for alkaline phosphatase (Figure 5a). In 3 individual clonal fiPSC lines there was high expression of feline Nanog, which was not detectable in the original fibroblasts (Figure 5b). Additionally cells were positive for the surface marker SSEA1 (Figure 5c-e), negative for SSEA4 (Figure 5f-h) and positive for nuclear transcription factors Oct4 (Figure 5i-k), Sox2 (Figure 5l-n) and Nanog (Figure 5o-q). fiPSC could be passaged >20 times without losing proliferative potential, maintained the above phenotype and had a normal karyotype (Figure 6).

Feline iPSCs are capable of *in vitro* differentiation into derivatives of the three germ layers

We next sought to assess whether the fiPSC had the potential to differentiate into cells representative of the 3 germ layers. fiPSCs seeded onto a low attachment surface in the absence of fLIF formed embryoid bodies (EBs, Figure 7a). Outgrowth cells from EBs attached to gelatin-coated plates displayed multiple morphologies (Figure 7b). Immunostaining revealed that subsets of these cells expressed the mesodermal markers smooth muscle actin (Figure 7d) and α -sarcomeric actin (Figure 7f), ectodermal marker β III-tubulin (Figure 7h) and endodermal marker SOX17 (Figure 7j). None of these markers were expressed in the undifferentiated iPSCs (Figure 7 c, e, g and i). Together these results indicated the *in vitro* pluripotency of fiPSCs.

Feline iPSCs continue to express viral transgenes

Feline iPSCs showed expression of the viral transgenes in the infected fibroblasts and in three iPSC lines at passage 7 and passage 22 (Figure 8). Expression levels for Oct4 and Sox2 were higher in the iPSC lines compared to the infected fibroblasts ($p < 0.05$). Expression of cMyc was higher in passage 7 iPSCs compared to infected fibroblasts ($p < 0.05$). Nanog expression was lower in iPSCs compared with the infected pool, however this difference was not significant ($p = 0.252$ for passage 7 and $p = 0.720$ for passage 22). There was no difference in expression levels between passage 7 and passage 22 iPSCs for any of the transgenes ($p = 0.993$ for Oct4, $p = 0.997$ for Sox2, $p = 0.223$ for Klf4, $p = 1.00$ for cMyc and $p = 0.881$ for Nanog).

Discussion

In this study we describe the first generation of induced pluripotent stem cells from the domestic cat. We present a number of optimisation steps required for successful iPSC generation and anticipate these results will advance feline medical research similar to the advances made in human medical research, particularly for genetic diseases [32,33]. These new technologies have been widely applied in human neurological and cardiac diseases, where patient-specific iPSC lines act as powerful tools as *in vitro* disease models [34–36]. These include new gene therapy strategies for iPSC-derived cellular models of hypertrophic cardiomyopathy [37]. Our work provides the initial steps to realising these advancements for feline patients and enhances the potential of cats as large animal models for analogous human disease where prospective therapies can be rapidly translated from the feline to the human clinic.

A major barrier to inducing pluripotency is the efficient delivery and expression of pluripotent genes (Oct4, Sox2, Klf4, cMyc and Nanog; OSKMN) [38]. There are few reports on viral delivery of genes in domestic cat cells [39,40]. Initially we trialled feline iPSC generation using a Sendai virus based delivery of OSKM (Cytotune 2.0, Thermo Fisher), however although the virus was able to infect feline cells, iPSC colonies never appeared despite several attempts with various modifications (data not shown). We then selected an MMLV-based retrovirus delivery system as this has been previously reported for the generation of iPSCs from wild feline species [19,20]. Additionally this allowed us to add Nanog into the reprogramming factors, as this is required for iPSC generation in wild felids [20]. This system yielded low transfection efficiencies when replicating

conditions used for mouse or human cells. We hypothesised that this was due to slow growth of feline fibroblasts in DMEM/10% FBS media since MMLV retrovirus is only able to integrate into cells that are undergoing mitosis [41]. In order to increase the proliferation of the fibroblasts, media was supplemented with bFGF [42] together with seeding cells 24 h prior to transfection and at low density to increase the MOI. This led to a maximum GFP expression of about 99%. This is in line with retroviral based delivery methods for iPSC generation in other species [8,11,20]. We further demonstrated that feline cells are able to express the transduced human genes at the mRNA and protein level.

It was also necessary to optimise the conditions for iPSC colony emergence. We observed mesenchymal-to-epithelial transition (MET) of fibroblasts within 2-3 days of viral transfection, which is thought to be one of the earliest stages in the reprogramming process [43]. Under conditions used for human iPSC generation (including KSR and bFGF), these epithelial-shaped cells reverted back to a mesenchymal morphology. We also observed that if MEFs were added after day 5, no colonies emerged. Since MET was occurring within a few days, it appeared that reprogramming in feline cells was following a similar timeline as for mouse iPSCs [44], which form quicker than human iPSCs [45]. We therefore added iMEFs to the transfected cells and included feline LIF as early as one day after virus was removed which yielded colonies with typical ESC and iPSC morphology [46].

These fiPSCs expressed Oct4, Sox2, Nanog and SSEA1 but not SSEA4 similar to mouse ES and iPS cells [47]. Further characterisation of the fiPSCs showed they were able to

differentiate into cells representing the three germ layers and had a normal karyotype after >20 passages. Nanog, a marker of cellular reprogramming [48], was also activated.

Culture conditions for iPSCs from different species show considerable variation between publications, which makes determining the optimal conditions for a novel species challenging [15]. This difference may be due to the developmental stages represented by iPSCs [49]. For example, mouse iPSCs represent a 'naive' type pluripotent stem cell, and have a domed and phase-bright appearance [50]. However human iPSCs represent a 'primed' pluripotent cell, similar to mouse epiblast cells, with flattened colony morphology [51]. Both types can be converted into the other type by adjusting culture conditions [52,53]. The naive type relies on LIF for maintenance of pluripotency and self-renewal, whereas the primed type is dependent on bFGF [54]. It is less clear where iPSCs derived from other species fit along this spectrum, with iPSCs derived from monkeys, pigs, horses, cows, goats and dogs fitting both definitions [11,13,14,16,18,55–57]. With respect to our feline iPSCs, they closely resemble the naive type, which is substantiated by their morphology and surface expression of SSEA1 and not SSEA4, as is the case for mouse iPSCs. Specifically feline cells relied on feline LIF, with murine LIF unable to sustain alkaline phosphatase expression, and this is likely due to the differences between feline and murine LIF protein and receptors [58].

Although we describe conditions for feline iPSCs, there is potential for the system to be further optimised. For example, the use of FBS and iMEFs led to inconsistent colony morphology and behaviour. Further optimisation may identify other growth factors or

molecules that would enable a more stable phenotype and facilitate feeder cell removal. The suboptimal culture conditions in our study may have led to the persistent transgene expression that we observed in the feline iPSCs [59]. In human and mouse studies transgene expression is expected to cease at around passage 10-15, which was not the case for the iPSCs in this study [60]. This is common issue with iPSCs derived from domestic animals and where expression of inserted factors has been directly measured the majority of reports demonstrate persistent transgene expression [15], as is also the case for wild felid iPSCs [19,20]. Although these cells would be useful for *in vitro* drug discovery, continued transgene expression creates an issue for clinical application since implantation of these cells may cause tumorigenesis [6], therefore transgene-free feline iPSCs would be a useful next step, and this has been the process for human iPSCs.

In conclusion, we sought to establish a platform to generate the first iPSCs from domestic cats with the expectation that these will advance feline medicine. Outcomes from their application to the understanding and management of disease is not limited to the cat since similarities of genetic pathologies between cats and humans will provide invaluable information for human conditions where similar clinical translation is more difficult and outcomes take longer.

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Author Disclosure Statement

The authors declare no competing financial interests.

References

1. Stadtfeld M and K Hochedlinger (2010). Induced pluripotency: history, mechanisms, and applications. *Genes Dev* 24: 2239–2263.
2. Takahashi K and S Yamanaka (2006). Induction of Pluripotent Stem Cells from Mouse Embryonic and Adult Fibroblast Cultures by Defined Factors. *Cell* 126: 663–676.
3. Takahashi K, K Tanabe, M Ohnuki, M Narita, T Ichisaka, K Tomoda and S Yamanaka (2007). Induction of Pluripotent Stem Cells from Adult Human Fibroblasts by Defined Factors. *Cell* 131: 861–872.
4. Prondzynski M, E Krämer, SD Laufer, A Shibamiya, O Pless, F Flenner, OJ Müller, J Münch, C Redwood, A Hansen, M Patten, T Eschenhagen, G Mearini and L Carrier (2017). Evaluation of MYBPC3 trans-Splicing and Gene Replacement as Therapeutic Options in Human iPSC-Derived Cardiomyocytes. *Mol Ther - Nucleic Acids* 7: 475–486.
5. Matsu E, PW Burrige, K-H Yu, JH Ahrens, V Termglinchan, H Wu, C Liu, P Shukla, N Sayed, JM Churko, N Shao, NA Woo, AS Chao, JD Gold, I Karakikes, MP Snyder and JC Wu (2016). Transcriptome Profiling of Patient-Specific Human iPSC-Cardiomyocytes Predicts Individual Drug Safety and Efficacy Responses In Vitro. *Cell Stem Cell* 19: 311–325.
6. Gunaseeli I, MX Doss, C Antzelevitch, J Hescheler and A Sachinidis (2010). Induced pluripotent stem cells as a model for accelerated patient- and disease-specific drug discovery. *Curr Med Chem* 17: 759–766.
7. Cai B, S Sun, Z Li, X Zhang, Y Ke, J Yang and X Li (2018). Application of CRISPR/Cas9 technologies combined with iPSCs in the study and treatment of retinal

degenerative diseases. *Hum Genet* 137: 679–688.

8. Takahashi K and S Yamanaka (2006). Induction of Pluripotent Stem Cells from Mouse Embryonic and Adult Fibroblast Cultures by Defined Factors. *Cell* 126: 663–676.

9. Yu J, MA Vodyanik, K Smuga-Otto, J Antosiewicz-Bourget, JL Frane, S Tian, J Nie, GA Jonsdottir, V Ruotti, R Stewart, II Slukvin and JA Thomson (2007). Induced Pluripotent Stem Cell Lines Derived from Human Somatic Cells. *Science* (80-) 318: 1917–1920.

10. Beers J, KL Linask, JA Chen, LI Siniscalchi, Y Lin, W Zheng, M Rao and G Chen (2015). A cost-effective and efficient reprogramming platform for large-scale production of integration-free human induced pluripotent stem cells in chemically defined culture. *Sci Rep* 5: 11319.

11. Song H, H Li, M Huang, D Xu, C Gu, Z Wang, F Dong and F Wang (2013). Induced pluripotent stem cells from goat fibroblasts. *Mol Reprod Dev* 80: 1009–1017.

12. Li Y, M Cang, AS Lee, K Zhang and D Liu (2011). Reprogramming of Sheep Fibroblasts into Pluripotency under a Drug-Inducible Expression of Mouse-Derived Defined Factors. *PLoS One* 6: e15947.

13. Kawaguchi T, T Tsukiyama, K Kimura, S Matsuyama, N Minami, M Yamada and H Imai (2015). Generation of Naïve Bovine Induced Pluripotent Stem Cells Using PiggyBac Transposition of Doxycycline-Inducible Transcription Factors. *PLoS One* 10: e0135403.

14. Baird AEG, T Barsby and DJ Guest (2015). Derivation of Canine Induced Pluripotent Stem Cells. *Reprod Domest Anim* 50: 669–676.

15. Paterson YZ, C Kafarnik and DJ Guest (2017). Characterization of companion

animal pluripotent stem cells. *Cytom Part A* 93A: 137–148.

16. Breton A, R Sharma, AC Diaz, AG Parham, A Graham, C Neil, CB Whitelaw, E Milne and FX Donadeu (2013). Derivation and Characterization of Induced Pluripotent Stem Cells from Equine Fibroblasts. *Stem Cells Dev* 22: 611–621.

17. Kumar D, T Anand, KP Singh, MK Singh, RA Shah, MS Chauhan, P Palta, SK Singla and RS Manik (2011). Derivation of buffalo embryonic stem-like cells from in vitro-produced blastocysts on homologous and heterologous feeder cells. *J Assist Reprod Genet* 28: 679–688.

18. Ma Y, T Yu, Y Cai and H Wang (2018). Preserving self-renewal of porcine pluripotent stem cells in serum-free 3i culture condition and independent of LIF and b-FGF cytokines. *Cell Death Discov* 4: 21.

19. Verma R, MK Holland, P Temple-Smith and PJ Verma (2012). Inducing pluripotency in somatic cells from the snow leopard (*Panthera uncia*), an endangered felid. *Theriogenology* 77: 220-228.e2.

20. Verma R, J Liu, MK Holland, P Temple-Smith, M Williamson and PJ Verma (2013). Nanog Is an Essential Factor for Induction of Pluripotency in Somatic Cells from Endangered Felids. *Biores Open Access* 2: 72–76.

21. Hoenig M (2006). The cat as a model for human nutrition and disease. *Curr Opin Clin Nutr Metab Care* 9: 584–588.

22. Gurda BL, AM Bradbury and CH Vite (2017). Canine and feline models of human genetic diseases and their contributions to advancing clinical therapies. *Yale J Biol Med* 90: 417–431.

23. Narfström K, K Holland Deckman and M Menotti-Raymond (2011). The Domestic

Cat as a Large Animal Model for Characterization of Disease and Therapeutic Intervention in Hereditary Retinal Blindness. *J Ophthalmol* 2011: 1–8.

24. Chandler M, S Cunningham, EM Lund, C Khanna, R Naramore, A Patel and MJ Day (2017). Obesity and Associated Comorbidities in People and Companion Animals: A One Health Perspective. *J Comp Pathol* 156: 296–309.

25. Maron BJ and PR Fox (2015). Hypertrophic cardiomyopathy in man and cats. *J Vet Cardiol* 17: S6–S9.

26. Ripoll Vera T, L Monserrat Iglesias, M Hermida Prieto, M Ortiz, I Rodriguez Garcia, N Govea Callizo, C Gómez Navarro, J Rosell Andreo, JM Gámez Martínez, G Pons Lladó, D Cremer Luengos and J Torres Marqués (2010). The R820W mutation in the MYBPC3 gene, associated with hypertrophic cardiomyopathy in cats, causes hypertrophic cardiomyopathy and left ventricular non-compaction in humans. *Int J Cardiol* 145: 405–407.

27. Payne JR, DC Brodbelt and V Luis Fuentes (2015). Cardiomyopathy prevalence in 780 apparently healthy cats in rehoming centres (the CatScan study). *J Vet Cardiol* 17: S244–S257.

28. Malik N (2013). A Review of the Methods for Human iPSC Derivation. 997: 23–33.

29. Yu X, G Jin, X Yin, S Cho, J Jeon, S Lee and I Kong (2008). Isolation and characterization of embryonic stem-like cells derived from in vivo-produced cat blastocysts. *Mol Reprod Dev* 75: 1426–1432.

30. Gómez MC, MA Serrano, CE Pope, JA Jenkins and MN Biancardi (2010). Derivation of cat embryonic stem-like cells from in vitro -produced blastocysts on homologous and heterologous feeder cells. *THE* 74: 498–515.

31. Clark KC, FA Fierro, EM Ko, NJ Walker, B Arzi, CG Tepper, H Dahlenburg, A Cicchetto, A Kol, L Marsh, WJ Murphy, N Fazel and DL Borjesson (2017). Human and feline adipose-derived mesenchymal stem cells have comparable phenotype, immunomodulatory functions, and transcriptome. *Stem Cell Res Ther* 8: 69.
32. Ebert AD, P Liang and JC Wu (2012). Induced pluripotent stem cells as a disease modeling and drug screening platform. *J Cardiovasc Pharmacol* 60: 408–416.
33. Kumar S, J Blangero and JE Curran (2018). Induced Pluripotent Stem Cells in Disease Modeling and Gene Identification. In *Disease Gene Identification: Methods and Protocols*, DiStefano, JK, ed. (New York, NY: Springer New York), pp. 17–38.
34. Engle SJ, L Blaha and RJ Kleiman (2018). Best Practices for Translational Disease Modeling Using Human iPSC-Derived Neurons. *Neuron* 100: 783–797.
35. Karakikes I, M Ameen, V Termglinchan and JC Wu (2015). Human Induced Pluripotent Stem Cell-Derived Cardiomyocytes: Insights into Molecular, Cellular, and Functional Phenotypes. *Circ Res* 117: 80–88.
36. Du X and JM Parent (2015). Using Patient-Derived Induced Pluripotent Stem Cells to Model and Treat Epilepsies. *Curr Neurol Neurosci Rep* 15: 71.
37. Prondzynski M, G Mearini and L Carrier (2019). Gene therapy strategies in the treatment of hypertrophic cardiomyopathy. *Eur J Physiol* 471: 807–815.
38. Maherali N and K Hochedlinger (2008). Guidelines and Techniques for the Generation of Induced Pluripotent Stem Cells. *Cell Stem Cell* 3: 595–605.
39. Olyslaegers DAJ, LMB Desmarets, A Dedeurwaerder, HL Dewerchin and HJ Nauwynck (2013). Generation and characterization of feline arterial and venous endothelial cell lines for the study of the vascular endothelium. *BMC Vet Res* 9: 1.

40. Ocklind A, SYK Yousufzai, S Ghosh, M Coca-Prados, J St Jernschantz and AA Abdel-Latif (1995). Immortalization of cat iris sphincter smooth muscle cells by SV40 virus: Growth, morphological, biochemical and pharmacological characteristics. *Exp Eye Res* 61: 535–545.
41. Miller DG, MA Adam and AD Miller (1990). Gene transfer by retrovirus vectors occurs only in cells that are actively replicating at the time of infection. *Mol Cell Biol* 10: 4239–4242.
42. Nawrocka D, K Kornicka, J Szydlarska and K Marycz (2017). Basic Fibroblast Growth Factor Inhibits Apoptosis and Promotes Proliferation of Adipose-Derived Mesenchymal Stromal Cells Isolated from Patients with Type 2 Diabetes by Reducing Cellular Oxidative Stress. *Oxid Med Cell Longev* 2017: 1–22.
43. Riaz N, SL Wolden, DY Gelblum and J Eric (2016). Mechanisms underlying the formation of induced pluripotent stem cells. *118*: 6072–6078.
44. Esteban MA, T Wang, B Qin, J Yang, D Qin, J Cai, W Li, Z Weng, J Chen, S Ni, K Chen, Y Li, X Liu, J Xu, S Zhang, F Li, W He, K Labuda, Y Song, A Peterbauer, S Wolbank, H Redl, M Zhong, D Cai, L Zeng and D Pei (2010). Vitamin C Enhances the Generation of Mouse and Human Induced Pluripotent Stem Cells. *Cell Stem Cell* 6: 71–79.
45. Lorenzo IM, A Fleischer and D Bachiller (2013). Generation of Mouse and Human Induced Pluripotent Stem Cells (iPSC) from Primary Somatic Cells. *Stem Cell Rev Reports* 9: 435–450.
46. Pei Y, L Yue, W Zhang, Y Wang, B Wen, L Zhong, J Xiang, J Li, S Zhang, H Wang, H Mu, Q Wei and J Han (2015). Improvement in Mouse iPSC Induction by Rab32

Reveals the Importance of Lipid Metabolism during Reprogramming. *Sci Rep* 5: 1–12.

47. Brambrink T, R Foreman, GG Welstead, CJ Lengner, M Wernig, H Suh and R Jaenisch (2008). Sequential expression of pluripotency markers during direct reprogramming of mouse somatic cells. *Cell Stem Cell* 2: 151–159.

48. Olariu V, C Lökvist and K Sneppen (2016). Nanog, Oct4 and Tet1 interplay in establishing pluripotency. *Sci Rep* 6: 1–11.

49. Nichols J and A Smith (2009). Naive and Primed Pluripotent States. *Cell Stem Cell* 4: 487–492.

50. Weinberger L, M Ayyash, N Novershtern and JH Hanna (2016). Dynamic stem cell states: Naive to primed pluripotency in rodents and humans. *Nat Rev Mol Cell Biol* 17: 155–169.

51. Sakaki-Yumoto M, J Liu, M Ramalho-Santos, N Yoshida and R Derynck (2013). Smad2 Is essential for maintenance of the human and mouse primed pluripotent stem cell state. *J Biol Chem* 288: 18546–18560.

52. Gafni O, L Weinberger, AA Mansour, YS Manor, E Chomsky, D Ben-Yosef, Y Kalma, S Viukov, I Maza, A Zviran, Y Rais, Z Shipony, Z Mukamel, V Krupalnik, M Zerbib, S Geula, I Caspi, D Schneir, T Shwartz, S Gilad, D Amann-Zalcenstein, S Benjamin, I Amit, A Tanay, R Massarwa, N Novershtern and JH Hanna (2013). Derivation of novel human ground state naive pluripotent stem cells. *Nature* 504: 282–286.

53. Hirai H, M Firpo and N Kikyo (2012). Establishment of LIF-dependent human iPS cells closely related to basic FGF-dependent authentic iPS cells. *PLoS One* 7: 1–10.

54. Nichols J and A Smith (2009). Naive and Primed Pluripotent States. *Cell Stem Cell*

4: 487–492.

55. Zhang Y, C Wei, P Zhang, X Li, T Liu, Y Pu, Y Li, Z Cao, H Cao, Y Liu, X Zhang and Y Zhang (2014). Efficient reprogramming of naïve-like induced pluripotent stem cells from porcine adipose-derived stem cells with a feeder-independent and serum-free system. *PLoS One* 9: e85089.

56. Fang R, K Liu, Y Zhao, H Li, D Zhu, Y Du, C Xiang, X Li, H Liu, Z Miao, X Zhang, Y Shi, W Yang, J Xu and H Deng (2014). Generation of naive induced pluripotent stem cells from rhesus monkey fibroblasts. *Cell Stem Cell* 15: 488–497.

57. Tai D, P Liu, J Gao, M Jin, T Xu, Y Zuo, H Liang and D Liu (2015). Generation of Arbas Cashmere Goat Induced Pluripotent Stem Cells Through Fibroblast Reprogramming. *Cell Reprogram* 17: 297–305.

58. Kanegi R, S Hatoya, Y Tsujimoto, S Takenaka, T Nishimura, V Wijewardana, K Sugiura, M Takahashi, N Kawate, H Tamada and T Inaba (2015). Production of feline leukemia inhibitory factor with biological activity in *Escherichia coli*. *Theriogenology* 86: 604–611.

59. Hu K (2014). All Roads Lead to Induced Pluripotent Stem Cells: The Technologies of iPSC Generation. *Stem Cells Dev* 23: 1285–1300.

60. Hotta A and J Ellis (2008). Retroviral vector silencing during iPS cell induction: An epigenetic beacon that signals distinct pluripotent states. *J Cell Biochem* 105: 940–948.