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Role of platelets as mediators that link inflammation and thrombosis in atherosclerosis

Platelets and the Immune Response

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ABSTRACT

Platelets, crucial mediators of the acute complications of atherosclerosis that cause life-threatening ischemic events at late stages of the disease, are also key effectors of inflammation throughout plaque development through their interaction with endothelial and immune cells in the injured vessel wall. During the first steps of atherosclerosis, blood inflammatory leukocytes interact with the damaged endothelium in areas rich in platelet aggregates. In late stages of the disease, platelets secrete several inflammatory molecules, even without forming aggregates. These molecules exacerbate the inflammation and induce the transition from chronic to acute disease, featuring increased instability of the atherosclerotic lesion that results in plaque rupture and thrombosis. Moreover, platelets play an important role in vascular wall remodeling induced by chronic inflammation by controlling vascular cell differentiation and proliferation. In this review we discuss the role of platelets as cell mediators that link inflammation and thrombosis in atherosclerotic disease and their potential in the development of new therapeutic tools to fight cardiovascular disease.

INTRODUCTION

The incidence and prevalence of cardiovascular disease has increased significantly in recent years, due at least in part to the progressive aging of the population. According to the World Health Organization (WHO), by 2020 cardiovascular disease will become the leading cause of disability and death in the world [1, 2]. In most cases, myocardial infarction and stroke are the consequence of atherosclerotic plaque rupture and thrombus formation. Atherothrombosis is regulated by both genetic and environmental factors (e.g., dyslipidemia, hypertension, smoking, diabetes and obesity) [3, 4]. The development of atherosclerotic lesions is the result both of lipid accumulation in the sub-endothelial space in arteries and of a complex process of chronic inflammation characterized by endothelial dysfunction, leukocyte infiltration and activation of Platelets induce differentiation of human CD34+ progenitor cells into foam cells and endothelial cells vascular smooth muscle cells. Importantly, activated smooth muscle cells within the injured vessel wall undergo a de-differentiation process, acquiring a highly proliferative and migratory phenotype [5-7].

Circulating platelets in peripheral blood play a major role in maintaining blood homeostasis. Reports in the last decade have described the secretion by platelets of pro-inflammatory molecules that exacerbate the inflammatory response in the atherosclerotic plaque [8-10]. This event occurs during the initial injury to the endothelium as well as during later stages when the atherosclerotic plaque is destabilized [11, 12]. Platelets also interact directly with other cells of the immune system in physiological and pathological conditions [10]. In the context of atherosclerosis, platelets can adhere to endothelial cells and contribute to the recruitment of leukocytes involved in the local vascular inflammation [13, 14]. In this review we discuss the involvement of platelets in the initiation and development of the inflammatory process of atherosclerosis. We also describe the most relevant platelet molecules implicated in the interaction with immune cells and their activation toward the inflammatory phenotype.

INFLAMMATION, PLATELETS AND ATHEROSCLEROSIS

It is today well-established that numerous cellular and molecular inflammatory components participate during all stages of atherosclerosis, from early lesion development to vulnerable plaque formation [15, 16]. Atherosclerotic lesions typically have a prominent infiltrate of immune cells, composed mainly of monocytes/macrophages, T cells, dendritic cells and neutrophils. These cells are recruited from the bloodstream at sites of endothelial injury and locally produce a plethora of regulatory molecules that contribute to the onset and progression of atherosclerosis [17-19]. At late stages of the disease, monocytes/macrophages accumulate in the atheromatous lesion and secrete large quantities of extracellular proteases, including serine proteases, cathepsin and metalloproteinases, that increase plaque instability [20]. Moreover, apoptosis of neointimal macrophages provokes the release of collagen degrading proteases and tissue factor (TF), further promoting plaque vulnerability [21-23]. Immune cells also promote plaque instability by producing cathepsins and metalloproteinases that degrade interstitial collagen [24, 25]. Stable and vulnerable plaques differ in the composition of their inflammatory infiltrates [26], and current studies therefore focus on characterizing the immune cells localized at the plaque in order to identify markers of vulnerable plaques [27]. These markers would allow the identification of high risk patients before they start to develop an acute ischemic event [28, 29].

As indicated before, platelets play a major role in vascular inflammation through the production and release of pro-inflammatory mediators and by interacting with endothelial cells, leukocytes and vascular smooth muscle cells. Using the atherosclerosis-prone apolipoprotein E-deficient mouse model (ApoE^{-/-}), it has been shown that activated platelets and platelet-leukocyte/monocyte aggregates promote formation of atherosclerotic lesions, suggesting their atherogenic potential [30]. This may be due in part to increased cell proliferation, a key process in the early stages of atherosclerosis both in animal models and in humans [31]. Indeed, platelet-derived growth factor (PDGF) down regulates the expression of the growth suppressor p27,

whose genetic ablation in ApoE^{-/-} mice promotes excessive proliferation of vascular smooth muscle cells and neointimal macrophages and exacerbates atherosclerosis [32-35]. Another pro-atherogenic factor released by platelets is the chemokine stromal-cell derived factor-1 (SDF-1), which recruits hematopoietic precursors to incipient lesions where they proliferate and differentiate into new inflammatory cells [36]. In addition, after platelets get activated and form aggregates, they increase the secretion of other potentially pro-atherogenic molecules, such as growth-regulated oncogene-a (GRO-a), platelet factor-4 (PF-4), epithelial-neutrophil activating peptide (ENA-78), interleukin-8 (IL-8), monocyte chemo attractant protein-1 (MCP-1), macrophage inflammatory protein 1 α (MIP-1 α) and RANTES [37]. It is interesting to note that thrombin-mediated platelet activation is not required for the early development of atherosclerosis in ApoE^{-/-} mice, suggesting that, if platelet activation is required for plaque formation in this animal model, platelet activators other than thrombin suffice [38]. For instance, P-selectin expression is critical for monocyte recruitment to sites of neointima formation after arterial injury in ApoE^{-/-} mice and its complete absence attenuates plaque area in this experimental model (ref 39). Moreover, platelet P-selectin expression is associated with atherosclerotic wall thickness in carotid artery in humans (ref 40).

Platelets also intensify the inflammatory process at all stages of atherosclerosis by expressing membrane molecules such as intercellular adhesion molecule-2 (ICAM-2), P-selectin, CD95L and CD40L. These molecules regulate several biological functions in the vessel wall, including cellular adhesion and aggregation, chemotaxis, survival and differentiation, and angiogenesis [41, 42].

It has also been shown that immunoglobulin complexes increase platelet expression of inflammatory molecules, such as soluble CD40L (sCD40L) and RANTES, in the absence of aggregation [43]. Moreover, platelet activation also promotes a conformational change in C-reactive protein (CRP), shifting it from the pentameric to the monomeric form, which is highly

abundant in inflammatory settings and which may play a role in the pathogenesis of atherosclerosis [44]. Recently, Kirbis *et al* have shown that increased levels of monomeric CRP can predict the risk of acute coronary syndrome independently of conventional cardiovascular risk factors [45].

PLATELET COMPONENTS THAT REGULATE THE INFLAMMATORY PROCESS

Platelets have been identified as regulators of the inflammatory processes that control both the initiation of the atherosclerotic lesion and plaque instability at late stages of disease progression (Figure 1) [46, 47]. This regulation is mediated by factors that are produced by activated platelets, such as CD40L, P-selectin, RANTES and toll-like receptors (TLRs).

CD40L. Immune and endothelial cells express CD40, a member of the superfamily of tumor necrosis factor (TNF) receptors. The ligand CD40L is similarly expressed in the plasma membranes of endothelial cells, T-lymphocytes and platelets [48-50]. However, platelets are the main source of the soluble form, sCD40L, contributing 95% of its circulating levels [51]. High levels of sCD40L have been associated with platelet activation together with increased numbers of neutrophils *in vivo*, suggesting a prognostic value in patients with advanced atherosclerosis and metabolic syndrome, and potential use as a biomarker of thrombosis [52, 53].

The major contributors to the release of circulating sCD40L from CD40L on the platelet surface are metalloprotease-2 [54] and glycoprotein (GP) Ib/IX/V complex (GPIb/IX/V), which induces sCD40L through the production of thromboxane A₂ (TXA₂) in patients with atherosclerosis [55]. sCD40L significantly increases platelet activation and aggregation through CD40-dependent TRAF-2/Rac1/p38 MAPK signaling [56], while blockade of this pathway with anti-CD40L antibodies can prevent or delay atherosclerosis progression [57, 58].

CD40/CD40L has been shown to induce macrophage-mediated liberation of TF, favoring a pro-thrombotic stage [59]. sCD40L is also able to bind macrophage 1 antigen (Mac-1), an integrin

expressed on monocytes/macrophages [60], resulting in the secretion of TF, pro-inflammatory cytokines (e.g., IL-1 β , IL-6, IL-8, and TNF- α) and myeloperoxidases [61]. CD40L can also induce maturation of dendritic cells, with a positive regulation of co-stimulatory molecules and the production of IL-12p40 [62]. CD40L also increases T CD8+ cell responses *in vitro* and *in vivo* [63], as well as the release of IgG from B lymphocytes [64]. CD40-CD40L interaction in neutrophils is PI3K-dependent and induces the production of reactive oxygen species, establishing a positive feedback loop for platelet activation by the redox environment [65]. In endothelial cells, sCD40L induces the expression of cell adhesion molecules (ICAM-1, VCAM-1 and E-selectin) and the secretion of the pro-angiogenic chemokines MCP-1 and IL-8 [66, 67], and reduces the expression of thrombomodulin, thus promoting a pro-thrombotic state [68]. sCD40L also activates vascular smooth cells and fibroblasts [69, 70], two cell types that play an important role in the pathogenesis of atherosclerosis [71, 72].

P-selectin. P-selectin is a cell surface adhesion molecule stored in the Weibel-Palade bodies of endothelial cells and in α -granules in platelets [73]. P-selectin localizes to the cell surface of endothelial cells upon granule exocytosis and plays an essential role in the initial recruitment of leukocytes to sites of injury during inflammation by interacting with P-selectin glycoprotein ligand (PSGL1) [74, 75]. P-selectin expressed by neointimal macrophages may also contribute to the inflammatory response during atherosclerosis development [76-78]. P-selectin expression in platelets is mainly regulated by Gas6 [79] and low levels of fibrinogen [80]. Platelet-derived P-selectin seems to contribute to atherosclerotic lesion development [81] and arterial thrombogenesis by forming large stable platelet-leukocyte aggregates [82]. Notably, platelet-derived P-selectin, but not endothelial P-selectin, also plays a crucial role in the development of neointima formation after angioplasty/air desiccation injury of the mouse carotid artery [83].

sP-selectin is a soluble form of P-selectin produced *in vivo* by platelets either from alternative mRNA splicing that generates an isoform that lacks the transmembrane domain, or from

proteolytic cleavage of the membrane-bound form [84]. Evidence exists that sP-selectin is a biomarker of vascular disease, since its levels are increased in patients with acute coronary syndrome [85] or hypertension [86]. Moreover, measurement of circulating sP-selectin has been suggested for remote testing of platelet function in patients treated with clopidogrel and aspirin [87].

Mice engineered to produce abnormally high plasma levels of sP-selectin exhibit enlarged infarcts in an ischemic stroke model, and increased susceptibility to atherosclerosis development in the ApoE^{-/-} genetic background [88]. In addition to its role in cell adhesion, the interaction of P-selectin with PSGL1 is essential for the secretion of von Willebrand Factor by endothelial cells [89]. P-selectin also activates leukocyte Mac-1 and later antigen-4 (VLA-4), which intensify inflammation and facilitate adhesion of circulating platelets to the vascular endothelium [90]. Moreover, the P-selectin-PSGL1 interaction induces the “inside-out” activation of integrins $\alpha_L\beta_2$ and $\alpha_M\beta_2$ in leukocytes via signaling through Src kinases and Naf1 [91]. This process promotes cell-cell and cell-extracellular matrix interactions, since $\alpha_M\beta_2$ binds several ligands (including iC3b, fibrinogen and heparin) and $\alpha_L\beta_2$ binds ICAM-1 [92]. P-selectin-induced recruitment of neutrophils to the injured endothelium is primarily mediated by the activation of integrins β_2 and β_3 (CD11b/CD18 and CD41/CD61), further promoting the inflammatory response [93, 94]. Moreover, PSGL1 activation in monocytes and neutrophils induces superoxide anion production [95].

RANTES. Activated platelets secrete the chemokine RANTES, which binds to the CCR1 chemokine receptor in leukocytes and promotes their interaction with the endothelium in the inflamed vasculature [96]. In addition, RANTES facilitates the recruitment of low density lipoproteins (LDLs) to the sub-endothelial space. LDLs undergo oxidation that contributes to the inflammatory response and to the progressive increase of plaque vulnerability [97, 98]. Nanomolar concentrations of RANTES promote chemotaxis through the interaction with G

protein-coupled receptors (GPCRs) [99]. In contrast, at micromolar concentrations RANTES promotes the formation of aggregates that contribute to cell activation (proliferation, apoptosis and cytokine secretion) through mechanisms independent of GPCRs [99]. Cytoplasmic levels of RANTES correlate with CRP and fibrinogen levels in middle aged patients with cardiovascular risk factors, suggesting the potential of RANTES as a biomarker of atherosclerosis and associated cardiovascular disease [100, 101].

The function of RANTES in atherosclerosis is related to other platelet-derived molecules. For example, interactions between RANTES and PF-4 amplify the effect of PF-4 on monocytes by heterophilic interactions [102, 103]. Also, deposition of RANTES on the endothelium is promoted by platelet P-selectin [104], promoting the expression of the pro-atherogenic molecule MCP-1 by inflammatory monocytes [105].

Toll-like receptors (TLRs). TLRs are the main regulators of the adaptive and innate immune responses [106], and play important roles in atherosclerosis and pathological myocardial remodeling [107]. TLR4 and TLR9 are prominently expressed in the cytoplasm of human platelets [108], and their expression levels are doubled in activated platelets [109]. TLRs are activated by heat-shock proteins, components of the extracellular matrix, fibrinogen and myeloid related protein 8/14, resulting in the secretion of cytokines, chemokines and molecules related to the transition to an unstable plaque [110, 111]. It has been shown that lipopolysaccharide stimulates platelet secretion and potentiates platelet aggregation via TLR4/MyD88 and the cGMP-dependent protein kinase pathway [112]. The binding of lipopolysaccharides to TLR4 also activates platelets, leading to the secretion of IL-1 β and activation of their membrane-bound integrins GPIIb and IIIa [113]. In turn, active GPIIb and IIIa in platelets stimulate the secretion of MCP-1 and the expression of ICAM-1 and $\alpha\beta$ 3 by endothelial cells, thus promoting monocyte and neutrophil recruitment [114, 115]. In addition, TLR4 activation increases platelet release of sCD40L and platelet-activating factor 4 (PAF4) [116]. However, another study suggests that stimulation of non-

platelet TLR2 and TLR4 during the advanced stages of atherosclerotic disease reduces the secretion of MIP-1 and RANTES in ApoE^{-/-} mice [117]. Stimulation of TLR2 with Pam3CSK4, a synthetic agonist of TLR2/TLR1, activates GPIIb/IIIa and augments the expression of P-selectin on the platelet surface [118]. In addition, TLR2 activation promotes Ca²⁺ mobilization, TXA₂ production and platelet aggregation mediated by the activation of P2X1 and ADP receptors [119]. The TLR2 signaling pathway further modulates the adhesion between platelets and immune cells and remodeling of their cytoskeleton [120].

PLATELET COMPONENTS THAT CONTROL CELL DIFFERENTIATION

Recent studies have shown that platelets are also involved in the differentiation of immune and endothelial precursor cells. For example, in co-cultures with platelets, CD34⁺ stem cells differentiate into CD68-immunoreactive macrophages capable of forming foam cells [121]. This interaction of bone marrow cells with platelets involves the intervention of P-selectin and the GPIIb integrin [122]. SDF-1 secretion by activated platelets contributes to vascular remodeling through the recruitment and differentiation of bone marrow precursors [123, 124]. This process is mediated by CXCR4, CD184 and vascular endothelial growth factor receptor-1 (VEGFR-1) [125]. This SDF-1-dependent mechanism favors neo-vascularization of ischemic organs in which platelets accumulate (e.g. heart, liver and brain), thus contributing to their functional and structural repair [126]. Nevertheless, SDF-1 production by activated platelets can also trigger noxious responses by inducing a pro-inflammatory phenotype in hematopoietic precursors recruited to the arterial wall [127].

CONCLUSIONS

In addition to their well-recognized role in promoting thrombus formation after atherosclerotic plaque rupture, platelets exert pro-atherogenic actions by exacerbating inflammation at all stages of atherosclerosis development. This is achieved in part by facilitating the interaction between immune and endothelial cells through molecules such as sCD40L, P-selectin, RANTES, TLRs,

and SDF-1, even in the absence of platelet aggregation. These new findings provide evidence that platelet activation is an attractive therapeutic target for prevention of the transition from chronic to acute inflammation and the subsequent formation of atherosclerotic plaque and thrombosis.

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Declaration of interest

The authors report no declarations of interest.

REFERENCES

1. Mackay J, Mensah GA. The atlas of heart disease and stroke. Geneva : World Health Organization, 2004.
2. AHA Statistical Fact Sheet. International Cardiovascular Disease Statistics. American Heart Association, 2003.
3. Reddy KS, Yusuf S. Emerging epidemic of cardiovascular disease in developing countries. *Circulation* 1998;97:596-601.
4. Marenberg ME, Risch N, Berkman LF, Floderus B, de Faire U. Genetic susceptibility to death from coronary heart disease in a study of twins. *N Engl J Med* 1994;330:1041-6.
5. Inoue K, Kawahara K, Biswas KK, Ando K, Mitsudo K, Nobuyoshi M, Maruyama I. HMGB1 expression by activated vascular smooth muscle cells in advanced human atherosclerosis plaques. *Cardiovasc Pathol* 2007;16:136-43.
6. Andres V. Control of vascular cell proliferation and migration by cyclin-dependent kinase signalling: new perspectives and therapeutic potential. *Cardiovasc Res* 2004;63:11-21.
7. Braun-Dullaeus RC, Mann MJ, Seay U, Zhang L, von Der Leyen HE, Morris RE, Dzau VJ. Cell cycle protein expression in vascular smooth muscle cells in vitro and in vivo is regulated through phosphatidylinositol 3-kinase and mammalian target of rapamycin. *Arterioscler Thromb Vasc Biol* 2001;21:1152-1158.
8. von Hundelshausen P, Weber C. Platelets as immune cells: bridging inflammation and cardiovascular disease. *Circ Res* 2007;100:27-40.
9. Wagner DD, Burger PC. Platelets in inflammation and thrombosis. *Arterioscler Thromb Vasc Biol* 2003;23:2131-7.
10. Lievens D, Zerneck A, Seijkens T, Soehnlein O, Beckers L, Munnix IC, Wijnands E, et al. Platelet CD40L mediates thrombotic and inflammatory processes in atherosclerosis. *Blood* 2010;116:4317-27.

11. Massberg S, Schürzinger K, Lorenz M, Konrad I, Schulz C, Plesnila N, Kennerknecht E, et al. Platelet adhesion via glycoprotein IIb integrin is critical for atheroprogession and focal cerebral ischemia: an in vivo study in mice lacking glycoprotein IIb. *Circulation* 2005;112:1180-8.
12. Aukrust P, Halvorsen B, Ueland T, Michelsen AE, Skjelland M, Gullestad L, Yndestad A, et al. Activated platelets and atherosclerosis. *Expert Rev Cardiovasc Ther* 2010;8:1297-1307.
13. Nishijima K, Kiryu J, Tsujikawa A, Miyamoto K, Honjo M, Tanihara H, Nonaka A, et al. Platelets adhering to the vascular wall mediate postischemic leukocyte-endothelial cell interactions in retinal microcirculation. *Invest Ophthalmol Vis Sci* 2004;45:977-84.
14. Ishikawa M, Cooper D, Arumugam TV, Zhang JH, Nanda A, Granger DN. Platelet-leukocyte-endothelial cell interactions after middle cerebral artery occlusion and reperfusion. *J Cereb Blood Flow Metab* 2004;24:907-15.
15. van der Wal AC, Becker AE, van der Loos CM, Das PK. Site of intimal rupture or erosion of thrombosed coronary atherosclerotic plaques is characterized by an inflammatory process irrespective of the dominant plaque morphology. *Circulation* 1994;89:36-44.
16. Libby P, Ridker PM, Maseri A. Inflammation and atherosclerosis. *Circulation* 2002 5;105:1135-1143.
17. Matijevic N, Wu KK, Howard AG, Wasserman B, Wang WY, Folsom AR, Sharrett AR. Association of blood monocyte and platelet markers with carotid artery characteristics: the atherosclerosis risk in communities carotid MRI study. *Cerebrovasc Dis* 2011;31:552-558.
18. Gómez M, Sanz-González SM, Abu Nabah YN, Lamana A, Sánchez-Madrid F, Andrés V. Atherosclerosis development in apolipoprotein E-null mice deficient for CD69. *Cardiovasc Res* 2009;81:197-205.

19. Rong S, Cao Q, Liu M, Seo J, Jia L, Boudyguina E, Gebre AK, et al. Macrophage 12/15 lipoxygenase expression increases plasma and hepatic lipid levels and exacerbates atherosclerosis. *J Lipid Res* 2012 [Epub ahead of print].
20. Newby AC. Metalloproteinase expression in monocytes and macrophages and its relationship to atherosclerotic plaque instability. *Arterioscler ThrombVasc Biol* 2008;28:2108-2114.
21. Hutter R, Valdiviezo C, Sauter BV, Savontaus M, Chereshnev I, Carrick FE, Bauriedel G, et al. Caspase-3 and tissue factor expression in lipid-rich plaque macrophages: evidence for apoptosis as link between inflammation and atherothrombosis. *Circulation* 2004;109:2001-2008.
22. Shah PK, Falk E, Badimon JJ, Fernandez-Ortiz A, Mailhac A, Villareal-Levy G, Fallon JT, et al. Human monocyte-derived macrophages induce collagen breakdown in fibrous caps of atherosclerotic plaques. Potential role of matrix-degrading metalloproteinases and implications for plaque rupture. *Circulation* 1995;92:1565-9.
23. Meisel SR, Xu XP, Edgington TS, Cercek B, Ong J, Kaul S, Shah PK. Dose-dependent modulation of tissue factor protein and procoagulant activity in human monocyte-derived macrophages by oxidized low density lipoprotein. *J Atheroscler Thromb* 2011;18:596-603.
24. Barascuk N, Skjøt-Arkil H, Register TC, Larsen L, Byrjalsen I, Christiansen C, Karsdal MA. Human macrophage foam cells degrade atherosclerotic plaques through cathepsin K mediated processes. *BMC Cardiovasc Disord* 2010;10:19.
25. Galis ZS, Sukhova GK, Lark MW, Libby P. Increased expression of matrix metalloproteinases and matrix degrading activity in vulnerable regions of human atherosclerotic plaques. *J Clin Invest* 1994;94:2493-503.
26. Tavora FR, Ripple M, Li L, Burke AP. Monocytes and neutrophils expressing myeloperoxidase occur in fibrous caps and thrombi in unstable coronary plaques. *BMC Cardiovasc Disord* 2009;9:27.

27. Segers D, Helderma F, Cheng C, van Damme LC, Tempel D, Boersma E, Serruys PW, et al. Gelatinolytic activity in atherosclerotic plaques is highly localized and is associated with both macrophages and smooth muscle cells in vivo. *Circulation* 2007;115:609-16.
28. Galis ZS, Sukhova GK, Lark MW, Libby P. Increased expression of matrix metalloproteinases and matrix degrading activity in vulnerable regions of human atherosclerotic plaques. *J Clin Invest* 1994;94:2493-503.
29. Loftus IM, Naylor AR, Goodall S, Crowther M, Jones L, Bell PR, Thompson MM. Increased matrix metalloproteinase-9 activity in unstable carotid plaques. A potential role in acute plaque disruption. *Stroke* 2000;31:40-7.
30. Huo Y, Schober A, Forlow SB, Smith DF, Hyman MC, Jung S, Littman DR, et al. Circulating activated platelets exacerbate atherosclerosis in mice deficient in apolipoprotein E. *Nat Med* 2003;9:61-7.
31. Fuster JJ, Fernández P, González-Navarro H, Silvestre C, Nabah YN, Andrés V. Control of cell proliferation in atherosclerosis: insights from animal models and human studies. *Cardiovasc Res* 2010;86:254-264.
32. Díez-Juan A, Andrés V. The growth suppressor p27(Kip1) protects against diet-induced atherosclerosis. *FASEB J* 2001;15:1989-1995.
33. Castro C, Díez-Juan A, Cortés MJ, Andrés V. Distinct regulation of mitogen-activated protein kinases and p27Kip1 in smooth muscle cells from different vascular beds. A potential role in establishing regional phenotypic variance. *J Biol Chem* 2003;278:4482-4490.
34. Sakakibara K, Kubota K, Worku B, Ryer EJ, Miller JP, Koff A, Kent KC, et al. PDGF-BB regulates p27 expression through ERK-dependent RNA turn-over in vascular smooth muscle cells. *J Biol Chem* 2005;280:25470-7.

35. Díez-Juan A, Pérez P, Aracil M, Sancho D, Bernad A, Sánchez-Madrid F, Andrés V. Selective inactivation of p27(Kip1) in hematopoietic progenitor cells increases neointimal macrophage proliferation and accelerates atherosclerosis. *Blood* 2004;103:158-161.
36. Wragg A, Mellad JA, Beltran LE, Konoplyannikov M, San H, Boozer S, Deans RJ, et al. VEGFR1/CXCR4-positive progenitor cells modulate local inflammation and augment tissue perfusion by a SDF-1-dependent mechanism. *J Mol Med (Berl)* 2008;86:1221-32.
37. Blair P, Flaumenhaft R. Platelet alpha-granules: basic biology and clinical correlates. *Blood Rev* 2009;23:177-189.
38. Hamilton JR, Cornelissen I, Mountford JK, Coughlin SR. Atherosclerosis proceeds independently of thrombin-induced platelet activation in ApoE^{-/-} mice. *Atherosclerosis* 2009;205:427-32.
39. Manka D, Collins RG, Ley K, Beaudet AL, Sarembock IJ. Absence of p-selectin, but not intercellular adhesion molecule-1, attenuates neointimal growth after arterial injury in apolipoprotein e-deficient mice. *Circulation* 2001;103:1000-5.
40. Koyama H, Maeno T, Fukumoto S, Shoji T, Yamane T, Yokoyama H, Emoto M, et al. Platelet P-selectin expression is associated with atherosclerotic wall thickness in carotid artery in humans. *Circulation* 2003;108:524-9.
41. Schoenwaelder SM, Yuan Y, Josefsson EC, White MJ, Yao Y, Mason KD, O'Reilly LA, et al. Two distinct pathways regulate platelet phosphatidylserine exposure and procoagulant function. *Blood* 2009;114:663-666.
42. Italiano JE Jr, Richardson JL, Patel-Hett S, Battinelli E, Zaslavsky A, Short S, Ryeom S, et al. Angiogenesis is regulated by a novel mechanism: pro- and antiangiogenic proteins are organized into separate platelet alpha granules and differentially released. *Blood* 2008;111:1227-1233.
43. Antczak AJ, Singh N, Gay SR, Worth RG. IgG-complex stimulated platelets: a source of sCD40L and RANTES in initiation of inflammatory cascade. *Cell Immunol* 2010;263:129-133.

44. Eisenhardt SU, Habersberger J, Peter K. Monomeric C-reactive protein generation on activated platelets: the missing link between inflammation and atherothrombotic risk. *Trends Cardiovasc Med* 2009;19:232-237.
45. Kirbis S, Breskvar UD, Sabovic M, Zupan I, Sinkovic A. Inflammation markers in patients with coronary artery disease--comparison of intracoronary and systemic levels. *Wien KlinWochenschr* 2010;122 Suppl 2:31-4.
46. Ding S, Zhang M, Zhao Y, Chen W, Yao G, Zhang C, Zhang P, et al. The role of carotid plaque vulnerability and inflammation in the pathogenesis of acute ischemic stroke. *Am J Med S* 2008;336:27-31.
47. Schulz C, Penz S, Hoffmann C, Langer H, Gillitzer A, Schneider S, Brandl R, et al. Platelet GPVI binds to collagenous structures in the core region of human atheromatous plaque and is critical for atheroprogession in vivo. *Basic Res Cardiol* 2008;103:356-67.
48. Vaitaitis G, Waid D, Wagner D. The Expanding Role of TNF-Receptor Super Family Member CD40 (tnfrsf5) in Autoimmune Disease: Focus on Th40 Cells. *Current Immunology Reviews* 2010;6:130-136.
49. Kotowicz K, Dixon GL, Klein NJ, Peters MJ, Callard RE. Biological function of CD40 on human endothelial cells: costimulation with CD40 ligand and interleukin-4 selectively induces expression of vascular cell adhesion molecule-1 and P-selectin resulting in preferential adhesion of lymphocytes. *Immunology* 2000;100:441-8.
50. Koguchi Y, Buenafe AC, Thauland TJ, Gardell JL, Bivins-Smith ER, Jacoby DB, Slifka MK, et al. Preformed CD40L Is Stored in Th1, Th2, Th17, and T Follicular Helper Cells as Well as CD48 Thymocytes and Invariant NKT Cells but Not in Treg Cells. *PLoS One* 2012;7:e31296.
51. Missiou A, Wolf D, Platzer I, Ernst S, Walter C, Rudolf P, Zirlik K, et al. CD40L induces inflammation and adipogenesis in adipose cells--a potential link between metabolic and cardiovascular disease. *ThrombHaemost* 2010;103:788-796.

52. Palomo IG, Jaramillo JC, Alarcón ML, Gutiérrez CL, Moore-Carrasco R, Segovia FM, Leiva EM, et al. Increased concentrations of soluble vascular cell adhesion molecule-1 and soluble CD40L in subjects with metabolic syndrome. *Mol Med Report* 2009;2:481-485.
53. Setianto BY, Hartopo AB, Gharini PP, Anggrahini DW, Irawan B. Circulating soluble CD40 ligand mediates the interaction between neutrophils and platelets in acute coronary syndrome. *Heart Vessels* 2010;25:282-287.
54. Reinboldt S, Wenzel F, Rauch BH, Hohlfeld T, Grandoch M, Fischer JW, Weber AA. Preliminary evidence for a matrix metalloproteinase-2 (MMP-2)-dependent shedding of soluble CD40 ligand (sCD40L) from activated platelets. *Platelets* 2009;20:441-444.
55. Santilli F, Davì G, Consoli A, Cipollone F, Mezzetti A, Falco A, Taraborelli T, et al. Thromboxane-dependent CD40 ligand release in type 2 diabetes mellitus. *J Am CollCardiol* 2006;47:391-397.
56. Yacoub D, Hachem A, Théorêt JF, Gillis MA, Mourad W, Merhi Y. Enhanced levels of soluble CD40 ligand exacerbate platelet aggregation and thrombus formation through a CD40-dependent tumor necrosis factor receptor-associated factor-2/Rac1/p38 mitogen-activated protein kinase signaling pathway. *ArteriosclerThrombVasc Biol* 2010;30:2424-2433.
57. Schönbeck U, Sukhova GK, Shimizu K, Mach F, Libby P. Inhibition of CD40 signaling limits evolution of established atherosclerosis in mice. *Proc Natl Acad Sci USA* 2000;97:7458–7463.
58. Lutgens E, Cleutjens KB, Heeneman S, Koteliansky VE, Burkly LC, Daemen MJ. Both early and delayed anti-CD40L antibody treatment induces a stable plaque phenotype. *Proc Natl Acad Sci USA* 2000;97:7464–7469.
59. Mach F, Schönbeck U, Bonnefoy JY, Pober JS, Libby P. Activation of monocyte/macrophage functions related to acute atheroma complication by ligation of CD40: induction of collagenase, stromelysin, and tissue factor. *Circulation* 1997;96:396-9.

60. Zirlik A, Maier C, Gerdes N, MacFarlane L, Soosairajah J, Bavendiek U, Ahrens I, et al. CD40 ligand mediates inflammation independently of CD40 by interaction with Mac-1. *Circulation* 2007;115:1571-1580.
61. Léveillé C, Bouillon M, Guo W, Bolduc J, Sharif-Askari E, El-Fakhry Y, Reyes-Moreno C, et al. CD40 ligand binds to alpha5beta1 integrin and triggers cell signaling. *J Biol Chem* 2007;282:5143-5151.
62. Czapiga M, Kirk AD, Lekstrom-Himes J. Platelets deliver costimulatory signals to antigen-presenting cells: a potential bridge between injury and immune activation. *ExpHematol* 2004;32:135-139.
63. Sprague DL, Elzey BD, Crist SA, Waldschmidt TJ, Jensen RJ, Ratliff TL. Platelet-mediated modulation of adaptive immunity: unique delivery of CD154 signal by platelet-derived membrane vesicles. *Blood* 2008;111:5028-5036.
64. Elzey BD, Grant JF, Sinn HW, Nieswandt B, Waldschmidt TJ, Ratliff TL. Cooperation between platelet-derived CD154 and CD4+ T cells for enhanced germinal center formation. *J Leukoc Biol* 2005;78:80-84.
65. Vanichakarn P, Blair P, Wu C, Freedman JE, Chakrabarti S. Neutrophil CD40 enhances platelet-mediated inflammation. *Thromb Res* 2008;122:346-358.
66. Karmann K, Hughes CC, Schechner J, Fanslow WC, Pober JS. CD40 on human endothelial cells: inducibility by cytokines and functional regulation of adhesion molecule expression. *Proc Natl Acad Sci USA* 1995;92:4342-6.
67. Henn V, Slupsky JR, Gräfe M, Anagnostopoulos I, Förster R, Müller-Berghaus G, Kroczeck RA. CD40 ligand on activated platelets triggers an inflammatory reaction of endothelial cells. *Nature*. 1998;391:591-4.
68. Miller DL, Yaron R, Yellin MJ. CD40L-CD40 interactions regulate endothelial cell surface tissue factor and thrombomodulin expression. *J Leukoc Biol* 1998;63:373-9.

69. Chai H, Aghaie K, Zhou W. Soluble CD40 ligand induces human coronary artery smooth muscle cells proliferation and migration. *Surgery* 2009;146:5-11.
70. Vogel J, West G, Sturm A, Levine A, Fiocchi C. Essential role of the CD40 pathway in T-cell-mediated induction of chemokines and cell adhesion molecules by human intestinal fibroblasts (HIF) and microvascular endothelial cells (HIMEC). *Gastroenterology* 2001;120:A192.
71. Yellin MJ, Winikoff S, Fortune SM, Baum D, Crow MK, Lederman S, Chess L. Ligation of CD40 on fibroblasts induces CD54 (ICAM-1) and CD106 (VCAM-1) up-regulation and IL-6 production and proliferation. *J Leukoc Biol* 1995;58:209-16.
72. Mach F, Schönbeck U, Sukhova GK, Bourcier T, Bonnefoy JY, Pober JS, Libby P. Functional CD40 ligand is expressed on human vascular endothelial cells, smooth muscle cells, and macrophages: implications for CD40-CD40 ligand signaling in atherosclerosis. *Proc Natl Acad Sci USA* 1997;94:1931-6.
73. McEver RP, Beckstead JH, Moore KL, Marshall-Carlson L, Bainton DF. GMP-140, a platelet α -granule membrane protein, is also synthesized by vascular endothelial cells and is localized in Weibel-Palade bodies. *J Clin Invest* 1989;84:92-99.
74. Ley K, Laudanna C, Cybulsky MI, Nourshargh S. Getting to the site of inflammation: the leukocyte adhesion cascade updated. *Nat Rev Immunol* 2007;7:678-689.
75. Ludwig RJ, Schultz JE, Boehncke WH, Podda M, Tandi C, Krombach F, Baatz H, et al. Activated, not resting, platelets increase leukocyte rolling in murine skin utilizing a distinct set of adhesion molecules. *J Invest Dermatol* 2004;122:830-6.
76. Li G, Sanders JM, Phan ET, Ley K, Sarembock IJ. Arterial macrophages and regenerating endothelial cells express P-selectin in atherosclerosis-prone apolipoprotein E-deficient mice. *Am J Pathol* 2005;167:1511-1518.
77. Phillips JW, Barringhaus KG, Sanders JM, Hesselbacher SE, Czarnik AC, Manka D, Vestweber D, et al. Single injection of P-selectin or P-selectin glycoprotein ligand-1 monoclonal

antibody blocks neointima formation after arterial injury in apolipoprotein E-deficient mice. *Circulation* 2003;107:2244-9.

78. Schulz C, Schäfer A, Stolla M, Kerstan S, Lorenz M, von Brühl ML, Schiemann M, et al. Chemokine fractalkine mediates leukocyte recruitment to inflammatory endothelial cells in flowing whole blood: a critical role for P-selectin expressed on activated platelets. *Circulation* 2007;116:764-773.

79. Tjwa M, Bellido-Martin L, Lin Y, Lutgens E, Plaisance S, Bono F, Delesque-Touchard N, et al. Gas6 promotes inflammation by enhancing interactions between endothelial cells, platelets, and leukocytes. *Blood* 2008;111:4096-4105.

80. Yang H, Lang S, Zhai Z, Li L, Kahr WH, Chen P, Brkić J, et al. Fibrinogen is required for maintenance of platelet intracellular and cell-surface P-selectin expression. *Blood* 2009;114:425-436.

81. Burger PC, Wagner DD. Platelet P-selectin facilitates atherosclerotic lesion development. *Blood*. 2003;101:2661-6.

82. Yokoyama S, Ikeda H, Haramaki N, Yasukawa H, Murohara T, Imaizumi T. Platelet P-selectin plays an important role in arterial thrombogenesis by forming large stable platelet-leukocyte aggregates. *J Am Coll Cardiol* 2005;45:1280-6.

83. Wang K, Zhou X, Zhou Z, Mal N, Fan L, Zhang M, Lincoff AM, et al. Platelet, not endothelial, P-selectin is required for neointimal formation after vascular injury. *Arterioscler Thromb Vasc Biol* 2005;25:1584-9.

84. Ishiwata N, Takio K, Katayama M, Watanabe K, Titani K, Ikeda Y, Handa M. Alternatively spliced isoform of P-selectin is present in vivo as a soluble molecule. *J BiolChem* 1994;269:23708-23715.

85. Stellos K, Bigalke B, Stakos D, Henkelmann N, Gawaz M. Platelet-bound P-selectin expression in patients with coronary artery disease: impact on clinical presentation and

myocardial necrosis, and effect of diabetes mellitus and anti-platelet medication. *J ThrombHaemost* 2010;8:205-207.

86. Preston RA, Coffey JO, Materson BJ, Ledford M, Alonso AB. Elevated platelet P-selectin expression and platelet activation in high risk patients with uncontrolled severe hypertension. *Atherosclerosis* 2007;192:148-154.

87. Fox SC, May JA, Shah A, Neubert U, Heptinstall S. Measurement of platelet P-selectin for remote testing of platelet function during treatment with clopidogrel and/or aspirin. *Platelets* 2009;20:250-9.

88. Kisucka J, Chauhan AK, Zhao BQ, Patten IS, Yesilaltay A, Krieger M, Wagner DD. Elevated levels of soluble P-selectin in mice alter blood-brain barrier function, exacerbate stroke, and promote atherosclerosis. *Blood* 2009;113:6015-6022.

89. Dole VS, Bergmeier W, Patten IS, Hirahashi J, Mayadas TN, Wagner DD. PSGL-1 regulates platelet P-selectin-mediated endothelial activation and shedding of P-selectin from activated platelets. *ThrombHaemost* 2007;98:806-812.

90. Kuckleburg CJ, Yates CM, Kalia N, Zhao Y, Nash GB, Watson SP, Rainger GE. Endothelial cell-borne platelet bridges selectively recruit monocytes in human and mouse models of vascular inflammation. *Cardiovasc Res* 2011;91:134-141.

91. Wang HB, Wang JT, Zhang L, Geng ZH, Xu WL, Xu T, Huo Y, et al. P-selectin primes leukocyte integrin activation during inflammation. *Nat Immunol* 2007;8:882-892.

92. Luo BH, Carman CV, Springer TA. Structural basis of integrin regulation and signaling. *Annu Rev Immunol* 2007;25:619-647.

93. Zarbock A, Polanowska-Grabowska RK, Ley K. Platelet-neutrophil-interactions: linking hemostasis and inflammation. *Blood Rev* 2007;21:99-111.

94. Brown KK, Henson PM, Maclouf J, Moyle M, Ely JA, Worthen GS. Neutrophil-platelet adhesion: relative roles of platelet P-selectin and neutrophil beta2 (DC18) integrins. *Am J Respir Cell Mol Biol* 1998;18:100-10.
95. Miner JJ, Xia L, Yago T, Kappelmayer J, Liu Z, Klopocki AG, Shao B, et al. Separable requirements for cytoplasmic domain of PSGL-1 in leukocyte rolling and signaling under flow. *Blood* 2008;112:2035-2045.
96. Weber C, Weber KS, Klier C, Gu S, Wank R, Horuk R, Nelson PJ. Specialized roles of the chemokine receptors CCR1 and CCR5 in the recruitment of monocytes and T(H)1-like/CD45RO(+) T cells. *Blood* 2001;97:1144-6.
97. Nishi K, Itabe H, Uno M, Kitazato KT, Horiguchi H, Shinno K, Nagahiro S. Oxidized LDL in carotid plaques and plasma associates with plaque instability. *Arterioscler Thromb Vasc Biol* 2002;22:1649-54.
98. Virani SS, Nambi V, Hoogeveen R, Wasserman BA, Coresh J, Gonzalez F, Chambless LE, et al. Relationship between circulating levels of RANTES (regulated on activation, normal T-cell expressed, and secreted) and carotid plaque characteristics: the Atherosclerosis Risk in Communities (ARIC) Carotid MRI Study. *Eur Heart J* 2011;32:459-468.
99. Terao S, Yilmaz G, Stokes KY, Russell J, Ishikawa M, Kawase T, Granger DN. Blood cell-derived RANTES mediates cerebral microvascular dysfunction, inflammation, and tissue injury after focal ischemia-reperfusion. *Stroke* 2008;39:2560-2570.
100. Koh SJ, Kim JY, Hyun YJ, Park SH, Chae JS, Park S, Kim JS, et al. Association of serum RANTES concentrations with established cardiovascular risk markers in middle-aged subjects. *Int J Cardiol* 2009;132:102-108.
101. Dénes A, Humphreys N, Lane TE, Grecis R, Rothwell N. Chronic systemic infection exacerbates ischemic brain damage via a CCL5 (regulated on activation, normal T-cell expressed and secreted)-mediated proinflammatory response in mice. *J Neurosci* 2010;30:10086-10095.

102. Koenen RR, von Hundelshausen P, Nesmelova IV, Zerneck A, Liehn EA, Sarabi A, Kramp BK, et al. Disrupting functional interactions between platelet chemokines inhibits atherosclerosis in hyperlipidemic mice. *Nat Med* 2009;15:97-103.
103. von Hundelshausen P, Koenen RR, Sack M, Mause SF, Adriaens W, Proudfoot AE, Hackeng TM, et al. Heterophilic interactions of platelet factor 4 and RANTES promote monocyte arrest on endothelium. *Blood* 2005;105:924-30.
104. Schober A, Manka D, von Hundelshausen P, Huo Y, Hanrath P, Sarembock IJ, Ley K, et al. Deposition of platelet RANTES triggering monocyte recruitment requires P-selectin and is involved in neointima formation after arterial injury. *Circulation* 2002;106:1523-9.
105. Weyrich AS, Elstad MR, McEver RP, McIntyre TM, Moore KL, Morrissey JH, Prescott SM, et al. Activated platelets signal chemokine synthesis by human monocytes. *J Clin Invest* 1996;97:1525-34.
106. Leulier F, Lemaitre B. Toll-like receptors--taking an evolutionary approach. *Nat Rev Genet* 2008;9:165-178.
107. Lin E, Freedman JE, Beaulieu LM. Innate immunity and toll-like receptor antagonists: a potential role in the treatment of cardiovascular diseases. *Cardiovasc Ther* 2009;27:117-123.
108. Cognasse F, Hamzeh H, Chavarin P, Acquart S, Genin C, Garraud O. Evidence of Toll-like receptor molecules on human platelets. *Immunol Cell Biol* 2005;83:196-198.
109. Chearwae W, Bright JJ. 15-deoxy-Delta (12,14)-prostaglandin J(2) and curcumin modulate the expression of toll-like receptors 4 and 9 in autoimmune T lymphocytes. *J Clin Immunol* 2008;28:558-70.
110. Erridge C. Endogenous ligands of TLR2 and TLR4: agonists or assistants?. *J Leukoc Biol* 2010;87:989-999.

111. Wyss CA, Neidhart M, Altwegg L, Spanaus KS, Yonekawa K, Wischnewsky MB, Corti R, et al. Cellular actors, Toll-like receptors, and local cytokine profile in acute coronary syndromes. *Eur Heart J* 2010;31:1457-1469.
112. Zhang G, Han J, Welch EJ, Ye RD, Voyno-Yasenetskaya TA, Malik AB, Du X, et al. Lipopolysaccharide stimulates platelet secretion and potentiates platelet aggregation via TLR4/MyD88 and the cGMP-dependent protein kinase pathway. *J Immunol* 2009;182:7997-8004.
113. Brown GT, McIntyre TM. Lipopolysaccharide signaling without a nucleus: kinase cascades stimulate platelet shedding of proinflammatory IL-1 β -rich microparticles. *J Immunol* 2011;186:5489-5496.
114. Linden MD, Jackson DE. Platelets: pleiotropic roles in atherogenesis and atherothrombosis. *Int J Biochem Cell Biol* 2010;42:1762-1766.
115. Gawaz M, Brand K, Dickfeld T, Pogatsa-Murray G, Page S, Bogner C, Koch W, et al. Platelets induce alterations of chemotactic and adhesive properties of endothelial cells mediated through an interleukin-1-dependent mechanism. Implications for atherogenesis. *Atherosclerosis* 2000;148:75-85.
116. Cognasse F, Hamzeh-Cognasse H, Lafarge S, Delezay O, Pozzetto B, McNicol A, Garraud O. Toll-like receptor 4 ligand can differentially modulate the release of cytokines by human platelets. *Br J Haematol* 2008;141:84-91.
117. Schoneveld AH, Hofer I, Sluijter JP, Laman JD, de Kleijn DP, Pasterkamp G. Atherosclerotic lesion development and Toll like receptor 2 and 4 responsiveness. *Atherosclerosis* 2008;197:95-104.
118. Blair P, Rex S, Vitseva O, Beaulieu L, Tanriverdi K, Chakrabarti S, Hayashi C, et al. Stimulation of Toll-like receptor 2 in human platelets induces a thromboinflammatory response through activation of phosphoinositide 3-kinase. *Circ Res* 2009;104:346-354.

119. Kälvegren H, Skoglund C, Helldahl C, Lerm M, Grenegård M, Bengtsson T. Toll-like receptor 2 stimulation of platelets is mediated by purinergic P2X1-dependent Ca²⁺ mobilisation, cyclooxygenase and purinergic P2Y1 and P2Y12 receptor activation. *ThrombHaemost* 2010;103:398-407.
120. Rex S, Beaulieu LM, Perlman DH, Vitseva O, Blair PS, McComb ME, Costello CE, et al. Immune versus thrombotic stimulation of platelets differentially regulates signalling pathways, intracellular protein-protein interactions, and alpha-granule release. *ThrombHaemost* 2009;102:97-110.
121. Daub K, Langer H, Seizer P, Stellos K, May AE, Goyal P, Bigalke B, et al. Platelets induce differentiation of human CD34+ progenitor cells into foam cells and endothelial cells. *FASEB J* 2006;20:2559-2561.
122. Massberg S, Konrad I, Schürzinger K, Lorenz M, Schneider S, Zohlnhoefer D, Hoppe K, et al. Platelets secrete stromal cell-derived factor 1alpha and recruit bone marrow-derived progenitor cells to arterial thrombi in vivo. *J Exp Med* 2006;203:1221-33.
123. Stellos K, Seizer P, Bigalke B, Daub K, Geisler T, Gawaz M. Platelet aggregates-induced human CD34+ progenitor cell proliferation and differentiation to macrophages and foam cells is mediated by stromal cell derived factor 1 in vitro. *SeminThrombHemost* 2010;36:139-145.
124. Rafii DC, Psaila B, Butler J, Jin DK, Lyden D. Regulation of vasculogenesis by platelet-mediated recruitment of bone marrow-derived cells. *ArteriosclerThrombVasc Biol* 2008;28:217-222.
125. Petit I, Jin D, Rafii S. The SDF-1-CXCR4 signaling pathway: a molecular hub modulating neo-angiogenesis. *Trends Immunol* 2007;28:299-307.
126. Stellos K, Langer H, Daub K, Schoenberger T, Gauss A, Geisler T, Bigalke B, et al. Platelet-derived stromal cell-derived factor-1 regulates adhesion and promotes differentiation of human CD34+ cells to endothelial progenitor cells. *Circulation* 2008;117:206-215.

127. Gawaz M, Stellos K, Langer HF. Platelets modulate atherogenesis and progression of atherosclerotic plaques via interaction with progenitor and dendritic cells. *J ThrombHaemost* 2008;6:235-242.

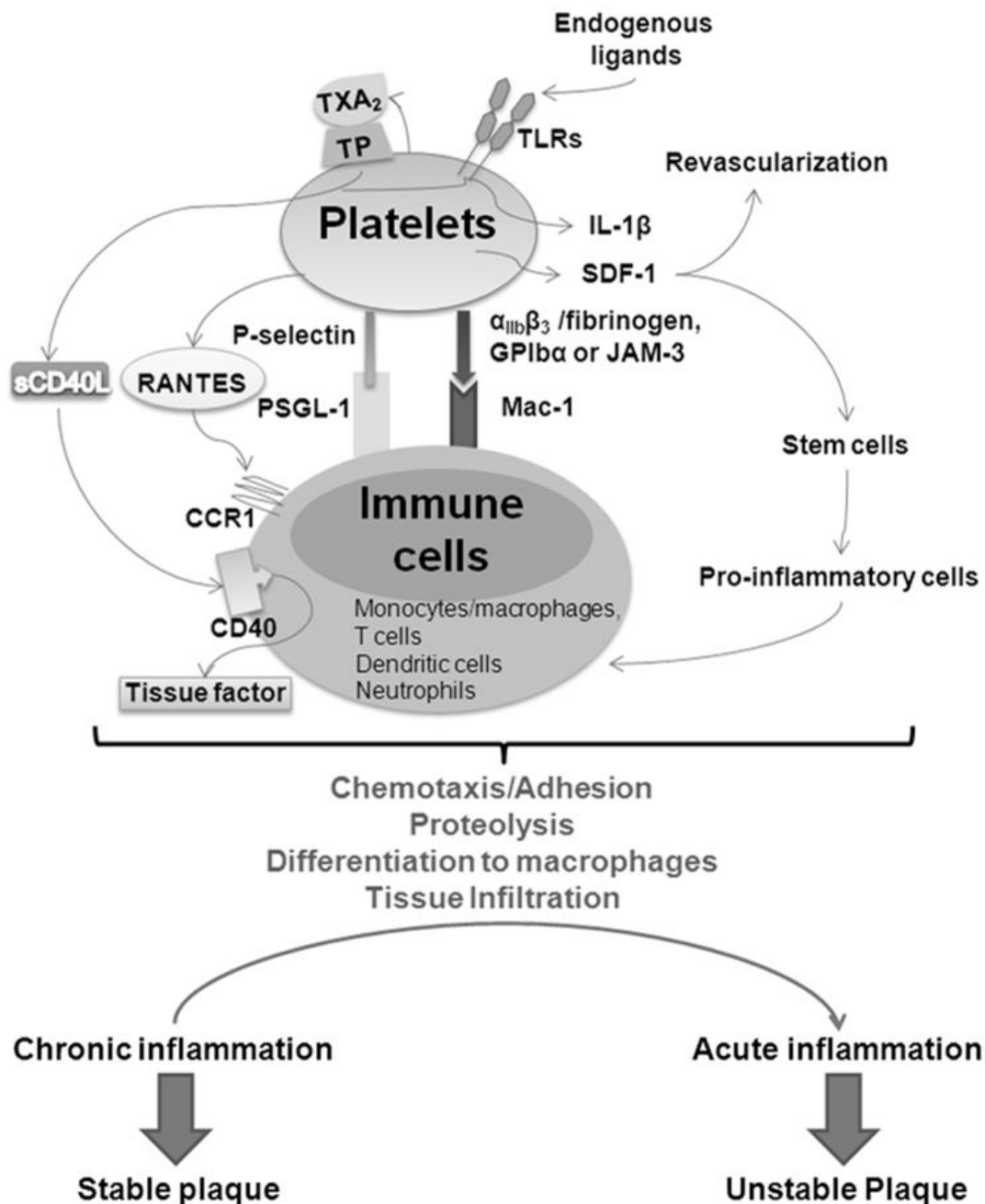


Figure 1. Role of platelets in the vascular inflammatory response associated with atherosclerosis. Platelets promote inflammation in the arterial wall in part through interaction with immune cells. Even without forming aggregates, platelets produce inflammatory molecules (sCD40L, P-selectin, RANTES and TLRs) that contribute to all stages of atherosclerosis, from chronic phases of atheroma initiation and growth to acute disease characterized by plaque rupture and ensuing thrombosis and ischemic events (myocardial infarction or stroke). Through the recruitment and differentiation of progenitor cells mobilized from the bone marrow, SDF-1 produced by platelets can have both beneficial effects (enhancement of angiogenesis) and harmful effects (exacerbation of inflammation) in the artery wall. CCR1, Chemokine (C-C motif) receptor 1; JAM-3, Junctional adhesion molecule 3; TXA₂, Thromboxane A₂; TP, Thromboxane A₂ receptor; TLRs, Toll-like receptors; IL-1β, Interleukin-1 beta; SDF-1, Stroma-cell derived factor-1; GPIbα, glycoprotein Iba; PSGL-1, P-Selectin Glycoprotein Ligand 1; Mac-1, Macrophage 1 antigen.