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pH-Responsive Nano-Self-Assemblies of the Anticancer Drug 2-Hydroxyoleic Acid

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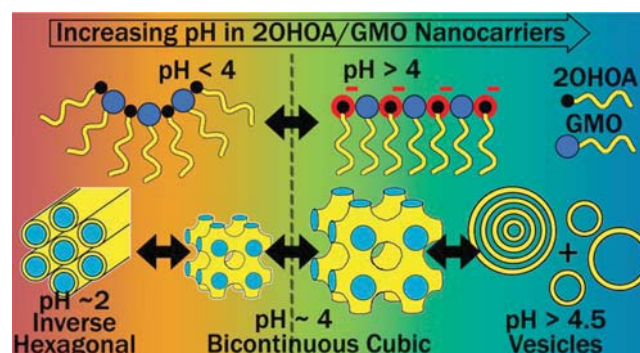
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Supporting Information

ABSTRACT: pH-responsive lipid nanocarriers have the potential to selectively target the acidic extracellular pH environment of cancer tissues and may further improve the efficacy of chemotherapeutics by minimizing their toxic side-effects. Here, we present the design and characterization of pH-sensitive nano-self-assemblies of the poorly water-soluble anticancer drug 2-hydroxyoleic acid (2OHOA) with glycerol monooleate (GMO). pH-triggered nanostructural transformations from 2OHOA/GMO nanoparticles with an internal inverse hexagonal structure (hexosomes) at pH around 2.0–3.0, via nanocarriers with an internal bicontinuous cubic structure (cubosomes) at pH 2.0–4.5, to vesicles at pH 4.5–7.4 were observed with synchrotron small-angle X-ray scattering, and cryogenic transmission electron microscopy. ζ -potential measurements highlight that the pH-driven deprotonation of the carboxylic group of 2OHOA, and the resulting charge-repulsions at the lipid–water interface account for these nanostructural alterations. The study provides detailed insight into the pH-dependent self-assembly of 2OHOA with GMO in excess buffer at physiologically relevant pH values, and discusses the effects of pH alterations on modulating their nanostructure. The results may guide the further development of pH-responsive anticancer nanocarriers for the targeted delivery of chemotherapeutics to the local microenvironment of tumor cells.



■ INTRODUCTION

Cancer is one of the major public health problems worldwide.^{1–3} Along with the current cancer therapies, including surgical intervention and radiation therapy, chemotherapy is characterized as the least invasive cancer treatment approach with a clinical success in tumor shrinking and cancer relapse reduction.^{4–6} However, chemotherapy is often associated with side-effects caused by the off-site toxicity due to the lack of drug specificity.⁵ Hence, the design of novel and more efficient cancer nanomedicines that can selectively target the tumor sites, avoiding off-target side-effects, may improve current therapies and also patients' quality of life.^{5,7–10} Among different suggested nanomedicines, pH-triggered chemotherapeutic delivery systems may be of interest in cancer treatment, as the extracellular microenvironment of tumor cells with pH of around 5.5–6.5 is more acidic than that of healthy cells with pH of 7.4.^{10–12} The occurrence of such acidic local tumor microenvironment is attributed to an increased glycolysis rate, a higher production level of lactic acid, and an insufficient vasculature for the removal of acidic byproducts.^{11,13–18}

This study describes the formation and characterization of pH-responsive self-assembled nanocarriers based on glycerol

monooleate (GMO), and the anticancer drug 2-hydroxyoleic acid (2OHOA). The molecular structures of both lipids are presented in Figure 1. The poorly water-soluble anticancer drug 2OHOA is a derivative of oleic acid (OA) with an additional hydroxyl group on the α -carbon of the fatty acid backbone.¹⁹ It is a potent anticancer agent that induces cell

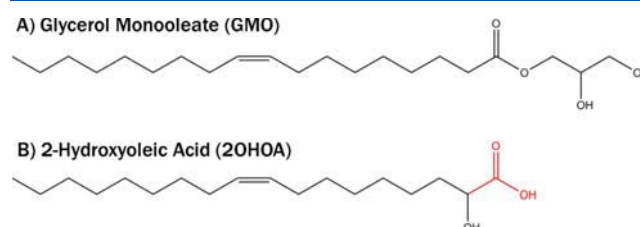


Figure 1. Molecular structures of the lipids: (A) glycerol monooleate (GMO) and (B) 2-hydroxyoleic acid (2OHOA). The carboxylic group of 2OHOA is highlighted in red color in panel B.

cycle arrest and apoptosis, cellular differentiation, and autophagy in a wide range of human cancer cells, including leukemia, glioma, lung, colon, and breast cancer cells.^{19–27}

GMO-based nanocarriers have been previously used for enhancing the solubilization of poorly water-soluble therapeutic agents.^{28–35} Among these nanocarriers, dispersions of inverse bicontinuous cubic, hexagonal, and micellar cubic structures (cubosomes, hexosomes, and micellar cubosomes, respectively), emulsified microemulsions, vesicles, and sponge phases are gaining increasing attention for their drug delivery applications. The biological relevance and the relatively large extent of the internal lipid–water interfacial area render these colloidal nanosystems attractive for delivering hydrophilic, hydrophobic, and amphiphilic drugs and peptide molecules.^{36–48} For targeted delivery applications, they can be surface nanoengineered to respond to an external stimulus, such as pH, for triggering the release of therapeutic molecules or enhancing the interactions with cells on demand, owing to structural alterations in these nanocarriers.^{33,49–55} Despite the attractiveness of cubosomes, hexosomes, and related non-lamellar liquid crystalline nanoparticles in the development of injectable drug nanocarriers, there is still to date no FDA-approved formulation based on these nanostructured emulsions.^{56,57} Nevertheless, recent *in vivo* studies showed that these nanoparticles are promising candidates in the development of nanocarriers for anticancer drug delivery.^{58,59}

The major goal of this study is to investigate the composition and pH dependence of 2OHOA/GMO nano-self-assemblies and gain further insight into their pH-triggered structural, morphological, and size alterations by using synchrotron small-angle X-ray scattering (SAXS), cryogenic transmission electron microscopy (cryo-TEM), and ζ -potential measurements. An improved understanding of the structural mechanism involved in these pH-triggered transitions is an important keystone toward the development of pH-responsive anticancer nanomedicines. Based on the presented experimental findings, the pH-induced phase transformations in the interiors of 2OHOA/GMO nanoparticles are most likely attributed to the protonation/deprotonation of the carboxylic group of 2OHOA at the lipid–water interface. The presented results shed light on the molecular interactions during the self-assembly of a cell membrane active anticancer agent with a biologically relevant lipid in excess buffer and could potentially be applied for the future design of pH-guided nanocarriers for tumor-targeted delivery of anticancer drugs.

■ EXPERIMENTAL SECTION

Preparation of 2OHOA/GMO Nanodispersions. Sodium salt of 2-hydroxyoleic acid (2OHOA; Avanti polar lipids, Alabama, U.S.A.) and glycerol monooleate (GMO; $\geq 90\%$ purity, Riken Vitamin co., Ltd. company, Japan) were weighed out in glass vials at 3/7 and 1/1 2OHOA/GMO mass ratios and dispersed in 149 mM Dulbecco's Phosphate-buffered saline (PBS) at pH 7.4 (Sigma-Aldrich, Poole, UK), containing 2 mg/mL Pluronic F127 (gift from BASF SE, Ludwigshafen, Germany), to achieve a total lipid concentration of 20 mg/mL. These raw lipid emulsions were then homogenized by means of ultrasonication (Ultrasonic Processor Qsonica 500, Qsonica LLC, Newtown, CT, U.S.A.) for 4 min in pulse mode (3 s pulse, 5 s break) at 27% of its maximum power (500 W). The pH of the prepared nanodispersions was then adjusted with 1 M NaOH ($\geq 99\%$ purity, Carl Roth GmbH, Karlsruhe, Germany) or 1 M HCl prepared from 37% HCl stock solution (ACS reagent grade, Sigma-Aldrich, Buchs, Switzerland) and allowed to equilibrate at 25 °C for at least 1 h before carrying out the planned experiments. In this study, the pH of the

continuous aqueous medium of the dispersion with 1/1 2OHOA/GMO mass ratio was first gradually decreased from 7.4 to 2.0 and then increased back to 5.0, while taking out samples at different selected pH points for SAXS structural characterization. This pH cycle was applied to study whether the structural alterations associated with changing pH can be reversed. Double distilled water was used for the preparation of all samples. All ingredients were used without further purification.

Synchrotron Small-Angle X-ray Scattering (SAXS). SAXS patterns were recorded at the Austrian SAXS beamline at the synchrotron light source ELETTRA (Trieste, Italy). An X-ray beam with a wavelength of 1.54 Å (8 keV) was used, with a sample-to-detector distance of 1314 mm, covering a q -range from about 0.18 to 5.00 nm⁻¹, where q is the length of the scattering vector, defined by $q = 4\pi/\lambda \sin(\theta/2)$, λ being the wavelength, and θ the scattering angle. The samples were sealed in thin-walled quartz capillaries and five frames with an exposure time of 20 s per frame were collected and averaged at room temperature. The 2D SAXS patterns were acquired using a Pilatus3 1 M detector (Dectris Ltd., Baden, Switzerland; active area 169 × 179 mm² with a pixel size of 172 × 172 μm^2), integrated into one-dimensional (1-D) scattering function $I(q)$ using Fit2D⁶⁰ (European Synchrotron Radiation Facility, Grenoble, France) and then analyzed with IGOR pro (Wavemetrics, Inc., Lake Oswego, U.S.A.). The scattering curves were plotted as a function of intensity, $I(q)$, averaged over the five repeated exposures versus q . The scattering from PBS was subtracted as background from all measurements before further data analysis. The calculation of the lattice parameters of the corresponding lyotropic nonlamellar liquid crystalline phases is described in SI (eqs S2–S4).

Cryogenic-Transmission Electron Microscopy (cryo-TEM). The morphological characterization of F127-stabilized 2OHOA/GMO nanoparticles was done in their vitrified states by cryo-TEM imaging. Briefly, lacey carbon 300 mesh copper grid (Ted Pella Inc., CA, U.S.A.) enforced by a silicon monoxide coating was subjected to glow discharge treatment and treated with 3 μL of the sample. The excess sample on the grid was then blotted with filter paper at a blotting time of 3.5 s, a blotting force 0, temperature 25 °C, and 100% humidity (FEI Vitrobot IV, Holland) and was rapidly plunged into liquid nitrogen-cooled ethane close to its melting point (−174 °C). A Gatan 626 cryo-holder (Gatan, U.K.) was used to transfer the plunged grid with the samples in a frozen glassy state into the Tecnai G2 20 transmission electron microscope (FEI, Holland). The images were digitally recorded at a voltage of 200 kV, under low-dose conditions ($\sim 5 \text{ e}/\text{\AA}^2\text{s}$), and at a magnification of $\times 29\,000$.

ζ -Potential Measurements. The ζ -potential of the 2OHOA/GMO nanodispersions was measured at different pH using Zetasizer Nano ZS equipped with a 633 nm laser (Malvern Instruments, Worcestershire, U.K.). Prior to measurements at 25 °C and 173° scattering angle, the samples were diluted 1/100 in PBS and their pH was readjusted. ζ -potential was calculated by Zetasizer Software 7.11 (Malvern Instruments) using Smoluchowski equation:⁶¹

$$\mu_e = \frac{\epsilon_r \epsilon_0 \zeta}{\eta} \quad (1)$$

where μ_e is the electrophoretic mobility, ϵ_r is the dielectric constant of water, ϵ_0 is the permittivity of vacuum, and η is the viscosity of the solvent (water). The measurements were done in triplicate and averaged. Estimations of 2OHOA's pK_a^{app} from the ζ -potential values are described in the SI.

■ RESULTS AND DISCUSSION

The GMO dispersed in water forms cubosomes with an internal bicontinuous cubic phase of the $Pn3m$ or $Im3m$ type symmetry depending on the F127 stabilizer concentration.^{62,63} The integration of the anticancer drug 2OHOA into this dispersion led to composition- and pH-dependent nano-structural transformations in the nanocarriers. Figure 2 demonstrates the pH-dependent self-assembly of the binary

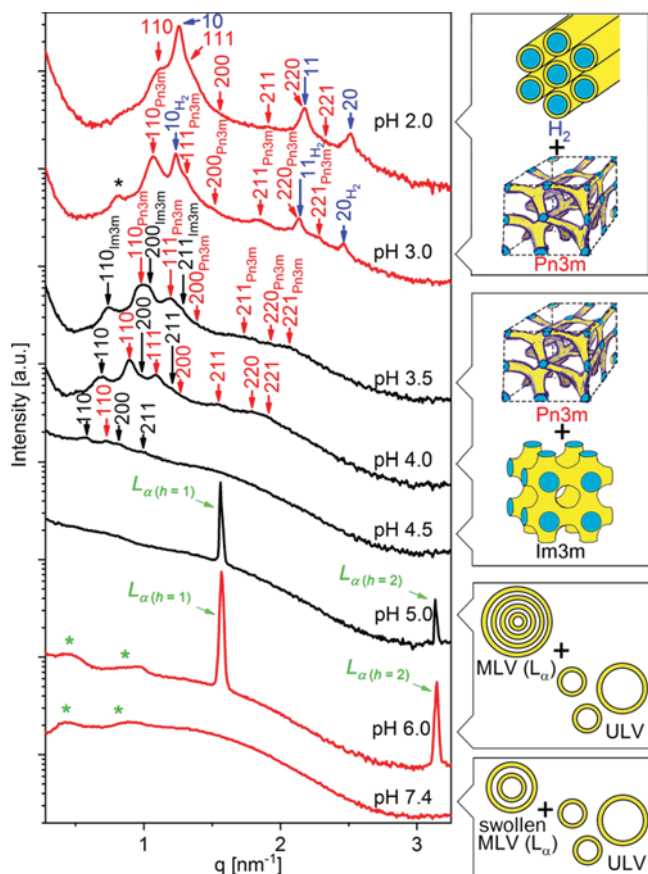


Figure 2. SAXS patterns for the F127-stabilized nano-self-assemblies prepared at 2OHOA/GMO mass ratio of 1/1 as the pH of the dispersion was gradually decreased from 7.4 to 2.0 (red curves) and then increased back to 5.0 (black curves). The characteristic Bragg peaks and the corresponding Miller indices of the cubic $Pn3m$ (red), $Im3m$ (black), and H_2 (blue) phases are marked with arrows. Low intensity peak attributed to a potential coexistence of $Im3m$ phase is marked with a black asterisk at pH 3.0. The first two Bragg reflections from the interlamellar distance of the MLVs are marked with $L_{\alpha}(h=1)$ and $L_{\alpha}(h=2)$ (green). Broad correlation peaks most likely corresponding to weak bilayer correlations in a coexisting population of swollen MLVs are marked with green asterisks.

2OHOA/GMO self-assemblies in excess buffer at a mass ratio of 1/1. The appearance of three dominating Bragg reflections in the SAXS curves at q values of 1.26, 2.17, and 2.51 nm^{-1} , corresponding to a peak spacing-ratio of $1:\sqrt{3}:\sqrt{4}$, indicate the formation of hexosomes with an inverse hexagonal (H_2) phase at pH 2.0. The corresponding lattice parameter, a_{H_2} , is 5.8 nm. Additional weak reflections, occurring most prominently around $q \approx 1.1 \text{ nm}^{-1}$, indicate a coexisting bicontinuous cubic structure with $Pn3m$ symmetry having a lattice parameter, a_{Pn3m} , of 8.0 nm. These results demonstrate that the 2OHOA mostly integrates into the hydrophobic domains of the self-assembled nanostructure at pH 2.0. This integration increases the apparent volume of the hydrophobic domains, which ultimately leads to the nanostructural transition from cubosomes to hexosomes. This observation agrees with theoretical considerations from the critical packing parameter model (see SI). The trend is also consistent with 2OHOA-induced lamellar-nonlamellar structural transitions in dielaidoylphosphatidylethanolamine dispersion.^{24,64}

Upon increasing the pH from 2.0 to 3.0, the Bragg peaks of the H_2 phase shifted to lower q values, corresponding to an increase in a_{H_2} to 5.9 nm (Figure 2). In addition to the characteristic reflections of the H_2 phase, additional Bragg peaks with peak-ratios of $\sqrt{2}:\sqrt{3}:\sqrt{4}:\sqrt{6}:\sqrt{8}:\sqrt{9}$ were detected. These peaks could be indexed as the (110), (111), (200), (211), (220), and (221) reflection of the bicontinuous $Pn3m$ -type cubic structure with $a_{Pn3m} = 8.3 \text{ nm}$. In this phase assignment, it should be noted that the characteristic reflection (220) of the cubic $Pn3m$ phase at pH 2.0 and 3.0 appears to overlap with the characteristic reflection (11) of the coexisting H_2 phase. A single peak around $q \approx 0.8 \text{ nm}^{-1}$ was also observed, likely resulting from traces of a coexisting $Im3m$ -type bicontinuous cubic structure. The identification of this coexisting $Im3m$ structure was based on its gradual evolvement with a further increase in pH. At pH 3.5, the Bragg reflections of the H_2 phase diminished and peaks indicating a coexistence of $Pn3m$ and $Im3m$ phase were observed. The corresponding lattice parameters for the biphasic $Pn3m/Im3m$ feature were 9.1 and 12.0 nm, respectively. Further increase in pH to 4.0 caused a shift in the peak positions of the $Pn3m$ and $Im3m$ phases to lower q values, resulting in an increase in a_{Pn3m} to 9.9 nm and a_{Im3m} to 12.9 nm. At pH 4.5, the lattice parameters of $Pn3m$ and $Im3m$ increased to 12.3 and 15.3 nm, respectively. The ratio of a_{Im3m}/a_{Pn3m} was calculated and found to be 1.32, 1.30, and 1.24 at pH 3.5, 4.0, and 4.5. This ratio is consistent with the reported theoretical Bonnet ratio of 1.279 for these structures^{36,65} (Table S1). At pH 5.0 and 6.0, two Bragg peaks at q values of about 1.56 and 3.12 nm^{-1} were observed, showing the formation of multilamellar vesicles (MLVs) with an interlamellar d -spacing of about 4.0 nm, in agreement with the previously reported bilayer dimensions in GMO-based self-assemblies.^{66,67} At pH 7.4, the diffuse-dominating scattering patterns indicated the formation of unilamellar vesicles (ULVs). The additional broad correlation peaks in the low- q region of the SAXS curves at pH 6.0 and 7.4, with maxima around $q \approx 0.45$ and 0.90 nm^{-1} are most likely attributed to weak bilayer correlations in a coexisting population of swollen bi- or multilamellar vesicles with an estimated d of around $\sim 14 \text{ nm}$. The formation of highly swollen lamellar structures with similar bilayer-to-bilayer spacing and low repeat numbers was also previously reported for nano-self-assemblies of GMO with the anionic phospholipid dioleoyl-phosphatidylglycerol.⁶⁷

To evaluate the potential to reverse the colloidal transformations in this system, the pH in the sample at 1/1 2OHOA/GMO mass ratio was decreased from pH 7.4, where vesicles were the dominating species, to pH 2.0 where cubosomes and hexosomes dominated (Figure 2). The following pH increase to 5.0 in this sample led to the reformation of vesicles, demonstrating the pH-induced transformations from nonlamellar structures to vesicles and back upon circulating the pH of the continuous water phase. The pH-triggered swelling of nonlamellar liquid crystalline structures and the gradual nonlamellar-lamellar transitions have been previously reported for self-assemblies containing the analogue OA^{50,52,69,70} and other fatty acids.^{49,71}

The cryo-TEM images of the 2OHOA/GMO nano-self-assemblies at 1/1 mass ratio confirmed the pH-triggered nanostructural transitions observed by SAXS. Hexosomes were observed at pH 2.0, and internally structured nanoparticles coexisting with vesicles were found at pH 4.0 (Figure 3A,B). At pH 6.0, the cryo-TEM image (Figure 3C) indicated the

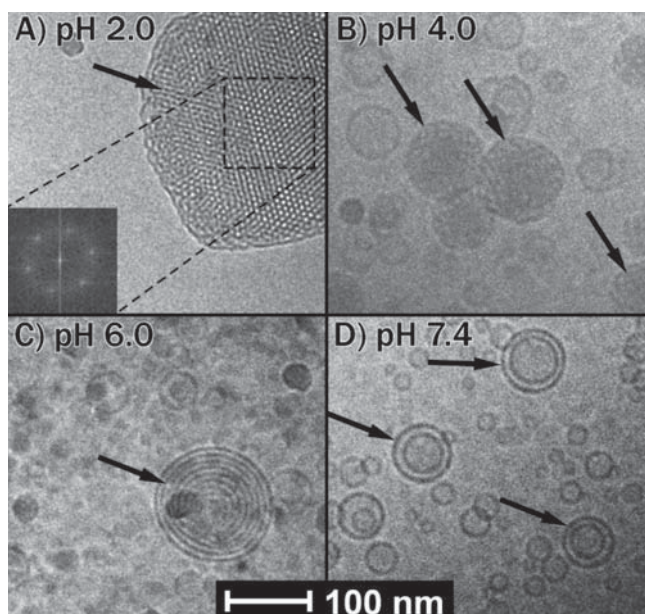


Figure 3. Representative cryo-TEM images of the nanodispersions prepared at 2OHOA/GMO mass ratio of 1/1 and the following pH: (A) pH 2.0, (B) pH 4.0, (C) pH 6.0, and (D) pH 7.4. The insert in panel A shows the fast Fourier transformation (FFT) applied to the structured part of the image indicating the formation of hexosomes. Colloidal transitions from (A) hexosomes to (B) cubosomes, and (C) MLVs (marked with black arrows) coexisting with relatively small nano-self-assemblies were observed with increasing pH. Small ULVs of variable sizes from around 100 to 15 nm in diameter were observed at pH 7.4, coexisting with multilamellar vesicles (marked with black arrows) with a bilayer-to-bilayer distance of around ~ 10 nm (D).

presence of MLVs coexisting with smaller ULVs. The image at pH 7.4 (Figure 3D) was dominated by ULVs mostly below 100 nm in diameter, coexisting with bilamellar vesicles. The bilayer-to-bilayer distance of around 10 nm in the cryo-TEM of multilamellar vesicles at pH 7.4 (Figure 3D) correlates reasonably well with the estimated interlamellar distance from SAXS (Figure 2).

Figure 4 shows the SAXS patterns for the 2OHOA/GMO nano-self-assemblies at a 2OHOA/GMO mass ratio of 3/7, in the pH range of 2.0–7.4. At pH 2.0, the SAXS curve shows the characteristic peak-spacing of the $Pn3m$ type cubic phase with a_{Pn3m} of 8.39 nm. Hence, compared to the dispersion that was prepared at a mass ratio of 1/1, the 2OHOA content in this sample is insufficient to induce a $Pn3m$ -to- H_2 phase transition even at pH 2.0. On increasing pH to 3.0 and 3.5, the calculated lattice parameter of the cubic $Pn3m$ phase increased to 8.6 and 9.3 nm, respectively. However, three additional Bragg peaks at q values of 0.72, 1.02, and 1.25 nm^{-1} with peak spacing-ratio of $\sqrt{2}:\sqrt{4}:\sqrt{6}$ were observed at pH 3.5, which can be attributed to a coexisting $Im3m$ phase with a_{Im3m} of 12.3 nm. The calculated ratio between the lattice parameters of $Im3m$ and $Pn3m$ phases was 1.33 and fairly consistent with the theoretical Bonnet ratio (Table S1). The SAXS curve at pH 4.0 was dominated by the $Im3m$ phase with a_{Im3m} of about 14.6 nm, as the three $Im3m$ peaks were shifted to lower q values. Further increase in pH to 4.5–7.4 led to the loss of the highly ordered nonlamellar structures and the appearance of diffusive SAXS curves, indicating a transformation to vesicles. The correlation peaks observed in the low- q region of curves at pH 6.0 and 7.4, with their maxima around 0.48 and 0.96 nm^{-1} , may correspond

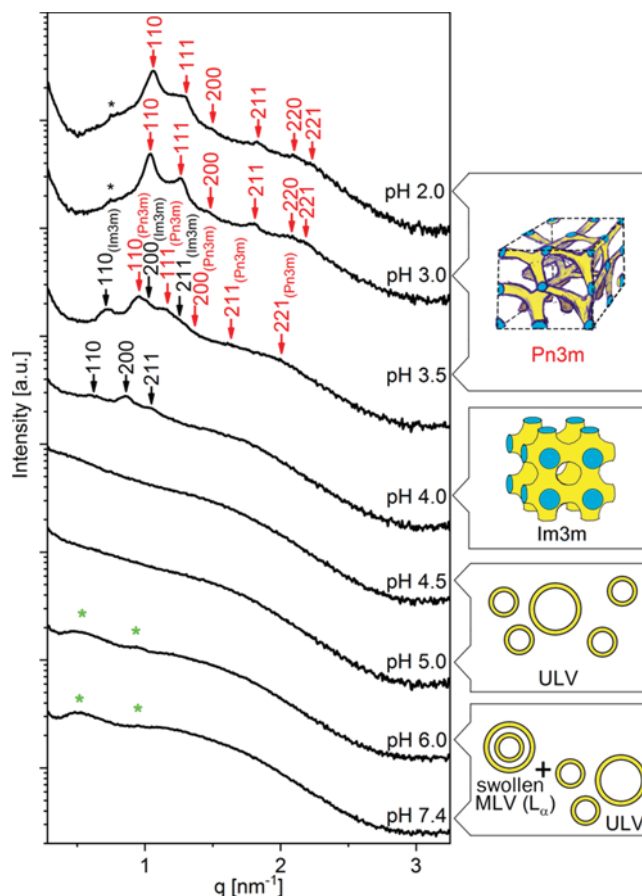


Figure 4. SAXS patterns of the F127-stabilized nanoself-assemblies prepared at 2OHOA/GMO mass ratio of 3/7 and pH in the range of 2.0–7.4. The Bragg peaks of the cubic $Pn3m$ and $Im3m$ phases and their corresponding Miller indices are indicated with red and black arrows, respectively. Low intensity peaks potentially from a coexisting $Im3m$ phase are marked with black asterisks at pH of 2.0 and 3.0. Broad correlation peaks, most likely corresponding to the weak bilayer correlations in coexisting swollen MLVs, are marked with green asterisks at pH 6.0 and 7.4.

to the weak bilayer correlations of a coexisting population of highly swollen MLVs with an estimated d of around 13 nm in agreement with the discussions above.

The structural transformations of the 2OHOA/GMO nano-self-assemblies upon increasing pH from 2.0 to 7.4 are attributed to the charge repulsions between the gradually deprotonating carboxylic groups of 2OHOA embedded at the lipid–water interface. Such repulsions would increase the effective headgroup area of the embedded 2OHOA molecules at the lipid–water interface, modifying its spontaneous curvature. Figure 5 summarizes the pH-induced phase transitions and changes in the lattice parameters for all phases detected on increasing pH from 2.0 to 4.5 for both 2OHOA/GMO samples prepared at mass ratios of 3/7 and 1/1. The cubic $Im3m$ phases in both 2OHOA/GMO dispersions summarized in Table S1 have smaller lattice parameters at pH 3.0 and 3.5 than pristine GMO cubosomes with $a_{Im3m} = 13.1$ nm around the same pH (see Figure S1). This is consistent with the integration of 2OHOA into the hydrophobic domains of the structure at this pH value. On the other hand, upon reaching pH 4.0 and 4.5 in dispersions with 2OHOA/GMO mass ratios of 3/7 and 1/1, respectively, the a_{Im3m} was found to be greater than that of GMO cubosomes in

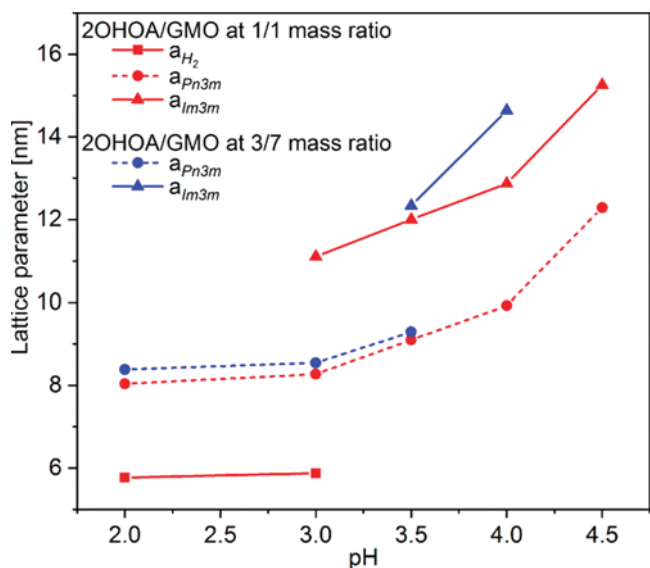


Figure 5. pH-dependent changes in the lattice parameters of the H_2 (■), $Pn3m$ (●), and $Im3m$ (▲) phases as derived from SAXS patterns of the two samples prepared at 1/1 (red) and 3/7 (blue) 2OHOA/GMO mass ratios presented in Figures 2 and 4. For numerical values see Table S1.

absence of 2OHOA (Figure S1). The later dispersion, measured as control, showed only a slight structural alteration in response to pH change between 3.0 and 7.0. The increase in a_{Im3m} from 13.1 nm at pH 3.0 to 13.5 nm at pH 7.0 may be attributed to the potential presence of trace concentrations of OA as a hydrolysis product in the sample.⁶⁸ However, this change in lattice parameter in the 2OHOA free cubosomes is insignificant compared to that in the 2OHOA/GMO system reported above. The larger pH-induced increase in the lattice parameters of the internal liquid crystalline phases of the 2OHOA/GMO nanocarriers at pH 4.0–4.5, as compared to that at pH 3.0–3.5, reflected a higher degree of 2OHOA deprotonation and indicated the closer proximity to the apparent pK_a (pK_a^{app}) of the embedded 2OHOA (Table S1). Upon increasing the pH to >5.0 , the 2OHOA/GMO cubosomes eventually transformed into vesicles.

The pH effect on the ζ -potential of 2OHOA/GMO nano-self-assemblies is demonstrated in Figure 6. At pH 2.0, the measurements for the dispersions with 1/1 and 3/7 2OHOA/GMO mass ratios indicated the occurrence of rather neutral surfaces with ζ -potential values of -0.4 ± 0.7 and -3.8 ± 2.9 mV, respectively. The ζ -potential of these dispersions was observed to gradually decrease upon increasing pH, with the largest drop in ζ -potential occurring around pH 6.0, and reaching -30.9 ± 4.8 and -29.1 ± 3.5 mV at pH 9.0 for the samples with 1/1 and 3/7 2OHOA/GMO mass ratios, respectively. This decrease in ζ -potential is mostly caused by the deprotonation of 2OHOA, and correlated well with the discussed nanostructural transitions observed by SAXS and cryo-TEM.

For the system at 2OHOA/GMO mass ratio of 1/1, the decrease in the ζ -potential from -5.8 ± 2.8 to -31.3 ± 1.7 mV on increasing pH from 5.0 to pH 7.5 could be associated with an increase in the electrostatic repulsions between the negatively charged 2OHOA/GMO bilayers in the MLVs. This charge repulsion between bilayer sheets could then induce the formation of the swollen MLVs with bilayer–

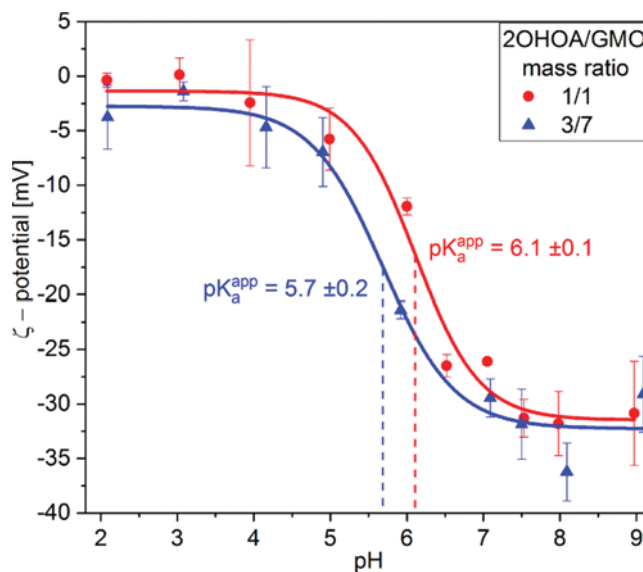


Figure 6. pH-dependent ζ -potential values for the dispersions prepared at 1/1 (red) and 3/7 (blue) 2OHOA/GMO mass ratios. Fits of eq S5 to the data (see S1) are represented as lines and used to estimate the apparent pK_a of 2OHOA in the samples prepared at two different lipid compositions.

bilayer distances above 10 nm at pH 6.0 and 7.4, as reported above.

The pK_a^{app} values of 2OHOA in the nano-self-assemblies at 2OHOA/GMO mass ratios of 1/1 and 3/7 were calculated from the pH-dependent ζ -potential values (see Figure 6 and S1). The resulting pK_a^{app} values are 6.1 ± 0.1 and 5.7 ± 0.2 , respectively. These pK_a^{app} values are slightly larger than the previously reported pK_a^{app} of 5.4 for 2OHOA in Triton X-100 micelles.⁶⁴ This may be caused by the different 2OHOA content in the 2OHOA/GMO nano-self-assemblies (2OHOA/lipid mass ratios of 1/1 and 3/7) as compared to a mass ratio 1/9 in 2OHOA/Triton X-100 micelles. The higher concentration of ionizable carboxylic groups on the oil–water interface increases the negative surface-charge density, which could enhance the accumulation of protons, leading to an increase in pK_a^{app} .^{52,72} The results suggest that the pK_a^{app} of 2OHOA in the designed 2OHOA/GMO nanocarriers may be controlled through changes in the lipid composition. This could potentially be used to tune the phase boundaries of these self-assemblies to desired pH ranges for specific delivery applications.

The detected pH-triggered structural transformations in this study can be of interest in the development of 2OHOA/GMO nanocarriers for tumor-targeted delivery applications. In the investigated nanocarriers, the major changes in the protonation state of 2OHOA occurred within the typical pH range of the extracellular tumor environment of pH 5.5–6.5.^{11,12} The modifications of the 2OHOA/GMO ratio in these nanocarriers was further found to modulate the pK_a^{app} of 2OHOA. Thus, the nanocarriers could be fine-tuned to obtain pH-triggered phase transitions at specific pH values by optimizing the lipid composition. Additionally, the introduction of hydrophobic additives, such as vitamin E or triglycerides, could increase the hydrophobic tail volume.^{73–78} This could also shift the phase boundaries between cubosomes and vesicles to higher pH values, even close to the physiological pH of 7.4. The reported pH-triggered nanostructural transformations could also trigger

specific interactions of the nanocarriers with the cells, similar to previous observations on antimicrobial peptide nanocarriers.⁷⁰ This would improve the targeted delivery of this anticancer agent and may focus its exposure to the acidic tumor microenvironment, minimizing off-target side-effects. Further in vitro and in vivo studies are required to determine the practical application of these nano-self-assemblies in cancer therapy.

■ CONCLUSIONS

The design and characterization of pH-responsive nanocarriers based on mixtures of 2OHOA with GMO are presented. In a concentration and pH dependent manner, 2OHOA was found to actively participate and modulate the structural features of GMO dispersion. The protonation state of the embedded 2OHOA at the lipid–water interfacial area dictated the structural and morphological characteristics of 2OHOA/GMO nanocarriers. At pH <4.0, the mostly protonated 2OHOA integrated into the hydrophobic domains of the cubosomes, shrinking their internal channel dimensions and promoting the formation of the H_2 phase. However, at pH ≥ 4.0 , the gradual deprotonation of the 2OHOA molecules on increasing pH resulted in electrostatic repulsions among its deprotonated carboxyl-headgroups at the lipid–water interface. This caused the swelling of the cubic $Pn3m$ and $Im3m$ phases and eventually, at pH ≥ 4.5 the colloidal transformation from cubosomes to vesicles.

The results contribute to the fundamental understanding of the self-assembly of pH-responsive and surfactant-like lipids. The presented experimental findings on the designed pH-responsive 2OHOA/GMO nano-self-assemblies could guide the development of advanced cancer nanomedicines for targeted delivery of chemotherapeutics.

■ ASSOCIATED CONTENT

● Supporting Information

The Supporting Information is available

Details on the critical packing parameter model, SAXS data analysis, and calculation of the pK_a^{app} from the ζ -potential data (PDF)

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Notes

The authors declare no competing financial interest.

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■ ABBREVIATIONS

2-OHOA	2-hydroxyoleic acid
GMO	glycerol monooleate
SAXS	small-angle X-ray scattering
cryo-TEM	cryogenic-transmission electron microscopy
H_2	inverted type hexagonal phase
PBS	phosphate buffer saline
MLVs	multilamellar vesicles
ULVs	unilamellar vesicles

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