

Research Article

Early Effects of Hyperbaric Oxygen on Inducible Nitric Oxide Synthase Activity/Expression in Lymphocytes of Type 1 Diabetes Patients: A Prospective Pilot Study

Ivana Resanovic ¹, Zoran Gluvcic ², Bozidarka Zaric,¹ Emina Sudar-Milovanovic ¹, Aleksandra Jovanovic ¹, Davorka Milacic,³ Radmilo Isakovic,³ and Esmar R. Isenovic ¹

¹Institute of Nuclear Sciences Vinca, University of Belgrade, Laboratory of Radiobiology and Molecular Genetics, Belgrade, Serbia

²Clinic for Internal Medicine, Zemun Clinical Hospital, School of Medicine, University of Belgrade, Belgrade, Serbia

³Department of Hyperbaric Medicine, Zemun Clinical Hospital, Belgrade, Serbia

Correspondence should be addressed to Ivana Resanovic; ivana_resanovic@yahoo.com and Esmar R. Isenovic; isenovic@yahoo.com

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This study aimed at examining the early effects of hyperbaric oxygen therapy (HBOT) on inducible nitric oxide synthase (iNOS) activity/expression in lymphocytes of type 1 diabetes mellitus (T1DM) patients. A group of 19 patients (mean age: 63 ± 2.1) with T1DM and with the peripheral arterial disease were included in this study. Patients were exposed to 10 sessions of HBOT in the duration of 1 h to 100% oxygen inhalation at 2.4 ATA. Blood samples were collected for the plasma C-reactive protein (CRP), plasma free fatty acid (FFA), serum nitrite/nitrate, and serum arginase activity measurements. Expression of iNOS and phosphorylation of p65 subunit of nuclear factor- κ B (NF κ B-p65), extracellular-regulated kinases 1/2 (ERK1/2), and protein kinase B (Akt) were examined in lymphocyte lysates by Western blot. After exposure to HBOT, plasma CRP and FFA were significantly decreased ($p < 0.001$). Protein expression of iNOS and serum nitrite/nitrate levels were decreased ($p < 0.01$), while serum arginase activity was increased ($p < 0.05$) versus before exposure to HBOT. Increased phosphorylation of NF κ B-p65 at Ser⁵³⁶ ($p < 0.05$) and decreased level of NF κ B-p65 protein ($p < 0.001$) in lymphocytes of T1DM patients were observed after HBOT. Decreased phosphorylation of ERK1/2 ($p < 0.05$) and Akt ($p < 0.05$) was detected after HBOT. Our results indicate that exposure to HBO decreased iNOS activity/expression via decreasing phosphorylation of ERK1/2 and Akt followed by decreased activity of NF κ B.

1. Introduction

Diabetes mellitus (DM) type 1 (T1DM) is a multifactorial autoimmune disease associated with significant morbidity and mortality related to microvascular and macrovascular complications [1, 2]. T1DM is associated with abnormal synthesis of nitric oxide (NO) via activation of inducible NO synthase (iNOS) [3, 4], and an increased level of iNOS enzyme is associated with DM-related vascular complications [5–8]. The iNOS gene expression is stimulated through activation of transcription factors, such as nuclear factor- κ B (NF κ B). Furthermore, induction of iNOS is mediated via stimulation of NF κ B, by different stimuli

including extracellular-regulated kinases (ERK1/2) and protein kinase B (Akt) [9, 10].

Systemic hyperbaric oxygen (HBO) therapy (HBOT) has been proposed as a medical treatment for DM patients with the developed peripheral arterial disease. HBOT is defined as therapeutic inhalation of 100% oxygen in the elevated pressure controlled conditions, which induces micro- and macrovascular hemodynamic changes [11]. An increase in oxygen arterial partial pressure in hyperbaric conditions promotes better solubility of plasma oxygen. This further results in the preservation of vitality of tissues, reversibly damaged by atherosclerosis-induced ischemia, simultaneously with microcirculation restoration [12]. Exposure

to HBO decreases iNOS activity/expression followed by reduction of NO generation, which implies that this may be mechanisms responsible for the anti-inflammatory effect of HBOT [13, 14]. However, the exact molecular mechanism by which HBOT reduces inflammation remains still unclear.

We hypothesized that HBO affects iNOS regulation through inhibition of the NF κ B activation, by a mechanism that involves phosphorylation of ERK1/2 and Akt. The aim of this prospective pilot study was twofold: (1) to investigate the early effects of HBOT on iNOS activity/expression and (2) to investigate the involvement of Akt and ERK1/2 in the regulation of iNOS activity/expression, in T1DM patients exposed to HBO.

2. Material and Methods

2.1. Chemicals and Reagents. Lymphocyte separation media (LSM), Lymphosep, was obtained from BioWest S.A.S (Nuaille, France). Protease inhibitor (Complete, Ultra, Mini, EDTA-free) and phosphatase inhibitor (PhosStop) cocktails were obtained from Roche (Mannheim, Germany). Luminol and p-coumaric acid were obtained from Sigma-Aldrich Corporation (St. Louis, MO, USA). The commercially available kit for determination of total cholesterol Beckman enzymatic reagent kit was purchased from Beckman Coulter (Brea, CA, USA). The nitrate/nitrite colourimetric assay kit was purchased from Cayman Chemical (Ann Arbor, MI, USA). For measuring the activity of arginase, we used L-arginine monohydrochloride obtained from Kemika (Zagreb, Croatia) and alpha-isonitrosopropiophenone (ISPF) obtained from Sigma-Aldrich Corporation (St. Louis, MO, USA). The polyclonal rabbit anti-iNOS and anti-NF κ B-p65, monoclonal mouse anti-actin antibody, secondary anti-mouse, and anti-rabbit IgG horseradish peroxidase-linked antibodies were obtained from Santa Cruz Biotechnology Inc. (Santa Cruz, CA, USA). The following antibodies were obtained from Cell Signaling Technology Inc. (Danvers, MA, US): anti-phospho-NF κ B-p65 (Ser⁵³⁶), anti-phospho-ERK1/2 (Thr²⁰²/Tyr²⁰⁴), anti-total ERK1/2, anti-phospho-Akt (Ser⁴⁷³), and anti-total Akt antibodies. The enhanced chemiluminescent reagent was obtained from Amersham, GE Healthcare (Buckinghamshire, UK).

2.2. Subjects and Methods. After obtaining the Ethical Committee of Zemun Clinical Hospital approval and written informed consent, 19 type 1 DM (T1DM) patients with no obvious DM ulcers, inflammatory redness on lower extremities, and contraindications for HBOT were included in this prospective pilot study during 2017, in Zemun Clinical Hospital, Zemun, Serbia. The study inclusion criterion was the presence of T1DM with the peripheral arterial disease. A total of 10 sessions of HBOT were applied as one session per day, for two weeks, in the duration of 1 h of 100% oxygen inhalation at 2.4 ATA. The demographic properties and medical history of all patients were recorded on the first visit, including gender, type of diabetes, current diabetes condition, hypertension, and other concomitant therapy of study interest (aspirin, statins, fibrates, antiplatelet therapy, angiotensin-converting enzyme inhibitors, angiotensin

receptor blockers, calcium-channel blockers, and beta blockers). The peripheral arterial disease was confirmed by the standard questionnaire. Blood samples were collected, and lymphocytes were isolated and stored at -20°C . Half of the collected blood was transferred to EDTA-containing vacutainer tubes, incubated on ice for one hour, and subsequently centrifuged for 15 min at $2000 \times g$. Obtained supernatants (plasma) were stored at -20°C before analysis. Serums were isolated by incubation of blood at room temperature for 30 min without anticoagulants, followed by 15 min centrifugation at $1800 \times g$. Serums were stored at -20°C until further biochemical analysis. Glycosylated hemoglobin A1c (HbA_{1c}), C-reactive protein (CRP), free fatty acids (FFA), nitrite/nitrate levels, and arginase activity were measured before and after completion of HBOT. Complete blood count, total cholesterol, and triglyceride (TG) were measured before HBOT.

2.3. Measurement of Serum HbA_{1c} Levels. The measurement of HbA_{1c} was carried out using an immunological method with an antibody to the beta chain of HbA_{1c} on Roche cobas c 501 analyzer (Roche Diagnostics, Indianapolis, IN, USA). The HbA_{1c} results are expressed as a percentage of glycosylated hemoglobin (%).

2.4. Measurement of Plasma TG and Total Cholesterol Concentration. The concentration of TG was measured using a commercially available kit according to the manufacturer's guidelines using a Roche cobas c 501 analyzer (Roche Diagnostics, Indianapolis, IN, USA). The concentration of total cholesterol was measured by standardized enzymatic colour test using Beckman Coulter Olympus AU400 analyzer (Brea, CA, USA) [15]. Total cholesterol and TG concentrations were expressed as mmol/L.

2.5. Measurement of Plasma CRP and FFA Concentrations. The concentration of CRP in the plasma was measured by the immunoturbidimetric method using a commercially available kit (system reagent for the quantitative determination of CRP in human plasma), following manufacturer guidelines, on Roche cobas c 501 analyzer (Roche Diagnostics, Indianapolis, IN, USA). CRP concentrations were expressed in mg/L and presented as % of CRP before HBOT.

The FFA concentration was measured in the plasma containing EDTA, using a modified version of the Duncombe method [16]. The principle of the method is that extracted FFA in chloroform, in the presence of an appropriate reagent (aqueous solution of $\text{Cu}(\text{NO}_3)_2 \times 3\text{H}_2\text{O}$ with triethanolamine, pH 7.8), forms salts of copper, which in contact with diethyldithiocarbamate builds a yellow complex compound with a maximum absorbance at 436 nm [15]. The concentrations of FFA were expressed in mmol/L and presented as % of FFA before HBOT.

2.6. Measurement of Serum Nitrite/Nitrate Concentration and Arginase Activity. The concentration of the serum nitrite and nitrate was measured by using a nitrate/nitrite colourimetric assay kit (Cayman Chemical, Ann Arbor, MI, USA) according to the manufacturer's protocol [15]. The concentrations of nitrite/nitrate were expressed in mmol/L and presented as % of nitrite/nitrate before HBOT.

Arginase activity was measured spectrophotometrically in serum, according to the method of Corraliza et al. [17]. Arginase is the enzyme that converts L-arginine to urea and L-ornithine; thereby, the principle of this method is based on measurement of the amount of generated urea which is directly proportional to arginase activity. Briefly, aliquots of serum were incubated for 10 min at 55°C in complete assay mixture lacking arginine. The reaction was initiated by addition of L-arginine, and incubation was continued at 37°C for 1 h. The reaction was terminated by heating at 100°C for 45 min. Values of arginase activity were expressed in nmol/min/mg of proteins and presented as % of arginase activity before HBOT.

2.7. Isolation of Lymphocytes from Peripheral Blood. The heparinized whole blood samples from each patient were used for the isolation of lymphocytes with LSM, according to the method of Boyum [18]. The lymphocytes forming a layer directly above the LSM were isolated and washed twice with phosphate-buffered saline. Each wash was followed by centrifugation at $1200 \times g$ for 10 min. Finally, the supernatant was removed and stored at -70°C for further analysis.

2.8. Isolation of Proteins from Lymphocytes. Whole cell extracts were prepared by suspending cell pellets in ice-cold buffer for protein isolation (150 mM sodium chloride (NaCl), 20 mM tris(hydroxymethyl)aminomethane (TRIS), 2 mM ethylenediaminetetraacetic acid (EDTA), 2 mM dithiothreitol (DTT), 1% nonionic detergent Triton X-100, and 10% glycerol) containing phosphatase and protease inhibitor cocktails. Lymphocytes were lysed by rotation at $+4^\circ\text{C}$ for a period of 1 h. Lysed samples were then centrifuged for 30 min at $14000 \times g$ at $+4^\circ\text{C}$. Protein concentration was determined by the Lowry method [19]. The supernatants were stored at -70°C for further analysis.

2.9. SDS-PAGE and Western Blotting. The lysate proteins (40 $\mu\text{g}/\text{lane}$) were separated by 10% or 12% SDS-PAGE and transferred to PVDF membranes as previously described [20]. Membranes were blocked with 5% bovine serum albumin and probed with the following antibodies: anti-iNOS, NF κ B-p65 (Ser⁵³⁶), anti-NF κ B-p65, anti-phospho-Akt (Ser⁴⁷³), anti-total Akt, anti-phospho-ERK1/2 (Thr²⁰²/Tyr²⁰⁴), and anti-total ERK1/2. The membranes were washed and incubated with the appropriate secondary HRP-conjugated antibody and after that used for detection with the enhanced chemiluminescent reagent. Anti-actin monoclonal antibodies were used as a loading control. The obtained signals were quantified using ImageJ software (NIH, USA).

2.10. Statistical Analysis. Values are expressed as means \pm SEM. SPSS 18.0 for Windows (SPSS Inc., Chicago, IL, USA) was used for all statistical calculations. Analysis of data was evaluated using two-tailed Student's *t*-test. The statistical significance level was set at $p < 0.05$.

TABLE 1: Anthropometric and metabolic parameters of T1DM patients. Results are presented as mean \pm SEM: HBOT: hyperbaric oxygen therapy; T1DM: type 1 diabetes mellitus; BMI: body mass index; HbA1c: glycosylated hemoglobin; WBC: white blood cells; RBC: red blood cells; TG: triglyceride.

Parameter	Mean \pm SE (min–max)	Reference range
Age (years)	63 \pm 2.1 (50–80)	
Weight (kg)	82.6 \pm 2.9 (62–99)	
High (m)	1.74 \pm 0.02 (1.6–1.8)	
BMI (kg/m)	27.4 \pm 0.9 (21.8–33.7)	
HbA1c (%)	8.6 \pm 0.08 (6.1–12.7)	4.8–5.9
Hemoglobin (g/L)	136 \pm 3.4 (111–169)	120–175
Hematocrit (%)	40 \pm 0.1 (30–50)	35–50
WBC ($\times 10^3/\text{L}$)	8.3 \pm 0.5 (3.4–11.3)	3.5–10.5
RBC ($\times 10^6/\text{L}$)	4.7 \pm 0.2 (3.9–6.6)	3.9–5.7
Platelets ($\times 10^3/\text{L}$)	264 \pm 26.7 (127–393)	150–450
Cholesterol (mmol/L)	5 \pm 0.4 (3.25–7.9)	5–6.2
TG (mmol/L)	2.3 \pm 0.4 (0.7–4.0)	1.7–2.2

3. Results

The anthropometric parameters including the metabolic parameters of T1DM patients at admission are presented in Table 1.

Results presented on Figure 1(a) show a significant decrease (by 67%; $p < 0.001$) of plasma CRP concentration and plasma FFA concentration (by 46%; $p < 0.001$) after exposure to HBOT (Figure 1(b)).

The serum nitrite/nitrate concentration (Figure 2(a)) was decreased by 29% (before HBOT = 100%; after HBOT = 71 \pm 8.9%; $p < 0.01$), while serum arginase activity was increased by 32% after exposure to HBOT (before HBOT = 100%; after HBOT = 132 \pm 15.1%; $p < 0.05$) (Figure 2(b)). To assess a decrease in iNOS synthesis, protein samples from cell lysates were analyzed by Western blotting (Figure 2(c)). HBOT significantly decreased the iNOS protein level (before HBOT = 1-fold; after HBOT = 0.87 \pm 0.05-fold, $p < 0.01$).

Since the promoter of iNOS gene is the binding site for NF κ B transcription factor [21], we further examined whether NF κ B is involved in HBOT downregulation of iNOS activity/expression by measuring p65 subunit level of NF κ B (Figure 3). Furthermore, we also assessed the effect of HBOT of the phosphorylation state of NF κ B-p65 at Ser⁵³⁶ by the immunoblotting with the phospho-specific NF κ B-p65 antibody that recognizes NF κ B-p65 only when phosphorylated at Ser⁵³⁶.

HBOT leads to a significant decrease of the level of the total form of NF κ B-p65 by 0.23-fold (before HBOT = 1-fold; after HBOT = 0.77 \pm 0.07-fold, $p < 0.01$) (Figure 3(a)), as well as the level of the phosphorylated form of NF κ B-p65 by 0.3-fold (before HBOT = 1-fold; after HBOT = 0.70 \pm 0.13-fold, $p < 0.05$) (Figure 3(b)). However, HBOT increased the ratio between the phosphorylated and total forms of NF κ B-p65 in the lysate of T1DM patients' lymphocytes (before HBOT = 1-fold; after HBOT = 1.42 \pm 0.17-fold, $p < 0.05$) (Figure 3(c)).

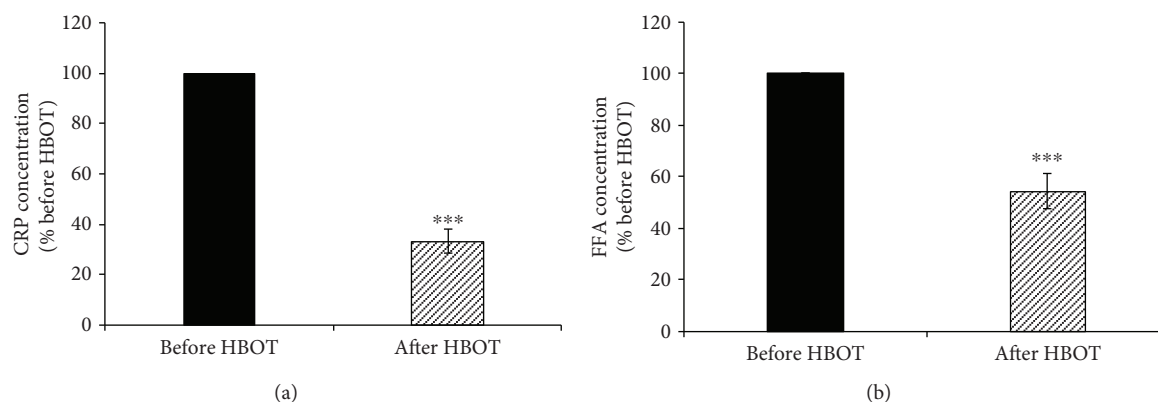


FIGURE 1: Effects of HBOT on plasma CRP and FFA levels. (a) The plasma level of CRP; (b) the plasma level of FFA. Results are expressed as % of the value obtained before HBOT and represent mean \pm SEM ($n = 15$); *** $p < 0.001$. CRP: C-reactive protein; FFA: free fatty acid. Other abbreviations are under Table 1.

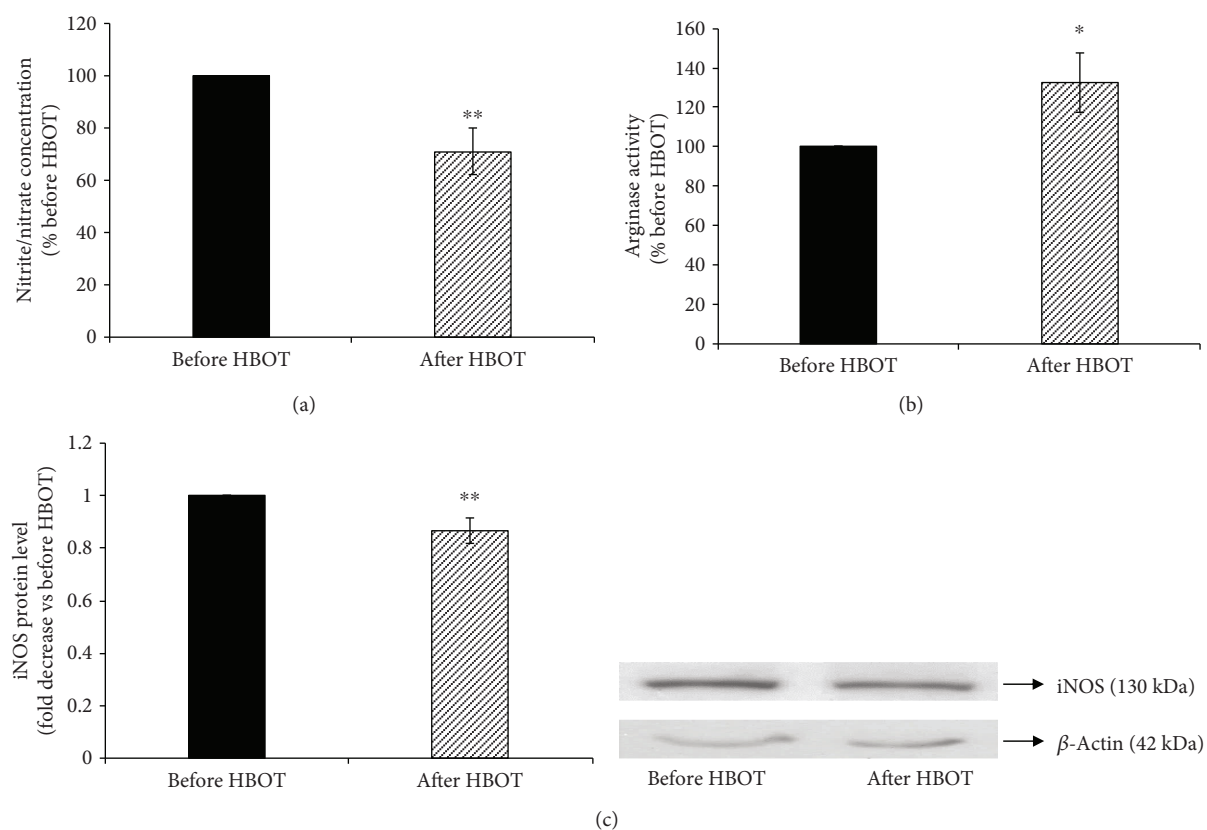


FIGURE 2: Changes in the serum nitrite/nitrate concentration, arginase activity, and iNOS protein level in lymphocytes after HBO exposure. (a) The serum nitrite/nitrate concentration was expressed as a % before HBOT; (b) the serum arginase activity was expressed as a % before HBOT; (c) level of iNOS protein in lymphocytes, normalized to β -actin and expressed as fold change before HBOT (arbitrary control set at 1). Inset: representative Western blots ($n = 11 - 13$); ** $p < 0.01$; * $p < 0.05$. iNOS: inducible nitric oxide synthase. Other abbreviations are under Table 1.

Since we have previously shown that the stimulation of iNOS also involves activation of ERK1/2 phosphorylation [22] and activation of Akt phosphorylation [23], we next examined whether ERK1/2 and Akt are involved in decreased iNOS activity/expression after the exposure to HBOT. Therefore, we assessed the effect of HBOT of the phosphorylation

state of ERK1/2 (Figure 4) and Akt (Figure 5) by the immunoblotting with phospho-specific ERK1/2 and Akt antibodies that recognize ERK1/2 and Akt only when ERK is phosphorylated at Thr²⁰²/Tyr²⁰⁴ and Akt at Ser⁴⁷³, respectively.

HBOT decreased the total ERK1/2 protein level (before HBOT = 1-fold; after HBOT = 0.89 ± 0.05 -fold; $p < 0.05$)

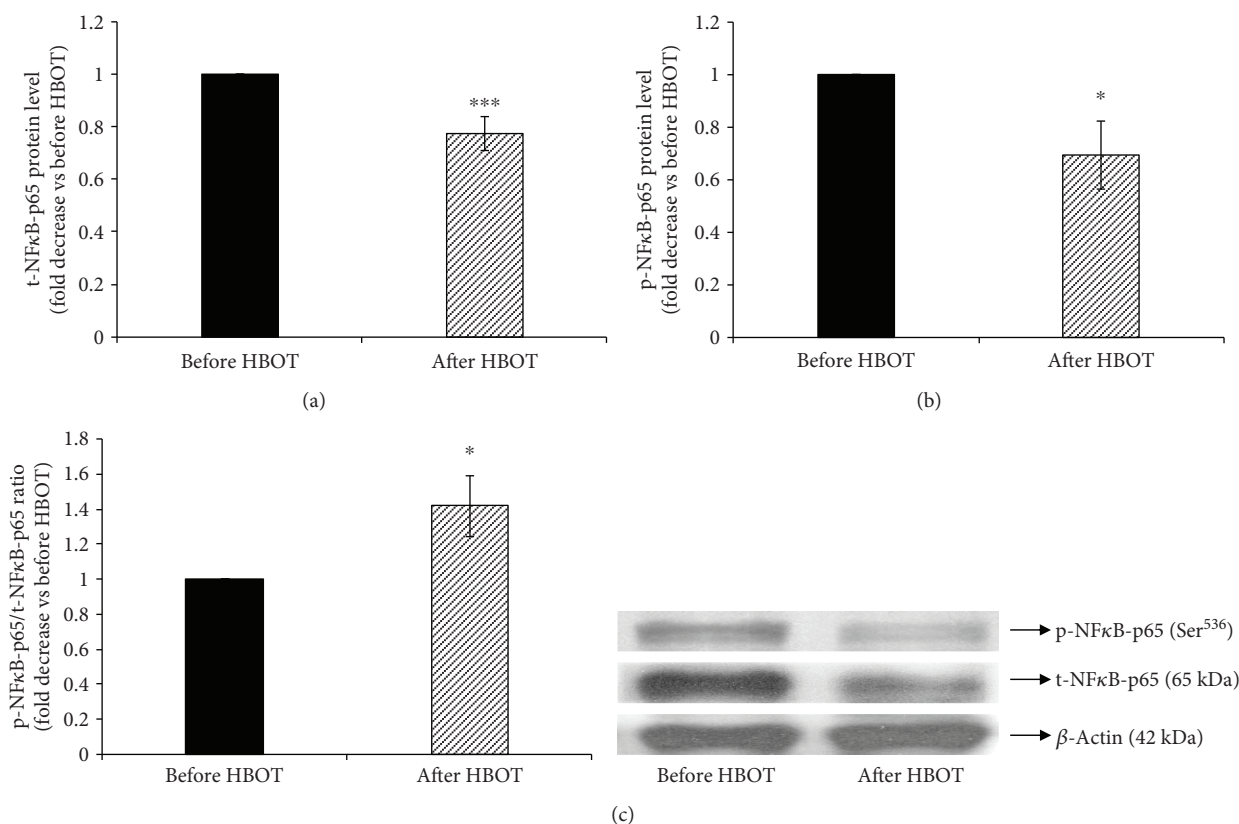


FIGURE 3: Phosphorylation of NFκB-p65 at Ser⁵³⁶ in lymphocytes after HBO exposure. (a) Level of t-NFκB-p65 protein in lymphocytes, normalized to β-actin and expressed as fold change before HBOT (arbitrary control set at 1); (b) level of p-NFκB-p65 (Ser⁵³⁶) protein in lymphocytes, normalized to β-actin and expressed as fold change before HBOT (arbitrary control set at 1); (c) ratio of p-NFκB-p65/t-NFκB-p65 expressed as fold change before HBOT (arbitrary control set at 1). Inset: representative Western blots ($n = 5 - 15$); ** $p < 0.01$; * $p < 0.05$; p-NFκB-p65: the phosphorylated form of p65 subunit of nuclear factor-κB; t-NFκB-p65: the total form p65 subunit of nuclear factor-κB. Other abbreviations are under Table 1.

(Figure 4(a)) and total Akt protein level, normalized to β-actin (before HBOT = 1-fold; after HBOT = 0.89 ± 0.04 -fold; $p < 0.05$) (Figure 5(a)). Similarly, HBOT decreased phosphorylation of ERK1/2 (Figures 4(b) and 4(c)) (before HBOT = 1-fold; after HBOT = 0.79 ± 0.06 -fold; $p < 0.01$) and phosphorylation of Akt (Figures 5(b) and 5(c)) (before HBOT = 1-fold; after HBOT = 0.57 ± 0.11 -fold; $p < 0.01$). These results suggest that exposure to HBOT significantly decreases iNOS activity/expression and indicate that ERK1/2 and Akt are involved in HBOT-downregulated iNOS activity/expression in T1DM patients.

4. Discussion

The principal findings of our study included the observation that in T1DM patients, HBOT downregulates the iNOS activity/expression by decreasing phosphorylation of ERK1/2 and Akt followed by a decreased activation of NFκB. For this study, we have used HBOT as a medical treatment for T1DM patients with the developed peripheral arterial disease, with the purpose to reduce inflammation and to promote microcirculation.

Biochemical profile of DM is commonly presented by elevated levels of HbA_{1c}, CRP, FFA, and NO which provide

an environment of oxidative stress and inflammation [24]. HbA_{1c} is a reliable biomarker for the diagnosis and prognosis of DM and correlates well with the risk of long-term diabetes complications [25]. Results from our study show that the level of HbA_{1c} was $8.6 \pm 0.08\%$, which indicates a higher risk of DM-associated complications [26].

Elevated FFA, which is common in the diabetic population [27], may trigger systemic inflammation and impair NO bioavailability, leading to impaired endothelium-dependent and insulin-mediated vasodilation [28]. Increase in FFA level is associated with an increase in NO level and upregulation of iNOS mRNA as well as the reduction of insulin output in cultured prediabetic rat pancreatic islets [29]. Also, increased plasma FFA levels are accompanied by increased expression of several NFκB-dependent cytokines, leading to chronic inflammatory processes [30]. In our study, we observed a significant decrease in FFA after HBOT in T1DM patients. These results are consistent with data gained by Teshigawara et al. [31]. Teshigawara et al. showed that FFA concentration was significantly decreased after HBOT, implying that HBOT could have beneficial effects on lipid metabolism. Clinical manifestation of inflammation is characterized by the increase of acute-phase proteins, among which prominent place has CRP [32, 33]. The level of CRP is a well-known marker of

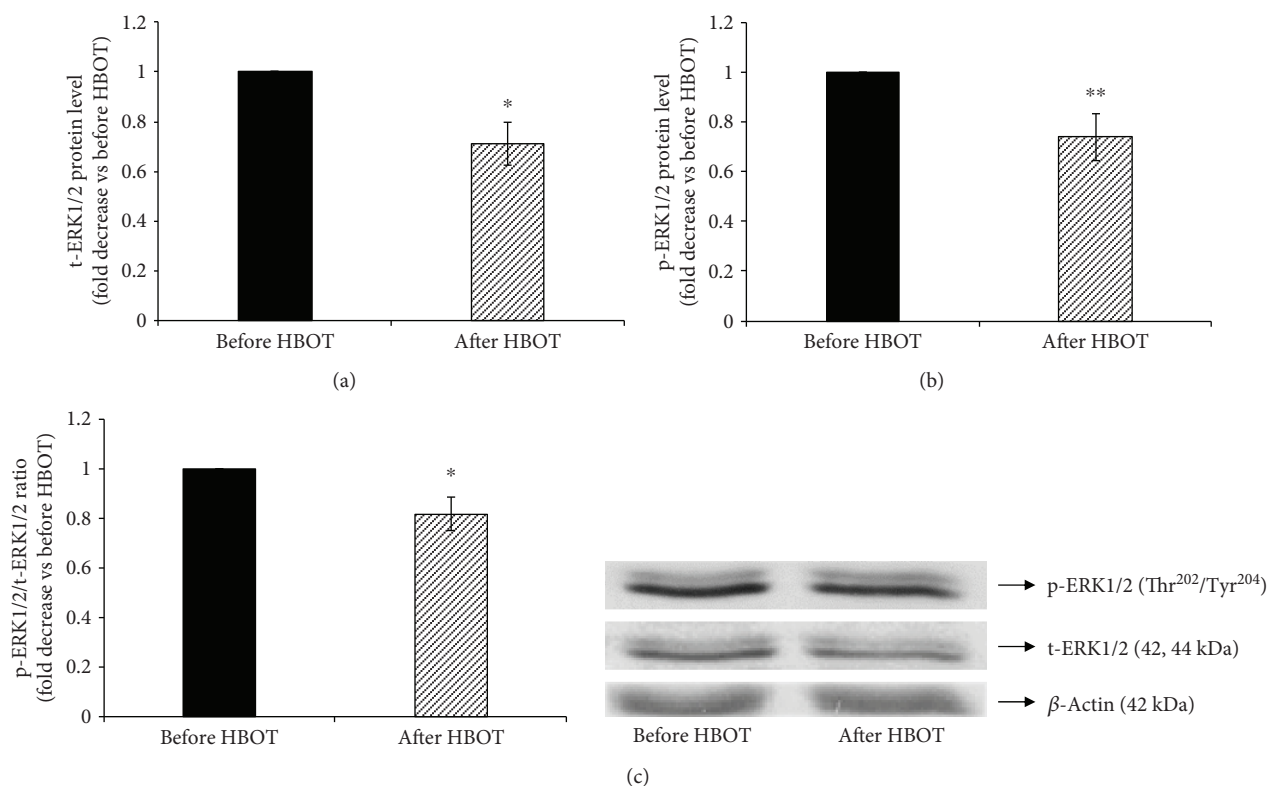


FIGURE 4: Phosphorylation of ERK1/2 at Thr²⁰² and Tyr²⁰⁴ in lymphocytes after HBO exposure. (a) Level of t-ERK1/2 protein in lymphocytes, normalized to β -actin and expressed as fold change before HBOT (arbitrary control set at 1); (b) level of p-ERK1/2/(Thr²⁰²/Tyr²⁰⁴) protein in lymphocytes, normalized to β -actin and expressed as fold change before HBOT (arbitrary control set at 1); (c) ratio of p-ERK1/2/t-ERK1/2 expressed as fold change before HBOT (arbitrary control set at 1). Inset: representative Western blots ($n = 14$); ** $p < 0.01$; * $p < 0.05$; p-ERK1/2: the phosphorylated form of extracellular signal-regulated kinases 1 and 2; t-ERK1/2: the total form of extracellular signal-regulated kinases 1 and 2. Other abbreviations are under Table 1.

inflammation [32], and it is reported that CRP may stimulate iNOS expression and NO production [34]. Our results show that exposure to HBOT significantly reduces the level of CRP in T1DM patients. Thus, the results from our study suggest that HBOT leads to a reduction of inflammation and subsequent progression of atherosclerosis [35]. iNOS is an important mediator of inflammation and may be the critical link between metabolic disorders and inflammation [36].

iNOS expression occurs after induction as a response to cytokines and other proinflammatory agents. After induction, iNOS continuously produces 100- to 1000-fold more NO than constitutive NOS [37, 38]. Excessive NO production favors the formation of peroxynitrite and thus contributes to tissue injury and reduced NO bioavailability [39, 40]. In our study, we have observed that HBOT leads to a decrease in iNOS expression in lymphocytes and consequently decreases the level of the serum nitrite/nitrate concentration. Our results also show that HBOT increased serum arginase activity, and increased arginase activity may reduce the substrate availability of NOS, since NOS and arginase use the same substrate, the amino acid L-arginine, and consequently could reduce nitrite/nitrate concentration [41].

Moreover, numerous literature data reported that activities of iNOS and arginase are regulated reciprocally by cytokines and that the inhibition of iNOS leads to the subsequent

decrease in the NO production that is associated with increased arginase activity [42–44]. Elevated levels of glucose in diabetes may enhance NO production through increased expression of endothelial NOS (eNOS) and iNOS gene and protein levels [8, 45–47]. Although iNOS/eNOS might be responsible for the release of NO from endothelial cells [47], numerous literature data reported that hyperglycaemia increased the NO level mostly through activation of iNOS [6, 48–50]. Thus, the decreased serum nitrite/nitrate concentration suggests that decreased iNOS and increased arginase activity after exposure to HBOT in our study may be due to decreased iNOS expression. This is in accordance with other results [51–56]. Pedoto et al. have also observed in their study that HBO pretreatment reduces lipopolysaccharide-induced iNOS expression [51]. Also, HBOT reduces the level of exhaled NO for 1 h in human subjects [55]. Similarly, rats exposed to HBOT for 7 days show significantly decreased NOS activity [13], while rabbits exposed to HBOT for 5 days have decreased expression of iNOS [52].

The results from our study reveal that HBOT significantly increased phosphorylation of NF κ B-p65 at Ser⁵³⁶, while it decreased the level of NF κ B-p65 protein in lymphocytes of T1DM patients. Our results are in agreement with the results of Meng et al. who have demonstrated that

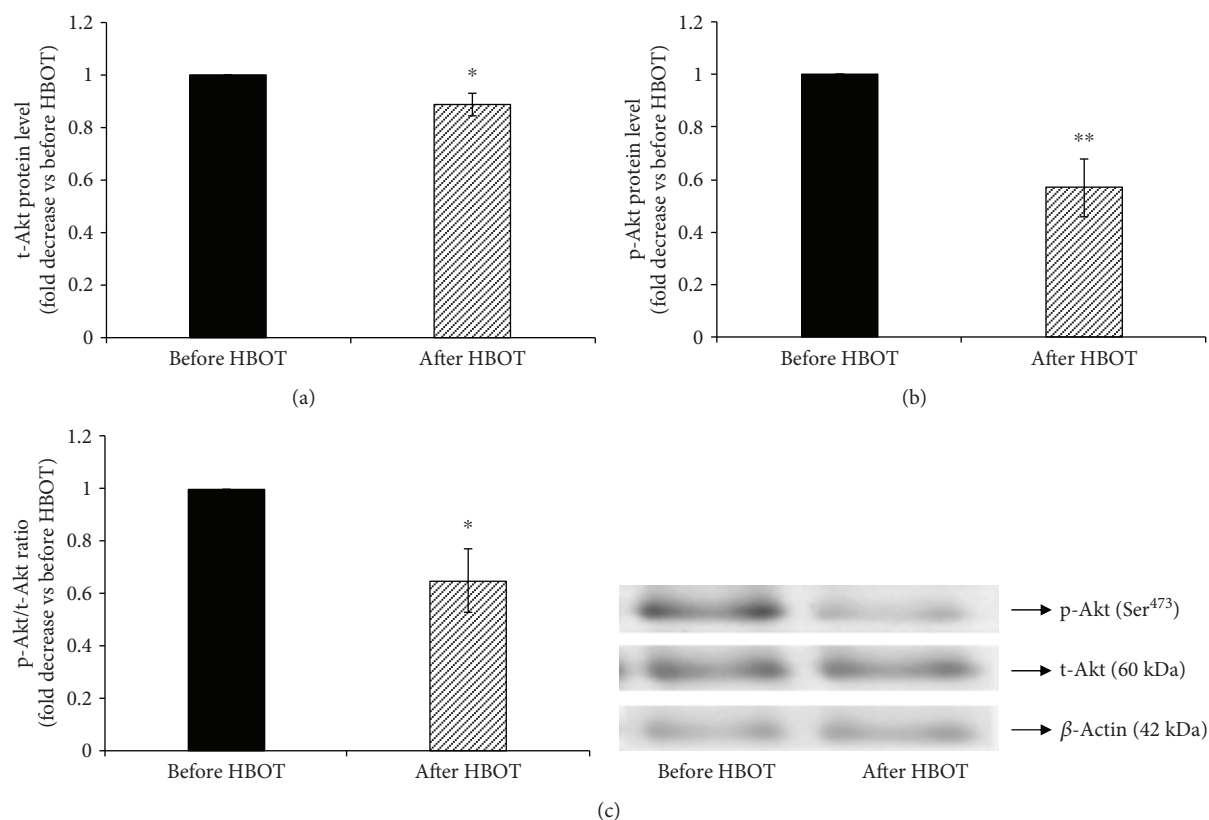


FIGURE 5: Phosphorylation of Akt at Ser⁴⁷³ in lymphocytes after HBO exposure. (a) Level of t-Akt protein in lymphocytes, normalized to β -actin and expressed as fold change before HBOT (arbitrary control set at 1); (b) level of p-Akt (Ser⁴⁷³) protein in lymphocytes, expressed as fold change before HBOT (arbitrary control set at 1); (c) ratio of p-Akt/t-Akt expressed as fold change before HBOT (arbitrary control set at 1). Inset: representative Western blots ($n = 7$); ** $p < 0.01$; * $p < 0.05$; p-Akt: the phosphorylated form of protein kinase B; t-Akt: the total form of protein kinase B. Other abbreviations are under Table 1.

exposure of rats to HBO significantly reduces the activation of the NF κ B protein [57]. Mattioli et al. reported that phosphorylation of NF κ B-p65 at Ser⁵³⁶ negatively regulates the import of NF κ B-p65 to the nucleus [58]. Additionally, phosphorylation of NF κ B-p65 at Ser⁵³⁶ plays an important role in promoting the proteasomal degradation of NF κ B-p65, thus leading to transcriptional termination of NF κ B-dependent genes [59], and phosphorylation of NF κ B-p65 at Ser⁵³⁶ regulates the ability of nuclear I κ B α to inhibit the NF κ B binding to promoters in human leukocytes and thereby inhibit transcription of genes in a selective manner [60]. Furthermore, phosphorylation of NF κ B-p65 at Ser⁵³⁶ increases turnover of NF κ B-p65, thereby inhibiting NF κ B activity in macrophages and subsequently reducing inflammation [61]. Since iNOS expression is regulated primarily by control of gene transcription, a possible explanation for the decrease of iNOS protein level after HBOT may be that decreased activation of transcription factor NF κ B-p65 subunit, which has a binding site on the iNOS gene promoter, consequentially leads to reduced iNOS expression. Here, we demonstrate that HBOT causes downregulation of NF κ B-p65 protein and this decrease could be because of the decreased cytokine level after exposure to HBO [62]. Thus, we assume that HBOT downregulates iNOS activity/expression by a mechanism involving NF κ B in combination with decreased CRP

concentrations. Although CRP by itself did not affect NO synthesis, CRP further enhanced cytokine-evoked increases in iNOS mRNA protein levels [34]. Also, we postulate that decreased iNOS expression is associated with decreased expression of FFA translocase, a cluster of differentiation 36 (CD36), and a reduction in the level of FFA after exposure to HBOT.

ERK1/2 is involved in the regulation of iNOS activation [63–66]. Increased ERK1/2 phosphorylation upregulates both NF κ B activity and iNOS protein expression [64, 67–70], while inhibition of ERK1/2 phosphorylation downregulates the expression of cytokine-induced NF κ B-activation [71]. Furthermore, inhibition of ERK1/2 decreases iNOS gene expression [71]. In our study, HBOT significantly decreases the phosphorylation of ERK 1/2, and this decrease could be due to the decreased NF κ B activity [9]. Our results are in accordance with the study of Niu et al. [72]. Niu et al. reported that HBO suppressed the phosphorylation of ERK1/2 and decreased the synthesis of NO. Therefore, we assume that decreases in ERK1/2 phosphorylation after exposure to HBOT could lead to the reduction of NF κ B activation and consequentially reduced expression of iNOS protein.

Akt is an upstream activator of NF κ B, whereby phosphorylated Akt (or ERK1/2) activates NF κ B inhibitor kinase

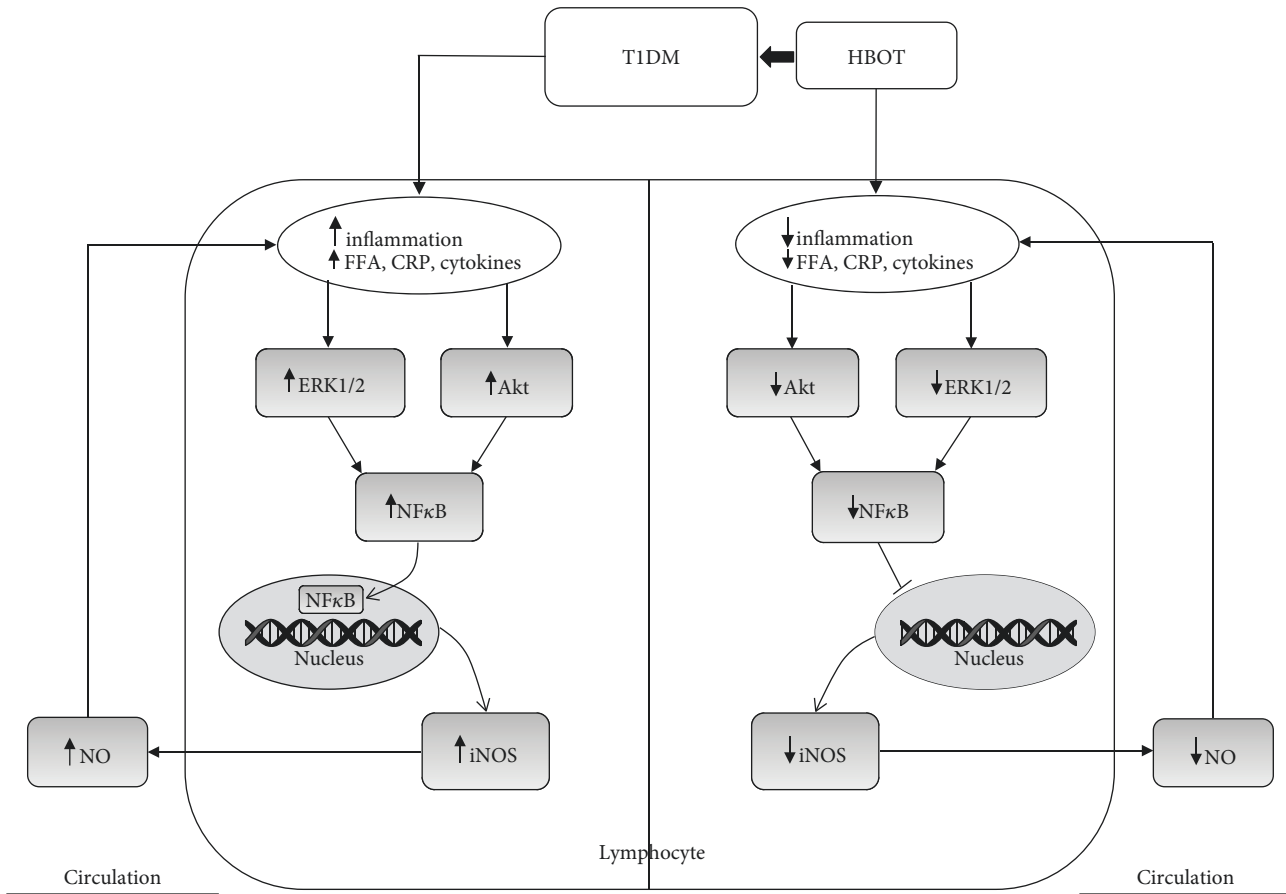


FIGURE 6: The proposed mechanism by which HBOT affects regulation of iNOS activity/expression and nitrite/nitrate production in T1DM patients. iNOS: inducible nitric oxide synthase; NF κ B-p65: the p65 subunit of nuclear factor- κ B; NO: nitric oxide; ERK1/2: extracellular signal-regulated kinases 1 and 2; Akt: protein kinase B; CRP: C-reactive protein; FFA: free fatty acid. Other abbreviations are under Table 1.

and consequently leads to transcription and translation of iNOS [68, 73]. Hattori et al. reported that the activation of NF κ B is suppressed by inhibitors of Akt [74]. In our study, HBOT decreased Akt phosphorylation, indicating that activation of Akt is involved in mediating the effect of HBOT on the regulation of iNOS. Our results are in accordance with the study of Felderhoff-Mueser et al. who reported that hyperoxia reduces phosphorylation of Akt at Ser⁴⁷³ [75]. Since the phosphorylation of Akt is necessary for NF κ B activation, we suggest that a decrease in Akt phosphorylation leads to NF κ B downregulation and a decrease in iNOS protein expression.

To explain our results, we proposed the following model (Figure 6). Altered level of FFA, CRP, and cytokines caused by T1DM induces inflammation which influences the iNOS/NO pathway and thus promotes vascular complications by attenuating insulin action. This is likely a consequence of increased ERK1/2/Akt activation, which further upregulates NF κ B. There are several possibilities to explain the effect of HBOT in the modulation of iNOS. It is possible that HBOT leads to the decreased level of CRP, FFA, and cytokines, and consequently this leads to decreased activation of ERK1/2 and Akt activation and thus decreases the activity of NF κ B, and subsequently iNOS or NF κ B activation by cytokines in T1DM is achieved only through Akt.

Also, NF κ B-p65 activation could be involved in ERK1/2 activation [9].

In accordance to the promising results of this study that pointed out the early effects of HBOT on some molecular aspects of inflammation in T1DM patients, we are planning to carry out another study that will be performed on a more significant sample size with inclusion of patients suffering from metabolic syndrome and type 2 DM as well healthy control subjects. The currently presented prospective pilot study is based on the number of administered HBOT sessions included in one HBOT course and the exploration of some early inflammatory molecular effects of HBOT after one course of exposure. As the positive effects are detected in our study, the evaluation of long-term HBOT effects (e.g., after 1 or 5 years) regarding the molecular basis of the anti-inflammatory effects of the therapy is needed. Furthermore, there is a possibility to test the effects of different HBOT administration protocols and to select the most appropriate model (i.e., study plan could be performed either through the increase in the number of sessions involved in HBOT course or by shortening of the period between successive HBOT courses). Besides evaluating the molecular mechanisms involved in the reduction of inflammation, it is also necessary to design prediction models for major cardiovascular and cerebrovascular atherosclerotic events and to examine

the influence of various confounding variables that will be of great interest and contribution in the planning of primary and secondary prevention of major atherosclerotic accidents in diabetic patients and in improving patients' quality of life.

The limitation of our study is a small number of T1DM patients exposed to HBOT. The other limitation of our study is that we did not directly measure the level of cytokines and we did not measure the level of NO in lymphocytes and the level of NF κ B in the lymphocyte nucleus.

In conclusion, in the present study, we demonstrate for the first time that HBOT causes downregulation of iNOS activity/expression by a mechanism involving ERK1/2, Akt, and NF κ B, in T1DM patients. The results from our study suggest that HBOT has a vital role in the reduction of inflammation in T1DM patients. Future studies on a larger population size are needed to elucidate the molecular mechanism by which HBOT regulates iNOS activity/expression in T1DM patients with vascular complications.

Abbreviations

Akt:	Protein kinase B
CD36:	Cluster of differentiation 36
CRP:	C-reactive protein
DM:	Diabetes mellitus
DTT:	Dithiothreitol
EDTA:	Ethylenediaminetetraacetic acid
eNOS:	Endothelial NOS
ERK1/2:	Extracellular signal-regulated kinases 1/2
FFA:	Free fatty acids
HbA _{1c} :	Glycosylated hemoglobin A1c
HBO:	Hyperbaric oxygen
HBOT:	Hyperbaric oxygen therapy
iNOS:	Inducible nitric oxide synthase
ISPF:	Alpha-isonitrosopropiophenone
LSM:	Lymphocyte separation media
NaCl:	Sodium chloride
NF κ B-p65:	p65 subunit of nuclear factor- κ B
NO:	Nitric oxide
T1DM:	Type 1 diabetes mellitus
TES:	N-[Tris(hydroxymethyl)methyl]-2-aminoethanesulfonic acid
TG:	Triglyceride
TRIS:	Tris(hydroxymethyl)aminomethane.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Disclosure

Results presented in this work are part of the PhD thesis of MSc Ivana Resanovic.

Conflicts of Interest

The authors confirm that this article content has no conflict of interest.

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References

- [1] S. Rahman, T. Rahman, A. A. S. Ismail, and A. R. A. Rashid, "Diabetes-associated macrovasculopathy: pathophysiology and pathogenesis," *Diabetes, Obesity & Metabolism*, vol. 9, no. 6, pp. 767–780, 2007.
- [2] W. T. Cade, "Diabetes-related microvascular and macrovascular diseases in the physical therapy setting," *Physical Therapy*, vol. 88, no. 11, pp. 1322–1335, 2008.
- [3] S. B. Williams, J. A. Cusco, M. A. Roddy, M. T. Johnstone, and M. A. Creager, "Impaired nitric oxide-mediated vasodilation in patients with non-insulin-dependent diabetes mellitus," *Journal of the American College of Cardiology*, vol. 27, no. 3, pp. 567–574, 1996.
- [4] T. S. Assmann, L. A. Brondani, A. P. Bouças et al., "Nitric oxide levels in patients with diabetes mellitus: a systematic review and meta-analysis," *Nitric Oxide*, vol. 61, pp. 1–9, 2016.
- [5] C. Rask-Madsen and G. L. King, "Mechanisms of disease: endothelial dysfunction in insulin resistance and diabetes," *Nature Clinical Practice. Endocrinology & Metabolism*, vol. 3, no. 1, pp. 46–56, 2007.
- [6] M. Fujimoto, N. Shimizu, K. Kunii, J. A. J. Martyn, K. Ueki, and M. Kaneki, "A role for iNOS in fasting hyperglycemia and impaired insulin signaling in the liver of obese diabetic mice," *Diabetes*, vol. 54, no. 5, pp. 1340–1348, 2005.
- [7] P. R. Nagareddy, H. Soliman, G. Lin et al., "Selective inhibition of protein kinase C β_2 attenuates inducible nitric oxide synthase-mediated cardiovascular abnormalities in streptozotocin-induced diabetic rats," *Diabetes*, vol. 58, no. 10, pp. 2355–2364, 2009.
- [8] P. Yang, Y. Cao, and H. Li, "Hyperglycemia induces inducible nitric oxide synthase gene expression and consequent nitrosative stress via c-Jun N-terminal kinase activation," *American Journal of Obstetrics and Gynecology*, vol. 203, no. 2, pp. 185.e5–185.e11, 2010.
- [9] K. M. Ahmed, S. Dong, M. Fan, and J. J. Li, "Nuclear factor- κ B p65 inhibits mitogen-activated protein kinase signaling pathway in radioresistant breast cancer cells," *Molecular Cancer Research*, vol. 4, no. 12, pp. 945–955, 2006.
- [10] H. C. Dan, M. J. Cooper, P. C. Cogswell, J. A. Duncan, J. P. Y. Ting, and A. S. Baldwin, "Akt-dependent regulation of NF- κ B is controlled by mTOR and Raptor in association with IKK," *Genes & Development*, vol. 22, no. 11, pp. 1490–1500, 2008.
- [11] K. K. Jain, "Physiological, and biochemical aspects of hyperbaric oxygenation," in *Textbook of Hyperbaric Medicine*, K.

- K. Jain, Ed., Hogrefe & Huber Publishers, Cambridge, MA, USA, 5th edition, 2009.
- [12] B. Ratzenhofer-Komenda, J. Niinikoski, and M. Hamilton-Farell, "Physical and pathophysiological bases of hyperbaric oxygen therapy," in *Handbook on Hyperbaric Medicine*, pp. 14–159, Springer, Dordrecht, Netherlands, 2006.
 - [13] D. Rachmilewitz, F. Karmeli, E. Okon, I. Rubenstein, and O. S. Better, "Hyperbaric oxygen: a novel modality to ameliorate experimental colitis," *Gut*, vol. 43, no. 4, pp. 512–518, 1998.
 - [14] G. Han, L. Li, and L. Meng, "Effects of hyperbaric oxygen on pain-related behaviours and nitric oxide synthase expression in a rat model of neuropathic pain," *Pain Research & Management*, vol. 18, no. 3, pp. 137–141, 2013.
 - [15] J. Stanimirovic, M. Obradovic, A. Jovanovic et al., "A high fat diet induces sex-specific differences in hepatic lipid metabolism and nitrite/nitrate in rats," *Nitric Oxide*, vol. 54, pp. 51–59, 2016.
 - [16] W. G. Duncombe, "The colorimetric micro-determination of non-esterified fatty acids in plasma," *Clinica Chimica Acta*, vol. 9, no. 2, pp. 122–125, 1964.
 - [17] I. M. Corraliza, M. L. Campo, G. Soler, and M. Modollell, "Determination of arginase activity in macrophages: a micro-method," *Journal of Immunological Methods*, vol. 174, no. 1–2, pp. 231–235, 1994.
 - [18] A. Boyum, "Isolation of mononuclear cells and granulocytes from human blood. Isolation of mononuclear cells by one centrifugation, and of granulocytes by combining centrifugation and sedimentation at 1 g," *Scandinavian Journal of Clinical and Laboratory Investigation. Supplementum*, vol. 97, pp. 77–89, 1968.
 - [19] O. H. Lowry, N. Rosebrough, A. Farr, and R. Randall, "Protein measurement with the Folin phenol reagent," *The Journal of Biological Chemistry*, vol. 193, no. 1, pp. 265–275, 1951.
 - [20] U. K. Laemmli, "Cleavage of structural proteins during the assembly of the head of bacteriophage T4," *Nature*, vol. 227, no. 5259, pp. 680–685, 1970.
 - [21] S. Negi, E. V. Kharshiing, and R. Sharma, "NO way! Is nitric oxide level in tomato regulated by a mammalian IKK/NF- κ B like signaling pathway?," *Plant Signaling & Behavior*, vol. 6, no. 7, pp. 1049–1052, 2014.
 - [22] E. R. Isenovic, D. B. Jacobs, M. H. Kedees et al., "Angiotensin II regulation of the Na⁺ pump involves the phosphatidylinositol-3 kinase and p42/44 mitogen-activated protein kinase signaling pathways in vascular smooth muscle cells," *Endocrinology*, vol. 145, no. 3, pp. 1151–1160, 2004.
 - [23] E. R. Isenovic, Y. Meng, A. Divald, N. Milivojevic, and J. R. Sowers, "Role of phosphatidylinositol 3-kinase/Akt pathway in angiotensin II and insulin-like growth factor-1 modulation of nitric oxide synthase in vascular smooth muscle cells," *Endocrine*, vol. 19, no. 3, pp. 287–292, 2002.
 - [24] S. R. Thom, "Oxidative stress is fundamental to hyperbaric oxygen therapy," *Journal of Applied Physiology*, vol. 106, no. 3, pp. 988–995, 2009.
 - [25] S. I. Sherwani, H. A. Khan, A. Ekhezaimy, A. Masood, and M. K. Sakharkar, "Significance of HbA1c test in diagnosis and prognosis of diabetic patients," *Biomarker Insights*, vol. 11, pp. 95–104, 2016.
 - [26] S. A. Virk, K. C. Donaghue, Y. H. Cho et al., "Association between HbA1c variability and risk of microvascular complications in adolescents with type 1 diabetes," *The Journal of Clinical Endocrinology and Metabolism*, vol. 101, no. 9, pp. 3257–3263, 2016.
 - [27] R. N. Bergman and M. Ader, "Free fatty acids and pathogenesis of type 2 diabetes mellitus," *Trends in Endocrinology and Metabolism*, vol. 11, no. 9, pp. 351–356, 2000.
 - [28] D. Tousoulis, N. Papageorgiou, E. Androulakis et al., "Diabetes mellitus-associated vascular impairment: novel circulating biomarkers and therapeutic approaches," *Journal of the American College of Cardiology*, vol. 62, no. 8, pp. 667–676, 2013.
 - [29] M. Shimabukuro, M. Ohneda, Y. Lee, and R. H. Unger, "Role of nitric oxide in obesity-induced beta cell disease," *The Journal of Clinical Investigation*, vol. 100, no. 2, pp. 290–295, 1997.
 - [30] P. Barma, S. Bhattacharya, A. Bhattacharya et al., "Lipid induced overexpression of NF- κ B in skeletal muscle cells is linked to insulin resistance," *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*, vol. 1792, no. 3, pp. 190–200, 2009.
 - [31] K. Teshigawara, T. Hosaka, M. Yamaguchi et al., "Long-term treatment with hyperbaric air improves hyperlipidemia of db/db mice," *The Journal of Medical Investigation*, vol. 57, no. 3,4, pp. 224–231, 2010.
 - [32] R. Nesto, "C-reactive protein, its role in inflammation, type 2 diabetes and cardiovascular disease, and the effects of insulin-sensitizing treatment with thiazolidinediones," *Diabetic Medicine*, vol. 21, no. 8, pp. 810–817, 2004.
 - [33] L. Marseglia, S. Manti, G. D'Angelo et al., "Oxidative stress in obesity: a critical component in human diseases," *International Journal of Molecular Sciences*, vol. 16, no. 1, pp. 378–400, 2015.
 - [34] U. Ikeda, Y. Maeda, K. Yamamoto, and K. Shimada, "C-reactive protein augments inducible nitric oxide synthase expression in cytokine-stimulated cardiac myocytes," *Cardiovascular Research*, vol. 56, no. 1, pp. 86–92, 2002.
 - [35] D. A. Rossignol, L. W. Rossignol, S. J. James, S. Melnyk, and E. Mumper, "The effects of hyperbaric oxygen therapy on oxidative stress, inflammation, and symptoms in children with autism: an open-label pilot study," *BMC Pediatrics*, vol. 7, no. 1, p. 36, 2007.
 - [36] J. Stanimirovic, M. Obradovic, S. Zafirovic et al., "Effects of altered hepatic lipid metabolism on regulation of hepatic iNOS," *Clinical Lipidology*, vol. 10, no. 2, pp. 167–175, 2015.
 - [37] S. S. Soskić, B. D. Dobutović, E. M. Sudar et al., "Regulation of inducible nitric oxide synthase (iNOS) and its potential role in insulin resistance, diabetes and heart failure," *Open Cardiovascular Medicine Journal*, vol. 5, no. 1, pp. 153–163, 2011.
 - [38] C. Nathan and Q. W. Xie, "Nitric oxide synthases: roles, tolls, and controls," *Cell*, vol. 78, no. 6, pp. 915–918, 1994.
 - [39] S. Cuzzocrea, E. Mazzon, L. Dugo, R. di Paola, A. P. Caputi, and D. Salvemini, "Superoxide: a key player in hypertension," *The FASEB Journal*, vol. 18, no. 1, pp. 94–101, 2004.
 - [40] S. S. A. El Gawad, M. M. Yossof, A. A. Wafa Soliman, A. A. Abd El-Aziz, F. B. El-Shahat, and A. K. Selim, "Expression of inducible nitric oxide synthase gene in diabetic and non-diabetic coronary artery disease patients," *Researcher*, vol. 3, no. 6, pp. 40–48, 2011.
 - [41] C. V. Suschek, O. Schnorr, and V. Kolb-Bachofen, "The role of iNOS in chronic inflammatory processes in vivo: is it damage-promoting, protective, or active at all?," *Current Molecular Medicine*, vol. 4, no. 7, pp. 763–775, 2004.

- [42] M. Mori and T. Gotoh, "Regulation of nitric oxide production by arginine metabolic enzymes," *Biochemical and Biophysical Research Communications*, vol. 275, no. 3, pp. 715–719, 2000.
- [43] M. Modolell, I. M. Corraliza, F. Link, G. Soler, and K. Eichmann, "Reciprocal regulation of the nitric oxide synthase/arginase balance in mouse bone marrow-derived macrophages by TH1 and TH2 cytokines," *European Journal of Immunology*, vol. 25, no. 4, pp. 1101–1104, 1995.
- [44] C. R. Morris, M. Poljakovic, L. Lavrisha, L. Machado, F. A. Kuypers, and S. M. Morris Jr., "Decreased arginine bioavailability and increased serum arginase activity in asthma," *American Journal of Respiratory and Critical Care Medicine*, vol. 170, no. 2, pp. 148–153, 2004.
- [45] F. Cosentino, K. Hishikawa, Z. S. Katusic, and T. F. Luscher, "High glucose increases nitric oxide synthase expression and superoxide anion generation in human aortic endothelial cells," *Circulation*, vol. 96, no. 1, pp. 25–28, 1997.
- [46] S. CAI, J. KHOO, and K. M. CHANNON, "Augmented BH4 by gene transfer restores nitric oxide synthase function in hyperglycemic human endothelial cells," *Cardiovascular Research*, vol. 65, no. 4, pp. 823–831, 2005.
- [47] R. Adela, S. K. Nethi, P. K. Bagul et al., "Hyperglycaemia enhances nitric oxide production in diabetes: a study from South Indian patients," *PLoS One*, vol. 10, no. 4, article e0125270, 2015.
- [48] H. Sugita, M. Kaneki, E. Tokunaga et al., "Inducible nitric oxide synthase plays a role in LPS-induced hyperglycemia and insulin resistance," *American Journal of Physiology. Endocrinology and Metabolism*, vol. 282, no. 2, pp. E386–E394, 2002.
- [49] J. Tanaka, L. Qiang, A. S. Banks et al., "Foxo1 links hyperglycemia to LDL oxidation and endothelial nitric oxide synthase dysfunction in vascular endothelial cells," *Diabetes*, vol. 58, no. 10, pp. 2344–2354, 2009.
- [50] N. di Pietro, P. di Tomo, S. di Silvestre et al., "Increased iNOS activity in vascular smooth muscle cells from diabetic rats: potential role of Ca²⁺/calmodulin-dependent protein kinase II delta 2 (CaMKII δ_2)," *Atherosclerosis*, vol. 226, no. 1, pp. 88–94, 2013.
- [51] A. Pedoto, J. Nandi, Z. J. Yang et al., "Beneficial effect of hyperbaric oxygen pretreatment on lipopolysaccharide-induced shock in rats," *Clinical and Experimental Pharmacology & Physiology*, vol. 30, no. 7, pp. 482–488, 2003.
- [52] L.-J. Yuan, S. W. N. Ueng, S.-S. Lin, W.-L. Yeh, C.-Y. Yang, and P. Y. Lin, "Attenuation of apoptosis and enhancement of proteoglycan synthesis in rabbit cartilage defects by hyperbaric oxygen treatment are related to the suppression of nitric oxide production," *Journal of Orthopaedic Research*, vol. 22, no. 5, pp. 1126–1134, 2004.
- [53] B. J. Kudchodkar, A. Pierce, and L. Dory, "Chronic hyperbaric oxygen treatment elicits an anti-oxidant response and attenuates atherosclerosis in apoE knockout mice," *Atherosclerosis*, vol. 193, no. 1, pp. 28–35, 2007.
- [54] B. J. Kudchodkar, J. Wilson, A. Lacko, and L. Dory, "Hyperbaric oxygen reduces the progression and accelerates the regression of atherosclerosis in rabbits," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 20, no. 6, pp. 1637–1643, 2000.
- [55] Z. A. Puthuchery, J. Liu, M. Bennett, B. Trytko, S. Chow, and P. S. Thomas, "Exhaled nitric oxide is decreased by exposure to the hyperbaric oxygen therapy environment," *Mediators of Inflammation*, vol. 2006, Article ID 72620, 6 pages, 2006.
- [56] C. Luongo, F. Imperatore, S. Cuzzocrea et al., "Effects of hyperbaric oxygen exposure on a zymosan-induced shock model," *Critical Care Medicine*, vol. 26, no. 12, pp. 1972–1976, 1998.
- [57] X.-E. Meng, Y. Zhang, N. Li et al., "Hyperbaric oxygen alleviates secondary brain injury after trauma through inhibition of TLR4/NF- κ B signaling pathway," *Medical Science Monitor*, vol. 22, pp. 284–288, 2016.
- [58] I. Mattioli, A. Sebald, C. Bucher et al., "Transient and selective NF- κ B p65 serine 536 phosphorylation induced by T cell costimulation is mediated by I κ B kinase β and controls the kinetics of p65 nuclear import," *Journal of Immunology*, vol. 172, no. 10, pp. 6336–6344, 2004.
- [59] S. Sacconi, I. Marazzi, A. A. Beg, and G. Natoli, "Degradation of promoter-bound p65/RelA is essential for the prompt termination of the nuclear factor κ B response," *The Journal of Experimental Medicine*, vol. 200, no. 1, pp. 107–113, 2004.
- [60] C. C. Ghosh, S. Ramaswami, A. Juvekar et al., "Gene-specific repression of proinflammatory cytokines in stimulated human macrophages by nuclear I κ B α ," *Journal of Immunology*, vol. 185, no. 6, pp. 3685–3693, 2010.
- [61] T. Lawrence, M. Bebién, G. Y. Liu, V. Nizet, and M. Karin, "IKK α limits macrophage NF- κ B activation and contributes to the resolution of inflammation," *Nature*, vol. 434, no. 7037, pp. 1138–1143, 2005.
- [62] R. M. Benson, L. M. Minter, B. A. Osborne, and E. V. Granowitz, "Hyperbaric oxygen inhibits stimulus-induced proinflammatory cytokine synthesis by human blood-derived monocyte-macrophages," *Clinical and Experimental Immunology*, vol. 134, no. 1, pp. 57–62, 2003.
- [63] E. Sudar, B. Dobutovic, S. Soskic et al., "Regulation of inducible nitric oxide synthase activity/expression in rat hearts from ghrelin-treated rats," *Journal of Physiology and Biochemistry*, vol. 67, no. 2, pp. 195–204, 2011.
- [64] E. Sudar Milovanovic, A. Jovanovic, M. Misirkic-Marjanovic, L. Vucicevic, K. Janjetovic, and E. R. Isenovic, "Effects of intracerebroventricularly (ICV) injected ghrelin on cardiac inducible nitric oxide synthase activity/expression in obese rats," *Experimental and Clinical Endocrinology & Diabetes*, vol. 123, no. 10, pp. 581–588, 2015.
- [65] S. Zafirovic, M. Obradovic, E. Sudar-Milovanovic et al., "17 β -estradiol protects against the effects of a high fat diet on cardiac glucose, lipid and nitric oxide metabolism in rats," *Molecular and Cellular Endocrinology*, vol. 446, pp. 12–20, 2017.
- [66] A. Jovanovic, E. Sudar-Milovanovic, M. Obradovic et al., "Influence of a high-fat diet on cardiac iNOS in female rats," *Current Vascular Pharmacology*, vol. 15, no. 5, pp. 491–500, 2017.
- [67] J. A. Ellerhorst, S. Ekmekcioglu, M. K. Johnson, C. P. Cooke, M. M. Johnson, and E. A. Grimm, "Regulation of iNOS by the p44/42 mitogen-activated protein kinase pathway in human melanoma," *Oncogene*, vol. 25, no. 28, pp. 3956–3962, 2006.
- [68] C. H. Hou, J. Lin, S. C. Huang, S. M. Hou, and C. H. Tang, "Ultrasound stimulates NF- κ B activation and iNOS expression via the Ras/Raf/MEK/ERK signaling pathway in cultured preosteoblasts," *Journal of Cellular Physiology*, vol. 220, no. 1, pp. 196–203, 2009.
- [69] J. W. Kim and C. Kim, "Inhibition of LPS-induced NO production by taurine chloramine in macrophages is mediated through Ras-ERK-NF- κ B," *Biochemical Pharmacology*, vol. 70, no. 9, pp. 1352–1360, 2005.

- [70] Z. Xiao, T. Wang, H. Qin, C. Huang, Y. Feng, and Y. Xia, "Endoplasmic reticulum Ca^{2+} release modulates endothelial nitric-oxide synthase via extracellular signal-regulated kinase (ERK) 1/2-mediated serine 635 phosphorylation," *The Journal of Biological Chemistry*, vol. 286, no. 22, pp. 20100–20108, 2011.
- [71] Z. Xu, B. R. Wang, X. Wang et al., "ERK1/2 and p38 mitogen-activated protein kinase mediate iNOS-induced spinal neuron degeneration after acute traumatic spinal cord injury," *Life Sciences*, vol. 79, no. 20, pp. 1895–1905, 2006.
- [72] C. C. Niu, S. S. Lin, L. J. Yuan et al., "Hyperbaric oxygen treatment suppresses MAPK signaling and mitochondrial apoptotic pathway in degenerated human intervertebral disc cells," *Journal of Orthopaedic Research*, vol. 31, no. 2, pp. 204–209, 2013.
- [73] M. R. Gwinn and V. Vallyathan, "Respiratory burst: role in signal transduction in alveolar macrophages," *Journal of Toxicology and Environmental Health. Part B, Critical Reviews*, vol. 9, no. 1, pp. 27–39, 2006.
- [74] Y. Hattori, S. Hattori, and K. Kasai, "Lipopolysaccharide activates Akt in vascular smooth muscle cells resulting in induction of inducible nitric oxide synthase through nuclear factor-kappa B activation," *European Journal of Pharmacology*, vol. 481, no. 2-3, pp. 153–158, 2003.
- [75] U. Felderhoff-Mueser, P. Bittigau, M. Siffringer et al., "Oxygen causes cell death in the developing brain," *Neurobiology of Disease*, vol. 17, no. 2, pp. 273–282, 2004.



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