

Contribution of the vertical movement of dissolved organic carbon to carbon allocation in two distinct soil types under Castanopsis fargesii Franch. and C. carlesii (Hemsl.) Hayata forests

Youtao Si, Li Xiong, Yuehmin Chen, Jinmao Zhu, Jinsheng Xie, Ren Gao,

Yusheng Yang

► To cite this version:

Youtao Si, Li Xiong, Yuehmin Chen, Jinmao Zhu, Jinsheng Xie, et al.. Contribution of the vertical movement of dissolved organic carbon to carbon allocation in two distinct soil types under Castanopsis fargesii Franch. and C. carlesii (Hemsl.) Hayata forests. Annals of Forest Science, Springer Verlag/EDP Sciences, 2018, 75 (3), pp.79. 10.1007/s13595-018-0756-0. hal-02267135

HAL Id: hal-02267135 https://hal.archives-ouvertes.fr/hal-02267135

Submitted on 19 Aug 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

RESEARCH PAPER



Contribution of the vertical movement of dissolved organic carbon to carbon allocation in two distinct soil types under *Castanopsis fargesii* Franch. and *C. carlesii* (Hemsl.) Hayata forests

Youtao Si^{1,2} · Li Xiong^{1,3} · Yuehmin Chen^{1,2} · Jinmao Zhu^{1,2} · Jinsheng Xie^{1,2} · Ren Gao^{1,2} · Yusheng Yang^{1,2}

Received: 21 July 2017 / Accepted: 6 July 2018 / Published online: 13 August 2018 © INRA and Springer-Verlag France SAS, part of Springer Nature 2018

Abstract

• *Key message* The vertical transport of dissolved organic carbon (DOC) is an important determinant of carbon distribution across a soil profile. The transport of DOC down a soil profile can be largely influenced by incoming DOC and soil organic carbon (SOC) levels, which insulate DOC from adsorption processes regulated by soil texture and Fe/Al mineralogy.

• *Context* Uncertainties about how soil properties affect DOC transport through the soil profile require study because soils can differ strongly with respect to texture or Fe/Al mineralogy and yet retain similar quantities of DOC.

• *Aims* This study aimed to assess the role of incoming DOC and native SOC in regulating DOC migration in soils and investigate the contribution of DOC movement to SOC allocation.

• *Methods* We leached a standard DOC solution extracted from *Castanopsis carlesii* litter through two distinct soil types, using two leaching strategies: single leaching and sequential leaching. The two soil types under a natural *Castanopsis carlesii* (Hemsl.) Hayata forest and a natural *Castanopsis fargesii* Franch. forest, respectively, differ strongly with respect to soil texture, Fe/Al oxide abundances, and SOC nature.

Handling Editor: Ana Rincón
Contribution of the co-authors
Dr. Youtao Si participated in every step of the study and especially put effort in experimental design, DOM characterization, data analysis, and finally writing the paper.
Li Xiong performed part of the experiment.
Yuehmin Chen supervised the work and wrote part of the paper.
Jinmao Zhu ran part of the data analysis.
Jinsheng Xie provided advices on how to write the paper.
Ren Gao coordinated the research project.
Yusheng Yang supervised the work.

☑ Yuehmin Chen ymchen@fjnu.edu.cn

⊠ Yusheng Yang geoyys@fjnu.edu.cn

> Youtao Si yt.si@fjnu.edu.cn

Li Xiong 1326765030@qq.com

Jinmao Zhu jmzhu@fjnu.edu.cn

Jinsheng Xie jshxie@163.com Ren Gao r.gao@hotmail.com

- ¹ State Key Laboratory for Subtropical Mountain Ecology of the Ministry of Science and Technology and Fujian Province, Fujian Normal University, Fuzhou 350007, China
- ² Institute of Geography, Fujian Normal University, Fuzhou 350007, China
- ³ Institute of Resource, Ecosystem and Environment of Agriculture, and Center of Climate Change and Agriculture, Nanjing Agriculture University, Nanjing 210095, China



🖄 Springer

• *Results* With single leaching, where each of six soil layers making up an entire 0–100-cm soil depth profile received single doses of standard DOC solution, deeper soil layers retained more DOC than upper soil layers, with native SOC largely masking the effects of soil texture and Fe/Al mineralogy on DOC migration. Following sequential leaching, where a sixfold larger amount of standard DOC solution sequentially percolated through the six soil layers, the upper soil layers generally retained more DOC than deeper layers. Nevertheless, in sequential leaching, desorption-induced transfer of carbon from upper soil layers to deeper soil layers resulted in greater total carbon retention than in single leaching.

• *Conclusion* Forest subsoils (40–100 cm) are well below C saturation, but DOC vertical movement from top soils only transfers limited organic carbon to them. However, DOC vertical movement may greatly alter SOC allocation along the top soil profile (0–40 cm), with part of outer sphere native SOC displaced by incoming DOC and migrating downwards, which is a natural way to preserve SOC.

Keywords Dissolved organic carbon · Adsorption · Desorption · Soil organic carbon allocation

1 Introduction

Soil organic carbon (SOC) is a sensitive indicator of soil quality and a large component of the global carbon budget. Dissolved organic carbon (DOC) is the most reactive component of SOC because it is highly mobile and contains diverse functional groups. It is a major vehicle for the movement of carbon from surface to deep soils and contributes significantly to SOC dynamics and sequestration (Kalbitz et al. 2000; Tipping et al. 2012; Toosi et al. 2012). It was reported that 25–98% of SOC is associated with the fine soil fraction, and the dominant mechanism is chemically sorptive protection of DOC (Basile-Doelsch et al. 2007; Jagadamma et al. 2014).

Early work on DOC chemistry assumed that only surfacereactive (highly sorptive) substances such as phenolic acids were preferentially removed from incoming DOC by adsorption, whereas no desorption of native SOC occurred (Guggenberger and Kaiser 2003; Jason and Gregory 2001). A cycling downwards model was then proposed, in which, besides adsorption, desorption of microbial processed carbon by fresh plant-derived carbon also drives DOC movement (Kaiser and Kalbitz 2012). Scott and Rothstein (2014) used DOC fractionation to show that DOC collected from deeper soils included desorbed SOC from upper soils and corroborated that desorption was involved in net sorption dynamics across a soil profile. The idea of potential solubility of sorbed organic carbon has now been widely incorporated into hydrological modeling (Dusek et al. 2017; Yurova et al. 2008).

DOC adsorption and desorption occur throughout the soil profile, and the balance of these processes depends on interactions between soil surfaces and incoming DOC. Forest soil properties like soil texture and abundance of Fe/Al oxides may influence these interactions (Kaiser and Guggenberger 2000; Kothawala et al. 2009; Schmidt et al. 2011). Nevertheless, the extent to which soil properties impact DOC migration is uncertain. Scott and Rothstein (2014) found that six contrasting types of forest soil retained similar quantities of DOC when supplied with a common DOC solution, despite wide variation

🖄 Springer 🔳



in textural properties (clay content ranged from 0.1 to 5.2%) and Fe/Al mineralogy (iron hydrous oxide content ranged from 2.9 to 11.6%). Top soils (typically 0–40 cm deep) with higher SOC contents often have limited binding sites, but in the fields, they often show higher DOC retention than subsoils (typically 40–100 cm deep) which contain more clays and Fe/Al oxides (Kalbitz and Kaiser 2008; Kothawala et al. 2009; Lilienfein et al. 2004).

Solving this paradox will require more fully understanding the nature of SOC and the covering of mineral soil surfaces, which contrasts juvenile minerals. It has been demonstrated that SOC can impede exogenous DOC from reacting with soil minerals (Guggenberger and Kaiser 2003), thereby reducing the sorptive capacity of soil particles and perhaps causing soils that differ with respect to texture or mineralogy to exhibit similar retention rates. Inner sphere SOC is directly associated with minerals, and reduces the availability of binding sites on the soil surface, and so is it typically associated with long residence times. In contrast, outer sphere SOC is loosely bound and interacts more readily with exogenous DOC (Guggenberger and Kaiser 2003; Kleber et al. 2015). The outer sphere SOC either adsorbs exogenous DOC or is displaced by the DOC, depending on the nature of the exogenous DOC and the outer sphere SOC itself. Hot waterextractable organic carbon (HEOC) often represents the most reactive part of the outer sphere SOC (Bu et al. 2010), with total SOC amount and chemistry determining outer sphere SOC amount and chemistry, which in turn can determine interactions with DOC inputs. Critically, few studies have explored the role of SOC in regulating the vertical movement of DOC through soils with contrasting chemical and physical properties.

A second entry point into solving the above paradox is understanding how the amount and quality of incoming DOC affect adsorption and desorption dynamics across a soil profile. The more highly sorptive DOC is preferentially retained in upper soil layers, which diminishes the quantity and sorptive quality of the DOC that enters deeper soil layers and so reduces retention rates in deeper soils (Guggenberger and Kaiser 2003; Kothawala et al. 2009). There is, however, very little information on how DOC chemistry and amount affect retention dynamics across a soil profile. We suggest that it is possible to examine the factors that affect DOC retention rates by examining retention efficiency and sorption/ desorption dynamics across a soil profile by subjecting contrasting soil profiles and different soil layers within each profile to fixed amounts of a common DOC.

Organic matter fractionation into hydrophobic and hydrophilic components is a standard method for analyzing organic carbon quality in aqueous solutions. It is widely believed that hydrophobic compounds are more sorptive than hydrophilic compounds (Gu et al. 1994; Kaiser and Guggenberger 2000). Hydrophilic compounds are more likely to be displaced or remain in soil solution and move into deeper soils. Therefore, their concentration would increase with increasing soil depth (Yano et al. 2005). Ultraviolet-visible (UV) and fluorescence spectroscopy have recently been applied in ecological studies as simple but efficient ways to characterize DOC (Bu et al. 2010; Bu et al. 2011; Choe et al. 2010; Colombo et al. 2014; Dai et al. 2006; Foresti et al. 2003; Kim and Hyun 2014; Kleber et al. 2011). The aromaticity index (AI), obtained by dividing the UV-specific absorbance at 254 nm by DOC concentration, can reflect the content of aromatic compounds, whose conjugated π electron system is highly sorptive (Corvasce et al. 2006; Kalbitz et al. 2003; McKnight et al. 1992; Weishaar et al. 2003). The humification index (HIX) calculated from fluorescence spectra is indicative of the microbially processed molecules, such as products derived from microbial decomposition or the condensation of simple structural components (Michel et al. 2006; Ohno 2002; Zsolnay et al. 1999). The combination of fractionation and spectroscopic characterization can provide a more comprehensive view of DOC chemistry.

In this study, we investigated DOC leaching dynamics and chemical changes through two types of soils, each spanning the 0-100-cm depth range and physically divided into six fixed but separated depth layers. Two leaching strategies were used: (i) single leaching, in which each soil layer received a fixed amount of a standard DOC solution and (ii) sequential leaching, where a sixfold larger amount of a standard DOC solution percolated successively through each of the six layers. The standard DOC solution was extracted from the Castanopsis carlesii litter. The two soil types are from a natural C. carlesii forest and a natural C. fargesii forest in midsubtropical China, respectively. The two soil types were chosen to demonstrate the effect of native soil conditions on DOC leaching dynamics. Besides significant differences in soil texture and Fe/Al oxide abundances, differing vegetation composition between the two forest stands results in distinct SOC nature, which are expected to significantly influence DOC dynamics. In addition, because many natural C. fargesii stands have been converted to *C. carlesii* plantations, using DOC extracted from the *C. carlesii* litter as the standard source of DOC could also provide insight into how forest conversion would affect SOC allocation along depth by altered aboveground DOC input. Combining organic matter fractionation and spectroscopic characterization, this study aimed to (1) assess the roles of incoming DOC and native SOC chemistry in regulating soil DOC migration and (2) investigate the contribution of DOC adsorption/desorption to SOC allocation.

2 Materials and methods

2.1 Study area

Two dominant types of forest stands in Fujian Province of subtropical China ($25^{\circ} 29' \sim 27^{\circ} 07'$ N, $116^{\circ} 22' \sim 118^{\circ} 39'$ E) were selected for soil sampling. Both stands are located in a maritime monsoon climate zone where the mean annual temperature is about 19 °C. One stand is a natural *C. carlesii* forest, located in the *Castanopsis kawakamii* Hayata Nature Reserve of Sanming county ($26^{\circ} 10'$ N, $117^{\circ} 27'$ E). This region has a mean annual precipitation of 1749 mm. Soils in this area are classified as sandy clay loam Ferralic Cambisols (hereafter, FC) according to FAO/UNESCO. The other stand is a natural *C. fargesii* forest located in Ninghua county ($26^{\circ} 19'$ N, $116^{\circ} 49'$ E). This region has a mean annual precipitation of 1688 mm and soils are classified as Humic Alfisols (hereafter, HA).

2.2 Soil sampling and soil core preparation

To obtain a wide range of soil conditions, soils were sampled down to 100-cm depth. In July 2015, we selected three sampling points in each of the two stand types with a distance of \geq 10 m between points. At each point, we collected six soil segments at 0-10, 10-20, 20-40, 40-60, 60-80, and 80-100 cm deep. Visible litter and rocks were removed and the soils were sieved (mesh size 2 mm). Portions of the soil samples were analyzed for soil pH, SOC content, particle size distribution, and Fe/Al contents. The remainder of each sample was air-dried and split into two portions-one to extract HEOC (see section 2.5) and the other packed into 4 cm \times 10 cm (diameter \times length) PVC columns equipped with glass fibers at the bottom to prevent soil loss during leaching. Each PVC column was filled with 151-g air-dried soils to ensure uniform soil core density (1.2 g cm⁻³). Disturbed soil cores rather than intact soil cores were used in this study to reduce soil heterogeneity. All soil cores were kept in a refrigerator for 2 weeks to equilibrate before leaching to eliminate postdisturbance effects.



2.3 The standard DOC solution

To isolate the effects of soil properties on leachate chemistry, we created a standard DOC solution for initial DOC input. In July 2015, newly fallen, undecomposed litter from *C. carlesii* was collected, air-dried, and mixed. The standard DOC solution was prepared by extracting the chemically homogeneous litters with ultrapure water (litter mass: ultrapure water volume = 1:100) for 24 h at room temperature and filtering the extract through 0.45-µm polyethersulfone membranes. The extraction time and dilution ratio were chosen in an attempt to extract all the soluble ingredients. The standard DOC solution was freshly made before each use. The concentration of DOC in the standard solution was 209.1 ± 8.6 mg C L⁻¹ (n = 12), which is comparable to DOC concentrations in Oi and Oe horizon extracts (Yano et al. 2005). Its pH was 5.6 ± 0.3, with the hydrophobic compound fraction constituting 80.5 ± 2.2%.

2.4 Soil core leaching

Two leaching methods were used in this study. With single leaching, every soil core was equally and individually supplied with 151-mL standard solution (Fig. 1, left). With sequential leaching, 906 mL (6×151 mL) of the standard DOC solution was first percolated through the 0–10-cm soils, followed by the leachate through the 10–20-cm layer, and so on until application to the last 80–100-cm soil depth (Fig. 1, right). The standard DOC solution was applied at an input rate of 2 mL/min. Portions of all leachates were collected, filtered through a 0.45-µm polyethersulfone membrane, and promptly analyzed.

2.5 Hot water-extractable organic carbon

The air-dried soils (equal to soils for PVC package, see section 2.2) were extracted for HEOC, which was considered as the most reactive part of native SOC. Forty-milliliter ultrapure water and 8-g soils were added to a 50-mL centrifuge tube, then shaken at 80 °C for 1 h. The suspension was centrifuged at 3500 rpm for 15 min and the supernatant was filtered through a 0.45- μ m polyethersulfone membrane for immediate analysis.

2.6 Analytical methods

SOC content was determined with a soil elemental analyzer (Vario MAX-1, Elementar, Germany). Soil pH was determined in 1:2.5 soil:water suspension (McLean 1982). Soil particle size distribution was determined by the hydrometer method (Zhang et al. 2000). The citrate-dithionite-bicarbonate method was used to determine the content of free Fe and Al oxides (Fe_d, Al_d) (Mehra and Jackson 1960). The contents of amorphous Fe and Al oxides (Fe_o, Al_o) were estimated by extraction with ammonium oxalate at pH 3.0 (McKeague 1978).

🖄 Springer





Fig. 1 Schematic representation of single leaching (left) and sequential leaching (right)

All DOC samples were analyzed within 8 h of preparation. The organic carbon concentrations of all DOC solutions (including the standard solutions, HEOC, and leachates) were determined on a TOC-VCPH/CPN (Shimadzu, Japan). The differences in total DOC contents between incoming solutions and leachates are regarded as total retention. The standard DOC solutions and leachates were fractionated into hydrophobic and hydrophilic organics using XAD-8 exchange resin at pH = 2. Organic carbon absorbed onto XAD-8 was defined as the hydrophobic fraction, not absorbed as hydrophilic fraction (Leenheer 1981). The differences in hydrophobic DOC contents between incoming solutions and leachates are regarded as hydrophobic retention.

Spectroscopic characterization was conducted for all DOC solutions. Fluorescence emission spectra were recorded with a Hitachi F-7000 (Japan, λ_{ex} 254 nm, slit 10 nm; λ_{em} 300–480 nm, slit 10 nm; scanning speed 1200 nm min⁻¹). The ratio of the peak area in the Σ (435–480)-nm range to the peak area in the Σ (300–345)-nm range was defined as the humification index (HIX) (Zsolnay et al. 1999). UV spectra were obtained on a UV-2450 (Shimadzu, Japan) in cuvettes with 1-cm edge length. Absorption at 254 nm was used to calculate the aromaticity index (AI) according to the equation AI = (UV₂₅₄ DOC concentration) × 100 (Zsolnay 2003).

2.7 Statistical analyses

For each soil type, basic soil properties (such as sand, silt, clay) or DOC traits (total retention, hydrophobic retention,

Table 1Selected characteristics of Ferralic Cambisols (FC) and Humic Alfisols (HA) soils (mean \pm SD, n = 3)

Soil depth (cm)	рН	Soil organic carbon (g kg ^{-1})	Sand (%) (0.25–0.05 mm)	Silt (%) (0.05–0.002 mm)	Clay (%) (< 0.002 mm)	$ \begin{array}{c} Fe_d - Fe_o \\ (g \ kg^{-1}) \end{array} $	$\begin{array}{c} Al_d \\ (g \ kg^{-1}) \end{array}$
FC soils							
0~10	$4.33\pm0.18aA$	$28.24\pm2.49aB$	$60.5\pm0.17aA$	$13.82\pm0.66bB$	$25.69\pm0.56aA$	$21.56\pm0.28aA$	$8.04\pm0.43aA$
10~20	$4.37\pm0.13aA$	$15.27\pm1.57 bA$	$60.2\pm1.62aA$	$12.20\pm1.96aB$	$27.60 \pm 1.55 aA$	$24.19 \pm 1.55 bA$	$10.11 \pm 1.64 bcA$
20~40	$4.53\pm0.11 aA$	$6.58\pm0.98cA$	$52.4\pm0.59 bA$	$15.58 \pm 1.27 \text{cB}$	$32.04 \pm 1.08 bA$	$27.82 \pm 1.01 \text{cA}$	$8.95 \pm 1.00 abA$
40~60	$4.46\pm0.18aA$	$3.14\pm0.84dA$	$51.2\pm1.82 bA$	$15.62\pm1.11\text{cB}$	$33.22\pm2.02 bcA$	$28.63 \pm 1.44 cdA$	$10.37\pm0.87 bcA$
60~80	$4.46\pm0.04aA$	$2.44\pm0.25~dB$	$50.4\pm0.41 bcA$	$14.78\pm0.53 bcB$	$34.80\pm0.84cA$	$30.34 \pm 1.39 dA \\$	$9.79 \pm 1.25 bcA$
80~100	$4.41\pm0.28aA$	$2.29\pm0.40 dA$	$48.9 \pm 1.94 \text{cA}$	$16.25\pm1.23\text{cB}$	$34.82\pm2.66cA$	$32.72\pm2.06eA$	$11.02\pm0.63cA$
HA soils							
0~10	$4.64\pm0.14aA$	$61.81\pm3.18aA$	$37.29\pm10.15aB$	$43.55\pm8.65aA$	$19.17 \pm 1.80 aB$	$8.59\pm0.65aB$	$1.44\pm0.14aB$
10~20	$4.73\pm0.11abB$	$16.59\pm4.18bA$	$34.97\pm10.19bB$	$43.97 \pm 7.83 abA$	$21.06 \pm 2.58 abB$	$10.09\pm0.67abB$	$1.48\pm0.14abB$
20~40	$4.71\pm0.14abA$	$6.50\pm1.31\text{cA}$	$33.55\pm9.83 bcB$	$43.55\pm 6.01 bA$	$22.90\pm3.83 bcB$	$11.32\pm0.57bB$	$1.60\pm0.05abB$
40~60	$4.88\pm0.17abB$	$4.21\pm0.68cA$	$33.34\pm9.72cB$	$42.81\pm5.82\text{cA}$	$23.85\pm3.94 bcB$	$13.96\pm0.83cB$	$1.72\pm0.08 bcB$
60~80	$4.89\pm0.17abB$	$3.83\pm0.43cA$	$32.62\pm9.74cB$	$41.31\pm3.75\text{dA}$	$26.07\pm 6.19 cdA$	$16.90\pm1.79~\mathrm{dB}$	$1.90\pm0.24cB$
80~100	$4.93\pm0.13bB$	$3.13\pm0.51 cA$	$30.27 \pm 11.34 \text{ dB}$	$42.31\pm6.02dA$	$27.42\pm5.39 dA$	$18.09\pm2.98~\mathrm{dB}$	$2.34\pm0.15~dB$

Lowercase letters following the data in the same column indicate significant difference across depths within one soil type; capital letters show significant difference between two soil types at the same depth. The significance level was set at 0.05

leachate HIX, and leachate AI) across all depths were compared with analysis of variance (ANOVA) in a randomized block design. Soil depth was treated as fixed effect and sampling point (i.e., block, three blocks in each soil type) was included as the random factor. The least significant difference (LSD) method was applied for means comparisons in all ANOVA. Soil properties for two different soil types at the same depth were compared with a t test for independent samples. The Pearson correlation coefficients (r value) were calculated for total retention, hydrophobic retention, leachate AI, and leachate HIX. Multiple linear regressions were performed to assess the relationship between individual traits of single leaching (dependent variables: total retention, hydrophobic retention, leachate AI, and leachate HIX) and multiple soil properties (independent variables: soil pH, SOC, sand, silt, clay, Fe_d, Al_d, Fe_o, Al_o, Fe_d-Fe_o, HEOC quantity, HEOC AI, HEOC HIX). The stepwise method was used to select significant independent variables.

All statistical analyses were performed using SPSS (SPSS 20 for Windows, SPSS Inc. Chicago, IL, USA). Relative proportion of DOC input, DOC retention, and SOC of each soil horizon during sequential leaching were calculated as individual value of each horizon divided by the sum of six (for 0–100-cm FC soil) or five (for 10–100-cm HA soil) horizons.

Data availability The datasets generated and/or analyzed during the current study are available in the figshare repository (Si 2018; https://figshare.com/s/3b4864e304ef0bed3fec; https://doi.org/10.6084/m9.figshare.6632942). The dataset was not peer-reviewed.

3 Results

3.1 Soil properties

Between the two soil types, four out of six layers differed significantly in pH (Table 1). FC soils were more acidic and had significantly higher Fe_d - Fe_o , Al_d , sand, and clay contents but significantly lower silt contents than HA. SOC content decreased with depth for both soil types, and HA soils had higher SOC content than FC soils (Table 1). HEOC of all FC soil layers, except for the 10–20-cm segments, had lower AI values than those of HA. HEOC of FC soils had higher HIX than those of HA soils (Figs. 1 and 2, blank columns).

3.2 DOC retention

3.2.1 Single leaching

With single leaching, total retention and hydrophobic retention increased with depth within each soil type (Fig. 4). Different from all the other soil segments, the 0–10-cm HA soils showed negative total retention and negative hydrophobic retention (Fig. 4, HA). FC soils retained more DOC than HA soils (total retention 160.8 vs. 78.3 mg C kg⁻¹ soil, hydrophobic retention 148.7 vs. 76.6 mg C kg⁻¹ soil). For both soil types, total retention was significantly positively related to hydrophobic retention, but significantly negatively related to leachate HIX (Table 2).



Deringer

Fig. 2 AI (special UV absorption) of leachates from soil cores (black columns for single leaching and shadow columns for sequential leaching) and AI of HEOC (blank columns) for Ferralic Cambisols (FC) and Humic Alfisols (HA) soils. Error bars represent standard deviation (n = 3). Lowercase letters over the same type of columns denote significant difference between horizons within each single Figure. Capital letters show significant difference between FC soils and HA soils, and only soil horizons of the same depth range are compared. The significance level was set at 0.05. Horizontal lines at AI = 1.79 indicate the AI of the standard DOC input



3.2.2 Sequential leaching

With sequential leaching, the quantity of DOC input to various horizons decreased with depth for each soil type (Fig. 5). Total retention and hydrophobic retention in FC soil decreased with depth (Fig. 4, FC). A sixfold higher amount of DOC input with sequential leaching made the 0–10-cm HA horizon release 17 times more DOC compared to single leaching, while retention over 10–100-cm HA soil layers was positive throughout but decreased along the soil profile (Fig. 4, HA). On average, FC soils retained more DOC than HA soils (total retention 168.2 vs. 153.2 mg C kg⁻¹ soil, hydrophobic retention 142.8 vs. 133.7 mg C kg⁻¹ soil).

🖄 Springer



3.3 UV absorption

Movement through soils reduced AI of DOC solutions (Fig. 2). In single leaching, leachates from deeper soil layers had lower AI than those from upper soil layers, and AI of leachates from 0 to 40-cm HA soils were significantly higher than those from 0 to 40-cm FC soils (Fig. 2, FC). In sequential leaching, by the time the percolating solutions reached the 60-cm depth, their AI had dropped significantly compared with that of the standard DOC, whereas the AI of the solution percolating through the 60–100-cm layers only slightly decreased. Notably, leachates from the 0–10-cm HA soil layers had the highest AI (1.6) but declined sharply (0.8) after they passed through 10–20-cm soils (Fig. 2, HA).

Table 2 The Pearson correlation coefficients between total retention, hydrophobic retention, leachate AI, and leachate HIX. Only data from single leaching were used. For Ferralic Cambisols (FC) soils, data from all six soil layers were included and n = 18; for Humic Alfisols (HA) soils, only data from five layers within 10–100 cm were included and n = 15; the top layer was excluded due to the net release of DOC

	Total retention	Hydrophobic retention	Leachate AI	Leachate HIX
FC soils				
Total retention	1			
Hydrophobic retention	0.961**	1		
Leachate AI	-0.401	-0.385	1	
Leachate HIX	-0.774 **	-0.724 **	0.693**	1
HA soils				
Total retention	1			
Hydrophobic retention	0.984**	1		
Leachate AI	-0.967 **	-0.969 **	1	
Leachate HIX	-0.901**	-0.896**	0.898**	1

***p* < 0.01

3.4 Fluorescence spectroscopic analysis

As shown in Fig. 3, leachates always had lower HIX than incoming solutions. For each soil type in single leaching (Fig. 3 FC), the HIX of leachates of the four layers at the 0–60-cm depth differed significantly, whereas that of the leachates out of the three layers at the 40–100-cm depth showed no significant difference. Leachates from the 40–100-cm HA soil layers had significantly higher HIX than those from the 40–100-cm FC soil layers.

For each soil type in sequential leaching (Fig. 3, HA), HIX of leachates from six layers significantly differed from each other. Leachates from HA soils had slightly lower HIX than those from FC soils.

3.5 Multiple linear regression

To isolate the effect of soil conditions on leaching dynamics, only data from single leaching was used to carry out the regression analysis. Data about 0–10-cm HA soils was excluded due to net release. DOC movement through FC soils was predominantly influenced by SOC content, which was the only significant variable predicting total and hydrophobic retention. DOC movement through 10–100-cm HA soils was mainly affected by HEOC AI or HEOC HIX (see relative contribution, Table 3).

4 Discussion

4.1 Single leaching

By single leaching, we demonstrated that within a soil type, deeper soils had higher retentive capacity than upper soils.

Lower SOC concentrations of deep versus surface soils may mean deep soils are further from carbon saturation than surface soils. Hydrophobic fraction accounted for the majority of total retention (Fig. 4) and they were significantly positively correlated (Table 2), consistent with the findings of Kaiser and Zech (1997). Aromatic substances, especially polycyclic aromatic compounds, are highly conjugated and have active π electrons that readily react with Fe/Al oxides and hydroxides (Weishaar et al. 2003). These substances are highly reactive and are preferentially retained at the soil surface. The lower AI of leachates out of deeper soils thus means more retention. HIX can indicate the percentage of microbe-processed organic carbon in solutions (Zsolnay et al. 1999). Lower HIX of leachates out of 40-100-cm soils meant these soils released less native SOC and retained more incoming DOC than 0-40-cm soils (significantly negative correlation between leachate HIX and total retention, Table 2).

SOC of FC soils greatly masked the effect of soil texture and Fe/Al mineralogy, because SOC is the only variable incorporated in the model to predict total retention or hydrophobic retention by FC soils, as seen in Table 3. FC soils were collected from a natural C. carlesii forest, whose SOC is structurally similar to our standard DOC and has been accumulated for hundreds of years, which makes sense as this litter source has been contributing to SOC formation for hundreds of years, perhaps explaining SOC effects on retention patterns (Table 3). Lack of SOC might result in similar retention, as the 60-80-cm and 80-100-cm FC soil layers showed similar and overall strong sorptive capacities with minimal SOC coverage, despite significant difference in their Fe_d-Fe_o contents, consistent with Scott and Rothstein (2014). Dynamic exchange between incoming DOC and outer sphere SOC could be seen from the FC leachate AI, which was influenced by HEOC AI (Table 3). This implies that outer sphere SOC was displaced into leachates. Moreover, leachate AI was generally lower than HEOC AI (Fig. 2, FC), suggesting that higher aromatic substances were more competitive for soil surfaces. The amount of displaced SOC could be estimated by leachate HIX, which was positively controlled by SOC content (Table 3), suggesting that larger SOC content was associated with larger releases of SOC.

HA soils were collected from a natural *C. fargesii* stand, whose SOC nature should be different from that of the *C. carlesii* litter used to produce the standard DOC. As expected, we saw greater interaction between the standard DOC and outer sphere HA SOC, with stronger desorption and consequent less retention compared to FC soils. The strongest desorption due to the highest SOC content led to net DOC release from the top 10-cm HA soils. This provided unambiguous evidence for dynamic exchange and competition between incoming DOC and native SOC for soil surfaces. Displacement of native



Fig. 3 Humification index (HIX) of leachates from soil cores (black columns for single leaching and shadow columns for sequential leaching) and HIX of hot waterextractable soil organic carbon (HEOC) (blank columns) for Ferralic Cambisols (FC) and Humic Alfisols (HA) soils. Error bars represent standard deviation (n = 3). Lowercase letters over the same type of columns denote significant difference between horizons within each single figure. Capital letters show significant difference between FC soils and HA soils, and only soil horizons of the same depth range are compared. The significance level was set at 0.05. Horizontal lines at HIX = 1.50 indicate the HIX of the standard DOC input



SOC from other HA soil layers could be deduced from (i) leachate AI from the top 20-cm HA soils which was higher than leachate AI of the top 20-cm FC soils, despite the two kinds of soils having similar HEOC AI, and (ii) although HEOC HIX of 40–100-cm HA soils was less than or comparable to that of 40–100-cm FC soils, leachates out of the former soils showed significantly higher HIX than leachates out of the latter soils, indicating the former released much more microbe-processed SOC. For HA, outer sphere SOC therefore regulated DOC movement to a greater extent than in FC soils. This is in line with the fact that HEOC HIX and/or HEOC AI were included in the regression models predicting total retention, hydrophobic retention, and leachate HIX by 10–100-cm HA soils (Table 3).

 $\underline{\textcircled{O}}$ Springer



4.2 Sequential leaching

In contrast to single leaching, deeper soils retained less DOC than upper soils with sequential leaching (except for the 0–10cm HA soil horizon). As we have already demonstrated that deeper soils had stronger retentive capacities than upper soils, the retentive pattern for sequential leaching was clearly attributed to the changes in quantity and sorptive quality of incoming DOC.

Although decreasing with depth, DOC input to 0–80-cm FC and 0–100-cm HA soils for sequential leaching was still more than DOC input to these soils for single leaching (Fig. 5). Nevertheless, the 10–80-cm FC and the 60–100-cm HA soil layers retained less DOC with sequential leaching versus single leaching (Fig. 4). This suggests that reduced

 Table 3
 Multiple linear regression model to assess the relationship between individual leaching traits (dependent variables, including total retention, hydrophobic retention, leachate AI, and leachate HIX) and soil properties (independent variables, including soil pH, SOC, sand, silt, clay,
 Fe_d, Al_d, Fe_o, Al_o, Fe_d-Fe_o, HEOC quantity, HEOC AI, HEOC HIX). The stepwise method was used to select significant independent variables. VIF (variation inflation factor) of selected variables were all below 7.2

	Adjusted R^2 (%)	Relative contribution of independant variables
FC soils (of 0–100-cm depth range)		
Total retention = $-1.573 \times SOC + 176.031$	62.1	SOC:100%
Hydrophobic retention = $-1.104 \times SOC + 159.390$	49.3	SOC:100%
Leachate AI = $0.492 \times \text{HEOC AI} - 0.003 \times \text{sand} - 0.032 \text{ Al}_{d} + 1.977$	85.8	HEOC AI: 56.9%, sand: 26.6%, Ald: 16.5%
Leachate HIX = $0.018 \times \text{SOC} + 0.004 \times \text{sand} - 1.783$	96.8	SOC: 49.1%, sand: 50.9%
HA soils (of 10–100-cm depth range)		
Total retention = $-29.447 \times \text{HEOC AI} + 2.579 \times \text{Fe}_{d} - \text{Fe}_{o} + 90.371$	81.3	HEOC AI: 49.2%, Fe _d -Fe _o : 50.8%
$\begin{array}{l} Hydrophobic \ retention = - \ 14.492 \times HEOC \\ HIX + 1.488 \times Fe_d\text{-}Fe_o + 0.123 \times silt - 22.812 \times HEOC \ AI + 50.507 \end{array}$	90.2	HEOC HIX: 28.7%, Fe _d -Fe _o : 33.6%, silt: 9.0%, HEOC AI: 28.7%
Leachate AI = $0.015 \times \text{SOC} - 0.021 \times \text{Fe}_d + 0.735$	85.7	SOC: 54.0%, Fed: 46.0%
Leachate HIX = $0.187 \times \text{HEOC}$ HIX + $0.218 \times \text{HEOC}$ AI + 0.331	81.6	HEOC HIX: 54.8%, HEOC AI: 45.2%

quantities of DOC input do not necessarily result in less retention. Declining affinity of DOC input along soil depth was thus the key factor explaining the retentive pattern of sequential leaching.

Reduction in the affinity of incoming DOC to deeper soils for soil surfaces was attributed to (i) the quantity and percentage of hydrophobic compounds of input solutions decreased with depth (Guggenberger and Kaiser 2003; Kaiser and Kalbitz 2012). (ii) DOC input to deeper soils had lower AI (Fig. 2, shadow columns). By the time the percolating solution reached the 60-cm depth, AI had declined drastically compared to the standard DOC (Fig. 2). Subsequently, the 60-100-cm soils only slightly decreased the AI of the percolating solution, consistent with their small retention. (iii) Relative more weaker-binding compounds were input to deeper soils. Leachates from sequential leaching generally had higher HIX than those from single leaching. This suggested that leachates out of upper soils included relative more native fraction, which showed weaker affinity for soil surfaces and accumulated in the leachate solutions.

Despite six times greater DOC input than that of single leaching, the total retention by top 10-cm FC soils increased only by a factor of 4.2. This might be due to the already high SOC concentration and the chemical similarity between the standard DOC and native FC SOC. This implies that retention amount should not be considered linear with DOC input as assumed by some models (Tipping et al. 2012). However, sequential leaching widened the gap in SOC between the two uppermost FC horizons. The relative DOC retention by 0–10-cm FC soil layer was slightly higher than its relative SOC content (higher blank columns vs. shorter gray columns for this soil horizon in Fig. 5, FC). In contrast, the 10–20-cm horizon retained much less DOC compared to its relative SOC content. Under field conditions, rapid

degradation of SOC in the top 10-cm soils and migration of soil solution with rainfall, as well as carbon input from fine roots, would further transport organic carbon to 10–20-cm soils. The 20–100-cm FC soils also retained higher percentages of DOC than their native SOC content (Fig. 5, FC), probably because under field conditions these soils had little chance to receive so much DOC input as in our experiment and therefore sequestrated lower C.

The sixfold greater DOC input to the 0-10-cm HA soil layers increased the DOC release by a factor of 17 (Fig. 4). Obviously, this DOC release was caused not only due to scouring by the incoming DOC, but more importantly by desorption of native SOC. If scouring alone, six times of input would have produced at most a sixfold increase in the DOC release. Moreover, the AI of the leachates (1.6) from this soil horizon was highest of all leachates, exceeding HEOC AI and leachate AI of single leaching (Fig. 2). This implies that desorption-induced release of native SOC in sequential leaching was even more intensive than hot water extraction, and more aromatics were released. This caused much DOC input to the 10-40-cm HA horizons, and consequently, these soils retained the most DOC, with the leachate AI dropping sharply from 1.6 to 0.5 (Fig. 2, HA). Especially, the relative DOC retention by the 20-40-cm HA horizon was much greater than the relative SOC content in this layer (Fig. 5, HA). Due to the strong retention, the 40-100-cm soil layers retained lower percentages of DOC relative to SOC content.

4.3 Single leaching vs. sequential leaching

Despite the sorptive capacity of deeper soils was limited, the organic carbon sequestration was more in sequential leaching than in single leaching for each soil type, because (i) sequential leaching retained more hydrophilic fraction; (ii) leachates



Fig. 4 Amount of HEOC and DOC retained by Ferralic Cambisols (FC) and Humic Alfisols (HA) soils. Error bars represent standard deviation (n =3). Lowercase letters over the same type of columns denote significant difference between horizons within each single figure. Capital letters show significant difference between FC and HA, and only soil horizons of the same depth range are compared. The significance level was set at 0.05. Single leaching means soil core was equally and individually supplied with 151 mL (averagely 31.6 mg C) standard solution. Sequential leaching refers to that 906 mL (averagely 189. 5 mg C) standard DOC solution percolated through the 0-10-cm soils, followed by the leachate passing through the 10-20-cm layer, and so on until it penetrated the soil at 80-100-cm depth



from the four 20–100-cm horizons in sequential leaching had lower AI than those from single leaching (Fig. 2, black columns vs. shadow columns) and therefore retained more weaker-binding fraction than single leaching; and (iii) the HIX of the percolating solution steadily decreased during sequential leaching, so some of desorbed native SOC from the upper soil layers was retained by deeper soil layers. This desorption-induced transfer of organic carbon from upper soils to deeper soils shows that with time weaker-binding organic carbon will be associated with deeper soils, which is a natural way to preserve SOC, in agreement with the cycling downwards model (Kaiser and Kalbitz 2012).

4.4 Ecological significance

Our results do not reflect actual DOC dynamics of in situ soils for obvious reasons—we relied on modified soils and we applied DOC in a way that does not reflect actual hydrological processes. The sorptive capacity of soil surfaces for DOC shown in laboratory studies may not control the transport of DOC in soils if macropore fluxes dominate under field conditions. Soil macropores can facilitate DOC solution flow, either downward or lateral, and often serve as hotspots with high microbial activity, rapid organic matter mineralization and more soluble microbial products (Kuzyakov and

 $\underline{\textcircled{O}}$ Springer



Fig. 5 Proportion of DOC input, retention, and SOC of each soil horizon during sequential leaching for Ferralic Cambisols (FC) and Humic Alfisols (HA) soils. Error bars represent standard deviation (n = 3). Horizontal lines indicate DOC input during single leaching. Proportions for FC soils were calculated as individual data of each horizon divided by sum of all six horizons; proportions for HA soils were calculated as individual data of each horizon divided by sum of five horizons between 10 and 100 cm. Lowercase letters over the same type of columns denote significant difference between horizons within one soil type



Blagodatskaya 2015). However, this study focused on the sorptive capacity of soil surfaces and reduced the influence of macropore by packing disturbed soils to PVC columns.

Traditional aboveground litter decomposition studies considered only the remaining recalcitrant particulate litter residue as contributing stable SOC and underestimated DOC fluxes to the mineral soil (Berg and Mcclaugherty 2008). Cotrufo et al. (2015) identified the importance of DOC leaching from litter followed by high microbial incorporation, i.e., the DOCmicrobial pathway, for the formation of mineral-associated SOC at the early stage of litter decomposition. While they found no new SOC formation from recently decomposed litter in soils below 20-cm soil depth, our results highlighted that aboveground DOC input can actually alter SOC processes and composition to the 100-cm depth. This is particularly important for forest conversion. DOC from *C. carlesii* litter would displace much native *C. fargesii* SOC of 0–10-cm depth range and move them downwards, which greatly alter the SOC nature of 10–40-cm soils and subsequent microbial community structure and activity. This desorption-induced transfer could explain why the oldest organic matter being located in the youngest part of the soil profile (the deep mineral subsoil) and why subsoil DOC is of microbial origin. Another possible fate of the displaced SOC from upper soil layers is the potential to be leached into aquatic environments, which is of course not good to SOC accumulation. Such desorptioninduced SOC loss might explain rapid SOC loss during early stages of post-harvest stand growth (Chen et al. 2004; Chen et



🖄 Springer

al. 2010). While we focused on the role of DOC and SOC nature in controlling DOC adsorption/desorption in this study, other environmental variables may be equally important. A warmer atmosphere may lead to higher microbial activity and more microbial products and might promote SOC desorption and DOC migration to deeper soils (Allison et al. 2010). Nitrogen deposition could also affect DOC movement by altering litter decomposition rates and DOC input rates to soils (Knorr et al. 2005).

Although soil DOC dynamics has been incorporated into some mechanistic SOC models (Jason and Gregory 2001; Sanderman et al. 2008; Tipping et al. 2012), our results can still provide useful information to optimize such models. First, through single leaching, this study revealed that deeper soil horizons have stronger sorptive capacities than upper horizons. However, models, such as DyDOC (Tipping et al. 2012), do not include a sorptive capacity factor for different soil horizons. Secondly, a sixfold higher amount of DOC input to the 0-10-cm soils (both FC and HA) with sequential leaching did not result in six times the retention or release compared with single leaching, but Tipping et al. (2012) assumed there was a linear relationship between retention and DOC input. Thirdly, Sanderman et al. (2008) proposed a "potentially exchangeable or reactive soil carbon pool" responsible for the composition of soil DOC. Beyond this concept, our results further demonstrate that the size of this pool is highly dependent on the nature of the incoming DOC.

5 Conclusion

This study highlighted the importance of the native SOC and incoming DOC in regulating DOC leaching dynamics. By single leaching, we demonstrated deeper soil layers had stronger sorptive capacities than upper soil layers, and SOC or the HEOC was the most important factors regulating DOC movement through soils. With sequential leaching, however, changing DOC quantity and chemistry with declining affinity for soil surfaces increasingly restricted carbon sequestration in deeper soils. Nevertheless, desorption-induced transfer of organic carbon from upper to deeper soils ensured that sequential leaching always retained more organic carbon than single leaching, and DOC vertical movement greatly altered SOC formation and loss along the top soil profile. Clearly, our results demonstrate a need for soil biogeochemistry and hydrology models to more accurately address DOC and SOC dynamics and interactions.

Acknowledgments We thank Dr. Minhuang Wang and Maokui Lv for their assistance with multivariate analyses. We are also very grateful to Dr. Christian P. Giardina, from Institute of Pacific Islands Forestry of the USA, for helping to edit our writing.

🖄 Springer



Funding This project was funded by National Natural Science Foundation of China (Nos. 31570606, U1505233, and 31100467), and Natural Science Foundation of Fujian Province (No. 2015J01120).

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Allison SD, Wallenstein MD, Bradford MA (2010) Soil-carbon response to warming dependent on microbial physiology. Nat Geosci 3:336– 340
- Basile-Doelsch I, Amundson R, Stone WEE, Borschneck D, Bottero JY, Moustier S, Masin F, Colin F (2007) Mineral control of carbon pools in a volcanic soil horizon. Geoderma 137:477–489
- Berg B, Mcclaugherty C (2008) Plant litter: decomposition, humus formation, carbon sequestration. Springer
- Bu X, Wang L, Ma W, Yu X, McDowell WH, Ruan H (2010) Spectroscopic characterization of hot-water extractable organic matter from soils under four different vegetation types along an elevation gradient in the Wuyi Mountains. Geoderma 159:139–146
- Bu XL, Ding JM, Wang LM, Yu XN, Huang W, Ruan HH (2011) Biodegradation and chemical characteristics of hot-water extractable organic matter from soils under four different vegetation types in the Wuyi Mountains, southeastern China. Eur J Soil Biol 47:102–107
- Chen CR, Xu ZH, Mathers NJ (2004) Soil carbon pools in adjacent natural and plantation forests of subtropical Australia. Soil Sci Soc Am J 68:282–291
- Chen FS, Zeng DH, Fahey TJ, Liao PF (2010) Organic carbon in soil physical fractions under different-aged plantations of Mongolian pine in semi-arid region of Northeast China. Appl Soil Ecol 44: 42–48
- Choe E, Meer F, Rossiter D, Salm C, Kim KW (2010) An alternate method for Fourier transform infrared (FTIR) spectroscopic determination of soil nitrate using derivative analysis and sample treatments. Water Air Soil Pollut 206:129–137
- Colombo C, Palumbo G, Di Iorio E, Sellitto VM, Comolli R, Stellacci AM, Castrignanò A (2014) Soil organic carbon variation in Alpine landscape (Northern Italy) as evaluated by diffuse reflectance spectroscopy. Soil Sci Soc Am J 78:794–804
- Corvasce M, Zsolnay A, D'Orazio V, Lopez R, Miano TM (2006) Characterization of water extractable organic matter in a deep soil profile. Chemosphere 62:1583–1590
- Cotrufo MF, Soong JL, Horton AJ, Campbell EE, Haddix Michelle L, Wall DH, Parton WJ (2015) Formation of soil organic matter via biochemical and physical pathways of litter mass loss. Nat Geosci 8: 776–781
- Dai J, Ran W, Xing B, Gu M, Wang L (2006) Characterization of fulvic acid fractions obtained by sequential extractions with pH buffers, water, and ethanol from paddy soils. Geoderma 135:284–295
- Dusek J, Vogel T, Dohnal M, Barth JAC, Sanda M, Marx A, Jankovec J (2017) Dynamics of dissolved organic carbon in hillslope discharge: modeling and challenges. J Hydrol 546:309–325
- Foresti E, Gazzano M, Gualtieri AF, Lesci IG, Lunelli B, Pecchini G, Renna E, Roveri N (2003) Determination of low levels of free fibres of chrysotile in contaminated soils by X-ray diffraction and FTIR spectroscopy. Anal Bioanal Chem 376:653–658
- Gu B, Schmitt J, Chen Z, Liang L, McCarthy JF (1994) Adsorption and desorption of natural organic matter on iron oxide: mechanisms and models. Environ Sci Technol 28:38–46

- Guggenberger G, Kaiser K (2003) Dissolved organic matter in soil: challenging the paradigm of sorptive preservation. Geoderma 113:293–310
- Jagadamma S, Mayes MA, Zinn YL, Gisladottir G, Russell AE (2014) Sorption of organic carbon compounds to the fine fraction of surface and subsurface soils. Geoderma 213:79–86
- Jason CN, Gregory PA (2001) Dissolved organic carbon in terrestrial ecosystems: synthesis and a model. Ecosystems 4:29–48
- Kaiser K, Guggenberger G (2000) The role of DOM sorption to mineral surfaces in the preservation of organic matter in soils. Org Geochem 31:711–725
- Kaiser K, Kalbitz K (2012) Cycling downwards dissolved organic matter in soils. Soil Biol Biochem 52:29–32
- Kaiser K, Zech W (1997) Competitive sorption of dissolved organic matter fractions to soils and related mineral phases. Soil Sci Soc Am J 61:64–69
- Kalbitz K, Kaiser K (2008) Contribution of dissolved organic matter to carbon storage in forest mineral soils. J Plant Nutr Soil Sci 171:52–60
- Kalbitz K, Solinger S, Park J-H, Michalzik B, Matzner E (2000) Controls on the dynamics of dissolved organic matter in soils: a review. Soil Sci 165:277–304
- Kalbitz K, Schmerwitz J, Schwesig D, Matzner E (2003) Biodegradation of soil-derived dissolved organic matter as related to its properties. Geoderma 113:273–291
- Kim M, Hyun S (2014) Effect of surface coordination on 2,4-D sorption by kaolinite from methanol/water mixture. Chemosphere 103:329– 335
- Kleber M, Nico PS, Plante A, Filley T, Kramer M, Swanston C, Sollins P (2011) Old and stable soil organic matter is not necessarily chemically recalcitrant: implications for modeling concepts and temperature sensitivity. Glob Chang Biol 17:1097–1107
- Kleber M, Eusterhues K, Keiluweit M, Mikutta C, Mikutta R, Nico PS (2015) Chapter one - mineral–organic associations: formation, properties, and relevance in soil environments. In: Donald LS (ed) Advances in agronomy, volume 130. Academic Press, pp 1–140
- Knorr M, Frey SD, Curtis PS (2005) Nitrogen additions and litter decomposition: a meta-analysis. Ecology 86:3252–3257
- Kothawala DN, Moore TR, Hendershot WH (2009) Soil properties controlling the adsorption of dissolved organic carbon to mineral soils. Soil Sci Soc Am J 73:1831–1842
- Kuzyakov Y, Blagodatskaya E (2015) Microbial hotspots and hot moments in soil: concept & review. Soil Biol Biochem 83:184–199
- Leenheer JA (1981) Comprehensive approach to preparative isolation and fractionation of dissolved organic carbon from natural waters and wastewaters. Environ Sci Technol 15:578–587
- Lilienfein J, Qualls RG, Uselman SM, Bridgham SD (2004) Adsorption of dissolved organic carbon and nitrogen in soils of a weathering chronosequence. Soil Sci Soc Am J 68:292–305
- McKeague JA (1978) Manual on soil sampling and methods of analysis. Canadian Society of Soil Science, Canada
- McKnight DM, Bencala KE, Zellweger GW, Aiken GR, Feder GL, Thorn KA (1992) Sorption of dissolved organic carbon by hydrous aluminum and iron oxides occurring at the confluence of Deer Creek with the Snake River, Summit County, Colorado. Environ Sci Technol 26:1388–1396

- McLean EO (1982) Soil pH and lime requirement. In: Page AL, Miller RH, Keeney DR (eds) Methods of soil analysis: part 2. Chemical and Microbiological Methods. American Society of Agronomy, Soil Sci Soc Am, Madison, WI, pp 199–224
- Mehra OP, Jackson ML (1960) Iron oxide removal from soils and clays by dithionite-cetrate systems buffered with sodium bicarbonate. Clay Clay Miner 7:317–327
- Michel K, Matzner E, Dignac M-F, Kögel-Knabner I (2006) Properties of dissolved organic matter related to soil organic matter quality and nitrogen additions in Norway spruce forest floors. Geoderma 130: 250–264
- Ohno T (2002) Fluorescence inner-filtering correction for determining the humification index of dissolved organic matter. Environ Sci Technol 36:742–746
- Sanderman J, Baldock JA, Amundson R (2008) Dissolved organic carbon chemistry and dynamics in contrasting forest and grassland soils. Biogeochemistry 89:181–198
- Schmidt MWI, Torn MS, Abiven S, Dittmar T, Guggenberger G, Janssens IA, Kleber M, Kögel-Knabner I, Lehmann J, Manning DAC, Nannipieri P, Rasse DP, Weiner S, Trumbore SE (2011) Persistence of soil organic matter as an ecosystem property. Nature 478:49–56
- Scott EE, Rothstein DE (2014) The dynamic exchange of dissolved organic matter percolating through six diverse soils. Soil Biol Biochem 69:83–92
- Si, Y (2018) The vertical movement of dissolved organic carbon through two distinct soil types under Castanopsis fargesii Franch. and C. carlesii (Hemsl.) Hayata forests. V6. figshare. [Dataset]. https:// doi.org/10.6084/m9.figshare.6632942 [Accessed 07 July 2018]
- Tipping E, Chamberlain PM, Fröberg M, Hanson PJ, Jardine PM (2012) Simulation of carbon cycling, including dissolved organic carbon transport, in forest soil locally enriched with ¹⁴C. Biogeochemistry 108:91–107
- Toosi ER, Doane TA, Horwath WR (2012) Abiotic solubilization of soil organic matter, a less-seen aspect of dissolved organic matter production. Soil Biol Biochem 50:12–21
- Weishaar JL, Aiken GR, Bergamaschi BA, Fram MS, Fujii R, Mopper K (2003) Evaluation of specific ultraviolet absorbance as an indicator of the chemical composition and reactivity of dissolved organic carbon. Environ Sci Technol 37:4702–4708
- Yano Y, Lajtha K, Sollins P, Caldwell BA (2005) Chemistry and dynamics of dissolved organic matter in a temperate coniferous forest on andic soils: effects of litter quality. Ecosystems 8:286–300
- Yurova A, Sirin A, Buffam I, Bishop K, Laudon H (2008) Modeling the dissolved organic carbon output from a boreal mire using the convection-dispersion equation: importance of representing sorption. Water Resour Res 44:W07411
- Zhang W, Yang G, Tu X (2000) Forest soil analytical methods. Standards Press of China, China
- Zsolnay Á (2003) Dissolved organic matter: artefacts, definitions, and functions. Geoderma 113:187–209
- Zsolnay A, Baigar E, Jimenez M, Steinweg B, Saccomandi F (1999)
 Differentiating with fluorescence spectroscopy the sources of dissolved organic matter in soils subjected to drying. Chemosphere 38: 45–50

