

RESEARCH ARTICLE

Open Access

Ubiquitous presence of piscidin-1 in Atlantic cod as evidenced by immunolocalisation

Jareeporn Ruangrsri¹, Jorge M O Fernandes¹, Jan H W M Rombout^{1,2}, Monica F Brinchmann¹ and Viswanath Kiron^{1*}

Abstract

Background: Antimicrobial peptides (AMPs), the natural antibiotics bestowed upon all forms of life, consist of small molecular weight proteins with a broad spectrum antimicrobial activity against a variety of pathogenic microorganisms. Piscidins are one of the AMP families that are imperative for the innate defence mechanisms of teleosts. Atlantic cod, a basal fish belonging to the superorder Paracanthopterygii also possesses multiple piscidin peptides. Two piscidin paralogues (*pis1* and *pis2*) and a novel alternative splice variant of *pis2* of this fish were previously described by us. To shed light on other potent roles of these molecules, now we have mapped the distribution of piscidin 1 (Pis1), in different tissues and organs of cod through immunohistochemistry (IHC) employing an affinity purified polyclonal antibody specific to Pis1.

Results: Various cell types and tissues of Atlantic cod including those from the immune organs of naïve fish are armed with Pis1 peptide. Different types of the blood leucocytes and phagocytic cells among the leucocytes examined gave a relatively strong indication of Pis1 immunopositivity. In addition, other cell types such as hematopoietic cells, epithelial cells and multi-granular cells located in the mucosal and hematopoietic tissues were also Pis1-immunoreactive. More interestingly, chondrocytes appear to produce Pis1 and this is the first report on the presence of an AMP in cartilage tissue of fish. Furthermore, Pis1 immunopositivity was detected in other tissues and organs of naïve fish including neural tissues, exocrine and endocrine glands, compound gland cells, excretory kidney, intestinal and respiratory epithelial cells, swim bladder, skin and hypodermis layer, myosepta, liver, heart, eye and oocytes.

Conclusions: Pis1 peptide is produced by various cell types located in different tissues and organs of Atlantic cod. It is present in all immune-related organs of naïve fish and the elevated peptide expression following phagocytosis strongly suggest their involvement in innate defence. Further, its widespread occurrence in non-immune tissues and organs of apparently healthy fish implies that piscidin may have other functions in addition to its role as an immune effector molecule.

Keywords: Piscidin, Antimicrobial peptide, Innate immunity, Multi-functionality, *Gadus morhua*

Background

Atlantic cod (*Gadus morhua* L.) is a demersal fish that is widely distributed in the North Atlantic region, the Baltic Sea and the Barents Sea. Commercial production of this fish has been undertaken mainly by Norway, though fraught with several challenges. There has been great interest in understanding the immune system of this fish species. It has been confirmed recently that cod has a unique immune architecture compared to other vertebrates as they

are devoid of genes for major histocompatibility complex (MHC) II, cluster of differentiation 4 (CD4) and invariant chain (Ii) [1]; all of them are attributed to a normal functioning of adaptive immunity. Earlier studies [2-4] that examined the antibody responses of Atlantic cod have revealed that cod relies more on innate than adaptive defence mechanisms. On the other hand, cod exhibits an incredible ability to defend itself against pathogens [5]. The effective functioning of the innate immune system could be due to the presence of a number of MHC I loci and the unique organization of Toll-like receptor (TLR) families in the genome [1,6]. Furthermore, our contribution to the knowledge on the innate immune components of Atlantic

* Correspondence: kiron.viswanath@uin.no

¹Faculty of Biosciences and Aquaculture, University of Nordland, 8049, Bodø, Norway

Full list of author information is available at the end of the article

cod is that several tissues of the fish are armed with a battery of peptides with antimicrobial activity [7].

Antimicrobial peptides (AMPs), the natural antibiotics bestowed upon all forms of life, consist of small molecular weight proteins with a broad spectrum antimicrobial activity against a variety of pathogenic microorganisms [8]. Several fish AMPs, are described as essential innate defence molecules [9,10]. Piscidins are one of the most potent AMPs found in both freshwater and marine teleosts [11-17] and their antimicrobial properties enable them to inhibit the growth of bacteria, fungi, viruses and parasites [14,18-21]. Immunohistochemical studies have shown that various cell types in different tissues and organs, particularly the interface tissues that are in constant interaction with the environment (e.g. gills, skin, alimentary tract) and the hematopoietic tissues are involved in the production of piscidin peptides [13,14,22,23].

Recently two piscidin paralogues (*pis1* and *pis2*) and a novel alternative splice variant of piscidin (*pis2b*) [17,24] were described for cod. Moreover, we have reported that *pis* genes of Atlantic cod have undergone structural diversifications through positive selection [17]. Our additional studies have shed light on the variation in their gene expressions in different tissues of adult fish and during developmental stages and on the broad antibacterial properties of the synthetic peptides of Atlantic cod piscidin [24]. To further understand the potent role of piscidin peptides in Atlantic cod, immunohistochemistry (IHC) was used to identify tissue and cell distribution of *Pis1*, using an anti-*Pis1* antibody.

Methods

Anti-*Pis1* antibody

Affinity-purified rabbit polyclonal anti-*Pis1* antibody raised against the whole mature peptide sequence of Atlantic cod *Pis1*, prepared on demand (GenScript, New Jersey, USA) was used in the present study. The peptide antigen corresponding to C-FIHIIIGWISHGVRAIHR AIHG was used later for polyclonal antibody production following the manufacturer's procedures (GenScript). Briefly, synthetic peptide was conjugated to keyhole limpet hemocyanin (KLH) and then injected into rabbits. The antiserum was then affinity purified by running over a column having the 23-mer *Pis1* fragment conjugated to cyanogen bromide-activated agarose as an immunosorbent. The resulting titer of the affinity-purified antibody was 1:64,000, confirmed by ELISA. The peptide-specific antibody had less than 1 % cross-reactivity in the ELISA, where 1 % cross-reactivity is 100 times more antibody than is required to produce the same optical density with either free KLH, conjugated KLH, or free peptide that shares less than three amino acids in the sequence (according to GenScript). Furthermore, through Western

blot analysis it was determined that anti-*Pis1* antibody did not react with either synthetic *Pis2* or *Pis2b* peptide (1 µg of *Pis2*; FLHHIVGLIHHGLSLFGDRAD or *Pis2b*; FLHHIVGLIHHGKLDMYRSNN) of Atlantic cod, while its immunoreactivity was strong with 1 µg of *Pis1* peptide (Additional file 1: Figure S1).

Fish and their maintenance

Apparently healthy hatchery produced Atlantic cod were maintained in the indoor facilities at the research station of University of Nordland, Bodø, Norway. The fish (200-300 g) selected for the study were from among those reared at 7-8°C in 2800 l fiberglass tanks and fed daily on a commercial feed (Amber Neptun; Skretting AS, Norway). All animal handling protocols were approved by the National Animal Research Authority (Forsøksdyruttvalget, FDU; id numbers: 2453 and 3207) in Norway.

Leucocyte preparation and fixation

Experimental fish were anesthetized using a sub-lethal dose (100 mg·l⁻¹) of tricaine methanesulphonate (MS-222, Argent Chemical Laboratories, Washington, USA) and immediately euthanised. Blood was collected from the caudal vein using a heparinised (Heparin 150 Unit·ml⁻¹, Sigma, Missouri, USA) syringe. Following aseptic procedures, freshly dissected head kidney and spleen were put in L-15 medium (Leibovitz, Sigma) containing 2 % fetal bovine serum (Sigma) and 100 U·ml⁻¹ heparin. Subsequently, the tissues were placed on a 100 µm filter and teased gently with a sterile syringe plunger to prepare cell suspensions in the medium contained in a Petri dish. The cells collected were transferred to a 15 ml sterile tube. In order to achieve a good quality harvest of leucocytes from the peripheral blood, head kidney or spleen cells, their cell suspensions were centrifuged at 1000g for 5 min at 4°C. Thereafter the buffy coat of leucocytes was put into a new 1.5 ml sterile tube and was resuspended in L-15 medium to get an approximate concentration of 10⁶ cells·ml⁻¹.

An aliquot of the blood sample was applied onto a poly-L-lysine coated glass slide (VWR International BVBA, Leuven, Belgium), smeared and air dried for about 2 h. The slides were then fixed in 100 % grade methanol for few minutes, air dried and kept at 4°C until use. The remaining blood, head kidney or spleen leucocytes were mixed with latex beads (Fluoresbrite[®] YG Carboxylate Microspheres 1.0 µm, Polysciences Europe GmbH, Germany) at a concentration of 5 × 10⁷ beads·ml⁻¹ cells, to get an approximate concentration of 50 beads per cell. They were incubated at 14°C for 4 h, after which each sample was applied onto a poly-L-lysine coated glass slide, smeared, dried and preserved using the same procedure as mentioned above.

Tissue fixation and paraffin section

Different tissues and organs - dorsal skin, gill, peritoneal tissue, head kidney, trunk kidney, spleen, swim bladder, liver, pyloric caeca, intestine, rectum, heart and ovary - were sampled and fixed overnight in 4 % paraformaldehyde prepared in phosphate buffer saline (PBS, pH 7.4) treated with 0.1 % diethylpyrocarbonate (Sigma). Standard histological procedures were adopted to process these samples and embed them in paraffin (Paraplast® Tissue Embedding Media, Pennsylvania, USA). Later the paraffin-blocked tissues were used to prepare 4-5 µm sections employing a microtome (Shandon Finesse, Thermo Scientific, Barrington, USA) and these sections were then mounted on poly-L-lysine coated slides. The slides were subsequently incubated overnight at 50°C and kept at room temperature until they were used in IHC analysis.

Specificity of the anti-Pis1 antibody

The specificity of the anti-Pis1 antibody was confirmed by Western blot analysis using synthetic piscidins and tissue extracts as test samples, following the procedure reported by Corrales et al. [23]. In tissue sections, the antibody specificity was determined by incubating the serial section samples from the fry stage of cod in different preparations of primary antibody. These preparations were: (i) untreated anti-Pis1 antibody (the positive assay), (ii) antibody pre-incubated with the cod Pis1 peptide, (iii) dilution buffer (1.5 % bovine serum albumin (BSA) in 0.1 M PBS) alone, (iv) antibody pre-incubated with a related peptide (cod Pis2) or (v) a non-related peptide (cecropin P1) to check cross-reactivity or (vi) an anti-cod galectin antibody, to detect false positive reactions in some test tissues and organs. The pre-incubation of antibody with each peptide was performed following the procedure reported by Sompuram et al. [25]. Briefly, undiluted anti-Pis1 antibody (727 µg·ml⁻¹, according to GenScript) was mixed with an equal volume of each peptide solution, maintaining a ratio of 1:2 for the concentration of the antibody and the peptide, and incubated at 37°C for 45 min. The pre-absorbed antibody was later diluted in dilution buffer to make the final concentration of ~14.5 µg·ml⁻¹ before being used for IHC.

Immunohistochemistry

IHC was performed on both smear and section samples, using procedures of Mulero et al. [22], but with some modifications. Briefly, paraffin sections prepared as described under tissue fixation were dewaxed with xylene followed by rehydration through decreasing gradients of ethyl alcohol. In the standardization steps of IHC analyses, two different antigen retrieval (AR) solutions, which are widely used for antigen retrieval in IHC studies [26], were evaluated - citrate buffer (10 mM sodium citrate, 0.05 %

Tween 20, pH 6.0) and Tris-EDTA buffer (10 mM Tris-Base, 1 mM EDTA solution, pH ~9), at high (autoclave at 100°C, 10 min) and low (water bath at 65°C, 1 h) temperatures. Immunostaining of Pis1 in the tissue sections retrieved with citrate buffer at both low and high temperatures was faint, while the sections retrieved with Tris-EDTA buffer gave stronger Pis1 immunoreactivity and a desirable light background at high temperature. The last set of conditions was chosen for the IHC studies. The antigenic epitope of rehydrated specimens were retrieved by immersing slides in a staining dish containing Tris-EDTA buffer. The dish with the slides was autoclaved at 100°C for 10 min and subsequently sections were kept at room temperature for 20 min.

The tissue sections and all the leucocyte samples referred to previously were washed thrice for 5 min each time with de-ionized water. They were later incubated with 3 % hydrogen peroxide in 100 % methanol for 10 min at room temperature to quench endogenous peroxidase activity. This step was followed by washing each slide three times with washing buffer (1.5 % Tween 20 in 0.1 M PBS) for 10 min and incubating the slides in blocking solution (5 % BSA in 0.1 M PBS) for 1 h at room temperature. After removing the blocking solution the specimens were incubated overnight at 4°C with primary antibody at a dilution of 1:50 (~14.5 µg·ml⁻¹, the best titer that gave good immunostaining and light background from among the different dilutions tested: 1:25, 1:50, 1:100, 1:400 and 1:800) with the dilution buffer. Sections were also incubated with a 1:50 dilution of Pis-1 pre-incubated antibody or with the dilution buffer alone (omitting primary antibody). After rinsing for 10 min with washing buffer (3 times), sections were incubated for 30 min with HRP conjugated secondary anti-rabbit antibody (Santa Cruz Biotechnology, California, USA) at 1:800 (the best titer which produced good immunostaining and light background from among those tested: 1:400, 1:800, 1:1200 and 1:2000) dilution in buffer. Following another three-step washing cycle as mentioned earlier with the buffer, the presence of the primary antibody bound to the tissue antigen was detected using liquid-stable 3,3'-diaminobenzidine tetrahydrochloride (DAB, Sigma) solution as a dye substrate for alkaline phosphatase. The desired signal level was achieved after 3–10 min of incubation. Sections were then slightly counterstained in hematoxylin (MerckKgaA, Darmstadt, Germany), dehydrated and mounted with Eukitt mounting medium (Eukitt O. Kindler GmbH, Freiburg, Germany). The slides were observed under a binocular microscope equipped with a Leica camera (Leica Microsystem, Germany).

Results

Anti-Pis1 antibody is specific for Atlantic cod Pis1 peptide
Western blot analysis indicated that Pis1-antibody was immunoreactive only with Pis1 and with peptides that have

molecular weight similar to that of Pis1 (Additional file 1: Figure S1). In tissue sections, we observed the positive reaction to Pis1 in various cells and tissues treated with anti-Pis1 antibody (Figure 1a and c). Pis1-immunoreactivity was also seen in serial sections and cells treated with antibody pre-incubated with either the most relevant peptide (Pis2) or the irrelevant peptide (cecropin P1; Figure not presented). On the other hand, Pis1-immunoreactivity was absent in all the control samples treated either with the antibody pre-incubated with Pis1-specific peptide or when the dilution buffer alone was used (Figure 1b and d). For all the IHC analyses that followed, negative control samples were set up by treating sections with the dilution buffer alone. In the test sample treated with the anti-cod galectin antibody, immunopositivity was found to vary depending on cells and tissues. For instance, no positive chondrocytes were seen in all cartilaginous tissues, while most of the cells in the gill epithelium were strongly immunostained compared to the anti-Pis1 antibody treated sample (Figure 1e).

Pis1 is produced by leucocytes, phagocytic cells and hematopoietic tissues

Treating the samples with anti-Pis1 antibody revealed the various cell types residing in different tissues and organs of cod that are involved in Pis1 peptide production. Some immunopositive signals were also found in regions that form the cellular network of some of the tested tissues.

Among the leucocytes isolated from the peripheral blood an oval thrombocyte-like cell, a round cell and a large cell with granular and lobed nuclei were stained positive for the anti-Pis1 antibody (Figure 2a). IHC analyses of leucocytes pre-incubated with latex beads revealed that phagocytes were able to produce Pis1. Some of the leucocytes that had phagocytosed beads were Pis1-immunoreactive (Figure 2b). Immunoreactive phagocytes were abundant in head kidney, and few in spleen cell suspensions (Figure 2c, d). Furthermore, among the head kidney leucocytes, the cells with high bead uptake exhibited a stronger Pis1-immunoreactivity than those with a lower bead uptake (Figure 2c).

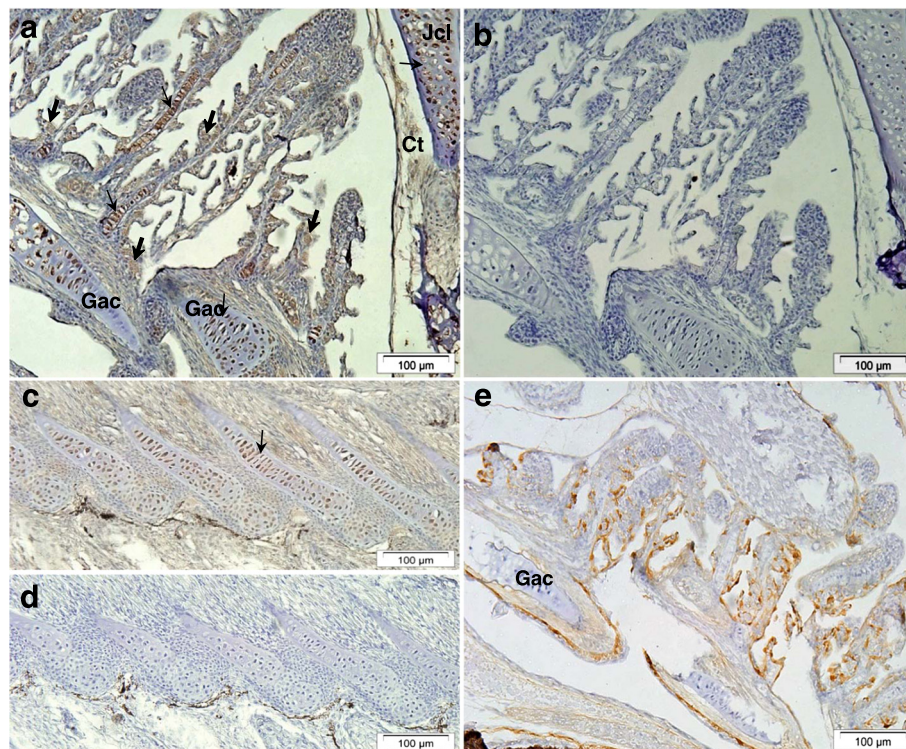


Figure 1 Immunohistochemistry performed on paraffin sections of healthy Atlantic cod fry. (a) Gill section treated with anti-Pis1 antibody showing immunopositive multi-granular cells (thick arrows) of the gills and a number of chondrocytes (thin arrow) in cartilaginous tissues of gill filament, gill arch (Gac), jaw cartilage (Jcl), and a weak positive reaction for the connective tissue (Ct). (b) Control serial section of (a) treated with Pis1 pre-incubated antibody indicate that all cells and tissues were not immunoreactive. (c) Radial cartilage of anal fin, chondrocytes (arrow) and connective tissue of muscle fibers (*) were Pis1-immunoreactive; (d) control serial section of (c) treated with Pis1 pre-incubated antibody. (e) Gill section treated with anti-cod galectin antibody showing immunoreactive cells scattered throughout the epithelial tissues, while all the chondrocytes were not immunostained.

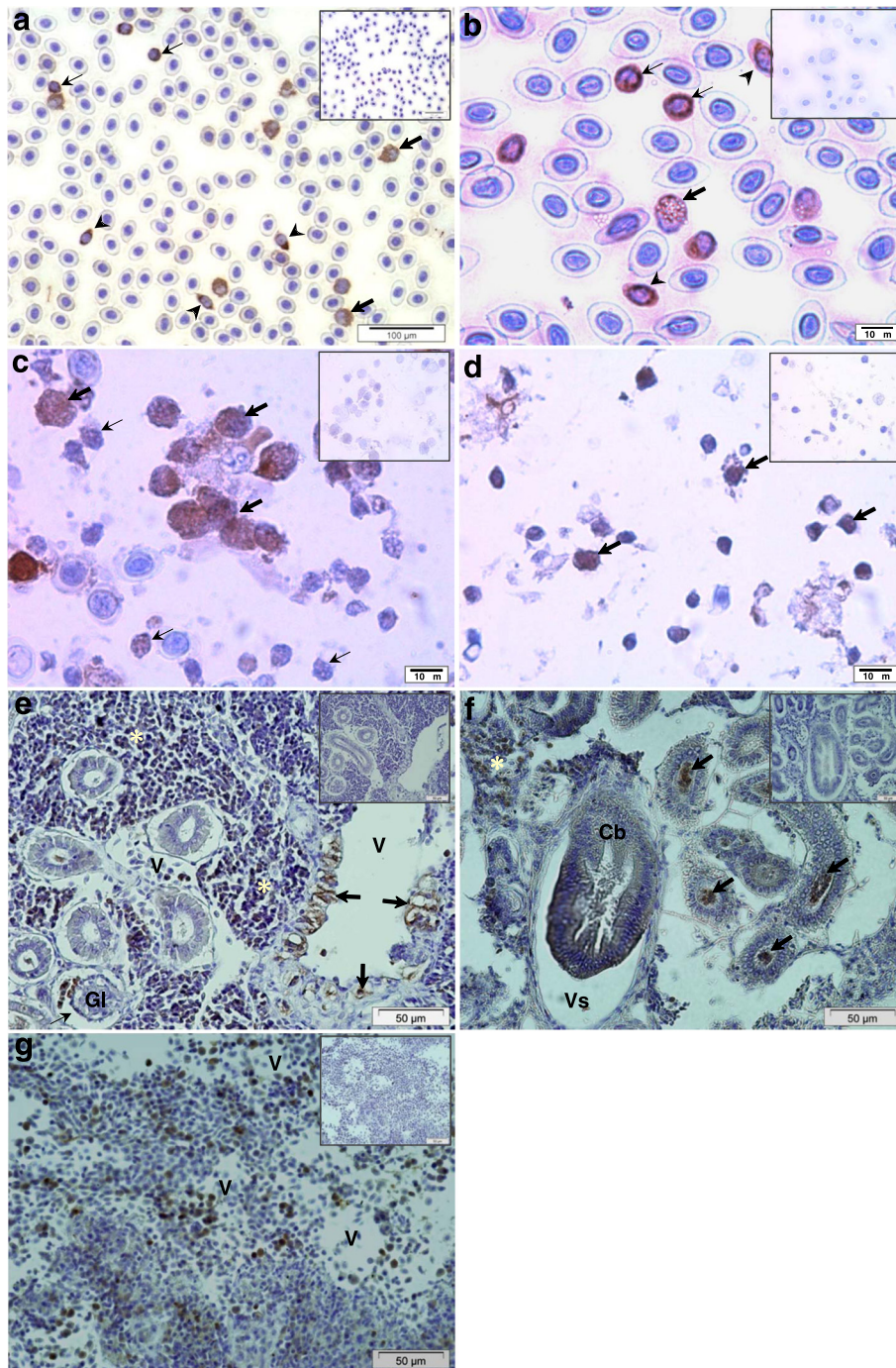


Figure 2 Immunohistochemistry on Atlantic cod leucocytes and different hematopoietic tissues. (a) The cytoplasm of different leucocyte types, including round cells (thin arrows), large cells with granular and lobed nuclei (thick arrows) and thrombocyte-like cells (arrow heads) were Pis1-immunoreactive (brown colour). Leucocytes incubated with latex beads (b, c and d). (b) Blood: some bead-containing phagocytes (thick arrows), round cells (thin arrows) and thrombocyte-like cells (arrow heads) were Pis1-immunoreactive. (c) Head kidney: numerous leucocytes, including phagocytes were Pis1-immunoreactive, phagocytes with intense staining had high bead uptake (thick arrows) than those with a low bead uptake (thin arrows). (d) Spleen: most of the leucocytes including phagocytes (arrows) were Pis1-immunoreactive. (e) Head kidney: numerous vascular tissue (arrows) and some cells in the hematopoietic tissues (*), glomerulus (Gl) and blood vessel (V) were Pis1-immunoreactive; note that glomerulus is surrounded by Bowman's capsule space (unstained - thin arrow). (f) Trunk kidney: cells in hematopoietic tissue (*), cytoplasm of columnar epithelial cells of collecting tubule (Cb), cells and detached elements in the lumen of renal tubules (arrows) were Pis1-immunoreactive. (g) Spleen: numerous hematopoietic cells were Pis1-immunoreactive (v: blood vessel). Negative control (inset) of the cell samples and serial sections treated with dilution buffer was Pis1-immunonegative (Larger images are seen in Additional file 1: Figure S2).

In the head kidney sections, immunopositive signals were prevalent in the hematopoietic tissue, and also in the cells of glomerulus and adjacent vascular tissues (Figure 2e). In trunk kidney many Pis1-immunoreactive cells were also observed in the hematopoietic tissue, and in the cytoplasm of the columnar epithelial cells of collecting tubules and in cell debris located in the lumen of renal tubules (Figure 2f). In the spleen, numerous hematopoietic cells were also Pis1-immunoreactive, particularly those residing close to the blood vessels (Figure 2g).

Pis1 is produced by mucosal tissues

In the dorsal skin sections, Pis1-immunoreactivity was observed in the epithelial and basal cells of the epidermal layer (Figure 3a). Some skin epidermal tissues, collected from the vicinity of eyes, were also found to be immunopositive (Figure 3b).

In the gill section, respiratory epithelial cells and multi-granular cells of primary and secondary lamellae were Pis1-immunoreactive (Figure 3c). Some multi-granular cells of the adductor muscle were also positive (Figure 3d). Interestingly, Pis1-immunoreactivity of chondrocytes was strong in the cartilaginous tissues of both filaments and gill arches (Figure 3e-f).

In the gastrointestinal tract, Pis1-immunoreactivity was weak in the cytoplasm of epithelial cells of the caecal mucosal fold, and in some cells and connective tissues surrounding the caecae (Figure 4a). In the intestine, epithelial columnar cells, multi-granular cells in the lamina propria and submucosal tissues, and cytoplasm of columnar cells in compound glands [27] were immunostained, but goblet cells were Pis1-immunonegative (Figure 4b-d).

Mucosal epithelial cells of the swim bladder were only slightly Pis1-immunostained, while the intensity of staining was stronger in numerous neural cells of the swim bladder wall (Figure 4e). In addition, a strong positive reaction was observed in the fibroblasts, the cellular network of submucosal tissue and in the secretory cells of the gas gland, while the signal was weak in the rete mirabile (Figure 4f-g).

Pis1 is produced by neuronal cells, exocrine and endocrine glands

The presence of Pis1 peptide was evident in the cluster of nerve cells found in the swim bladder wall as well as in the nervous system of some other organs. A large cluster of immunopositive perikarya was located on the surface of the head kidney in the parasympathetic ganglia (Figure 5A-a). In the ocular region, cartilaginous tissue, and various cells and tissues of retina - photoreceptor cells, inner nuclear layer, and nerves and nerve tissues - showed Pis1-immunoreaction (Figure 5A-b). In addition, some positive cells were also found in the choroid rete mirabile (not shown).

Examination of the sections of intestine and kidney revealed that exocrine and endocrine glands are also involved in the production of Pis1 peptide. In pancreas, most of the exocrine acinar cells exhibited strong Pis1-immunoreactivity (Figure 5B-a). In corpuscles of Staninus located on the surface tissue of both head and trunk kidneys, Pis1-immunoreactive secretory cells were found (Figure 5B-b).

Pis1-immunoreactivity in other tissues and organs

Besides those mentioned above, some other tissues and organs were Pis1-immunoreactive. In the liver most of the hepatocytes and their cellular network were Pis1-immunostained (Figure 6a). Furthermore, the presence of this molecule was evident in the epithelial cells of gall bladder and bile ducts (Figure 6b-c). In heart, immunopositive fibroblast cells were observed in the bulbus arteriosus, while such a signal was seen neither in other parts of the heart tissue, nor in the control sample (Figure 6d).

In the peritoneum, Pis1-positivity was found in the connective tissues lining a layer of mesothelium (Figure 6e) and in cells and connective tissue of the skin hypodermal layer and around the muscle fibers (myosepta) (Figure 6f). Pis1-immunoreactivity was also noted in many stages of the oocytes; the intensity of immunostaining was stronger in the early or mid vitellogenic oocyte (Figure 6g). In addition to the gill cartilage chondrocytes mentioned earlier, other cartilaginous tissues in the radial cartilage of dorsal and anal fins, hyaline cartilage of the jaws and eyes were also Pis1-immunoreactive (see Figure 1a, c and Figure 5A-b). Further, the notochord of cod larvae was found to be immunoreactive as well (not shown).

Discussion

Distribution and localisation of Pis1 peptide, a natural antibiotic, was studied in different tissues of Atlantic cod using immunohistochemistry. The specificity of the antibody was confirmed in the serial section samples, by employing anti-Pis1 antibody that was pre-incubated with either synthetic Pis1 peptide or other non-related peptides. Through Western blot analysis we have shown that the anti-Pis1 antibody of Atlantic cod does not cross react with Pis2 or Pis2b of the same species. On the other hand, similar immunostaining patterns were observed in sections treated either with the anti-Pis1 antibody or pre-absorbed antibody with Pis2 peptide. The lack of cross-reactivity between anti-Pis1 antibody and Pis2 peptide could be attributed to their structural diversifications and low identity (36 % or 41 %) of Pis1 and Pis2 or Pis2b [24].

The specific antibody was used to further investigate the distribution of Pis1 peptide in fish tissues. There was sound cellular evidence on localization of Pis1 in some of the tissues, corresponding to the observations on *pis1* gene expression reported previously [24]. Various cell

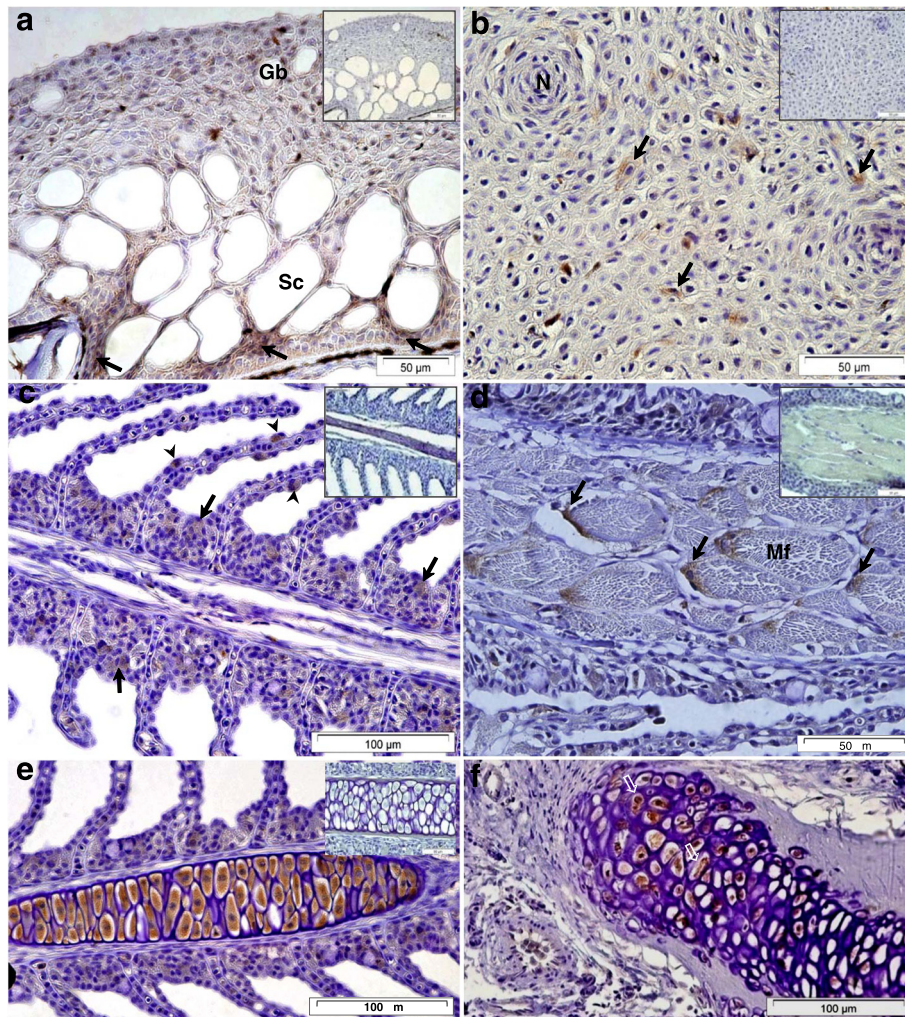


Figure 3 Immunohistochemical micrographs of different tissues of Atlantic cod. (a) Dorsal skin: numerous epithelial cells were Pis1-immunoreactive, but the basal cells (arrows) exhibited stronger immunoreactivity than the apical cells (Gb: goblet cell; Sc: sacular cell). (b) Epidermal layer of skin surrounding the eye - some Pis1-immunoreactivity was observed in the cytoplasm of epithelial cells (arrows), (N: cells of neuromast). (c) Gill filament: multi-granular cells of primary lamella (arrows) and epithelial cells of secondary lamella (arrow heads) were Pis1-immunoreactive. (d) Adductor muscle: multi-granular cells adjacent to the muscle fibers (Mf) were Pis1-immunostained. (e) Cartilaginous tissue of the primary lamella: all the chondrocytes (*) exhibited very strong Pis1-immunoreactivity. (f) Cartilaginous tissue of the gill filament: only chondrocytes (open arrows) were Pis1-immunostained (positive). Negative control (inset) of the samples treated in dilution buffer was Pis1-immunonegative (Larger images are seen in Additional file 1: Figure S3).

types, such as thrombocyte-like cells [28], multi-granular cells [29], hematopoietic cells [22] and epithelial cells from mucosal tissues [30] reacted positive to Pis1 antibody. This indicates that the molecule is probably associated with an immune function as these cells and tissues in fish are known to be involved in a dynamic immune defence network [9,29-34]. Furthermore, our observations strongly suggest that cod Pis1 activates the elimination of foreign bodies, via different mechanisms. An intracellular killing mechanism is clearly reflected through the presence of Pis1 in phagocytic cells (Figure 2). In addition, the strongly immunostained phagocytes with

high bead uptake indicate that Pis1 peptide is a component of the host defence mechanism during phagocytosis. The extracellular killing mechanism, on the other hand, could be supported by the detection of peptide outside the cells of various immune organs. Antimicrobial piscidins, which are part of the defence system in many fish species, are not produced by mucosal tissues alone, but also by cells residing in the spleen and head kidney [13,22,35]. Moreover, Mulero et al. [22] have demonstrated the ability of piscidins to eliminate invading bacteria through both intracellular and extracellular killing mechanisms.

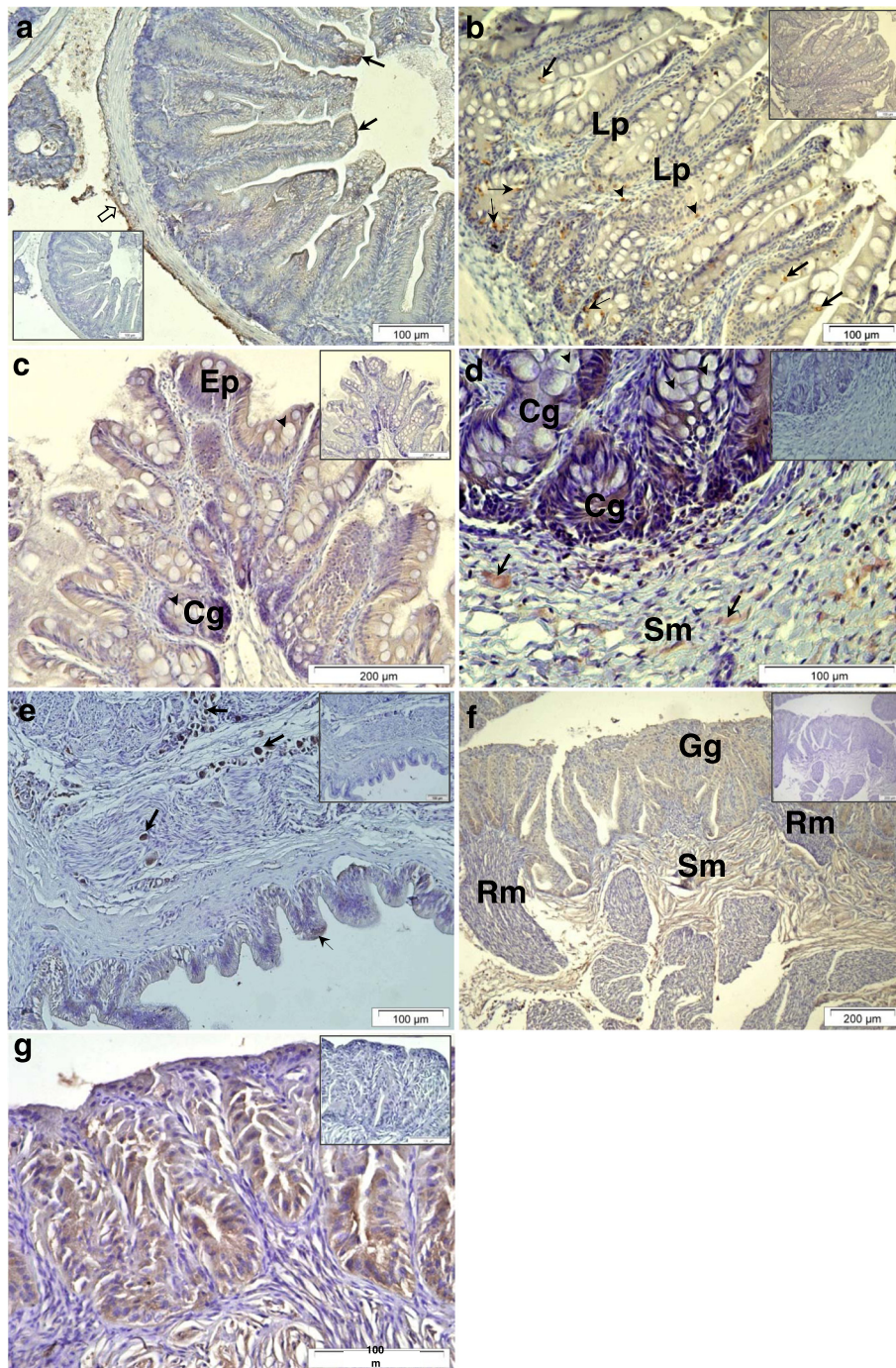


Figure 4 Immunohistochemical micrographs of the gastrointestinal tract and different parts of swim bladder of Atlantic cod. (a) Pyloric caeca: epithelial cells of the caecal mucosal fold (MuFo; arrows) were Pis1-immunoreactive, and some positivity was also observed in the surrounding tissues (open arrow) and between each caecal portion (*). (b) Proximal intestine: cells in mucosal epithelium (thick arrows) and compound glands (thin arrows), cells in lamina propria (Lp) and mucosal epithelium (arrow heads) were also Pis1-immunoreactive. (c and d) Rectum: cytoplasm of mucosal epithelium (Ep) and cytoplasm of columnar cells in compound glands (Cg) shown in (c) and (d), respectively were Pis1-immunoreactive, while goblet cells (arrow heads) in both mucosal epithelium and compound glands were immunonegative. Some positive multi-granular cells (arrows) in submucosa tissue (Sm) shown in (d) were also Pis1-positive. (e) A weak immunoreactivity was observed in mucosal epithelial cells (thin arrow), while a stronger positive signal was observed in the neuronal cell bodies (thick arrows). (f) Gas gland (Gg) and the connective tissue of the submucosa (Sm) were Pis1-immunoreactive, while a weaker positive signal was observed in the rete mirabile (Rm). (g) Higher magnification of gas gland showing the immunoreactivity of the gas gland cells. Negative control (inset) of the serial sections treated with dilution buffer was Pis1-immunonegative (Larger images are seen in Additional file 1: Figure S4).

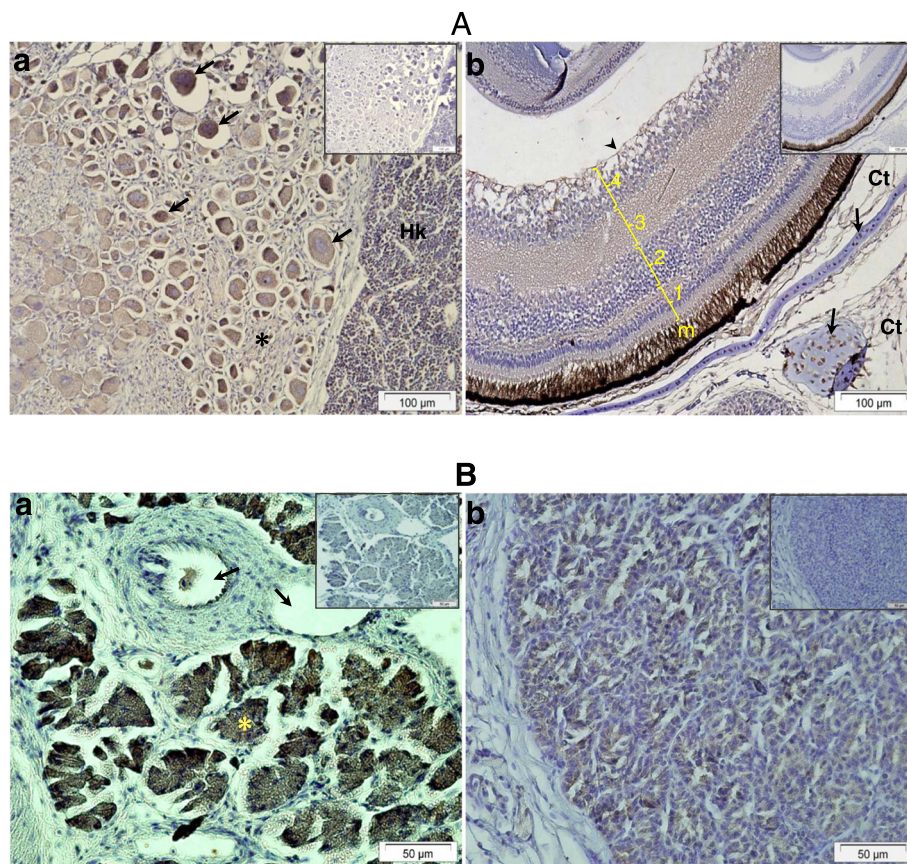


Figure 5 (5A) Immunohistochemical micrographs of different neural tissues (including eye) of Atlantic cod. (a) Parasympathetic ganglia: various sized neuronal cell bodies (arrows), cells and tissues in axons and dendrites (*) were Pis1-immunoreactive (Hk: head kidney's hematopoietic tissue). **(b)** Eye: Pis1-immunoreactivity was found in various cell types and tissues –including 1: sensory cells of the retina; 2: cells and tissues of inner nuclear layer; 3: inner plexiform layer; 4: ganglion cell and nerve fiber layer, and arrowhead shows the inner limiting membrane. The chondrocytes of the hyaline cartilage (arrows), connective tissues (Ct), and tissue of the lens (*) were also immunoreactive, (m: melanocytes). **(5B)** Section of exocrine and endocrine glands of Atlantic cod. **(a)** Exocrine pancreas: the cords of exocrine acinar cells (*) showing strong intensity of Pis1-immunoreactivity; note the presence of the large hepatic veins (arrows). **(b)** Corpuscles of Stannius: Pis1-immunoreactivity was found in the cytoplasm of most secretory cells. Negative control (inset) of the serial sections treated with dilution buffer was Pis1-immunonegative. (Larger images are seen in Additional file 1: Figure S5).

Another important observation in the present study is the discovery of the hitherto unreported information that piscidin is produced by chondrocytes of various cartilaginous tissues (Figure 1, 3 and 5A). In contrast, immunohistochemical studies on other teleost species have shown that the chondrocytes did not contain piscidins [13,14,23,35]. To our knowledge, more information on teleost chondrocytes does not exist. Nevertheless, studies on mammals have shown that this cell type not only produces extracellular matrix, but also acts as a central machinery to produce AMPs and other immune factors, thus contributing to host defence mechanisms in non-vascular cartilaginous tissues [36-38]. Therefore, chondrocytes of Atlantic cod are also likely to have similar potential function, but more investigations are needed to ascertain it. A strong Pis1-immunoreactivity was also observed in the swim bladder. This organ of zebrafish was found to express several

antimicrobial genes [39] and this seems to be the case as well for Atlantic cod [24,40]. As neither the genetic profiling of swim bladder nor its function in innate immune system has been described, additional studies need to be undertaken to determine the reason for the presence of AMPs or the role of AMPs in association with other established functions such as buoyancy, respiration and communication [41]. It should be noted that Silphaduang et al. [13] failed to detect piscidins in the swim bladder of hybrid striped bass, pointing to a species-dependent difference in functionality.

Atlantic cod has a unique immune system architecture compared to other vertebrates [1] and produce little or hardly any antibody upon vaccination [2,5]. Nevertheless, this fish copes with pathogen invasion [5], and its ability to protect itself could be linked to the effective innate immune components [1,3,6]. The data from this study as

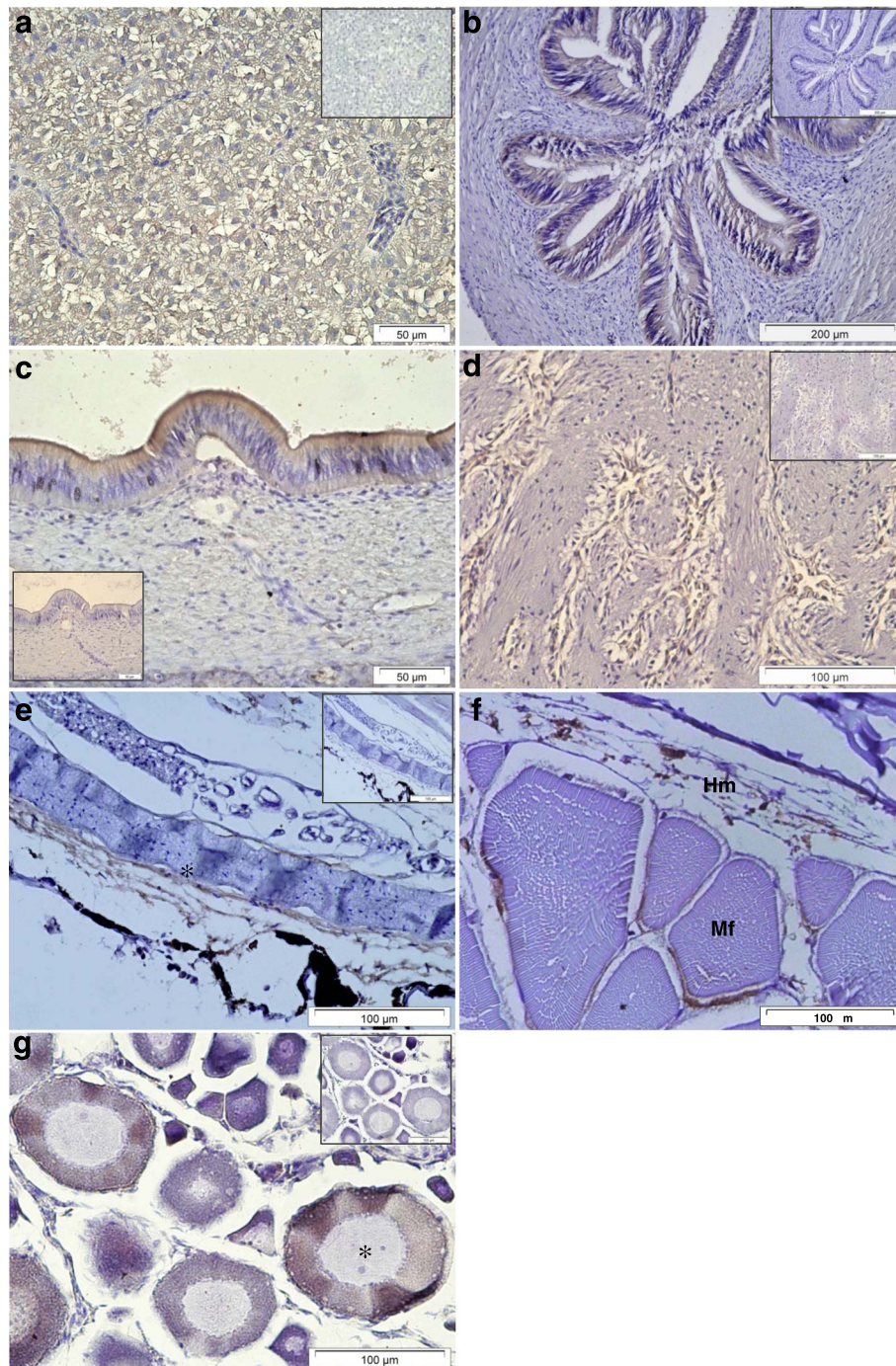


Figure 6 Immunohistochemical micrographs of different tissues of Atlantic cod. (a) Liver: all the hepatocyte cytoplasm and tissue that form cellular network were Pis1-immunoreactive. **(b)** Bile duct and **(c)** Gall bladder epithelium were Pis1-immunoreactive. **(d)** The tissue of bulbus arteriosus: numerous elongated fibrocytes and tissues that form cellular network were Pis1-immunoreactive. **(e)** Peritoneum: tissues that form cellular network lining a layer of mesothelium (*) were Pis1-immunostained. **(f)** Hypodermis layer (Hm) of the skin and muscle fibers (Mf): cells and connective tissues showing Pis1-positivity **(g)** Oocytes: various stages of the cells showing Pis1-immunoreactivity particularly the early or mid vitellogenic stage (*) exhibited strong positive signals. Negative control (inset) of the serial sections treated with dilution buffer was Pis1-immunonegative (Larger images are seen in Additional file 1: Figure S6).

well as from the previous study by Ruangsi et al. [7,24] strongly suggest that Atlantic cod possesses potent

antimicrobial components, particularly AMPs (piscidins), adding to the repertoire of its innate immune system.

In this study we investigated the presence of peptide in apparently healthy fish. The widespread occurrence of Pis1 suggests other possible functions of this peptide in fish, in addition to its roles against microorganisms. Pis1 may have a role in mediating cyto-protection as it was present in several tissues such as liver, bile duct and gall bladder, which are not anatomically exposed to a high pathogen pressure. Further, the high level of Pis1 peptide detected in swim bladder may signify its involvement in repair mechanisms of this tissue. In Atlantic cod, swim bladder damaged by a dramatic change of hydrostatic pressure was found to regain its function, possibly with the assistance of a membrane that lines its wall (cited by [42]). It has to be emphasized that wound healing properties of various AMPs have been reported in humans [43].

Cod Pis1 could also be a neurogenic peptide since its immunoreactivity was observed in neural tissue of some organs (Figure 4 and 5A). Little is known on AMPs and their function in nervous system of fish, however it has been demonstrated that nerve cells and or nerve tissues are never located far away from effector organs [30]. The crosstalk between nerves and immune system of the animal kingdom has long since been a topic of discussion, and it is now clear that there exists a bidirectional flow of information between the central nervous system and the immune system [44-47]. Furthermore, it has been shown that many neuropeptides and peptide hormones have similar characteristics, including antimicrobial properties [47,48]. Moreover, it is evident that together with the nervous system, the endocrine system of all animals including fish, participate in the maintenance of a steady physiologic state, homeostasis [30,47]. As Pis1 appeared in several glands (Figure 4 and 5B), this peptide may have a homeostatic role. Another fish AMP, beta-defensin (previously known as β -defensin) that was expressed highly in the pituitary and testis of orange spotted grouper is believed to maintain cellular homeostasis [49]. Apart from their antimicrobial properties, most of the hepcidins are known to maintain iron homeostasis in vertebrates including fish [50].

Involvement in osmoregulation and excretion are alternative functions which Pis1 may possess, as their presence was evident in kidney (especially trunk kidney), skin, gills, and corpuscles of Stannius of naïve fish. These organs are known to have crucial roles in the osmoregulatory and excretory mechanisms of fish besides their immune functions [30]. In addition, osmoregulatory signature of Pis1 peptide is evident from the immunoreactivity found in the rete mirabile of the swim bladder or the choroid rete mirabile of the fish eye. These unique parts are known to have key roles in oxygen secretion in the swim bladder and the eye of fish [51]. Osmoregulatory functions might also be attributed to peritoneum, hypodermis layer and myosepta where the Pis1-immunoreactivity was detected. The presence of

Psoriasin1, an AMP member that belongs to the calcium-binding protein super-family was detected in peritoneal tissue of horse [52]. Pis1 peptide was also found in all stages of the oocytes, their expression being greater in the advanced developmental stages.

Conclusion

It is evident that various cell types and tissues of Atlantic cod are armed with Pis1 peptide. Its constitutive presence in immune-related tissues of naïve fish suggests a significant role for this peptide in the innate immune system of Atlantic cod. In addition, the localisation of Pis1 in some of the non-immune tissues and organs confirmed our previous tissue distribution results at the molecular level [24] - suggesting other possible roles of this ubiquitous peptide and its capability to maintain homeostasis in Atlantic cod.

Additional file

Additional file 1: Figure S1. Tests to confirm the mono-specificity of anti-Pis1 antibody using tissue extracts from Atlantic cod and the synthetic piscidin peptide. Samples were separated on a 5–16 % acrylamide gel electrophoresis in the presence of sodium dodecyl sulfate (SDS-PAGE) running in Tris-Tricine buffer system, following the procedure of Ruangrsri et al. [7]. (A) Results from coomassie blue staining. (B) Western blot analysis of piscidins and tissue extracts that were transferred onto a PVDF membrane that was blocked for 1 h and incubated overnight at 4° C with anti-Pis1 antibody. Lanes are: 1 - protein marker, 2 - leucocytes from head kidney co-incubated with latex beads for 4 h, 3 - head kidney, 4 - blood, 5 - muscle, 6 - skin, 7 - ovary, 8 - synthetic Pis2b, 9 - synthetic Pis2 and 10 - synthetic Pis1. Anti-Pis antibody was immunoreactive only with Pis1 peptide (Lane 10), while moderate to faint immunoreactivity was detected also in some tissue extracts (Lane 2 and 7) at the positions that corresponded to a molecular weight similar to that of Pis1. Figure S2. Immunohistochemical micrographs of different tissues of Atlantic cod showing absence of immunoreactivity in the control samples of (a) blood, (b) blood mixed with latex beads, (c and d) head kidney and spleen leucocytes mixed with latex beads, (e, f and g) sections of head kidney, trunk kidney and spleen treated with dilution buffer. Figure S3. Immunohistochemical micrographs of control sections treated with dilution buffer, showing immunonegative reactions in tissues of Atlantic cod. (a) Dorsal skin, (b) epidermal layer of skin surrounding eye, (c) gill filament and chondrocytes and (d) adductor muscle. Figure S4. Immunohistochemical micrographs of control sections treated with dilution buffer, showing immunonegative reactions in tissues of Atlantic cod. (a) Pyloric caeca, (b) proximal intestine, (c) mucosal epithelium of rectum, (d) compound gland and submucosa tissue of rectum, (e) swim bladder wall and mucosal epithelium, (f) gas gland and (g) higher magnification of gas gland. Figure S5. Immunohistochemical micrographs of control sections treated with dilution buffer, showing immunonegative reactions in tissues. (A-a) Parasympathetic ganglia, (A-b) eye, (B-a) exocrine pancreas and (B-b) corpuscles of Stannius. Figure S6. Immunohistochemical micrographs of control sections treated with dilution buffer, showing immunonegative reactions in tissues. (a) Liver, (b) bile duct, (c) gall bladder epithelium, (d) bulbus arteriosus, (e) peritoneum and (f) oocytes.

Acknowledgements

This work was supported by the Research Council of Norway under the project 'Mucosal Immune System of Atlantic Cod - Creating a Knowledge Base' (184703). The authors would like to thank Carlos Lanes, for sharing the cod fry stage samples and Divya Kumari Hirkudel Vate for her help in preparation of some test slides for the study. Hilde Ribe and Ingvil Berg are thanked for their technical support. Jareeporn Ruangrsri would like to express

her gratitude to Prince of Songkla University (Thailand) for granting the PhD scholarship that enabled her to study at the University of Nordland, Norway.

Author details

¹Faculty of Biosciences and Aquaculture, University of Nordland, 8049, Bodø, Norway. ²Cell Biology and Immunology Group, Wageningen Institute of Animal Sciences, Wageningen University, Marijkeweg 40, 6709 PG, Wageningen, The Netherlands.

Authors' contributions

JR designed and performed the experiments, analyzed the data and wrote the manuscript. The experiments were overseen by JMOF, MFB, JHWMR and VK, who were also involved in the revision of the manuscript. All authors have read and approved the final manuscript.

Received: 8 December 2011 Accepted: 26 April 2012

Published: 26 April 2012

References

- Star B, Nederbragt AJ, Jentoft S, Grimholt U, Malmstrom M, Gregers TF, Rounge TB, Paulsen J, Solbakken MH, Sharma A, et al: **The genome sequence of Atlantic cod reveals a unique immune system.** *Nature* 2011, **477**:207–210.
- Lund V, Bordal S, Kjellsen O, Mikkelsen H, Schroder MB: **Comparison of antibody responses in Atlantic cod (*Gadus morhua* L.) to *Aeromonas salmonicida* and *Vibrio anguillarum*.** *Dev Comp Immunol* 2006, **30**:1145–1155.
- Magnadóttir B, Jónsdóttir H, Helgason S, Björnsson B, Solem ST, Pilstrom L: **Immune parameters of immunised cod (*Gadus morhua* L.).** *Fish Shellfish Immunol* 2001, **11**:75–89.
- Espelid S, Rødseth OM, Jørgensen TØ: **Vaccination experiments and studies of the humoral immune responses in cod, *Gadus morhua* L., to four strains of monoclonal-defined *Vibrio anguillarum*.** *J Fish Dis* 1991, **14**:185–197.
- Mikkelsen H, Lund V, Larsen R, Seppola M: **Vibriosis vaccines based on various sero-subgroups of *Vibrio anguillarum* O2 induce specific protection in Atlantic cod (*Gadus morhua* L.) juveniles.** *Fish Shellfish Immunol* 2011, **30**:330–339.
- Persson AC, Stet RJM, Pilstrom L: **Characterization of MHC class I and beta (2)-microglobulin sequences in Atlantic cod reveals an unusually high number of expressed class I genes.** *Immunogenetics* 1999, **50**:49–59.
- Ruangrsri J, Fernandes JMO, Brinchmann M, Kiron V: **Antimicrobial activity in the tissues of Atlantic cod (*Gadus morhua* L.).** *Fish Shellfish Immunol* 2010, **28**:879–886.
- Hancock RE: **Peptide antibiotics.** *Lancet* 1997, **349**:418–422.
- Smith VJ, Fernandes JMO: **Antimicrobial peptides of the innate immune system.** In *Fish Defenses*. Edited by Zaccane G, Marchalonis JJ, Schluter SF, Meseguer J, Kapoor BG. USA: Science Publishers; 2009:241–275.
- Noga EJ, Ullal AJ, Corrales J, Fernandes JMO: **Application of antimicrobial polypeptide host defenses to aquaculture: exploitation of downregulation and upregulation responses.** *Comp Biochem Physiol D* 2011, **6**:44–54.
- Sun BJ, Xie HX, Song Y, Nie P: **Gene structure of an antimicrobial peptide from mandarin fish, *Siniperca chuatsi* (Basilewsky), suggests that moronecidins and pleurocidins belong in one family: the piscidins.** *J Fish Dis* 2007, **30**:335–343.
- Salerno G, Parrinello N, Roch P, Cammarata M: **cDNA sequence and tissue expression of an antimicrobial peptide, dicentracin; a new component of the moronecidin family isolated from head kidney leukocytes of sea bass, *Dicentrarchus labrax*.** *Comp Biochem Physiol B* 2007, **146**:521–529.
- Silphaduang U, Colorni A, Noga EJ: **Evidence for widespread distribution of piscidin antimicrobial peptides in teleost fish.** *Dis Aquat Org* 2006, **72**:241–252.
- Silphaduang U, Noga EJ: **Peptide antibiotics in mast cells of fish.** *Nature* 2001, **414**:268–269.
- Yin ZX, He W, Chen WJ, Yan JH, Yang JN, Chan SM, He JG: **Cloning, expression and antimicrobial activity of an antimicrobial peptide, epinecidin-1, from the orange-spotted grouper, *Epinephelus coioides*.** *Aquaculture* 2006, **253**:204–211.
- Noga EJ, Silphaduang U, Park NG, Seo JK, Stephenson J, Kozowicz S: **Piscidin 4, a novel member of the piscidin family of antimicrobial peptides.** *Comp Biochem Physiol B* 2009, **152**:299–305.
- Fernandes JMO, Ruangrsri J, Kiron V: **Atlantic cod piscidin and its diversification through positive selection.** *PLoS One* 2010, **5**:e9501.
- Lauth X, Shike H, Burns JC, Westerman ME, Ostland VE, Carlberg JM, Van Olst JC, Nizet V, Taylor SW, Shimizu C, et al: **Discovery and characterization of two isoforms of moronecidin, a novel antimicrobial peptide from hybrid striped bass.** *J Biol Chem* 2002, **277**:5030–5039.
- Chinchar VG, Bryan L, Silphaduang U, Noga E, Wade D, Rollins-Smith L: **Inactivation of viruses infecting ectothermic animals by amphibian and piscine antimicrobial peptides.** *Virology* 2004, **323**:268–275.
- Sung WS, Lee J, Lee DG: **Fungicidal effect of piscidin on *Candida albicans*: pore formation in lipid vesicles and activity in fungal membranes.** *Biol Pharm Bull* 2008, **31**:1906–1910.
- Colorni A, Ullal A, Heinisch G, Noga EJ: **Activity of the antimicrobial polypeptide piscidin2 against fish ectoparasites.** *J Fish Dis* 2008, **31**:423–432.
- Mulero I, Noga EJ, Meseguer J, Garcia-Ayala A, Mulero V: **The antimicrobial peptides piscidins are stored in the granules of professional phagocytic granulocytes of fish and are delivered to the bacteria-containing phagosome upon phagocytosis.** *Dev Comp Immunol* 2008, **32**:1531–1538.
- Corrales J, Mulero I, Mulero V, Noga EJ: **Detection of antimicrobial peptides related to piscidin 4 in important aquacultured fish.** *Dev Comp Immunol* 2010, **34**:331–343.
- Ruangrsri J, Salger SA, Caipang CMA, Kiron V, Fernandes JMO: **Differential expression and biological activity of two piscidin paralogues and a novel splice variant in Atlantic cod (*Gadus morhua* L.).** *Fish Shellfish Immunol* 2012, **32**:396–406.
- Sompuram SR, Kodala V, Zhang K, Ramanathan H, Radcliffe G, Falb P, Bogen SA: **A novel quality control slide for quantitative immunohistochemistry testing.** *J Histochem Cytochem* 2002, **50**:1425–1434.
- Shi SR, Cote RJ, Taylor CR: **Antigen retrieval immunohistochemistry: past, present, and future.** *J Histochem Cytochem* 1997, **45**:327–343.
- Marrison MC: **Histology of the Atlantic cod, *Gadus morhua*: an atlas.** Part One. Digestive Tract and Associated organs. Canada: Can Spec Publ Fis Aquat Sci; 1987.
- Claver JA, Quaglia AIE: **Comparative morphology, development, and function of blood cells in nonmammalian vertebrates.** *J Exot Pet Med* 2009, **18**:87–97.
- Reite OB, Evensen Ø: **Inflammatory cells of teleostean fish: a review focusing on mast cells/eosinophilic granule cells and rodlet cells.** *Fish Shellfish Immunol* 2006, **20**:192–208.
- Genten F, Terwinghe E, Danguy A: **Atlas of Fish Histology.** USA: Science Publishers; 2009.
- Tort L, Balasch J, Mackenzie S: **Fish immune system: a crossroads between innate and adaptive responses.** *Immunologia* 2003, **23**:277–286.
- Pakers SGM, Uppalapati S, Meyer W, Maderson P, Sell AF, Kruse C, Paus R: **Fish matters: the relevance of fish skin biology to investigative dermatology.** *Exp Dermatol* 2010, **19**:313–324.
- Ellis AE: **Innate host defense mechanisms of fish against viruses and bacteria.** *Dev Comp Immunol* 2001, **25**:827–839.
- Rombout JH, Abelli L, Picchiatti S, Scapigliati G, Kiron V: **Teleost intestinal immunology.** *Fish Shellfish Immunol* 2011, **31**:616–626.
- Dezfuli BS, Pironi F, Giari L, Noga EJ: **Immunocytochemical localization of piscidin in mast cells of infected seabass gill.** *Fish Shellfish Immunol* 2010, **28**:476–482.
- Varoga D, Pufe T, Mentlein R, Kohrs S, Grohmann S, Tillmann B, Hassenpflug J, Paulsen F: **Expression and regulation of antimicrobial peptides in articular joints.** *Ann Anat* 2005, **187**:499–508.
- Warnke PH, Russo PAJ, Hopfenzie M, Kurz B, Becker ST, Sherry E, Springer I, Sivananthan S: **Antimicrobial peptide immunity protects human nasal and auricular cartilage against infection.** *J Craniofac Surg* 2010, **21**:198–201.
- Macrory L, Vaughan-Thomas A, Clegg PD, Innes JF: **An exploration of the ability of tepoxalin to ameliorate the degradation of articular cartilage in a canine *in vitro* model.** *BMC Vet Res* 2009, **5**:25.
- Oehlers SH, Flores MV, Chen T, Hall CJ, Crosier KE, Crosier PS: **Topographical distribution of antimicrobial genes in the zebrafish intestine.** *Dev Comp Immunol* 2011, **35**:385–391.
- Ruangrsri J, Kiron V, Jeppinamogeri L, Fernandes JMO: **A novel beta defensin of Atlantic cod.** submitted.
- Nilsson S: **Nervous control of fish swimbladders.** *Acta Histochem* 2009, **111**:176–184.
- van der Kooij J, Righton D, Strand E, Michalsen K, Thorsteinsson V, Svedäng H, Neat FC, Neuenfeldt S: **Life under pressure: insights from electronic data-storage tags into cod swimbladder function.** *ICES J Mar Sci* 2007, **64**:1293–1301.

43. Steintraesser L, Koehler T, Jacobsen F, Daigeler A, Goertz O, Langer S, Kesting M, Steinau H, Eriksson E, Hirsch T: **Host defense peptides in wound healing.** *Mol Med* 2008, **14**:528–537.
44. Kaplin A, Bartner S: **Reciprocal communication between the nervous and immune systems: crosstalk, back-talk and motivational speeches.** *Int Rev Psychiatr* 2006, **17**:439–441.
45. Salzet M, Vieau D, Day R: **Crosstalk between nervous and immune systems through the animal kingdom: focus on opioids.** *Trends Neurosci* 2000, **23**:550–555.
46. Zhang N, Oppenheim JJ: **Crosstalk between chemokines and neuronal receptors bridges immune and nervous systems.** *J Leukocyte Biol* 2005, **78**:1210–1214.
47. Brogden KA, Guthmiller JM, Salzet M, Zasloff M: **The nervous system and innate immunity: the neuropeptide connection.** *Nat Immunol* 2005, **6**:558–564.
48. El Karim IA, Linden GJ, Orr DF, Lundy FT: **Antimicrobial activity of neuropeptides against a range of micro-organisms from skin, oral, respiratory and gastrointestinal tract sites.** *Neuroimmunology* 2008, **200**:11–16.
49. Jin JY, Zhou L, Wang Y, Li Z, Zhao JG, Zhang QY, Gui JF: **Antibacterial and antiviral roles of a fish beta-defensin expressed both in pituitary and testis.** *PLoS One* 2010, **5**:e12883.
50. Rodrigues PNS, Vázquez-Dorado S, Neves JV, Wilson JM: **Dual function of fish hepcidin: Response to experimental iron overload and bacterial infection in sea bass (*Dicentrarchus labrax*).** *Dev Comp Immunol* 2006, **30**:1156–1167.
51. Berenbrink M: **Historical reconstructions of evolving physiological complexity: O₂ secretion in the eye and swimbladder of fishes.** *J Exp Biol* 2007, **210**:1641–1652.
52. Bruhn O, Grotzinger J, Cascorbi I, Jung S: **Antimicrobial peptides and proteins of the horse-insights into a well-armed organism.** *Vet Res* 2011, **42**:98.

doi:10.1186/1746-6148-8-46

Cite this article as: Ruangrsri et al.: Ubiquitous presence of piscidin-1 in Atlantic cod as evidenced by immunolocalisation. *BMC Veterinary Research* 2012 **8**:46.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

