1	Changing r	oatterns of	growth in s	a changing	nlanet: h	ow a shift i	n nhenology
T	Changing P		gi un ma	a changing	plance. It	ow a shirt i	n phenology

- 2 affects critical life-history traits in annual fishes
- 3
- 4 Daniel García¹, Carl Smith^{2,3,4}, Emanuel Machín¹, Marcelo Loureiro¹ and Martin
- 5 Reichard^{2,*}
- 6

⁷ ¹Departamento de Ecología y Evolución, Instituto de Biología, Facultad de Ciencias,

- 8 Universidad de la República, Iguá 4225, Montevideo, Uruguay
- 9 ²The Czech Academy of Sciences, Institute of Vertebrate Biology, Květná 8, Brno,
- 10 Czech Republic
- ³Department of Ecology & Vertebrate Zoology, University of Łódź, Banacha 12/16,
- 12 90-237 Łódź, Poland

13 ⁴School of Biology and Bell-Pettigrew Museum of Natural History, University of St

14 Andrews, St Andrews KY16 8LB, UK

15

- 16 ORCID: MR (0000-0002-9306-0074)
- 17 *Corresponding author: M. Reichard: The Czech Academy of Sciences, Institute of
- 18 Vertebrate Biology, Květná 8, 603 65, Brno, Czech Republic, phone: +420 543 422
- 19 522, e-mail: <u>reichard@ivb.cz</u>

20

21 Keywords: desiccation, developmental temperature, drought, killifish, phenology

23 Summary

Under current climate change trends, freshwater wetlands in the temperate
 Neotropics are predicted to undergo major alternations in precipitation regime.

26 2. We investigated consequences of altered seasonal patterns in precipitation on
27 replicated populations of annual killifish, using a field survey. Annual killifish are
28 endemic to ephemeral pools and highly endangered through disruptions to wetlands.

3. In 2015, the Pampa region of western Uruguay experienced an unprecedented midwinter desiccation of temporary pools that naturally contain water from autumn to late
spring and typically only desiccate in summer. In our study area, 72% of desiccated
pools were re-inundated from later rains and annual killifish hatched a second cohort in
some of them (53%).

4. Compared to the first cohort, the second cohort experienced lower water
temperatures during hatching and juvenile development, but higher temperatures during
the adult growth phase. The second cohort of fish developed more rapidly, with 40%
earlier attainment of asymptotic body size (14 vs. 23 weeks) that was significantly
smaller in one species (*Austrolebias bellottii*) but not in the other (*Austrolebias nigripinnis*).

40 5. Rapid juvenile development of second-cohort fish was associated with earlier
41 maturation and greater investment in reproductive tissue. The abbreviated life history
42 was adaptive because second-cohort fish lived shorter lives due to high summer
43 temperatures desiccating pools and extirpating the annual populations.

6. Annual fish demonstrated high developmental plasticity that buffered the negativeconsequences of a severely altered seasonal climatic cycle. The capacity of these annual

- 46 fishes to cope with alternations to their life cycle will depend on the ability of egg banks
- 47 in the sediment to tolerate an increased unpredictability of precipitation.

48 **1. INTRODUCTION**

49 Current climate change trends are rapidly driving modifications to the temporal and 50 spatial distribution of temperature and precipitation across the planet (Trenberth, 2011; 51 Walther et al., 2002). These changes can affect biodiversity at all levels, from individual 52 populations to biomes (Parmesan, 2006). Some of the most striking effects of climate 53 change are alterations in species phenologies; changes in timing of crucial life cycle 54 events. This effect may lead to cascading effects at the community level (Edwards & 55 Richardson, 2004; Gilman et al., 2010), through the responses of interacting species 56 (Bellard et al., 2012). Changes in plant phenology, for example, can affect pollinator 57 populations. Given their mutual dependency, altered plant phenology may ultimately 58 lead to the extinction of both pollinators and plants (Rafferty & Ives, 2010). However, 59 a range of other effects on the relationships between affiliate species can emerge 60 (Rafferty et al., 2014; Hassall et al., 2019), with largely unknown capacity for resilience 61 in some taxa (Hegg et al., 2013).

62 Changes in species phenology may also lead to changes in life history traits, 63 which determine the way an individual spreads its reproduction over its lifetime 64 (Forrest & Miller-Rushing, 2010). For example, delayed hatching time can lead to post-65 hatching development under altered environmental conditions, shifting growth and 66 maturation patterns, with potential consequences for the viability of subsequent 67 generations (Varpe, 2017). In Atlantic salmon (Salmo salar), date of emergence of 68 offspring has consequences for juvenile body size, with a significant decrease in 69 juvenile body size at the end of the growing season in individuals that emerged one 70 week later than the first cohort. This outcome may affect size-selective events, such as 71 winter mortality, predation risk and reproduction (Einum & Fleming, 2000). Other 72 ecological challenges can alter key life-history events. Interspecific resource competition can ultimately affect growth patterns and timing of sexual maturation
(Ferreira-Rodríguez *et al.*, 2018) to a degree that surpasses the effects of climate
(Rolinson *et al.*, 2016). Therefore, it is important to compare the relative importance of
one ecological challenge (climate change) with other potential modulators of lifehistory events.

78 The effects of altered phenology are likely to be especially pronounced in 79 ephemeral environments. Temporary habitats are subject to shifts between alternative 80 habitat states, often predictably following annual cycles of precipitation and 81 temperature (Varpe, 2017). Temporary pools are extreme environments that depend on 82 natural seasonal cycles between aquatic and desiccated phases (Polačik & Podrabsky, 83 2015). Among the organisms exploiting these environments, many are exclusively 84 adapted to the periodic phases of the habitat. In some cases, selection favours adaptation 85 to the regular pattern of aquatic phases, such as annual spring flooding (Lytle & Poff, 86 2004), though other species appear adapted to the unpredictability of erratic events, 87 such as desert flooding (Polačik & Podrabsky, 2015). As climate change modifies 88 precipitation regimes and increases the incidence of temporarily abbreviated rainfall 89 and concomitant extended periods of drought and heavy rainfall (Witze, 2018), 90 temporary pools experience dramatic change in their seasonal patterns of aquatic and 91 desiccated phases (Brooks, 2009). How these changes might alter the viability of 92 populations occupying these environments has yet to be explored.

Annual killifishes are a unique group of vertebrates that is exclusively adapted to live in temporary pools. In the Neotropics and Africa, killifishes (Cyprinodontiformes) have repeatedly adapted to life in ephemeral freshwater habitats (Furness, 2016), with three independent evolutionary transitions to adopt this life history in the family Rivulidae (Furness *et al.*, 2015a), a Neotropical clade of

98 Cyprinodontiformes fishes (Loureiro et al., 2018). Populations of annual fishes persist 99 in desiccated sediment in the form of embryos encased in eggs with a thickened chorion. 100 Development is halted in three diapause stages (Wourms, 1972) when the embryo is 101 particularly resilient to environmental stress (Podrabsky et al., 2001). Fish hatch when 102 their natal pool is filled with water. After hatching they grow rapidly and reach sexual 103 maturity within a few weeks (Vrtílek et al., 2018). Adults reproduce daily and the eggs 104 are deposited into the substrate where they survive until a subsequent aquatic phase. 105 The timing of embryo development is variable, with a marked differential propensity 106 of embryos to enter and leave diapause (Wourms, 1972; Podrabsky & Hand, 1999; 107 Furness et al., 2015b; Polačik et al., 2018), though it is broadly synchronised by 108 environmental cues (Domínguez-Castanedo et al., 2017), including temperature 109 (Podrabsky et al., 2010a), photoperiod (Levels & Denucé 1988) and relative humidity 110 (Podrabsky et al., 2010b).

111 Neotropical annual killifish are highly endangered. The major recognized threat 112 to their populations in southern South America (the Pampas biome) is habitat loss 113 associated with extensive agriculture and floodplain regulation (Volcan & Lanés, 2018). 114 In addition, annual fish depend on specific precipitation patterns and may be strictly 115 adapted to local natural cycles of pool inundation and desiccation. In southern South 116 America, the natural aquatic phase begins in the austral autumn (April), when 117 evaporation decreases and pools retain water from precipitation. The dry phase begins 118 in late spring (November), when evaporation increases and exceeds precipitation, and 119 habitats desiccate. Hence, unlike in the tropical range of annual fishes in Africa 120 (Reichard, 2016) and the Neotropics (Domínguez-Castanedo et al., 2013; Podrabsky et 121 al., 1997), there are no distinct rainy and dry seasons, and precipitation (with substrate

wetting and short-term accumulation of water) is relevant throughout the dry season,though precipitation does not lead to an aquatic phase during the summer months.

124 With recent changes in climatic conditions, we have earlier reported (García et 125 al., 2018) that the lower Río Negro floodplain (Villa Soriano district in western 126 Uruguay, Pampa vegetation region) experienced an unusual desiccation of temporary 127 pools in the middle of the aquatic phase (austral winter) in 2015, followed by an 128 unusually heavy precipitation that re-filled most of the desiccated pools. This event 129 triggered the hatching of a new cohort of annual fish in some pools (García et al., 2018). 130 Here, we tested how the alteration of seasonal patterns affected the growth, maturation 131 and fecundity of these anomalous annual fish populations. The region of South 132 American pampas, to which more than 40 species of annual killifish are endemic 133 (Froese & Pauly, 2018), experienced a moderate increase in temperature and more than 134 30% decrease in precipitation between 1976 and 1990 (Walther et al., 2002). This area 135 belongs to a region with the fastest estimated temperature increase (Loarie et al., 2009). 136 Periods of heavy rains are predicted to increase in the region of between 20 - 50% in 137 the next 90 years with a 3 °C increase in global temperature (Fisher & Knuti, 2015; Witze, 2018). 138

139 We investigated how the growth rate and sexual maturation of two annual 140 killifish species, Austrolebias bellottii and A. nigripinnis, were affected by non-141 seasonal conditions (a second cohort that hatched in mid-winter rather than in autumn) 142 and compared it with the effects of another ecological challenge, interspecific 143 competition. Growth and maturation are crucial life history traits for individual fitness 144 (Stearns, 1992) and population dynamics (Sæther & Bakke, 2000) and this is especially 145 pronounced in short-lived annual fishes (Wootton & Smith, 2015; Vrtílek et al., 2018). 146 We compared the growth and maturation of replicated A. bellottii populations

147 coexisting with A. nigripinnis and replicated A. bellottii populations without intra-guild 148 interspecific competition. Then, as the main goal of the study, we compared the growth 149 and development between the two cohorts of A. bellottii and the timing and size at 150 maturation of both cohorts and for each species. We predicted slower growth and later 151 sexual maturation in populations of A. bellottii coexisting with A. nigripinnis as a result 152 of interspecific competition between species occupying a similar ecological niche 153 (Arim et al., 2010). Based on life history theory, we further predicted that the second 154 cohort (with a shorter expected lifespan) would trade somatic growth for a greater 155 reproductive effort (Figueredo et al., 2006). Thus, individuals of the second cohort were 156 predicted to reach a smaller final size but mature earlier than the first cohort and make 157 a greater investment into reproduction than the first cohort, which was predicted to 158 divert more resources to a longer growth phase resulting in higher lifetime fecundity.

159

160 **2. Methods**

161 **2.1. Study area**

162 The study was conducted in the surroundings of Villa Soriano in Uruguay (33°23'56"S, 163 58°19'13"W) in 2015. This is a wetland area of a typical Pampa biome. It is 164 characterized by low altitude (1-7 m above sea level) and an abundance of shallow temporary freshwater pools. The pools are from 100 to over 2,000 m² in area, with a 165 166 maximum depth of 0.4 m. Annual killifish (three species) are regularly encountered in 167 the pools. Other fish species invade the pools sporadically, following flooding from the 168 adjacent Rio Negro. The study area encompassed approximately 53 km². For a detailed 169 description of the study area see García et al. (2018).

The study is based on sampling in 18 pools (Fig. 1). The annual fish that inhabit
the area are *Austrolebias bellottii*, *A. nigripinnis* and *A. elongatus*. Thirteen pools (72%)

were re-inundated but the second cohort of *A. bellotti* hatched only in 6 of 18 (33%) and *A. nigripinnis* in 7 pools (39%). No second cohort hatched after re-inundation in the 6 pools. The first cohort persisted throughout winter in XXX pools (*A. bellottii*) and YYY pools (*A. nigripinnis*). Given the rare occurrence of *A. elongatus* (0.23% of sampled fish in a total of 17% of the pools), this study focused on the other two species.

177

178 **2.2. Sampling and data collection**

179 Sampling started on 18 May 2015 and lasted until no further fish could be collected or 180 pools were dry (1 January 2016). Sampling was conducted every two weeks, except for 181 a 20-day interval between trip 7 (9–10 Aug) and trip 8 (29 Aug–1 Sep). For all pools, 182 fish hatching dates were estimated from otolith increments and the age of the fish was 183 calculated by adding the number of days between sampling trips. The first cohort 184 hatched in late April and the second cohort hatched in early August in some pools that 185 experienced mid-season desiccation (García et al., 2018). The pools were at least 0.3 186 km apart, except for pools 11D and 12D that were only 0.1 km apart but clearly separate. 187 Detailed descriptions of seasonal dynamics in population density and abundance are 188 described in García et al. (2019). In short, for the period of juvenile growth the pool 189 area was larger in the second cohort, but fish density was comparable between the 190 cohorts. Water temperature data were collected by data loggers (Hobo UA-002-08) 191 deployed in 12 pools on the first day of sampling. The loggers also provided the timing 192 of pool desiccation (Lanés et al., 2016; García et al., 2018).

Sampling was accomplished with a dip-net (200 x 150 mm, 2 mm mesh size)
that was swept parallel to the bottom. All available habitats in the pool were explored.
Sampling effort varied across sites due to wide variation in fish density and pool area.
Sampling effort was recorded as time spent collecting fish (median = 26 min, mean =

197 32, standard deviation = 23.8) and pool surface area. Pool area was measured in situ 198 during each sampling event by walking around the pool boundaries using a portable 199 GPS unit to map the outline of the pool surface (Garmin Etrex10). After sampling, all 200 annual fish collected were photographed on a tray (80 x 80 mm) with a wetted surface 201 and a reference scale. A total of 4,804 A. bellottii and 2,592 A. nigripinnis were 202 measured. Sample size for each pool, species, sex, and sampling event is given in 203 Supporting Information (Table S1). A random sample of 5 females was taken at each 204 sampling to estimate sexual maturation. Fish were euthanized with an overdose of clove 205 oil and stored in 4% formaldehyde for later dissection in the laboratory. All other fish 206 were then returned to the pool. Species and sex were determined from photographs on 207 the basis of external characters (fin size, shape and colouration). Size of fish was 208 measured from photographs using tpsDig2 software (v 2.1) (Rohlf, 2009) from the tip 209 of the snout to the end of the caudal fin (Total Length, TL). In the laboratory, a sample 210 of 5 females was measured for TL, weighed (W, to the nearest of 0.001g), dissected 211 and gonad mass (Wg) measured.

212 2.3. Data analysis

213 All analyses were completed in the R statistical environment (R Core Team, 2018). 214 First, we tested the effects of the presence of A. nigripinnis on the growth patterns of A. 215 bellottii. Austrolebias bellottii was present in all pools but A. nigripinnis occurred only 216 in 55% of pools (10 of 18). We used sex-specific Generalised Additive Mixed Models 217 (GAMM) with a Gaussian error distribution, with Coexistence (factor with two levels) 218 included as a fixed effect. The effect of age was modelled with a smoother term with 5 219 knots (k = 5), defined *a priori* based on the growth trajectory of *Austrolebias* (Volcan 220 et al., 2012) and allowing for potential selective disappearance of larger fish at the end 221 of the season (Reichard et al., 2018). The response variable was body size, measured 222 as TL. To account for dependency in the data from variation among pools, pool identity 223 was included as random effect. Two models were constructed that differed in how the 224 smoother term was modelled. First, a separate smoother was assigned for each 225 treatment level (coexistence with A. nigripinnis or not). Second, the same smoother was 226 used for both treatments. The fit of the models was compared using Akaike Information 227 Criterion (AIC). An AIC difference of >10 is considered a substantial improvement 228 in model fit. GAMM analysis was completed in the gamm4 package (Wood & Scheipl, 229 2017).

230 Similarly, we compared whether coexistence with A. nigripinnis affected time 231 to sexual maturity in A. bellottii females. Because a high proportion of fish were 232 immature, data were modelled using a zero-altered (hurdle) model, using R-INLA 233 (Rue et al., 2017). Zero-altered models are partitioned into two parts, with a binary 234 process modelling zeros and positive counts, and a second process modelling only 235 positive counts using a zero-truncated model (Hilbe, 2014). We modelled zero-236 truncated data using a Gaussian distribution (Zuur et al., 2014). This approach enabled 237 us to separately identify the environmental variables that predicted maturity (binary 238 part) and investment in gonad mass when gonads were present (zero-truncated part). 239 Fixed factors were Coexistence (two levels), body size (TL, log-transformed, 240 continuous predictor) and their interaction. In the model, the interaction between 241 Coexistence and TL measured the contribution of species coexistence to the timing of 242 sexual maturation relative to body size. This analysis was based on a contrast between 243 6 populations of A. bellottii without A. nigripinnis and 5 populations of A. bellottii 244 competing with A. nigripinnis, with four populations dropped during data exploration 245 due to the small number of females analysed (≤ 5).

246 The main analysis compared growth trajectories and sexual maturation of fish 247 from the first and second cohort, using the same approach as for testing the effect of 248 coexistence. We predicted, a priori, that the two species and sexes within species, 249 would differ in body size (Costa, 2006) and, consequently, we separated them before 250 analysis. We fitted sex-specific GAMMs with a Gaussian error distribution that 251 included the factor Cohort (two levels). Given that the coexistence of A. bellottii with 252 A. nigripinnis had no effect on growth and sexual maturation, this factor was not 253 included in the analysis. We modelled age either with separate smoothers for each 254 cohort (model 1) or a common smoother for both cohorts (model 2), with k = 5 in both 255 models. The fit of the two models was compared using AIC. The same analysis was 256 performed for both fish species.

To formally test the differences in the growth rates (rather than size at age) among cohorts, we calculated Specific Growth Rates (SGR) using the function:

259
$$SGR = (\ln(TL_x) - \ln(TL_{x-1}) \times \frac{100}{N_{days}})$$

Where TL_x is the Total Length for the date x, TL_{x-1} is the Total Length for the previous 260 261 date and N_{days} is the number of days between each date. SGR was estimated for each 262 species and each pool separately. We tested whether SGR differed between cohorts by 263 fitting a Gaussian Linear Model (LM) with age interval, cohort and their interaction as 264 factors. The response variable was population-specific SGR. Given the shorter lifespan 265 of the second cohort, we truncated age intervals for the first cohort to permit a full 266 interaction between the two factors. This analysis was completed for A. bellottii only 267 because we had insufficient estimates of population-specific SGR for the first cohort of 268 A. nigripinnis. For visualization, we extended estimated SGR for the first cohort over 269 their entire lifespan.

While growth curves are commonly used in analysing growth data, they suffer from temporal dependency. To formally accommodate temporal dependency in the data, we further modelled fish size using a random walk (RW2) trend model in a Bayesian framework using Integrated Nested Laplace Approximation (R-INLA; Rue *et al.*, 2017). Fish size was fitted for fish age following a gamma distribution with a log link function. The model was fitted as:

276
$$TL_{iik} \sim N(\mu_{iik}, \tau)$$

277
$$E(TL_{ijk}) = \mu_{ijk} \text{ and } var(TL_{ijk}) = \frac{\mu_{ijk}^2}{\tau}$$

$$\log\left(\mu_{ijk}\right) = \eta_{ijk}$$

279
$$\eta_{ijk} = \beta_1 + \beta_2 \times Sex_{ijk} \times \beta_3 \times Coh_{ijk} + \beta_4 \times Area_{ijk} + Age_j + Pool_k$$

$$280 \qquad \qquad Age_j \sim N(0, \sigma_{Age}^2)$$

281
$$Pool_k \sim N(0, \sigma_{Pool}^2)$$

282 Where TL_{ijk} is the size of fish *i* of age (Age) *j*. The variable Sex_{ijk} is a categorical 283 covariate with two levels (males, females) and Coh_{ijk} is a categorical covariate with two 284 levels (first or second cohort). The model also contained a linear effect for pool size 285 (Areaiik) as its extent varied over the season. The random intercept Pool was included 286 in the model to introduce a correlation structure between observations for fish collected 287 from the same pool with variance σ^2 , distributed normally and equal to 0. Uniform 288 default priors were put on model parameters and fitted estimates of fish size matched 289 observed data.

Time to sexual maturation and investment in reproduction were compared between cohorts using data on gonad mass (Wg) fitted with a zero-adjusted model with a gamma distribution (ZAG), like that used for interspecific competition contrasts. All final models were checked for overdispersion and model mis-specification followingthe guidelines of Zuur & Ieno (2016).

295

3. RESULTS

297 **3.1. Effect of interspecific competition**

298 There was no major effect of inter-specific competition on the growth of A. bellottii 299 (GAMM, males: F = 0.12, P = 0.77; females: F = 1.27, P = 0.260), with a clearly inferior 300 fit of the separate-smoothers model for females ($\Delta AIC = +256$). In males, the model 301 with separate smoothers for males provided a better fit ($\Delta AIC = -10.3$), though the principal difference was that the separate-smoothers model captured differences in size-302 303 selective mortality at the terminal phase of life rather than different growth rates among 304 A. bellottii populations coexisting with A. nigripinnis (Fig. 2). There was no effect of 305 coexistence with A. nigripinnis on the timing of sexual maturity of A. bellottii females (binary part of the model) or their allocation to gonads (zero-truncated part). Female A. 306 307 bellottii maturation and allocation to reproduction depended on body size but the 308 scaling of the relationship did not differ between coexistence treatment levels (Table 309 1).

310 **3.2. Differences between the two cohorts**

311 **3.2.1. Water temperature**

The first cohort experienced a temperature regime typical of temporary pools in the region. Water temperature decreased from warm (>20°C) to cold (<10°C) over the early life phase, remained relatively cold until the age of approximately 100 days and slowly climbed to 20°C again over the subsequent 100 days (Fig. 3). The second cohort hatched in relatively colder water that steadily increased in temperature and never fell
below 10°C (Fig. 3).

318 **3.2.2.** Growth trajectories

319 There were clear differences in growth trajectories of the first and second cohorts of 320 both species, except for A. nigripinnis females. The models with separate smoothers for 321 each cohort were clearly superior in A. bellottii males ($\Delta AIC = -118$), A. bellottii 322 females ($\Delta AIC = -246$) and A. nigripinnis males ($\Delta AIC = -39$), but only marginally in 323 A. nigripinnis females ($\Delta AIC = -2.52$). The growth curves were characterised by earlier 324 attainment of the growth asymptote in the second cohort (approximately 100 days) than 325 in the first cohort (approximately 150-170 days) (Fig. 4), resulting in a larger mean 326 asymptotic size in A. bellottii (males: F = 19.79, P < 0.001, Fig. 4a; females: F = 16.16, P < 0.001; Fig. 4b) but not *A. nigripinnis* (Fig. 4c, 4d). 327

Comparing specific growth rates (SGR) confirmed a steeper growth trajectory of second cohort fish over age-matched categories (LM: $F_{1,71} = 4.67$, P = 0.024) and much stronger decline in the SGR with age in the second cohort fish (age by cohort interaction: $F_{1,71} = 12.42$, P = 0.001; age: $F_{1,71} = 3.28$, P = 0.074) (Fig. 5). Overall, SGRs were initially higher in the second cohort but declined sharply with age. In contrast, the growth rates of the first cohort were relatively stable, though they varied among populations (Fig. 5).

335 **3.2.3. Body size**

Fitting a common smoother to growth data for *A. bellottii* using a gamma random walk trend model fitted using INLA demonstrated that sex differences in body size were higher in the second cohort (sex by cohort interaction in Table 2) and quantitatively confirmed that the second cohort grew more quickly (Cohort effect in Table 2).

340 **3.2.4. Sexual maturation**

Female maturation was strongly positively related to age (binomial occurrence model) with a steeper increase in the second-cohort females (Table 3a), implying that secondcohort fish reached sexual maturation at a younger age. In addition, the frequency model (magnitude of gonadal investment) demonstrated that the second-cohort females invested more heavily in gonad mass. Greater gonad mass was also recorded from pools with a larger area. As expected, gonad mass was positively associated with female age (Table 3b).

348

349 **4. Discussion**

350 The primary effects of climate change are alterations in precipitation and temperature 351 dynamics which have profound implications for species phenologies (Parmesan, 2006; 352 Ficetola & Maiorano, 2016). However, responses to climate change and its 353 consequences potentially vary greatly among species (Both et al., 2009; Colchero et al., 354 2019) and among particular interspecific interactions (Gilman et al., 2010; Kordas et 355 al., 2011; Hassall et al., 2019). We demonstrated previously that Austrolebias annual 356 fishes are able to hatch a new cohort in response to a secondary inundation following 357 an unexpected mid-winter (July) pool desiccation, though a second cohort hatched only 358 in a subset of pools (García et al., 2018). In the present study, we tested the 359 developmental consequences of late season hatching in an environment with a non-360 optimal temperature regime in A. bellottii. Overall, the changes associated with late 361 hatching demonstrated the capacity of A. bellottii to cope with shifted seasonality and 362 to adjust their life history to maximize reproductive success. Following non-seasonal 363 desiccation and subsequent re-filling of pools, 46% (6 of 13) A. bellottii populations

364 hatched a second cohort. The second-cohort juveniles developed in colder water (until 365 the age of approximately 50 days) but sustained accelerated growth and more rapid sexual maturation, along with a greater investment in fecundity. While this came at the 366 367 cost of smaller final size, the response was adaptive since life expectancy (and actual 368 lifespan) in the second cohort was shorter. The magnitude of response was substantial, 369 with no effects on growth and sexual maturation in populations subjected to another 370 ecological challenge, interspecific competition with A. nigripinnis, a related annual 371 killifish species that coexisted in several pools with A. bellottii. Austrolebias nigripinnis 372 also experienced accelerated growth in the second cohort, though the effect was 373 marginal in A. nigripinnis females and their asymptotic size was not smaller.

374 Differences in growth rate, final size and sexual maturation were most likely 375 shaped by the different temperature dynamics experienced by juvenile and adult A. 376 bellottii. The first cohort experienced typical seasonal dynamics in water temperature, 377 with hatching and the juvenile period during warm-water conditions in the austral 378 autumn (April), followed by a gradual decrease in temperature with adults living in a 379 low temperature environment during winter and experiencing increased mortality in 380 late spring (November) as the temperature again increased (García et al., 2019). Those 381 conditions are typical for most Austrolebias populations (Lanés et al., 2016). The 382 second cohort hatched in cold water with the temperature steadily increasing over the 383 season and exposing young adults (5 months old) to ambient temperatures exceeding 384 25°C (Fig. 3), which is considered the upper limit for the long-term survival of 385 Austrolebias (Lanés et al., 2016). In addition, the cohorts experienced different 386 photoperiods. While both cohorts hatched in a photoperiod of 10.5-11 hours of daylight, 387 this period decreased by 1 hour over two months of juvenile growth for the first cohort, 388 but increased by 2 hours for the second cohort, respectively (www.sunrise-sunset.org).

Increasing water temperature and photoperiod might have also increased productivity of the pools, further stimulating more rapid growth of the second cohort. Pools were generally larger after the second inundation, but killifish population densities were comparable (García *et al.*, 2019).

393 Laboratory studies have demonstrated that annual fishes are sensitive to 394 temperature regimes (Hsu & Chiu, 2009; Volcan et al., 2012; da Fonseca et al., 2013). 395 Austrolebias adloffii, a related species from the same region, exhibited a decreased 396 growth rate and shorter lifespan when a natural decrease in water temperature 397 fluctuation was interrupted. After rearing juveniles at 22°C, a decrease in temperature 398 (to 16°C) was associated with longer lifespan and sustained growth while control fish 399 that remained at 22°C ceased growing (Liu & Walford, 1966). Our field data are 400 consistent with laboratory studies of Austrolebias that reported earlier sexual maturity 401 and more rapid growth at higher temperature (25°C compared to 15°C in Austrolebias 402 viarius, and 22°C compared to 16°C in Austrolebias nigrofasciatus) (Errea & Danulat, 403 2001; Volcan et al., 2012). However, the positive correlation between water 404 temperature and juvenile development is reversed when temperature exceeds an 405 optimum of 21-24°C in Austrolebias wolterstorffi (Fonseca et al., 2013). High ambient 406 temperature during the juvenile growth phase comes at the cost of shorter lifespan 407 (Errea & Danulat, 2001). Thermal optima of Austrolebias are age-specific and 408 persistent high ambient temperature is costly to adult fish (Walford & Liu, 1965; Liu & 409 Walford, 1966).

We observed selective disappearance of larger individuals from populations during the terminal phases of growth (Fig. 4), though this may partly indicate selective predation (Reichard *et al.*, 2018) or limited endurance of large males contingent upon their investment in courtship and reproductive activity (Passos *et al.*, 2014) rather than 414 from a direct trade-off between rapid growth and longevity. Hence, while longevity of 415 annual fishes is associated with lower ambient temperature (Walford & Liu, 1965; 416 Valenzano et al., 2006; Hsu & Chiu, 2009), A. bellottii can cope with an altered 417 seasonal trend in water temperature and appears able to respond with an adaptive 418 increase in developmental rate and higher reproductive allocation. Second-cohort 419 females invested in early reproduction with earlier sexual maturation and greater 420 allocation to gonad mass, at the expense of a longer growth phase and, hence, greater 421 investment in future reproduction. This outcome is concordant with predictions from 422 life-history theory (Stearns, 1992), with cues indicating shorter life expectancy 423 probably based on ambient temperature.

424 Populations of annual fishes survive drying of their environment in egg banks 425 deposited in desiccated pool substrate (Polačik & Podrabsky, 2015). Propagule banks 426 are essential for the long-term persistence of communities in temporary environments 427 (Brock et al., 2003). In annual fishes, egg development is temporally staggered due to 428 facultative diapause stages. After fertilization, the embryos initially develop for a few 429 days and may be halted in Diapause I in the anoxic conditions of decaying pool 430 substrate. Upon resumption of development, embryos may respond to desiccation by 431 entering Diapause II and continue development if the substrate is moistened (Reichard 432 & Polačik, 2019). Importantly, different embryos respond to external cues to a different 433 degree (Furness, 2016; Polačik et al., 2017), resulting in the egg bank consisting of 434 embryos at various stages of development. In the study pools, hatching of the second 435 cohort was possible because the egg banks contained embryos ready to hatch at the time 436 of secondary inundation. While we term that group a new cohort, we acknowledge that 437 the fish may have hatched from eggs that were laid during the first inundation and may 438 effectively represent a second "generation". Interestingly, the length of embryonic

439 development has important consequences for post-hatching life. In African annual 440 killifish, rapidly developing embryos hatched to individuals that grew more quickly 441 (and were larger than individuals from slowly developing embryos from the age of 6 442 days), but lived shorter lives, and attained a smaller final size (Polačik et al., 2014). 443 While in that experiment, all fish experienced identical environmental conditions (i.e., 444 temperature, food supply, density), the outcomes were consistent with our results from 445 the wild. This finding suggests that there is an interplay between environmental 446 conditions and intrinsic developmental (potentially epigenetic) processes that give rise 447 to the expressed phenotype.

448 An outstanding question is how frequent the mid-season desiccation of 449 temporary pools in the pampas region was in the past, and how reliably the alternation 450 of flooding regime matches changes in the pattern of precipitation. While we have 451 shown that annual fish populations can respond to these changes with a secondary 452 hatching, a high frequency of mid-season desiccation could rapidly exhaust egg bank 453 reserves. Our dataset for the study region only encompasses 10 years of precipitation 454 and temperature data (data provided by National Meteorology Institute, INUMET). 455 This time series demonstrates unusually dry conditions from February to July 2015 456 (which resulted in desiccation) (Supporting Information, Fig. S1) even in the context 457 of the already less predictable rainfall pattern in the last decade (Ding et al., 2008). We 458 acknowledge that it is critical to understand whether the observed mid-season drought 459 was an exceptional and isolated event or represents a climatic trend and if so, how 460 repeated droughts will affect existing annual fish egg banks.

While climate change can alter the ecology of many species (Walther *et al.*,
2002; Evans & Wallenstein, 2014), potentially driving some towards extinction
(Thomas *et al.*, 2004), we show that *A. bellottii*, a representative species of a highly

464 endangered group of annual fishes, appears capable of coping with a sporadic seasonal 465 shift in rainfall patterns. This finding indicates that environmentally-triggered 466 phenotypic plasticity of life-history traits expressed by A. bellottii permits maintenance 467 of viable populations despite altered phenology (Nylin & Gottard, 1998, Colchero et 468 al., 2019). In fact, A. bellottii has a relatively wide distribution and some populations 469 reach the Río Paraguay basin in northern Argentina (García et al., 2012) where 470 precipitation seasonality is different to the study region and fish hatch when the water 471 temperature is warmer. Phenotypic plasticity is important for forecasting species 472 distributions in a climate change scenario (Valladares et al., 2014; Duputié et al., 2015) 473 and this is especially important for species with limited dispersal abilities such as annual 474 fishes (García et al., 2012).

475 Austrolebias species are the top predators in their communities (Laufer et al., 476 2009) and their potential disappearance from ephemeral pools may lead to a cascade of 477 effects (Paine, 1966; Baum & Worm, 2009) with unknown consequences for 478 community structure. While the primary effects of climate change (alternations in 479 precipitation and temperature dynamics) affect phenologies of certain species 480 (Parmesan, 2006; Ficetola & Maiorano, 2016), downstream consequences vary greatly 481 among communities (Both et al., 2009; Gilman et al., 2010; Kordas et al., 2011; Hassell 482 et al., 2019). For example, in the plankton communities of the North Sea, phenology of 483 primary producers (diatoms) is driven by photoperiod and hence unaffected by elevated 484 temperature while phenology of secondary (zooplankton) and tertiary (fish larvae) 485 consumers have moved forward by 10-45 days over a 45-year study period (Edwards 486 & Richardson, 2004). The resulting mismatch in trophic cascade has been implicated 487 in the decline of marine pelagic production, including commercially important fish 488 species (Edwards & Richardson, 2004). In contrast, a shift in the peak abundance of caterpillars was successfully compensated by modulation of egg-laying date and other
life history characteristics in pied flycatchers (*Ficedula hypoleuca*) for which
caterpillars represent the main food source (Both & Visser, 2005).

492 Freshwater wetlands inhabited by Austrolebias populations are important for 493 ecosystem function, including water purification and carbon retention (Baron et al., 494 2002). In our study region, freshwater wetlands inhabited by Austrolebias populations 495 are utilized for cattle ranching, often in proximity to agricultural plots. While cattle 496 ranching has no apparent negative effect on temporary pools, and Austrolebias 497 populations may even benefit from cattle ranching (Volcan et al., 2016), the major 498 threat to Austrolebias fishes is an expansion of rice or soybean cultivation (Volcan et 499 al., 2016; Volcan & Lanés, 2018). In Uruguay, agricultural development is primarily 500 associated with soybean, wheat and other crop, which have expanded as new 501 technologies have become available (Arbeletche & Gutiérrez, 2010; Dellazoppa, 2014). 502 The use of novel herbicides, for example, appears to alter the ability of Austrolebias to 503 cope with high water temperature, decreases their fecundity and affects diapausing 504 stages in the egg bank (Zebral et al., 2018). In southern Brazil, rice cultivation alters 505 the natural flooding regime, with consequences reported for anuran and invertebrate 506 communities (Cuhna et al., 2015; Ávila et al., 2015) and with likely negative impacts 507 on annual fish populations. At present, changes to land use appear to overshadow any 508 negative consequences of climate change for Austrolebias fishes (Volcan & Lanés, 509 2018), though the interaction of these effects, which are difficult to predict, may 510 represent their greatest threat.

511

512 Acknowledgements

513	Financial supp	ort came from A	Agencia Nacional d	le Investigación e	Innovación ((ANII
-----	----------------	-----------------	--------------------	--------------------	--------------	-------

- 514 FCE 2013 100380) to DG and Comisión Sectorial de Investigación Científica (CSIC
- 515 I+D 2014_7) to ML. DG holds a PhD scholarship from Agencia Nacional de
- 516 Investigación e Innovación (ANII, POS_NAC_2014_102280) and and from Comisión
- 517 Académica de Posgrado (CAP, UdelaR, BFPD_2018_1#26027733). DG and ML
- 518 belongs to Sistema Nacional de Investigadores (SNI) of Uruguay. MR was funded
- 519 through CSF 18-26284S. All local and national legal and ethical guidelines were
- 520 followed. The research was approved under reference number 240011-001885-13. We
- 521 thank to Patricia Magariños, Diego Díaz, Esteban Ortiz, Matej Polačik, Radim Blažek
- and Milan Vrtílek for the help in the field. We also thank to local land owners for
- 523 access on their properties, and local authorities of Villa Soriano for generous support
- 524 for the project. The authors declare no conflict of interest.
- 525

526 Data Availability Statement

527 Primary data are deposited at Figshare (doi: xxxx).

528 **References**

- 529 Arbeletche, P., & Gutiérrez, G. (2010). Crecimiento de la agricultura en Uruguay:
- 530 exclusión social o integración económica en redes. *Pampa* 6, 113-138.
- 531 Arim, M., Abades, S.R., Laufer, G., Loureiro, M., & Marquet, P. A. (2010). Food web
- 532 structure and body size: trophic position and resource acquisition. *Oikos*, 119(1),
- 533 147-153.
- 534 Ávila, A. C., Boelter, T., dos Santos, R. M., Stenert, C., Würdig, N. L., Rocha, O., &
- 535 Maltchik, L. (2015). The effects of different rice cultivation systems and ages on
- 536 resting stages of wetland invertebrates in southern Brazil. *Marine and Freshwater*
- 537 *Research*, 66(3), 276-285.
- 538 Baron, J. S., Poff, N. L., Angermeier, P. L., Dahm, C. N., Gleick, P. H., Hairston Jr.,
- N. G., ... Steinman, A. D. (2002). Meeting ecological and societal needs for
 freshwater. *Ecological Applications*, 12(5), 1247-1260.
- 541 Baum, J. K., & Worm, B. (2009). Cascading top down effects of changing oceanic
 542 predator abundances. *Journal of Animal Ecology*, 78(4), 699-714.
- 543 Bellard, C., Bertelsmeier, C., Leadley, P., Thuiller, W., & Courchamp, F. (2012).
- 544 Impacts of climate change on the future of biodiversity. *Ecology Letters*, 15(4),
- 545 365-377. DOI: 10.1111/j.1461-0248.2011.01736.x
- 546 Blažek, R., Polačik, M., & Reichard, M. (2013). Rapid growth, early maturation and
- 547 short generation time in African annual fishes. *EvoDevo*, 4(1), 1–7. DOI:
- 548 10.1186/2041-9139-4-24
- 549 Both, C., Van Asch, M., Bijlsma, R. G., Van Den Burg, A. B., & Visser, M. E.
- 550 (2009). Climate change and unequal phenological changes across four trophic
- 551 levels: Constraints or adaptations? *Journal of Animal Ecology*, 78(1), 73–83. DOI:
- 552 10.1111/j.1365-2656.2008.01458.x

- 553 Both, C., & Visser, M. E. (2005). The effect of climate change on the correlation
- between avian life history traits. *Global Change Biology*, 11(10), 1606-1613. DOI:

555 10.1111/j.1365-2486.2005.01038.x

- 556 Brock, M. A., Nielsen, D. L., Shiel, R. J., Green, J. D., & Langley, J. D. (2003).
- 557 Drought and aquatic community resilience: The role of eggs and seeds in
- sediments of temporary wetlands. *Freshwater Biology*, 48(7), 1207–1218. DOI:
- 559 10.1046/j.1365-2427.2003.01083.x
- 560 Brooks, R. T. (2009). Potential impacts of global climate change on the hydrology
- and ecology of ephemeral freshwater systems of the forests of the northeastern
- 562 United States. *Climatic Change*, 95(3-4), 469-483.
- 563 Cohen, J. M., Lajeunesse, M. J., & Rohr, J. R. (2018). A global synthesis of animal
- 564 phenological responses to climate change. *Nature Climate Change*, 8(3), 224–228
- 565 DOI: 10.1038/s41558-018-0067-3
- 566 Colchero, F., Jones, O., Conde, D. A., Hodgson, D., Zajitschek, F., Schmidt, B. R., ...
- 567 Gaillard, J. (2019). The diversity of population responses to environmental change.
- 568 *Ecology Letters*, 22, 342-353. DOI: 10.1111/ele.13195
- 569 Costa, W. J. E. M. (2006). The South American annual killifish genus Austrolebias
- 570 (Teleostei: Cyprinodontiformes: Rivulidae): phylogenetic relationships, descriptive
- 571 morphology and taxonomic revision. *Zootaxa*, (1213), 1-162.
- 572 Cunha, G. G., Stenert, C., Rolon, A., Mundstock, C. M., & Maltchik, L. (2015). Does
- 573 intensification of the rice cultivation cycle influence anuran diversity in rice fields?
- 574 *Wetlands ecology and management*, 23(4), 695-705.
- 575 Dellazoppa, R. (2014). Agro La revolución sorprendente. Montevideo, Editorial Fin
- 576 de Siglo.

- 577 Ding, R., Li, J., & Ha, K. J. (2008). Trends and interdecadal changes of weather
- 578 predictability during 1950s–1990s. *Journal of Geophysical Research:*
- 579 *Atmospheres*, 113(D24). DOI: 10.1029/2008JD010404
- 580 Domínguez-Castanedo, O., Mosqueda-Cabrera, M.Á., & Valdesalici, S. (2013). First
- 581 observations of annualism in *Millerichthys robustus* (Cyprinodontiformes:
- 582 Rivulidae). Ichthyological Exploration of Freshwaters, 24, 15-20.
- 583 Domínguez-Castanedo, O., Valdesalici, S., & Rosales-Torres, A. M. (2017).
- 584 Developmental ecology of annual killifish *Millerichthys robustus*
- 585 (Cyprinodontiformes: Cynolebiidae). *Developmental Dynamics*, 246(11), 802-806.
- 586 Duputié, A., Rutschmann, A., Ronce, O., & Chuine, I. (2015). Phenological plasticity
- 587 will not help all species adapt to climate change. *Global Change Biology*, 21(8),
- 588 3062-3073.
- 589 Edwards, M., & Richardson, A. J. (2004). The impact of climate change on the
- 590 phenology of the plankton community and trophic mismatch. *Nature*, 430(1977),
 591 881–884.
- 592 Einum S., & Fleming, I. A. (2000). Selection against late emergence and small
- 593 offspring in Atlantic salmo (*Salmo salar*). *Evolution*, 54(2), 628–639.
- 594 Evans, S. E., & Wallenstein, M. D. (2014). Climate change alters ecological strategies
 595 of soil bacteria. *Ecology Letters*, 17(2), 155-164.
- 596 Ferreira-Rodríguez, N., Sousa, R., & Pardo, I. (2018). Negative effects of Corbicula
- *fluminea* over native freshwater mussels. *Hydrobiologia*, *810*(1), 85-95.
- 598 Ficetola, G. F., & Maiorano, L. (2016). Contrasting effects of temperature and
- 599 precipitation change on amphibian phenology, abundance and performance.
- 600 *Oecologia*, 181(3), 683-693.

- 601 Figueredo, A. J., Vásquez, G., Brumbach, B. H., Schneider, S. M. R., Sefcek, J. A.,
- Tal, I. R., ... Jacobs, W. J. (2006). Consilience and Life History Theory: From
- 603 genes to brain to reproductive strategy. Developmental Review, 26(2006), 243–
- 604 275. DOI: 10.1016/j.dr.2006.02.002
- 605 Fischer, E. M., & Knutti, R. (2015). Anthropogenic contribution to global occurrence
- 606 of heavy-precipitation and high-temperature extremes. *Nature Climate Change*,

607 5(6), 560-565.

- 608 Fonseca, A. P. D., Volcan, M. V., Sampaio, L. A., Romano, L. A., & Robaldo, R.B.
- 609 (2013). Growth of Critically Endangered annual fish Austrolebias wolterstorffi
- 610 (Cyprinodontiformes: Rivulidae) at different temperatures. *Neotropical*
- 611 *Ichthyology*, 11(4), 837-844.
- 612 Forrest, J., & Miller-Rushing, A. J. (2010). Toward a synthetic understanding of the
- 613 role of phenology in ecology and evolution. *Philosophical Transactions of the*
- 614 *Royal Society B: Biological Sciences*, 365(1555), 3101–3112. DOI:
- 615 10.1098/rstb.2010.0145
- 616 Froese, R., & Pauly, D. (2018). FishBase, version February 2018.
- 617 Furness, A. I. (2016). The evolution of an annual life cycle in killifish: adaptation to
- 618 ephemeral aquatic environments through embryonic diapause. *Biological Reviews*
- 619 of the Cambridge Philosophical Society, 91(3), 796–812. DOI: 10.1111/brv.12194
- 620 Furness, A. I., Reznick, D. N., Springer, M. S., & Meredith, R. W. (2015).
- 621 Convergent evolution of alternative developmental trajectories associated with
- 622 diapause in African and South American killifish. *Proceedings of the Royal Society*
- 623 *B: Biological Sciences*, 282(1802), 1–9. DOI: 10.1098/rspb.2014.2189

- 624 García, D., Loureiro, M., Machín, E., & Reichard, M. (2018). Phenology of three
- 625 coexisting annual fish species: seasonal patterns in hatching dates. *Hydrobiologia*,
 626 809(1), 323-337.
- 627 García, D., Loureiro, M., Machín, E., & Reichard, M. (2019). Species coexistence and
- 628 sex ratio dynamics in annual fish assemblages in the lower Río Uruguay basin
- 629 Environmental Biology of Fish. DOI: 10.1007/s10641-019-00854-x
- 630 García, G., Gutiérrez, V., Vergara, J., Calviño, P., Duarte, A., & Loureiro, M. (2012).
- 631 Patterns of population differentiation in annual killifishes from the Paraná-
- 632 Uruguay-La Plata Basin: The role of vicariance and dispersal. *Journal of*
- 633 *Biogeography*, 39(9), 1707–1719. DOI: 10.1111/j.1365-2699.2012.02722.x
- 634 Gilman, S. E., Urban, M. C., Tewksbury, J., Gilchrist, G. W., & Holt, R. D. (2010). A
- 635 framework for community interactions under climate change. *Trends in Ecology*
- 636 *and Evolution*, 25(6), 325–331. DOI: 10.1016/j.tree.2010.03.002
- 637 Hassall, C., Billington, J., & Sherratt, T. N. (2019). Climate-induced phenological
- 638 shifts in a Batesian mimicry complex. *Proceedings of the National Academy of*
- 639 *Sciences*, 116(3), 929-933. DOI: 10.1073/pnas.1813367115
- 640 Hegg, J. C., Kennedy, B. P., Chittaro, P. M., & Zabel, R. W. (2013). Spatial
- 641 structuring of an evolving life-history strategy under altered environmental
- 642 conditions. *Oecologia*, 172(4), 1017–1029. DOI: 10.1007/s00442-012-2564-9
- 643 Hilbe, J. M. (2014). Bayesian Count Models. Wiley StatsRef: Statistics Reference
- 644 *Online*, 1-13. DOI: 10.1002/9781118445112.stat07963
- Hsu, C. Y., & Chiu, Y. C. (2009). Ambient temperature influences aging in an annual
- 646 fish (*Nothobranchius rachovii*). *Aging cell*, 8(6), 726-737.
- 647 Kordas, R. L., Harley, C. D., & O'Connor, M. I. (2011). Community ecology in a
- 648 warming world: the influence of temperature on interspecific interactions in marine

- 649 systems. Journal of Experimental Marine Biology and Ecology, 400(1-2), 218-226.
- 650 DOI: 10.1016/j.jembe.2011.02.029
- 651 Lanés, L. E. K., Godoy, R. S., Maltchik, L., Polačik, M., Blažek, R., Vrtílek, M.,
- 652 Reichard, M. (2016). Seasonal dynamics in community structure, abundance, body
- size and sex ratio in two species of Neotropical annual fishes. Journal of Fish
- 654 *Biology*, 89, 2345-2364.
- 655 Laufer, G., Arim, M., Loureiro, M., Piñeiro-Guerra, J. M., Clavijo-Baquet, S., &
- Fagúndez, C. (2009). Diet of four annual killifishes: an intra and interspecific

657 comparison. *Neotropical Ichthyology* 7, 77-86.

- 658 Levels P. J., & Denucé, J. M. (1988). Intrinsic variability in the frequency of
- 659 embryonic diapauses of the annual fish Nothobranchius korthausae, regulated by
- 660 light: dark cycle and temperature. *Environmental Biology of Fishes* 22, 211–223.
- 661 Liu, R. K., & Walford, R. L. (1966). Increased growth and life-span with lowered
- ambient temperature in the annual fish, *Cynolebias adloffi. Nature*, 212(5067),
- 663 1277.
- Loarie, S. R., Duffy, P. B., Hamilton, H., Asner, G. P., Field, C. B., & Ackerly, D. D.
- 665 (2009). The velocity of climate change. *Nature*, 462, 1052-1055. DOI:
- 666 10.1038/nature08649
- 667 Loureiro, M., de Sá, R., Serra, W. S., Alonso, F, Lanés, L. E. K., Volcan, M. V., ...
- 668 Garcia, G. (2018). Review of the family Rivulidae (Cyprinodontiformes,
- Aplocheiloidei) and a molecular and morphological phylogeny of the annual fish
- 670 genus *Austrolebias* Costa 1998. *Neotropical Ichthyology*, 16(3). DOI:
- 671 10.1590/1982-0224-20180007
- 672 Lytle, D. A., & Poff, N. L. (2004). Adaptation to natural flow regimes. Trends in
- 673 *Ecology & Evolution*, *19*(2), 94-100.

- 674 Mathias, A., & Chesson, P. (2013). Coexistence and evolutionary dynamics mediated
- by seasonal environmental variation in annual plant communities. *Theoretical*
- 676 *Population Biology*, 84(1), 56–71. DOI: 10.1016/j.tpb.2012.11.009
- Nylin, S., & Gotthard, K. (1998). Plasticity in life-history traits. *Annual review of entomology*, 43(1), 63-83.
- 679 Paine, R. T. (1966). Food web complexity and species diversity. *The American*
- 680 *Naturalist*, 100(910), 65-75.
- 681 Parmesan, C. (2006). Ecological and evolutionary responses to recent climate change.
- 682 *Annual Review of Ecology, Evolution, and Systematics*, 37(1), 637–669. DOI:
- 683 10.1146/annurev.ecolsys.37.091305.110100
- Passos, C., Tassino, B., Reyes, F., & Rosenthal, G. G. (2014). Seasonal variation in
- female mate choice and operational sex ratio in wild populations of an annual fish,
- 686 Austrolebias reicherti. PLoS ONE, 9(7). DOI: 10.1371/journal.pone.0101649
- 687 Podrabsky, J. E., Carpenter, J. F., & Hand, S. C. (2001). Survival of water stress in
- annual fish embryos: dehydration avoidance and egg envelope amyloid fibers.
- 689 American Journal of Physiology-Regulatory, Integrative and Comparative
- 690 *Physiology*, 280(1), R123-R131.
- 691 Podrabsky, J. E., Garrett I. D. F., & Kohl, Z. F. (2010a). Alternative developmental
- 692 pathways associated with diapause regulated by temperature and maternal
- 693 influences in embryos of the annual killifish *Austrofundulus limnaeus*. Journal of
- 694 *Experimental Biology*, 213(19), 3280–3288. DOI: 10.1242/jeb.045906
- 695 Podrabsky, J. E., & Hand, S. C. (1999). The bioenergetics of embryonic diapause in
- an annual killifish, *Austrofundulus limnaeus*. Journal of Experimental Biology,
- 697 202(19), 2567-2580.

- 698 Podrabsky, J. E., Hrbek, T., & Hand, S. C. (1997). Physical and chemical
- 699 characteristics of ephemeral pond habitats in the Maracaibo basin and Llanos
- region of Venezuela. *Hydrobiologia*, 362(1-3), 67-77.
- 701 Podrabsky, J. E., Tingaud-Sequeira, A., & Cerdá, J. (2010b). Metabolic dormancy and
- responses to environmental desiccation in fish embryos. In: Lubzens, E, J Cerdá,
- 703 M Clark (Eds), Dormancy and resistance in harsh environments. Berlin,
- Heidelberg, Springer. pp.: 203-226.
- 705 Polačik, M., & Podrabsky, J. E. (2015). Temporary environments. In: Riesch, R.,
- Tobler, M. & Plath, M. (Eds), *Extremophile fishes*. New York, Springer. pp.: 217–
- 707 245.
- 708 Polačik, M, & Reichard, M. (2010). Diet overlap among three sympatric African
- annual killifish species Nothobranchius spp. from Mozambique. Journal of Fish

710 Biology, 77(3), 754–768. DOI: 10.1111/j.1095-8649.2010.02717.x

- 711 Polačik, M., Reichard, M., & Vrtílek, M. (2018). Local variation in embryo
- development rate in annual fish. *Journal of Fish Biology*, 92, 1359-1370.
- 713 Polačik, M., Smith, C., Reichard, M. (2017). Maternal source of variability in the
- embryo development of an annual killifish. *Journal of Evolutionary Biology*, 30,
- 715 738-749. DOI:10.1111/jeb.13038
- 716 R Core Team (2018). R: A language and environment for statistical computing.
- 717 Vienna, Austria: R Foundation for Statistical Computing. (http://www. R-
- 718 project.org/).
- 719 Rafferty, N. E., Caradonna, P. J., & Bronstein, J. L. (2015). Phenological shifts and
- 720 the fate of mutualisms. *Oikos*, 124(1), 14–21. DOI: 10.1111/oik.01523
- 721 Rafferty, N. E., & Ives, A. R. (2011). Effects of experimental shifts in flowering
- phenology on plant-pollinator interactions. *Ecology Letters*, 14(1), 69–74.

- 723 Reichard, M. (2016). The evolutionary ecology of African annual fishes. In: Berois,
- 724 N, García, G, & de Sá, R (Eds). Annual Fishes: Life History Strategy, Diversity,
- 725 *and Evolution*. Boca Ratón, CRC Press. pp.: 133-158.
- Reichard, M., & Polačik, M. (2019). *Nothobranchius furzeri*, an 'instant' fish from an
 ephemeral habitat. *eLife*, 8, e41548.
- 728 Rollinson, C. R., Kaye, M. W., & Canham, C. D. (2016). Interspecific variation in
- growth responses to climate and competition of five eastern tree species. *Ecology*,
- 730 97(4), 1003-1011.
- Rohlf, F.J. (2009). tpsDig2, Version 2.1. State University of New York, Stony Brook.
- 732 Rue, H., Riebler, A., Sørbye, S. H., Illian, J. B., Simpson, D. P., & Lindgren, F. K.
- (2017). Bayesian computing with INLA: a review. *Annual Review of Statistics and its Application*, 4, 395-421.
- 735 Sæther, B. E., & Bakke, Ø. (2000). Avian life history variation and contribution of
- demographic traits to the population growth rate. *Ecology*, 81(3), 642-653.
- 737 Stearns, S. C. (1992). The evolution of life histories. New York, Oxford University
- 738 Press.
- 739 Thomas, C. D., Cameron, A., Green, R. E., Bakkenes, M., Beaumont, L. J.,
- 740 Collingham, Y. C., ... Williams, S. E. (2004). Extinction risk from climate
- 741 change. *Nature*, 427(6970), 145.
- 742 Trenberth, K. E. (2011). Changes in precipitation with climate change. *Climate*
- 743 *Research*, 47, 123–138. DOI: 10.3354/cr00953
- 744 Valenzano, D. R., Terzibasi, E., Cattaneo, A., Domenici, L., & Cellerino, A. (2006).
- 745 Temperature affects longevity and age related locomotor and cognitive decay in
- the short lived fish *Nothobranchius furzeri*. *Aging cell*, 5(3), 275-278.

- 747 Valladares, F., Matesanz, S., Guilhaumon, F., Araújo, M. B., Balaguer, L., Benito-
- 748 Garzón, M., ... Zavala, M. A. (2014). The effects of phenotypic plasticity and local
- adaptation on forecasts of species range shifts under climate change. *Ecology*
- 750 *Letters*, 17(11), 1351-1364.
- 751 Varpe, Ø. (2017). Life History Adaptations to Seasonality. Integrative and
- 752 *Comparative Biology*, 57(5), 943–960. DOI:10.1093/icb/icx123.
- 753 Volcan, M. V., Fonseca, A. P. D., Figueiredo, M. R. C., Sampaio, L. A., & Robaldo,
- R. B. (2012). Effect of temperature on growth of the threatened annual fish
- Austrolebias nigrofasciatus Costa & Cheffe 2001. Biota Neotropica, 12(4), 68-73.
- 756 Volcan, M. V., Gonçalves, Â. C., Lanés, L. E. K., & Guadagnin, D. L. (2016). Annual
- 757 fishes (Rivulidae) from Southern Brazil: A broad-scale assessment of their
- diversity and conservation. In: Berois, N., Garcia, G., de Sa, R. (Eds.) Annual
- 759 fishes: Life history strategy, diversity, and evolution. Boca Ratón, CRC Press. pp.:
- 760 185-206.
- 761 Volcan, M. V., & Lanés, L. E. K. (2018). Brazilian killifishes risk extinction. Science,
- 762 DOI: 10.1126/science.aau5930
- 763 Walford, R. L., & Liu, R, K. (1965). Husbandry, life span, and growth rate of the
- annual fish, *Cynolebias adloffi* E. Ahl. *Experimental Gerontology*, 1(2), 161-168.
- 765 Walther G. R., Post, E. Convey, P., Menzel, A., Parmesan, C., Beebee, T. J. C.,
- Fromentin, J. M., Hoegh-Guldbergs, O., & Bairlein, F. (2002). Ecological
- responses to recent climate change. *Nature*, 416, 389-395.
- 768 Witze, A. (2018). Why extreme rains are getting worse. *Nature*, 563, 458–460. DOI:
- 769 10.1038/d41586-018-07447-1

- Wood, S. & Scheipl, F. (2017). gamm4: Generalized Additive Mixed Models using
- 771 'mgcv' and 'lme4'. *R package version 0.2-5*, https://CRAN.R-
- project.org/package=gamm4.
- Wootton R. J., & Smith, C. (2015). *Reproductive biology of teleost fishes*. Oxford,
 Wiley.
- 775 Zebral, Y. D., Lansini, L. R., Costa, P. G., Roza, M., Bianchini, A., & Robaldo, R. B.
- 776 (2018). A glyphosate-based herbicide reduces fertility, embryonic upper thermal
- tolerance and alters embryonic diapause of the threatened annual fish *Austrolebias*
- nigrofasciatus. Chemosphere, 196, 260-269.
- 779 Zuur, A. F., & Ieno, E. N. (2016). A protocol for conducting and presenting results of
- regression type analyses. *Methods in Ecology and Evolution*, 7, 636-645.
- Zuur, A. F., Saveliev, A. A., & Ieno, E. N. (2014). *A beginner's guide to generalised additive mixed models with R.* Newburgh, Highland Statistics Ltd.
- 783
- 784

Table 1. Mean estimates, their standard errors (SE) and statistical significance for
gonad mass in *A. bellottii* as a function of coexistence with *A. nigripinnis*, body size
(TL, log-transformed) and their interaction, modelled using a zero-altered model with
pool identity included as a random term.

	(a) Occurrence model				(b) Frequency model			
Parameter	Estimate	SE	Z	Р	Estimate	SE	Z	Р
Intercept	-45.55	10.82	-4.21	< 0.001	-9.91	0.44	-22.71	<0.001
Coexistence	-51.95	33.75	-1.54	0.124	-0.80	0.69	-1.15	0.254
logTL	32.45	7.68	4.23	< 0.001	5.44	0.29	18.78	< 0.001
Coexist:TL	36.65	23.91	1.53	0.125	0.51	0.46	1.13	0.264

Table 2. Posterior mean estimates of *A. bellottii* TL modelled using a gamma random

walk trend model fitted using INLA. CrI is the 95% Bayesian credible interval.

794 Credible intervals that do not contain zero indicate a statistically important difference.

Sex is mature fish sex. *Cohort* is the cohort of fish to which an individual belonged.

Area is the surface area of the pool in which fish were captured.

Model parameter	Posterior mean	Lower CrI	Upper CrI
Intercept	1.077	0.989	1.164
Sex _(male)	0.119	0.106	0.132
Cohort _(second)	0.236	0.201	0.271
Area	-0.008	-0.024	0.007

Table 3. Posterior mean estimates for gonad mass in *A. bellottii* as a function of cohort,
age and pool area, modelled using a zero-altered gamma (ZAG) model with pool
included as a random term. CrI is the 95% Bayesian credible interval. Credible intervals
that do not contain zero in bold to indicate statistical importance.

	(a) Occurrence model			(b) Frequency model			
Parameter	Posterior	Lower	Upper	Posterior	Lower	Upper	
	mean	CrI	CrI	mean	CrI	CrI	
Intercept	1.47	0.15	2.80	3.08	2.70	3.43	
Cohort _(second)	2.11	0.43	3.97	1.37	0.84	1.93	
Age	3.83	2.83	4.95	0.86	0.75	0.99	
Area	-0.51	-1.29	0.21	0.27	0.11	0.44	

806 Figure Captions

807 Figure 1. Location of the study area (A), with individual pools indicated (B). 808 Symbols for individual pools designate presence of a single cohort that either survived 809 the entire season (open symbol) or perished when the pool desiccated in mid-winter 810 (grey symbol), and the presence of the first and second cohort (black symbol). Pools 811 with A. bellottii and A. nigripinnis are marked by triangles and pools without A. 812 nigripinnis are indicated by circles. Note that in two pools, the first cohort survived 813 despite apparent desiccation. 814 815 Figure 2. Growth trajectories of A. bellottii males (A) and females (B) in pools 816 without A. nigripinnis (blue diamonds) or coexisting with A. nigripinnis (red 817 triangles). Individual body size measurements are indicated (TL, in cm), along with 818 GAMM-fitted curves with separate smoothers for presence/absence of A. nigripinnis. 819 820 Figure 3. Water temperature experienced by the first (grey) and second (black) cohort

Bars indicate hatching period for each cohort. Note that values experienced by the
first and second cohort overlap and values experienced by adults from the first cohort
from the age of 110 days are the same as those experienced by the second cohort from
day 0.

of A. bellottii measured in situ using dataloggers. Daily mean values are presented.

826

821

Figure 4. Growth trajectories of the first (blue circles) and second (red triangles)
cohorts of *A. bellottii* males (A) and females (B) and *A. nigripinnis* males (C) and

829	females (D). Individual body size measurements are indicated (TL, in cm), along with
830	GAMM-fitted curves with separate smoothers for each cohort.
831	
832	Figure 5. Specific growth rates (SGR) of the first (blue circles) and second (red
833	triangles) cohort of A. bellottii (both sexes pooled). Note that formal analysis
834	compared only SGRs until Time 7 to avoid bias from unequal lifespan between the
835	two cohorts.
836	
837	Supporting Information Table S1. Sample size for each pool, species, sex, and
838	sampling event.
839	
840	Supporting Information Figure S1. Precipitation and temperature data for the study
841	area (Villa Soriano, Uruguay) comparing the year 2015 with 10 years database. Data

842 were provided by National Meteorology Institute, INUMET.