

LJMU Research Online

Babafemi, EO, David, OM, Oluduro, AO and Famurewa, O

Epidemiology of methicillin-resistant Staphylococcus aureus among hospitalized patients and apparently healthy individuals in Ekiti and Ondo States, Nigeria.

http://researchonline.ljmu.ac.uk/id/eprint/11198/

Article

Citation (please note it is advisable to refer to the publisher's version if you intend to cite from this work)

Babafemi, EO, David, OM, Oluduro, AO and Famurewa, O (2014) Epidemiology of methicillin-resistant Staphylococcus aureus among hospitalized patients and apparently healthy individuals in Ekiti and Ondo States. Nigeria. Journal of Pharmaceutical and Biomedical Sciences. 4 (11).

LJMU has developed LJMU Research Online for users to access the research output of the University more effectively. Copyright © and Moral Rights for the papers on this site are retained by the individual authors and/or other copyright owners. Users may download and/or print one copy of any article(s) in LJMU Research Online to facilitate their private study or for non-commercial research. You may not engage in further distribution of the material or use it for any profit-making activities or any commercial gain.

The version presented here may differ from the published version or from the version of the record. Please see the repository URL above for details on accessing the published version and note that access may require a subscription.

For more information please contact researchonline@ljmu.ac.uk



JOURNAL OF PHARMACEUTICAL AND BIOMEDICAL SCIENCES

Babafemi EO, David OM, Oluduro AO, Famurewa O. **Epidemiology of methicillin-resistant** *Staphylococcus aureus* among hospitalized patients and apparently healthy individuals in Ekiti and Ondo States, Nigeria. *J Pharm Biomed Sci* 2014;04(11):1125-1130.

The online version of this article, along with updated information and services, is located on the World Wide Web at: www.jpbms.info

Journal of Pharmaceutical and Biomedical Sciences (J Pharm Biomed Sci.), Member journal. Committee of Publication ethics (COPE) and Journal donation project (JDP).

Research article

Epidemiology of methicillin-resistant Staphylococcus aureus among hospitalized patients and apparently healthy individuals in Ekiti and Ondo States, Nigeria

Babafemi, E. O.¹, David, O. M.¹, Oluduro, A. O.² and Famurewa, O.^{1,*}

Affiliation:-

¹Department of Microbiology University of Ado-Ekiti, Nigeria.P.M.B.5363, Ado-Ekiti, Nigeria.

The name of the department(s) and institution(s) to which the work should be attributed:

Department of Microbiology, Ekiti State University, Ado-Ekiti, Nigeria, P.M.B. 5363, Ado-Ekiti, Nigeria Department of Microbiology, Obafemi Awolowo University, Ile-Ife, Nigeria

Address reprint requests to Famurewa, O.

Department of Microbiology Ekiti State University, Ado-Ekiti, Nigeria P.M.B. 5363, Ado-Ekiti, Nigeria or at ofamurewa@gmail.com

Article citation: Babafemi EO, David OM, Oluduro AO, Famurewa O. Epidemiology of methicillin-resistant Staphylococcus aureus among hospitalized patients and apparently healthy individuals in Ekiti and Ondo States, Nigeria. J Pharm Biomed Sci. 2014; 04(11):1125-1130. Available at www.jpbms.info

ABSTRACT

The incidence of methicillin-resistant *Staphylococcus* aureus (MRSA) in hospitalized patients and apparently healthy individuals was investigated in two western states of Nigeria using standard microbiological methods. One thousand and two

hundred non-repeat isolates of S. aureus were recovered from the subjects. At varying degrees the isolates were resistant to cotrimoxazole (54.8%), augmentin (36.9%), pefloxacin (35.9%), gentamycin (28.3%), erythromycin (24.9%), vancomycin (10.3%), ofloxacin (5.2%) and ciprofloxacin (0.3%). One hundred and fifty six (13.0%) were resistant to methicillin out of which 4.8% and 8.2% were from healthy individuals and patients respectively. There was no correlation between prevalence of MRSA and age or sex (p < 0.05). There was no correlation between the antibiotic resistance pattern in MRSA from healthy volunteers and patients (P < 0.05). A total of 9.0%, 12.2% and 21.2% of the MRSA were resistant to 3, 4 and 5 antibiotics respectively. Antibacterial activities of five biocides examined using agar diffusion method showed that 38.5%, 53.2%, 59.6%, 61.5% and 71.8% of the MRSA were not inhibited by Izal®, Morigad®, Septol®, Dettol® and Purit® respectively, at concentrations two times higher than the in-use concentration. This finding points to the fact that MRSA occurs among patients and in the communities in the study areas, which calls for a public health concern and awareness.

KEYWORDS: Hospital-acquired MRSA; community acquired MRSA; biocides, epidemiology; multiple antibiotic resistance.

INTRODUCTION

taphylococcus aureus is an opportunistic pathogen causing diseases and infections in both man and animal. It causes wide varieties of acute suppurative infections which include superficial, subcutaneous, sub-mucous infections^{47,13,38,2,32,50} characterized by

inflammation, necroses, and abscess formation^{5,7,31}.

Staphylococcus aureus is an extremely versatile and frequent pathogen of humans both in the community and in the hospitals 10,35,7,37. Despite standardized hygienic measures, the high rate of tolerance, adaptation to new environments and

²Department of Microbiology, ObafemiAwolowo University, Ile-Ife, Nigeria

increasing resistance to antibiotics are the major factors responsible for the recent upsurge in the rate of staphylococcal infections^{27,10,50}.

The acquisition of mecA gene by S. aureus has been confirmed to be responsible for its methicillin resistance²⁹. Methicillin-resistant *S. aureus* (MRSA) was first isolated in England and was initially considered a nosocomial pathogen, however in recent years, increasing numbers of MRSA strains have been isolated worldwide from patients with infections^{45,42,43,21,6}. community-acquired Staphylococcus aureus represents a major public health threat, as MRSA strains are the leading cause of nosocomially acquired infections. MRSA infections present a challenge to infection control and treatment strategies, resulting in increased morbidity, mortality, and length of hospitalization and health care costs44,28,6,38.

Kesah *et al.*²⁶ reported 21–30% MRSA in 8 African countries including Nigeria. Shittu et al.⁴⁸ also reported emergence of a variant of epidemic strain MRSA in South Western Nigeria. However, this study was aimed at isolating and determining of methicillin resistance among *S. aureus* strains from Ekiti and Ondo States, Nigeria.

MATERIALS AND METHODS

COLLECTION AND PROCESSING OF SAMPLES

One thousand and two hundred non-repeat samples of *Staphylococcus aureus* from various clinical specimens and apparently healthy volunteers were collected in Ekiti and Ondo States of Nigeria. Informed consent was obtained from all patients before the samples were collected and all experiments were performed in accordance with the international ethical standards. Samples were inoculated on Mannitol Salt Agar (MSA) and incubated at 35°C for 24 h. Colonies with brilliant yellow appearance were sub-cultured to get pure isolates. Pure isolates were identified by standard

methods described by Olutiola *et al.*³⁹ and Fawole and Oso¹⁸ while the results were interpreted according to Holts and collegues²³.

ANTIBIOTIC SENSITIVITY TESTING

The isolates were grown at 37 °C in Mueller-Hilton broth (Oxoid) for 18 h and diluted to an optical density of 0.1 (0.5 McFarland Standard) and stored at 4 °C. The disc diffusion method was used for susceptibility testing as described by Clinical and Laboratory Standard Institute, CLSI¹². The isolates were tested against the following commercial antibiotic with their concentrations amoxicillin (25), augmentin (30), ciprofloxacin (10), cotrimoxazole (25), erythromycin (5), gentamicin (10), ofloxacin (30), oxacillin (5), perfloxacin (5), and vancomycin (5). Oxford Staphylococcus aureus (NCTC 6571) was used as control strain. Multiple antibiotic resistance was determined for strains that are not susceptible to oxacillin, which is taken for methicillin resistance according to Onemu and Ophori⁴⁰. Agar dilution method was used to determine susceptibility of isolates to commonly used biocides.

STATISTICAL ANALYSIS OF DATA

Statistical analysis was done using SPSS (version 17) to determine frequency distribution, mean, harmonic mean, standard deviation, analysis of variance (ANOVA), Duncan Multiple Range and Pearson correlation coefficient.

RESULTS

The distribution of the S. aureus strains recovered from subjects is represented in Table 1. A total of 498 (41.5%) and 702 (56.5%) was recovered from both male and female subjects respectively. The antibiotic susceptibility test of 1,200 S. aureus is shown in Table 2. One hundred and fifty-six (13.0%) were resistant to methicillin antibiotic.

Table 1. The distribution of *S. aureus* recovered from the study areas.

State	Healthy (%)		Patients (%)		Total	Total	
	Male	Female	Male	Female	Male	Female	
Ekiti	154 (12.8)	284 (23.7)	114 (9.5)	48 (4.0)	268 (22.3)	332 (27.7)	
Ondo	191 (15.9)	308 (25.7)	39 (3.3)	62 (5.2)	230 (19.2)	370 (30.8)	
Total	345 (28.8)	592 (49.3)	153 (12.8)	110 (9.2)	498 (41.5)	702 (58.5)	

Antibiotic susceptibility test showed that most of the MRSA isolates were resistant to all the antibiotics tested in varying degrees. Resistance to cotrimoxazole was highest (87.2%) whereas it was least against ciprofloxacin (1.3%). This was followed by ofloxacin (37.2% resistance) and pefloxacin (46.8% resistance). One hundred and

thirty six (87.2%) strains being highest were resistant to cotrimoxazole, 130 (83.3%) to amoxicillin and 118 (75.6%) to vancomycin. There was no statistically significant difference between the antibiotic resistance pattern in MRSA from apparently healthy volunteers and patients (p<0.05).

Table 2. Distribution of Staphylococcus aureus according to antibiotics susceptibility testing.

Antibiotic	Ekiti (n = 600)				Ondo (n = 600)			
	Healthy (%)		Patients (%)		Healthy (%)		Patients (%)	
	R	S	R	S	R	S	R	S
Aug	159(13.3)	279 (23.3)	91 (7.6)	71 (5.9)	113 (9.4)	386 (32.2)	80 (6.7)	21 (1.8)
Amox	78 (6.5)	360 (30.0)	57 (4.8)	105 (8.8)	50 (4.2)	449 (37.4)	44 (3.7)	57 (4.8)
Cip	1 (0.08)	437 (36.4)	1 (0.08)	161 (13.4)	1 (0.08)	498 (41.5)	0 (0)	101 (8.4)
Cotr	211 (17.6)	227 (18.9)	109 (9.1)	53 (4.4)	251 (20.9)	248 (20.7)	87 (7.3)	14 (1.2)
Ery	91 (7.6)	347 (28.9)	53 (4.4)	109 (9.1)	84 (7.0)	415 (34.6)	71 (5.9)	30 (2.5)
Gen	125 (10.4)	313 (26.1)	97 (8.1)	65 (5.4)	83 (6.9)	416 (34.7)	35 (2.9)	66 (5.5)
Oflo	30 (2.5)	408 (3.4)	24 (2.0)	138 (11.5)	7 (0.6)	492 (41.0)	1 (0.08)	100 (8.3)
Oxa	9 (0.8)	429 (35.8)	51 (4.3)	111 (9.3)	49 (4.1)	450 (37.5)	47 (3.9)	54 (4.5)
Pef	202 (16.8)	236 (19.7)	67 (5.6)	95 (7.9)	118 (9.8)	381 (31.8)	44 (3.7)	57 (4.8)
Van	8 (0.7)	430 (35.8)	33 (2.81)	129 (10.8)	42 (3.5)	457 (38.1)	41 (3.4)	60 (5.0)

AMX=amoxicillin, AUG=augmentin, CIP=ciprofloxacin, COT=cotrimoxazole, ERY=erythromycin, GEN=gentamicin, OFL=ofloxacin, OXA=oxacillin, PEF=perfloxacin, VAN=vancomycin, S=Susceptible, R=resistance

Multiple antibiotic resistance (MAR) of MRSA strains was investigated. All the organisms showed multiple resistance in varying degrees ranging. Forty two (26.9%), 28 (18.0%) and 6 (3.9%) were

resistant to 6, 7 and 8 antibiotics among the MRSA strains respectively. None was resistant to all the 9 antibiotics tested (Table 3).

Table 3. Multiple Antibiotic Resistance (MAR) Strain among MRSA strains.

Number of Antibiotic	MRSA strain	Percentage
3	14	9.0
4	19	12.2
5	33	21.2
6	42	26.9
7	28	18.0
8	6	3.9

Most of the MRSA isolates tested against the various concentrations/dilutions of the commonly used disinfectants were resistant. Overall, 156 (100.0%) strains of MRSA from both Ekiti and Ondo States were resistant to all the disinfectants at the concentrations lower than the in-use concentrations recommended by manufacturers. At the recommended concentrations, 109 (69.9%) strains of MRSA from Ekiti and Ondo states were resistant to Izal® whereas 145 (92.9%), 145 (92.9%), 144 (92.3%) and 140 (89.7%) were resistant to Septol®, Dettol®, Purit® and Morigad® respectively.

At concentrations above the recommended in-use concentrations, the antimicrobial activity of the disinfectants revealed that 91 (58.3%) MRSA strains from or from Ekiti and Ondo were resistant to Izal®while 109 (69.9%), 126 (80.0%) and 130 (83.3%) resistant to Morigad®, Dettol® and Purit® respectively. Septol® was the least effective among

all the disinfectants with only 16.0~% of the isolates susceptible to it.

DISCUSSION

This study summarizes the results of a two-year surveillance in the two selected states of Nigeria. Most of the health care facilities surveyed are tertiary care specialist hospitals, tertiary institutions in Ekiti and Ondo States, Nigeria. A total of 1,200 isolates of S. aureus from various non-repeat clinical samples was recovered in the participating hospitals and communities out of which 156 (13.0%) were identified as MRSA. This was below that of Onemu and Ophori⁴⁰ that reported 79% resistance to methicillin among the clinical S. aureus isolates. This incidence of MRSA among the patients observed in this study was lower than the range between 24. % and 34.1 %recorded by Akujobi *et al.*⁵ among different cadres of healthcare workers in Nigeria. Onemu and Ophori⁴⁰ reported 79.0% while 34.1% recorded by

Akujobi et al.5 in Edo and Anambra States both in Nigeria. Fifty eight (37.2%), 23 (40.0%) and 98 (62.8%) MRSA strains were isolated from apparently healthy volunteers, apparently healthy hospital personnel/health care workers (HCWs) and patients respectively. The high incidence of MRSA among patients could be due to reasons ranging from being acquired from hospital workers who introduced the organism into hospital environment (as nosocomial infection) from the community, abuse/misuse of antibiotics and low immunity in patients as a result of disease conditions^{17,7,25,40}. This agrees with earlier reports by Abraham and collegues1. There was no statistically significant difference between the prevalence of MRSA and age or sex (p < 0.05).

The antibiotic sensitivity pattern of the isolates shows that ciprofloxacin was the most effective followed by ofloxacin, pefloxacin, gentamycin, erythromycin, augmentin, vancomycin, amoxycillin and cotrimoxazole in that order (Table 2). Statistically, there was no significant difference between the resistance to antibiotics among MRSA from healthy volunteers and patients (p < 10.05). The result of this study reveals the efficacy of members of floroquinolones over the other antibiotics. The study shows that the likes of cotrimoxazole. amoxycillin, vancomycin, augmentin and erythromycin, to a high degree, were an unreliable empirical choice of antibiotics for the treatment of infections with MRSA strains as the aetiological agents 17,46,49 .

Considering the general overview of the antibiotic susceptibility profile of the strains isolated in this study, the susceptibility of the MRSA strains to ciprofloxacin (98.7 %) may be due in part to it non-availability and exorbitant cost unlike the commonly used antibiotics which are cheaper and could easily be afforded. The newer and expensive antibiotics have undoubtedly high antibacterial efficacy against MRSA in this study.

The antibacterial activity of five commonly used biocides on the 156 MRSA strains showed the efficacy of the biocides used Izal®, Morigad®, Septol®, Purit® and Dettol® in that order on MRSA strains isolated. The finding in this study agrees with reports that resistance to benzalkonium chloride, a biocide, has been closely linked to oxacillin resistance in *S. aureus*^{33,24,16}.

It was reported that resistance to both biocides and antibiotics can be plasmid-mediated. Plasmidmediated resistance to biocides is a wellrecognized phenomenon. Such resistance to quaternary ammonium chlorides and other biocides has been identified in nosocomial pathogens^{51,14}. Resistance to antibiotics and biocides may be due to the common target site between the two antimicrobial agents which may bring about selection of mutants altered in such target by either agents or the emergence of crossresistance^{9,20,30}. It has been suggested that subtle differences in the biocide susceptibility of antibiotic-resistant strains might facilitate their selection and maintenance in the environment by low-sub-effective concentrations of the agents^{34,35,41,15} owing to indiscriminate and on inappropriate use11,19,22,16,24,41.

This study shows that there is a relatively high incidence of MRSA (13%) that are resistant to antibiotics and biocides in the study locations. A continuous surveillance is therefore recommended because of the public health importance of the organism and the high danger this may potent.

REFERENCES

- Abraham B, Jacob G, Pablo Y, Nechawa P, Nurith P, Ronif T, Hannah S, Klaris R, Miriam S, Francis S. Community acquired MRSA in institutionalized adults with development disabilities. *Emerg Infect Dis.* 2002; 8:966-969.
- 2. Adeleke SI, Asani MO. Urinary tract infection with children with nephritic syndrome in Kano, Nigeria. *Ann Afr Med.* 2009; 8:38-41.
- Ahmed MO, Elramalli AK, Amri SG, Abuzweda AR, Abouzeed YM. Isolation and screening of methicillin-resistant *Staphylococcus aureus* from health workers in Libyan hospitals. *East Meditrr Health J.* 2012; 18(1):37-42.
- Ako-Nai AK, Ogunniyi AD, Lamikanra A, Torimiro SE. The characterisation of clinical isolates of Staphylococcus aureus in Ile-Ife, Nigeria. J Med Microbiol. 1991; 34:109-112.
- Akujobi CN, Ilo IA, Egwuatu CC, Ezeanya CC. Prevalence of methicillin-resistant *Staphylococcus aureus* (MRSA) among healthcare workers in a tertiary institution in Nigeria. *Orient J Med.* 2013; 25(3-4):82-87.
- Anyanwu NCJ, Abdullahi IO, Ameh JB, Ella EE. Molecular detection of *PVL*, *msr*A genes and antibiotic susceptibility pattern of staphylococcus aureus from skin and soft tissue infections in Zaria, Nigeria. Scient J Microbiol. 2013: 2(2):43-52.
- Ateba NU, Schaumburg F, Adegnika AA, Kosters K, Möller T, Fernandes JF, Alabi A, Issifou S, Becker K, Grobusch MP, Kremsner PG, Lell B. Epidemiology and population structure of *Staphylococcus aureus* in various population groups from a rural and semi urban area in Gabon, Central Africa. *Acta Trop*. 2012; 124:42-47.
- 8. Boyce JM. MRSA in hospitals and long term care facilities: microbiology, epidemiology and preventive measures. *Infect Contr Hos Epidemiol*. 2005; 13:725-734.

ISSN NO- 2230 - 7885 CODEN JPBSCT NLM Title: J Pharm Biomed Sci.

- Castillo JA, Clapes P, Infante MR, Comas J, Manresa A. Comparative study of the antimicrobial activity of bis-(N{alpha}-caproyl-L-arginine)-1,3propanediamine dihydrochloride and chlorhexidinedihydrochloride against Staphylococcus aureus and Escherichia coli. J Antimicrob Chemother. 2006; 57:691-698.
- 10. Choi SC, Chow SY, Afra A. Nasal carriage of Staphylococcus aureus among healthy adults. J Microbiol Immunal Infect. 2006; 39:458-464.
- 11. Chopra I. Bacterial resistance to disinfectants, antiseptics and toxic metal ions. *Soc Appl Bacteriol Tech Ser.* 1991; 27:45-64.
- 12. CLSI. Performance Standards for Antimicrobial Disk Susceptibility Tests; Approved Standard—Eleventh Edition. M02-A11 32(1) Replaces M02-A10. 2012; 29(1).
- 13. Craig EZ, Bryon C, Mark AM, James EL, Adam A, Bruce KB. Community-acquired methicillin-resistant *Staphylococcus aureus* among Military Recruits. *Emerg Infect Dis.* 2004; 10:941-943.
- David OM, Fakayode IB, Famurewa O. Evaluation of the anti-enterococcal activity of disinfectants and medicated soaps on vancomycin-resistant Enterococcus faecalis strains. Ann Res Rev Biol. 2014; 4(3):509-519.
- 15. David OM, Ayeni D, Fakayode IB, Famurewa O. Evaluation of antibacterial properties of various hand sanitizers wipes used for cosmetic and hand hygiene purposes in Nigeria. *Microbiol Res Inter.* 2013; 1(2): 22-26.
- El-Mahmood AM, Doughari JH. Bacteriological examination of some diluted disinfectants routinely used in the Specialist Hospital Yola, Nigeria. *Afr. J. Pharm Pharmacol.* 2009; 3:185-1909.
- Esan CO. Molecular studies of resistant Staphylococcus aureus in Ekiti and Ondo States, Nigeria. Ph.D Thesis. University of Ado-Ekiti, Nigeria. 2011.
- Fawole MO, Oso BA. Laboratory Manual of Microbiology. Spectrum Books Limited, Ibadan. 2001: 127.
- Fayyaz M, Mirza IA, Ahmed Z, Abbasi SA, Hussain A and Ali S. (). *In vitro* susceptibility of chloramphenicol against methicillin-resistant *Staphylococcus aureus*. *J Coll Physic Surg Pak*. 2013; 23(9): 637-640.
- 20. Fraise AP. Biocide abuse and antimicrobial resistance A cause for concern? *J Antimicrob Chemother.* 2002; 49:1-12
- 21. Ghebremedhin B, Olugbosi MO, Raji AM, Layer F, Bakare RA, Konig B, Konig W. Emergence of a community-associated methicillin-resistant *Staphylococcus aureus* with unique resistance profile in Southwest of Nigeria. *J Clin Microbiol.* 2009; 47: 2975-2980.
- 22. Gilbert P, McBain AJ. Potential impact of increased use of biocides in consumer products on prevalence of antibiotic resistance. *Clin Microbiol Rev.* 2003; 16:189-208.
- 23. Holt JG, NR Krieg, PHA Sneath, JT Staley, and ST Williams. Bergey's Manual of Determinative

- Bacteriology, 9th Edn. Williams & Wilkins, Baltimore. 1994.
- 24. Karpanen TJ, Worthington T, Hendry ER, Conway BR, Lambert PA. Antimicrobial efficacy of chlorhexidine digluconate alone and in combination with eucalyptus oil, tea tree oil and thymol against planktonic and biofilm cultures of *Staphylococcus epidermidis. J Antimicrob Chemother.* 2008; 325:1-6.
- 25. Kejela T, Bacha K. Prevalence and antibiotic susceptibility pattern of methicillin-resistant *Staphylococcus aureus* (MRSA) among primary school children and prisoners in Jimma Town, Southwest Ethiopia. *Ann Clin Microbiol Antimicrob*. 2013: 24: 12:11
- Kesah C, Redjeb S.B., Odugbemi T.O., Boye C.S-B., Dosso M., NdinyaAchola J.O., Koulla-Shiro S, Benbachir M., Rahal K., Borg M (2003). Prevalence of methicillin-resistant Staphylococcus aureus in eight African hospitals and Malta. *Clin.Microbiol. Infect*. 9:153-156.
- 27. Kluytmans J, van Belkum A, Verbrugh H (1997). Nasal carriage of Staphylococcus aureus: epidemiology, underlying mechanisms, and associated risks. *Clin Microbiol Rev.* 10 (3): 505–20.
- 28. Kolman1, S, Arielly H, Paitan Y. Evaluation of single and double-locus real-time PCR assays for methicillin-resistant *Staphylococcus aureus* (MRSA) surveillance. *BMC Res Notes*. 2010; 3:110
- 29. Kumurya AS. Loss of the *mecA* gene during storage of methicillin-resistant *Staphylococcus aureus* isolates in Northwestern Nigeria. *J Public Health Epidemiol.* 2013; 5(10):410-415.
- 30. Little-John TG, Paulsen IT, Gillespie MT, Tennent JM, Midgley M, Jones IG. Substrate specificity and energetic of antiseptic and disinfectant resistance in *Staphylococcus aureus*. *FEMS Letters*. 1992; 95:259-266.
- 31. Monecke S, Ruppelt A, Wendlandt S, Schwarz S, Slickers P, Ehricht R, Jäckel SC. Genotyping of *Staphylococcus aureus* isolates from diseased poultry. *Vet Microbiol.* 2013; 162: 806-812.
- 32. Nadjia B, Mebrouk K. Phenotypic and genotypic characterization of *Staphylococcus aureus* agents of dairy cows' mastitis in Algeria. *J Appl Sci Res.* 2013; 9(1):86-93.
- 33. Noguchi N, Nakaminami H, Nishijima S, Kurokawa I, So H, Sasatsu M. Antimicrobial agent of susceptibilities and antiseptic resistance gene distribution among methicillin-resistant *Staphylococcus aureus* isolates from patients with impetigo and staphylococcal scalded skin syndrome. *J Clin Microbiol.* 44: 2119-2125.
- 34. Noguchi N, Suwa J, Narui K, Sasatsu M, Ito T, Hiramatsu K, Song JH. Susceptibilities to antiseptic agents and distribution of antiseptic-resistance genes *qacA/B* and *smr* of methicillin-resistant *Staphylococcus aureus* isolated in Asia during 1998 and 1999. *J Med Microbiol.* 2005; 54:557-565.
- 35. Nwakwo BOK, Abdulhadi S, Magagi A, Ihesiulor G. Methicillin resistant *Staphylococcus aureus* and their antibiotic susceptibility pattern in Kano,

ISSN NO- 2230 - 7885 CODEN JPBSCT NLM Title: J Pharm Biomed Sci.

- Nigeria. *Afr J Clin Experim.Microbiol.* 2010; 11(1):1595-689.
- 36. Ebrahimi A, Ghasemi M, Ghasemi B. Some Virulence Factors of Staphylococci Isolated From Wound and Skin Infections in Shahrekord, IR Iran. *Jundishapur J Microbiol.* 2014; 7(4):1-5.
- 37. Obajuluwa AF, Onaolapo JA, Oyi AR, Olayinka BO. Susceptibility profile of methicillin-resistant *Staphylococcus aureus* (MRSA) Isolates to antibiotics and methanolic extracts of parkia biglobosa (Jacq) Benth. *Brit J Pharma Res.* 2013; 3(4):587-596.
- 38. Odetoyin WB, Aboderin AO, Ikem RT, Kolawole BA, Oyelese AO. Asymptomatic bacteriuria in patients with diabetes mellitus in Ile-Ife, South-West, Nigeria. *East Afr Med J.* 2008; 85:18-23.
- 39. Olutiola PO, Famurewa O, Sonntag H-G. An Introduction to General Microbiology. Hygiene-Institut Der Universitat Heidelberg Federal Republic of Germany. 2001; 267.
- Onemu OŚ, Ophori EA. Prevalence of multi-drug resistant Staphylococcus aureus in clinical specimens obtained from patients attending the University of Benin Teaching Hospital, Benin City, Nigeria. J Nat Sci Res. 2013; 3(5):154-159.
- 41. Onochie CC, Chukwudi A, Alo MN, Onwa NC, Okonkwo EC and Afiukwa FN. Bacteriological examination of computer keyboards and mouse devices and their susceptibility patterns to disinfectants. *Amer J Microbiol.* 2013; 4(1):9-19.
- 42. Pallin DJ, Egan DJ, Pelletier AJ, Espinola JA, Hooper DC, Camargo CA. Increased US Emergency Department visits for skin and soft tissue infections, and changes in antibiotic choices, during the emergence of community-associated methicillinresistant Staphylococcus aureus. Ann Emerg Med. 2008; 51:291-298.
- 43. Popovich KJ, Weinstein RA, Hota B. Are community-associated methicillin- resistant *Staphylococcus aureus* (MRSA) strains replacing **Source of support:** None

- traditional nosocomial MRSA strains? *Clin Infect Dis.* 2008: 46:787-794.
- 44. Price CS, Williams A, Philips G, Dayton M, Smith W, Morgan S. *Staphylococcus aureus* nasal colonization in pre-operative orthopaedic out patients. *Clin Orthop Relat Res.* 2008; 466:2824-2847.
- 45. Ramdani-Bouguessa N, Bes M, Meugnier H, Forey F, Reverdy ME, Lina G, Vandenesch F, Tazir M, Etienne J. Detection of methicillin-resistant *Staphylococcus aureus* strains resistant to multiple antibiotics and carrying the Panton-Valentine leukocidin genes in an Algiers hospital. *Antimicrob Agents Chemother*. 2006; 50:1083-1085.
- 46. Sheikh AF, Mehdinejad M. Identification and determination of coagulase-negative Staphylococcus species and antimicrobial susceptibility pattern of isolates from clinical specimens. Afri J Microbiol Res. 2012; 6(8):1669-1674.
- 47. Shittu AO, Lin J, Kolawole DO. Antimicrobial susceptibility patterns of *Staphylococcus aureus* and characterization of MRSA in Southwestern Nigeria. *Wounds*. 2006; 18:77-84
- 48. Shittu AO, Lin J. Antimicrobial susceptibility pattern and characterization of clinical isolates of *Staphylococcus aureus* in Kwazulu-Natal province of South Africa. *BMC Infect Dis* 2006; 6:188-192.
- 49. Smith TL, Pearson ML, Wilcox KR. Emergence of Vancomycin resistance in *Staphylococcus aureus*. *NEJM*. 1999; 340:493-501.
- 50. Tula MY, Azih AV, Okojie RO. Antimicrobial susceptibility pattern and plasmid-mediated antibacterial resistance in *Staphylococcus aureus* and coagulase-negative staphylococci (CoNS). *Amer J Res Com.* 2013; 1(9):149-166.
- 51. Weber DJ, Rutala WA, Sickbert-Bennett EE. Outbreaks associated with contaminated antiseptics and disinfectants. *Antimicrob Agents Chemother*. 2007; 51:4217-4224.

Competing interest / Conflict of interest

The author(s) have no competing interests for financial support, publication of this research, patents and royalties through this collaborative research. All authors were equally involved in discussed research work. There is no financial conflict with the subject matter discussed in the manuscript.

Copyright © This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.