



Journal of Immunoassay and Immunochemistry

ISSN: 1532-1819 (Print) 1532-4230 (Online) Journal homepage: https://www.tandfonline.com/loi/ljii20

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To cite this article: Ebrahim Kouchaki, Hassan Nikoueinejad, Hossein Akbari, Shirin Azimi & Mohammad Behnam (2019): The investigation of relevancy between *PIAS1* and *PIAS2* gene expression and disease severity of multiple sclerosis, Journal of Immunoassay and Immunochemistry, DOI: <u>10.1080/15321819.2019.1613244</u>

To link to this article: https://doi.org/10.1080/15321819.2019.1613244



Published online: 13 May 2019.

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The investigation of relevancy between *PIAS1* and *PIAS2* gene expression and disease severity of multiple sclerosis

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ABSTRACT

Introduction: *PIAS1* and *PIAS2* (protein inhibitor of activated STAT 1,2) play key roles in the pathogenesis of autoimmune and inflammatory diseases. This study aims to evaluate the gene expression of these factors in multiple sclerosis (MS) patients compared to healthy individuals and correlate them with the severity of MS.

Materials and methods: Sixty participants, including 30 patients with MS and 30 healthy controls were studied. The expression of *PIAS1* and *PIAS2* genes in peripheral blood samples of all participants was measured by real-time PCR. The severity of MS was evaluated using the Expanded Disability Status Scale (EDSS). Finally, we evaluated the correlation between the expression of *PIAS1* and *PIAS2* genes with disease severity.

Results: The expression of *PIAS1* gene was increased in patients with MS compared to healthy subjects (*P* value<.001). Also, there was a significant correlation between the expression of *PIAS1* and *PIAS2* genes with disease severity according to EDSS. **Conclusion**: Our study suggests the expression of *PIAS1* and *PIAS2* genes as a prognostic and diagnostic marker in MS disease.

KEYWORDS

Multiple sclerosis; *PIAS1*; PIAS2; EDSS

Introduction

Multiple sclerosis (MS), a chronic autoimmune demyelinating disease of central nervous system,^[1] is mediated by autoreactive T cells^[2] and leads to progressive neurological disability.^[3] It is estimated that about 2.5 million people worldwide are diagnosed with MS, mainly young adults at the age between 20 and 40.^[4] The etiology of MS is yet unknown; however, a combination of immunological as well as environmental factors such as vitamin D deficiency, smoking,^[5] increase in

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cytomegalovirus, positivity of anti-CMV IgG and IgM antibodies,^[6] decreased *Helicobacter pylori*,^[7] imbalance of inflammatory and anti-inflammatory factors,^[8] and genetic factors^[9] are thought to be cause or contribute to MS. The classification of MS based on the severity of clinical symptoms includes: relapsing-remitting MS (RRMS), secondary progressive MS (SPMS), progressive relapsing MS (PRMS), and primary progressive MS (PPMS).^[8] Depending on the brain regions attacked by the immune system, the patients suffer from a variety of symptoms including muscle weakness, paresthesia, ataxia, and visual disturbances.^[4] Successful therapies target the immune system indicating such system plays an important role in the disease activity.

Different combinations of the Janus kinas/Signal Transducer and Activator of Transcription (JAK/STAT) factors, utilized by over 70 cytokines as well as growth factors,^[10] initiate and orchestrate innate and adaptive immune responses.^[9] The regulation of such pathway seems to be critical; and in this regard, the Protein Inhibitor of Activated STAT (PIAS) proteins contribute in endogenous negative regulation of JAK/STAT pathway.^[11] Actually, dysregulation of JAK/STAT pathway has pathologically been implicated in some autoimmune diseases such as MS.^[3]

PIAS family consists of *PIAS1*, *PIAS2* (also known as PIASx), *PIAS3*, and *PIAS4* (also known as PIASy).^[12,13] PIAS3 and PIAS2 interact with STAT3 and STAT4, respectively; while PIAS1 and PIASy interact with STAT1.^[13] It has been proposed that PIASs regulate the function of over 60 proteins through various molecular mechanisms^[14–16] including blocking the DNA binding activity of transcription factors, recruiting transcriptional co-receptors or coactivators, promoting protein somuylation, and sequestrating transcription factors to certain subnuclear structures where co-receptor complexes are enriched.^[14,15] This family also acts as potential regulators of cell proliferation, inflammatory responses,^[17,18] and tumor development.^[19] The PIAS family expression is known to be related to many inflammatory and autoimmune diseases such as CNS inflammation,^[20–22] several oncogene conditions,^[23] type 2 diabetes mellitus,^[24] and viral infections.^[25]

Considering the effects of PIAS protein family in inflammatory conditions, we proposed the probable relation of gene expression of PIAS1 and PIAS2 to the autoimmune disease of MS and its severity. Such investigation may reveal a prognostic role of PIAS1 and PIAS2 in MS and may also suggest new therapeutic clues.

Methods and materials

Study subjects

This case-control study was performed on 30 patients with MS and 30 healthy volunteers in the Neurology Clinic of Shahid Beheshti Hospital in

Kashan city, Iran. The inclusion criteria were MS patients with ages between 20 and 60, course of disease (RRMS, PPMS, SPMS, and PRMS) identified according to McDonald criteria.^[26] All patients were treated with IFN- β . Any other chronic inflammatory/autoimmune diseases, clinical relapse, and glu-cocorticoid therapy during the past 1 month were considered as exclusion criteria. Healthy volunteers had no any history of chronic inflammatory and autoimmune diseases. In addition, healthy participants were matched to the patients for age and gender. The protocol was approved by the ethics committee of Kashan University of Medical Science. Written informed consent was obtained from all participants.

Clinical assessment

The severity of MS was evaluated using the Expanded Disability Status Scale (EDSS), a clinician-administered assessment scale evaluating the functional systems of the central nervous system. Today, EDSS is used to describe disease progression in patients with MS and to assess the effectiveness of therapeutic interventions in clinical trials.^[8,27] It consists of ordinal rating system ranging from 0 (normal neurological status) to 10 (death due to MS) in 0.5 increments interval.^[28] Patients were divided into two groups of mild and severe according to their EDSS score. Patients with the score of 0.5–4.5 were placed in mild group and those with the EDSS score of 5–9.5 were placed in severe group. Patients in remission phase were enrolled in the study.

Laboratory procedures

Peripheral venous blood samples were collected from all participants. Fresh peripheral blood mononuclear cells were separated from 2 ml of anticoagulated blood by Ficoll–Hypaque (Lymphodex, Inno-Train, Germany) density gradient centrifugation. Total RNA was extracted from peripheral blood mononuclear cells using High Pure RNA Isolation Kit (Cat No: 11828665001, Roche Applied Science, Germany), and cDNA was synthesized from the extracted RNA using Transcriptor First Strand cDNA Synthesis Kit (Cat No: 04897030001, Roche Applied Science, Germany). The amounts of *PIAS1* and *PIAS2* gene expression were measured through Taqman primer probe Comparative CT method using ABI 7300 Real-Time PCR system. β -Actin housekeeping gene was used as endogenous control.

Statistical analysis

The statistical indices of the *PIAS1* and *PIAS2* gene expression were analyzed by independent t and chi-square tests. Using Pearson's correlation coefficient, we calculated the correlations between variables. All analyses were

performed using the SPSS 16 software. Data were expressed as mean \pm standard deviation (SD).

Results

The expression of *PIAS1* gene was 9.13 ± 1.96 and 6.77 ± 1.99 in MS patients and healthy controls, respectively. The expression of *PIAS1* gene was significantly higher in MS patients than in healthy subjects (P < .001). However, there was no significant difference in the expression of *PIAS2* gene between two groups (P = .39) (Table 1).

We found a significant correlation between the gene expression of *PIAS1* with EDSS score (R = 0.65, P < .001). We also found a significant correlation between the expression of *PIAS2* gene with EDSS score (R = 0.58, P < .001). The correlation between *PIAS1* and *PIAS2* gene expression was 0.32 (P = .012) (Table 2).

The expression of *PIAS1* gene was significantly higher in severe forms (10.3 ± 1.25) of MS than that in mild forms (8.55 ± 2.01) (P = .018). There was also a significant increase in the gene expression of PIAS2 according to EDSS score. The gene expression of PIAS2 was 7.55 ± 1.39 in mild forms of MS and 8.7 ± 1.25 in severe forms (P = .036) (Table 3).

Using receiver operating characteristic (ROC) curve and Area under Curve (AUC), we determined the sensitivity and specificity of both *PIAS1* and *PIAS2* gene expression as diagnostic markers of MS severity. In the best cutoff point of *PIAS1* gene expression = 7.5, sensitivity and specificity were 80% and 63.3%, respectively; and AUC was determined as 0.802. Considering

Variants		MS patients ($n = 30$)	Healthy controls ($n = 30$)	P value
Age (years)		38.11 ± 13.88	29.92 ± 9.52	0.019
Gender (male/female	e)	(5/25)	(9/21)	0.222
Disease duration (ye	ars)	6.62 ± 4.18	-	-
Type of disease	RRMS	23 (76.7%)	-	-
	PPMS	0 (0%)	-	
	SPMS	5 (16.7%)	-	
	PRMS	2 (6.7%)	-	
Type of treatment	No drug	2 (7.1%)	-	-
	Cinovex	20 (71.4%)	-	
	Rebief	5 (17.9%)	-	
	Others	1 (3.6%)	-	
Family history	Positive	4 (13.3%)	-	-
	Negative	26 (86.7%)	-	
Number of relapses		4.4 ± 4.1	-	-
Treatment duration (years)		3.58 ± 3.1	-	-
EDSS	Mild (0.5-4.5)	20 (66.6%)	-	-
	Severe (5–9.5)	10 (33.3%)	-	-
PIAS1		9.13 ± 1.96	6.77 ± 1.99	< 0.001
PIAS2		7.93 ± 1.43	7.53 ± 2.09	0.392

Table 1. Basic and clinical characteristic of MS patients and healthy subjects.

	PIAS1		PIAS2		
	Correlation coefficient	P value	Correlation coefficient	P value	
PIAS1	1	-	0.322	0.012	
PIAS2	0.322	0.012	1	-	
EDSS	0.656	< 0.001	0.588	0.001	
Number of relapses	0.374	0.042	0.314	0.091	
Treatment duration	0.294	0.121	0.225	0.241	
Duration of disease	0.308	0.098	0.112	0.557	
Age	0.346	0.012	-0.048	0.733	

 Table 2. Pearson's correlation coefficient between the expression of PIAS1 and PIAS2 genes with other variables in multiple sclerosis (MS) patients.

Table 3. PIAS1 and PIAS2 gene expression according to MS severity.

Variant	EDSS group	Number	$Mean\pmSD$	P value
PIAS1	Mild (0.5–4.5)	20	8.55 ± 2.012	0.018
	Severe (5–9.5)	10	10.30 ± 1.252	
PIAS2	Mild (0.5–4.5)	20	7.55 ± 1.395	0.036
	Severe (5–9.5)	10	8.7 ± 1.252	

Table 4. Sensitivity and specificity of PIAS1 and PIAS2 gene expression in ROC curve.

Inflammatory markers	Optimal cutoff point	Sensitivity	Specificity	LR+	LR-	AUC
PIAS1	7.5	80%	63.3%	2.17	3.16	0.802
PIAS2	6.5	80%	33.3%	1.19	1.665	0.554

the optimal cutoff point of *PIAS2* gene expression = 6.5, sensitivity was 80% and specificity was 33.3%, and AUC was determined as 0.554 (Table 4) (Figure 1).

Discussion

The present study aimed to evaluate the gene expression of *PIAS1* and *PIAS2* in MS patients. Rare studies evaluated PIASs in patients with MS. In a study by Catherine O'Doherty et al.,^[20] a significant relevancy between the gene expression of *PIAS1* and MS as well as responding to IFN-B treatment has been shown. Multi-allelic combinations analysis determined a JAK2-IL10RB-GBP1-PIAS1 combination was most significant. Evaluating the potential underlying biallelic patterns showed JAK2-IL10RB as the core element. However, the triplet including quartet with an additional PIAS1 allele or GBP1 allele demonstrated to be more influential.^[20] By evaluating RRMS peripheral blood mononuclear cells in the active and stable phases and its differences, a specific genomic signature for RRMS was determined, which PIAS-1were determined is related to the active phase of RRMS.^[29] However, in contrast to the results of these studies, there was no significant change in gene expression of PIAS genes between autism spectrum disorder patients and healthy subjects.^[30]



Figure 1. Sensitivity and specificity of PIAS1 and PIAS2 gene expression.

differences in diseases and its pathophysiology might explain the discrepancies among the results of studies.

Mechanistically, Members of PIAS family have been shown to interact with various STATs.^[14] PIAS3 and PIAS2 interact with STAT3 and STAT4, respectively; and PIAS1 and PIAS9 interact with STAT1. PIAS1 and PIAS3 exert their negative regulation by blocking the DNA binding of STAT1 and STAT3, respectively.^[13,15] On the other hand, PIAS2 and PIAS9 repress the transcriptional activity of STAT1 and STAT4 by recruiting co-repressor molecules such as histone deacetylases.^[13] Particularly, PIAS1 is a key regulator of the inflammation cascade of innate immunity.^[24] Such role of PIAS1 has been suggested in the control of autoimmune conditions such as insulin sensitivity in type 2 diabetes,^[24] essential thrombocytopenia,^[31] and primary hyperaldosteronism.^[32] PIAS1 overexpression has also been implicated in the regulation of several oncogenic conditions^[23] such as multiple myeloma,^[12] breast,^[33] prostate,^[34] colon,^[19] and lung cancer^[23] as well as infections of HIV^[35] and HSV1.^[36,37] In neurogenic tissues, PIAS1 expression is linked to neuronal plasticity as well as neuro-inflammation; and from this point of view, its expression may be associated with Huntington.^[22]

In line with the results of other study,^[29] our data showed an increased gene expression of PIAS1 in MS patients that was correlated to disease severity according to EDSS score. Considering the role of PIAS1 as an inhibitory factor of JAK-STAT pathway, we may conclude that the overexpression of PIAS1 in MS patients acts as a protective mechanism to decrease the disease severity.

Although, such mechanism would not be adequate to compensate the disease development, it may suggest a possible therapeutic clue in autoimmune condition of MS. Such association has also been shown in the case of other inflammatory cytokine inhibitors like SOCS1 and SOCS3.^[38,39] The relationship described above between PIAS1 and clinical conditions has also been shown in the case of PIAS2. For example, PIAS2 is overexpressed in several malignancies such as malignant melanoma^[40] as well as thyroid cancer^[41] and viral infections such as HBV and HCV.^[25,42] PIAS2 also plays some roles in neuronal system. A study suggested the novel role of PIAS2 somuylation in brain development and plasticity.^[43] From this point of view, PIAS2 alterations may contribute in some brain-related diseases such as schizophrenia.^[21] In a recent study, Vavougios et al. showed a significant relation between PIAS2 gene expression and MS disease.^[44] Unlike the latter study,^[44] we found no significant relation between gene expression of PIAS2 and MS disease. However, the gene expression of PIAS2 was associated to disease severity. According to EDSS score, PIAS2 gene expression was significantly higher in severe forms of MS than in mild forms. Our study showed a significant correlation between the expression of both PIAS1 and PIAS2 genes with disease severity according to EDSS score. This finding suggests PIAS1 and PIAS2 as predictors of severe forms of MS which is independent of sex, treatment duration, and the duration of the disease. We also demonstrated that the specificity of PIAS1 gene expression was higher than PIAS2 gene expression in the diagnosis of MS. It may reveal the potential role of PIAS1 as a new diagnostic factor for MS disease.

The limitation of our study was that we did not monitor the gene expression of PIAS1 and PIAS2 longitudinally. This limitation allowed just a cross-sectional analysis of their changes of only limited robustness. Other factors except for *PIAS1* and *PIAS2*, such as other PIAS genes may affect the MS severity which should be determined in future studies. Unfortunately, due to funding limitations, we did not assess the gene expression of other PIAS genes in the patients with MS. Moreover, further studies are needed to confirm those findings with larger sample size.

Conclusion

Our findings show the elevation of expression of *PIAS1* gene in patients with MS compared to healthy controls. We demonstrated that the *PIAS1* and *PIAS2* gene expression is correlated to disease severity according to EDSS. These findings may reveal a new diagnostic approach to MS severity and also suggests new therapeutic approaches to this disease.

Acknowledgments

This study was originated from the studies related to a research supported by Deputy of Research, Kashan University of Medical Sciences (kaums), Grant No. 96162.

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Declaration of interest

The authors declare no conflict of interest.

Authors' contributions

HN contributed in the conception or design of the work, analysis and drafting of the manuscript. EK, HA, and SA contributed in conception and manuscript drafting. The final version was confirmed by all authors for submission

References

- Höer, A.; Schiffhorst, G.; Zimmermann, A.; Fischaleck, J.; Gehrmann, L.; Ahrens, H.; Carl, G.; Sigel, K.-O.; Osowski, U.; Klein, M.; et al. Multiple Sclerosis in Germany: Data Analysis of Administrative Prevalence and Healthcare Delivery in the Statutory Health System. *BMC Health Serv. Res.* 2014, *14*(1), 381. DOI: 10.1186/1472-6963-14-381.
- [2] Olsson, T.; Zhi, W. W.; Höjeberg, B.; Kostulas, V.; Jiang, Y.; Anderson, G.; Ekre, H. P.; Link, H. Autoreactive T Lymphocytes in Multiple Sclerosis Determined by Antigen-Induced Secretion of Interferon-Gamma. *J. Clin. Invest.* 1990, *86*(3), 981–985. DOI: 10.1172/ JCI114800.
- [3] Hurtado-Guerrero, I.; Pinto-Medel, M. J.; Urbaneja, P.; Rodriguez-Bada, J. L.; León, A.; Guerrero, M.; Fernández, Ó.; Leyva, L.; Oliver-Martos, B.; Lenz, L. L. Activation of the Jak-Stat Signaling Pathway After in Vitro Stimulation with IFNss in Multiple Sclerosis Patients According to the Therapeutic Response to IFNss. *PLoS One.* 2017, *12*(1), e0170031. DOI: 10.1371/journal.pone.0170031.
- [4] Wetzels, S.; Wouters, K.; Miyata, T.; Scheijen, J. L.; Hendriks, J. J.; Schalkwijk, C. G.; Vanmierlo, T. Advanced Glycation Endproducts are Increased in the Animal Model of Multiple Sclerosis but Cannot Be Reduced by Pyridoxamine Treatment or Glyoxalase 1 Overexpression. *Int. J. Mol. Sci.* 2018, *19*, 5. DOI: 10.3390/ijms19051311.
- [5] Wingerchuk, D. M.; Environmental Factors in Multiple Sclerosis: Epstein-Barr Virus, Vitamin D, and Cigarette Smoking. *Mt. Sinai J. Med.* 2011, 78(2), 221–230. DOI: 10.1002/msj.v78.2.
- [6] Sanadgol N, Ramroodi N, Ahmadi GA, Komijani M, Moghtaderi A, Bouzari M; Rezaei M, Kardi MT, Dabiri S, Moradi M, et al. Prevalence of Cytomegalovirus Infection and Its Role in Total Immunoglobulin Pattern in Iranian Patients with Different Subtypes of Multiple Sclerosis. *Microbiol. Q. J. Microbiol. Sci.* 2011, 34(3), 263.
- [7] Mohebi, N.; Mamarabadi, M.; Moghaddasi, M. Relation of Helicobacter pylori Infection and Multiple Sclerosis in Iranian Patients. *Neurol. Int.* 2013, 5, 2. DOI: 10.4081/ni.2013.e10.
- [8] Kouchaki, E.; Tamtaji, O. R.; Dadgostar, E.; Karami, M.; Nikoueinejad, H.; Akbari, H. Correlation of Serum Levels of Il-33, Il-37, Soluble Form of Vascular Endothelial Growth Factor Receptor 2 (Vegfr2), and Circulatory Frequency of VEGFR2-expressing Cells with Multiple Sclerosis Severity. *Iran. J. Allergy Asthma Immunol.* 2017, 16(4), 329–337.
- [9] Benveniste, E. N.; Liu, Y.; McFarland, B. C.; Qin, H. Involvement of the Janus Kinase/ Signal Transducer and Activator of Transcription Signaling Pathway in Multiple Sclerosis and the Animal Model of Experimental Autoimmune Encephalomyelitis. *J. Interferon Cytokine Res.* 2014, 34(8), 577–588. DOI: 10.1089/jir.2014.0012.

- [10] Yan, Z.; Gibson, S. A.; Buckley, J. A.; Qin, H.; Benveniste, E. N. Role of the JAK/STAT Signaling Pathway in Regulation of Innate Immunity in Neuroinflammatory Diseases. *Clin. Immunol.* 2016, 189, 4–13. doi: 10.1016/j.clim.2016.09.014.
- [11] Fewings, N.; Gatt, P.; McKay, F.; Parnell, G.; Schibeci, S.; Edwards, J.; Basuki, M. A.; Goldinger, A.; Fabis-Pedrini, M. J.; Kermode, A. G.; et al. The Autoimmune Risk Gene ZMIZ1 Is a Vitamin D Responsive Marker of a Molecular Phenotype of Multiple Sclerosis. J. Autoimmun. 2017, 78, 57–69. DOI: 10.1016/j.jaut.2016.12.006.
- [12] Rabellino, A.; Andreani, C.; Scaglioni, P. P. The Role of PIAS SUMO E3-Ligases in Cancer. Cancer Res. 2017. DOI: 10.1158/0008-5472.CAN-16-2958.
- [13] Furqan, M.; Mukhi, N.; Lee, B.; Liu, D. Dysregulation of JAK-STAT Pathway in Hematological Malignancies and JAK Inhibitors for Clinical Application. *Biomarker Res.* 2013, 1(1), 1. DOI: 10.1186/2050-7771-1-5.
- [14] Shuai, K.; Regulation of Cytokine Signaling Pathways by PIAS Proteins. *Cell Res.* 2006, 16(2), 196. DOI: 10.1038/sj.cr.7310027.
- [15] Wang, R.; Huang, S.; Fu, X.; Huang, G.; Yan, X.; Yue, Z.; Chen, S.; Li, Y.; Xu, A. The Conserved Ancient Role of Chordate Pias as a Multilevel Repressor of the NF-κB Pathway. *Sci. Rep.* 2017, 7(1), 17063. DOI: 10.1038/s41598-017-16624-7.
- [16] Wang J, Sun Z, Zhang Z, Saadi I, Wang J, Li X, Gao S; Engle JJ; Kuburas A; Fu X; Yu W. Protein Inhibitors of Activated STAT (PIAS1 and PIASy) Differentially Regulate Pituitary Homeobox 2 (PITX2) Transcriptional Activity. *J. Biol. Chem.* 2013, M112, 374561.
- [17] Liu Y, Zhang Y-D, Guo L, Huang H-Y, Zhu H, Huang J-X; Liu Y; Zhou SR; Dang YJ; Li X; et al.. Protein Inhibitor of Activated Stat 1 (PIAS1) Is Identified as the Sumo E3 Ligase of CCAAT/Enhancer-Binding Protein (C/EBP) β During Adipogenesis. *Mol. Cell. Biol.* 2013, 33(22), 4606–4617. doi: 10.1128/MCB.00723-13.
- [18] Liu, B.; Shuai, K. Targeting the PIAS1 SUMO Ligase Pathway to Control Inflammation. *Trends Pharmacol. Sci.* 2008, 29(10), 505–509. DOI: 10.1016/j.tips.2008.07.008.
- [19] Coppola, D.; Parikh, V.; Boulware, D.; Blanck, G. Substantially Reduced Expression of PIAS1 Is Associated with Colon Cancer Development. J. Cancer Res. Clin. Oncol. 2009, 135(9), 1287–1291. DOI: 10.1007/s00432-009-0570-z.
- [20] O'Doherty, C.; Favorov, A.; Heggarty, S.; Graham, C.; Favorova, O.; Ochs, M.; Hawkins, S.; Hutchinson, M.; O'Rourke, K.; Vandenbroeck, K. Genetic Polymorphisms, Their Allele Combinations and IFN-β Treatment Response in Irish Multiple Sclerosis Patients. *Pharmacogenomics*. 2009, 10(7), 1177–1186. DOI: 10.2217/pgs.09.41.
- [21] Andrews, J. L.; Goodfellow, F. J.; Matosin, N.; Snelling, M. K.; Newell, K. A.; Huang, X.-F.; Fernandez-Enright, F. Alterations of Ubiquitin Related Proteins in the Pathology and Development of Schizophrenia: Evidence from Human and Animal Studies. *J. Psychiatr. Res.* 2017, *90*, 31–39. DOI: 10.1016/j.jpsychires.2017.01.009.
- [22] Ochaba, J.; Monteys, A. M.; O'Rourke, J. G.; Reidling, J. C.; Steffan, J. S.; Davidson, B. L.; Thompson, L. M. PIAS1 Regulates Mutant Huntingtin Accumulation and Huntington's Disease-Associated Phenotypes in Vivo. *Neuron.* 2016, *90*(3), 507–520. DOI: 10.1016/j. neuron.2016.03.016.
- [23] Rabellino, A.; Melegari, M.; Tompkins, V. S.; Chen, W.; Van Ness, B. G.; Teruya-Feldstein, J.; Conacci-Sorrell, M.; Janz, S.; Scaglioni, P. P. PIAS1 Promotes Lymphomagenesis through MYC Upregulation. *Cell Rep.* 2016, *15*(10), 2266–2278. DOI: 10.1016/j.celrep.2016.05.015.
- [24] Liu Y, Ge X, Dou X, Guo L, Liu Y, Zhou S-R; Wei XB; Qian SW, Huang HY; Xu CJ et al. Protein Inhibitor of Activated STAT 1 (PIAS1) Protects against Obesity-Induced Insulin Resistance by Inhibiting Inflammation Cascade in Adipose Tissue. *Diabetes*. 2015, 64(12), 4061–4074. doi: 10.2337/db15-0278.

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- [25] Pan, Y.; Dai, J.; Liao, Y.; Yu, Q. MicroRNA-137 Promotes Hepatitis B Virus Gene Expression and Replication Via Targeting the Protein Inhibitor of Activated Stat 2. *Die Pharmazie*. 2017, 72(9), 550–554.
- [26] Polman, C. H.; Reingold, S. C.; Banwell, B.; Clanet, M.; Cohen, J. A.; Filippi, M.; Fujihara, K.; Havrdova, E.; Hutchinson, M.; Kappos, L.; et al. Diagnostic Criteria for Multiple Sclerosis: 2010 Revisions to the McDonald Criteria. *Ann. Neurol.* 2011, 69(2), 292–302. DOI: 10.1002/ana.22366.
- [27] Salami, M.; Kouchaki, E.; Asemi, Z.; Tamtaji, O. R. How Probiotic Bacteria Influence the Motor and Mental Behaviors as Well as Immunological and Oxidative Biomarkers in Multiple Sclerosis? A Double Blind Clinical Trial. *J. Funct. Foods.* 2019, *52*, 8–13. DOI: 10.1016/j.jff.2018.10.023.
- [28] Meyer-Moock, S.; Feng, Y.-S.; Maeurer, M.; Dippel F-W, K. T.; Berg-Hansen, P.; Nygaard, G. O.; Sandvik, L.; Lie, B. A.; Celius, E. G.; Harbo, H. F. Systematic Literature Review and Validity Evaluation of the Expanded Disability Status Scale (EDSS) and the Multiple Sclerosis Functional Composite (MSFC) in Patients with Multiple Sclerosis. *BMC Neurol.* 2014, 14(1), 58. DOI: 10.1186/s12883-014-0196-x.
- [29] Ferrandi, C.; Richard, F.; Tavano, P.; Hauben, E.; Barbié, V.; Gotteland, J.-P.; Greco, B.; Fortunato, M.; Mariani, M. F.; Furlan, R.; et al. Characterization of Immune Cell Subsets During the Active Phase of Multiple Sclerosis Reveals Disease and c-Jun N-Terminal Kinase Pathway Biomarkers. *Mult. Scler. J.* 2011, *17*(1), 43–56. DOI: 10.1177/1352458510381258.
- [30] Eftekharian MM, Noroozi R, Omrani MD, Arsang-Jang S, Komaki A, Taheri M; Ghafouri-Fard S. Expression Analysis of Protein Inhibitor of Activated STAT (PIAS) Genes in Autistic Patients. *Adv. Neuroimmune Biol.* 2018, Preprint, 1–6. DOI: 10.3233/ NIB-180144.
- [31] Hsiao, H.; Liu, Y.; Yang, M.; Tsai, Y.; Liu, T.; Chang, C.; Lin, S.-F. Decreased Expression of PIAS1 and PIAS3 in Essential Thrombocythemia Patients. *Genet. Mol. Res.* 2013, 12(5617), 22. DOI: 10.4238/2013.November.18.10.
- [32] Scortegagna, M.; Berthon, A.; Settas, N.; Giannakou, A.; Garcia, G.; Li, J.-L.; Rives, M.; Georges, N.; Garcia-Bonnet, N.; Sylla, A. I.; et al. The E3 Ubiquitin Ligase Siah1 Regulates Adrenal Gland Organization and Aldosterone Secretion. *JCI Insight* 2017, 2, 23. DOI: 10.1172/jci.insight.88864.
- [33] Chanda, A.; Chan, A.; Deng, L.; Kornaga, E. N.; Enwere, E. K.; Morris, D. G.; Bonni, S.; Agoulnik, I. U. Identification of the SUMO E3 Ligase PIAS1 as a Potential Survival Biomarker in Breast Cancer. *PLoS One.* 2017, *12*(5), e0177639. DOI: 10.1371/journal. pone.0177639.
- [34] Hoefer, J.; Schäfer, G.; Klocker, H.; Erb, H. H.; Mills, I. G.; Hengst, L.; Puhr, M.; Culig, Z. PIAS1 Is Increased in Human Prostate Cancer and Enhances Proliferation through Inhibition of P21. Am. J. Pathol. 2012, 180(5), 2097–2107. DOI: 10.1016/j. ajpath.2012.01.026.
- [35] Sachdeva, M.; Sharma, A.; Arora, S. K. Increased Expression of Negative Regulators of Cytokine Signaling during Chronic HIV Disease Cause Functionally Exhausted State of Dendritic Cells. *Cytokine*. 2017, *91*, 118–123. DOI: 10.1016/j.cyto.2016.08.010.
- [36] Brown, J. R.; Conn, K. L.; Wasson, P.; Charman, M.; Tong, L.; Grant, K.; McFarlane, S.; Boutell, C. SUMO Ligase Protein Inhibitor of Activated STAT1 (PIAS1) Is a Constituent Promyelocytic Leukemia Nuclear Body Protein that Contributes to the Intrinsic Antiviral Immune Response to Herpes Simplex Virus 1. J. Virol. 2016, 90(13), 5939–5952. DOI: 10.1128/JVI.00426-16.
- [37] Conn KL, Wasson P, McFarlane S, Tong L, Brown JR, Grant KG; Domingues P; Boutell C. A Novel Role for Protein Inhibitor of Activated STAT 4 (PIAS4) in the

Restriction of Herpes Simplex Virus 1 (HSV-1) by the Cellular Intrinsic Antiviral Immune Response. J. Virol. 2016, JVI, 4807–4826.

- [38] Sedeño-Monge, V.; Arcega-Revilla, R.; Rojas-Morales, E.; Santos-López, G.; Perez-Garc ía, J. C.; Sosa-Jurado, F.; Vallejo-Ruiz, V.; Solis-Morales, C. L.; Aguilar-Rosas, S.; Reyes-Leyva, J. Quantitative Analysis of the Suppressors of Cytokine Signaling 1 and 3 in Peripheral Blood Leukocytes of Patients with Multiple Sclerosis. *J. Neuroimmunol.* 2014, 273(1–2), 117–119. DOI: 10.1016/j.jneuroim.2014.05.013.
- [39] Vandenbroeck, K.; Alvarez, J.; Swaminathan, B.; Alloza, I.; Matesanz, F.; Urcelay, E.; Comabella, M.; Alcina, A.; Fedetz, M.; Ortiz, M. A.; et al. A Cytokine Gene Screen Uncovers SOCS1 as Genetic Risk Factor for Multiple Sclerosis. *Genes Immun.* 2012, *13* (1), 21. DOI: 10.1038/gene.2011.44.
- [40] Ehlken, H.; Schadendorf, D.; Eichmüller, S. Humoral Immune Response against Melanoma Antigens Induced by Vaccination with Cytokine Gene-Modified Autologous Tumor Cells. Int. J. Cancer. 2004, 108(2), 307–313. DOI: 10.1002/ijc.11537.
- [41] Tuccilli, C.; Baldini, E.; Sorrenti, S.; Di, C. G.; Bosco, D.; Ascoli, V.; Mian, C.; Barollo, S.; Rendina, R.; Coccaro, C.; et al. Papillary Thyroid Cancer Is Characterized by Altered Expression of Genes Involved in the Sumoylation Process. *J. Biol. Regul. Homeost. Agents.* 2015, 29(3), 655–662.
- [42] Guo, J.; Chen, D.; Gao, X.; Hu, X.; Zhou, Y.; Wu, C.; Wang, Y.; Chen, J.; Pei, R.; Chen, X. Protein Inhibitor of Activated STAT2 Restricts HCV Replication by Modulating Viral Proteins Degradation. *Viruses.* 2017, 9(10), 285. DOI: 10.3390/ v9100285.
- [43] Shalizi, A.; Bilimoria, P. M.; Stegmüller, J.; Gaudillière, B.; Yang, Y.; Shuai, K.; Bonni, A. PIASx Is A Mef2 Sumo E3 Ligase That Promotes Postsynaptic Dendritic Morphogenesis. *J. Neurosci.* 2007, 27(37), 10037–10046. DOI: 10.1523/JNEUROSCI.0361-07.2007.
- [44] Vavougios, G. D.; Zarogiannis, S. G.; Krogfelt, K. A.; Gourgoulianis, K.; Mitsikostas, D. D.; Hadjigeorgiou, G. Novel Candidate Genes of the PARK7 Interactome as Mediators of Apoptosis and Acetylation in Multiple Sclerosis: An in Silico Analysis. *Mult. Scler. Relat. Disord.* 2018, 19, 8–14. DOI: 10.1016/j.msard.2017.10.013.