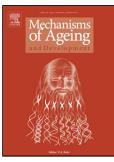
### Accepted Manuscript

Title: Exosome-based immunomodulation during aging: a nano-perspective on inflamm-aging

Author: Francesco Prattichizzo Luigina Micolucci Monica Cricca Sabrina De Carolis Emanuela Mensà Antonio Ceriello Antonio Domenico Procopio Massimiliano Bonafè Fabiola Olivieri



provided by Archiv

PII:	S0047-6374(16)30150-6
DOI:	http://dx.doi.org/doi:10.1016/j.mad.2017.02.008
Reference:	MAD 10932
To appear in:	Mechanisms of Ageing and Development
Received date:	19-8-2016
Revised date:	23-1-2017
Accepted date:	25-2-2017

Please cite this article as: Prattichizzo, F., Micolucci, L., Cricca, M., De Carolis, S., Mensgravea, E., Ceriello, A., Procopio, A.D., Bonafgravee, M., Olivieri, F.,Exosome-based immunomodulation during aging: a nanoperspective on inflamm-aging, *Mechanisms of Ageing and Development* (2017), http://dx.doi.org/10.1016/j.mad.2017.02.008

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Exosome-based immunomodulation during aging: a nano-perspective on inflamm-aging.

Francesco Prattichizzo<sup>1,2</sup>, Luigina Micolucci<sup>2</sup>, Monica Cricca<sup>3</sup>, Sabrina De Carolis<sup>3</sup>, Emanuela Mensà<sup>2</sup>, Antonio Ceriello<sup>4</sup>, Antonio Domenico Procopio<sup>2,5</sup>, Massimiliano Bonafè<sup>3\*</sup> and Fabiola Olivieri <sup>2,5\*</sup>

1- August Pi i Sunyer Biomedical Research Institute (IDIBAPS) and Centre of Biomedical Investigation on Diabetes and Associated Metabolic Disorders Network (CIBERDEM), 08036 Barcelona, Spain;

2- Department of Clinical and Molecular Sciences, DISCLIMO, Università Politecnica delle Marche, Ancona, Italy;

3- Department of Experimental, Diagnostic and Specialty Medicine, DIMES, University of Bologna, Bologna, Italy.

4- August Pi i Sunyer Biomedical Research Institute (IDIBAPS) and Centre of Biomedical Investigation on Diabetes and Associated Metabolic Disorders Network (CIBERDEM), 08036 Barcelona, Spain and Department of Cardiovascular and Metabolic Diseases, IRCCS Multimedica, Sesto San Giovanni, Milan, Italy;

5- Center of Clinical Pathology and Innovative Therapy, National Institute INRCA-IRCCS, Ancona, Italy;

### \*The authors contributed equally to the manuscript

### **Corresponding author**

Fabiola Olivieri, Ph.D,

Dept. of Clinical and Molecular Sciences (DISCLIMO),

Università Politecnica delle Marche, Ancona

Via Tronto 10/A - 60020 Ancona (Italy),

Ph: +39 071 220 6242, Fax: +39 071 220 6240

e-mail: f.olivieri@univpm.it

#### Abstract

Exosomes are nanovesicles formed by inward budding of endosomal membranes. They exert complex immunomodulatory effects on target cells, acting both as antigen-presenting vesicles and as shuttles for packets of information like proteins, coding and non-coding RNA, and nuclear and mitochondrial DNA fragments. Albeit different, all such functions seem to be encompassed in the adaptive mechanism mediating the complex interactions of the organism with a variety of stressors, providing both for defense and for the evolution of symbiotic relationships with others organisms (gut microbiota, bacteria, and viruses). Intriguingly, the newly deciphered human virome and exosome biogenesis seem to share some physical-chemical characteristics and molecular mechanisms. Exosomes are involved in immune system recognition of self from non-self throughout life: they are therefore ideal candidate to modulate inflamm-aging, the chronic, systemic, age-related pro-inflammatory status, which influence the development/progression of the most common age-related diseases (ARDs). Not surprisingly, recent evidence has documented exosomal alteration during aging and in association with ARDs, even though data in this field are still limited.

Here, we review current knowledge on exosome-based trafficking between immune cells and self/non-self cells (i.e. the virome), sketching a nano-perspective on inflamm-aging and on the mechanisms involved in health maintenance throughout life.

#### Introduction

Eukaryotic cells release a variety of extracellular vesicles (EVs) that differ in content and biophysical properties. EVs can be divided into three main classes according to their biogenesis: shedding microvesicles, apoptotic bodies, and exosomes. Exosomes derived from intracellular organelles, or multivesicular bodies (MVBs), are released by several different eukaryotic cells. Two distinct MVB pathways lead to lysosomal targeting and exosome secretion, respectively; the latter pathway is characterized by MVB fusion with the plasma membrane (**Buschow et al., 2009**). Consequently, exosomes contain molecules derived both from endosomes, like major histocompatibility complex (MHC) class II, transferrin receptor, and clathrins, and from the cell surface, including different receptor types (**Willms et al., 2016**).

The finding that exosomes also contain nucleic acids, such as fragments of nuclear and mitochondrial DNA, and coding and non-coding RNAs, has been a revolutionary biomedical discovery (Zhao et al., 2016; Huang et al., 2013; Silva and Melo, 2015; Thakur et al., 2014). Increasing evidence suggest that exosome-associated molecules reflect the pathophysiological status of releasing cells, and that their transfer may be an efficient way to induce metabolic changes in target cells (de Jong et al., 2012; Kooijmans et al., 2016). In support to this hypothesis, it should be noted that much of the load carried by exosomes consists of molecules exerting epigenetic actions (i.e. modifications that do not induce changes in the genetic code but rather in its decoding). Such molecules include: i) molecules that act directly on DNA by promoting covalent (e.g. DNA methylases and demethylases) or non-covalent binding (e.g. protein transcription factors) (Quian et al., 2015); (ii) agents that modulate DNA accessibility by promoting covalent binding to histones (e.g. histone methylases and acetylases); (iii) mRNAs that induce de novo protein synthesis in target cells; and (iv) microRNAs (miRNAs) and/or pre-miRNAs, which bind to mRNAs, modulating their translation (Smythies and Edelstein, 2012; Melo at al., 2014).

Since information exchange between cells is closely controlled by the immune system, it is highly unlikely for immune system cells to be mere viewers in the intense EV trafficking. The recent discovery that exosomes from damaged cells, including those enriched with DNA damage markers such as γH2AX and fragmented telomere repeat DNA, are able to elicit an inflammatory response (Wang and Lieberman, 2016) supports the notion that exosome content can exert an influence on inflammatory responses. Recent evidence shows that exosomal miRNAs can also modulate cellular response to endotoxins (Alexander et al., 2015), and participate in the exchanging of antigens from bacteria, parasites, and virus-infected cells (Marti and Johnson, 2016). In addition, exosomal alterations have been described during aging and in patients with the most common age-related diseases (ARDs) (Weilner et al., 2013; Xu and Tahara, 2013). Overall, age-related modifications in exosome amount and content could affect the crosstalk between organs and tissues and the signaling between immune system cells and self/non-self cells. However, despite the potential relevance of this topic to human health, data are still scarce and inconclusive.

Here, we hypothesize that age-related alterations in exosome-mediated communication may be involved in the immune system inflammatory activation that is associated with aging, i.e. inflammaging (Franceschi et al., 2000) and in age-related remodeling of immune system activity, i.e. immunosenescence (Fulop et al., 2015). Given their relevance to human health, the molecular mechanisms that link exosome trafficking to inflamm-aging and immune-senescence deserve further investigation.

#### **Exosomal trafficking**

A growing body of evidence suggests that exosomes transfer information not only in a paracrine fashion, but also through systemic mechanisms, providing a new pathway for the crosstalk among different cell types in a variety of pathophysiological conditions (**Ratajczak and Ratajczak, 2016**). As regards homing, the exosome integrin pattern appears to be the main determinant of exosome tropism, as in the case of exosomes released by cancer cells (**Hoshino A 2015**). Surface receptors

on exosomes may act as identifying signals, as suggested for glycan (**Batista et al., 2011**) and heparin sulfate proteoglycan signatures (**Christianson et al., 2013**). Exosomes communicate with target cells both through cell membrane-mediated signaling and/or through internalization, which depending on cell type may be clathrin/caveole-mediated endocytosis, phagocytosis, or macropinocytosis (**Urbanelli et al., 2013**).

The exosome-based crosstalk relies on the ability of exosomes to transfer "packets of information" to recipient cells more efficiently than "single instructions", and in a more cost-effective way. Notably, since exosome-borne information reaches target cells in remote locations, rather like a "message in a bottle", the activities of exosomes on target cells are likely to be determined not only by the composition of their cargo, but also by the metabolic status of target cells (**Ohno et al., 2016**).

Another function served by exosomes is to carry away harmful or unwanted material secreted by cells to preserve intracellular protein and RNA homeostasis (**Baixauli et al., 2014**). Notably, increased exosome release has been described in cells exposed to stress stimuli like glucose starvation and hypoxia, despite their unfavorable energetic status (**Garcia et al., 2015**). The exosome cargo may also exert beneficial effects on stressed target cells, as demonstrated by recent evidence that cancer cells can exploit it as a source of nutrients in nutrient deprivation or nutrient-stressed conditions (**Zhao et al., 2016**). Besides such extreme conditions, the functions exerted by exosomes are primarily related to the crosstalk among cells of different tissues, which necessarily involves immune system cells.

#### Exosomes as the interface between the virome and inflamm-aging

Even though exosomes are emerging as important constituents of the eukaryotic cell secretome, they seem to have been conserved during evolution, since bacteria and fungi also release microvesicles (**Mashburn and Whiteley, 2005; Deatherage and Cookson, 2012**). Parasite-derived EVs can transfer virulence factors and drug resistance markers, alter host cell gene expression, and

promote parasite adherence and host cell proliferation (Marti and Johnson, 2016). Moreover, infected cells can secrete exosomes that contain pathogenic compounds, which thus deliver noxious cargo to distant organs and tissues (Shimoda et al., 2016). Very recent data suggest that EVs are also a novel mode of viral propagation exploited by viruses to exit from cells non-lytically, to hide, and to manipulate the immune system (Altan-Bonnet, 2016). Several studies have documented a close similarity between exosome biogenesis, uptake, and secretion and the viral lifecycle molecular machinery (van Dongen et a., 2016, Meckes DG Jr, 2015; Nolte-'t Hoen et al., 2016). In keeping with their role as endogenous mRNA/miRNA carriers, EVs are exploited by viral RNA (e.g. human immunodeficiency virus and hepatitis C virus [HCV]) to propagate viral infection (Chahar et al., 2015). Indeed, EVs play a crucial role in HCV immune evasion, because serum-derived exosomes are infective and resistant to neutralizing antibodies (Liu et al., 2014b). Intriguingly, HCV-infected cells trigger an anti-viral response in neighboring infected cells, thus helping circumscribe and halt the infection. However, exosomes also play a pivotal role in some DNA virus infections, contributing to tumorigenesis, immune evasion, and viral latency (Schwab et al, 2015).

In particular, Epstein Barr virus (EBV) and herpes simplex virus 1 exploit exosomes both for spreading and/or curtailing viral mRNA and miRNA propagation throughout the organism (**Gutzeit et al., 2014: Han et al., 2016; Kalamvoki et al., 2014**). Interestingly, latent EBV infection induces increased secretion of several inflammatory factors, whereas lytic infections evade the antiviral inflammatory response. A recently discovered molecular mechanism underpinning exosome uptake by receptor-ligand interactions with recipient cells has demonstrated specificity for cargo delivery, suggesting an intriguing overlap of exosomes and viruses (**van Dongen et a., 2016**). For instance, B cell–derived exosomes released from EBV-infected B cells can deliver their content to B cells, not to T cells (**Gutzeit et al., 2014**).

The virome behaves in a similar way to the microbiome, i.e. it arouses the immune system, suppresses inflammation, and occupies the ecological niche, crowding out potential pathogens. Like exogenous viruses, endogenous viral-like sequences (Alu, Line, Herv) are also virome constituents

(Virgin, 2014). High-throughput metagenomic sequencing analysis has demonstrated that hundreds of human and non-human viruses, mostly species that are not reported to be pathogenic in humans, outnumber their pathogenic counterparts and dwell in healthy individuals, contributing to the human virome (Wylie et al., 2014). Furthermore, recent literature reports that some viral species (i.e. papillomavirus, polyomavirus, and anellovirus) populate ecological niches paralleling the microbiome behavior (Fancello et al., 2014). For instance, unpublished data from our lab indicate that papillomavirus DNA is found in exosomes from circulating serum and urine also in the absence of overt disease, thus strengthening the hypothesis that exosomes are involved in the interplay between virome and immunosenescence.

Overall, EVs and exosomes seem to play important but opposing roles in viral disease pathogenesis, both as "Trojan horses" and as "marathon soldiers" (**Figure 1**).

#### Exosome-mediated crosstalk between immune cells

An effective immune response requires the engagement of host receptors by pathogen-derived molecules and the stimulation of appropriate cellular responses. Surprisingly, exosomes can shuttle pathogenic molecules that exert opposite functions, serving as antigens of innate immune receptors to activate host defenses or promoting pathogen immune evasion (Schorey and Harding, 2016). Exosomes therefore provide a new, complex system for communication among immune cells, i.e. T and B lymphocytes and antigen-presenting cells (APCs) that is to be added to the mechanisms involving receptor-ligand cell-cell interactions or receptor-soluble ligand interaction. Exosomes have well-characterized immune presentation properties that are related to their formation in MVBs, where the antigen is bound to the MHC molecules. Exosomes secreted by APCs, such as B cells, mast cells, and mature and immature dendritic cells (DCs), contain MHC class I and II molecules (Pêche et al., 2006; Thery et al., 2001; Segura et al., 2005a) and are able to bind to specific T cell receptors (Eken et al., 2010); alternatively, they can be recaptured by DCs for antigen presentation (Morelli et al., 2004). Exosomes released from DCs have been shown to carry - besides peptide

MHC-I and -II - other T cell costimulatory molecules, such as CD80/B7.1 and CD86/B7.2, which exert a strong stimulation activity on the immune response (**Thery et al., 2002; 2009**).

DC-derived exosomes can also bind bacterial ligands for toll-like-receptor (TLR), thus acquiring the ability to induce strong activation of bystander DCs, enhancing proinflammatory cytokine secretion and stimulating the crosstalk with natural killer cells (**Sobo-Vujanovic et al., 2014**). Thus, exosomes play a key role in disseminating pathogen- as well as host-derived molecules during infections (**Schorey et al., 2015**). Overall, DC-released exosomes are able to induce an immune response by spreading MHC-antigen complexes to both CD4<sup>+</sup> and CD8<sup>+</sup> T lymphocytes, or by spreading TLR ligands to other DCs (**Iraci et al., 2016**). Adhesion molecules expressed on the exosome membrane, such as ICAM-1, can contribute to the interaction with lymphocyte function-associated antigen (LFA)-1 expressed by DCs and T lymphocytes (**Iraci et al., 2016**). Exosomes may stimulate immune system function also in allergic responses. DC-derived exosomes present allergenic antigens, inducing T cell T(H)2-like cytokine production in allergic donors (**Vallhov et al., 2015**). These findings suggest that such exosomes are involved in a potent mechanism that systemically alerts the host immune system to pathogen/allergen invasion.

The immune modulation action of exosomes includes not only antigen presentation, but also immune suppression (**Zhang et al., 2014**). Recent hypothesis suggest a role of exosomes to establish central tolerance, contributing to tissue-restricted antigen presentation within the thymic micromileus (**Skogberg et al., 2015**). When DC-derived exosomes are injected prior to skin graft implantation in animal models, exosomes from mature DCs can trigger effector T cell responses, leading to rapid graft rejection (**Segura et al., 2005b**), whereas exosomes from immature DCs inhibit anti-donor immune responses, significantly prolonging heart allograft survival (**Peche et al., 2003**). Moreover, oral administration of a protein antigen generates tolerosomes that induce tolerance when transferred to naive recipients (**Ostman et al., 2005**).

The contrasting immunomodulatory effects induced by exosomes, i.e. activation and inhibition of the immune response, may depend on the relative concentrations of specific subsets of antigen-

presenting exosomes and/or on the duration of exposure of immature immune cells to specific exosome subsets. The notion is strongly supported by data from in vitro and in vivo animal models, demonstrating the efficient alloantigen presentation and immunomodulatory abilities of exosomes in organ transplants (**Monguió-Tortajada et al., 2014**). Importantly, such activity could be directly mediated either by peptide-loaded MHC molecules or by accompanying epigenetic information, e.g. miRNAs, carried by antigen-loaded exosomes. For instance, exosomes derived from LPS-stimulated DCs contain sets of miRNAs that are capable of modulating endotoxin response in vivo (**Alexander et al., 2015**). Notably, exosomes containing miRNAs (i.e. miR-21) can directly activate intracellular TLR receptors, triggering a proinflammatory response (**Fabbri et al., 2012**).

A key notion in this context is that immune cell-derived exosomes can also modulate the phenotype of endothelial cells (ECs), thus promoting a systemic response (**Prattichizzo et al., 2016**). Monocyte-released exosomes can induce expression of adhesion molecules and cytokine secretion in ECs (**Tang et al., 2016**) or enhance their migratory properties (**Zhang et al., 2010**). On the other hand, exosomes derived from cardiomyocytes can transfect ECs to deliver signals that mediate heart repair after injury (**Yuan et al., 2016**).

It is conceivable that the aging process, by tilting the balance toward a persistent proinflammatory state, may compromise exosome-based communications among cells and immune system function.

#### Age-related changes in the circulating exosome pool

Aging is a complex phenotype associated with a variety of molecular and tissue alterations. Recent data from anti-aging and regenerative research indicate that microenvironmental and circulating factors deserve close attention (Almaça et al., 2014; Scudellari, 2015; Childs et al., 2015).

Our group has been providing evidence that sets of circulating miRNAs involved in the modulation of inflammation (thus designated inflammamiRs) are deregulated in aging and in ARDs (**Olivieri et al., 2015**). Circulating miRNAs are either exosome-borne or protein-bound, and both types can be functionally transferred to recipient cells (**Turchinovich et al., 2015**). The majority of miRNAs modulating inflammation, immunity, and aging pathways were seen to be exosome-associated

(Figure 2). These miRNA signatures are also involved in the modulation of insulin/IGF-1, mTOR, and other pathways relevant to aging and cellular senescence (Olivieri et al., 2015).

Accumulating senescent and/or pre-senescent cells during aging may influence the release and/or content of the circulating exosome pool. The progressive accrual of senescent cells during organismal aging could modulate the systemic exosome pool through two different mechanisms involving release by senescent cells of: i) a different amount of exosomes compared with younger cells, or ii) exosomes with a different content from exosomes derived from younger cells. The few available data support both hypotheses (Weilner et al., 2013), even though recent findings suggest that inflamm-aging and frailty do not result in an increased concentration of circulating EVs (Alberro et al., 2016). Evidence from cellular models suggests that exosomes released by senescent cells are more proinflammatory than those released by younger cells (Mitsuhashi et al., 2013). Moreover, even though plasma platelet-derived exosome number was lower in subjects older than 65 years than in younger ones, chemokine and HMGB1 levels were higher in the former subjects (Goetzl et al., 2016). These data support the hypothesis that macrophages and platelet-derived exosomes may contribute to the systemic spread of inflamm-aging. Interestingly, reduced phagocytic activity has been reported in monocytes from elderly donors both in mice and humans, demonstrating dysregulation of monocyte subpopulations with age (Bliederhaeuser et al., 2016). Significantly increased exosome release has been described in prostate cancer cells exposed to high clinical doses of radiation, which is a powerful mechanism inducing senescence through p53 activation (Lehmann et al., 2008). Interestingly, p53 pathway influences exosome formation in colorectal cancer cell lines (Sun et al., 2016) and an increased microvesicles shedding induced by pro-senescence stimuli was observed also in non-cancerous cell lines (Effenberger et al., 2014). These data lend support to the hypothesis that cellular senescence induced either by telomere attrition (i.e. replicative senescence) or by DNA damage (i.e. radiotherapy) may induce a p53dependent increase in the biogenesis of exosome-like vesicles and/or alter their cargo. Notably, senescence is associated with the acquisition of a proinflammatory and secretory phenotype -

designated SASP (senescence associated secretory phenotype) (Lecot et al., 2016) or SMS (senescence messaging secretome) (Kuilman and Peeper, 2009) - that promotes tumor progression (mostly by influencing the tumor-immune system crosstalk) and chemoresistance (Lecot et al., 2016; Di Mitri and Alimonti, 2016), and accelerates aging (Childs et al., 2015). Interestingly, p53 reactivation in advanced liver cancer has been associated with a functional SASP that was capable of attracting innate immune cells (neutrophils, macrophages, and natural killer cells), thus promoting the clearance of pre-malignant cells (Xue et al., 2007). Moreover, NF-kB, the main proinflammatory transcription factor and key SASP modulator (Salminen et al., 2012), has recently been involved in the modulation of exosome cargo. NF-kB -/- mice show an altered exosome content after exposure to a noxious stimulus (Yang et al., 2015). Furthermore, the SASP can transmit senescence to bystander cells (Acosta et al., 2013). Latent EBV infection induces increased secretion of several inflammatory factors, whereas lytic infections evade the antiviral inflammatory response. Interestingly, latent EBV infection in EBV-positive cells induces senescence and SASP acquisition in neighboring ECs, whereas lytic EBV infection abolishes this phenotype through downregulation of TNF- $\alpha$  secretion and consequent suppression of senescence transmission (Long et al., 2016). Moreover, EBV viral oncogene latent membrane protein 1 (LMP1) promotes endosomal-exosomal pathway trafficking, which in turn suppresses NF-kB activation (Verweij et al., 2011). These data sustain the hypothesis that viruses exploit evolutionarily conserved and aging-associated mechanisms to support their spread and/or survival, and that exosomal trafficking has a central role in this interplay.

Notably, senescent cells show lysosomal enzyme deregulation, i.e. increased  $\beta$ -galactosidase activity. It has been hypothesized that lysosome malfunction could be compensated for through release of potentially toxic cargo into EVs (**Eitan et al., 2016**). Interestingly, strong SASP suppression by rapamycin is associated with decreased  $\beta$ -galactosidase expression without cell cycle re-entry (**Laberge et al., 2015**), indicating that lysosomal  $\beta$ -galactosidase is a marker of secretory activity, rather than of cell cycle arrest (**Serrano 2015**).

Overall, a greater understanding of MVB trafficking to lysosomes and the plasma membrane is expected to provide insight into diseases where pathogenic proteins, lipids, or infectious agents accumulate in or outside cells (Eitan et al., 2016). Release by senescent cells of exosomes with abnormal cargo can exert different effects on cells living in a youthful or an aged milieu. It has been reported that peripheral exosome-mediated delivery of miRNAs from a youthful systemic milieu enhanced myelination in aging brain, mimicking the effect of youth on CNS myelination (Pusic and Kraig, 2014). In this regard, parabiosis experiments that connected the circulation of an aged and a young mouse have achieved a rejuvenation effect in several tissues of the aged mouse (Horrington et al., 1960, Scudellari, 2015). However, all efforts to identify the molecular agents exerting the rejuvenation effects in the young animal's plasma have not yet met with success (Reardon, 2015). Finally, lysosomes have been reported to contain molecules that increase *Caenorhabditis elegans* lifespan (Folick et al., 2015).

#### Exosomes and circulating inflammamiRs

It is still unclear whether exosome-associated non-coding RNA, such as miRNAs and long noncoding RNAs, is merely representative of the cell of origin or whether selective loading occurs (**Zhang et al., 2015b**). Evidence has been provided that a subset of miRNAs preferentially enter exosomes (**Guduric-Fuchs et al., 2012**) and that some miRNAs are overrepresented in exosomes compared with the cell of origin (**Goldie et al., 2014**). Neural sphingomyelinase 2 (nSMase2), sumoylated heterogeneous nuclear ribonucleoproteins (hnRNPs), the 3'-end of the miRNA sequence, and miRNA-induced silencing complex (miRISC) all appear to be key players in miRNA loading into exosomes (**Zhang et al., 2015b**).

However, since aging is a complex phenotype involving multiple tissues and organs, identifying a shared subset of miRNAs, i.e. inflammamiRs (**Olivieri et al., 2015**), capable of regulating the main age-related processes and pathways could be decisive to gain insights into organismal aging. Based on this hypothesis, we have combined all available data on the miRNAs relevant to aging,

inflammation, and immunity, focusing on those that have been demonstrated to be carried by exosomes. The results are depicted in **Figure 2.** The panel of miRNAs common to the three pathways includes some important inflammamiRs, i.e. miR-19b, -20a, -21, -126, -146a, and -155. A list of the chief targets of these inflammamiRs is reported in **table 1.** DNA damage response, oxidative stress, proteotoxic stress, mitochondrial dysfunction, senescence, inflamm-aging, and nutrient sensing pathways are all directly or indirectly affected by this miRNA panel, suggesting a key role for them in organismal aging.

Notably all the miRNAs in this panel share common features: i) an altered expression in senescent cells and/or in plasma/serum or in microparticles/exosomes from patients with the major ARDs; ii) the ability to modulate inflammatory pathways, and iii) the capacity to modulate bacterial and/or viral infections in immune cells (**Oliveri et al., 2015**).

In previous papers we have reported that the circulating plasma levels of miR-21 and miR-126 increase during aging in healthy subjects (Olivieri et al., 2012; Olivieri et al., 2014). Moreover, we have described altered circulating levels of these miRNAs in a variety of ARDs (Olivieri et al., 2015). Since circulating miRNA levels are affected by a variety of factors, they do not yet have clinical diagnostic/prognostic value for ARDs (Prattichizzo et al., 2015). In this context, exosomes could provide a more accurate source of miRNA-related information. Indeed, a recent pilot study has suggested a salivary exosomal miR, miR-24-3p, as a candidate biomarker of aging (Machida et al., 2015). Moreover, emerging evidence highlight the relevance of secreted and microvesicles contained miRNAs in tissues crosstalk, in the context of ARDs. Endothelial-miR-31 can be secreted by senescent cells inside MVs and can be taken-up by mesenchymal stem cells, inhibiting osteogenic differentiation by knocking down its target Frizzled-3 (Weilner et al., 2016). Further, miR-31 circulating levels increase during aging especially in osteoporotic patients, providing a proof of principle of the relevance of exosomal miRNAs in fostering a pro-aging environment (Weilner et al., 2016).

### **Future perspectives**

Aging is not a static, but rather a dynamic phenotype resulting from continuous interactions between genetic make-up and environmental factors, including the metabioma harbored in almost all body compartments. Since it involves the entire organism, it is conceivable that health preservation during aging is ensured by efficient tissue and organ crosstalk modulating the changes induced by such interactions. Based on this hypothesis, age-related health deterioration would stem from defective tissue and organ crosstalk, with molecules circulating in the bloodstream being the main culprits. What if the circulating factors that have been identified as modulators of the aging process were "hidden" inside exosomes? Exosomes transport packets of information, rather than a single instruction, that exert a synergistic effect on target cells. Among these quanta of information, there are those produced by the inhabitants of the healthy body, especially the virome constituents. Chronic exosome parabiosis experiments, like repeated administration to an old mouse of exosomes from a healthy young mouse, could shed light on this intriguing question. Future studies are required unravel the molecular mechanisms underpinning exosome modulation during aging and the effects of such deregulation on inflamm-aging/immunosenescence, to pave the way for innovative strategies, including virome manipulation, directed at slowing down inflamm-aging and postponing ARD development.

### **Competing interests**

The authors declare that they have no competing interests.

### Acknowledgments

The authors are grateful to Word Designs for the language revision (<u>www.silviamodena.com</u>).

### Funding

This work was supported by grants from the "Università Politecnica delle Marche" to ADP and FO.

 Table 1. mRNA targets and related pathways modulated by the exosome-carried miRs

 modulating Inflammation, Immunity and Aging

Figure 1. Exosomes can play opposing roles in viral disease pathogenesis

Figure 2. Venn diagram displaying the miRs related to Inflammation, Immunity, and Aging based on their circulating shuttles

Inner circles: exosome-associated miRs; Outer circles: circulating miRs associated with Ago-2, HDL, or other microparticles. Bold characters: miRs modulating at least two pathways

#### References

Acosta JC, Banito A, Wuestefeld T, et al. A complex secretory program orchestrated by the inflammasome controls paracrine senescence. Nat Cell Biol. 2013 Aug;15(8):978-90. doi: 10.1038/ncb2784. Epub 2013 Jun 16.

Alberro A, Sáenz-Cuesta M, Muñoz-Culla M, Mateo-Abad M, Gonzalez E, Carrasco-Garcia E, Araúzo-Bravo MJ, Matheu A, Vergara I, Otaegui D. Inflammaging and Frailty Status Do Not Result in an Increased Extracellular Vesicle Concentration in Circulation. Int J Mol Sci. 2016;17. pii: E1168. doi: 10.3390/ijms17071168.

Alexander M, Hu R, Runtsch MC, Kagele DA, Mosbruger TL, Tolmachova T, Seabra MC, Round JL, Ward DM, O'Connell RM. Exosome-delivered microRNAs modulate the inflammatory response to endotoxin. Nat Commun. 2015 Jun 18;6:7321. doi: 10.1038/ncomms8321.

Almaça J, Molina J, Arrojo E Drigo R, Abdulreda MH, Jeon WB, Berggren PO, Caicedo A, Nam HG. Young capillary vessels rejuvenate aged pancreatic islets. Proc Natl Acad Sci U S A. 2014 Dec 9;111(49):17612-7. doi: 10.1073/pnas.1414053111. Epub 2014 Nov 17.

Altan-Bonnet N. Extracellular vesicles are the Trojan horses of viral infection. Curr Opin Microbiol. 2016 May 24;32:77-81. doi: 10.1016/j.mib.2016.05.004.

Baixauli F, López-Otín C, Mittelbrunn M. Exosomes and autophagy: coordinated mechanisms for the maintenance of cellular fitness. Front Immunol. 2014 Aug 20;5:403. doi: 10.3389/fimmu.2014.00403. eCollection 2014.

Batista BS, Eng WS, Pilobello KT, Hendricks-Muñoz KD, Mahal LK. Identification of a conserved glycan signature for microvesicles. J Proteome Res (2011) 10:4624–3310.1021/pr200434y [

Beveridge NJ, Tooney PA, Carroll AP, Tran N, Cairns MJ. Down-regulation of miR-17 family expression in response to retinoic acid induced neuronal differentiation. Cell Signal. 2009 Dec;21(12):1837-45. doi: 10.1016/j.cellsig.2009.07.019. Epub 2009 Aug 8.

Bhat-Nakshatri P, Wang G, Collins NR, Thomson MJ, Geistlinger TR, Carroll JS, Brown M, Hammond S, Srour EF, Liu Y, Nakshatri H. Estradiol-regulated microRNAs control estradiol response in breast cancer cells. Nucleic Acids Res. 2009 Aug;37(14):4850-61. doi: 10.1093/nar/gkp500. Epub 2009 Jun 14.

Bhaumik D, Scott GK, Schokrpur S, Patil CK, Campisi J, Benz CC. Expression of microRNA-146 suppresses NF-kappaB activity with reduction of metastatic potential in breast cancer cells. Oncogene. 2008 Sep 18;27(42):5643-7. doi: 10.1038/onc.2008.171. Epub 2008 May 26.

Buschow SI, Nolte-'t Hoen EN, van Niel G, Pols MS, ten Broeke T, Lauwen M, Ossendorp F, Melief CJ, Raposo G, Wubbolts R, Wauben MH, Stoorvogel W. MHC II in dendritic cells is targeted to lysosomes or T cell-induced exosomes via distinct multivesicular body pathways. Traffic. 2009 Oct;10(10):1528-42. doi: 10.1111/j.1600-0854.2009.00963.x.

Chahar HS, Bao X, Casola A. Exosomes and Their Role in the Life Cycle and Pathogenesis of RNA Viruses. Viruses. 2015 Jun 19;7(6):3204-25. doi: 10.3390/v7062770

Childs BG, Durik M, Baker DJ, van Deursen JM. Cellular senescence in aging and age-related disease: from mechanisms to therapy. Nat Med. 2015 Dec;21(12):1424-35. doi: 10.1038/nm.4000. Review.

Christianson HC, Svensson KJ, van Kuppevelt TH, Li JP, Belting M. Cancer cell exosomes depend on cell-surface heparan sulfate proteoglycans for their internalization and functional activity. Proc Natl Acad Sci U S A. (2013) 110(43):17380–510.1073/pnas.1304266110

de Jong OG, Verhaar MC, Chen Y, Vader P, Gremmels H, Posthuma G, Schiffelers RM, Gucek M, van Balkom BW. Cellular stress conditions are reflected in the protein and RNA content of endothelial cell-derived exosomes. J Extracell Vesicles. 2012 Apr 16;1. doi: 10.3402/jev.v1i0.18396. eCollection 2012.

Di Mitri D, Alimonti A. Non-Cell-Autonomous Regulation of Cellular Senescence in Cancer. Trends Cell Biol. 2016 Mar;26(3):215-26. doi: 10.1016/j.tcb.2015.10.005. Epub 2015 Nov 9. Review.

Dinami R, Ercolani C, Petti E, et al. miR-155 drives telomere fragility in human breast cancer by targeting TRF1. Cancer Res. 2014 Aug 1;74(15):4145-56. doi: 10.1158/0008-5472.CAN-13-2038.

Effenberger T, von der Heyde J, Bartsch K, Garbers C, Schulze-Osthoff K, Chalaris A, Murphy G, Rose-John S, Rabe B. Senescence-associated release of transmembrane proteins involves proteolytic processing by ADAM17 and microvesicle shedding. FASEB J. 2014 Nov; 28(11): 4847-56. doi: 10.1096/fj.14-254565.

Eitan E, Suire C, Zhang S, Mattson MP. Impact of Lysosome Status on Extracellular Vesicle Content and Release. Ageing Res Rev. 2016 May 26. pii: S1568-1637(16)30077-0. doi: 10.1016/j.arr.2016.05.001.

Eken C, Martin PJ, Sadallah S, Treves S, Schaller M, Schifferli JA. Ectosomes released by polymorphonuclear neutrophils induce a MerTK-dependent anti-inflammatory pathway in macrophages. J Biol Chem. 2010 Dec 17;285(51):39914-21. doi: 10.1074/jbc.M110.126748.

Fabbri M, Paone A, Calore F, et al. MicroRNAs bind to Toll-like receptors to induce prometastatic inflammatory response. Proc Natl Acad Sci U S A. 2012 Jul 31;109(31):E2110-6. doi: 10.1073/pnas.1209414109. Epub 2012 Jul 2.

Felli N, Felicetti F, Lustri AM, Errico MC, Bottero L, Cannistraci A, De Feo A, Petrini M, Pedini F, Biffoni M, Alvino E, Negrini M, Ferracin M, Mattia G, Carè A. miR-126&126\* restored expressions play a tumor suppressor role by directly regulating ADAM9 and MMP7 in melanoma. PLoS One. 2013;8(2):e56824. doi: 10.1371/journal.pone.0056824. Epub 2013 Feb 21.

Fish JE, Santoro MM, Morton SU, Yu S, Yeh RF, Wythe JD, Ivey KN, Bruneau BG, Stainier DY, Srivastava D. miR-126 regulates angiogenic signaling and vascular integrity. Dev Cell. 2008 Aug;15(2):272-84. doi: 10.1016/j.devcel.2008.07.008.

Folick A, Oakley HD, Yu Y, Armstrong EH, Kumari M, Sanor L, Moore DD, Ortlund EA, Zechner R, Wang MC. Aging. Lysosomal signaling molecules regulate longevity in Caenorhabditis elegans. Science. 2015 Jan 2;347(6217):83-6. doi: 10.1126/science.1258857.

Franceschi C, Bonafè M, Valensin S, Olivieri F, De Luca M, Ottaviani E, De Benedictis G. Inflamm-aging. An evolutionary perspective on immunosenescence. Ann N Y Acad Sci. 2000 Jun;908:244-54.

Fancello L, Monteil S, Popgeorgiev N, Rivet R, Gouriet F, Fournier PE, Raoult D, Desnues C. Viral communities associated with human pericardial fluids in idiopathic pericarditis. PLoS One. 2014 Apr 1;9(4):e93367. doi: 10.1371/journal.pone.0093367. eCollection 2014.

Frühbeis C, Fröhlich D, Kuo WP, Krämer-Albers EM. Extracellular vesicles as mediators of neuron-glia communication. Front Cell Neurosci. 2013 Oct 30;7:182. doi: 10.3389/fncel.2013.00182.

Fulop T, Witkowski JM, Le Page A, Fortin C, Pawelec G, Larbi A. Intracellular signaling pathways: Targets to reverse immunosenescence. Clin Exp Immunol. 2016 Jul 1. doi: 10.1111/cei.12836.

Gabriely G, Wurdinger T, Kesari S, Esau CC, Burchard J, Linsley PS, Krichevsky AM. MicroRNA 21 promotes glioma invasion by targeting matrix metalloproteinase regulators. Mol Cell Biol. 2008 Sep;28(17):5369-80. doi: 10.1128/MCB.00479-08. Epub 2008 Jun 30.

Garcia NA, Ontoria-Oviedo I, González-King H, Diez-Juan A, Sepúlveda P.Glucose Starvation in Cardiomyocytes Enhances Exosome Secretion and Promotes Angiogenesis in Endothelial Cells. PLoS One. 2015 Sep 22;10(9):e0138849. doi: 10.1371/journal.pone.0138849. eCollection 2015.

Goldie BJ, Dun MD, Lin M, Smith ND, Verrills NM, Dayas CV, Cairns MJ. Activity-associated miRNA are packaged in Map1b-enriched exosomes released from depolarized neurons. Nucleic Acids Res. 2014 Aug;42(14):9195-208. doi: 10.1093/nar/gku594. Epub 2014 Jul 22.

Gibcus JH, Tan LP, Harms G, Schakel RN, de Jong D, Blokzijl T, Möller P, Poppema S, Kroesen BJ, van den Berg A. Hodgkin lymphoma cell lines are characterized by a specific miRNA expression profile. Neoplasia. 2009 Feb;11(2):167-76.

Gironella M, Seux M, Xie MJ, et al. Tumor protein 53-induced nuclear protein 1 expression is repressed by miR-155, and its restoration inhibits pancreatic tumor development. Proc Natl Acad Sci U S A. 2007 Oct 9;104(41):16170-5. Epub 2007 Oct 2.

Goetzl EJ, Goetzl L, Karliner JS, Tang N4, Pulliam L. Human plasma platelet-derived exosomes: effects of aspirin. FASEB J. 2016 May;30(5):2058-63. doi: 10.1096/fj.201500150R.

Guduric-Fuchs J, O'Connor A, Camp B, O'Neill CL, Medina RJ, Simpson DA. Selective extracellular vesicle-mediated export of an overlapping set of microRNAs from multiple cell types. BMC Genomics. 2012 Aug 1;13:357. doi: 10.1186/1471-2164-13-357.

Gutzeit C, Nagy N, Gentile M, Lyberg K, Gumz J, Vallhov H, Puga I, Klein E, Gabrielsson S, Cerutti A, Scheynius A. Exosomes derived from Burkitt's lymphoma cell lines induce proliferation,

differentiation, and class-switch recombination in B cells. J Immunol. 2014 Jun 15;192(12):5852-62. doi: 10.4049/jimmunol.1302068. Epub 2014 May 14.

Hackl M, Brunner S, Fortschegger K, et al. miR-17, miR-19b, miR-20a, and miR-106a are downregulated in human aging. Aging Cell. 2010 Apr;9(2):291-6. doi: 10.1111/j.1474-9726.2010.00549.x. Epub 2010 Jan 18.

Harris TA, Yamakuchi M, Ferlito M, Mendell JT, Lowenstein CJ. MicroRNA-126 regulates endothelial expression of vascular cell adhesion molecule 1. Proc Natl Acad Sci U S A. 2008 Feb 5;105(5):1516-21. doi: 10.1073/pnas.0707493105. Epub 2008 Jan 28.

Hébert SS, Horré K, Nicolaï L, Papadopoulou AS, Mandemakers W, Silahtaroglu AN, Kauppinen S, Delacourte A, De Strooper B. Loss of microRNA cluster miR-29a/b-1 in sporadic Alzheimer's disease correlates with increased BACE1/beta-secretase expression. Proc Natl Acad Sci U S A. 2008 Apr 29;105(17):6415-20. doi: 10.1073/pnas.0710263105. Epub 2008 Apr 23.

Han Z, Liu X, Chen X, Zhou X, Du T, Roizman B, Zhou G. miR-H28 and miR-H29 expressed late in productive infection are exported and restrict HSV-1 replication and spread in recipient cells. Proc Natl Acad Sci U S A. 2016 Feb 16;113(7):E894-901. doi: 10.1073/pnas.1525674113. Epub 2016 Feb 1.

Horrington EM, Pope F, Lunsford W, McCay CM. Age changes in the bones, blood pressure, and diseases of rats in parabiosis. Gerontologia. 1960;4:21-31.

Iraci N, Leonardi T, Gessler F, Vega B, Pluchino S. Focus on Extracellular Vesicles: Physiological Role and Signalling Properties of Extracellular Membrane Vesicles. Int J Mol Sci. 2016 Feb 6;17(2). pii: E171. doi: 10.3390/ijms17020171.

Jiang S, Zhang HW, Lu MH, He XH, Li Y, Gu H, Liu MF, Wang ED. MicroRNA-155 functions as an OncomiR in breast cancer by targeting the suppressor of cytokine signaling 1 gene. Cancer Res. 2010 Apr 15;70(8):3119-27. doi: 10.1158/0008-5472.CAN-09-4250. Epub 2010 Mar 30.

Jiao LR, Frampton AE, Jacob J, Pellegrino L, Krell J, Giamas G, Tsim N, Vlavianos P, Cohen P, Ahmad R, Keller A, Habib NA, Stebbing J, Castellano L. MicroRNAs targeting oncogenes are down-regulated in pancreatic malignant transformation from benign tumors. PLoS One. 2012;7(2):e32068. doi: 10.1371/journal.pone.0032068. Epub 2012 Feb 22.

Johnstone RM. Exosomes biological significance: a concise review. Blood Cells Mol Dis 2006; 36:315–21.

Jurkin J, Schichl YM, Koeffel R, Bauer T, Richter S, Konradi S, Gesslbauer B, Strobl H. miR-146a is differentially expressed by myeloid dendritic cell subsets and desensitizes cells to TLR2dependent activation. J Immunol. 2010 May 1;184(9):4955-65. doi: 10.4049/jimmunol.0903021. Epub 2010 Apr 7.

Kalamvoki M, Du T, Roizman B. Cells infected with herpes simplex virus 1 export to uninfected cells exosomes containing STING, viral mRNAs, and microRNAs. Proc Natl Acad Sci U S A. 2014 Nov 18;111(46):E4991-6. doi: 10.1073/pnas.1419338111. Epub 2014 Nov 3.

Kooijmans SA, Schiffelers RM, Zarovni N, Vago R. Modulation of tissue tropism and biological activity of exosomes and other extracellular vesicles: New nanotools for cancer treatment. Pharmacol Res. 2016 Jul 6;111:487-500. doi: 10.1016/j.phrs.2016.07.006.

Kong W, He L, Coppola M, Guo J, Esposito NN, Coppola D, Cheng JQ. MicroRNA-155 regulates cell survival, growth, and chemosensitivity by targeting FOXO3a in breast cancer. J Biol Chem. 2010 Jun 4;285(23):17869-79. doi: 10.1074/jbc.M110.101055. Epub 2010 Apr 6.

Kuilman T, Peeper DS. Senescence-messaging secretome: SMS-ing cellular stress. Nat Rev Cancer. 2009 Feb;9(2):81-94. doi: 10.1038/nrc2560. Epub 2009 Jan 9. Review.

Labbaye C, Spinello I, Quaranta MT, Pelosi E, Pasquini L, Petrucci E, Biffoni M, Nuzzolo ER, Billi M, Foà R, Brunetti E, Grignani F, Testa U, Peschle C. A three-step pathway comprising PLZF/miR-146a/CXCR4 controls megakaryopoiesis. Nat Cell Biol. 2008 Jul;10(7):788-801. doi: 10.1038/ncb1741. Epub 2008 Jun 22.

Laberge RM, Sun Y, Orjalo AV, et al. MTOR regulates the pro-tumorigenic senescence-associated secretory phenotype by promoting IL1A translation. Nat Cell Biol. 2015 Aug;17(8):1049-61. doi: 10.1038/ncb3195. Epub 2015 Jul 6.

Lecot P, Alimirah F, Desprez PY, Campisi J, Wiley C. Context-dependent effects of cellular senescence in cancer development. Br J Cancer. 2016 May 24;114(11):1180-4. doi: 10.1038/bjc.2016.115. Epub 2016 May 3.

Lehmann BD, Paine MS, Brooks AM, McCubrey JA, Renegar RH, Wang R, Terrian DM. Senescence-associated exosome release from human prostate cancer cells. Cancer Res. 2008 Oct 1;68(19):7864-71. doi: 10.1158/0008-5472.CAN-07-6538.

Liu B, Peng XC, Zheng XL, Wang J, Qin YW. MiR-126 restoration down-regulate VEGF and inhibit the growth of lung cancer cell lines in vitro and in vivo. Lung Cancer. 2009 Nov;66(2):169-75. doi: 10.1016/j.lungcan.2009.01.010. Epub 2009 Feb 14.

Liu SQ, Jiang S, Li C, Zhang B, Li QJ. miR-17-92 cluster targets phosphatase and tensin homology and Ikaros Family Zinc Finger 4 to promote TH17-mediated inflammation. J Biol Chem. 2014 May 2;289(18):12446-56. doi: 10.1074/jbc.M114.550723. Epub 2014 Mar 18.

Liu Z, Zhang X, Yu Q, He JJ. Exosome-associated hepatitis C virus in cell cultures and patient plasma. Biochem Biophys Res Commun. 2014b Dec 12;455(3-4):218-22. doi: 10.1016/j.bbrc.2014.10.146.

Long X, Li Y, Yang M, Huang L, Gong W, Kuang E. BZLF1 Attenuates Transmission of Inflammatory Paracrine Senescence in Epstein-Barr Virus-Infected Cells by Downregulating TNFα. J Virol. 2016 Jun 22. pii: JVI.00999-16. [Epub ahead of print]

Lundy SK, Klinker MW, Fox DA. Killer B lymphocytes and their fas ligand positive exosomes as inducers of immune tolerance. Front Immunol. 2015: 20;6:122. doi: 10.3389/fimmu.2015.00122. eCollection 2015.

Machida T, Tomofuji T, Ekuni D, Maruyama T, Yoneda T, Kawabata Y, Mizuno H, Miyai H, Kunitomo M, Morita M. MicroRNAs in Salivary Exosome as Potential Biomarkers of Aging. Int J Mol Sci. 2015 Sep 7;16(9):21294-309. doi: 10.3390/ijms160921294.

Marti M, Johnson PJ. Emerging roles for extracellular vesicles in parasitic infections. Curr Opin Microbiol. 2016 May 18;32:66-70. doi: 10.1016/j.mib.2016.04.008.

Mashburn LM, Whiteley M. Membrane vesicles traffic signals and facilitate group activities in a prokaryote. Nature 2005; 437:422–5.

Meckes DG Jr. Exosomal communication goes viral. J Virol. 2015 May;89(10):5200-3. doi: 10.1128/JVI.02470-14.

Meng F, Henson R, Wehbe-Janek H, Ghoshal K, Jacob ST, Patel T. MicroRNA-21 regulates expression of the PTEN tumor suppressor gene in human hepatocellular cancer. Gastroenterology. 2007 Aug;133(2):647-58. Epub 2007 May 21.

Mestdagh P, Boström AK, Impens F, et al. The miR-17-92 microRNA cluster regulates multiple components of the TGF- $\beta$  pathway in neuroblastoma. Mol Cell. 2010 Dec 10;40(5):762-73. doi: 10.1016/j.molcel.2010.11.038.

Mitsuhashi M, Taub DD, Kapogiannis D, Eitan E, Zukley L, Mattson MP, Ferrucci L, Schwartz JB, Goetzl EJ. Aging enhances release of exosomal cytokine mRNAs by Aβ1-42-stimulated macrophages. FASEB J. 2013 Dec;27(12):5141-50. doi: 10.1096/fj.13-238980. Epub 2013 Sep 6.

Monguió-Tortajada M, Lauzurica-Valdemoros R, Borràs FE. Tolerance in organ transplantation: from conventional immunosuppression to extracellular vesicles. Front Immunol. 2014 Sep 17;5:416. doi: 10.3389/fimmu.2014.00416. eCollection 2014.

Morelli AE, Larregina AT, Shufesky WJ, et al. Endocytosis, intracellular sorting, and processing of exosomes by dendritic cells. Blood 2004;104:3257–66.

Nolte-'t Hoen E, Cremer T, Gallo RC, Margolis LB. Extracellular vesicles and viruses: Are they close relatives? Proc Natl Acad Sci U S A. 2016 Jul 18. pii: 201605146

Oglesby IK, Bray IM, Chotirmall SH, Stallings RL, O'Neill SJ, McElvaney NG, Greene CM. miR-126 is downregulated in cystic fibrosis airway epithelial cells and regulates TOM1 expression. J Immunol. 2010 Feb 15;184(4):1702-9. doi: 10.4049/jimmunol.0902669. Epub 2010 Jan 18.

Ohno S, Drummen GP, Kuroda M. Focus on Extracellular Vesicles: Development of Extracellular Vesicle-Based Therapeutic Systems. Int J Mol Sci. 2016 Feb 6;17(2). pii: E172. doi: 10.3390/ijms17020172.

Olivieri F, Spazzafumo L, Santini G, et al. Age-related differences in the expression of circulating microRNAs: miR-21 as a new circulating marker of inflammaging. Mech Ageing Dev. 2012 Nov-Dec;133(11-12):675-85. doi: 10.1016/j.mad.2012.09.004. Epub 2012 Oct 2.

Olivieri F, Bonafè M, Spazzafumo L, et al. Age- and glycemia-related miR-126-3p levels in plasma and endothelial cells. Aging (Albany NY). 2014 Sep;6(9):771-87.

Olivieri F, Albertini MC, Orciani M, Ceka A, Cricca M, Procopio AD, Bonafè M. DNA damage response (DDR) and senescence: shuttled inflamma-miRNAs on the stage of inflamm-aging. Oncotarget. 2015 Nov 3;6(34):35509-21. doi: 10.18632/oncotarget.5899. Review.

Ostman S, Taube M, Telemo E. Tolerosome-induced oral tolerance is MHC dependent. Immunology. 2005 Dec;116(4):464-76.

Pêche H, Renaudin K, Beriou G, Merieau E, Amigorena S, Cuturi MC. Induction of tolerance by exosomes and short-term immunosuppression in a fully MHC-mismatched rat cardiac allograft model. Am J Transplant. 2006 Jul;6(7):1541-50.

Peche H, Heslan M, Usal C, Amigorena S, Cuturi MC. Presentation of donor major histocompatibility complex antigens by bone marrow dendritic cell-derived exosomes modulates allograft rejection. Transplantation 2003; 76: 1503–1510.

Philippe L, Alsaleh G, Suffert G, Meyer A, Georgel P, Sibilia J, Wachsmann D, Pfeffer S. TLR2 expression is regulated by microRNA miR-19 in rheumatoid fibroblast-like synoviocytes. J Immunol. 2012 Jan 1;188(1):454-61. doi: 10.4049/jimmunol.1102348. Epub 2011 Nov 21.

Pin AL, Houle F, Guillonneau M, Paquet ER, Simard MJ, Huot J. miR-20a represses endothelial cell migration by targeting MKK3 and inhibiting p38 MAP kinase activation in response to VEGF. Angiogenesis. 2012 Dec;15(4):593-608. doi: 10.1007/s10456-012-9283-z. Epub 2012 Jun 14.

Png KJ, Halberg N, Yoshida M, Tavazoie SF. A microRNA regulon that mediates endothelial recruitment and metastasis by cancer cells. Nature. 2011 Dec 14;481(7380):190-4. doi: 10.1038/nature10661.

Prattichizzo F, Giuliani A, Ceka A, Rippo MR, Bonfigli AR, Testa R, Procopio AD, Olivieri F. Epigenetic mechanisms of endothelial dysfunction in type 2 diabetes. Clin Epigenetics. 2015 May 23;7:56. doi: 10.1186/s13148-015-0090-4. eCollection 2015.

Prattichizzo F, Giuliani A, De Nigris V, Pujadas G, Ceka A, La Sala L, Genovese S, Testa R, Procopio AD, Olivieri F, Ceriello A. Extracellular microRNAs and endothelial hyperglycemic memory: a therapeutic opportunity? Diabetes Obes Metab. 2016 May 10. doi: 10.1111/dom.12688

Pusic AD, Kraig RP. Youth and environmental enrichment generate serum exosomes containing miR-219 that promote CNS myelination. Glia. 2014 Feb;62(2):284-99. doi: 10.1002/glia.22606

Qian Z, Shen Q, Yang X, Qiu Y, Zhang W. The Role of Extracellular Vesicles: An Epigenetic View of the Cancer Microenvironment. Biomed Res Int. 2015;2015:649161. doi: 10.1155/2015/649161. Epub 2015 Oct 25.

Ratajczak MZ, Ratajczak J. Horizontal transfer of RNA and proteins between cells by extracellular microvesicles: 14 years later. Clin Transl Med. 2016;5:7. doi: 10.1186/s40169-016-0087-4.

Reardon S. 'Young blood' anti-ageing mechanism called into question. doi:10.1038/nature.2015.17583

Salminen A, Kauppinen A, Kaarniranta K. Emerging role of NF-κB signaling in the induction of senescence-associated secretory phenotype (SASP). Cell Signal. 2012 Apr;24(4):835-45. doi: 10.1016/j.cellsig.2011.12.006. Epub 2011 Dec 11. Review.

Schorey JS, Harding CV. Extracellular vesicles and infectious diseases: new complexity to an old story. J Clin Invest. 2016 Apr 1;126(4):1181-9. doi: 10.1172/JCI81132.

Schorey JS, Cheng Y, Singh PP, Smith VL. Exosomes and other extracellular vesicles in hostpathogen interactions. EMBO Rep. 2015;16:24-43. doi: 10.15252/embr.201439363.

Schwab A, Meyering SS, Lepene B, Iordanskiy S, van Hoek ML, Hakami RM, Kashanchi F. Extracellular vesicles from infected cells: potential for direct pathogenesis. Front Microbiol. 2015 Oct 20;6:1132. doi: 10.3389/fmicb.2015.01132. eCollection 2015.

Scudellari M. Ageing research: Blood to blood. Nature. 2015 Jan 22;517(7535):426-9. doi: 10.1038/517426a. No abstract available.

Segura E, Nicco C, Lombard B et al. ICAM-1 on exosomes from mature dendritic cells is critical for efficient naive T-cell priming. Blood 2005; 106: 216–223.

Segura E, Amigorena S, Théry C. Mature dendritic cells secrete exosomes with strong ability to induce antigen-specific effector immune responses. Blood Cells Mol Dis. 2005 Sep-Oct;35(2):89-93.

Serrano M. The InflammTORy Powers of Senescence. Trends Cell Biol. 2015 Nov;25(11):634-6. doi: 10.1016/j.tcb.2015.09.011. Epub 2015 Oct 21.

Shimoda A, Ueda K, Nishiumi S, Murata-Kamiya N, Mukai SA, Sawada S, Azuma T, Hatakeyama M, Akiyoshi K. Exosomes as nanocarriers for systemic delivery of the Helicobacter pylori virulence factor CagA. Sci Rep. 2016 Jan 7;6:18346. doi: 10.1038/srep18346.

Si ML, Zhu S, Wu H, Lu Z, Wu F, Mo YY. miR-21-mediated tumor growth. Oncogene. 2007 Apr 26;26(19):2799-803. Epub 2006 Oct 30.

Silva M, Melo SA. Non-coding RNAs in Exosomes: New Players in Cancer Biology. Curr Genomics. 2015 Oct;16(5):295-303. doi: 10.2174/1389202916666150707154719

Skogberg G, Telemo E, Ekwall O. Exosomes in the Thymus: Antigen Transfer and Vesicles. Front Immunol. 2015 Jul 20;6:366. doi: 10.3389/fimmu.2015.00366. eCollection 2015

Smythies J, Edelstein L. Transsynaptic modality codes in the brain: possible involvement of synchronized spike timing, microRNAs, exosomes and epigenetic processes. Front Intregr Neurosci (2013) 2012(6):126.10.3389/fnint.2012.00126

Sobo-Vujanovic A, Munich S, Vujanovic NL. Dendritic-cell exosomes cross-present Toll-like receptor-ligands and activate bystander dendritic cells. Cell Immunol. 2014;289:119-27. doi: 10.1016/j.cellimm.2014.03.016.

Sun Y, Zheng W, Guo Z, Ju Q, Zhu L, Gao J, Zhou L, Liu F, Xu Y, Zhan Q, Zhou Z, Sun W, Zhao X. A novel TP53 pathway influences the HGS-mediated exosome formation in colorectal cancer. Sci Rep. 2016 Jun 17;6:28083. doi: 10.1038/srep28083.

Sylvestre Y, De Guire V, Querido E, Mukhopadhyay UK, Bourdeau V, Major F, Ferbeyre G, Chartrand P. An E2F/miR-20a autoregulatory feedback loop. J Biol Chem. 2007 Jan 26;282(4):2135-43. Epub 2006 Nov 29.

Taganov KD, Boldin MP, Chang KJ, Baltimore D. NF-kappaB-dependent induction of microRNA miR-146, an inhibitor targeted to signaling proteins of innate immune responses. Proc Natl Acad Sci U S A. 2006 Aug 15;103(33):12481-6. Epub 2006 Aug 2

Tang N, Sun B, Gupta A, Rempel H, Pulliam L. Monocyte exosomes induce adhesion molecules and cytokines via activation of NF-κB in endothelial cells. FASEB J. 2016 May 25. pii: fj.201600368RR. [Epub ahead of print]

Terao M, Fratelli M, Kurosaki M, Zanetti A, Guarnaccia V, Paroni G, Tsykin A, Lupi M, Gianni M, Goodall GJ, Garattini E. Induction of miR-21 by retinoic acid in estrogen receptor-positive breast carcinoma cells: biological correlates and molecular targets. J Biol Chem. 2011 Feb 4;286(5):4027-42. doi: 10.1074/jbc.M110.184994. Epub 2010 Dec 3.

Thakur BK, Zhang H, Becker A, et al. Double-stranded DNA in exosomes: a novel biomarker in cancer detection. Cell Res. 2014 Jun;24(6):766-9. doi: 10.1038/cr.2014.44

Théry C, Amigorena S. The cell biology of antigen presentation in dendritic cells. Curr Opin Immunol. 2001 Feb;13(1):45-51

Thery C, Zitvogel L, Amigorena S. Exosomes: composition, biogenesis and function. Nat Rev Immunol 2002; 2:569–79.

Thery C, Ostrowski M, Segura E. Membrane vesicles as conveyors of immune responses. Nat Rev Immunol 2009; 9:581–93.

Turchinovich A, Tonevitsky AG, Cho WC, Burwinkel B. Check and mate to exosomal extracellular miRNA: new lesson from a new approach. Front Mol Biosci. 2015 Apr 13;2:11. doi: 10.3389/fmolb.2015.00011. eCollection 2015.

Urbanelli L, Magini A, Buratta S, Brozzi A, Sagini K, Polchi A, Tancini B, Emiliani C. Signaling pathways in exosomes biogenesis, secretion and fate. Genes (Basel). 2013 Mar 28;4(2):152-70. doi: 10.3390/genes4020152.

Vallhov H, Gutzeit C, Hultenby K, Valenta R, Grönlund H, Scheynius A. Dendritic cell-derived exosomes carry the major cat allergen Fel d 1 and induce an allergic immune response. Allergy. 2015 Dec;70(12):1651-5. doi: 10.1111/all.12701. Epub 2015 Aug 25.

Van Dongen HM, Masoumi N, Witwer KW, Pegtel DM. Extracellular Vesicles Exploit Viral Entry Routes for Cargo Delivery. Microbiol Mol Biol Rev. 2016 Mar 2;80(2):369-86. doi: 10.1128/MMBR.00063-15.

Verweij FJ, van Eijndhoven MA, Hopmans ES, Vendrig T, Wurdinger T, Cahir-McFarland E, Kieff E, Geerts D, van der Kant R, Neefjes J, Middeldorp JM, Pegtel DM. LMP1 association with CD63

in endosomes and secretion via exosomes limits constitutive NF-κB activation. EMBO J. 2011 Jun 1;30(11):2115-29. doi: 10.1038/emboj.2011.123. Epub 2011 Apr 28.

Virgin HW. The virome in mammalian physiology and disease. Cell. 2014 Mar 27;157(1):142-50. doi: 10.1016/j.cell.2014.02.032.

Wang Z, Lieberman PM. The crosstalk of telomere dysfunction and inflammation through cell-free TERRA containing exosomes. RNA Biol. 2016 Jun 28:1-6.

Wang S, Zhang X, Ju Y, Zhao B, Yan X, Hu J, Shi L, Yang L, Ma Z, Chen L, Liu Y, Duan Z, Chen X, Meng S. MicroRNA-146a feedback suppresses T cell immune function by targeting Stat1 in patients with chronic hepatitis B. J Immunol. 2013 Jul 1;191(1):293-301. doi: 10.4049/jimmunol.1202100. Epub 2013 May 22.

Weilner S, Schraml E, Redl H, Grillari-Voglauer R, Grillari J. Secretion of microvesicular miRNAs in cellular and organismal aging. Exp Gerontol. 2013 Jul;48(7):626-33. doi: 10.1016/j.exger.2012.11.017.

Weilner S, Schraml E, Wieser M, Messner P, Schneider K, Wassermann K, Micutkova L, Fortschegger K, Maier AB, Westendorp R, Resch H, Wolbank S, Redl H, Jansen-Dürr P, Pietschmann P, Grillari-Voglauer R, Grillari J. Secreted microvesicular miR-31 inhibits osteogenic differentiation of mesenchymal stem cells. Aging Cell. 2016 Aug;15(4):744-54. doi: 10.1111/acel.12484.

Willms E, Johansson HJ, Mäger I, Lee Y, Blomberg KE, Sadik M, Alaarg A, Smith CI, Lehtiö J, El Andaloussi S, Wood MJ, Vader P. Cells release subpopulations of exosomes with distinct molecular and biological properties. Sci Rep. 2016 Mar 2;6:22519. doi: 10.1038/srep22519.

Wylie KM, Mihindukulasuriya KA, Zhou Y, Sodergren E, Storch GA, Weinstock GM. Metagenomic analysis of double-stranded DNA viruses in healthy adults. BMC Biol. 2014 Sep 10;12:71. doi: 10.1186/s12915-014-0071-7.

Xu D, Tahara H. The role of exosomes and microRNAs in senescence and aging. Adv Drug Deliv Rev. 2013 Mar;65(3):368-75. doi: 10.1016/j.addr.2012.07.010.

Xue W, Zender L, Miething C, Dickins RA, Hernando E, Krizhanovsky V, Cordon-Cardo C, Lowe SW. Senescence and tumour clearance is triggered by p53 restoration in murine liver carcinomas. Nature. 2007 Feb 8;445(7128):656-60. Epub 2007 Jan 24. Erratum in: Nature. 2011 May 26;473(7348):544.

Yang K, He YS, Wang XQ, Lu L, Chen QJ, Liu J, Sun Z, Shen WF. MiR-146a inhibits oxidized low-density lipoprotein-induced lipid accumulation and inflammatory response via targeting toll-like receptor 4. FEBS Lett. 2011 Mar 23;585(6):854-60. doi: 10.1016/j.febslet.2011.02.009. Epub 2011 Feb 15.

Yang JC, Lin MW, Rau CS, Jeng SF, Lu TH, Wu YC, Chen YC, Tzeng SL, Wu CJ, Hsieh CH. Altered exosomal protein expression in the serum of NF-κB knockout mice following skeletal muscle ischemia-reperfusion injury. J Biomed Sci. 2015 Jun 10;22:40. doi: 10.1186/s12929-015-0147-x.

Yin Q, McBride J, Fewell C, Lacey M, Wang X, Lin Z, Cameron J, Flemington EK. MicroRNA-155 is an Epstein-Barr virus-induced gene that modulates Epstein-Barr virus-regulated gene expression pathways. J Virol. 2008 Jun;82(11):5295-306. doi: 10.1128/JVI.02380-07. Epub 2008 Mar 26.

Yuan MJ, Maghsoudi T, Wang T. Exosomes Mediate the Intercellular Communication after Myocardial Infarction. Int J Med Sci. 2016;13(2):113-6. doi: 10.7150/ijms.14112. eCollection 2016.

Zhao S, Wang Y, Liang Y, Zhao M, Long H, Ding S, Yin H, Lu Q. MicroRNA-126 regulates DNA methylation in CD4+ T cells and contributes to systemic lupus erythematosus by targeting DNA methyltransferase 1. Arthritis Rheum. 2011 May;63(5):1376-86. doi: 10.1002/art.30196.

Zhao H, Yang L, Baddour J, et al. Tumor microenvironment derived exosomes pleiotropically modulate cancer cell metabolism. Elife. 2016 Feb 27;5. pii: e10250. doi: 10.7554/eLife.10250.

Zhang J, Du YY, Lin YF, Chen YT, Yang L, Wang HJ, Ma D. The cell growth suppressor, mir-126, targets IRS-1. Biochem Biophys Res Commun. 2008 Dec 5;377(1):136-40. doi: 10.1016/j.bbrc.2008.09.089. Epub 2008 Oct 1.

Zhang Y, Liu D, Chen X, et al. Secreted monocytic miR-150 enhances targeted endothelial cell migration. Mol Cell. 2010 Jul 9;39(1):133-44. doi: 10.1016/j.molcel.2010.06.010.

Zhang M, Liu Q, Mi S, Liang X, Zhang Z, Su X, Liu J, Chen Y, Wang M, Zhang Y, Guo F, Zhang Z, Yang R. Both miR-17-5p and miR-20a alleviate suppressive potential of myeloid-derived suppressor cells by modulating STAT3 expression. J Immunol. 2011 Apr 15;186(8):4716-24. doi: 10.4049/jimmunol.1002989. Epub 2011 Mar 7.

Zhang X, Ng WL, Wang P, Tian L, Werner E, Wang H, Doetsch P, Wang Y. MicroRNA-21 modulates the levels of reactive oxygen species by targeting SOD3 and TNFα. Cancer Res. 2012 Sep 15;72(18):4707-13. doi: 10.1158/0008-5472.CAN-12-0639. Epub 2012 Jul 25.

Zhang Y, Yang P, Sun T, et al. miR-126 and miR-126\* repress recruitment of mesenchymal stem cells and inflammatory monocytes to inhibit breast cancer metastasis. Nat Cell Biol. 2013 Mar;15(3):284-94. doi: 10.1038/ncb2690. Epub 2013 Feb 10.

Zhang B, Yin Y, Lai RC, Lim SK. Immunotherapeutic potential of extracellular vesicles. Front Immunol. 2014 Oct 22;5:518. doi: 10.3389/fimmu.2014.00518. eCollection 2014

Zhang L, Zhang S, Yao J, et al. Microenvironment-induced PTEN loss by exosomal microRNA primes brain metastasis outgrowth. Nature. 2015 Nov 5;527(7576):100-4. doi: 10.1038/nature15376. Epub 2015 Oct 19.

Zhang J, Li S, Li L, Li M, Guo C, Yao J, Mi S. Exosome and exosomal microRNA: trafficking, sorting, and function. Genomics Proteomics Bioinformatics. 2015 Feb;13(1):17-24. doi: 10.1016/j.gpb.2015.02.001. Epub 2015 Feb 24. Review.

Zhang Y, Zheng L, Ding Y, Li Q, Wang R, Liu T, Sun Q, Yang H, Peng S, Wang W, Chen L. MiR-20a Induces Cell Radioresistance by Activating the PTEN/PI3K/Akt Signaling Pathway in Hepatocellular Carcinoma. Int J Radiat Oncol Biol Phys. 2015 Aug 1;92(5):1132-40. doi: 10.1016/j.ijrobp.2015.04.007. Epub 2015 Apr 13.

Zhang B, Yeo RW, Tan KH, Lim SK. Focus on Extracellular Vesicles: Therapeutic Potential of Stem Cell-Derived Extracellular Vesicles. Int J Mol Sci. 2016;17(2). pii: E174. doi: 10.3390/ijms17020174.

Zhou J, Guo F, Wang G, Wang J, Zheng F, Guan X, Chang A, Zhang X, Dai C, Li S, Li X, Wang B. miR-20a regulates adipocyte differentiation by targeting lysine-specific demethylase 6b and transforming growth factor- $\beta$  signaling. Int J Obes (Lond). 2015 Aug;39(8):1282-91. doi: 10.1038/ijo.2015.43. Epub 2015 Mar 30.

Table 1. mRNA targets and related pathways modulated by the exosome-carried miRs modulating Inflammation, Immunity and Aging

MiRNA	mRNA targets	Related pathways	References
19b	BACE1, PTEN, TGFB-	Proteostasis, tumor	Hébert et al., 2008; Liu et al.,
	R2, TLR-2.	biology, senescence,	2014; Mestdagh et al., 2010;
		SASP, immune cytokine	Philippe et al., 2012.
		signaling, response	

		signaling, response	
20a	E2F1, E2F3, CDKN1A (p21), Stat3, TGFB-R2, PTEN, MAP3K12, MAP2K3.	Senescence, SASP, cytokine signaling, tumor biology, immune response	Sylvestre et al., 2007; Hackl et al., 2010; Zhang et al., 2011; Zhou et al., 2015; Zhang et al., 2015c; Pin et al., 2012; Beveridge et al., 2009.
21	IL-1B, ICAM1, SOD3, BCL2, TGFBR-2, TGFB- I, E2F1, PTEN.	Inflammasome, senescence, SASP, cytokine signaling, immune response, oxidative stress, tumor biology	Terao et al., 2011; Zhang et al., 2012; Si et al., 2007; Gabriely et al., 2008; Bhat- Nakshatri et al., 2009; Meng et al., 2007.
126	SPRED1, TOM1, VCAM1, IRS1, DNMT1, KRAS, VEGFA, IGFBP2, MMP7, CXCL12.	Senescence, SASP, cytokine signaling, immune response, insulin signaling, angiogenesis, epigenetic regulation, tumor biology	Fish et al., 2008; Oglesby et al., 2010; Zhang et al., 2008; Harris et al., 2008; Zhao et al., 2011; Jiao et al., 2012; Liu et al., 2009; Png et al., 2012; Felli et al., 2013; Zhang et al., 2013b.
146a	CXCR4, TLR-2, TLR-4, TRAF6, IRAK1, NFKB1, KIT, STAT1.	Senescence, SASP, cytokine signaling, immune response, tumor biology	Labbaye et al., 2008; Jurkin et al., 2010; Taganov et al., 2006; Bhaumik et al., 2008; Yang et al., 2011; Wang et al., 2013.
155	FOXO3a, TP53INP1, IKBKE, SMAD5, SOCS1, TRF1.	Telomere function, senescence, cytokine signaling, immune response, tumor biology	Kong et al., 2010; Gironella et al., 2007; Gibcus et al., 2009; Yin et al., 2008; Jiang et al., 2010; Dinami et al., 2014.

### Hightlights

Exosomes are involved in immune system recognition

Exosomal alteration was observed during aging and age-related diseases

Exosomes can play opposing roles in viral disease pathogenesis

MicroRNAs related to Inflammation, Immunity and Aging are exosomes-associated

Exosomes can contribute to inflammaging

