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# The Geographic Origin of *Helicobacter pylori* Influences the Association of the *homB* Gene with Gastric Cancer

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We found that South Korean *Helicobacter pylori* isolates predominantly carry *homB* at locus B and that there is no association between the *homB* allele and the *cagA* allele or the development of gastric cancer within this population. Uniquely, several East Asian strains carried multiple copies of the *hom* genes.

lelicobacter pylori colonizes the gastric mucosas of over 50% of the world's population (6, 13) and is the etiological agent of gastritis, duodenal ulcers, gastric ulcers, gastric adenocarcinoma, and mucosa-associated lymphoid tissue lymphoma (4, 5, 19, 20, 22). Due to this bacterium's association with gastric cancer, which is the second most common cause of cancer-associated death (14), the World Health Organization has classified H. pylori as a class I carcinogen (8). Gastric cancer mortality rates vary geographically; the highest rates are in East Asian countries like China, Japan, and South Korea, which also display high rates of *H. pylori* infection (7, 8, 21, 23). Clearly, gastric diseases are due, at least in part, to infection by *H. pylori*, and the ultimate disease developed appears to be affected by variability in *H. pylori* virulence factors. Recently, we presented detailed epidemiological studies of cagA and vacA from a collection of 260 isolates from South Korea (9, 10, 11). Our studies showed that there is a significant association between infection with H. pylori strains carrying the EPIYA-ABD cagA genotype and the development of gastric cancer (11). Moreover, the majority of H. pylori isolates encoded the most virulent CagA (EPIYA-ABD) and VacA (s1/i1/m1) proteins (9, 11). The polymorphisms in cagA and vacA, alone and in concert, impact the progression to severe gastric disease, but the impact of these two virulence factors alone is not sufficient to explain the vast discrepancy in gastric cancer rates between East Asian and Western populations. Thus, it is important to examine the impacts of different virulence factors among both Western and East Asian populations (9, 10, 11, 16).

In vitro, Helicobacter outer membrane B (HomB) promotes the secretion of the proinflammatory cytokine interleukin-8 (IL-8) and increases *H. pylori*'s ability to adhere to host cells (15). More importantly, homB presence is significantly associated with development of peptic ulcer disease in Portuguese children and young adults (15, 18) and with gastric cancer development and the presence of cagA in U.S. and Colombian populations (12). These findings suggest that the outer membrane protein HomB is a novel virulence factor. Thus, it and other members of the small paralogous family of hom adhesion molecules are currently being investigated. The two best-studied hom genes, homA and homB, are 90% identical at the nucleotide level (2, 3, 18). These homA and homB genes can be present at two different loci within the H. pylori genome: locus A and locus B. Strains can carry a single copy of one of the hom genes, a double copy of a single gene, a single copy of

each gene, or neither gene (15, 16). Previous studies suggest geographic differences, either in distribution, location, or copy number, of the *hom* genes in the genome and suggest that these differences influence any association with disease outcome (15, 16, 17).

In the present study, our collection of 260 South Korean isolates was assessed for any associations between the distribution of the *homA* or *homB* genes and disease state, as well as any associations between the *hom* genes and the different *cagA* and *vacA* alleles. The South Korean isolates include 115 isolates from patients diagnosed with gastrictis, 60 isolates from patients diagnosed with gastric ulcers, 55 isolates from patients diagnosed with duodenal ulcers, and 30 isolates from patients diagnosed with gastric cancer (9, 10, 11). A complete description of all strains can be found in Table S1 in the supplemental material.

To analyze the *hom* genotype of the South Korean *H. pylori* strains, the presence of the *homA* and/or *homB* gene(s) was identified by a single PCR with the *hom* primers hf and hr (Table 1 and Fig. 1). We successfully genotyped 225 samples for which we had complete epidemiological data for the *hom* genes (Fig. 2; see also Table S2 in the supplemental material). Of note, two strains showed an amplicon with a length that was intermediate (approximately 146 bp) of what was expected for either the *homA* PCR product (~128 bp) or the *homB* PCR product (~161 bp). This intermediate-length *hom* or *ihom* genotype has been previously described and shown to be due to random deletions and/or insertions within the *hom* genes (12).

Once the strains were genotyped for the presence of the two *hom* genes, we next sought to define the copy numbers and locations of the genes. This was accomplished through two additional PCRs (Fig. 1). The distribution of *homA/homB* is shown in Fig. 2. Within this pop-

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TABLE 1 Primer sequences

Primer name	Primer sequence	Reference
hf (F1-jhp0870/jhp0649)	AGAGGGTGTTTGAAACGCTCAATA	18
hr (R1-jhp0870/jhp0649)	GGTGAATTCTTCTGCGGTTTG	18
Af (F1-jhp0648/HP0709)	TAATTTCGCGCAAAAACATC	18
Ar (R1-jhp0650/HP0711)	ATTCCAGCGCCTAATGGAC	18
Bf (F1-jhp0869/HP0935)	AAGAGGATTGCGTGGTGGAGTTG	18
Br (R1-jhp0871/HP0936)	GGGTTGCCTTTGGGCTTGGA	18
K-Af	TGGAATATTGATATAAAGAAGTG	This study
K-Ar	GGGTTTAATAGGATGAGCCGC	This study
K-Bf	GATTTTCCCCACTCTTTTTATGG	This study
K-Br	GGTTTTTGTCCATGAACATGC	This study

ulation, 212 isolates carried a single *hom* gene at locus B (35, —/*homA*; 175, —/*homB*; and 2, —/*ihom*). This is in contrast to Western strains that carry a single *hom* gene at locus A 100% of the time (15, 16). Also, this distribution is different from what has been reported for Western

strains, which show a much more evenly distributed population of isolates carrying *homA* or *homB* (15, 16). In our population, three isolates were indeterminate for the presence of the *homB* gene at locus A but were positive for occupation at locus B (*homB*<sup>+/-</sup>/*homB*). These 3 *homB*<sup>+/-</sup>/*homB* isolates were included as *homB*-positive strains for the statistical analysis but were eliminated from the data set when assessing the impact of multiple copies of the *hom* genes. Six strains carried multiple *hom* genes (4 *homA/homA*, 1 *homB/homB*, and 1 *homA/homB*), which is again in contrast to previous studies that suggested that Western strains carry multiple copies of *homA* and/or *homB* but East Asian strains do not (15, 16). Our finding is perhaps not surprising since our collection of East Asian isolates is much larger than the collection previously examined.

Finally, four strains failed to amplify either *hom* gene at either locus A or locus B and therefore were considered *hom* negative.

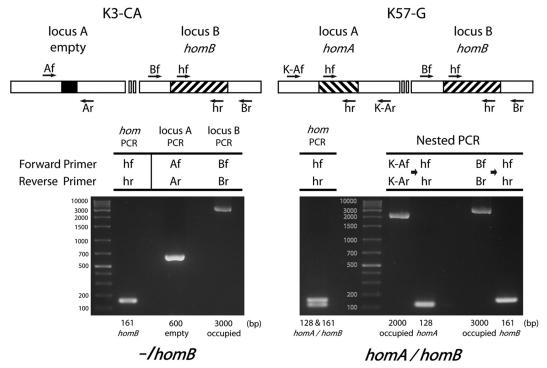


FIG 1 Genotyping of the hom genes at the respective loci. (Top) Schematic representation of the two loci where the hom genes are traditionally found, locus A and locus B. The annealing positions (arrows) and names of the primers used in this study are shown. The presence of a hom gene in a particular locus is depicted by the presence of a dashed box. (Bottom) The strains were genotyped for the hom gene by a single PCR with the hf and hr primers. A PCR amplicon of 128 bp indicates the presence of the homA gene, and an amplicon of 161 bp denotes the presence of the homB gene. In order to determine the location (locus A or B) of the hom gene, two additional PCRs were performed. To amplify locus A, primers Af and Ar were used, and to amplify locus B, the Bf and Br primers were used, as previously described (18). If an indeterminate result was obtained from the PCR using the Af and Ar primers or Bf and Br primers, another PCR with the K-Af and K-Ar primers or K-Bf and K-Br primers (Table 1) was performed. These K-Af/K-Ar and K-Bf/K-Br primers were designed according to the genome sequences of the Korean H. pylori strains HP51 and HP52 (GenBank accession numbers CP000012 and CP001680, respectively). For locus A, a resulting amplicon of 300 to 900 bp indicates that locus A is empty, whereas the presence of a 2,000- to 2,500-bp amplicon confirms that locus A is occupied by a hom gene. In the case of locus B, a 1,300- to 1,800-bp amplicon denotes that B is empty and the presence of a 2,500- to 4,000-bp amplicon indicates that locus B is occupied by a hom gene. (Left) For the K3-CA strain, the PCR with the hf and hr primers yielded a single amplicon of 161 bp (second lane), denoting the presence of the homB gene. The PCR with the Af and Ar primers (amplifying locus A) yielded a 600-bp product, indicating that locus A is empty (third lane), whereas a 3,000-bp amplicon produced from the PCR using the Bf and Br primers indicates that locus B is occupied (fourth lane). These results indicate that K3-CA has a genotype of -/homB. (Right) For K57-G and any strains that carried both homA and homB, an additional set of nested PCRs was also performed. First, PCRs with the hf and hr primers yielded two differently sized amplicons, a 128-bp amplicon, indicating the presence of a homA allele, and a 161-bp amplicon, indicating the presence of a homB allele (first lane). Next, a PCR using the Af and Ar primers or the K-Af and K-Ar primers yielded a 2,000-bp amplicon, which denotes an occupied locus. In the case of K57-G (right, third lane), the K-Af and K-Ar primers were used to amplify locus A because it showed no band for PCR using the Af/Ar primers. This PCR product was then purified and used as the template in a PCR using the hf and hr primers (fourth lane). This PCR yielded a 128-bp amplicon, which indicates that homA is located at locus A. Next, a PCR was performed using the Bf and Br primers, which yielded a 3,000-bp product (fifth lane). This again indicates an occupied locus B, and this PCR product was purified and used as the template in a PCR using the hf and hr primers. This PCR yielded a 161-bp amplicon (right, sixth lane), which indicates that homB is located at locus B. Thereby, these results indicate that K57-G has a genotype of homA/homB.

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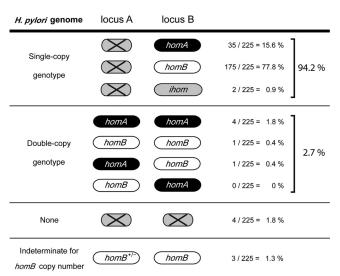


FIG 2 A schematic of the distribution of *hom* genes at the respective loci within this South Korean population is shown.  $homB^{+/-}$  indicates that the amplification of locus A was unsuccessful. Therefore, the strains are homB positive and there is at least a single copy of homB found at locus B, but it cannot be determined whether or not they have two copies of homB. These strains were included as homB-positive strains for the statistical analysis but were eliminated from the data set when assessing the impact of multiple copies of the hom genes. To the right of the schematic, the percentage of the overall population of each individual genotype is indicated.

For these four —/— strains, PCR amplification of locus A and locus B yielded products that were indicative of an empty locus. However, each of the four —/— strains indicated the presence of a hom gene through the hom PCR amplification: two had homA, one had ihom, and one had homA and homB. This suggests that for these strains, homA or homB is presumably located at an alternate unknown location within the genome. It is interesting to speculate that perhaps homA or homB may be carried in the normal locus for the virtually unstudied homC or homD gene. Further study is clearly necessary to elucidate whether the location of the homA/homB genes corresponds to a functional difference and whether homA/homB can be located at other locations besides locus A and locus B. Of note, no strains carrying homA/—, homB/—, or homB/homA alleles were found.

We next assessed whether there was an association between the distribution of the individual *hom* genes and the disease state. A complete breakdown of the relationship between the *hom* alleles and the disease state is provided in Tables S1 and S2 in the supplemental material. The Fisher exact test was used to analyze two-way associations with SAS version 9.1 software (SAS Institute Inc., Cary, NC). There was no association between the distribution of the *hom* gene and the disease state among this population of East Asian strains (P = 0.9978), a result which is in direct contrast to the association found in Western populations (12, 15). In fact, there was no association regardless of which disease states were assessed (Table 2).

Since there is a statistical association between the presence of *homB* and gastric cancer in Western populations, which carry the single *hom* gene at locus A, and not in East Asian populations, which carry the single *hom* gene at locus B, these data perhaps suggest that the location of the *hom* gene within the genome is important. Genes carried at one particular locus could be ex-

pressed at greater levels; the promoters of *homA* and *homB* may differ enough to influence transcriptional levels of each gene, or the different loci within the genome may provide the ability to bind and influence overall levels of the *hom* transcript for different enhancers/inhibitors of each *hom* gene.

Recently, it has become clear that individual virulence factors interact in order to impact H. pylori pathogenesis (1, 9, 10, 24). Since *homB* is associated with *cagA* within Western populations (12, 15), we next assessed the distribution of *hom* genes in combination with the *cagA* alleles, the *vacA* alleles, and the disease state. A complete breakdown of the strains based on these factors is provided in Table S3 in the supplemental material. We first assessed if there was any association between the distribution of the hom alleles and the different cagA alleles (a canonical EPIYA-ABD versus all other EPIYA motifs). We found that there was no association between the distribution of the hom alleles and the different cagA alleles (P = 0.0872). Furthermore, when each gene was considered separately, there was no association between the distribution of homA (P = 0.6139) or that of homB (P = 0.2217)across the different *cagA* alleles. We next analyzed the association between the distribution of the hom alleles and the different vacA alleles (see Table S3 in the supplemental material) and found that the distribution of vacA alleles among the two hom genes was statistically significant (P = 0.0142). This association was dependent only on the *homB* allele (P = 0.0275), since there was no association between the distribution of the homA allele and that of the *vacA* allele (P = 0.3955). The overall association between the vacA alleles and the hom alleles was also influenced by the distribution of cagA alleles; the association was present in the non-EPIYA-ABD population (P = 0.0319) but not in the EPIYA-ABD population (P = 0.1014). Due to this difference, we used loglinear modeling to determine if there was a three-way association between the *cagA*, *vacA*, and *hom* alleles. However, no association between these three virulence factors was identified (P = 0.681). Another interesting aspect of this *vacA/hom* association was that it appeared to be influenced by the age of the patient; the association only became evident in the population above 60 years of age (P =0.0076). A higher-order association between the cagA, vacA, and hom alleles and disease states was also assessed, but no significant associations existed between the virulence factors and the disease state.

TABLE 2 Significance of the associations between the distributions of *homA/homB* and the disease states

	<i>P</i> value for distribution of:		
Disease comparison	hom <sup>a</sup>	homA	homB
Across all diseases	0.9978	0.9802	0.9335
Gastritis vs all other diseases	0.8953	0.7215	0.7311
Duodenal ulcers vs all other diseases	1.0000	1.0000	1.0000
Gastric ulcers vs all other diseases	0.9219	1.0000	1.0000
Gastric cancer vs all other gastric diseases	0.8805	0.7931	0.6121
Peptic ulcers (both duodenal and gastric ulcers) vs gastritis and gastric cancer	1.0000	1.0000	1.0000
More severe diseases (gastric ulcer and gastric cancer) vs less severe diseases (gastritis and duodenal ulcer)	0.9379	0.8481	0.7146
Gastritis vs peptic ulcers vs gastric cancer	0.9803	0.9084	0.7894

<sup>&</sup>lt;sup>a</sup> hom represents the distribution of the homA, homB, and hom-negative strains for the different disease states listed.

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In conclusion, this is the first study to assess the association between the presence of the homB gene and gastric cancer in a population of predominantly East Asian strains; we found that the impact of the *homB* allele on disease is geographically dependent. In Western strains, there is a more even distribution of the homA and homB genes, while in East Asian strains, homB is more common (Fig. 2) (15, 16). Moreover, Western strains carry a single hom gene at locus A, whereas East Asian strains carry a single hom gene at locus B (Fig. 2) (15, 16). This study was the first to identify the presence of any East Asian isolates that carry multiple copies of the hom genes. Interestingly, in this population, no association between the presence of homB and the progression to gastric cancer was found (Table 2), suggesting that a hierarchy of virulence factors exists and that virulence factors have different impacts on disease based on the presence of other virulence factor polymorphisms. Within East Asian strains, EPIYA-ABD CagA appears to be the "master" virulence factor. En masse, these data exemplify the need for information about the presence and function of different virulence factors within different populations and the need to develop geographically tailored treatment regimens.

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