Gross N-cycling Rates in Ephemeral Wetlands

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Abstract

Ephemeral wetlands or depressions in hummocky landscapes have high levels of C, N, and soil moisture, often leading to high nutrient cycling activity. However, measuring soil nitrate and ammonium pools is typically a poor indication of N-cycling activity or of the soil N that is available for other processes such as N₂O emissions. This study used stable ¹⁵N isotope dilution techniques in cultivated and uncultivated ephemeral wetlands in central Saskatchewan to quantify land use effects on gross mineralization and nitrification rates. In-field incubation experiments were repeated in early May, mid-June and late July. There was a clear land use effect on inorganic soil N levels, with significantly less NH₄⁺ and more NO₃⁻ in the cultivated wetland soils. However, the rates of NH₄⁺ mineralization and NO₃⁻ nitrification were similar for both land uses, indicating similar substrate availability but different N-consuming processes. Both N pools turned over in as little as 1-2 d, highlighting the ineffectuality of measuring inorganic N pools as a predictor for N availability in these soils.

Introduction

Hummocky till landscapes are the dominant geomorphic surface in the Prairie Pothole region, which extends from East-Central Alberta through to North-Central Iowa. These landscapes contain a very high number of wetlands – for example, our research site near St. Denis, Saskatchewan, Canada contains 216 wetlands in an area of 3.84 km² (Hogan and Conly, 2002) – interspersed within the cultivated upland portions of the landscape. Many of the wetlands have been cleared of natural vegetation and are now used for annual crop production. This conversion from native wetland to cultivated wetlands has presumably altered the rates of fundamental N processes but to date little or no information is available on the rates of N-cycling processes in these landscape positions.

Cropped landscapes receive nitrogen (N) inputs in the form of inorganic and/or organic fertilizers, and these inputs have a significant impact on local and regional N cycling, N gas losses, and N leaching (Burke et al., 2002). N-cycling processes of particular interest for both agricultural and environmental concerns are: 1) mineralization, which produces NH_4^+ from the soil organic N pool and 2) nitrification, which produces NO_3^- from the NH_4^+ and organic N pools and emits nitrous oxide (N₂O) as a by-product. Nitrous oxide is a greenhouse gas with 296 times the global warming potential of CO_2 . In Canada, agriculture accounts for up to 70% of anthropogenic N₂O emissions (Janzen et al., 1998).

Typical assessments of N pool status and activity include "snapshot" tests of soil NH_4^+ or NO_3^- pools at one or more times in the growing season, or measures of net mineralization and nitrification (i.e., the net increase or decrease in NH_4^+ or NO_3^- over a given period of time). However, researchers are increasingly recognizing that these measures provide limited information about N-cycling activity in the soil and are turning to stable isotope dilution techniques to measure gross rates of NH_4^+ and NO_3^- production (Booth et al., 2005; Davidson et al., 1991; Hart et al., 1994). In US ecosystems with more temperate climates than the Prairie Pothole region, lower slopes have been shown to have higher rates of both gross NH_4^+ mineralization and immobilization (Verchot et al., 2002) and lower rates of gross nitrification compared to upper slopes (Corre et al., 2002), emphasizing the significance of landform position in N-cycling processes. Research in the Pothole region of Saskatchewan has shown that lower slopes have higher SOC stores and lower N_2O emissions in uncultivated soils compared to cultivated ones, emphasizing the overlying effects of land use on landform patterns (Corre et al., 1999; Pennock, 2003).

Examining N-cycling processes in the context of land use will improve our ability to manage agricultural landscapes to minimize N losses, advancing sustainable agriculture while addressing concerns surrounding greenhouse gas emissions and global change. This study used stable ¹⁵N isotope techniques in cultivated and uncultivated ephemeral wetlands in central Saskatchewan to quantify gross mineralization and nitrification by land use. These measurements will be compared with the more ubiquitous measurements of soil inorganic N levels.

Methods

This project was carried out in the St. Denis National Wildlife Area (SDNWA) (Latitude: 52°12', Longitude: 106°5') of central Saskatchewan, Canada. The SDNWA is in the Dark Brown soil climatic zone and is dominated by hummocky terrain with loamy unsorted glacial till (Weyburn association) parent materials; slope classes range from 10 to 15% (Miller et al., 1985). Within the SDNWA, five ephemeral wetlands were selected: 3 uncultivated wetlands (2 were cultivated prior to 1968) and 2 cultivated wetlands. Ephemeral wetlands are those depressions in hummocky landscapes that contain standing water in the spring, but typically dry up during the growing season (Hayashi et al., 1998). The cultivated wetlands were direct-seeded to wheat on May 8, 2005. Granular fertilizer containing 72% 46-0-0 and 28% 11-52-0 was placed with seed at a rate of 135 kg ha⁻¹.

Each of the 5 wetlands was sampled three times throughout the 2005 growing season: May 4 (post-snowmelt), June 16 (early growing season), and July 26 (mid-growing season). Within each uncultivated wetland, sampling was focused in the riparian or willow ring soils, not the pond centers. For the cultivated wetlands, sampling was from the analogous concave footslope elements (Pennock et al., 1987). Three sub-sampling points were selected for each wetland ($5 \times 3 = 15$ points in total). On a given sampling date, five intact cores were taken per sub-sampling point: one for bulk density and gravimetric soil moisture measurements and four for the isotope dilution field incubation experiment. Cores (5-cm I.D.) were taken to a 15-cm depth, corresponding to the average thickness of the A horizon, using a slide hammer sampler equipped with a removable plastic liner (AMS Signature Series).

Of the four isotope-dilution cores, one pair was labeled with $(^{15}NH_4)_2SO_4$ (for gross NH₄ mineralization) and the other pair was labeled with $K^{15}NO_3$ (for gross nitrification); each solution contained 30 µg N ml⁻¹ at 98% ¹⁵N enrichment. Labeling was done using seven 2-ml injections per core with an 18-gauge side-port spinal needle after Davidson et al. (1991). Solution injections increased the gravimetric soil moisture by 7±2%. With the exception of those uncultivated cores with the lowest initial concentrations of soil NO₃⁻ (where NO₃⁻ injections as much as doubled the soil NO₃⁻ levels) adding inorganic N did not significantly increase the soil N levels relative to the inherent background variability observed within each wetland.

One core of each pair was broken up within 1 h of injection (t=0), sub-sampled and extracted using 2*M* KCl (on site). The other core was capped and buried on site for 24 h before extraction. The KCl extracts were analyzed for inorganic N concentration colorimetrically using a Technicon AutoAnalyzer (Technicon Industrial Systems, 1978) and analyzed for ¹⁵NH₄ and ¹⁵NO₃ using the modified diffusion procedure outlined in Bedard-Haughn et al. (2004). The differences in N concentration and ¹⁵N content between the t=0 and t=24 h samples were used to calculate gross mineralization and nitrification according to Hart et al. (1994). Turnover rate, the rate at which a given inorganic N pool replaces itself, was calculated as the NH₄⁺ pool size divided by the gross mineralization rate for NH₄⁺ turnover, and as the NO₃⁻ pool size divided by the gross nitrification rate for NO₃⁻ turnover.

Results

Soil NH_4^+ levels in the uncultivated wetlands (mg NH_4^+ -N kg⁻¹ soil) were significantly higher than cultivated wetlands for all experimental dates (Fig. 1). Over the course of the growing season, there was a small increase in the amount of NH_4^+ measured in the cultivated soils but no significant change in the uncultivated soils. This land use difference in soil NH_4^+ levels was not reflected in gross mineralization levels. Gross mineralization levels increased similarly for both land uses from May to July (Fig. 2). The NH_4^+ -N turnover time reflected the land use differences in soil NH_4^+ , with average turnover times of 1-2 days for smaller NH_4^+ pools in the cultivated soils and 2-5 days for the uncultivated soils. Turnover rates for both land uses tended to become more rapid through the season as both production and consumption rates increased (Table 1).

		Days to turnover (S.D.)	
		$\mathbf{NH_4}^+$	NO ₃
Uncultivated	May	2.8 (1.0)	0.5 (0.4)
	June	4.9 (4.1)	1.5 (1.2)
	July	2.7 (2.9)	1.2 (0.6)
Cultivated	May	1.5 (1.1)	3.4 (1.0)
	June	1.6 (1.3)	1.8 (1.0)
	July	0.6 (0.3)	1.7 (1.0)

Table 1. Turnover rates for NH_4^+ and NO_3^- pools in cultivated vs. uncultivated wetlands.



Figure 1. Soil NH₄⁺ levels in cultivated vs. uncultivated ephemeral wetlands.



Figure 2. Gross NH₄⁺ mineralization rates in cultivated vs. uncultivated ephemeral wetlands.

The land use effect for soil NO₃⁻ (mg NO₃⁻-N kg⁻¹ soil) was inverse that observed for soil NH₄⁺: cultivated soils had significantly greater soil NO₃⁻ than uncultivated soils (Fig. 3). Mean soil NO₃⁻ for cultivated soils was similar for May and June, but lower in July. Mean soil NO₃⁻ for uncultivated soils increased through the season, resulting in equal soil NO₃⁻ for both soils in July. There was no land use effect on gross nitrification levels and nitrification rates increased for both soils from May to July (Fig. 4). As was observed for NH₄⁺, soil NO₃⁻ turnover reflected soil NO₃⁻ levels, with more rapid turnover (≤ 1 d) occurring in the uncultivated soils with their lower soil NO₃⁻ levels (Table 1). However, unlike the seasonal decrease observed for the NH₄⁺ turnover, NO₃⁻ turnover became slower for the uncultivated soils (from 0.5 to 1.5 d) and more rapid for the cultivated soils (from 3.5 to 2 d) between the beginning and end of the growing season, reflecting soil NO₃⁻ levels over the same time period.



Figure 3. Soil NO₃⁻ levels in cultivated vs. uncultivated ephemeral wetlands.



Figure 4. Gross nitrification rates in cultivated vs. uncultivated ephemeral wetlands.

Discussion

The N-production and -consumption rates reported for these ephemeral wetlands were within the range of previously-reported values for agricultural and grassland soils (Booth et al., 2005). However, there were significant differences in the size of the NH_4^+ and NO_3^- pools between cultivated and uncultivated soils despite similar gross mineralization and nitrification rates, suggesting mechanistic N-cycling differences under the two ecosystems. From the similar gross N-production rates, we can infer that neither mineralization nor nitrification is substrate-limited in either soil. From the different soil inorganic-N levels, however, we can infer that the fate or cycling mechanism for the produced N differs between soils.

For example, the higher levels of soil NH_4^+ in uncultivated soils may indicate a greater occurrence of heterotrophic nitrification under the associated willow ring or riparian grassland vegetation. Uncultivated soils are more likely to have a significant fungal biomass and hence, a greater tendency towards heterotrophic nitrification whereby the biomass oxidizes both organic and inorganic substrate to produce NO_3^- (Paul and Clark, 1996). In contrast, the cultivated wetlands will be dominated by bacterial biomass and hence autotrophic nitrification will dominate. Therefore for both soils to produce the comparable amounts of NO_3^- measured in the gross nitrification experiment, nitrifiers in the cultivated soil would consume more NH_4^+ whereas those in the uncultivated soil would consume both NH_4^+ and organic N to meet their needs.

Similarly, differences in the size of the soil NO_3^- pool may be attributable to differences in NO_3^- consuming processes. There was significantly higher gross NO_3^- consumption in the uncultivated soils throughout the growing season and consumption rates increased at a rate

similar to gross nitrification rates, resulting in minimal increases in extractable soil NO_3^- through the season. Given the absence of plant uptake or leaching as a loss mechanism in the incubated cores, NO_3^- reduction is likely the primary pathway for uncultivated NO_3^- consumption. Based on ¹⁵N measurements of the NH_4^+ extracted from ¹⁵NO₃⁻ labeled cores in the uncultivated soils, there was a small amount of dissimilatory nitrate reduction to ammonium (DNRA) taking place, but the dominant NO_3^- consumption mechanism was likely reduction by denitrification. Previous authors have also noted that NO_3^- -N does not tend to accumulate under native grass vegetation (Malo et al., 2005).

Dividing the N pool size by its corresponding gross N-production rate gives an estimation of how rapidly that pool turns over or replaces itself. For these soils, the smaller the pool, the more rapidly it turned over. The NH_4^+ pool in the cultivated soils and the NO_3^- pool in the uncultivated soils both turned over in approximately 1 d, comparable to previously reported turnover rates in a range of soils (Booth et al., 2005). This rapid turnover further demonstrates the limited utility of a "snapshot" of extractable NH_4^+ and NO_3^- as indicators of either plant-available N for agricultural production or, in the case of NO_3^- , denitrification substrate for N_2O emissions.

There was a clear seasonal effect for gross N-producing and N-consuming processes. Booth et al. (2005) noted a strong positive relationship between mineralization, NH_4^+ assimilation and nitrification rates across a range of ecosystems. A positive relationship was also noted in our soils when all data were considered. However, when examined on a date-by-date basis, the relationship did not persist. In these soils, the correlation was apparently more a function of a similar response of the N-producing processes to the increase in soil temperature through the growing season (Bengtson et al., 2005), with rates of both processes increasing from May to July. Booth et al. (2005) noted that the temperature-nitrification relationship depends in part on the availability of NH₄⁺. Therefore, when gross mineralization increases with soil temperature, gross nitrification will also increase, as long as no other NH4⁺-assimilating or consuming process dominates. Schimel and Bennett (2004) suggest that N-producing, -consuming, and -emitting processes are most likely occurring in close proximity in soil microsites and are controlled in part by N availability. Nitrogen-producing and -consuming processes may be similarly activated by enzymes when the intracellular N content of associated microorganisms reaches critically low levels (Bengtson et al., 2005). Further investigation into the landform and land use impacts on these N-cycling feedback mechanisms is necessary.

Conclusion

Land use has had a significant impact on the soil NH₄⁺ and NO₃⁻ levels in these ephemeral wetlands but no influence on gross mineralization and nitrification rates. Instead, the primary effect of land use on N-cycling processes appears to be in the mechanistic details, where each land use favors a different suite of processes carried out by a specific group of microorganisms. The assemblage of dominant microbial players present under a given land use appears to be more important than other environmental factors previously assumed to be decisive, including fertilization and WFPS. The complexity of the interrelationships among the N-producing, N-consuming, and N-emitting processes highlights the need for greater knowledge of the structure and lability of the soil organic N pool and of defining the microbial communities responsible for N cycling under different land use regimes.

References

- Bedard-Haughn, A., Tate, K.W., van Kessel, C., 2004. Using ¹⁵N to quantify vegetative buffer effectiveness for sequestering N in runoff. Journal of Environmental Quality 33, 2252-2262.
- Bengtson, P., Falkengren-Grerup, U., Bengtsson, G., 2005. Relieving substrate limitation-soil moisture and temperature determine gross N transformation rates. Oikos 111, 81-90.
- Booth, M.S., Stark, J.M., Rastetter, E., 2005. Controls on nitrogen cycling in terrestrial ecosystems: A synthetic analysis of literature data. Ecological Monographs 75, 139-157.
- Burke, I.C., Lauenroth, W.K., Cunfer, G., Barrett, J.E., Mosier, A., Lowe, P., 2002. Nitrogen in the central grasslands region of the United States. Bioscience 52, 813-823.
- Corre, M.D., Pennock, D.J., Van Kessel, C., Elliott, D.K., 1999. Estimation of annual nitrous oxide emissions from a transitional grassland-forest region in Saskatchewan, Canada. Biogeochemistry 44, 29-49.
- Corre, M.D., Schnabel, R.R., Stout, W.L., 2002. Spatial and seasonal variation of gross nitrogen transformations and microbial biomass in a Northeastern US grassland. Soil Biology & Biochemistry 34, 445-457.
- Davidson, E.A., Hart, S.C., Shanks, C.A., Firestone, M.K., 1991. Measuring gross nitrogen mineralization, immobilization, and nitrification by N-15 isotopic pool dilution in intact soil cores. Journal of Soil Science 42, 335-349.
- Hart, S.C., Stark, J.M., Davidson, E.A., Firestone, M.K., 1994. Nitrogen mineralization, immobilization, and nitrification. In: R. Weaver (Editor), Methods of Soil Analysis, Part 2. Microbiological and Biochemical Properties. SSSA Book Series, No. 5. American Society of Agronomy, Madison, WI, pp. 985-1019.
- Hayashi, M., van der Kamp, G., Rudolph, D.L., 1998. Water and solute transfer between a prairie wetland and adjacent uplands, 1. Water balance. Journal of Hydrology 207, 42-55.
- Janzen, H.H., Desjardins, R.L., Asselin, J.M.R., Grace, B. (Editors), 1998. The Health of Our Air: Toward Sustainable Agriculture in Canada. Publication 1981/E Research Branch. Research Branch, Agriculture and Agri-Food Canada, Ottawa, Ontario, Canada, 98 pp.
- Malo, D.D., Schumacher, T.E., Doolittle, J.J., 2005. Long-term cultivation impacts on selected soil properties in the northern Great Plains. Soil & Tillage Research 81, 277-291.
- Miller, J.J., Acton, D.F., St. Arnaud, R.J., 1985. The effect of groundwater on soil formation in a morainal landscape in Saskatchewan. Canadian Journal of Soil Science 65, 293-307.
- Paul, E.A., Clark, F.E., 1996. Soil microbiology and biochemistry. Academic Press, San Diego, CA, USA.
- Pennock, D.J., 2003. Terrain attributes, landform segmentation, and soil redistribution. Soil & Tillage Research 69, 15-26.
- Pennock, D.J., Zebarth, B.J., Dejong, E., 1987. Landform classification and soil distribution in hummocky terrain, Saskatchewan, Canada. Geoderma 40, 297-315.
- Schimel, J.P., Bennett, J., 2004. Nitrogen mineralization: Challenges of a changing paradigm. Ecology 85, 591-602.
- Verchot, L.V., Groffman, P.M., Frank, D.A., 2002. Landscape versus ungulate control of gross mineralization and gross nitrification in semi-arid grasslands of Yellowstone National Park. Soil Biology & Biochemistry 34, 1691-1699.