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Title: UBC-Nepal Expedition: Haemoconcentration underlies the reductions in cerebral blood flow observed during acclimatization to high-altitude

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Running Title: Impact of haemoconcentration on cerebral blood flow at high-altitude

Abstract: At high-altitude, increases in haematocrit (Hct) are achieved through altitude-

induced diuresis and erythropoiesis, both of which result in increased arterial oxygen content (CaO₂). Given the impact alterations in Hct have on CaO₂, haemoconcentration has been hypothesized to partly mediate the attenuation of the initial elevation in cerebral blood flow (CBF) at high-altitude. To test this hypothesis, healthy males (n=13) ascended to 5050 m over nine days without the aid of prophylactic acclimatization medications. Following one-week of acclimatization at 5050 m, participants were haemodiluted by rapid saline infusion (2.10{plus minus}0.28 L) to return Hct towards pre-acclimatized levels. Arterial blood gases, Hct, global CBF (duplex ultrasound), and haemodynamic variables were measured following initial arrival to 5050 m, and after one-week of acclimatization at high-altitude, prior to and following the haemodilution protocol. Following one-week at 5050m, Hct increased from 42.5{plus minus}2.5 to 49.6{plus minus}2.5 % (P<0.001), and was subsequently reduced to 45.6{plus minus}2.3 % (P<0.001) following haemodilution. Global CBF decreased from 844{plus minus}160 to 619{plus minus}136 mL/min (P=0.033) following one-week of acclimatization and increased to 714{plus minus}204 mL/min (P=0.045) following haemodilution. Despite the significant changes in Hct, and thus CaO₂, cerebral oxygen delivery was unchanged at all time points. Furthermore, these observations occurred in the absence of any changes in mean arterial blood pressure, cardiac output, arterial blood pH, or oxygen saturation pre- and post-haemodilution. These data highlight the influence of Hct in the regulation of CBF and are the first to demonstrate experimentally that haemoconcentration contributes to the reduction in CBF during acclimatization to altitude.

New Findings: 1. What is the central question of this study? To evaluate the degree to which increases in haematocrit alters cerebral blood flow and cerebral oxygen delivery during acclimatization to high-altitude 2. What is the main finding and its importance? Through haemodilution, we determined that, following one week of acclimatization, the primary mechanism contributing to the cerebral blood flow acclimatization response is generated by increases in haemoglobin and haematocrit, while the remaining contribution to the cerebral blood flow acclimatization response is likely attributable to ventilatory acclimatization

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41 New Findings

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48

49 ABSTRACT

50 At high-altitude, increases in haematocrit (Hct) are achieved through altitude-induced diuresis
51 and erythropoiesis, both of which result in increased arterial oxygen content (CaO_2). Given the
52 impact alterations in Hct have on CaO_2 , haemoconcentration has been hypothesized to partly
53 mediate the attenuation of the initial elevation in cerebral blood flow (CBF) at high-altitude. To
54 test this hypothesis, healthy males ($n=13$) ascended to 5050 m over nine days without the aid of
55 prophylactic acclimatization medications. Following one-week of acclimatization at 5050 m,
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57 acclimatized levels. Arterial blood gases, Hct, global CBF (duplex ultrasound), and
58 haemodynamic variables were measured following initial arrival to 5050 m, and after one-week
59 of acclimatization at high-altitude, prior to and following the haemodilution protocol. Following
60 one-week at 5050m, Hct increased from 42.5 ± 2.5 to 49.6 ± 2.5 % ($P < 0.001$), and was
61 subsequently reduced to 45.6 ± 2.3 % ($P < 0.001$) following haemodilution. Global CBF decreased
62 from 844 ± 160 to 619 ± 136 mL/min ($P = 0.033$) following one-week of acclimatization and

63 increased to 714 ± 204 mL/min ($P=0.045$) following haemodilution. Despite the significant
64 changes in Hct, and thus CaO_2 , cerebral oxygen delivery was unchanged at all time points.
65 Furthermore, these observations occurred in the absence of any changes in mean arterial blood
66 pressure, cardiac output, arterial blood pH, or oxygen saturation pre- and post-haemodilution.
67 These data highlight the influence of Hct in the regulation of CBF and are the first to demonstrate
68 experimentally that haemoconcentration contributes to the reduction in CBF during
69 acclimatization to altitude.

70

71 INTRODUCTION

72 A reduction in the partial pressure of atmospheric oxygen results in hypoxemia, whereby
73 necessary cerebrovascular responses are required to maintain adequate cerebral oxygen delivery
74 (CDO_2). Upon ascent to high-altitude, cerebral blood flow (CBF) increases in proportion to the
75 degree of hypoxemia to compensate for the reduced arterial oxygen content (CaO_2) (Severinghaus
76 *et al.*, 1966; Ainslie & Subudhi, 2014). Within two- to three-days following initial arrival to high-
77 altitude, CBF begins to decrease, returning to sea-level values typically between 7-14 days at a
78 given altitude (Willie *et al.*, 2014), yet no mechanism(s) have been clearly ascribed to this pattern
79 of CBF changes. It is well established that hypoxic CBF regulation is driven by changes in CaO_2
80 to maintain consistent CDO_2 during arterial hypoxemia at sea-level [reviewed in: (Hoiland *et al.*,
81 2016)]. This regulation of blood flow is similar to that of the systemic circulation during exercise
82 (Roach *et al.*, 1999; Gonzalez-Alonso *et al.*, 2001), whereby flow is coupled to CaO_2 , but not the
83 partial pressure of arterial oxygen (PaO_2). Thus, the pattern of CBF changes observed at high-
84 altitude are also likely regulated by changes in CaO_2 (Ainslie & Subudhi, 2014; Hoiland *et al.*,

85 2018). The primary factors influencing CaO_2 at high-altitude, and by association CBF, are
86 changes in arterial oxygen saturation (SaO_2), hematocrit (Hct). This is in addition to the well
87 documented influence of the partial pressure of carbon dioxide (PaCO_2) and pH on CBF.
88 Alterations in cerebrovascular reactivity to CaO_2 and/or PaCO_2 may also underlie changes in CBF
89 at altitude (Lucas *et al.*, 2011; Willie *et al.*, 2015) although their contribution to CBF changes
90 during acclimatization is likely modest at best [reviewed in: (Hoiland *et al.*, 2018)].

91 Early increases in Hct occur through high-altitude induced diuresis whereby elevated
92 arterial pH stimulates bicarbonate to be excreted by the kidneys through the urine, reducing
93 plasma volume. This response partially mitigates the initial reductions in CaO_2 (Pugh, 1964; Ryan
94 *et al.*, 2014; Siebenmann *et al.*, 2017). Thereafter, following approximately one-week at altitude,
95 Hct increases further through erythropoiesis (Siebenmann *et al.*, 2017). While PaO_2 and SaO_2
96 increase and PaCO_2 decreases as a result of ventilatory acclimatization [reviewed in: (Hoiland *et*
97 *al.*, 2018)], previous research has speculated that increased Hct provides a greater input into the
98 changes in CaO_2 observed at altitude. Indeed, estimations indicate Hct may account for 60-70%
99 of the increase in CaO_2 during acclimatization, and therefore possess a commensurate influence
100 on the reduction in CBF observed across acclimatization (Hoiland *et al.*, 2018). Importantly,
101 CaO_2 regulates hypoxic vasodilation via changes in $\text{PaO}_2/\text{SaO}_2$ as well as independent of
102 alterations in $\text{PaO}_2/\text{SaO}_2$ such as when Hct is altered [reviewed in: (Hoiland *et al.*, 2016)].
103 However, this regulation of CBF by Hct appears unrelated to changes in blood viscosity [(Brown
104 & Marshall, 1985; reviewed in: Hoiland *et al* 2016)]. This understanding of the fundamental
105 importance of CaO_2 in the regulation of hypoxic cerebral vasodilation has been long standing
106 (Brown *et al.*, 1985); however, the influence of Hct on CBF during acclimatization at altitude due
107 to its relationship with CaO_2 has never been experimentally determined.

108 The aim of this study was to evaluate the degree to which Hct alters CBF and CDO₂ by
109 acutely haemodiluting subjects to pre-acclimatized Hct concentrations observed upon initial
110 arrival to 5050 m. We hypothesized that following approximately seven days of acclimatization at
111 5050 m above sea-level, CBF would be significantly decreased from initial arrival values
112 concomitant with an increase in Hct. Further, we hypothesized that following haemodilution, due
113 to the experimentally isolated reduction in Hct, CBF would concomitantly increase and thus be
114 partially restored to near pre-acclimatized levels.

115

116 **METHODS**

117

118 *Ethical approval*

119 All participants gave written informed consent prior to participating. This study was
120 approved by the University of British Columbia Clinical Research Ethics Board, the Nepal Health
121 Research Council, and conformed to the standards set by the *Declaration of Helsinki* (except
122 registry in a database) and the Canadian Government Tri-council Policy Statement (TCPS2) for
123 integrity in research.

124

125 *Participants*

126 Thirteen healthy male participants (BMI: $23.2 \pm 2.1 \text{ kg} \cdot \text{m}^{-2}$, Age: 27 ± 5 years) were
127 recruited to participate in this study. Participants were recruited at the University of British
128 Columbia's Okanagan campus and were part of the research team. This project was part of a
129 larger expedition to the EV-K2-CNR Italian Pyramid Research laboratory in the Khumbu Valley,

130 Nepal at 5050 m that took place September to November 2016 (Willie *et al.*, 2018). Although
131 participants were recruited to other projects at the Pyramid Laboratory, all arrived to high-altitude
132 at the same time and were tested >5 half-lives following any drug interventions. Further, all
133 participants abstained from exercise, caffeine, and alcohol for 24 hours prior to testing and
134 arrived having fasted for >4 hours. All participants were born and lived at or near sea-level and
135 were free of cardiovascular, respiratory and neurological diseases and were non-smokers. One
136 participant was taking Pentasa®, a medication used to treat inflammatory bowel disease,
137 throughout the entire study.

138

139 *Experimental Overview*

140 The research team travelled to Nepal to begin the ascent to high-altitude. All participants
141 spent 3-9 days in Kathmandu (1400 m) prior to flying to Lukla (2860 m) to begin the trek to the
142 EV-K2-CNR Pyramid Research Laboratory (5050 m). Ascent to the Pyramid Laboratory took
143 place over a slow and safe 9-day trek without the use of any acute mountain sickness
144 prophylactics
145 (e.g., acetazolamide). Participants spent one night in Monjo (2800 m), three nights in Namche
146 Bazaar (3400 m), one night in Deboche (3820 m), and then three nights in Pheriche (4371 m)
147 followed by the final trekking day to the Pyramid Laboratory (5050 m). Due to the length of
148 testing per participant, and hours available for testing at the remote laboratory, testing was limited
149 to two participants per day. As such, participants underwent the haemodilution protocol across a
150 small range of relative acclimatization with testing conducted on days 4-10 (avg. 7) at the
151 Pyramid Laboratory (**Figure 1**). For more details on the ascent see (Willie *et al.*, 2018).

152 The day following initial arrival to the pyramid lab (i.e. day 1), subjects lay supine for 20
153 minutes of quiet rest, after which CBF, echocardiography and arterial blood samples were
154 acquired. A minimum of 24 hours prior to the main experimental protocol participants' blood
155 volumes were measured using the Carbon Monoxide Rebreathe technique (Schmidt & Prommer,
156 2005). This protocol is detailed in the following section (*Experimental Measures*). The following
157 day, participants completed the experimental protocol prior to and following haemodilution. After
158 20 minutes of quiet supine rest participants were cannulated, following which, CBF, cardiac
159 output (Q), and radial arterial blood measures were acquired. A hypervolemic haemodilution
160 protocol was performed in an attempt to return Hct and plasma volume to pre-acclimatized values
161 observed upon initial arrival at high-altitude (i.e., Hct \approx 45 %) through rapid saline infusion (0.9
162 % NaCl). Prior to expedition testing, pilot work was conducted to determine the adequate
163 infusion volume of saline required to elicit the desired decrease in Hct. A bolus infusion of saline
164 equal to between 30-35 % of the participants total blood volume (avg 2.10 ± 0.28 L) was infused
165 over 0.5-1 hr.

166

167 *Experimental Measures*

168 A radial artery catheter (20-gauge; Arrow, Markham, ON, Canada) was inserted into the
169 left radial artery under local anesthesia (Lidocaine, 1.0%) and ultrasound guidance using sterile
170 technique. The arterial catheter was attached to an inline waste-less sampling system (Edwards
171 Lifesciences, VAMP, CA, USA) and a pressure transducer that was placed at the height of the
172 right atrium (TruWave transducer). Following cannulation, subjects rested quietly for 20-minutes,
173 after which an arterial blood sample was taken, and analyzed (ABL90 FLEX, Radiometer,
174 Copenhagen, Denmark).

175 Internal carotid artery (ICA) and vertebral artery (VA) image acquisition was obtained
176 using a 10 MHz multi-frequency linear array probe attached to a high-resolution ultrasound
177 machine (15L4, Terason t3200, Burlington, MA, USA). Arterial diameter was measured with B-
178 mode imaging while pulse-wave mode simultaneously measured blood velocity within the vessel.
179 Image location was selected on an individual basis in order to ensure clear, highly reproducible
180 measures with VA imaging occurring between cervical vertebrae 4 and 5, 5 and 6 or proximal to
181 entry into the vertebral column and ICA imaging taken ≥ 1.5 cm distal to the common carotid
182 bifurcation. The sum of both ICA and VA flow multiplied by two was used to estimate global
183 cerebral blood flow (gCBF). Cerebrovascular reactivity to CaO_2 was also assessed using the
184 concurrent ultrasound blood flow values and arterial blood samples. CaO_2 was calculated from
185 the arterial blood sample utilizing the arterial SpO_2 , PaO_2 , and [Hb] ($\text{CaO}_2 = (1.34 \times [\text{Hb}] \times \text{SaO}_2)$
186 $+ (0.003 \times \text{PaO}_2)$).

187 Upon initial arrival (day 1), blood pressure was assessed using an automated blood
188 pressure cuff (HEM775CAN, Omron Healthcare). During the haemodilution protocol, blood
189 pressure was assessed continuously by means of intra-radial pressure through the radial arterial
190 catheter and was corrected to manual brachial blood pressure measurements (i.e. auscultation). A
191 portable ultrasound (Vivid Q, GE Healthcare, Piscataway, NJ, USA) was used for the
192 echocardiography assessment of cardiac output (Q). Participants were rolled into the left lateral
193 decubitus position whereby a trained cardiac sonographer acquired parasternal and apical images.
194 Stroke volume (SV) was calculated from the diameter of the left ventricular outflow tract
195 diameter and the velocity-time integral from a five-chamber view (Lang *et al.*, 2015). Reliability
196 values for a range of echocardiographic variables from our group have previously been reported

197 (Stembridge *et al.*, 2014). Heart rate was obtained from the R-R intervals recorded from a three
198 lead ECG and multiplied by SV to calculate Q.

199

200 *Assessment of total blood volume*

201 Total blood volume was determined using the previously validated carbon monoxide
202 rebreathing method, as described in detail elsewhere (Schmidt & Prommer, 2005). Using a
203 custom glass spirometer (Blood Tec) and a 5-liter reservoir bag of 100 % O₂, participants were
204 instructed to maximally inhale a specific volume of carbon monoxide (0.8 mL · kg⁻¹) which was
205 inserted into the apparatus mid inspiration. Participants then held their breath at maximal
206 inspiration for 10 seconds before rebreathing on the apparatus for two-minutes. A venous blood
207 sample was obtained at baseline and seven minutes following onset of the carbon monoxide
208 rebreathing to determine total haemoglobin [Hb] and carboxyhaemoglobin (ABL 90;
209 Radiometer). Expired carbon monoxide was measured using a portable carbon monoxide analyzer
210 (Dräger Pac 3500; Draeger Safety) at baseline and at four- and seven-minutes post rebreath
211 onset. Total red cell volume assessed by this measure was used to accurately calculate total blood
212 volume pre- and post- haemodilution in conjunction with Hct %. Blood volume post
213 haemodilution was calculated using the initial blood volume, and Hct % recorded from arterial
214 blood samples pre- and post- haemodilution (i.e. $Hct_{Initial} \times \text{blood volume}_{Initial} = Hct_{Final} \times \text{blood}$
215 volume_{Final}).

216

217 *Data and Statistical Analyses*

218 Statistical analyses were performed using IBM SPSS 24. All data is reported as mean \pm
219 standard deviation. Linear mixed effects models, with repeated measures compound symmetry
220 covariance structures, were used to determine significant changes in CBF, arterial blood gases,
221 and cardiovascular variables from initial arrival and pre- vs post-haemodilution (fixed factor:
222 time, random factor: subjects). Estimated CBF was calculated using the slope of the response
223 between gCBF and Hct, pre- to post-haemodilution, multiplied by the change in Hct required to
224 elicit a full reversal of haemoconcentration to initial arrival levels in individual participants. The
225 “estimated” CBF was compared to initial arrival using a paired t-test. A linear mixed effects
226 model analysis with repeated measures (compound symmetry covariance structure) was used to
227 determine cerebrovascular reactivity to changes in CaO_2 during the acclimatization process (day 1
228 to pre hemo; condition 1) and during haemodilution (pre hemo to post hemo; condition 2). The
229 dependent variable was gCBF, while CaO_2 and condition were fixed effects. Subjects were
230 included as a random effect and MAP and PaCO_2 were added as co-variates, which improved the
231 model fit (-2 Log Likelihood). This was repeated for ICA and VA flow as well. Relative changes
232 in ICA and VA flow and reactivity were compared using a t-test following acclimatization and
233 haemodilution to examine regional CBF differences. Finally, linear regression analysis was
234 performed between both ΔgCBF and gCBF reactivity versus days spent at altitude to delineate the
235 effect of testing day. Significance was set at $P < 0.05$ for all statistical analyses.

236

237 RESULTS

238 *Arterial blood content changes*

239 Arterial blood metrics are presented in **Table 1**. Following one-week of acclimatization at
240 5050 m, [Hb] and Hct were elevated and then subsequently decreased following haemodilution.
241 However, [Hb] ($P<0.01$) and, Hct ($P<0.001$), were not reduced to their initial arrival levels and
242 remained slightly elevated following haemodilution. Both PaO_2 and SaO_2 increased following
243 acclimatization (SaO_2 , $P<0.01$; PaO_2 , $P<0.001$) and remained unchanged following
244 haemodilution (SaO_2 , $P=0.78$; PaO_2 , $P=0.79$). Though PaCO_2 significantly decreased following
245 acclimatization and following the haemodilution protocol, pH was only altered (increased)
246 following acclimatization, but remaining unchanged with haemodilution ($P=0.39$).

247

248 *Cardiovascular variables*

249 Resting cardiovascular data are presented in **Table 2**. Following acclimatization mean
250 arterial pressure (MAP) was elevated ($P=0.03$), but did not increase further with haemodilution
251 ($P=0.32$). Following haemodilution MAP was higher compared to initial arrival ($P<0.01$). Heart
252 rate did not decrease following acclimatization ($P=0.29$), but was reduced following
253 haemodilution compared to initial arrival ($P=0.03$). Notably, heart rate did not change pre- to
254 post-haemodilution ($P=0.15$). Stroke volume decreased following acclimatization ($P<0.01$) and
255 then increased following hemodilution ($P=0.02$). Following haemodilution, SV was not
256 *statistically* different compared to initial arrival ($P=0.051$). These changes ultimately led to a
257 reduction in Q following acclimatization ($P<0.01$), whereby Q remained below initial arrival

258 values following haemodilution ($P=0.01$). However, Q was not altered following haemodilution
259 ($P=0.62$).

260

261 *CBF Results*

262 Cerebrovascular results are presented in **Figure 2**. Following approximately one-week at
263 5050 m gCBF decreased by $20.9 \pm 23.6\%$ from 843.8 ± 160.2 to 619.6 ± 135.9 mL \cdot min⁻¹
264 ($P<0.001$). This decrease was mediated by reductions in blood flow through both the ICA and VA
265 ($-14.2 \pm 23.2\%$; $P<0.01$ and $-22.3 \pm 33.2\%$; $P<0.01$, respectively) (**Figure 2**). Following the
266 haemodilution trial, gCBF was elevated by $15.3 \pm 18.8\%$ to 714.1 ± 203.5 mL \cdot min⁻¹ ($P=0.045$)
267 due to an increase in ICA blood flow ($+15.7 \pm 19.7\%$; $P=0.03$) while blood flow in the VA only
268 increased marginally and non-significantly ($+15.5 \pm 28.4\%$; $P=0.305$). Post haemodilution, gCBF
269 remained significantly lower compared to initial arrival at high-altitude ($P=0.033$). At each time
270 point, gCBF was regulated to the extent that CDO₂ was unaltered across acclimatization ($126.2 \pm$
271 25.5 vs. 114.4 ± 22.7 mL \cdot min⁻¹) and hemodilution (121.6 ± 30.2 mL \cdot min⁻¹; main effect:
272 $P=0.302$). No regional differences were observed between the relative changes in ICA and VA
273 flow following acclimatization (-14.20 ± 23.15 vs -22.34 ± 33.16 %; $P=0.10$) or following
274 haemodilution (15.72 ± 19.67 vs 15.52 ± 28.36 %; $P=0.98$). Linear regression analysis indicated
275 there was no relationship between Δ gCBF and day of testing ($R^2=0.191$; $P=0.135$).

276

277 *Blood volume*

278 Blood volume data prior to and following haemodilution are presented in **Table 3**. Pre
279 haemodilution, individual blood volumes ranged from 5.47 to 7.75 L with a mean of 6.18 ± 0.83

280 L. Of this, red cell volume accounted for 3.07 ± 0.49 L while plasma volume accounted for 3.11
281 ± 0.40 L. Assuming blood volume and hematocrit were relatively unchanged within 24 hrs
282 following the carbon monoxide rebreath assessment, blood volume post haemodilution was
283 calculated based on the pre-haemodilution blood data. From these calculations, the mean increase
284 in plasma volume and thus total blood volume observed following saline infusion was 0.54 ± 0.14
285 L. Notably, hydration status, as assessed by arterial blood osmolarity was not different pre- to
286 post-haemodilution (288.14 ± 2.21 vs 289.00 ± 2.11 mmol \cdot Kg⁻¹) and falls within euhydration
287 ranges (Armstrong et al., 2010).

288

289 *Mathematical estimation of CBF with a fully normalized Hct*

290 Despite the large saline infusion utilized (2.10 ± 0.28 L), the haemodilution protocol
291 failed to fully revert Hct to initial arrival values (**Table 1**) and estimations based on total Hb mass
292 and Hct concentration indicate on average 540 mL of saline remained within the vasculature at
293 the time of measurement. Thus, an estimation of the CBF that would occur at a fully reversed
294 haemoconcentration was utilized. The group mean regression for Δ gCBF (mL \cdot min⁻¹) per Δ Hct
295 (%) was 22.4 mL \cdot min⁻¹ \cdot Hct⁻¹ with individual values ranging from -17.96 to 82.33 mL \cdot min⁻¹ \cdot
296 Hct⁻¹. Utilizing these reactivity slopes on an individual subject basis, we calculated that CBF
297 would have increased to 790.5 mL \cdot min⁻¹ had Hct been reduced by an additional 3.1% and fully
298 restored to the initial arrival value of ~42.5 %. Cumulatively, addition of this theoretical change
299 in Hct leads to an overall 165.46 ml \cdot min⁻¹ increase in gCBF from pre- to post-hemodilution.
300 This is inclusive of the experimentally elicited increase and additional estimated change (**Figure**
301 **2**). While accounting for the change in CBF with full normalization of Hct statistically eliminates

302 the difference between initial arrival gCBF and that of the theoretical post-hemo gCBF ($843.8 \pm$
 303 160.2 vs. $790.5 \pm 266.8 \text{ mL} \cdot \text{min}^{-1}$; $P=0.62$), this increase in gCBF is not sufficient to explain the
 304 entire CBF response during acclimatization, which was a $224.13 \text{ ml} \cdot \text{min}^{-1}$ decrease. Indeed,
 305 these changes in gCBF are significantly different (224.1 ± 189.2 vs. $165.5 \pm 212.1 \text{ mL} \cdot \text{min}^{-1}$;
 306 $P=0.02$).

307

308 *CBF reactivity to CaO₂*

309 Reactivity values from the linear mixed effects modelling are reported as mean \pm SD
 310 (**Figure 3**). Following one-week of acclimatization and haemodilution gCBF reactivity was
 311 unchanged (-58.7 ± 51.2 vs $-64.5 \pm 56.2 [\Delta(\text{mL} \cdot \text{min}^{-1} \cdot \text{mL}^{-1} \cdot \text{dL}^{-1})]$ $P=0.72$). Additionally there
 312 was no change in ICA reactivity to CaO₂ (-18.1 ± 18.7 vs $-22.9 \pm 20.2 [\Delta(\text{mL} \cdot \text{min}^{-1} \cdot \text{mL}^{-1} \cdot \text{dL}^{-1})]$;
 313 $P=0.43$) or VA reactivity (-10.3 ± 12.6 vs $-8.0 \pm 14.1 [\Delta(\text{mL} \cdot \text{min}^{-1} \cdot \text{mL}^{-1} \cdot \text{dL}^{-1})]$;
 314 $P=0.56$) between acclimatization and haemodilution. Additionally, there were no differences detected in
 315 relative ICA and VA reactivity to CaO₂ (9.87 ± 6.49 and $9.11 \pm 5.43 [\Delta(\% \cdot \text{mL}^{-1} \cdot \text{dL}^{-1})]$,
 316 respectively) following haemodilution ($P=0.97$). No regional differences between relative ICA
 317 and VA reactivity to CaO₂ were observed following acclimatization (-1.80 ± 22.14 vs $-9.18 \pm$
 318 $29.43 [\Delta(\% \cdot \text{mL}^{-1} \cdot \text{dL}^{-1})]$; $P=0.07$) or haemodilution (-1.08 ± 29.58 vs $9.05 \pm 15.33 [\Delta(\% \cdot \text{mL}^{-1}$
 319 $\cdot \text{dL}^{-1})]$; $P=0.23$). Linear regression analysis indicated there was no relationship between gCBF
 320 reactivity and day of testing ($R^2=0.209$; $P=0.116$).

321

322

323 DISCUSSION

324 This study aimed to mechanistically examine the degree to which haemoconcentration at
325 high-altitude contributes to changes in CBF during acclimatization. The main findings of this
326 study were that; 1) Similar to previous studies, elevations in CaO_2 through increases in Hct, as
327 well as PaO_2 , occurred following approximately one-week at high-altitude, resulting in a
328 reduction in CBF but maintained CDO_2 (Severinghaus *et al.*, 1966; Huang *et al.*, 1987; Jensen *et*
329 *al.*, 1990; Baumgartner *et al.*, 1994; Lucas *et al.*, 2011; Willie *et al.*, 2014; Subudhi *et al.*, 2014);
330 2) Following haemodilution, an absolute reduction in Hct by ~4 % resulted in a significant
331 increase in gCBF; however, this response was blunted compared to that during acclimatization
332 where CBF did not return to initial arrival values. Further, changes in CBF following
333 haemodilution and mathematical extension of this response indicate that haemoconcentration is
334 responsible for ~74% of the decrease in CBF observed during acclimatization to high-altitude.
335 This finding is further supported when the tight coupling of CBF to CaO_2 is considered (Brown *et*
336 *al.*, 1985). Indeed, CBF both during acclimatization and following haemodilution appears to be
337 tightly dictated by changes in CaO_2 . These findings imply that though ventilatory acclimatization
338 and elevations in haematocrit occur concomitantly during acclimatization, the
339 haemoconcentration response contributes to the CBF response during acclimatization to a greater
340 degree at high-altitude.

341

342 *Cerebral Blood Flow*

343 Increased CBF in response to a reduction in CaO_2 has been observed in previous studies,
344 which demonstrates the brain's ability to maintain CDO_2 during hypoxemia (Ainslie *et al.*, 2014).

345 This relationship is further evidenced by the reduction in CBF that occurs concomitant to
346 increased CaO_2 with acclimatization to high-altitude [reviewed in: (Ainslie & Subudhi, 2014)].
347 Therefore, it appears that CBF is regulated to maintain convective oxygen delivery to the brain.
348 However, the relative contribution that haemoconcentration has on the reduction in CBF observed
349 across acclimatization had not yet been investigated.

350 Notably, due to the rapid diffusion of saline out of the intravascular space (Greenfield *et*
351 *al.*, 1989), Hct was not fully reversed to values observed upon initial arrival to high-altitude.
352 However, as Hct has a linear inverse relationship with CBF through its relationship with CaO_2
353 (Brown & Marshall, 1985; Ainslie *et al.*, 2014), the fully reversed response can be calculated by
354 extrapolating this linear inverse relationship (**Figure 2**). Following mathematical correction to
355 estimate the influence of a full reversal of haemoconcentration (see “*Mathematical estimation of*
356 *CBF with a fully normalized Hct*”), gCBF did not completely return to initial arrival values
357 despite the lack of statistical difference between pre-acclimatization and the theoretical post-
358 haemodilution CBF. The increase in gCBF following haemodilution (and mathematical extension
359 of this increase) explained 74% of the reduction in gCBF observed following acclimatization.
360 This suggests that the majority of the reduction in gCBF following one-week of acclimatization is
361 mediated through increases in [Hb] and Hct, and their subsequent influence on CaO_2 . This is in
362 agreement with recent estimations made by our group (Hoiland *et al.*, 2018).

363 Cerebrovascular hypoxic reactivity did not differ between acclimatization and
364 haemodilution, highlighting previous findings demonstrating that CBF is primarily governed by
365 changes in CaO_2 and suggests that the difference in the CBF responses between acclimatization
366 and haemodilution are not due to alterations in hypoxic reactivity. Therefore, it stands to reason
367 that the difference between initial arrival CBF and following haemodilution (and correction for

368 full restoration of Hct) is primarily a result of ventilatory acclimatization whereby SaO₂, PaO₂,
369 and pH are also elevated from initial arrival at high-altitude. Indeed, SaO₂ and PaO₂ were 5.0%
370 and 6.1 mmHg higher following acclimatization, respectively, while pH was increased by 0.03
371 units (**Table 1**). Given SaO₂, PaO₂, and pH were unaltered during haemodilution, their influence
372 on cerebral vasomotor tone would have been unaltered and likely explains the difference between
373 initial arrival and post-haemodilution CBF.

374 Differences in regional CBF have been noted at high-altitude and in hypoxia with an
375 apparent preference of flow to the posterior regions of the brain which houses the primary centers
376 for regulating physiological function upon arrival to high-altitude (Subudhi *et al.*, 2014;
377 Feddersen *et al.*, 2015) and during exposure to normobaric hypoxia (Willie *et al.*, 2012). Of note,
378 we did not observe any significant regional differences between relative changes in flow
379 following acclimatization or haemodilution. Further, no differences were observed in the hypoxic
380 reactivity between the ICA and VA. This finding agrees with previous work from our group
381 conducted at the same altitude in subjects using acetazolamide (Willie *et al.*, 2014). Differences
382 between these findings and previous research showing regional flow disparities may be related to
383 the methodology of data collection, primarily the blood flow measurement (transcranial Doppler
384 ultrasound vs duplex ultrasound) and the severity and mode of exposure to the hypoxic stimulus
385 (Hoiland *et al.*, 2018; Willie *et al.*, 2018) whereby a more severe step change or exposure may
386 necessitate regional blood flow prioritization to the posterior circulation as a form of survival
387 response. Further, an important consideration is that while these regional differences appear to be
388 more prevalent at highly localized levels (Binks *et al.*, 2008; Lawley *et al.*, 2017), bulk flow
389 measures at the VA and ICA may fail to detect these differences in some studies given that flow
390 measures at these sites (VA & ICA) represent the summation of multiple discrete brain regions.

391 Further, differences in reactivity that have been noted at high-altitude are thus likely not
392 attributable to changes in Hct based on our results.

393 Following acclimatization, PaO_2 and SaO_2 increased through well documented
394 mechanisms of ventilatory acclimatization [reviewed in; (Hoiland *et al.*, 2018)]. Both were
395 unaltered following haemodilution, isolating any effects of reversing ventilatory acclimatization
396 induced blood gas changes from our CBF changes attributed to acclimatization/Hct. Thus, the
397 persisting influence of SaO_2 and PaO_2 may represent the remaining stimulus for lower CBF post
398 haemodilution compared to initial arrival. Further, while pH was unaltered following
399 haemodilution, both pre- and post-haemodilution pH were higher than initial arrival. This may
400 also be driving a reduction in CBF and indicate our results may in fact be underestimating (albeit
401 modestly) the influence of haemoconcentration on CBF at altitude.

402

403

404 **Methodological considerations**

405

406 *Effect of viscosity on CBF*

407 Alterations in Hct occurring both chronically during acclimatization (increases) and
408 acutely during haemodilution (decreases) will lead to concomitant changes in whole blood
409 viscosity, which may subsequently affect flow through the vasculature. However, experimental
410 study has to date refuted this [(Brown & Marshall, 1985) reviewed in (Hoiland *et al.*, 2016)].
411 Indeed, a reduction in blood viscosity through plasma exchange in patients with paraproteinemia,
412 whereby CaO_2 and PaCO_2 were constant, does not alter CBF (Brown & Marshall, 1985). While at

413 odds with basic physical principles (i.e., Poiseuille's Law), it is important to consider that the
414 cerebral vasculature is a complex network of compliant vessels conveying a non-newtonian fluid.
415 Thus, the conditions do not reflect those in which Poiseuille's Law was defined. Further,
416 alterations in blood viscosity can influence vascular paracrine signalling. To speculate, reductions
417 in blood viscosity would reduce the direct resistive effects of blood flow through the vessel,
418 however, the shear stress stimuli induced by viscosity would also be reduced, potentially limiting
419 dilation through reduced stimulation of shear dependent pathways (Melkumyants, Balashov, &
420 Khayutin, 1989). This would act to increase vascular resistance. Thus, it is likely that a balance
421 exists between these two stimuli, possibly explaining why viscosity has been previously shown to
422 have a negligible effect on CBF (Brown & Marshall, 1985). Indeed, that haemodilution leads to
423 greater increases in CBF than blood flow to other vascular beds (Crystal & Salem, 2002; Van
424 Bommel et al., 2002) indicate the increases in CBF observed during haemodilution reflect active
425 vascular regulation.

426

427 *Effect of PaCO₂ and pH*

428 As expected, PaCO₂ was significantly reduced following one-week of acclimatization at
429 high-altitude due to ventilatory acclimatization (Rahn & Otis, 1949). However, we also observed
430 a 2.6 mmHg reduction in PaCO₂ following haemodilution, though both PaO₂ and SaO₂ were
431 unchanged. This decrease in PaCO₂ in the presence of unchanged PaO₂ and SaO₂ has been
432 reported in a previous saline infusion study (Prisk *et al.*, 2010) and is likely due to the dilutional
433 acidosis effect of the saline solution (Muir *et al.*, 1975). Notably, at least at sea level, PaCO₂ has

434 been shown to effect ICA and VA flow by ~6-8 % per 1 mmHg change in PaCO₂ (Willie *et al.*,
435 2012;
436 Hoiland *et al.*, 2015). These observations indicate that, upon correction for this small alteration in
437 PaCO₂, the CBF response would be greater following haemodilution, suggesting that the
438 contribution of Hct on the CBF response would be more substantial. However, as PaCO₂
439 primarily alters CBF through changes in arterial pH - which notably was not different pre to post
440 haemodilution - the difference in PaCO₂ pre-to post-haemodilution likely had a negligible effect.

441

442 *Blood volume expansion*

443 Due to the logistical constraints and remote nature of the high-altitude expedition, a
444 hypervolemic hemodilution protocol using saline infusion was utilized as opposed to a
445 normovolemic haemodilution protocol in which blood volume would be maintained. However,
446 this may be most appropriate as the initial increases in Hct observed at altitude are due to a
447 reduced plasma volume (Siebenmann *et al.*, 2015), thus our intervention manipulated Hct in the
448 same manner as the environmental stress of high-altitude hypoxia. Though the volume of fluid
449 utilized in this protocol was relatively large in relation to the overall blood volume of
450 participants, the crystalloid properties of the fluid infused and the time at rest between measures
451 resulted in a large portion of the infused saline leaving the vasculature (Greenfield *et al.*, 1989).
452 This is primarily evidenced by the relatively marginal reduction in hematocrit compared to
453 volume infused and blood volume data which indicates only 540 mL of saline remained within
454 the vasculature at the time of CBF measurement. Further, if increases in blood volume resulted in
455 alterations in CBF, this would be expected to coincide with alterations in cardiac parameters, (i.e.

456 increased Q). Of note, neither of these variables were significantly elevated by hemodilution
457 suggesting there was no direct impact of volume expansion on CBF changes in this study.

458

459

460 **CONCLUSIONS**

461 This study was the first to experimentally investigate the degree to which CBF
462 acclimatization is driven by haemoconcentration. Through haemodilution, we were able to
463 determine that, following one-week of acclimatization, the primary mechanism contributing to the
464 CBF response during acclimatization response is generated by diuresis and erythropoiesis-
465 mediated increases in [Hb] and Hct, while the remaining contribution to the CBF response during
466 acclimatization response is likely attributable to ventilatory acclimatization.

467

468

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566

567 **AUTHOR CONTRIBUTIONS**

568 C.A.H., P.N.A., & R.L.H. conceived the study design. C.A.H., P.N.A., J.C.T., H.H.C., A.P.,
569 M.S., A.W., E.D., M.G.R., M.T., C.G., A.S., D.B.M., & R.L.H. were involved in data collection.
570 C.A.H., P.N.A., J.C.T., M.S., A.L.D., & R.L.H. were involved in data analyses and interpretation.
571 C.A.H., P.N.A., & R.L.H. drafted the manuscript. All authors critically reviewed and approved
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573

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582

583 CONFLICT OF INTEREST

584 The authors declare no conflicts of interest, financial or otherwise.

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592 TABLES

593

594 **Table 1. Arterial blood gases at high altitude.**

	Arrival at 5050m	Pre-Haemodilution	Post-Haemodilution
CaO ₂ (mL · dL)	15.4 ± 1.6	18.6 ± 1.3*	17.2 ± 1.0*†
Hct (%)	42.5 ± 2.5	49.6 ± 2.5*	45.6 ± 2.3*†
[Hb] (g · dL)	14.3 ± 0.8	16.2 ± 0.8*	14.9 ± 0.8*†

PaCO ₂ (mmHg)	29.6 ± 1.8	23.6 ± 2.1*	21.0 ± 1.5*†
pH	7.47 ± 0.02	7.50 ± 0.02*	7.49 ± 0.02*
PaO ₂ (mmHg)	40.6 ± 4.7	46.7 ± 4.2*	47.0 ± 3.4*
SaO ₂ (%)	78.3 ± 5.6	83.3 ± 4.1*	83.7 ± 3.5*

595 * significant difference from arrival at 5050m

596 † significant difference between pre- and post-haemodilution

597 **Table 2. Cardiovascular parameters upon arrival to high-altitude and pre- and post-**
 598 **haemodilution**

	Arrival at 5050m	Pre-Haemodilution	Post-Haemodilution
MAP (mmHg)	99±12	108±10*	111±9*
HR (bpm)	65±18	58±11	53±12*
SV (mL)	69±13	54±11*	63±8.8†
Q (L · min ⁻¹)	4.45±1.52	3.09±0.65*	3.28±0.65

599 * significant difference from arrival at 5050m

600 † significant difference between pre- and post-haemodilution

601 **Table 3. Individual blood volume data pre- and post-haemodilution**

<i>Subject</i>	Pre-Haemodilution				Post-Haemodilution			
	Hct (%)	Hct Volume (L)	Plasma Volume (L)	Blood Volume (L)	Hct (%)	Hct Volume (L)	Plasma Volume (L)	Blood Volume (L)
1	47.4	2.661	2.953	5.613	43.4	2.661	3.470	6.131
2	53.2	2.911	2.561	5.472	48.9	2.911	2.989	5.900
3	50.2	2.569	2.549	5.118	46.6	2.569	2.923	5.492
4	50.8	3.555	3.443	6.997	46.1	3.555	4.110	7.665
5	49	3.799	3.954	7.753	45.4	3.799	4.550	8.349
6	49	2.787	2.900	5.687	45.5	2.787	3.325	6.111
7	49.8	2.927	2.951	5.878	45.2	2.927	3.522	6.449
8	50.6	3.607	3.521	7.128	45.4	3.607	4.288	7.895
9	50.8	3.474	3.365	6.839	45.6	3.474	4.094	7.569
10	50.1	2.896	2.884	5.780	46.8	2.896	3.271	6.167
11	52.9	3.695	3.290	6.985	49.3	3.695	3.747	7.442
12	43.3	2.411	3.157	5.568	40.5	2.411	3.572	5.983
13	47	2.604	2.936	5.540	43.6	2.604	3.619	6.223
MEAN	49.5	3.069	3.113	6.181	45.6	3.069	3.652	6.721
SD	2.6	0.486	0.396	0.834	2.3	0.486	0.492	0.924

602 Individual blood volume data pre- and post-haemodilution. Plasma volume and total blood
603 volume are derived from the Hct % and total Hct volume measured using a CO re-breath test
604 performed one-day prior to the haemodilution protocol.

605 **FIGURES**

606

607

608 **Figure 1. Experimental overview of the study protocol.** Participants ascended to 5050 m over a
609 9-day period involving rapid ascent by plane to 2860 m, and ambulatory ascent onward to 5050
610 m. Cerebral blood flow (CBF), arterial blood gases (ABGs), blood pressure (BP), cardiac output
611 (Q) and heart rate (HR) were recorded at all three time points.

612

613

614 **Figure 2. Cerebral blood flow during acclimatization and following haemodilution.** At the
615 arrival time point, sample sizes were reduced for the internal carotid artery (ICA; n=11), vertebral
616 artery (VA; n=11), and global cerebral blood flow (gCBF; n=10). Data was successfully collected
617 in all participants (n=13) following one-week of acclimatization at 5050 m (pre-hemo) and
618 following hypervolemic haemodilution (post-hemo). “Estimated” data (denoted by the grey
619 background) are based on theoretical calculations for a fully normalized Hct (see “*Theoretical*
620 *calculation of CBF with a fully normalized Hct*”). Blood flow was calculated to assume a
621 complete reversal of haematocrit to initial arrival values (i.e. greater extent of haemodilution).
622 Open circles represent individual data points with dotted lines tracking within subject changes.
623 Black horizontal lines represent the average flow at each time point. * signifies a significant
624 difference from arrival at 5050m. † signifies a significant difference pre- to post- haemodilution.
625 Significance is set a
626 $P < 0.05$

627

628

629 **Figure 3. Cerebrovascular reactivity to acclimatization and haemodilution.** Reactivity to
630 changes in CaO_2 was not different across acclimatization (Initial arrival to pre-hemo) compared
631 to during haemodilution (pre-hemo to post-hemo). gCBF, global cerebral blood flow; ICA,
632 internal carotid artery; VA, vertebral artery. No differences in reactivity were observed between
633 the ICA and VA at any time point. Data are presented as mean \pm SD.

634





