

Non-invasive assessment of changes in liver fibrosis via liver stiffness measurement in patients with chronic hepatitis B: impact of antiviral treatment on fibrosis regression

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Abstract

Background Liver stiffness measurement (LSM) can assess liver fibrosis in patients with chronic hepatitis B (CHB). We evaluated whether LSM can be used to assess changes in liver fibrosis during antiviral treatment using nucleos(t)ide analogs in patients with CHB.

Methods We recruited 41 patients with CHB who had significant liver fibrosis, normal or slightly elevated serum alanine aminotransferase (ALT) levels ($\leq 2 \times$ upper limit of normal), and detectable serum hepatitis B virus DNA before antiviral treatment. Patients in Group 1 ($n = 23$) and Group 2 ($n = 18$) underwent follow-up LSM after antiviral treatment for 1 and 2 years, respectively.

Results The mean age, ALT and LSM value of all patients (34 men and 7 women) before antiviral treatment were

46.6 ± 9.5 years, 40.6 ± 17.2 IU/L and 12.9 ± 8.6 kPa, respectively. Hepatitis B e antigen (HBeAg) was detected in 31 patients (75.6%). Fibrosis stage was F2 in 12 (29.3%), F3 in 6 (14.6%) and F4 in 23 (56.1%) patients. After antiviral treatment, LSM values and DNA positivity decreased significantly as compared to baseline ($P = 0.018$ and $P < 0.001$ in Group 1; $P = 0.017$ and $P < 0.001$ in Group 2, respectively), whereas ALT levels were unchanged ($P = 0.063$ in Group 1; $P = 0.082$ in Group 2).

Conclusions Our preliminary data suggest that LSM can be used to assess liver fibrosis regression after antiviral treatment using nucleos(t)ide analogs in patients with CHB.

Keywords Alanine aminotransferase · Chronic hepatitis B · Liver fibrosis · Liver stiffness measurement · Nucleos(t)ide analog · Transient elastography

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Abbreviations

LB	Liver biopsy
HBV	Hepatitis B virus
HCV	Hepatitis C virus
CHB	Chronic hepatitis B
CHC	Chronic hepatitis C
CLD	Chronic liver disease
LSM	Liver stiffness measurement
ALT	Alanine aminotransferase
AST	Aspartate aminotransferase
ULN	Upper limit of normal
HBsAg	Hepatitis B surface antigen
HBeAg	Hepatitis B e antigen
kPa	Kilopascals
IQR	Interquartile range
API	Age–platelet count index

APRI	Aspartate aminotransferase to platelet ratio index
ASPRI	Age–spleen–platelet ratio index
SVR	Sustained viral response

Introduction

Chronic hepatitis B (CHB) is a chronic inflammatory condition that results in the formation of fibrous tissue and may lead to architectural distortion and persistent damage of the liver. Until recently, liver fibrosis and cirrhosis were generally considered to be irreversible [1, 2]. However, several studies using serial liver biopsies (LBs) have reported that several therapeutic interventions can produce histological improvement, including liver fibrosis regression [3–7]. Among these therapeutic interventions, nucleos(t)ide analogs that suppress hepatitis B virus (HBV) replication eliminated HBV DNA in 30–70% of cases and were significantly associated with liver fibrosis regression by reducing liver inflammation and cellular damage [8–11].

Because the prognosis and management of patients with CHB, as well as other chronic liver diseases (CLDs), depend strongly on the degree of liver fibrosis, the assessment of liver fibrosis regression is helpful to clinicians [12]. However, follow-up LB for confirming liver fibrosis regression during or after antiviral treatment is troublesome except consenting subjects. Therefore, a non-invasive method to assess liver fibrosis regression in patients with CHB who have undergone antiviral treatment would be highly useful.

Liver stiffness measurement (LSM) using FibroScan® allows the accurate assessment of liver fibrosis in patients with CHB and chronic hepatitis C (CHC) [13–15]. Although there have been several longitudinal investigations of patients with CHC [16–18], no longitudinal follow-up study has examined patients with CHB receiving antiviral treatment. Therefore, aims of our study were to evaluate whether LSM, rather than LB, can be used to assess changes in liver fibrosis during antiviral treatment using nucleos(t)ide analogs in patients with CHB and to compare the performance of LSM and other non-invasive methods in assessing liver fibrosis regression.

Patients and methods

Patients

In this retrospective cohort study, all data were collected from the database of Severance Hospital, Yonsei University

College of Medicine, Seoul, Korea. Between January and August 2007, we recruited 41 patients with CHB who showed significant liver fibrosis (\geq stage F2 according to the METAVIR scoring system) on LB, normal or slightly elevated serum alanine aminotransferase (ALT) levels [$<2 \times$ upper limit of normal (ULN)], detectable serum HBV DNA by hybridization capture assay before anti-viral treatment [19], and no ALT fluctuation ($\geq 2 \times$ ULN) during antiviral treatment. The study protocol conformed to the ethical guidelines of the 1975 Helsinki declaration and was approved by our independent institutional review board.

Patients with other CLDs, such as liver cancer, co-infection with hepatitis C virus (HCV), or human immunodeficiency virus and those with co-morbidities associated with HBV, such as non-alcoholic steatohepatitis, primary sclerosing cholangitis, or primary biliary cirrhosis, were excluded. Patients with a history of alcohol ingestion over 40 g/day for more than 5 years, previous liver resection surgery, liver transplantation, or evidence of cardiac or renal failure (defined as a serum creatinine levels ≥ 1.5 mg/dL) were also excluded. Patients with an unreliable LSM [<10 successful acquisitions, a success rate $<60\%$, an interquartile range (IQR) over median ratio lower than 30%, or measurement on a different day from LB; $n = 4$] or an unsuitable LB for fibrosis staging (<15 mm or 6 portal tracts; $n = 2$) were also excluded.

From the medical records, we collected data on age, gender, body mass index, liver histology and LSM values. The following laboratory parameters were also examined in all patients at the time of LB and LSM: ALT, total bilirubin, serum albumin, platelet count and prothrombin time (international normalized ratio). Hepatitis B surface antigen (HBsAg), HBeAg and antibodies were measured using standard enzyme-linked immunosorbent assays (Abbott Diagnostics, Abbott Park, IL, USA). HBV DNA levels were measured using a hybridization capture assay (Digene Diagnostics, Gaithersburg, MD, USA).

Patients in Group 1 ($n = 23$) and Group 2 ($n = 18$) underwent follow-up LSM and LB (optional) after antiviral treatment for 1 and 2 years, respectively. The mean interval from the starting day of antiviral treatment to follow-up LSM and LB was 12.2 ± 1.2 months in Group 1 and 24.4 ± 0.8 months in Group 2. Nucleos(t)ide analogs used for antiviral treatment included lamivudine ($n = 16$; 11 patients in Group 1 and 5 in Group 2), adefovir ($n = 10$; 5 in each group) and entecavir ($n = 15$; 7 in Group 1 and 8 in Group 2). The selection of a nucleos(t)ide analog was made after considering the availability of national insurance coverage for the antiviral agent, patient demand, or the decision of the attending clinician. The mean duration from the day of baseline LSM to the start of antiviral treatment was 5.7 ± 2.4 days.

Table 1 Calculation method of noninvasive model

Models	Calculation method
API	Age (years): <30 = 0, 30–39 = 1, 40–49 = 2, 50–59 = 3, 60–69 = 4, ≥70 = 5 Platelet count (10^9 L^{-1}): ≥225 = 0, 200–224 = 1, 175–199 = 2, 150–174 = 3; 125–149 = 4, <125 = 5 API is the sum of the above (possible value 0–10)
APRI	$[(\text{AST}/\text{ULN})/\text{platelet count } (10^9 \text{ L}^{-1})] \times 100$
SPRI	Spleen size (cm)/platelet count (10^9 L^{-1}) × 100
ASPRI	Age (years): <30 = 0, 30–39 = 1, 40–49 = 2, 50–59 = 3, 60–69 = 4, ≥70 = 5 ASPRI is the sum of age and SPRI

API age–platelet index [22], APRI aspartate aminotransferase to platelet ratio index [23], AST aspartate aminotransferase, ASPRI age–spleen–platelet ratio index [24]

Liver stiffness measurement

Liver stiffness measurement using FibroScan® was performed according to previously described methods [20, 21]. Briefly, an ultrasound transducer probe is mounted on the axis of a vibrator. Vibrations of mild amplitude (1 mm) and low frequency (50 Hz) are transmitted by the transducer, inducing an elastic shear wave that propagates through underlying tissues. Pulse-echo ultrasound acquisitions are used to follow the propagation of the shear wave and to measure its velocity, which is directly related to tissue stiffness (the elastic modulus): the stiffer the tissue, the faster the shear wave propagates. LSM measures liver stiffness in a volume that approximates a cylinder of 1-cm wide and 4-cm long, 25–65 mm below the skin surface. This volume is at least 100 times larger than that of a biopsy sample and is, therefore, far more representative of the hepatic parenchyma.

Ten valid measurements were performed on each patient. The success rate was calculated as the number of valid measurements divided by the total number of measurements. The results were expressed in kilopascals (kPa). IQR was defined as an index of intrinsic variability of LSM corresponding to the interval around LSM result containing 50% of the valid measurements between the 25th and 75th percentiles. The median value was considered representative of the elastic modulus of the liver. Only procedures with ten valid measurements, a success rate of at least 60% and an IQR over median ratio lower than 30% were considered reliable.

Other non-invasive methods of assessing liver fibrosis

The age–platelet index (API), aspartate aminotransferase (AST) to platelet ratio index (APRI) and age–spleen–platelet ratio index (ASPRI) were also evaluated for comparison with LSM results. The API, ARPI and ASPRI were calculated according to Poynard, Wai and Kim et al., respectively (Table 1) [22–24]. ALT level was measured using an automated chemistry analyzer (Hitachi 7600,

Tokyo, Japan). According to the manufacturer's instructions, the ULN for ALT was determined to be 40 IU/L.

Evaluation of liver histology

The indications for LB included an assessment of the severity of liver fibrosis and inflammation before antiviral treatment. LB was performed twice using 16-gauge gun biopsy sheathed cutting needle (spring loaded) (TSK Laboratory, Tokyo, Japan) on the right lobe of the liver under local anesthesia and ultrasound guidance. LB specimens were fixed in formalin and embedded in paraffin. 4-μm-thick sections were stained with hematoxylin and eosin and Masson's trichrome. All liver tissue samples were evaluated by an experienced hepatopathologist (YN Park) who was blinded to the patients' clinical histories.

Liver histology was evaluated according to the METAVIR scoring system [25]. Fibrosis was staged on the following scale of 0–4: F0, no fibrosis; F1, portal fibrosis without septa; F2, portal fibrosis and a few septa; F3, numerous septa without cirrhosis; and F4, cirrhosis. The mean length of the liver samples before antiviral treatment was 18.5 ± 1.6 mm.

Statistical analysis

Patient characteristics are presented as mean \pm standard deviation. The independent *t* test and Fisher's exact test were used to compare the baseline characteristics of Groups 1 and 2. The paired sample *t* test and McNemar test were used to compare the values pre- and post-antiviral treatment in each patient and to identify variables that showed a significant change during antiviral treatment. Two-way analysis of variance test was performed to identify the effects of each antiviral agent on the changes in LSM value during the antiviral treatment. A two-sided $P < 0.05$ was considered to be statistically significant. All statistical analyses were performed with SPSS 12.0 (SPSS, Chicago, IL, USA).

Table 2 Baseline characteristics at the time of LSM and LB

Variables	Total (n = 41)	Group 1 (n = 23)	Group 2 (n = 18)	P value
Male gender	34 (82.9)	19 (82.6)	15 (83.3)	NS
Age (years)	46.6 ± 9.5	45.9 ± 11.4	47.6 ± 6.5	NS
Body mass index (kg/m ²)	23.7 ± 2.5	23.7 ± 2.3	23.6 ± 2.8	NS
Platelet count (10 ⁹ L ⁻¹)	171 ± 52	170 ± 55	172 ± 51	NS
Albumin (g/dL)	4.3 ± 0.3	4.4 ± 0.2	4.2 ± 0.3	0.072
Total bilirubin (mg/dL)	0.8 ± 0.3	0.9 ± 0.4	0.8 ± 0.4	NS
ALT (IU/L)	40.6 ± 17.2	39.7 ± 15.0	41.9 ± 20.2	NS
HBeAg positivity	31 (75.6)	15 (65.2)	16 (88.9)	NS
HBV DNA positivity	41 (100.0)	23 (100.0)	18 (100.0)	NS
Prothrombin time (INR)	1.02 ± 0.09	1.03 ± 0.08	1.00 ± 0.10	NS
Fibrosis at initial biopsy				
F2	12 (29.3)	5 (21.7)	7 (38.9)	NS
F3	6 (14.6)	3 (13.1)	3 (16.7)	
F4	23 (56.1)	15 (65.2)	8 (44.4)	
LSM value (kPa)	12.9 ± 8.6	13.7 ± 8.0	12.1 ± 9.6	NS
Interquartile range (kPa)	1.6 ± 1.5	2.1 ± 1.7	1.9 ± 1.1	NS
Success rate (%)	97.4 ± 6.7	97.0 ± 8.4	98 ± 3.9	NS

Results

Baseline characteristics

The baseline clinical and histological characteristics of all patients at the time of LSM and LB are shown in Table 2. The mean age, ALT level and LSM value of all patients (34 men and 7 women) before antiviral treatment were 46.6 ± 9.5 years, 40.6 ± 17.2 IU/L and 12.9 ± 8.6 kPa, respectively. HBV DNA was detectable in all patients using a hybridization capture assay, and HBeAg was detected in 31 patients (75.6%) before antiviral treatment. The fibrosis stage was F2 in 12 (29.3%), F3 in 6 (14.6%) and F4 in 23 (56.1%) patients. There were no significantly different variables between Groups 1 and 2. Among variables, only fibrosis stage was significantly correlated to LSM values before antiviral treatment (Spearman correlation coefficient, $r = 0.576$ and $P = 0.041$, data not shown).

Comparison between the values at pre- and post-antiviral treatment

Pre- and post-antiviral treatment laboratory findings, LSM values and several non-invasive models for assessing liver fibrosis are summarized in Table 3. LSM values and HBV DNA positivity decreased significantly after antiviral treatment compared to baseline ($P = 0.018$ and $P < 0.001$ in Group 1; $P = 0.017$ and $P < 0.001$ in Group 2, respectively), whereas ALT levels were unchanged ($P = 0.063$ in Group 1, $P = 0.082$ in Group 2, respectively). The API,

Table 3 Comparison between pre- and post-antiviral treatment

	Group 1 (n = 23)		P value
	Pre	Post	
Body mass index (kg/m ²)	23.7 ± 2.3	23.5 ± 2.2	NS
ALT (IU/L)	39.6 ± 15.0	35.9 ± 14.0	0.063
Total bilirubin (mg/dL)	0.8 ± 0.3	0.85 ± 0.3	NS
Prothrombin time (INR)	1.03 ± 0.09	1.03 ± 0.06	NS
Platelet count (10 ⁹ L ⁻¹)	171 ± 20	173 ± 24	NS
LSM value (kPa)	13.7 ± 7.9	11.3 ± 5.3	0.018
	11.7 (4.1–36.3)	11.8 (3.8–20.9)	
HBV DNA positivity	23 (100)	1 (0.5)	<0.001
HBeAg positivity	15 (65.2)	15 (65.2)	NS
API	4.69 ± 2.26	4.55 ± 1.89	NS
APRI	0.85 ± 0.73	0.83 ± 0.80	NS
ASPRI	8.82 ± 3.74	8.80 ± 2.96	NS

APRI and ASPRI also failed to show a significant difference between pre- and post-antiviral treatment values. Variations in LSM value and ALT level are shown in Figs. 1 (Group 1) and 2 (Group 2).

Details of Group 2 patients who underwent follow-up LB

Follow-up LB was performed in four patients (nos. 8, 10, 12 and 13) in Group 2 (Table 4). ALT levels and LSM values tended to decrease, all patients were HBV DNA negative and one patient (no. 12) showed HBeAg seroconversion

Fig. 1 Variation in LSM values and ALT levels between pre- and post-antiviral treatment in patients with CHB who underwent follow-up LSM after 1 year of antiviral treatment using nucleos(t)ide analogs (Group 1). *LSM* liver stiffness measurement, *ALT* alanine aminotransferase, *LB* liver biopsy

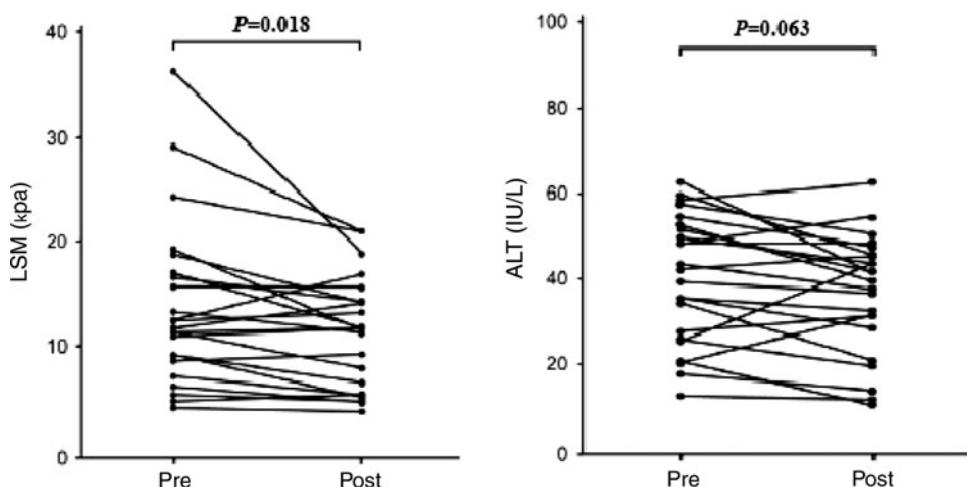
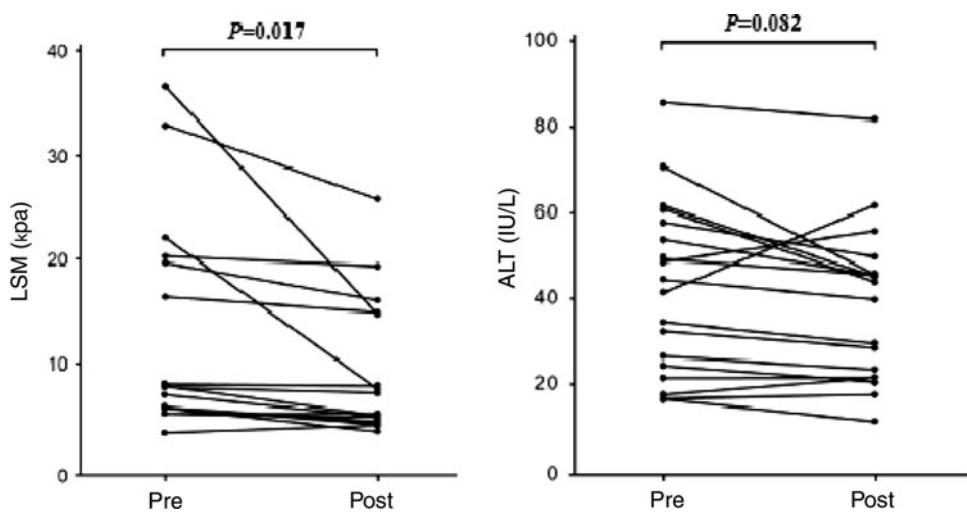


Fig. 2 Variation in LSM values and ALT levels between pre- and post-antiviral treatment in patients with CHB who underwent follow-up LSM after 2 years of antiviral treatment using nucleos(t)ide analogs (Group 2). *LSM* liver stiffness measurement, *ALT* alanine aminotransferase, *LB* liver biopsy



after antiviral treatment. Three patients (nos. 8, 10 and 13) showed liver fibrosis regression from F3 to F2 or from F2 to F1, whereas one patient (no. 12) showed no change in fibrosis stage. However, patient no. 12 showed slightly decreased portal fibrosis within the same fibrosis stage (F2) as compared to baseline LB. Figure 3 shows histological changes of patient no. 8 after 2 years of antiviral treatment.

Discussion

Because the degree of liver fibrosis determines prognosis and treatment strategy for patients with CHB and CHC [12], liver fibrosis regression during antiviral treatment can provide additional information.

Liver biopsy has been the ‘gold standard’ for assessing liver fibrosis, and is frequently performed as a baseline evaluation of liver fibrosis before antiviral treatment [26]. However, LB is rarely performed to confirm liver fibrosis regression during antiviral treatment, except in a few studies that investigated liver fibrosis regression during

antiviral treatment using serial LBs [27, 28]. Therefore, a non-invasive method to assess liver fibrosis regression during antiviral treatment would enable clinicians to re-evaluate prognoses and treatment strategies or to conduct comparative studies on the efficacy of antiviral agents.

Currently, several simple non-invasive serologic models for predicting liver fibrosis are available [22–24]. However, the ability of these models to predict liver fibrosis regression during antiviral treatment has not been investigated. According to our results, the available simple non-invasive serologic markers do not appear to be useful in detecting liver fibrosis regression during antiviral treatment when compared with LSM. This result can be explained in part by the confounding effects of extra-hepatic conditions and the superior performance to predict liver fibrosis of LSM has been confirmed in previous studies [20, 29]. Because other markers, such as hyaluronic acid and type IV collagen were not available in this retrospective study, further studies with these markers will be needed.

Recently, LSM has been accepted as a non-invasive tool for assessing liver fibrosis. However, most investigations of

Table 4 Laboratory and histologic findings of four patients in Group 2 who underwent follow-up liver biopsy

	Patient #8		Patient #10		Patient #12		Patient #13	
	Pre	Post	Pre	Post	Pre	Post	Pre	Post
Antiviral agent	Entecavir	Entecavir	Entecavir	Entecavir	Lamivudine			
ALT (IU/L)	32	17	57	65	26	25	60	56
HBV DNA (pg/mL)	3.2	<0.5	13.3	<0.5	3.4	<0.5	23.3	<0.5
HBeAg	Pos	Pos	Pos	Pos	Pos	Neg	Pos	Pos
Platelet count (10^9 L^{-1})	187	190	236	220	221	200	231	220
Serum albumin (g/dL)	4.0	4.5	4.6	5.0	4.3	4.1	4.3	4.2
Prothrombin time (INR)	1.18	0.97	0.85	0.98	0.99	0.89	0.80	0.92
API	4	5	3	4	4	4	2	4
APRI	0.41	0.40	0.31	0.35	0.29	0.30	0.41	0.48
ASPRI	8.42	9.40	6.39	6.30	6.85	6.80	5.68	6.48
LSM value (kPa)	7.9	4.8	6.7	4.5	6.0	4.9	7.8	6.2
Fibrosis grade	F3	F2	F2	F1	F2	F2	F2	F1

LSM conducted a one-time analysis using demographic and laboratory data obtained at the time of LB. Therefore, there is a chance that the dynamic changes in the biochemical tests before and after LB, especially ALT level, were not incorporated in the analysis. Several recent studies addressed this limitation by acquiring serial LSM values and concluded that ALT level has a significant impact on LSM [30–33]. Therefore, it has become more evident that LSM can be influenced by high ALT levels. Accordingly, a more recent study conducted in Hong Kong suggested that ALT level should be incorporated into the algorithms for diagnostic and treatment strategies in patients with CHB [34]. Therefore, failure to consider ALT levels may lead to less reliable or confusing results.

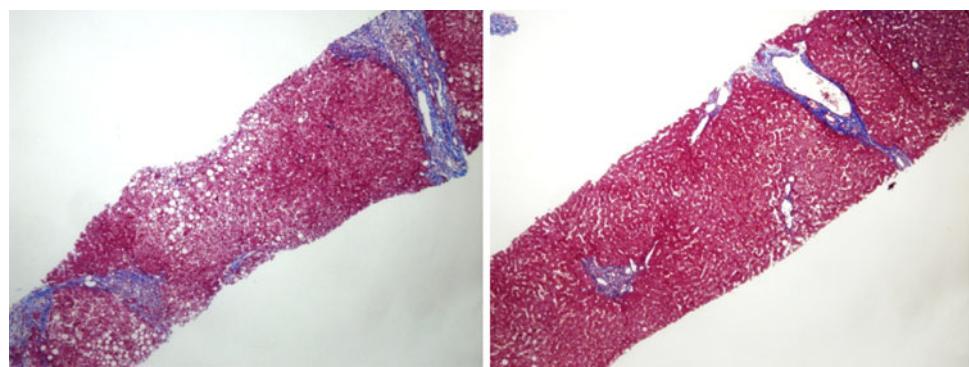
Among those who showed detectable HBV DNA in the hybridization capture assay and persistently normal or slightly elevated serum ALT levels ($<2 \times \text{ULN}$), a large proportion had significant histology on LB examination,

which is an indication for antiviral treatment in South Korea [29, 35]. Recruiting these patients enabled us to exclude the confounding effects of high ALT levels and specifically examine the performance of LSM in assessing liver fibrosis regression during antiviral treatment [30–33].

Several teams have investigated dynamic changes in LSM values during antiviral treatment in patients with CHC who had achieved SVR. However, there is no published longitudinal study for patients with CHB. De Ledinghen et al. [16] evaluated liver fibrosis regression using LSM and FibroTest in a very long-term follow-up of HCV responders and concluded that LSM and FibroTest are useful tools for the non-invasive evaluation of liver fibrosis and monitoring the histological response after antiviral treatment in HCV patients. Hezoda et al. [17] reported that LSM value was significantly, but modestly, reduced at the end of follow-up in patients with CHC who achieved SVR in response to peginterferon and alpha-ribavirin combination treatment. Colletao et al. [18] also investigated liver fibrosis among long-term responders (patients with CHC and CHB) to antiviral treatment, and concluded that LSM may reliably substitute for LB to confirm liver fibrosis regression. More recent study investigated the changes of LSM values during HCV treatment and concluded that irrespective of the virological response, treatment for HCV infection is associated with an improvement of LSM values [36].

In this study, we controlled the confounding effects of high ALT levels by recruiting patients with CHB showing persistently normal or minimally elevated ALT levels before antiviral treatment and persistent viral suppression after antiviral treatment. The reduction in ALT levels after antiviral treatment failed to show statistical significance ($P = 0.063$ in Group 1 and $P = 0.082$ in Group 2), whereas the reduction in LSM values predicted liver fibrosis regression significantly. These results were consistent when our study population was analyzed altogether considering small sample size (ALT, $P = 0.068$ and LSM, $P = 0.010$, data not shown). Furthermore, three patients (nos. 8, 10 and 13) in the 2 years of antiviral treatment group (Group 2)

Fig. 3 Regression of liver fibrosis from stage F3–F2 in patient no. 8. *Left* septal collagenous fibrosis of moderate thickness is present before antiviral treatment. *Right* almost all of the portal tracts reveal periportal fibrosis with occasional long slender fibrosis only (incipient septa) after 2 years of antiviral treatment (elastic trichrome, original magnification $\times 40$)



underwent optional LB; data from these procedures were used to confirm the performance of LSM in predicting liver fibrosis regression. A previous study reported that cirrhotic nodules appear to enlarge by expansion against septa as well as by lysis of septa and nodules surrounded by thin septa coalesce first, giving rise to macronodules or large regenerative nodules [3]. Accordingly, decreased LSM value after antiviral treatment in our current study implied fibrosis regression by virtue of lowered volume of total fibrosis in liver after long-term antiviral treatment.

Extrapolating our results, we might be able to consider LSM value at the time at which ALT level falls below $2 \times \text{ULN}$ after antiviral treatment as a baseline and use this value to predict subsequent liver fibrosis regression in patients with high ALT levels ($\geq 2 \times \text{ULN}$) before antiviral treatment. We previously showed that LSM values required additional time to normalize even after ALT levels had normalized in patients with acute hepatitis [37]. Therefore, if the interval between the decline in ALT level (to $< 2 \times \text{ULN}$) and the stabilization of LSM value can be validated in future investigations [38], LSM could also be used to predict liver fibrosis regression in patients with high ALT levels ($\geq 2 \times \text{ULN}$) before antiviral treatment.

An additional analysis to investigate the changes in ALT levels and LSM values according to each antiviral agent showed that ALT level did not change significantly in responses to any of the examined antiviral agents ($P = 0.186$ for lamivudine, $P = 0.129$ for adefovir and $P = 0.260$ for entecavir), whereas LSM decreased significantly in patients receiving adefovir and entecavir, but not lamivudine ($P = 0.026$, $P = 0.014$ and $P = 0.101$, respectively; data not shown). In addition, no significant difference was observed regarding the effects of each antiviral agent and HBeAg status on LSM value after antiviral treatment (all not significant; data not shown).

We are aware of several limitations in our study. First, sample size was small and there is no control group (i.e. no antiviral treatment), which is required to compare the change in LSM values from baseline to follow-up. Second, follow-up LB data are necessary for an exact comparison with follow-up LSM and original LB. However, follow-up LB was optional in this retrospective study and data were finally obtained for only four patients from Group 2. Third, we only recruited patients showing normal or slightly elevated ALT before antiviral treatment and no ALT fluctuation during antiviral treatment. Therefore, the clinical usefulness of LSM should be validated by further investigations which enroll patients with high ALT before antiviral treatment and those with fluctuating ALT during antiviral treatment and perform paired LB after long-term antiviral treatment.

In conclusion, our preliminary data showed that antiviral treatment in patients with CHB was associated with a

decrease in LSM values and suggest that LSM might be used to assess liver fibrosis regression after antiviral treatment using nucleos(t)ide analogs in patients with CHB who demonstrate ALT levels $< 2 \times \text{ULN}$ before antiviral treatment and persistent viral suppression during the course of antiviral treatment. However, large-scaled randomized prospective study with full paired LB and long-term follow-up will be needed in the near future to validate our conclusions.

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