

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21

Silicon and zinc biogeochemical cycles coupled through the Southern Ocean

Derek Vance^{1*}, Susan H. Little², Gregory F. de Souza¹, Samar P. Khatiwala³, Maeve C. Lohan⁴, Rob Middag⁵

¹Institute of Geochemistry and Petrology, Department of Earth Sciences, ETH Zürich, Clausiusstrasse 25, 8092 Zürich.

²Department of Earth Science and Engineering, Imperial College London, South Kensington Campus, Exhibition Road, London, SW7 2AZ, UK.

³Department of Earth Sciences, South Parks Road, Oxford, OX1 3AN, UK.

⁴School of Ocean and Earth Sciences, National Oceanography Centre, University of Southampton, Southampton, UK.

⁵Royal Netherlands Institute for Sea Research (Royal NIOZ), P.O. Box 59, 1790 AB Den Burg, Texel, The Netherlands.

22 **Zinc is vital for the physiology of oceanic phytoplankton. The striking similarity of the depth**
23 **profiles of zinc to those of silicate suggests that the uptake of both elements into the opaline**
24 **frustules of diatoms, and their regeneration from these frustules, should be coupled. However,**
25 **the zinc content of diatom opal is negligible, and zinc is taken up into and regenerated from**
26 **the organic parts of diatom cells. Thus, since opaline frustules dissolve deep in the water**
27 **column whilst organic material is regenerated in the shallow subsurface ocean, there is little**
28 **reason to expect the observed close similarity between zinc and silicate, and the dissimilarity**
29 **between zinc and phosphate. Here we combine observations with simulations using a three-**
30 **dimensional model of ocean circulation and biogeochemistry to show that the coupled**
31 **distribution of zinc and silicate, as well as the decoupling of zinc and phosphate, can arise in**
32 **the absence of mechanistic links between the uptake of zinc and silicate, and despite**
33 **contrasting regeneration length-scales. Our simulations indicate that the oceanic zinc**
34 **distribution is, in fact, a natural result of the interaction between ocean biogeochemistry and**
35 **the physical circulation through the Southern Ocean hub. Our analysis demonstrates the**
36 **importance of uptake stoichiometry in controlling ocean biogeochemistry, and the utility of**
37 **global-scale elemental co-variation in the ocean in understanding these controls.**

38
39 The spatial and vertical distributions of macro- (e.g. phosphate, nitrate, silicate) and metal micro-
40 nutrients (e.g. iron, zinc) control sequestration of carbon by the ocean's biosphere. Zinc plays vital
41 physiological roles in oceanic phytoplankton (e.g., ref 1) and Zn contents of phytoplankton cells are
42 of the same order as the essential trace element, iron²⁻⁴. As a result, and like the major nutrients (P,
43 N, Si), Zn shows extreme depletions in the surface ocean and enrichments at depth (Fig. 1, see Fig.
44 2 for locations and sources of all data used in the paper), a distribution that is often conceptualised
45 in terms of one-dimensional vertical cycling involving uptake in the photic zone and regeneration
46 beneath. Though the extent to which surface Zn concentrations limit primary productivity is still
47 unclear (e.g., ref 12), the processes that control the removal of Zn from the photic zone, its

48 regeneration in the deep ocean, and its return to the surface via the ocean circulation, are a key part
49 of the dynamics of oceanic nutrient chemistry.

50

51 A remarkable feature of global ocean nutrient profiles (Fig. 1) is the tight co-variation of Zn and Si
52 in all three major ocean basins. Both elements increase to maxima for each profile in the abyssal
53 ocean, in marked contrast to the intermediate-depth maximum for PO₄. Though this coupling of Zn
54 to Si has been repeatedly noted⁵⁻⁷, its origin has been a puzzle. Rationales for the deep maximum in
55 Si relative to organic-associated nutrients, nitrate and phosphate, have emphasised two key
56 processes: (1) vertical cycling with a greater regeneration length-scale of diatom opal relative to
57 organic matter (e.g., 13, 14); (2) rapid depletion of Si relative to organic-associated nutrients in the
58 surface Southern Ocean, creating distinct water mass signatures that are exported to the rest of the
59 global ocean^{15,16}. For Zn it has been tentatively suggested that uptake of Zn and Si may be linked,
60 and that increased Si uptake by diatoms due to Fe limitation may explain co-variation of Zn and Si
61 observed in high-latitude nutriclines¹⁷. The first of these two views would suggest that the oceanic
62 cycle of Zn is dominated by uptake into and regeneration from diatom opal. However, culturing
63 studies¹⁸ have demonstrated that the Zn content of diatom opal is negligible (1-3% of the total
64 cellular inventory), and that the Zn/Si ratio of diatom opal is nearly two orders of magnitude lower
65 than observed in the deep ocean^{5,18}. This is confirmed by synchrotron micro-XRF data^{2,3} from both
66 cultured and natural diatom cells, which clearly show that Zn is co-located with phosphorus in the
67 organic matter of diatom cells and not in their opaline frustules, and that Zn is regenerated from this
68 organic material in the upper ocean with phosphate, and not with Si from opal^{19,20}.

69

70 **Diatom uptake in the Southern Ocean couples Zn and Si**

71 We suggest that the solution to this paradox lies in the biogeochemistry and physical oceanography
72 of the Southern Ocean. Water upwelled at the Antarctic divergence, and moved northward at the

73 surface by Ekman transport to the Sub-Antarctic zone, is stripped of Si and Zn much faster than
74 PO_4 (Fig. 3a, b). Thus, both Zn and Si concentrations drop by a factor of about 40 over the surface
75 transect shown in Fig. 3a, whereas PO_4 drops by only a factor of 3. This remarkable difference in
76 relative drawdown rates is consistent with the ecological dominance of diatoms in this region (e.g.
77 refs 22, 23), and their peculiarly high Zn/P uptake ratio, up to an order of magnitude greater than
78 the average for oceanic phytoplankton³. The Zn- and Si-depleted Sub-Antarctic surface layer is the
79 ultimate source of Sub-Antarctic Mode Water (SAMW), and it is the northward transport of this
80 water mass that, we suggest, sets the low dissolved Si and Zn concentrations, and low Si/ PO_4 and
81 Zn/ PO_4 ratios, of most of the global upper ocean (Fig. 1). The data in Fig. 3a also imply a rather
82 constant ratio for the removal of Zn and Si from the dissolved pool of the surface Southern Ocean,
83 despite the fact that Si is taken up into opal and Zn into organic material.

84

85 What is the ultimate fate of the large amounts of Zn and Si relative to PO_4 removed from the
86 surface Southern Ocean? The Sub-Antarctic winter mixed layer from which SAMW forms is of the
87 order of 200-400m thick (e.g., ref 24), beneath which the Southern Ocean water column is
88 vigorously mixed and rather homogeneous (e.g., ref 25), including for Si, Zn and PO_4 ¹¹. Thus, if
89 diatom cells are exported beneath about 200-400m, their high Zn/P and Si/P characteristics will be
90 imparted to the deep Southern Ocean regardless of the exact cellular location (opal or organic
91 matter) of Zn, Si and P, making the different regeneration length-scales of these particulate phases
92 much less relevant here than elsewhere in the ocean. This export, which may be aided by the rapid
93 blooming and equally rapid population collapse that is characteristic of diatom ecology (e.g., refs
94 22, 23), results in a Zn and Si deficit relative to PO_4 in the mixed layer, which is the source of upper
95 ocean water masses such as AAIW and SAMW.

96

97 In summary, the hypothesis we put forward here consists of three components: (1) extreme
98 drawdown of Zn and Si relative to PO_4 in the surface Southern Ocean (Fig. 2a) consistent with the
99 known stoichiometry of Southern Ocean diatoms³, thus setting the biogeochemical signature of the
100 surface Southern Ocean and the upper ocean water masses derived from it; (2) export of diatom
101 cells below the winter mixed layer depth, transferring both Zn and Si at high Zn/P and Si/P to the
102 deep Southern Ocean, despite being located in two different components of the diatom cell with
103 different regeneration length-scales, thus setting the biogeochemical signatures of deep and abyssal
104 Southern Ocean-derived water masses (Fig. 1); and (3) lateral export of these water mass signatures
105 to the low latitude oceans, as previously proposed to control Si- NO_3 nutrient systematics^{15,16}. The
106 imprint of these processes can be seen in Fig. 3c. Zinc- PO_4 data for the Atlantic sector of the
107 Antarctic zone of the Southern Ocean (including the surface transect in Fig. 3a and the entire water
108 column at 67°S, black triangles¹¹) indicate co-variation along a line with a slope of about 8 mmol
109 mol^{-1} , consistent with a control by diatom uptake and regeneration. Data for stations proximal to the
110 Southern Ocean, at 40°S in the South Atlantic (entire water column, blue squares⁷) clearly show
111 two different behaviours, with the deep ocean lying close to the Antarctic zone data, and the upper
112 ocean data reflecting the much lower slope expected for water masses sourced in the mixed layer of
113 the Southern Ocean, from which Zn has been stripped. The red dashed lines indicate, schematically,
114 how water mass mixing in locations more distal from the Southern Ocean sources would confound
115 these clear distinctions.

116

117 **The Southern Ocean uptake hypothesis tested in an ocean model**

118 To provide a quantitative test of these ideas, and taking advantage of the computational efficiency
119 of the transport matrix method²⁶, we performed a series of 11 sensitivity simulations (see Methods
120 and Supplementary Information) using an ocean general circulation model coupled offline to a
121 biogeochemical model of P, Si and Zn cycling. In the biogeochemical model, PO_4 and Si cycling

122 are treated completely independently, whilst Zn cycling is explicitly tied to that of P. Zinc in the
123 oceanic dissolved pool is present as both inorganic Zn (Zn^{2+}) and complexed with an organic ligand
124 (ZnL). Culturing studies²⁷ have shown that Zn uptake by phytoplankton, and their Zn/P ratios, can
125 be parameterised using the quantitative dependence of Zn uptake on free Zn concentrations, the
126 approach we adopt here. It is currently unclear exactly *why* Southern Ocean diatoms exhibit high
127 Zn/P uptake ratios, but the high values seen in data³ emerge from the model simply from the fact
128 that upwelled water in the Southern Ocean contains high concentrations of Zn so that not all of it is
129 complexed by organic ligands, a finding that is also supported by data²⁸.

130

131 Although the lengthscale of Zn regeneration is identical to P in these simulations, and though its
132 cycling is entirely decoupled from that of Si, our model reproduces the observed near-linear
133 correlation between Zn and Si at the global scale (Fig. 4c). It also reproduces the slow increase with
134 depth in Zn concentrations through the upper ocean at low latitudes (Fig. 1), despite the fact that Zn
135 is regenerated in the shallow subsurface in the model. The mechanism behind this model behaviour
136 is indicated by our model's sensitivity to the Zn/P uptake ratio in the Southern Ocean (see
137 Supplementary Information for sensitivity tests): as the average Zn/P ratio of uptake in the Southern
138 Ocean increases, the Zn distribution changes from being closely correlated with that of PO_4 (at Zn/P
139 uptake ratios of $\sim 1 \text{ mmol mol}^{-1}$) to being very similar to the large-scale Si distribution (at values
140 above $\sim 4.5 \text{ mmol mol}^{-1}$).

141

142 The key feature of the global Zn- PO_4 covariation (Fig. 4d) is the curvature in the model array. This
143 arises because of the partitioning of the global oceans into the two broad regimes seen in Fig. 3c: an
144 upper ocean that is severely depleted in Zn (and Si) relative to PO_4 as a result of the stripping of Zn
145 (and Si) from the surface Southern Ocean, and the export of this water to the upper ocean globally;
146 and a deep and abyssal ocean that is dominated by water masses originating in the deep Southern

147 Ocean, containing the regenerated counterpart of this surface uptake process. The model is less
148 good at representing the physical mixing of these end-members outside the Southern Ocean (see
149 schematic trajectories in Fig. 3c), so that it underestimates the scatter in the data in Fig. 4b,d.

150

151 We emphasise that our aim here is to provide an explanation for a first-order feature of global ocean
152 Zn-Si-P distributions and, as such, we have deliberately kept the model simple, attempting to
153 restrict the processes it represents to those that are well-established. Thus, Zn uptake is tied to that
154 of P as dictated by the results of culturing experiments, and there are no mechanistic links between
155 the uptake of Zn and Si. We have parameterised Zn speciation in the context of a single ligand
156 model (see Supplementary Information). The model does not consider the potential impacts of
157 variations in metal uptake stoichiometry across different taxonomic groups. It does not consider the
158 potential effects that Fe limitation may have on diatom physiology. Despite this simplicity, the
159 model is very successful in reproducing global dissolved nutrient distributions. Invoking further
160 complexity, such as a significant role for previously-proposed vertical processes such as scavenging
161 of Zn by particulate material or a second organic Zn pool associated with opal^{11,29,30} is not required
162 to reproduce first-order global Zn-Si-PO₄ relationships. We have also avoided tuning the model to
163 improve the fit to the data. For example, the slope of the model Zn-Si correlation on Fig. 4c is about
164 12% below that defined by the data, at 0.056 mmol mol⁻¹ versus 0.064 mmol mol⁻¹. The simulated
165 slope is entirely dependent on assumed model values for the average oceanic Zn and Si
166 concentrations, drawn from the literature (see Supplementary Information). The former, in
167 particular, given the comparative lack of data coverage, is certainly only an estimate at this stage.

168

169 Our analysis of Zn-Si-PO₄ systematics in the global ocean has important implications for both Zn
170 itself and for oceanic trace elements more broadly. The implication of our proposal is that Southern
171 Ocean diatoms dominate oceanic Zn cycling, transferring a large fraction of the oceanic pool to the

172 abyssal ocean and trapping it there, in a direct analogy with Si trapping^{15,16}. The finding that Zn
173 distributions in the global ocean are so profoundly influenced by the stoichiometry of uptake in the
174 Southern Ocean has relevance for studies of other trace metals, their oceanic distributions, and
175 relationships between different nutrients. For example, the “kink” in Zn-PO₄ is reminiscent of the
176 well established “kink” in the relationship between the trace metal cadmium (Cd) and PO₄, whose
177 precise origin is much debated³¹⁻³³. Phytoplankton uptake stoichiometry, speciation of metals in the
178 photic zone, and their systematics in regions of the ocean that are hubs for the physical circulation,
179 are likely as important for other metals as they are for Zn in determining global ocean distributions.
180 The implication is that oceanic metal micronutrient distributions are set by a combination of
181 Southern Ocean ecology and physical circulation, as for major nutrients^{15,16}. Therefore, changes in
182 the Southern Ocean forced by past and future climate change will have global impact.

183 **References**

- 184 1. Morel, F.M.M., Milligan, A.J. & Saito, M.A. Marine bioinorganic chemistry: the role of trace
185 metals in the oceanic cycles of major nutrients. *Treatise in Geochemistry* **8**, 123-150 (2014).
- 186 2. Twining, B.S. *et al.* Quantifying trace elements in individual aquatic protest cells with a
187 synchrotron X-ray fluorescence microprobe. *Anal. Chem.* **75**, 3806-3816 (2003).
- 188 3. Twining, B.S. & Baines, S.B. The trace metal composition of marine phytoplankton. *Ann. Rev.*
189 *Mar. Sci.* **5**, 191-215 (2013).
- 190 4. Boyd, P.W. & Ellwood, M.C. The biogeochemical cycle of iron in the ocean. *Nature Geoscience*
191 **3**, 675-682 (2010).
- 192 5. Bruland, K.W. Oceanographic distributions of cadmium, zinc, nickel, and copper in the North
193 Pacific. *Earth Planet. Sci. Lett.* **47**, 176-198 (1980).
- 194 6. Martin, J.H., Gordon, R.M., Fitzwater, S. & Broenkow, W.W. VERTEX: phytoplankton/iron
195 studies in the Gulf of Alaska. *Deep-Sea Res.* **36**, 649-680 (1989).
- 196 7. Wyatt, N.J. *et al.* Biogeochemical cycling of dissolved zinc along the GEOTRACES South
197 Atlantic transect GA10 at 40°C. *Glob. Biogeochem. Cycles* **28**, 44-56 (2014).
- 198 8. You, Y. Intermediate water circulation and ventilation of the Indian Ocean derived from water-
199 mass contributions. *J. Mar.Res.* **56**, 1029-1067 (1998).
- 200 9. Vu, H.T.D. & Sohrin, Y. Diverse stoichiometry of dissolved trace metals in the Indian Ocean.
201 *Sci. Rep.* **3**, paper 1745. (2013).
- 202 10. Geotraces Intermediate Data Product. <http://www.bodc.ac.uk/geotraces/data/idp2014>.
- 203 11. Zhao, Y., Vance, D., Abouchami, W. & de Baar, H.J.W. Biogeochemical cycling of zinc and
204 its isotopes in the Southern Ocean. *Geochim. Cosmochim. Acta* **125**, 653-672 (2014).
- 205 12. Moore, C.M. *et al.* Processes and patterns of oceanic nutrient limitation. *Nature Geosci.* **6**, 701-
206 710. (2013).

- 207 13. Berelson, W.M. The flux of particulate organic carbon into the ocean interior: a comparison of
208 four US JGOFS regional studies. *Oceanogr.* **14**, 59-67 (2001).
- 209 14. Ragueneau, O., Dittert, N., Pondaven, P., Treguer, P. & Corrin, L. Si/C decoupling in the world
210 ocean: is the Southern Ocean different? *Deep-Sea Research II* **49**, 3127-3154 (2002).
- 211 15. Sarmiento, J.L., Gruber, N., Brzezinski, M.A. & Dunne, J.P. High latitude controls of
212 thermocline nutrients and low latitude biological productivity. *Nature* **427**, 56-60 (2004).
- 213 16. Sarmiento, J.L. *et al.* Deep ocean biogeochemistry of silicic acid and nitrate. *Global*
214 *Biogeochem. Cycles* **21**, B1S90, 10.1029/2006GB002720 (2007).
- 215 17. Sunda, W.G. & Huntsman, S.A. Effect of Zn, Mn, and Fe on Cd accumulation in
216 phytoplankton: implications for oceanic Cd cycling. *Limnol. Oceanogr.* **45**, 1501-1516 (2000).
- 217 *Limnol. Oceanogr.* **59**, 689-704 (2014).
- 218 18. Ellwood, M.C. & Hunter, K.A. The incorporation of zinc and iron in the frustule of the marine
219 diatom *Thalassiosira pseudonana*. *Limnol. Oceanogr.* **45**, 1517-1524 (2000).
- 220 19. Lee, B.-G. & Fisher, N.S. Release rates of trace elements and protein from decomposing
221 planktonic debris. I. Phytoplankton debris. *J. Mar. Res.* **51**, 391-421 (1993).
- 222 20. Twining, B.S. *et al.* Differential remineralization of major and trace elements in sinking
223 diatoms.
- 224 21. Schlitzer, R. Electronic atlas of WOCE hydrographic and tracer data now available. *EOS Trans.*
225 *AGU* **81**, 45 (2000).
- 226 22. Armbrust, E.V. The life of diatoms in the world's oceans. *Nature* **459**, 185-192 (2009).
- 227 23. Assmy, P. *et al.* Thick-shelled grazer-protected diatoms decouple ocean carbon and silicon cycle
228 in the iron-limited Antarctic Circumpolar Current. *Proc. Natl. Acad. Sci.* **51**, 20633-20638 (2013).
- 229 24. Sallée, J. B., Wienders, N., Speer, K. & Morrow, R. Formation of subantarctic mode water in
230 the southeastern Indian Ocean. *Ocean Dyn.* **56**, 525-54. (2006).

- 231 25. Marshall, J. & Speer, K. Closure of the meridional overturning circulation in the Southern
232 Ocean. *Nature Geosci.* **5**, 171-180 (2012).
- 233 26. Khatiwala, S., Visbeck, M. & Cane, M.A. Accelerated simulation of passive tracers in ocean
234 circulation models. *Ocean Modelling* **9**, 51-69 (2005).
- 235 27. Sunda, W.G. & Huntsman, S.A. Feedback interactions between zinc and phytoplankton in
236 seawater. *Limnol. and Oceanogr.* **37**, 25-40 (1992).
- 237 28. Baars, O. & Croot, P.L. The speciation of dissolved zinc in the Atlantic sector of the Southern
238 Ocean. *Deep Sea Res. II* **58**, 2720-2732 (2011).
- 239 29. Lohan, M.C., Statham, P.J. & Crawford, D.W. (2002) Total dissolved zinc in the upper water
240 column of the subarctic North East Pacific. *Deep-Sea Res. II* **49**, 5793-5808 (2002).
- 241 30. John, S.G. & Conway, T.M. A role for scavenging in the marine biogeochemical cycling of zinc
242 and zinc isotopes. *Earth Planet. Sci. Lett.* **394**, 159-167 (2014).
- 243 31. Elderfield, H. & Rickaby, R.E.M. Oceanic Cd/O ratio and nutrient utilization in the Southern
244 Ocean. *Nature* **405**, 305-210 (2000).
- 245 32. Cullen, J.T. On the nonlinear relationship between dissolved cadmium and phosphate in the
246 modern global ocean: could chronic iron limitation of phytoplankton growth cause the kink?
247 *Limnol. Oceanogr.* **51**, 1369-1380 (2006).
- 248 33. Quay, P., Cullen, J.T., Landing, W.M. & Morton, P. Processes controlling the distributions of
249 Cd and PO₄ in the ocean. *Glob. Biogeochem. Cyc.* **29**, 830-841. (2015)

250

251 Correspondence and requests for materials to: derek.vance@erdw.ethz.ch.

252

253 **Acknowledgements**

254 This research was supported by ETH Zürich and Swiss National Science Foundation (SNF) Grant
255 200021-153087/1 to DV. SHL is supported by a Leverhulme Early Career Fellowship and GFdS by
256 a Marie Skłodowska-Curie Fellowship.

257

258 **Author contributions**

259 DV and SHL conceived the study. DV wrote the first draft of the paper. GdS constructed the
260 biogeochemical model in collaboration of SPK, conceived and carried out the sensitivity
261 simulations, and analysed the model output. MCL and RM were responsible for the Atlantic data in
262 the GEOTRACES Intermediate Data Product and used in the figures. All authors read and
263 commented on the paper.

264

265 **Competing financial interests**

266 The authors declare no competing financial interests.

267

268 **Figure captions**

269

270 **Figure 1: Example depth profiles of Zn, Si and PO₄ in the three main ocean basins**

271 The striking similarity in the depth distributions of Zn and Si stands in marked contrast to the
272 dissimilarity with PO₄. The presence of Sub-Antarctic Mode Water (SAMW, potential density
273 anomaly $\sigma_{\theta} = 26.80^{15}$) and Antarctic Intermediate Water (AAIW^{7,8}) in the South Atlantic and
274 equatorial Indian Oceans is reflected in the low Si and Zn concentrations compared to P. In the
275 North Pacific these depth levels are occupied by other water masses. Locations of stations depicted,
276 as well as data sources, are given in Fig. 2.

277

278 **Figure 2: Location map**

279 The locations of the stations from which nutrient data are depicted in Figs. 1, 3 and 4. Pacific data
280 from Bruland et al.⁵ (Station 17) and VERTEX profiles⁶. Atlantic (GA02 and GA10⁷) and Indian
281 (GI04⁹) data from the GEOTRACES 2014 Intermediate Data Product¹⁰. Southern Ocean (IPY) data
282 from Zhao et al.¹¹.

283

284 **Figure 3: Contrasting variability in nutrient uptake in the Atlantic sector of the Southern**
285 **Ocean**

286 **a,b:** The strong depletion of Zn and Si versus moderate depletion of PO₄ as water is moved
287 northward from the upwelling zone (data from dashed black line in Fig. 2^{7,11}). Physical features
288 identified using WOCE hydrographic data along the zero meridian²¹: AD, Antarctic Divergence;
289 PF, Polar Front; SAF, Sub-Antarctic Front; AZ, Antarctic Zone, PFZ, Polar Frontal Zone; SAZ,
290 Sub-Antarctic Zone. **c:** Distribution of Zn and PO₄ in the Atlantic sector of the Southern Ocean.
291 Antarctic Zone data¹¹ track removal of Zn and PO₄ at a very high ratio (Zn/P = 8 mmol mol⁻¹).
292 South Atlantic 40°S data⁷ show contrasting Zn/P ratios, extremely low (0.3 mmol mol⁻¹) in the
293 upper ocean, and high in the deep. Dashed red lines show possible trajectories for mixing between
294 end-members on this diagram.

295

296 **Figure 4: Coupled major and micro-nutrient distributions in the global ocean**

297 **a,b:** Global Zn-Si-PO₄ systematics in all three ocean basins. Black arrows in **b** as in Fig 3c. Data
298 plotted for all stations on each section shown in Fig. 2 (see caption for data sources), and for the
299 entire water column. **c,d:** The same data as in **a,b** (grey), with the results of a three-dimensional
300 model of ocean circulation and biogeochemistry superimposed (Simulation 11, see Supplementary

301 Information; colours represent the volume-weighted relative frequency of model cells with the
302 given Zn-Si or Zn-PO₄ characteristics).
303

304 **Methods**

305 *Physical model:* Ocean general circulation model simulations were carried out using the transport
306 matrix method (TMM) of Khatiwala et al.²⁶ using transport matrices (TMs) derived from a coarse-
307 resolution version of the MITgcm³⁴ with $2.8^\circ \times 2.8^\circ$ lateral resolution and 15 vertical levels forced
308 with monthly mean climatological fluxes of momentum, heat and freshwater and with weak
309 restoring of surface temperature and salinity to the Levitus climatology³⁵. Our sensitivity
310 simulations were carried out using constant annual-mean circulation fields derived from the
311 equilibrium state of the model after 5000 yr integration; thus, seasonal variability is not represented
312 in our simulations. The physical model is coupled to a biogeochemical model that simulates the
313 internal oceanic cycling of phosphorus (as phosphate, PO_4 , and dissolved organic phosphorus,
314 DOP), zinc (Zn) and silicate (Si). Biogeochemical model simulations were initialized with constant
315 nutrient tracer fields (ocean-mean concentrations of $2.17 \mu\text{M}$, 5.4 nM and $92 \mu\text{M}$ for PO_4 , Zn and Si
316 respectively) and integrated forward for 5000 model years.

317 *Biogeochemical model:* The nutrient-cycling model that forms the basis of our simulations is based
318 on the formulation developed for the OCMIP-2 project³⁶ and uses PO_4 as its nutrient currency.
319 Phosphate uptake in the surface ocean is driven by restoring surface PO_4 concentrations^{37,38} towards
320 the objectively-analysed annual-mean PO_4 field of World Ocean Atlas 2013³⁹ with a restoring
321 timescale of 36.5 days. A fraction of PO_4 taken up is immediately shunted towards DOP, whilst the
322 remaining fraction is exported as an implicit particulate flux to depth, which remineralises
323 following a power-law dependency on depth^{40,41}. Any particulate flux reaching the bottom of the
324 water column remineralises within the bottom ocean cell. Dissolved organic phosphorus is carried
325 passively with the circulation and decays back to PO_4 with a first-order rate constant of $(0.5 \text{ year})^{-1}$.
326 The Zn-cycling model is explicitly coupled to the P-cycling model above. Zinc uptake in the
327 surface ocean is tied to PO_4 uptake via a dimensionless stoichiometric parameter $r_{\text{Zn:P}}$ (see *Zn*
328 *uptake parameterization* below). This surface-ocean uptake drives an implicit export flux of

329 particulate Zn, which remineralises identically to the implicit particulate P flux. The organic
330 speciation of dissolved Zn is represented implicitly by assuming a constant ligand concentration of
331 1.2 nM with a conditional stability constant of 10^{10} M^{-1} , which reduces the calculation of the
332 concentration of free Zn (i.e. organically and inorganically uncomplexed Zn^{2+}) to the solution of a
333 quadratic equation (see Supplementary Information). Varying biogeochemical behaviours of the
334 Zn-cycling model are achieved by the formulation of the stoichiometric parameter $r_{\text{Zn:P}}$, as detailed
335 below.

336 Our silicon cycling model is conceptually very similar to our PO_4 -cycling model, and to the Si-
337 cycling model of de Souza et al.⁴². Dissolved Si uptake in the surface ocean ($\leq 120\text{m}$ water depth) is
338 driven by restoring surface Si concentrations towards the objectively-analysed annual-mean Si field
339 of World Ocean Atlas 2013 with a restoring timescale of 36.5 days. This surface-ocean uptake
340 drives an implicit particulate export flux, which dissolves following an exponential dependency on
341 depth with a length-scale of 1000 m (ref. 41). Any particulate flux reaching the bottom of the water
342 column remineralises within the bottom ocean cell. It is important to note that in our model
343 formulation, Si cycling is entirely biogeochemically independent of PO_4 and Zn cycling.

344 **Zn uptake parameterization**

345 Details of the parameterization of Zn uptake, including the rationale behind choices made in
346 defining this parameterization, can be found in the Supplementary Information. Broadly, our
347 biogeochemical model represents the plasticity in the Zn:P stoichiometry of uptake by
348 phytoplankton (diatoms and coccolithophorids) observed in culture experiments by Sunda and
349 Hunstman²⁷. These authors observed that the Zn:P uptake ratio varies non-linearly with the
350 concentration of free Zn (Zn^{2+}) in the culturing medium, a dependency with the following
351 functional form (called the “two-site model”):

$$352 \quad r_{\text{Zn:P}} = \frac{a_{\text{Zn}} \cdot \text{Zn}^{2+}}{b_{\text{Zn}} + \text{Zn}^{2+}} + c_{\text{Zn}} \cdot \text{Zn}^{2+} \quad (\text{Eqn. 1})$$

353 where $r_{Zn:P}$ is the molar Zn:P uptake ratio, and parameters a_{Zn} , b_{Zn} and c_{Zn} are constants analogous
354 to maximum uptake rate, half-saturation constant and non-saturable uptake rate, respectively. The
355 culture data of Sunda and Huntsman²⁶ were used to delineate the extent of variability of these
356 parameters in our suite of sensitivity simulations, as detailed in the Supplementary Information.
357 Parameters and key statistical metrics for the 11 sensitivity simulations carried out are listed in
358 Table S1, together with a detailed discussion of the results.

359

360 **Data availability.** The data that support the findings of this study are available at
361 <http://www.bodc.ac.uk/geotraces/data/idp2014>.

362

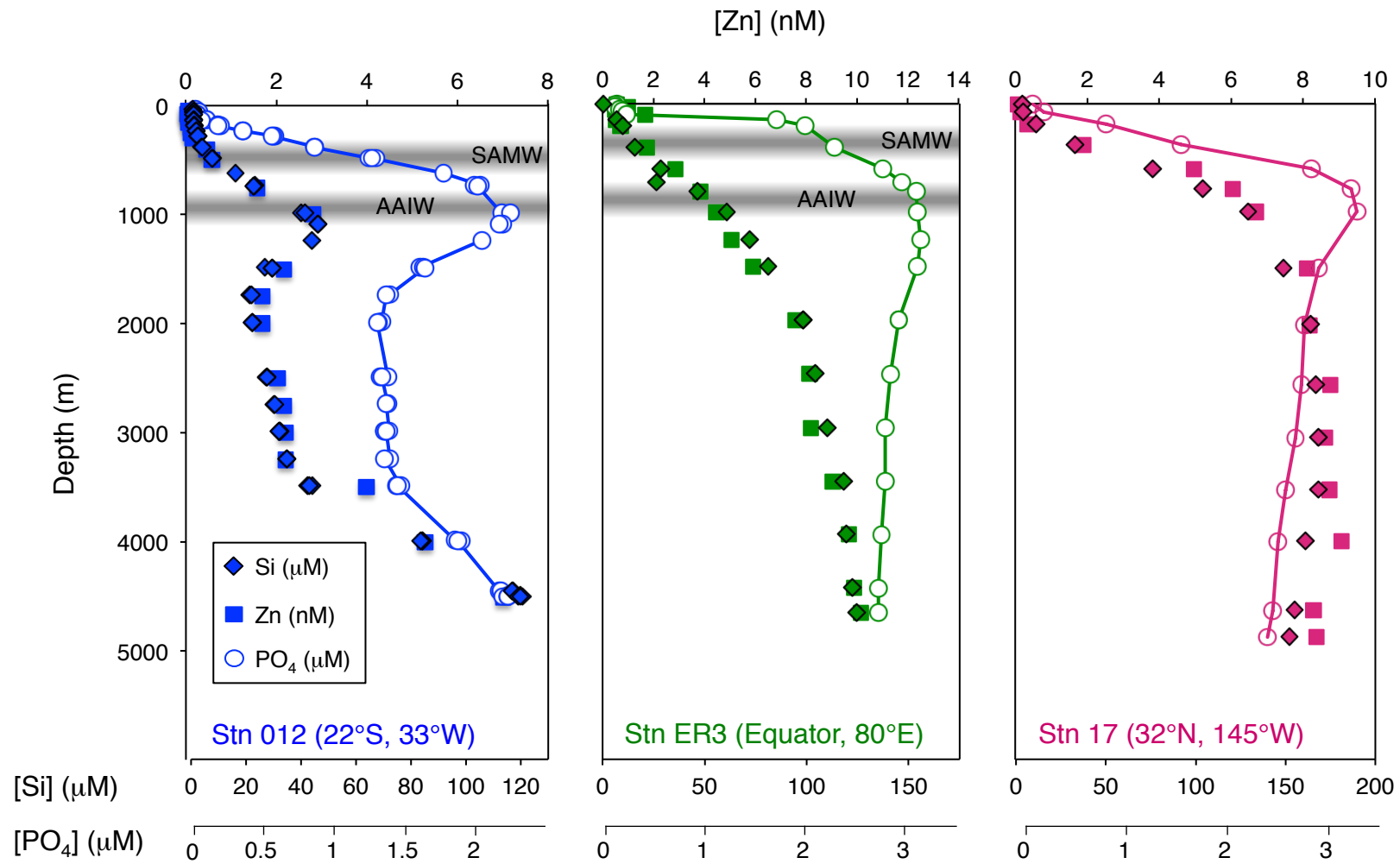
363 **Code availability.** The TMM code is available on GitHub
364 (<https://github.com/samarkhatiwala/tmm>), and the biogeochemical code can be provided on request.

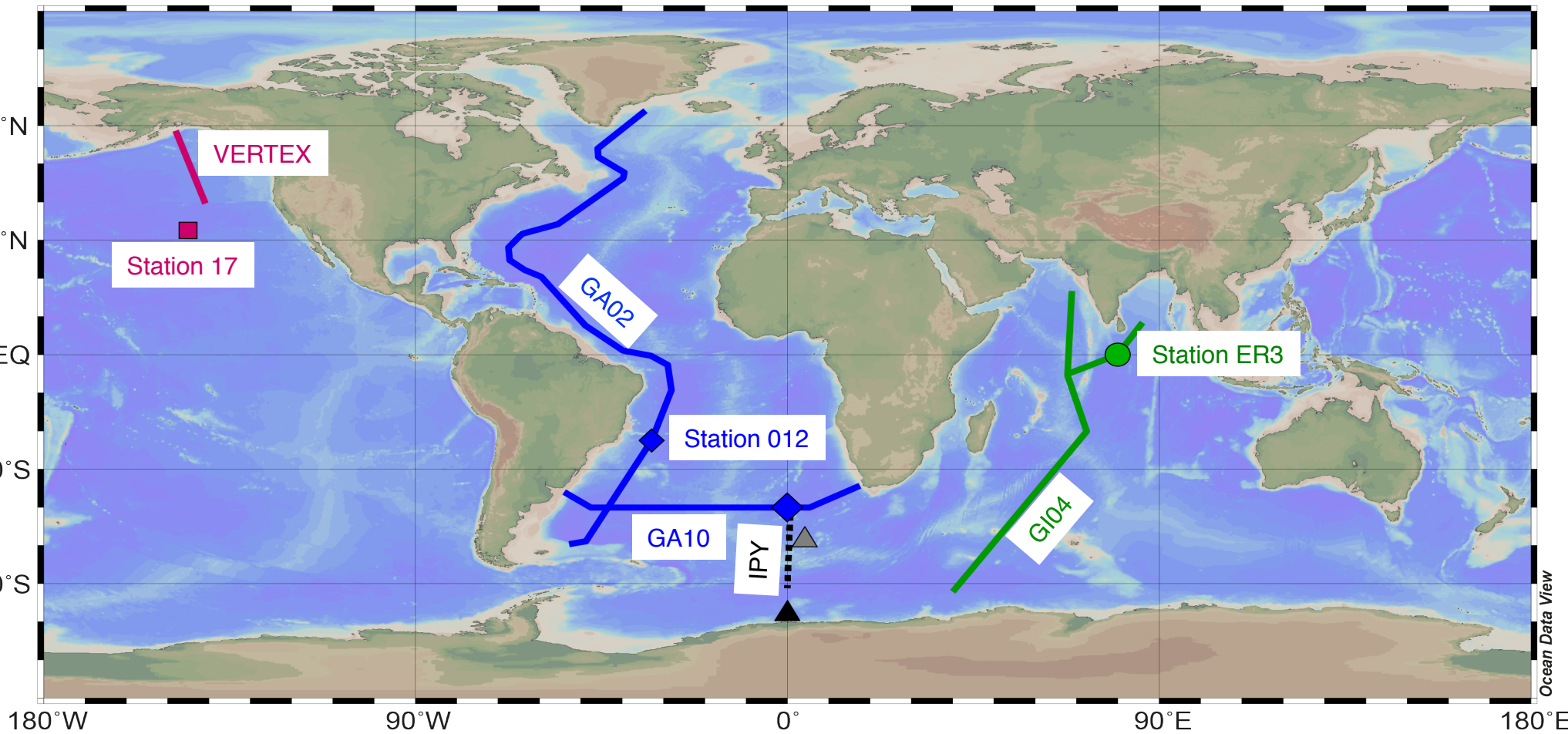
365

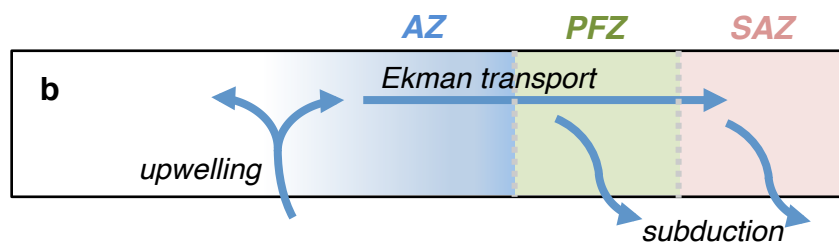
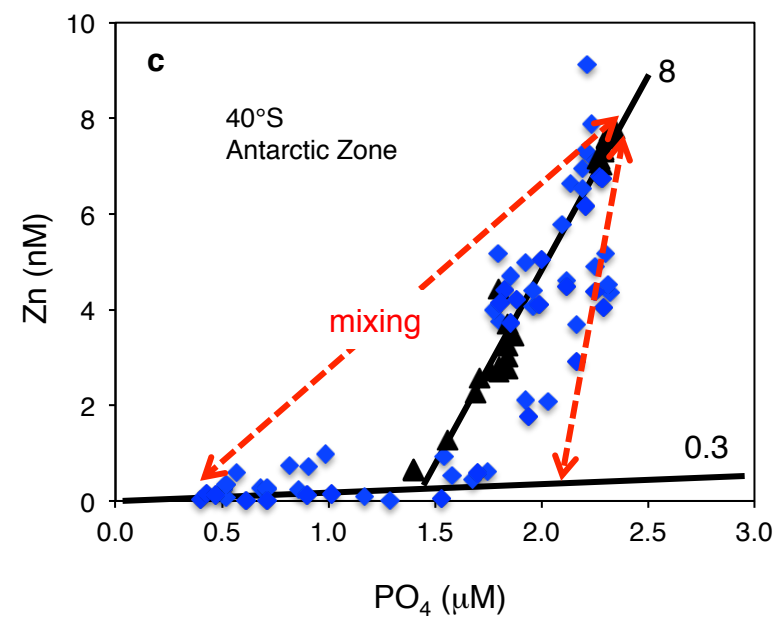
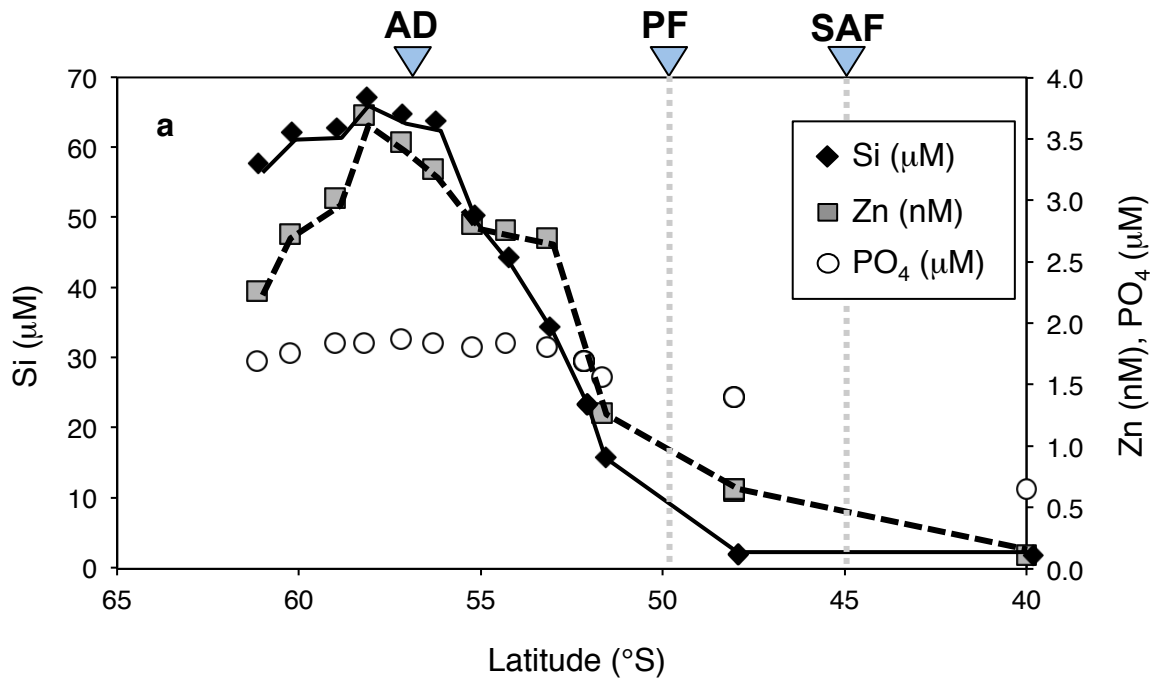
366 **References**

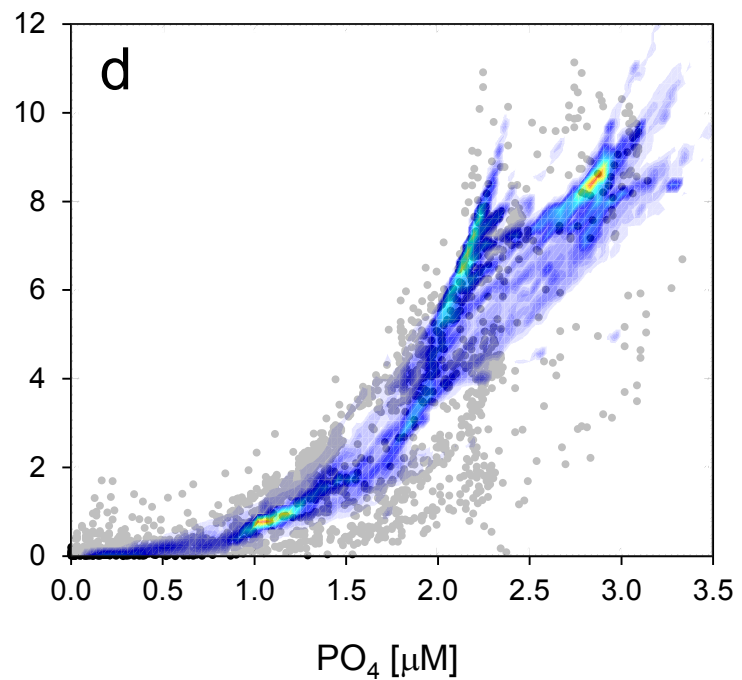
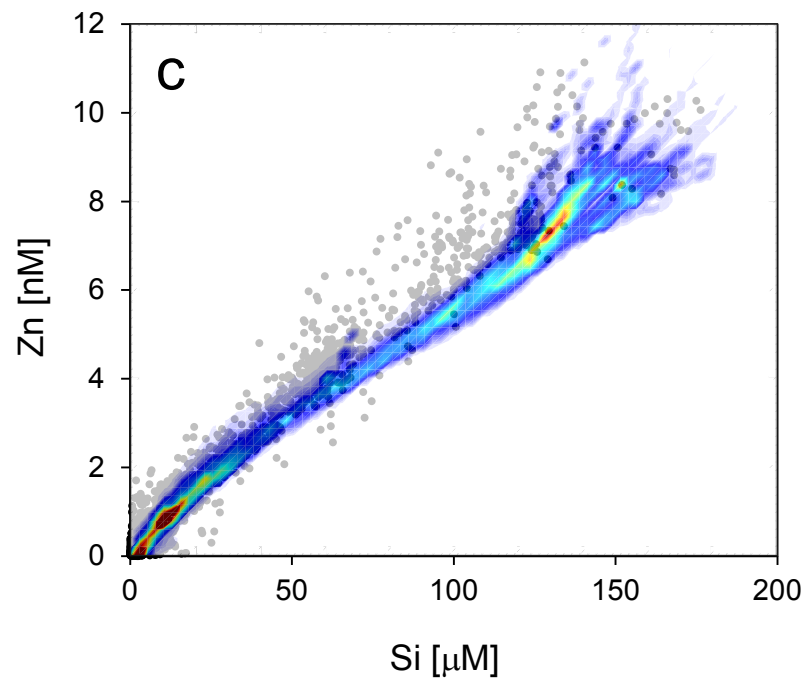
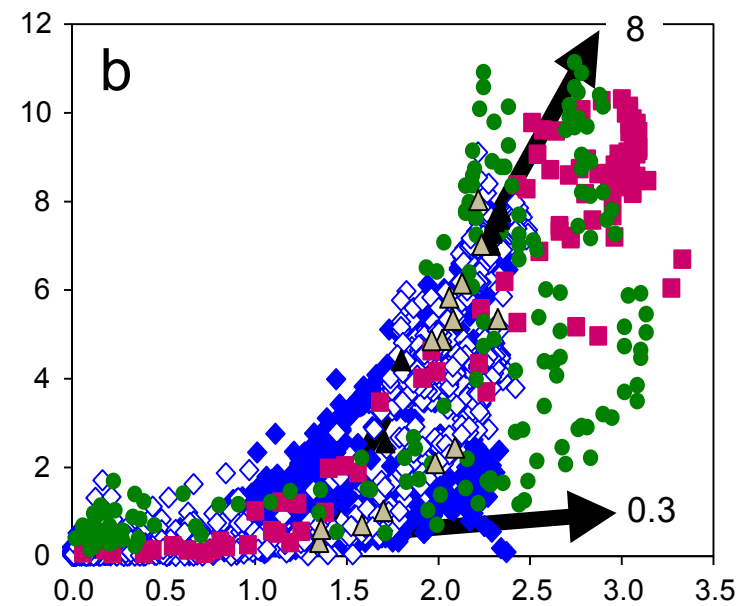
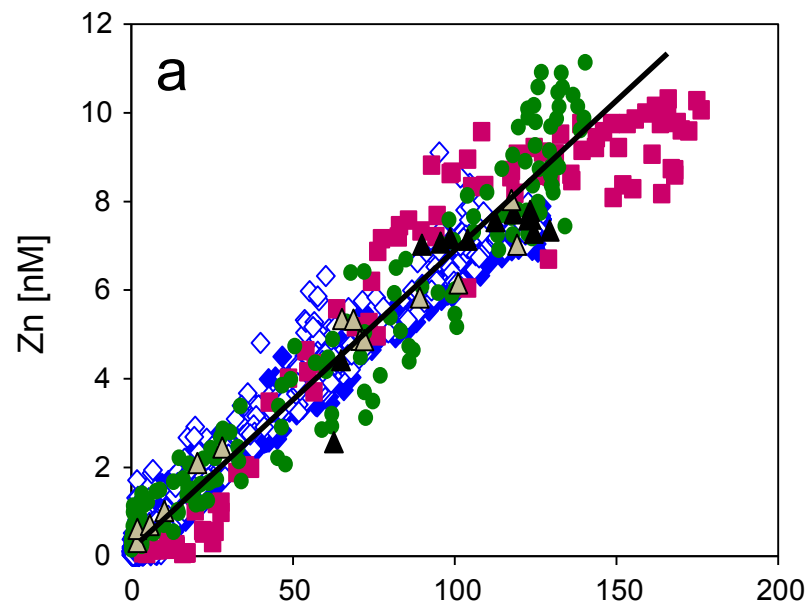
- 367 34. Marshall, J., Adcroft, A., Hill, C., Perelman, L. & Heisey, C. A finite-volume, incompressible
368 Navier-Stokes model for studies of the ocean on parallel computers. *J. Geophys. Res. Oceans* **102**,
369 5733-5752 (1997).
- 370 35. Levitus, S., et al. World Ocean Database 1998, NOAA Atlas NESDIS 18, NOAA, Silver
371 Spring, MD (1998).
- 372 36. Najjar, R.G., et al. Impact of circulation on export production, dissolved organic matter, and
373 dissolved oxygen in the ocean: Results from Phase II of the Ocean Carbon-cycle Model
374 Intercomparison Project (OCMIP-2). *Glob. Biogeochem. Cyc.* **21**, GB3007, doi:
375 10.1029/2006gb002857 (2007).

- 376 37. Najjar, R.G., Sarmiento, J.L. & Toggweiler, J.R.. Downward transport and fate of organic
377 matter in the ocean: Simulations with a general circulation model. *Glob. Biogeochem. Cyc.* **6**, 45-76
378 (1992).
- 379 38. Anderson, L.A. & Sarmiento, J.L. Global ocean phosphate and oxygen simulations. *Glob.*
380 *Biogeochem. Cyc.* **9**, 621-636 (1995).
- 381 39. Garcia, H.E., Locarnini, R.A., Boyer, T.P., Antonov, J.I., Baranova, O.K., Zweng, M.M.,
382 Reagan, J.R. & Johnson, D.R.. World Ocean Atlas 2013, Volume 4: Dissolved Inorganic Nutrients
383 (phosphate, nitrate, silicate), Levitus, S. (Ed.), NOAA Atlas NESDIS 76, Silver Spring, MD (2013).
- 384 40. Martin, J.H., Knauer, G.A., Karl, D.M. & Broenkow, W.W.. VERTEX: carbon cycling in the
385 northeast Pacific. *Deep Sea Res.* **34**, 267-285 (1987).
- 386 41. Berelson, W.M. The flux of particulate organic carbon into the ocean interior: a comparison of
387 four US JGOFS regional studies. *Oceanogr.* **14**, 59-67 (2001).
- 388 42. de Souza, G.F., Slater, R.D., Dunne, J.P. & Sarmiento, J.L. Deconvolving the controls on the
389 deep ocean's silicon stable isotope distribution. *Earth Planet. Sci. Lett.* **398**, 66-76 (2014).









SUPPLEMENTARY INFORMATION TO

Silicon and zinc biogeochemical cycles coupled through the Southern Ocean

Derek Vance, Susan H. Little, Gregory F. de Souza, Samar P. Khatiwala, Maeve C. Lohan, Rob Middag

1. Description of the physical and biogeochemical model framework

Ocean general circulation model simulations were carried out using the transport matrix method (TMM) of Khatiwala et al. (2005), which allows the efficient offline simulation of passive tracers in the ocean by making use of transport matrices (TMs) derived from a full three-dimensional ocean general circulation model. Here, we use TMs derived from a coarse-resolution version of the MITgcm (Marshall et al., 1997) with $2.8^\circ \times 2.8^\circ$ lateral resolution and 15 vertical levels, forced with monthly mean climatological fluxes of momentum, heat and freshwater and with weak restoring of surface temperature and salinity to the Levitus climatology (Levitus et al., 1998). Simulations were carried out with annual-mean circulation fields derived from the equilibrium state of the model after 5000 yr integration; thus, seasonal variability is not represented in our simulations. This physical model is coupled to a biogeochemical model that simulates the internal oceanic cycling of phosphorus (as phosphate, PO_4 , and dissolved organic phosphorus, DOP), zinc (Zn) and silica (Si). Biogeochemical model simulations were initialized with constant nutrient tracer fields and integrated forward for 5000 model years; the output for model year 5000 is presented herein.

PO₄ cycling: The nutrient-cycling model that forms the basis of our simulations is based on the formulation developed for the OCMIP-2 project (Najjar et al., 2007) and uses PO_4 as its nutrient currency. Phosphate uptake in the surface ocean ($\leq 120\text{m}$ water depth) is driven by restoring surface PO_4 concentrations (Najjar et al., 1992; Anderson and Sarmiento, 1995) towards the objectively-analysed annual-mean PO_4 field of World Ocean Atlas 2013 (Garcia et al., 2013) with a restoring timescale of 36.5 days. A fraction (0.67) of PO_4 taken up is immediately shunted towards the production of DOP, whilst the remaining fraction is exported as an implicit particulate flux to depth, which remineralises following a power-law dependency on depth with an exponent of -0.858 (Martin et al., 1987; Berelson, 2001). Any particulate flux reaching the bottom of the water column remineralises within the bottom ocean cell, since we only represent internal oceanic cycling with no sources (or sinks) into (or out of) the ocean. Dissolved organic phosphorus is carried passively with the circulation and decays back to PO_4 with a first-order rate constant of $(0.5 \text{ year})^{-1}$. Mean ocean phosphorus concentration is set to $2.17 \mu\text{M}$ (calculated from World Ocean Atlas 2013; Garcia et al., 2013), all of which is in the form of PO_4 at $t=0$; at equilibrium, mean ocean PO_4 concentration is $2.164 \mu\text{M}$, whilst mean DOP concentration is $0.006 \mu\text{M}$.

Zn cycling: In order to test our mechanistic hypotheses regarding Zn cycling in the ocean, the Zn-cycling model is explicitly coupled to the PO_4 -cycling model above. Zn uptake in the surface ocean is related to PO_4 uptake by a dimensionless stoichiometric parameter $r_{\text{Zn:P}}$. This surface-ocean uptake

drives an implicit export flux of particulate Zn, which remineralises identically to the implicit particulate P flux, i.e. following a power-law dependency on depth with an exponent of -0.858 . Any particulate flux reaching the bottom of the water column remineralises within the bottom ocean cell. The organic speciation of dissolved Zn is represented implicitly by assuming a constant ligand concentration of 1.2 nM with a conditional stability constant of 10^{10} M^{-1} , which reduces the calculation of the concentrations of free Zn (i.e. organically- and inorganically uncomplexed Zn^{2+}) to the solution of a quadratic equation (see below). Varying biogeochemical behaviours of the Zn-cycling model are achieved by the formulation of the stoichiometric parameter $r_{\text{Zn:P}}$, as detailed below. Mean ocean Zn concentration is set to 5.4 nM (Chester and Jickells, 2012).

Si cycling: The silicon cycling model is conceptually very similar to the PO_4 -cycling model and the Si-cycling model of de Souza et al. (2014, 2015). Dissolved Si uptake in the surface ocean ($\leq 120\text{m}$ water depth) is driven by restoring surface Si concentrations towards the objectively-analysed annual-mean Si field of World Ocean Atlas 2013 with a restoring timescale of 36.5 days. This surface-ocean uptake drives an implicit particulate export flux, which dissolves following an exponential dependency on depth with a length-scale of 1000 m (de Souza et al., 2014; de Souza et al., 2015). Any particulate flux reaching the bottom of the water column remineralises within the bottom ocean cell. Mean ocean Si concentration is set to $92 \text{ }\mu\text{M}$ (calculated from World Ocean Atlas 2013; Garcia et al., 2013). It is important to note that in our model formulation, Si cycling is *entirely biogeochemically independent* of PO_4 and Zn cycling.

2. Parameterisation of Zn uptake

2.1. Rationale

The hypothesis proposed in this study is based on the observation that Southern Ocean diatoms possess a Zn quota (relative to P) that is 5 to 13 times higher than that of average oceanic phytoplankton. Export of organic matter with such an elevated Zn:P ratio in the Southern Ocean could thus lead to a decoupling of the large-scale distributions of Zn and PO_4 in a similar manner to that proposed by Sarmiento et al. (2004) for Si, without requiring any mechanistic coupling between Zn and Si.

A potential explanation of the high Zn quota of Southern Ocean phytoplankton may lie in Sunda and Huntsman's (1992) observation of the dependence of phytoplankton Zn uptake (and Zn quota) on the ambient concentration of free Zn (i.e. Zn^{2+}). From the limited observations of Zn speciation, the surface waters of the Southern Ocean represent the only euphotic zone where total dissolved Zn concentrations exceed those of the chelating organic ligand, allowing Zn^{2+} concentrations to rise (Baars and Croot, 2011). In our simulations, we assess the influence of such a Zn^{2+} -dependence of Zn uptake on the large-scale distribution of Zn. We do this via two idealized models of Zn uptake, in which this uptake is either (a) a linear function of total dissolved Zn concentration (model version

LIN) or (b) a non-linear function of Zn^{2+} concentration as in Sunda and Huntsman (1992; model version NONLIN).

2.2. Model description

In both Zn model versions used herein, the implicit assumption is that oceanic concentration of the chelating ligand (denoted L) is constant at 1.2 nM. The existing sparse observations of Zn-complexing ligand concentrations in the ocean (Donat and Bruland, 1990; Bruland, 1989; Ellwood and van den Berg, 2000; Ellwood, 2004; Lohan et al., 2005; Baars and Croot, 2011) do bear witness to some variability, although only to a limited extent (approximately 0.6–2.4 nM) and without any systematic variation.

Model version LIN was motivated by (i) the wish to maximise model simplicity in order to ease interpretation and (ii) the observation that the non-saturating Michaelis-Menten dependence of Zn uptake on Zn^{2+} (see model NONLIN below), together with the simple quadratic dependence of Zn^{2+} on total dissolved Zn (given constant L) combine to produce a near-linear dependence of Zn uptake on total dissolved Zn (see Fig. S1), making this formulation a good first approximation. In this model version, the stoichiometric parameter $r_{Zn:P}$ (mol/mol) that couples the uptake of Zn to that of PO_4 is given by:

$$r_{Zn:P} = m_{Zn} \cdot Zn^T \quad (\text{Eqn. 1})$$

where Zn^T is total dissolved Zn and m_{Zn} is a prescribed slope. The Zn uptake term is then simply calculated as:

$$J_{Zn}^{uptake} = r_{Zn:P} \cdot J_{PO_4}^{uptake} \quad (\text{Eqn. 2})$$

Model version NONLIN was motivated by (i) the wish to maximise fidelity to observations of Zn uptake in culture experiments (Sunda and Huntsman, 1992), and (ii) the inability of LIN to reproduce the location and position of the latitudinal gradient in surface Zn concentrations (see Section 3.2). Following the formulation of Sunda and Hunstman, in this model version the stoichiometric parameter that couples Zn uptake to PO_4 uptake is a non-saturating Michaelis-Menten-type function of Zn^{2+} , i.e. the sum of a Michaelis-Menten function and a linear dependency (the so-called “two-site” model):

$$r_{Zn:P} = \frac{a_{Zn} \cdot Zn^{2+}}{b_{Zn} + Zn^{2+}} + c_{Zn} \cdot Zn^{2+} \quad (\text{Eqn. 3})$$

where parameters a_{Zn} , b_{Zn} and c_{Zn} are constants analogous to maximum uptake rate, half-saturation constant and non-saturable uptake rate, respectively. The concentration of Zn^{2+} is calculated from the implicit concentration of non-ligand-bound Zn (Zn') following the simple relationship:

$$Zn^{2+} = Zn' / \alpha \quad (\text{Eqn. 4})$$

where $\alpha = 2.1$ is the inorganic side-reaction coefficient for Zn in seawater (Ellwood and Van den Berg, 2000). In its turn, Zn' is calculated as the positive root of the quadratic equation that follows from the mass balance of Zn and the chelating ligand L:

$$K_L \cdot (Zn')^2 + (K_L L^T - K_L Zn^T + 1) \cdot Zn' - Zn^T = 0 \quad (\text{Eqn. 5})$$

where $K_L = 10^{10} \text{ M}^{-1}$ is the conditional stability constant for the chelation of Zn by the ligand L, and $L^T = 1.2 \text{ nM}$ is the concentration of total ligand L (i.e., both unbound and Zn-bound L). Finally, as in model LIN above, the Zn uptake term is related to PO_4 uptake following Eqn. 2.

2.3. Uptake parameter choices

In model version NONLIN, a range of values for the parameters a_{Zn} , b_{Zn} and c_{Zn} in Eqn. 3 was obtained by fitting this function to culture data from five diatom and coccolithophorid species/clones from Sunda and Huntsman (1992). Of these parameter sets, the values for the diatom *Thalassiosira oceanica* produce an uptake dependence that lies squarely in the centre of the species variability (Fig. S2). In model version LIN, the standard value of m_{Zn} in Eqn. 1 was set so as to roughly reproduce the dependence of rZn:P on ZnT in Eqn. 3 when using the parameters for *T. oceanica*. Parameters and key statistical metrics for the 11 sensitivity simulations carried out are listed in Table S1.

3. Results

3.1. Global metrics

In the following, we illustrate the degree of similarity between tracer fields using Taylor diagrams (Taylor, 2001), which provide a concise statistical summary of the similarity between two three-dimensional fields as a single point (defined by the normalised centred root mean square (RMS) difference, correlation coefficient, and relative standard deviation of these two fields) on a polar-style graph. We use these diagrams both to quantify model skill at reproducing the global distributions of PO_4 and Si as represented by World Ocean Atlas 2013 (WOA13), as well as to compare the similarity of the global oceanic Zn fields simulated by our two model variants (and corresponding parameter-sensitivity simulations). For all diagrams shown herein, tracer fields are first interpolated to the model grid if required (e.g. for the WOA13 fields), and statistics calculated after weighting the tracer concentration by the corresponding model cell volume (Taylor, 2001).

Figure S3 compares the results of our PO_4 and Si cycling models to the corresponding three-dimensional tracer fields from WOA13. Both models reproduce the large-scale oceanic macronutrient patterns with good fidelity: the correlation coefficient R between model output and WOA13 is 0.87 for PO_4 and 0.92 for Si, whilst the corresponding normalised centred RMS difference \hat{E}' is 0.57 for PO_4 and 0.41 for Si. The PO_4 field simulated by our OCMIP-2-type PO_4 -DOP cycling model compares favourably with other ocean biogeochemistry general circulation models of this type (see e.g. Gnanadesikan et al., 2004).

Figure S4 presents a sensitivity analysis for our Zn cycling model variants, in order to illustrate underlying systematics that produce a Zn distribution that mimics either that of PO_4 or of Si. In order to do this most clearly, Fig. S4 compares the model Zn tracer field with *simulated* PO_4 and Si tracer fields scaled by the ratio between the mean ocean concentrations of these species (i.e. Zn is compared

with $(\overline{Zn}/\overline{PO_4}) \cdot PO_4$ and $(\overline{Zn}/\overline{Si}) \cdot Si$, where the overbar denotes an average over the entire ocean), in order to explore the mechanistic systematics internal to the model (see also Table S1). Model formulations and parameter choices that produce a more Si-like distribution of Zn will thus exhibit a lower RMS error and plot closer towards the “bullseye” of the Taylor diagram. Figure S4 shows that, for both formulations of the dependency of Zn uptake on Zn concentration (i.e. model versions LIN and NONLIN), differing choices of parameter values can lead to very different large-scale Zn distributions. Furthermore, it shows that certain choices of model formulation and parameter values produce Zn distributions that are *very* similar to the Si field (red symbols close to normalised standard deviation of 1 and with correlation coefficients >0.95 ; e.g. Simulation 11), whilst other choices may instead produce PO_4 -like distributions (green symbol for Simulation 1 in the same region).

3.2. Surface-ocean metrics

The Taylor diagrams discussed above are an excellent way of quantifying global similarity in tracer fields. However, due to the fact that cells are weighted by their volume in constructing the relevant statistics, these diagrams are not very sensitive to changes in the surface and upper ocean, whose volume and nutrient concentrations are small relative to those of the deep ocean. Thus, in the following, we focus on a few surface ocean metrics to illustrate the sensitivity of our Zn-cycling model to uptake formulation and parameter choices.

Figure S5 shows the zonal average surface Zn concentration produced by the various sensitivity simulations for which global metrics are presented in Fig. S4 above. Note that whilst model PO_4 and Si distributions are prescribed by the surface-restoring production scheme used by the respective models, the simulated surface Zn field is free to evolve based on the interplay between Zn uptake (which is dependent on PO_4 uptake via Eqn. 2), regeneration and the physical circulation. The fields simulated by model variants LIN (Fig. S5a) and NONLIN (Fig. S5b) are compared to an “ideal” Si-like Zn field that scales as $(\overline{Zn}/\overline{Si}) \cdot Si$ (i.e. one that is linearly correlated with model Si). In Fig. S5a, it can be seen that variant LIN struggles to simultaneously reproduce the low concentrations at mid- to low-latitudes, high concentrations in the subpolar and polar oceans, and sharp latitudinal surface gradient centered at around $55^\circ S$ of the “ideal Zn” field. The sensitivity simulation that produces the most Si-like Zn distribution at the global scale (Simulation 6) strongly underestimates surface Zn concentrations south of about $60^\circ S$ (by about 30%), but does place the zonally-averaged Southern Ocean surface Zn gradient at approximately the right position. In comparison, the model variant NONLIN (Fig. S5b) is capable of not only producing the sharp Zn concentrations differences between the low- and high-latitude surface oceans, but of maintaining this gradient even when the zonally-averaged Southern Ocean surface Zn gradient is moved southwards towards (but not quite reaching) $55^\circ S$. The best-performing sensitivity simulation of this model variant (Simulation 11) produces surface Southern Ocean Zn concentrations within a few percent of the zonally averaged “ideal Zn” and places the strong latitudinal Zn gradient only a few degrees too far north. Relative to this “ideal Zn”, the model does appear to underestimate low-latitude surface Zn concentrations, although observations

show that Zn concentrations in the low-latitude ocean can be extremely low (e.g. <0.1 nM in the low-latitude Atlantic Ocean; Wyatt et al., 2014).

By combining the simulated Zn fields shown in Fig. S5 with Eqns. 3-5, we can diagnose the Zn:P of uptake simulated by these model variants (Fig. S6). From a comparison of Fig. S6 with Figs. S4 and S5, it is clear that the sensitivity simulations in which there is great latitudinal plasticity in Zn:PO₄ uptake ratio, especially with a sharp gradient between the mid- to low-latitude and the subpolar/polar oceans poleward of 40°S, are the most successful at producing a surface and global-ocean Zn distribution that is similar to that of Si. The variability of Zn:PO₄ uptake simulated in our sensitivity simulations is well within the observed plasticity of Zn:P as compiled, for instance, by Twining and Baines (2013).

4. Conclusions

Our sensitivity simulations illustrate the wide variety of elemental distributions that can be simulated as the result of varying a few model parameters that control the stoichiometry of elemental uptake in the surface ocean. The Zn distributions simulated by our suite of models thus ranges from being similar to PO₄ to being tightly and near-linearly correlated with Si (Fig. S4). The particular importance of Southern Ocean uptake stoichiometry in controlling this variation is shown systematically in Fig. S7, which plots the export-weighted mean Zn:PO₄ uptake ratio in the Southern Ocean ($>50^\circ\text{S}$) against the normalised centred RMS difference between the Zn fields simulated by our 11 sensitivity simulations and the simulated Si and PO₄ fields (scaled by the appropriate ocean-mean elemental ratio, as detailed in Section 3.1). It can be seen that elevated Zn:PO₄ uptake in the Southern Ocean systematically results in a more “Si-like” global Zn distribution, regardless of the specifics of our model formulation (LIN vs. NONLIN). As discussed in Section 3.2, the simulation that best reproduces both global (Fig. S4) and surface (Fig. S5) Zn distributions is Simulation 11 (Table S1), a version of model variant NONLIN in which the Zn:PO₄ of uptake increases rapidly at low Zn²⁺ and almost saturates thereafter; this feature seems to produce a sharp gradient in Zn:PO₄ of uptake across the subpolar oceans (Fig. S6) and may be responsible for the model’s skill at reproducing the sharp surface gradient in Zn *concentrations* here (Fig. S5). It is this simulation for which we show results in the main text (Fig. 3).

At this juncture it is worth explicitly noting once again that the similarity in Zn and Si fields in our simulations comes about *without* any coupling between Zn and Si in the models and *despite* the difference in their regeneration lengthscales, since Zn is cycled identically to PO₄ in our models. Rather, this similarity is driven by the geographically collocated, but mechanistically unrelated, strong uptake of Zn and Si by Southern Ocean phytoplankton communities.

Table S1. Details of the sensitivity simulations performed, including key parameter values and normalised centred RMS difference (\hat{E}') of the appropriately scaled model Zn field to simulated Si and PO₄ fields (see Section 3.1).

Simulation	Model variant	Description	Key parameter values			\hat{E}'		
			m_{Zn} [mol:mol/ μ M]	a_{Zn} [mol:mol]	b_{Zn} [μ M]	c_{Zn} [mol:mol/ μ M]	vs. Si	vs. PO ₄
1	LIN	0.25× standard slope					0.53	0.16
2	LIN	0.5 standard slope					0.43	0.37
3	LIN	Standard slope (Eqn. 1 fit to <i>T. oceanica</i>)					0.34	0.59
4	LIN	1.5× standard slope					0.30	0.70
5	LIN	2× standard slope					0.27	0.77
6	LIN	3× standard slope					0.26	0.86
7	NONLIN	Eqn. 3 fit to <i>E. huxleyi</i> A1383	2.4×10^{-3}	4.0×10^{-5}	4.7		0.30	0.71
8	NONLIN	Eqn. 3 fit to <i>T. weissflogii</i>	1.6×10^{-3}	1.7×10^{-5}	4.0		0.32	0.66
9	NONLIN	Eqn. 3 fit to <i>T. oceanica</i>	1.25×10^{-3}	1.0×10^{-5}	2.9		0.35	0.6
10	NONLIN	Eqn. 3 fit to <i>T. pseudonana</i>	1.2×10^{-3}	8.0×10^{-6}	1.7		0.39	0.52
11	NONLIN	Eqn. 3 fit to <i>E. huxleyi</i> BT6	6.0×10^{-3}	3.0×10^{-5}	0.32		0.26	0.81

Fig. S1

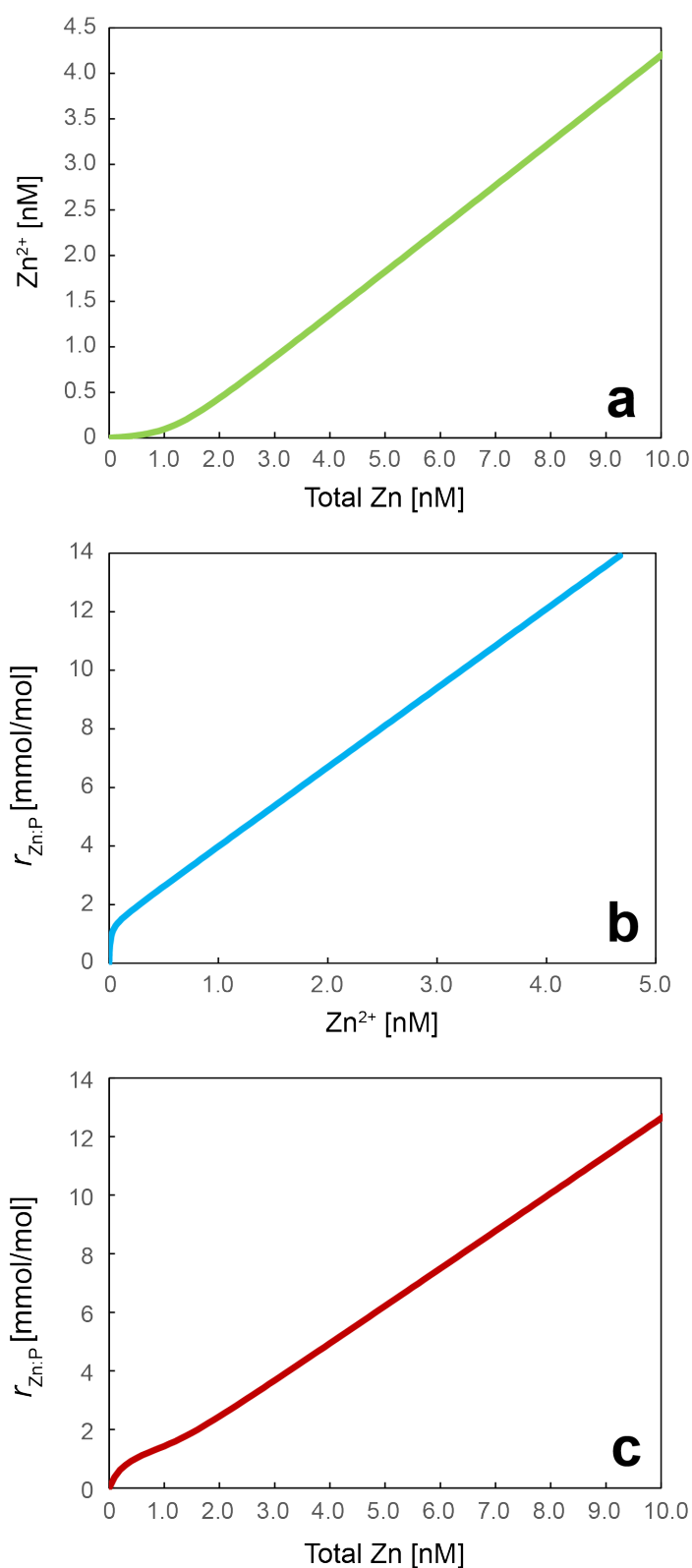


Fig. S1: Systematics of Zn speciation and Zn:P uptake. Relationship between (a) concentration of Zn^{2+} and total Zn in the presence of a chelating organic ligand (ligand concentration 1.2 nM, conditional stability constant $10^{10} M^{-1}$) and (b) the non-linear “two-site” dependence of P-specific Zn uptake $r_{Zn:P}$ on Zn^{2+} concentration, based on culture data of Sunda and Huntsman (1992). These two dependencies combine to produce the near-linear dependence of $r_{Zn:P}$ on total Zn in panel c.

Fig. S2

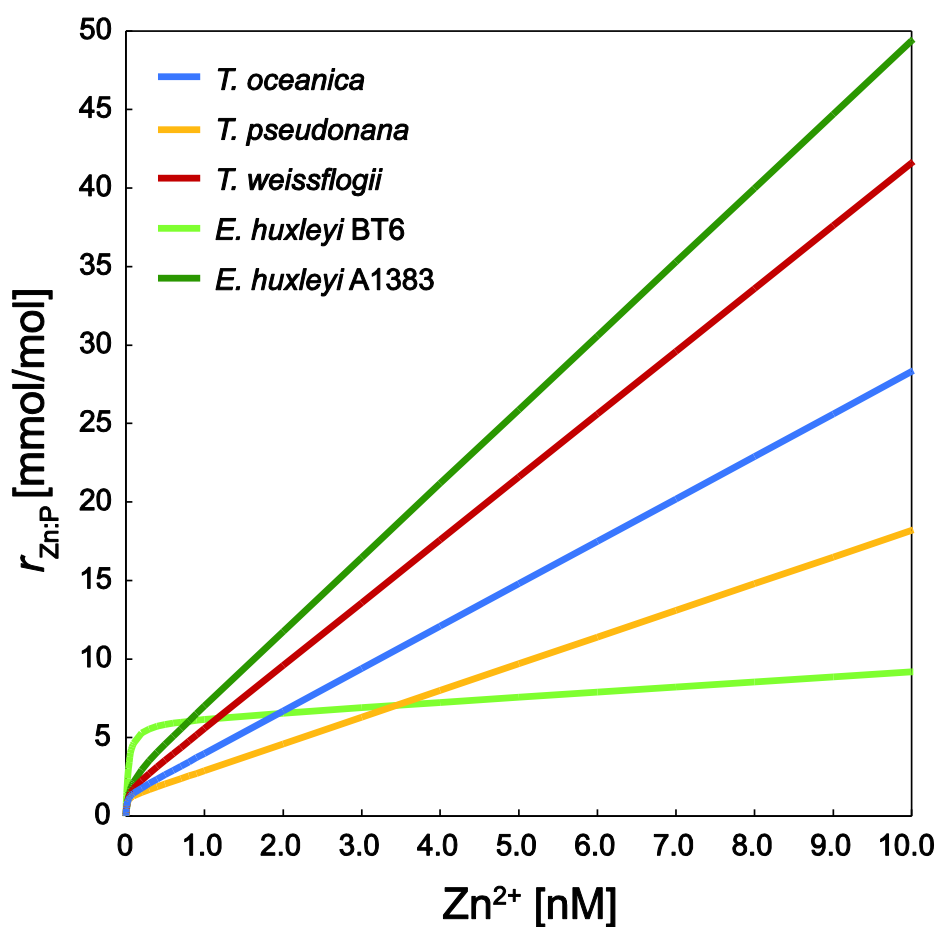


Fig. S2: Variability in specific Zn uptake dependency in phytoplankton. “Two-site” model curves (Eqn. 3) fit to culture data from Sunda and Huntsman (1992). Considerable variability is observed, both between genera and between clones of the same species. We apply this large observed variability in the dependence of $r_{Zn:P}$ on Zn^{2+} concentrations in 5 parameter sensitivity simulations of the model variant NONLIN in order to assess the extent to which surface uptake processes may affect the large-scale Zn distribution.

Fig. S3

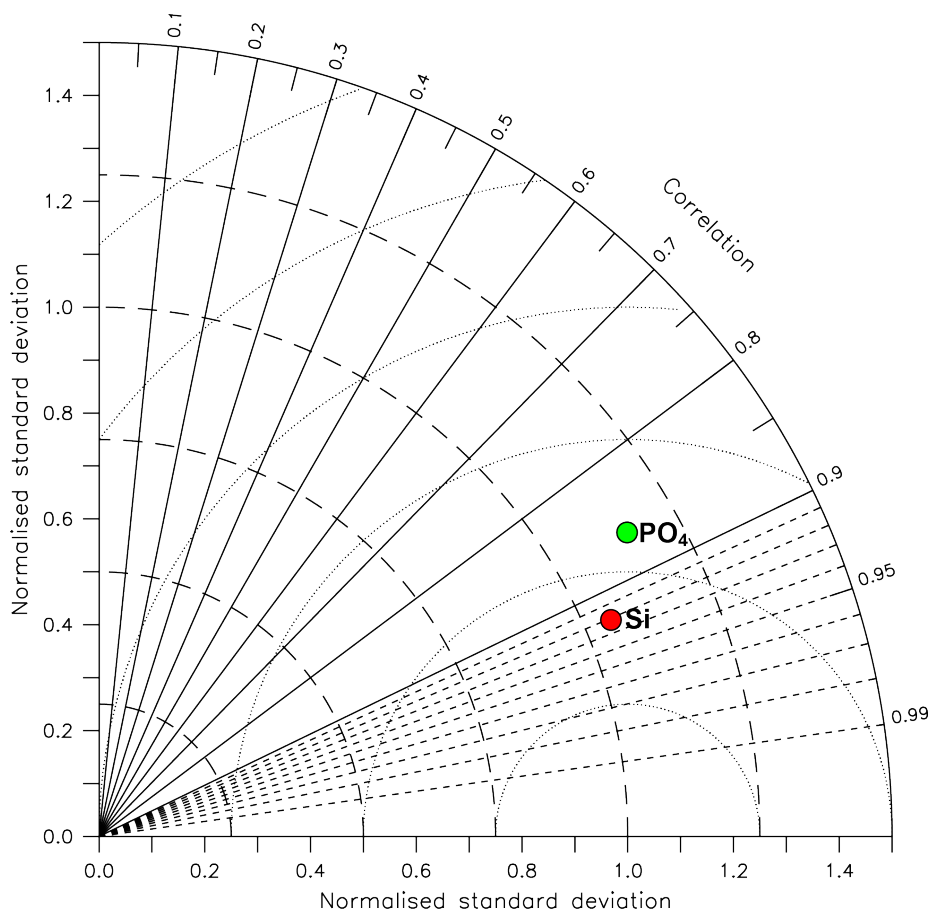


Fig. S3: Evaluation of PO₄ and Si model skill. Volume-weighted Taylor diagram comparing the three-dimensional simulated PO₄ and Si tracer fields to the corresponding objectively-analysed fields in World Ocean Atlas 2013 (Garcia et al., 2013). Both model fields exhibit good fidelity to the data product, with normalised standard deviations close to 1 and correlation coefficients above 0.85.

Fig. S4

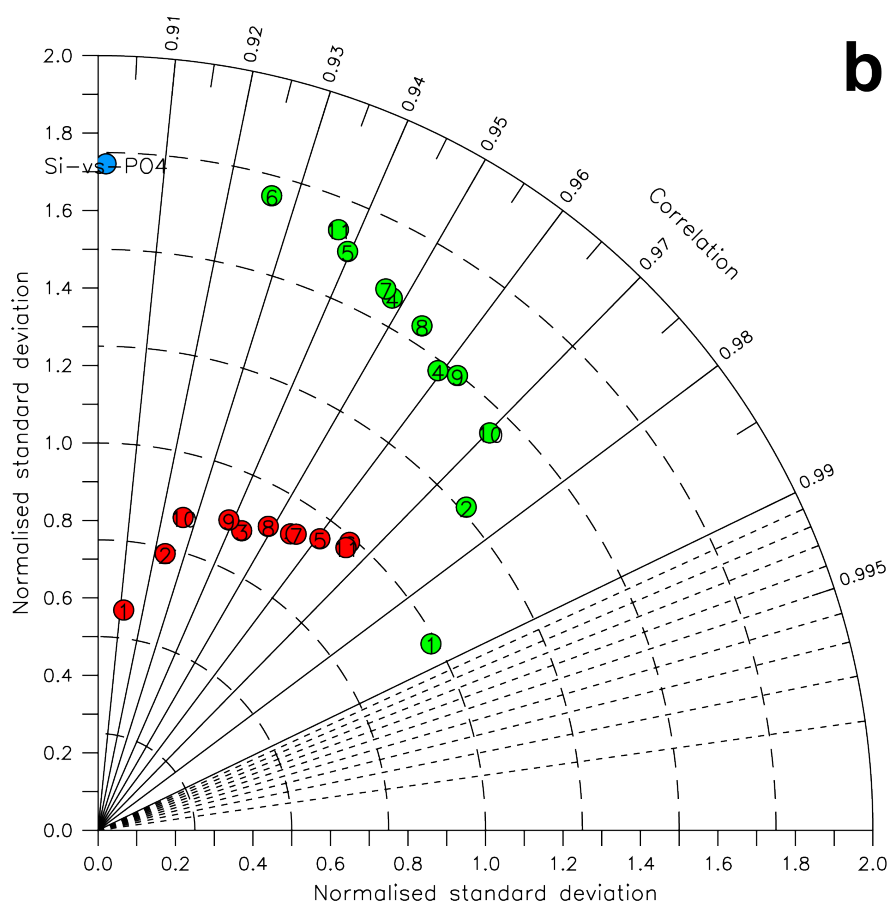
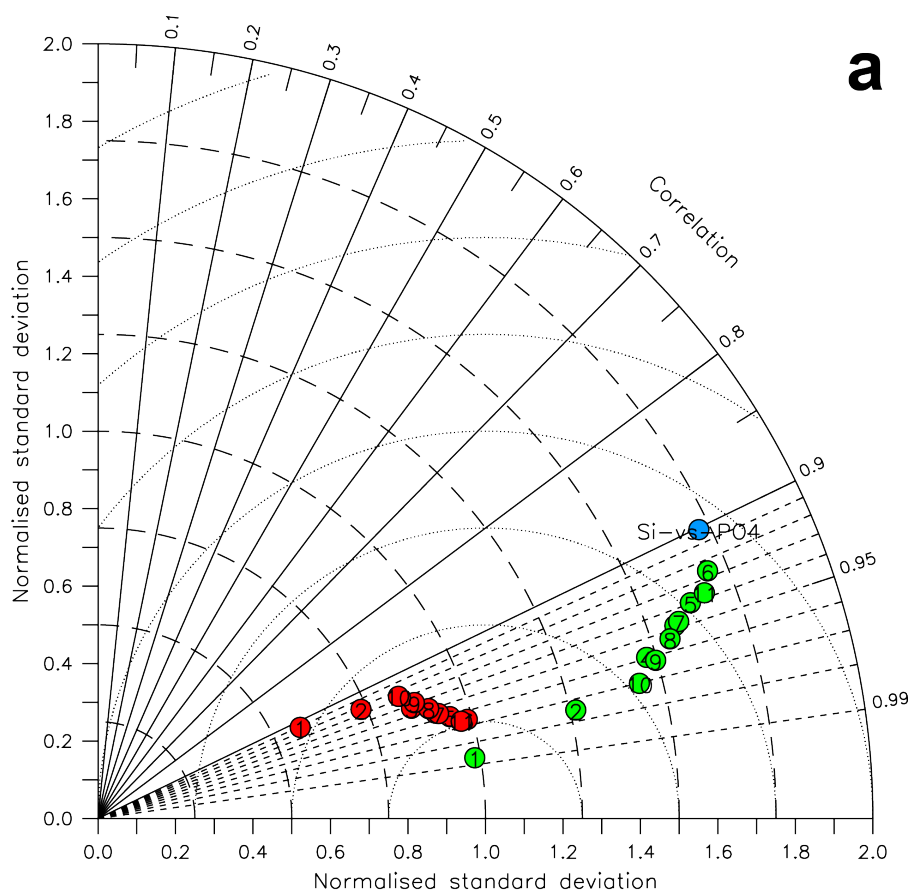


Fig. S4: Comparison of simulated Zn distributions to model Si and PO₄ fields. Each circle in the volume-weighted Taylor diagram above represents a parameter-sensitivity simulation of the Zn cycling model variants LIN (6 simulations) and NONLIN (5 simulations) and compares the appropriately scaled three-dimensional Zn field to the simulated Si field (red circles) or PO₄ field (green circles; see Section 3.1). The blue circle is included as reference and illustrates the degree of similarity between the appropriately scaled simulated Si and PO₄ fields. It can be seen that, depending on parameter choices in either of the two model variants, the simulated Zn distribution may be either similar to the Si tracer field (red circles close to a normalised standard deviation of 1 and with high correlation coefficient) or to the PO₄ tracer field (green circle in the same region). Panel *b* presents the same results as panel *a* with an enlarged scale for the correlation coefficient (0.9 – 1.0) for better clarity. See Table S1 for details of the simulations.

Fig. S5

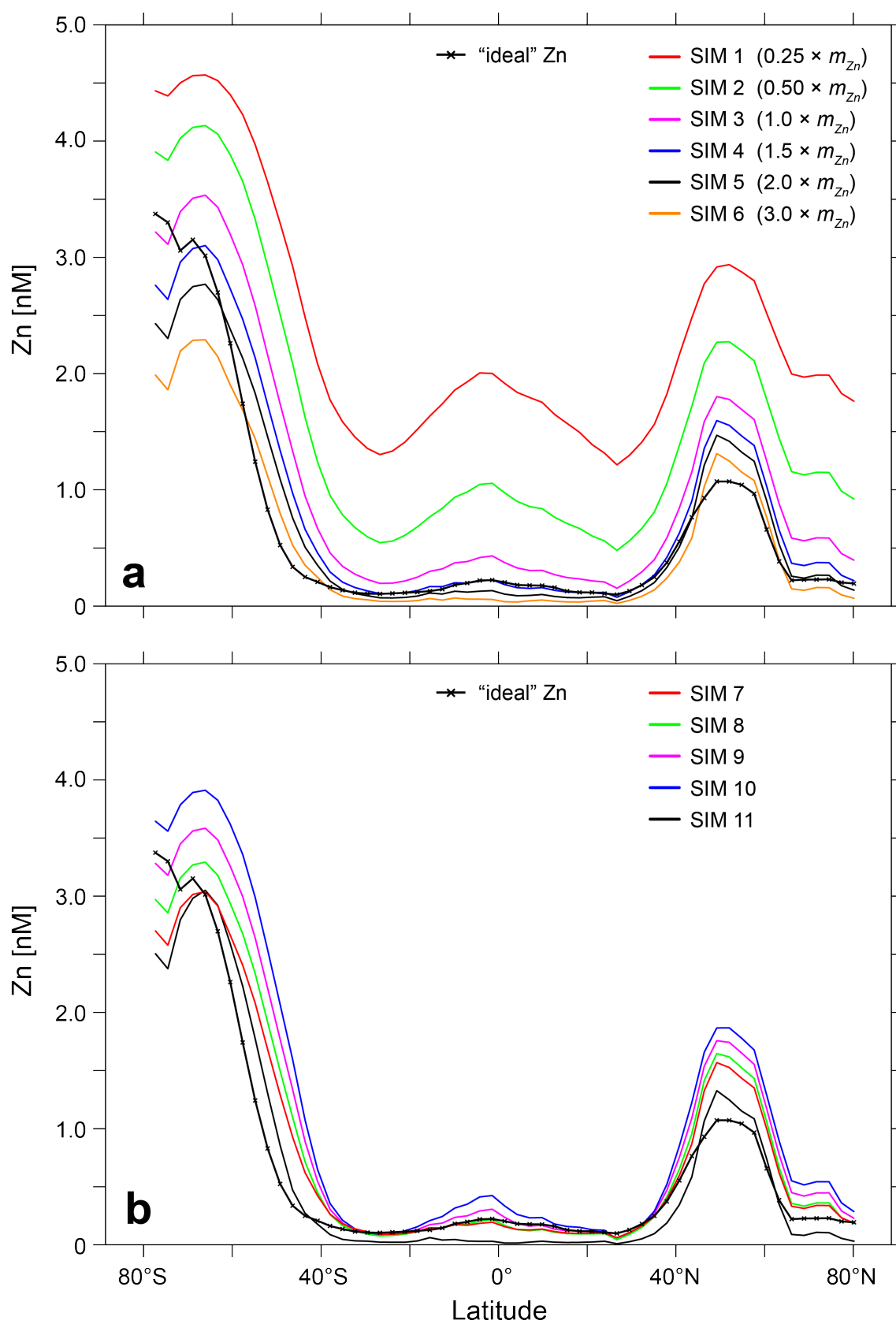


Fig. S5: Zonal mean surface Zn concentration. Results from the parameter sensitivity simulations of the two model variants (a) LIN and (b) NONLIN are shown relative to an hypothetical "ideal" Zn distribution that scales linearly with Si. See Table S1 for details of the simulations.

Fig. S6

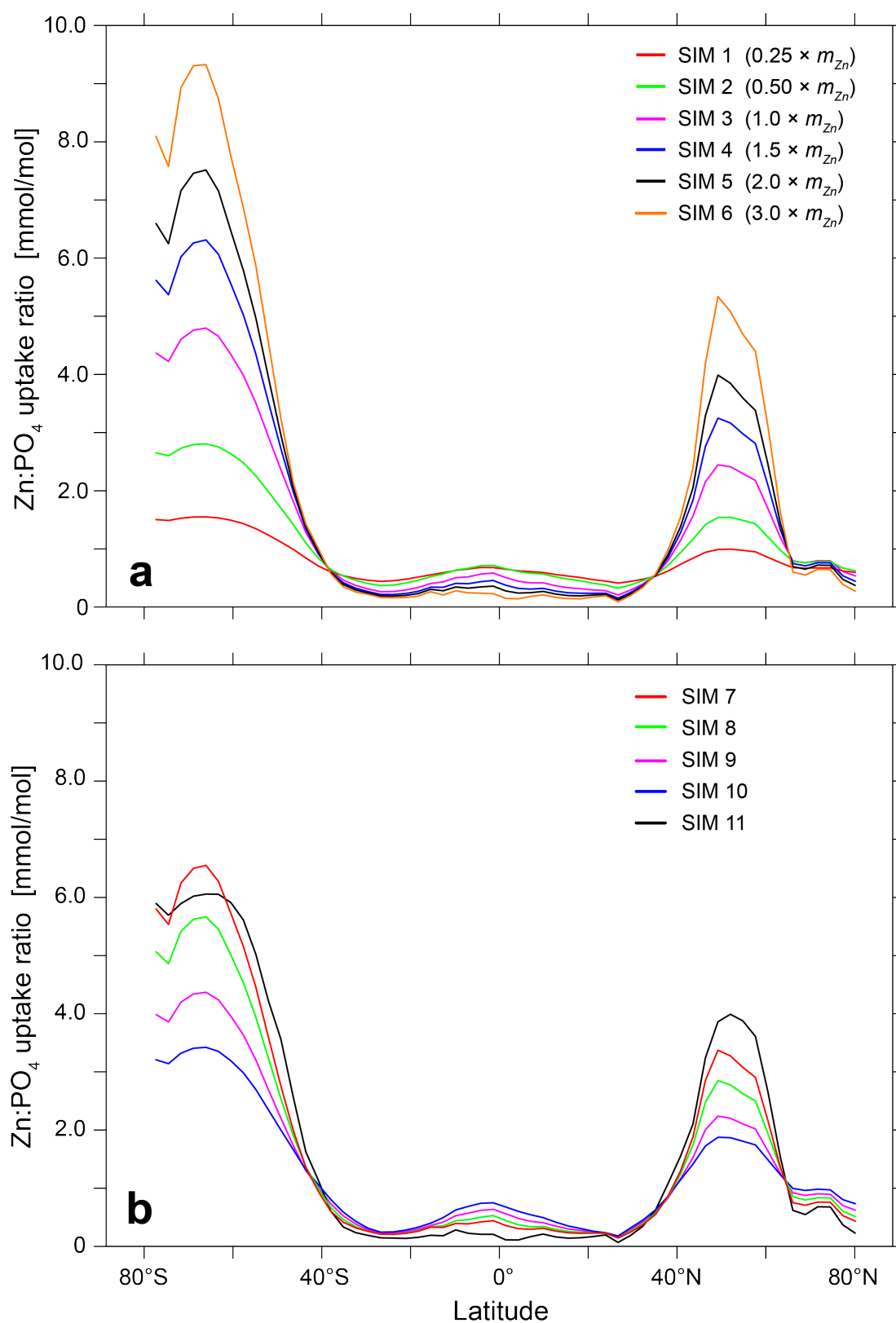


Fig. S&: Zonal mean Zn:PO₄ uptake ratio. Both model variants (a) LIN and (b) NONLIN can exhibit strong latitudinal variability in Zn:PO₄ uptake depending on the choice of parameter values. Comparison with Figs. S4 and S5 above reveals that simulations with greater latitudinal gradients in Zn:PO₄ of uptake produce surface *and* global ocean Zn distributions that more closely resemble the Si distribution (see also Fig. 3 of the main text). See Table S1 for details of the simulations.

Fig. S7

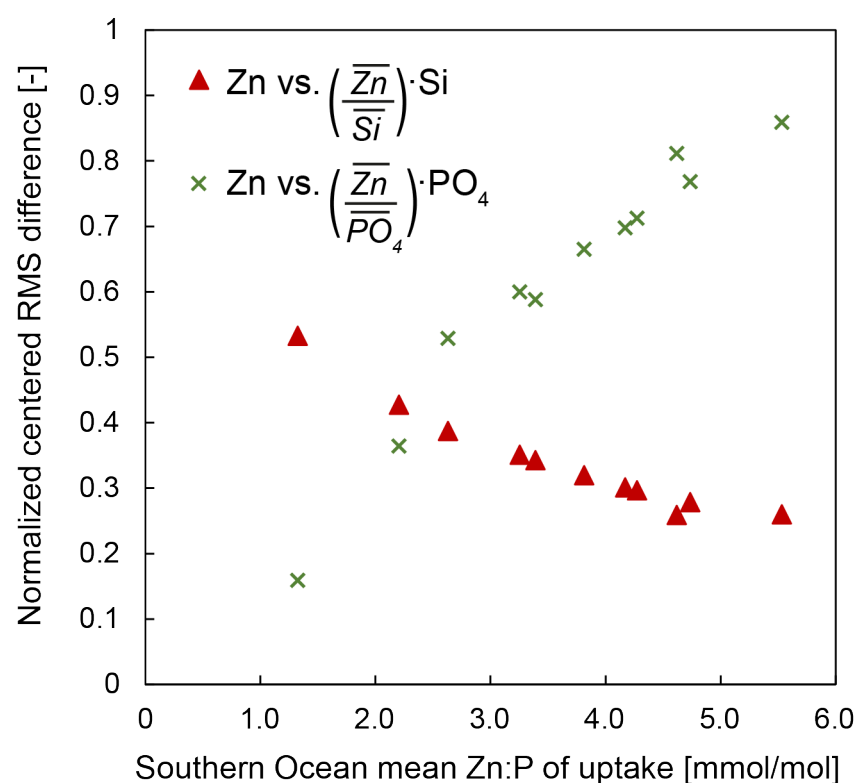


Fig. S7: Systematics of the control of Southern Ocean uptake stoichiometry on global Zn distribution. As the export-weighted mean Zn:P uptake ratio in the Southern Ocean (>50°S) increases in our simulations, the simulated Zn field becomes increasingly “Si-like”, whilst its similarity to the simulated PO₄ field decreases, as reflected by the increase in normalised centred RMS difference \hat{E}' between the two fields. Export-weighted means were calculated as the ratio of the total Zn export flux south of 50°S to that of P.

References

- Anderson, L.A., Sarmiento, J.L., 1995. Global ocean phosphate and oxygen simulations. *Glob. Biogeochem. Cyc.* 9, 621-636.
- Baars, O., Croot, P.L., 2011. The speciation of dissolved zinc in the Atlantic sector of the Southern Ocean. *Deep Sea Res. II* 58, 2720-2732.
- Berelson, W.M., 2001. The flux of particulate organic carbon into the ocean interior: a comparison of four US JGOFS regional studies. *Oceanogr.* 14, 59-67.
- Bruland, K.W., 1989. Complexation of zinc by natural organic ligands in the central North Pacific. *Limnol. Oceanogr.* 34, 269-285.
- Chester, R., Jickells, T., 2012. *Marine Geochemistry*, 3rd ed. Wiley-Blackwell, Chichester.
- de Souza, G.F., Slater, R.D., Dunne, J.P., Sarmiento, J.L., 2014. Deconvolving the controls on the deep ocean's silicon stable isotope distribution. *Earth Planet. Sci. Lett.* 398, 66-76.
- de Souza, G.F., Slater, R.D., Hain, M.P., Brzezinski, M.A., Sarmiento, J.L., 2015. Distal and proximal controls on the silicon stable isotope signature of North Atlantic Deep Water. *Earth Planet. Sci. Lett.* 432, 342-353.
- Donat, J.R., Bruland, K.W., 1989. A comparison of two voltametric techniques for determining zinc speciation in Northeast Pacific Ocean waters. *Mar. Chem.* 28, 301-323.
- Ellwood, M.J., 2004. Zinc and cadmium speciation in sub-Antarctic waters east of New Zealand. *Mar. Chem.* 87, 37-58.
- Ellwood, M.J., van den Berg, C.M.G., 2000. Zinc speciation in the Northeastern Atlantic Ocean. *Mar. Chem.* 68, 295-306.
- Garcia, H.E., Locarnini, R.A., Boyer, T.P., Antonov, J.I., Baranova, O.K., Zweng, M.M., Reagan, J.R., Johnson, D.R., 2013. *World Ocean Atlas 2013, Volume 4: Dissolved Inorganic Nutrients (phosphate, nitrate, silicate)*, Levitus, S. (Ed.), *NOAA Atlas NESDIS 76*, Silver Spring, MD.
- Gnanadesikan, A., Dunne, J., Key, R., Matsumoto, K., Sarmiento, J.L., Slater, R., Swathi, P.S., 2004. Oceanic ventilation and biogeochemical cycling: Understanding the physical mechanisms that produce realistic distributions of tracers and productivity. *Glob. Biogeochem. Cyc.* 18, doi:10.1029/2003GB002097.
- Khatiwal, S., Visbeck, M., Cane, M.A., 2005. Accelerated simulation of passive tracers in ocean circulation models. *Ocean Modell.* 9, 51-69.
- Levitus, S., et al., 1998. *World Ocean Database 1998, NOAA Atlas NESDIS 18*, NOAA, Silver Spring, MD.
- Lohan, M.C., Crawford, D.W., Purdie, D.A., Statham, P.J., 2005. Iron and zinc enrichments in the northeastern subarctic Pacific: ligand production and zinc availability in response to phytoplankton growth. *Limnol. Oceanogr.* 50, 1428-1437.
- Marshall, J., Adcroft, A., Hill, C., Perelman, L., Heisey, C., 1997. A finite-volume, incompressible Navier-Stokes model for studies of the ocean on parallel computers. *J. Geophys. Res. Oceans* 102, 5733-5752.
- Martin, J.H., Knauer, G.A., Karl, D.M., Broenkow, W.W., 1987. VERTEX: carbon cycling in the northeast Pacific. *Deep Sea Res.* 34, 267-285.
- Najjar, R.G., Sarmiento, J.L., Toggweiler, J.R., 1992. Downward transport and fate of organic matter in the ocean: Simulations with a general circulation model. *Glob. Biogeochem. Cyc.* 6, 45-76.
- Najjar, R.G., et al., 2007. Impact of circulation on export production, dissolved organic matter, and dissolved oxygen in the ocean: Results from Phase II of the Ocean Carbon-cycle Model Intercomparison Project (OCMIP-2). *Glob. Biogeochem. Cyc.* 21, doi: 10.1029/2006gb002857.
- Sarmiento, J.L., Gruber, N., Brzezinski, M.A., Dunne, J.P., 2004. High latitude controls of thermocline nutrients and low latitude biological productivity. *Nature* 427, 56-60.
- Sunda, W.G., Huntsman, S.A., 1992. Feedback interactions between zinc and phytoplankton in seawater. *Limnol. Oceanogr.* 37, 25-40.
- Taylor, K.E., 2001. Summarizing multiple aspects of model performance in a single diagram. *J. Geophys. Res. Atm.* 106, 7183-7192.
- Twining, B.S., Baines, S.B., 2013. The trace metal composition of marine phytoplankton. *Ann. Rev. Mar. Sci.* 5, 191-215.
- Wyatt, N.J., Milne, A., Woodward, E.M.S., Rees, A.P., Browning, T.J., Bouman, H.A., Worsfold, P.J., Lohan, M.C., 2014. Biogeochemical cycling of dissolved zinc along the GEOTRACES South Atlantic transect GA10 at 40°S. *Glob. Biogeochem. Cyc.* 28, doi: 10.1002/2013gb004637.