fragrep: Efficient Search for Fragmented Patterns in Genomic Sequences

Axel Mosig, Katrin Sameith and Peter F. Stadler

Bioinformatics Group, Department of Computer Science, University of Leipzig, Kreuzstrasse 7b, Leipzig, D-04103, Germany and Interdisciplinary Center for Bioinformatics, University of Leipzig, Kreuzstrasse 7b, Leipzig, D-04103, Germany

ABSTRACT

Summary: Many classes of non-coding RNAs (including yRNAs, vaultRNAs, RNAse P and MRP RNA, as well as a novel class recently discovered in *Dictyostelium discoideum*) can be characterized by a pattern of short but well conserved sequence elements that are separated by a poorly conserved regions of sometimes highly variable lenght. Local alignment algorithms such as blast are therefore ill-suited suitable for the discovery new homologs of such ncRNAs in genomic sequences. The fragrep tool instead implements an efficient algorithm for detecting pattern fragments that occur in a given order. For each pattern a mismatches tolerance and bounds on the length of the intervening sequences can be specified separately for each pattern fragment.

Availability: The program fragrep can be downloaded from http://www.bioinf.uni-leipzig.de/Software/fragrep/.

Contact: Axel Mosig, Tel: ++49 341 14951 31, Fax: ++49 341 14951 19, axel@bioinf.uni-leipzig.de

Methods for detecting non-coding RNAs (ncRNAs) in genomic sequence data has been a topic of intense research. While techniques for detecting protein-coding genes can rely on universal characteristics such as start and stop codons, the triplet amino acid code or ribosome binding sites, there are no corresponding characteristics known in ncRNAs. Computational tools for ncRNAs detection are therefore restricted to one or few particular classes of RNA. Some classes, such as yRNAs and vaultR-NAs, contain stem or loop regions with well-conserved sequence patterns (Farris et al., 1999; Teunissen et al., 2000; Kickhoefer et al., 2003)). These characteristics are used by fragrep.

Supppose that our ncRNA of interest contains k conserved sequence fragments, denoted by C_1,\ldots,C_k , which occur in a given order in a set of known examples. In practice, the C_i are obtained as the consensus sequences of conserved blocks in a multiple alignment. Scanning a genome T for these blocks, we expect to find a non-conserved sequence segment X_i between any two fragments C_i and C_{i+1} . The fragrep solves he

problem of determining whether there are sequences X_1, \ldots, X_{k-1} such that $C_1 X_1 C_2 X_2 \ldots X_{k-1} C_k$ is a substring of T. Additionally, fragrep can take into account two further aspects:

Gap length bounds: For each X_i , the user can specify lower and upper bounds, denoted by ℓ_i and u_i , respectively, for the length of X_i , so that only matches satisfying $\ell_i \leq |X_i| \leq u_i$ will be taken into account by fragrep. Mismatches: The fragments C_i do not need to match the corresponding sequence part of T exactly; the user can specify a number of mismatches m_i . Denoting C_i' as the fragment C_i modified by at most m_i many arbitrary mismatches, fragrep will report occurences of some $C_1'X_1C_2'X_2\ldots X_{k-1}C_k'$ as well.

We start by computing all occurences of the most informative fragment C_a , i.e., the fragment that is least likely to occur as a random subsequence of the genome T. Then a neighborhood defined by the bounds ℓ_i on the lengths of the intervening sequences X_i is searched for the other fragments C_i , $i \neq a$. From this position information a graph G is constructed such that paths of length k in G correspond to occurrences of C_1, \ldots, C_k in the given order under the specified mismatch and gap length constraints. These paths can be found easily by means of dynamic programming. Starting with the most informative sequence C_a rather than C_1 increases the efficiency of the search and in practice leads to a significant speedup, in particular when short or ambiguous fragments are part of the pattern. The C implementation of fragrep has been optimized in several algorithmic details to improve the runtime.

We used fragrep to studying the evolution of a class of ncRNAs in the the slime mold *Dictyostelium discoideum* that was discovered in an experimental survey by Aspegren *et al.* (2004). We searched the genomic sequence (Fey *et al.*, 2004) for *type-I ncRNAs* using the following simple pattern:

0 0 GTTGRCCTTACAGCAA 2 0 120 GTCAACTG 2

The first two columns contain the minimal and maximal

distance before a the pattern fragment (always 0 for the first fragment, of course), the last column is the maximal number of mismatches that is tolerated in each fragment. We recovered 45 candidates of which 34 are sufficiently similar to the experimentally determined sequences to be alignable. 11 very divergent sequences were not included in the further analysis. A neighborjoining tree summarizing both known sequences and the novel candidates detected by fragrep is displayed as Fig. 1. We find the the class-I ncRNAs are located in small clusters in all 6 chromosomes. Interestingly, there are two subclasses, denoted by A and B, that alternate in the larger clusters, even though their direction on the chromosomes does not seem to follow a simple rule.

In order to evaluate the performance of the algorthm underlying fragrep, we used a query derived from yRNA alignments in Farris *et al.* (1999) to scan the whole human genome. The query consisted of four fragments whose length varide between 5 and 17 nucleotides. Scanning all chromosomes of the human genome took less than 20 minutes on a standard desktop computer with a 2.4GHz processor and 1GB main memory; further results from scanning the human as well as the mouse and the rat genome are given in the folowing table.

Genome	H.sap.	M.musc.	R.nov.
size in Mb	2,980	2,561	2,640
runtime (mm:ss)	19:43	15:44	15:48
# cand. matches	44	1	1

These examples demonstrate that fragrep can be used for systematic surveys of eukaryotic genomes. The application of standard multiple alignment tools such as ClustalW or dialign to a relatively small set of representatives of an ncRNA class can be used to determine conserved sequence patterns, which can be turned into fragrep queries in a straightforward manner. The fragrep tool can then be employed to find additional members of the ncRNA family in related genomes. This approach yields significant matches where other sequence search tools such as blast fail to report useful results, while structure based approaches, such as infernal are too costly. Of course, fragrep is not limited to ncRNA detection; the search for specific constellations of transcription factor binding sites is another potential application.

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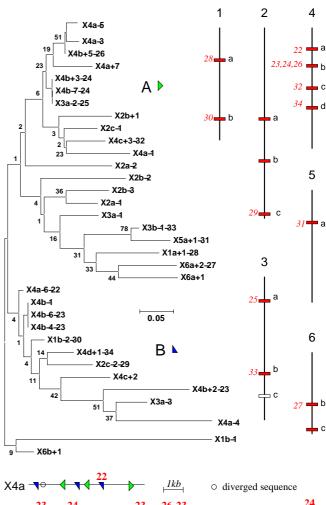




Fig. 1. Type-I ncRNAs from *Dictyostelium discoideum*. Red numbers are the DdR- numbers of the expressed RNAs from the experimental survey by Aspegren *et al.* (2004). The sequences appear in clusters on all chromosomes (right). The phylogenetic tree (left, neighborjoining method) suggests that there are two major subgroups, labeled A and B. Below the organization of the two largest clusters X4a and X4b located at chromosome 4. Note that type A and type B copies alternate. The other type-I ncRNA clusters consist of not more than three sequences.

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