



Cell Cycle

ISSN: 1538-4101 (Print) 1551-4005 (Online) Journal homepage: https://www.tandfonline.com/loi/kccy20

A Novel Role for Hedgehog in T-Cell Receptor Signaling: Implications for Development and Immunity

Nicola J. Rowbotham, Ariadne L. Hager-Theodorides, Anna L. Furmanski & **Tessa Crompton**

To cite this article: Nicola J. Rowbotham, Ariadne L. Hager-Theodorides, Anna L. Furmanski & Tessa Crompton (2007) A Novel Role for Hedgehog in T-Cell Receptor Signaling: Implications for Development and Immunity, Cell Cycle, 6:17, 2138-2142, DOI: 10.4161/cc.6.17.4644

To link to this article: https://doi.org/10.4161/cc.6.17.4644

\mathbf{O}

Copyright © 2007 Landes Bioscience



Published online: 07 Sep 2007.

-	
	1.
t.	v 1
_	

Submit your article to this journal 🗹

Article views: 292



View related articles 🗹



Citing articles: 16 View citing articles 🖸

Perspective

A Novel Role for Hedgehog in T-Cell Receptor Signaling

Implications for Development and Immunity

Nicola J. Rowbotham^{1,2} Ariadne L. Hager-Theodorides¹ Anna L. Furmanski¹ Tessa Crompton^{1,2}

¹Immunobiology Unit; UCL Institute of Child Health; London, UK

²Division of Cell and Molecular Biology; Imperial College London; London, UK

*Correspondence to: Tessa Crompton; Immunobiology Unit; UCL Institute of Child Health; 30 Guilford Street; London WC1N 1EH UK; Tel.: +44.0.207.905.2893; Fax: +44.0.207.813.8494; Email: t.crompton@ich.ucl.ac.uk

Original manuscript submitted: 06/27/07 Manuscript accepted: 06/27/07

Previously published online as a *Cell Cycle* E-publication: http://www.landesbioscience.com/journals/cc/article/4644

KEY WORDS

hedgehog signalling, Gli2, Shh, T cell, thymocyte development, ERK phosphorylation, TCR, T cell activation, IL-2, proliferation

ACKNOWLEDGEMENTS

This work was supported by the MRC, Leukaemia Research Fund, BBSRC and the Wellcome Trust.

ABSTRACT

The Hedgehog (Hh) signaling pathway is a key regulator of both embryonic development and homeostasis of adult tissues, including thymus and blood. In the thymus, Hh signals for differentiation, survival and proliferation in the early stages of T cell development, before TCR gene rearrangement. Our recent data has shown that Hh signaling also modulates T cell receptor (TCR) signal strength in more mature T lineage cells. We showed that constitutive activation of the Hh pathway in thymocytes (by transgenic expression of the transcriptional activator form of Gli2) decreased TCR signal strength with profound consequences for the thymus-allowing self-reactive T cells to escape deletion and altering T cell CD4/CD8 lineage decisions. In contrast, in the Sonic Hh deficient thymus, TCR signaling was increased, again influencing both TCR repertoire selection and CD4/8 lineage commitment. In peripheral T cells, the transcriptional changes induced by activation of the Hh signaling pathway lead to reduced T cell activation. Hh signaling also attenuated ERK phosphorylation and proliferation in mature T cells on TCR ligation. Modulation of TCR signal strength by Hh pathway activation has importance for immunity as the presence or absence of Hh in the environment in which a T cell is activated would shape the immune response.

INTRODUCTION

The hedgehog (Hh) family of secreted intercellular signaling molecules (Sonic, Indian and Desert Hh) modulate target gene expression via the Gli transcription factors. Hh proteins are essential morphogens during embryogenesis and the homeostasis of adult tissues,¹⁻⁴ and can influence differentiation, cell cycle progression and survival.^{1,5} In a classical model, a morphogen specifies cell fate by establishment of a morphogen gradient, in which its movement from a polarised source provides positional information for patterning and differentiation in a solid tissue. In addition to its competence to transduce the signal, the response of the receiving cell depends on the concentration and duration of the signal received and therefore on its position in the tissue. In contrast to organogenesis of solid tissues, the immune system is 'fluid', made up of multiple cell types that move through the body and interact, differentiate and respond in many different environments. Despite this apparent difference, Hh signaling has also turned out to be an important regulator of the immune system.^{2,6-12}

The Gli transcription factors (Gli1, Gli2 and Gli3) are key to the interpretation of the Hh gradient by the receiving cell.^{1,5} They have distinct temporal and tissue-specific expression patterns and functions. Gli2 is necessary to initiate the first transcriptional changes induced by Hh signaling.¹³ Gli1 is solely an activator of transcription, but Gli2 and Gli3 can be processed to activate or repress target-gene transcription, according to the presence or absence of Hh respectively.¹³⁻¹⁶ Gli functions are affected by their cellular context, and are dependent both on cell type and the Hh environment of that cell.^{17,18} The interpretation of the Hh signal in a given cell relies on the balance of activator and repressor forms of Gli proteins produced.

HH SIGNALING IN THE THYMUS

The specialised environment of the thymus supports the maturation of haematopoietic progenitors into functional T lymphocytes and this involves bi-directional signaling between the thymic epithelium and developing thymocytes. During T cell development, thymocytes pass through stages that can be defined by the expression of cell surface markers: CD4⁻CD8⁻ double negative (DN) cells differentiate into CD4⁺CD8⁺ double

positive (DP) thymocytes, which mature to become CD4 single positive (SP) and CD8 SP thymocytes. Signaling by a functional pre-T cell receptor (pre-TCR) is necessary for differentiation from DN to DP cell.¹⁹ Maturation from DP to SP cell requires the expression of a functional, MHC-restricted $\alpha\beta$ TCR and involves TCR repertoire selection.^{20,21} Positive selection of thymocytes that express TCR with appropriate affinity for self-MHC molecules ensures functional self-restriction, while negative selection of thymocytes that express TCR with high affinity for self removes overtly self-reactive T cell clones; therefore cell fate is at least in part determined by the strength of the signal transduced by the TCR.²¹ Mature T cells then exit the thymus to populate peripheral lymphoid organs where they can be activated on TCR binding to peptide antigen presented by major histocompatibility complex (MHC) molecules. CD4 T cells recognise antigen as peptide in the context of self MHC Class II molecules, whereas CD8 T cells recognise antigen as peptide in the context of self MHC Class I molecules.

Hh signaling is important in T cell development, in both humans and mice.^{2,6,7,9-12} Shh is expressed by thymic epithelial cells and components of the Hh signaling pathway are expressed in the lymphoid and stromal compartments of the thymus.^{2,6,9,11,12} Hh signaling is involved in the homeostasis of DN cells, differentiation from DN1 to $DN2^{6,7,10,11}$ and the DN to DP transition.^{2,7,10,11}

We have recently investigated the function of Hh signaling at later stages of murine thymocyte development. We have shown that Hh signaling is involved in the maturation of immature DP cells to mature SP thymocytes and that it can affect TCR repertoire selection and CD4/8 lineage choice.¹²

HH SIGNALING ALTERS TCR REPERTOIRE SELECTION AND CD4:CD8 RATIO IN DEVELOPING THYMOCYTES

We produced transgenic mice with a transcriptional activator form of the Gli2 protein $(\text{Gli2}\Delta N_2)^{14}$ under the control of the T-lineage-restricted Lck promoter.^{22,23} This Lck-Gli2 ΔN_2 transgene induces the transcriptional events normally caused by active Hh signaling, allowing us to assess the effect of constitutive activation of the Hh signaling pathway specifically in T cells.

Expression of the Lck-Gli2 ΔN_2 transgene decreased the production of CD4 SP cells in the thymus, and reduced the CD4:CD8 ratio. Treatment of wild type (WT) thymus explants with recombinant Shh protein (r-Shh) also decreased the CD4:CD8 ratio, confirming that Hh can influence the CD4/8 lineage decision. Conversely, analysis of Shh^{-/-} thymi showed increased production of CD4 SP cells and the CD4:CD8 ratio, confirming that Hh signaling, and Shh in particular, does affect CD4/8 lineage commitment under physiological conditions (Fig. 1A). Commitment of the DP population to the CD4 or CD8 lineage is complicated by the fact that the mature CD4 and CD8 cells must express TCR restricted by the appropriate MHC molecule (by Class I in the case of CD8 SP cells, and by Class II in the case of CD4 SP cells). Numerous models have been proposed to explain how the DP population commit to the CD4 or CD8 lineage,²⁴ and TCR signal strength during repertoire selection has been shown to influence SP lineage commitment, with a stronger signal favouring differentiation to CD4 SP.²⁵⁻²⁷ The fact that manipulation of the Hh signal received by developing thymocytes influenced the CD4/8 lineage decision therefore raised the possibility that Hh was modulating TCR signal strength, and in this way affecting the lineage decision. Interestingly, we found that cell surface expression of CD5, which correlates with TCR signal



Figure 1. Summary of Hedgehog pathway activity in CD4/8 lineage commitment, TCR repertoire selection and peripheral T cell activation. (A) Reduced Hh signaling (i.e., in Shh^{-/-} thymus) promotes CD4 lineage commitment resulting in increased CD4:CD8 ratio (left). In contrast increased Hh signaling (i.e., in transgenic Lck-Gli2AN₂ thymocytes with constitutively active Hh signaling, or in thymus explants treated in vitro with r-Shh) results in reduced CD4:CD8 ratio (right). (B) Hh signaling negatively influences both positive and negative selection. Deletion of CD8 thymocytes expressing the transgenic TCR recognizing the male antigen HY (in the context of MHC class I) is impaired in Lck-Gli2 ΔN_2 male thymus but enhanced in Shh^{-/-} male thymus (left). Positive selection of Lck-Gli2 ΔN_2 CD8 cells expressing the HY-TCR is also negatively affected in the female thymus whereas positive selection of HY-TCR transgenic thymocytes is promoted in Shh^{-/-} female thymus. (C) Hedgehog signaling antagonizes in vitro T cell activation. Proliferation and activation of peripheral Lck-Gli2AN₂ T cells was inhibited. (D) Proposed model for possible role of Hh signaling and T cell activation during tissue repair or tumor development. Damaged tissues or tumors might produce and secrete Hh proteins that can signal to lymphoid cells present. We predict that Hh pathway signaling would limit T cell activation in response to Hh secretion by damaged tissues and tumors.

strength,²⁸ was decreased in the Lck-Gli2 ΔN_2 transgenic thymocytes, and after treatment of WT thymi with r-Shh. This reduction in cell surface CD5 expression therefore supported the idea that the reduced proportion of CD4 SP cells found in the Lck-Gli2 ΔN_2 transgenic, and after treatment of WT thymi with r-Shh, is a result of Hh signaling acting to reduce TCR signal strength.

To test this hypothesis, we used the HY-TCR transgenic model that allowed the study of positive and negative selection of CD8 SP cells.²⁹ HY-TCR transgenic T cells express a TCR that recognises a male antigen, restricted by a Class I MHC molecule. In male HY mice, negative selection is highly efficient with deletion of the male-reactive CD8 and DP population.²⁹ When we added the Lck-Gli2 Δ N₂ transgene to male mice positive for the HY-TCR we found a small population of HY-TCR positive (auto-reactive) CD8 SP cells that had escaped deletion.¹² The HY-TCR model can also be used to assess positive selection to the CD8 lineage in female mice.²⁹ In HY-TCR females with the Lck-Gli2 Δ N₂ transgene, we observed reduced positive selection of thymocytes expressing the transgenic TCR to the CD8 SP lineage (Fig. 1B).

To confirm the physiological significance of this observation we analysed Shh deficient HY-TCR mice. Again, analysis of Shh deficiency showed the opposite effect to the Lck-Gli2 ΔN_2 transgenic, with more efficient deletion in males and increased positive selection in females.¹² The altered selection observed using the HY-TCR model thus showed that Shh signaling does act to reduce TCR signal strength in developing thymocytes during TCR repertoire selection (Fig. 1B).

Shh is secreted by thymus epithelial cells scattered in the medulla and at the corticomedullary junction,^{2,6,9} where TCR repertoire selection takes place.³⁰ Proximity to the Shh source would dictate the strength of TCR signal received by a given cell, allowing the architecture of the thymus to influence the T cell repertoire by setting up a Hh gradient. As developing thymocytes move through the thymus they will encounter different strengths of Shh and therefore the outcome of TCR-ligation for positive and negative selection and CD4/CD8 lineage decisions will be dependent on their position in the thymus relative to the Shh signal. Not all medullary epithelial cells express Shh,^{2,6,9} so it is likely that Shh-secreting-epithelial cells are specialised for particular functions, setting up 'niches' to promote positive selection and commitment to the CD8 lineage.

The way in which the transcriptional changes induced by Hh pathway activation modulate the TCR signal during repertoire selection are unknown, and further investigation is required.

HH SIGNALING ATTENUATES PERIPHERAL T-CELL ACTIVATION AND PROLIFERATION

On antigen recognition in the periphery, naïve T cells are stimulated to clonally expand and differentiate into effector cells. This requires costimulation from antigen presenting cells (APC) and outcome is influenced by cytokines, the microenvironment and the strength of the TCR signal received.³¹

Peripheral T cell activation and proliferation require two distinct signaling events. The TCR-CD3 complex recognises antigen presented as peptide bound to MHC molecules on the surface of APC, and signals for T cell activation. In addition, for full T cell activation a costimulatory signal is required, transduced by binding of CD28 on the T cell to CD80/CD86 on the APC. These events at the cell surface trigger intracellular signaling: Immunoreceptor tyrosine-based activation motifs (ITAMs) on CD3 polypeptides are phosphorylated by Lck causing recruitment and activation of ZAP-70. This results in the activation of multiple signaling cascades including the MAPKinase, PKC, DAG Kinase, PI3Kinase and Calcineurin pathways, leading to the activation of transcription factors such as NF κ B, NFAT and AP-1. These transcription factors bind to and activate transcription of genes necessary for T cell activation,

including IL-2.³²⁻³⁴ Secretion of IL-2, a potent T cell growth factor, and upregulation of IL-2 Receptor expression by the T cell, allow autocrine control of T cell proliferation.

The strength of signal transduced by the naïve T cell on TCR binding, as determined by antigen concentration and affinity and duration of the interaction, influences further T cell maturation to specialised effector populations.³⁵ The interaction between the T cell and the APC is enhanced by adhesion molecules and integrins. These are differentially expressed depending on the activation and differentiation status of the cell, and this is itself regulated by cytokines and signals from the environment.³⁶ The final outcome of the activating signals is therefore additionally dependent on the microenvironment of both the T cell and the APC. One possible influence on T cell activation could be Hh secretion at the site of activation. The role of Shh in peripheral T cell activation and function, however, is controversial.^{6,37-39} We therefore used the Lck-Gli2 Δ N₂ transgenic model to investigate the effect of Hh pathway activation on T cell activation.

In these experiments, in vitro activation of Lck-Gli2 ΔN_2 transgenic T cells by ligation of TCR and CD28, attenuated T cell activation, indicating that the Hh signaling pathway negatively affects TCR-induced T cell activation.¹² The presence of the Lck-Gli2 ΔN_2 transgene also seriously affected the ability of T cells to proliferate in response to TCR and CD28 ligation¹² (Fig. 1C).

Hh signaling has long been linked to proliferation, with an increase in Hh signaling generally found to enhance proliferation (for example in the development of limb, taste buds, neural crest and thymus).^{4,6,10,40-44} Consistent with Hh promoting cell expansion, the first Gli protein to be discovered, Gli1, was initially identified as an amplified gene and potential oncogene in a human glioma line,^{45,46} and aberrant Hh signaling has been implicated in many common malignant tumors such as small cell lung carcinoma, pancreatic, stomach and prostate cancer.⁴⁷⁻⁵⁰

By analysis of loss-of-function mutants, T lineage studies have shown that Hh signaling promotes proliferation of early DN thymocytes.^{6,10} In contrast, we have recently found that constitutive activation of the Hh pathway inhibits the proliferation of mature T cells.¹² The effect of Hh signaling on proliferation has also appeared ambiguous in other systems, in particular in the retina.⁵¹ In chick and mouse retina Shh activation resulted in increased BrdU incorporation suggesting that Hh signaling promotes proliferation.⁵²⁻⁵⁴ However, Hh mutant Zebrafish exhibited prolonged retinal proliferation due to the inability of precursor cells to exit the cell cycle.⁵⁵ Recent work has suggested that Hh signaling may effect stem, progenitor and mature cells differently, with Hh signaling either promoting proliferation or pushing cells out of the cell cycle resulting in reduced proliferative ability depending on the state of differentiation of the cell.⁵⁶ The T cell lineage seems to provide another example of Hh activation promoting proliferation of the progenitor cell, but limiting the proliferation of the more differentiated mature cell. This difference in outcome could reflect differences in the intracellular context in which the Hh signal is transduced, leading to the transcriptional regulation of distinct sets of target genes. Alternatively, the inhibition of TCR-induced mature T cell proliferation could be the result of reduction in TCR signal strength by the pathway, rather than a direct effect on cell cycle regulator target genes. While early thymocyte progenitors must expand rapidly to fill the DP pool, and do not yet express a TCR, in differentiated T cells proliferation must only occur as a consequence of appropriate activation in response to antigen, after stimulation through their TCR. Thus, quite different Hh signaling in the T lineage, in accordance with maturity.

ERK PHOSPHORYLATION AND HH SIGNALING

To dissect further the effect of Hh signaling on TCR signal strength, peripheral T cell activation and proliferation, we followed the kinetics of MAPkinase pathway activation by measuring ERK phosphorylation after TCR-induced activation by Western blot.⁵⁷ Presence of the Lck-Gli2 ΔN_2 transgene led to much delayed and reduced levels of ERK phosphorylation, which was further confirmed to be T-cell specific using a FACS-based assay.¹²

This reduced ERK phosphorylation observed in our transgenic T cells is of particular interest, as previous accounts of Hh pathway involvement in other tissues are often linked to improved ERK phosphorylation.58-61 For example, addition of Shh during retinal regeneration, led to an increase in ERK phosphorylation indirectly via the FGF/FGFR pathway.⁵⁹ The mechanism by which the transcriptional events induced by Hh signaling reduced TCR-induced ERK phosphorylation is unknown, however it must be proximal to TCR ligation, as ERK phosphorylation is an immediate consequence of TCR ligation.62

The MAPKinase pathway is a key regulator of cell proliferation.⁵⁷ ERK activity must be sustained late into G1 to enable successful S-phase entry⁶³ and inhibitors of ERK phosphorylation have been shown to inhibit the proliferation of many cell types including T cells.⁵⁷ A reduction in ERK phosphorylation leads to a decrease in the transcription of ubiquitous mediators of cell cycle progression and in the case of activating T cells, also of IL-234,57,64 any of which could lead to the observed attenuation of proliferation in the Lck-Gli2AN2 cells. In our experiments, addition of IL-2 to activated cultures restored proliferation of Lck-Gli $2\Delta N_2$ T cells to WT levels, suggesting that a reduction in IL-2 gene transcription (caused by attenuated MAPkinase activity⁶⁴) is responsible for the proliferative defect. The fact the Lck-Gli2AN2 T cells can proliferate in response to IL-2 demonstrated that the transcriptional changes induced by active Hh signaling did not inherently inhibit T cell division.

Thus, we favour a model in which the decreased proliferation and ERK activation of mature T cells on TCR ligation, caused by constitutive Hh signaling, is a consequence of reduced TCR signal strength mediated by Hh-dependent transcription, rather than a direct effect of Hh on the cells' proliferation per se.

IMPLICATIONS FOR IMMUNITY AND DISEASE

Modulation of TCR signal strength by Hh has profound implications for immunity. Presence of Hh would alter the threshold required for T cell activation, and therefore determine the outcome of the naïve T cell's interaction with antigen, with consequences for anergy, regulation, tolerance and inflammation. This has significance for immune responses, as localised sources of Hh in the T cell microenvironment could influence the outcome of TCR signaling. In the future, it will be important to test this hypothesis in mouse models and in human disease.

Hh proteins are expressed in skin, gut and lung.¹ These tissues are subject to recurrent immune challenge, as they are the sites of entry of external pathogens from the environment, from food and air. They are also subject to inflammatory and autoimmune diseases. Hh expression in these tissues during renewal or remodelling after infection or tissue-damage could function to dampen down the immune response, protecting against the induction of inflammatory

processes initiate proliferation, consistent with the opposing effects of or autoimmune diseases (Fig. 1D). By reducing lymphocyte responses to antigen, Hh signaling may also be involved in immune evasion by tumours. Aberrant Hh signaling is common in many cancers^{5,17} and secretion of Hh by a tumour could potentially diminish the immune response to that tumour, allowing escape from immune surveillance.

Changes in Hh signaling could be involved in the aetiology of autoimmune disease and mutations in components of the pathway could increase disease susceptibility, either by allowing self-reactive T cells to escape negative selection in the thymus, or by reducing the TCR signal threshold for mature T cell activation. In the thymus, mutations increasing either Shh secretion by epithelial cells, or signal interpretation by thymocytes, would allow those self-reactive clones that would normally be deleted, to mature and escape into the peripheral lymphoid organs, potentially leading to autoimmune disease. In contrast, mutations leading to decreased Hh signal in peripheral T cells (via decreased secretion in the environment or signal interpretation by the T cell) would increase the TCR signal, potentially leading to inappropriate T cell activation.

The role of Hh signaling in T cells (and immunity) is a new field, so there is still much work to be done to dissect its functions and gain insight into the molecular mechanisms. It will be important to understand how Hh modulates the TCR signal, and to investigate the effect of Hh gradient on the signal. It will also be interesting to test if Hh signaling regulates other components of the immune system.

References

- 1. Ingham PW, McMahon AP. Hedgehog signaling in animal development: Paradigms and principles. Genes Dev 2001; 15:3059-87.
- 2. Outram SV, Varas A, Pepicelli CV, Crompton T. Hedgehog signaling regulates differentiation from double-negative to double-positive thymocyte. Immunity 2000; 13:187-97.
- Bhardwaj G, Murdoch B, Wu D, Baker DP, Williams KP, Chadwick K, Ling LE, Karanu FN, Bhatia M. Sonic hedgehog induces the proliferation of primitive human hematopoietic cells via BMP regulation. Nat Immunol 2001; 2:172-80.
- Gill PS, Rosenblum ND. Control of murine kidney development by sonic hedgehog and its GLI effectors. Cell Cycle 2006; 5:1426-30.
- 5. Taipale J, Beachy PA. The Hedgehog and Wnt signaling pathways in cancer. Nature 2001; 411:349-54.
- 6. El Andaloussi A, Graves S, Meng F, Mandal M, Mashayekhi M, Aifantis I. Hedgehog signaling controls thymocyte progenitor homeostasis and differentiation in the thymus. Nat Immunol 2006; 7:418-26.
- 7. Gutierrez-Frias C, Sacedon R, Hernandez-Lopez C, Cejalvo T, Crompton T, Zapata AG, Varas A, Vicente A. Sonic hedgehog regulates early human thymocyte differentiation by counteracting the IL-7-induced development of CD34+ precursor cells. J Immunol 2004; 173:5046-53
- 8. Sacedon R, Diez B, Nunez V, Hernandez-Lopez C, Gutierrez-Frias C, Cejalvo T, Outram SV, Crompton T, Zapata AG, Vicente A, Varas A. Sonic hedgehog is produced by follicular dendritic cells and protects germinal center B cells from apoptosis. J Immunol 2005; 174:1456-61.
- 9. Sacedon R, Varas A, Hernandez-Lopez C, Gutierrez-deFrias C, Crompton T, Zapata AG, Vicente A. Expression of hedgehog proteins in the human thymus. J Histochem Cytochem 2003; 51:1557-66.
- 10. Shah DK, Hager-Theodorides AL, Outram SV, Ross SE, Varas A, Crompton T. Reduced thymocyte development in sonic hedgehog knockout embryos. J Immunol 2004; 172:2296-306
- 11. Hager-Theodorides AL, Dessens JT, Outram SV, Crompton T. The transcription factor Gli3 regulates differentiation of fetal CD4- CD8- double-negative thymocytes. Blood 2005; 106:1296-304
- 12. Rowbotham NJ, Hager-Theodorides AL, Cebecauer M, Shah DK, Drakopoulou E, Dyson J, Outram SV, Crompton T. Activation of the Hedgehog signaling pathway in T-lineage cells inhibits TCR repertoire selection in the thymus and peripheral T-cell activation. Blood 2007: 109:3757-66.
- 13. Bai CB, Auerbach W, Lee JS, Stephen D, Joyner AL. Gli2, but not Gli1, is required for initial Shh signaling and ectopic activation of the Shh pathway. Development 2002; 129:4753-61.
- 14. Sasaki H, Nishizaki Y, Hui C, Nakafuku M, Kondoh H. Regulation of Gli2 and Gli3 activities by an amino-terminal repression domain: Implication of Gli2 and Gli3 as primary mediators of Shh signaling. Development 1999; 126:3915-24.
- 15. Park HL, Bai C, Platt KA, Matise MP, Beeghly A, Hui CC, Nakashima M, Joyner AL. Mouse Gli1 mutants are viable but have defects in SHH signaling in combination with a Gli2 mutation. Development 2000; 127:1593-605.

- te Welscher P, Fernandez-Teran M, Ros MA, Zeller R. Mutual genetic antagonism involving GLI3 and dHAND prepatterns the vertebrate limb bud mesenchyme prior to SHH signaling. Genes Dev 2002; 16:421-6.
- 17. Matise MP, Joyner AL. Gli genes in development and cancer. Oncogene 1999; 18:7852-9.
- Koebernick K, Pieler T. Gli-type zinc finger proteins as bipotential transducers of Hedgehog signaling. Differentiation 2002; 70:69-76.
- Mallick CA, Dudley EC, Viney JL, Owen MJ, Hayday AC. Rearrangement and diversity of T cell receptor beta chain genes in thymocytes: A critical role for the beta chain in development. Cell 1993; 73:513-9.
- 20. Germain RN. T-cell development and the CD4-CD8 lineage decision. Nat Rev Immunol 2002; 2:309-22.
- 21. von Boehmer H. Shaping the T cell repertoire. J Immunol 2006; 176:3-4.
- Buckland J, Pennington DJ, Bruno L, Owen MJ. Coordination of the expression of the protein tyrosine kinase p56(lck) with the pre-T cell receptor during thymocyte development. European Journal of Immunology 2000; 30:8-18.
- Chaffin KE, Beals CR, Wilkie TM, Forbush KA, Simon MI, Perlmutter RM. Dissection of thymocyte signaling pathways by in vivo expression of pertussis toxin ADP-ribosyltransferase. Embo J 1990; 9:3821-9.
- He X, Kappes DJ. CD4/CD8 lineage commitment: Light at the end of the tunnel? Current Opinion in Immunology 2006; 18:135-42.
- Hernandez-Hoyos G, Sohn SJ, Rothenberg EV, Alberola-Ila J. Lck activity controls CD4/ CD8 T cell lineage commitment. Immunity 2000; 12:313-22.
- Bommhardt U, Basson MA, Krummrei U, Zamoyska R. Activation of the extracellular signal-related kinase/mitogen-activated protein kinase pathway discriminates CD4 versus CD8 lineage commitment in the thymus. J Immunol 1999; 163:715-22.
- 27. Kappes DJ, He X, He X. CD4-CD8 lineage commitment: An inside view. Nat Immunol 2005; 6:761-6.
- Azzam HS, DeJarnette JB, Huang K, Emmons R, Park CS, Sommers CL, El-Khoury D, Shores EW, Love PE. Fine tuning of TCR signaling by CD5. J Immunol 2001; 166:5464-72.
- Kisielow P, Teh HS, Bluthmann H, von Boehmer H. Positive selection of antigen-specific T cells in thymus by restricting MHC molecules. Nature 1988; 335:730-3.
- Petrie HT. Role of thymic organ structure and stromal composition in steady-state postnatal T-cell production. Immunol Rev 2002; 189:8-19.
- Leitenberg D, Bottomly K. Regulation of naive T cell differentiation by varying the potency of TCR signal transduction. Semin Immunol 1999; 11:283-92.
- Berridge MJ. Lymphocyte activation in health and disease. Crit Rev Immunol 1997; 17:155-78.
- Borde M, Barrington RA, Heissmeyer V, Carroll MC, Rao A. Transcriptional basis of lymphocyte tolerance. Immunol Rev 2006; 210:105-19.
- Jain J, Loh C, Rao A. Transcriptional regulation of the *IL-2* gene. Curr Opin Immunol 1995; 7:333-42.
- Hayes SM, Love PE. Strength of signal: A fundamental mechanism for cell fate specification. Immunol Rev 2006; 209:170-5.
- Dailey MO. Expression of T lymphocyte adhesion molecules: Regulation during antigen-induced T cell activation and differentiation. Critical Reviews in Immunology 1998; 18:153-84.
- Lowrey JA, Stewart GA, Lindey S, Hoyne GF, Dallman MJ, Howie SE, Lamb JR. Sonic hedgehog promotes cell cycle progression in activated peripheral CD4(+) T lymphocytes. J Immunol 2002; 169:1869-75.
- Stewart GA, Lowrey JA, Wakelin SJ, Fitch PM, Lindey S, Dallman MJ, Lamb JR, Howie SE. Sonic hedgehog signaling modulates activation of and cytokine production by human peripheral CD4+ T cells. J Immunol 2002; 169:5451-7.
- Chan VS, Chau SY, Tian L, Chen Y, Kwong SK, Quackenbush J, Dallman M, Lamb J, Tam PK. Sonic hedgehog promotes CD4+ T lymphocyte proliferation and modulates the expression of a subset of CD28-targeted genes. Int Immunol 2006; 18:1627-36.
- Lai K, Kaspar BK, Gage FH, Schaffer DV. Sonic hedgehog regulates adult neural progenitor proliferation in vitro and in vivo. Nature Neuroscience 2003; 6:21-7.
- Fu M, Lui VC, Sham MH, Pachnis V, Tam PK. Sonic hedgehog regulates the proliferation, differentiation, and migration of enteric neural crest cells in gut. J Cell Biol 2004; 166:673-84.
- McGlinn E, Tabin CJ. Mechanistic insight into how Shh patterns the vertebrate limb. Curr Opin Genet Dev 2006; 16:426-32.
- Miura H, Kusakabe Y, Harada S. Cell lineage and differentiation in taste buds. Arch Histol Cytol 2006; 69:209-25.
- Knoepfler PS, Kenney AM. Neural precursor cycling at sonic speed: N-Myc pedals, GSK-3 brakes. Cell Cycle 2006; 5:47-52.
- Kinzler KW, Bigner SH, Bigner DD, Trent JM, Law ML, O'Brien SJ, Wong AJ, Vogelstein B. Identification of an amplified, highly expressed gene in a human glioma. Science 1987; 236:70-3.
- 46. Kinzler KW, Ruppert JM, Bigner SH, Vogelstein B. The *GLI* gene is a member of the Kruppel family of zinc finger proteins. Nature 1988; 332:371-4.
- Karhadkar SS, Bova GS, Abdallah N, Dhara S, Gardner D, Maitra A, Isaacs JT, Berman DM, Beachy PA. Hedgehog signaling in prostate regeneration, neoplasia and metastasis. Nature 2004; 431:707-12.
- Berman DM, Karhadkar SS, Maitra A, Montes De Oca R, Gerstenblith MR, Briggs K, Parker AR, Shimada Y, Eshleman JR, Watkins DN, Beachy PA. Widespread requirement for Hedgehog ligand stimulation in growth of digestive tract tumours. Nature 2003; 425:846-51.

- 49. Thayer SP, di Magliano MP, Heiser PW, Nielsen CM, Roberts DJ, Lauwers GY, Qi YP, Gysin S, Fernandez-del Castillo C, Yajnik V, Antoniu B, McMahon M, Warshaw AL, Hebrok M. Hedgehog is an early and late mediator of pancreatic cancer tumorigenesis. Nature 20003; 425:851-6.
- Watkins DN, Berman DM, Burkholder SG, Wang B, Beachy PA, Baylin SB. Hedgehog signaling within airway epithelial progenitors and in small-cell lung cancer. Nature 2003; 422:313-7.
- Agathocleous M, Locker M, Harris WA, Perron M. A general role of hedgehog in the regulation of proliferation. Cell Cycle 2007; 6:156-9.
- Jensen AM, Wallace VA. Expression of Sonic hedgehog and its putative role as a precursor cell mitogen in the developing mouse retina. Development 1997; 124:363-71.
- Moshiri A, McGuire CR, Reh TA. Sonic hedgehog regulates proliferation of the retinal ciliary marginal zone in posthatch chicks. Dev Dyn 2005; 233:66-75.
- Wang Y, Dakubo GD, Thurig S, Mazerolle CJ, Wallace VA. Retinal ganglion cell-derived sonic hedgehog locally controls proliferation and the timing of RGC development in the embryonic mouse retina. Development 2005; 132:5103-13.
- 55. Neumann CJ. Hedgehogs as negative regulators of the cell cycle. Cell Cycle 2005; 4:1139-40.
- Locker M, Agathocleous M, Amato MA, Parain K, Harris WA, Perron M. Hedgehog signaling and the retina: Insights into the mechanisms controlling the proliferative properties of neural precursors. Genes Dev 2006; 20:3036-48.
- Meloche S, Pouyssegur J. The ERK1/2 mitogen-activated protein kinase pathway as a master regulator of the G₁- to S-phase transition. Oncogene 2007; 26:3227-39.
- Xie J, Aszterbaum M, Zhang X, Bonifas JM, Zachary C, Epstein E, McCormick F. A role of PDGFRalpha in basal cell carcinoma proliferation. Proceedings of the National Academy of Sciences of the United States of America 2001; 98:9255-9.
- Spence JR, Aycinena JC, Del Rio-Tsonis K. Fibroblast growth factor-hedgehog interdependence during retina regeneration. Dev Dyn 2007; 236:1161-74.
- Osawa H, Ohnishi H, Takano K, Noguti T, Mashima H, Hoshino H, Kita H, Sato K, Matsui H, Sugano K. Sonic hedgehog stimulates the proliferation of rat gastric mucosal cells through ERK activation by elevating intracellular calcium concentration. Biochem Biophys Res Commun 2006; 344:680-7.
- Riobo NA, Lu K, Emerson Jr CP. Hedgehog signal transduction: Signal integration and cross talk in development and cancer. Cell Cycle 2006; 5:1612-5.
- Alberola-Ila J, Hernandez-Hoyos G. The Ras/MAPK cascade and the control of positive selection. Immunol Rev 2003; 191:79-96.
- Yamamoto T, Ebisuya M, Ashida F, Okamoto K, Yonehara S, Nishida E. Continuous ERK activation downregulates antiproliferative genes throughout G₁ phase to allow cell-cycle progression. Curr Biol 2006; 16:1171-82.
- Kemp ML, Wille L, Lewis CL, Nicholson LB, Lauffenburger DA. Quantitative network signal combinations downstream of TCR activation can predict IL-2 production response. J Immunol 2007; 178:4984-92.