1 *Review*

2 Omega-3 PUFA metabolism and brain modifications during aging

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14 Abstract:

In Canada, 5.5 million (16% of Canadians) adults are >65 years old and projections suggest this 15 number will be approximately 20% of Canadians by 2024. A major concern regarding old age is a 16 decline in health, especially if this entails a loss of self-sufficiency and independence caused by a 17 decline in cognition. The brain contains 60% of fat and is one of the most concentrated organs in long 18 chain omega-3 fatty acids such as docosahexaenoic acid (DHA). During aging, there are physiological 19 modifications in the metabolism of lipids that could also have consequences on brain structure and 20 levels of DHA. This review will hence discuss the physiological modifications in the metabolism of 21 lipids during aging with a focus on long chain omega-3 and omega-6 fatty acids and also outline the 22 structural and functional modifications of the brain during aging including brain lipid modifications 23 24 and its relation to higher levels of DHA and cognition. Therefore, in this review, we outline the importance of collecting more data on the biology of aging since it might highly improve our 25 understanding about what are «normal» modifications occurring during aging and what can become 26 27 pathological.

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33 1. Introduction

34 Almost every country in the world experiences an aging population, and this population is expected to be 35 one of the most significant forces shaping our economy and society in the next 20-30 years. A major concern 36 about old age, both at the individual and societal levels, is a decline in health, especially if this means a loss of self-sufficiency and independence. Increasing research aimed at promoting healthy aging is actually 37 38 ongoing but one of the major hurdles is to define the biology of aging. Aging in humans refers to a 39 multidimensional process of physical, psychological, and social changes. Therefore, it follows that fundamental knowledge on the biological processes occurring during aging may help to design 40 environmental strategies aimed at promoting healthy biological aging. Thus, there is a need for better 41 prevention strategies, but one major gap in this field is a need to better understand what the biological 42 modifications are, also called geroscience, since this field is relatively new. One of the strategies to promote 43 healthy aging is the consumption of one or two fish meals each week ¹⁻³. Normally, the intake of fish 44 positively correlates with increased plasma and erythrocyte omega-3 fatty acids (n-3 FA), likely with 45 eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) concentrations in a time- and dose-46 dependent manner⁴⁻⁶. EPA and DHA have to be provided through the diet because their synthesis from their 47 precursor alpha-linolenic acid (ALA) is extremely limited in humans ⁷. However, over the 20th century, the 48 dietary fat consumption has drastically changed with an increased level of omega-6 fatty acids such as 49 linoleic acid (LA) from 2.79% to 7.21% of energy. This shift in our dietary fat intake was largely due to our 50 dependence on new food production methodologies, including soybean oil⁸. 51

The link between our dietary fat intake and the incidence of chronic diseases has been largely debated over 52 the last 20 years. Our research group is mainly focused on prevention of cognitive decline, so the focus of 53 this review paper, with respect to chronic diseases, will be on cognition. This link between dietary fat intake 54 and the risk of cognitive decline has been the focus of many review papers overs the last 10-15 years ⁹⁻¹¹. 55 One of the most recent reviews supports a positive association between dietary and blood n-6: n-3 ratio and 56 cognitive decline and incidence of dementia, as evaluated on 14 human studies including 7 prospective 57 studies ¹². A recent meta-analysis on 11 cohort studies evaluated the association between 299 metabolites 58 and general cognitive ability and dementia. They reported that higher DHA levels in blood were associated 59 with higher cognitive function in 22,887 individuals ¹³. Hence, it seems that more elevated concentration of 60 n-3 FA in the blood is associated with lower cognitive decline and perhaps lower risk of other chronic 61 diseases. However, our group showed that for older participants, plasma EPA and DHA kinetics are 62 dysregulated and this will likely lower the capacity of older adults to incorporate EPA and DHA in organs 63

and tissues. Usually, a fish oil supplementation increases the level of EPA and DHA in the plasma or 64 erythrocytes but in those aged >70 years old, we don't know whether this process is efficient. There is no 65 clear definition or parameters to define an old vs. a young participant. Most of the studies used the median 66 of age in their participants group or a continuous age ae range. Following from the information summarized 67 above, this paper will review some of the metabolism modifications occurring during aging with a focus on 68 lipid metabolism. By reviewing these evidences, we will also expose how these modifications might limit 69 incorporations of n-3 FA in membranes of cells with a focus on the brain because it is one of the most 70 71 enriched organs in DHA.

72 2. Lipid and fatty acid metabolism differences during aging

Generally speaking, there are differences in the lipid and fatty acid metabolism occurring during aging and these modifications are considered totally normal and part of the aging process. These processes include the transport of fatty acids after their intake and their transit to the different organs and tissues that are modified during aging. This section will review some of these modifications.

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2.1.Normal transport of fatty acids from dietary intake to their circulation in the blood:

In Western adults, the diet is composed of 30 to 40% of lipids, of which 92 to 96% are long chain 78 79 fatty acid esterified to a glycerol thus constituting what is the main form of dietary lipid: triglycerides 80 (TG) ¹⁴. Whole-body homeostasis requires fine-tuning of fatty acid transport and utilization by metabolically active tissues ¹⁵. Because of their regulatory roles in cellular fatty acid uptake and 81 utilization, membrane apolipoprotein receptors and fatty acid transporters form an integral part of this 82 homeostatic system. As a result, imbalances in lipid metabolism likely will influence the functioning 83 of fatty acid transporters and their protein levels. Lipids are not soluble in water and necessitate 84 incorporation into amphiphilic molecules called lipoproteins to circulate in the blood. Hence, 85 following ingestion of TG, they will be hydrolysed at their ester bonds by gastric and pancreatic 86 lipases into two non-esterified fatty acids (NEFA) and one monoacylglycerol (MAG) with the fatty 87 acid being in the Sn-2 position ¹⁶. Both forms of lipids are passively transported into enterocytes ¹⁷ via 88 diffusion or transporters such as "Fatty Acid Transport Proteins" (FATPs) and "Cluster of 89 Differentiation 36" (CD36)¹⁸. Dietary lipids are efficiently digested and absorbed by the enterocytes 90 19. 91

92 Once inside the intestinal cells, enzymes convert the NEFAs and MAG back into TG ²⁰. These 93 will be integrated in chylomicrons and exported to the lymphatic system via the Golgi apparatus ²¹.

The chylomicrons, now rich in exogenous triglycerides, join the bloodstream via the thoracic duct and 94 get transported to the peripheral tissues such as muscle and fat cells. In the bloodstream, lipoprotein 95 lipase (LPL) gets activated when it detects an apolipoprotein C II (apoC-II)²² on the surface of the 96 chylomicrons. The role of lipoprotein lipase is to hydrolyse the ester bonds of TGs in chylomicrons ²² 97 to release NEFAs into the bloodstream where there will be an uptake by nearby cells. The loss of TGs 98 will result in a decrease in size of chylomicrons and leave chylomicrons constituents available for the 99 synthesis of native HDL disks ²³. Remnant chylomicrons rich in cholesteryl esters will be captured by 100 endocytosis by hepatocytes receptors such as LDL receptor (LDLR)²² and LDL receptor-related 101 (https://onlinelibrary-wiley-(LRP) 102 protein

103 com.ezproxy.usherbrooke.ca/doi/abs/10.1002/%28SICI%291096-

9136%28199708%2914%3A3%2B%3CS75%3A%3AAID-DIA449%3E3.0.CO%3B2-9). The liver can then use 104 the endogenous TG and cholesteryl esters to form the very low density lipoprotein (VLDL)²⁴. These 105 lipoproteins will be directed to peripheral tissues. Following a loss of TG, there will be a decrease in 106 VLDL density ²⁵. With the action of lipoprotein lipase, VLDL will then become intermediate density 107 lipoprotein (IDL). With the action of hepatic lipase ²⁵ IDL becomes low density lipoprotein (LDL). 108 LDLs carry cholesterol to tissues ²⁶. LDL will be captured by their receptor, LDLR which are found 109 on cell membranes, where it will be eliminated from the bloodstream by endocytosis ²⁶. LDL 110 cholesterol will be recovered in the cell. An excess of cholesterol in the tissues will cause an inhibition 111 of transcription of the genes responsible for the formation of the LDLR ²⁷. It thus reduces the uptake 112 of LDL by the cells and these LDLs will remain in circulation. The remaining LDLs in the circulation 113 are more likely to be oxidized ²⁸ which will thereafter contribute to the development of atherosclerotic 114 plaque²⁸. 115

116 **2.2** Lipoprotein metabolism modification during aging

During aging, the metabolism of lipids is modified and causes an increase of plasma lipids. For instance, the fasting plasma levels of VLDL, TG, LDL and cholesterol ²⁹ are significantly higher in the elderly ³⁰. <u>Higher levels of lipid and cholesterol can be the source of many health problems such as cardiovascular</u> <u>disease and diabetes (REF =http://diabetes.diabetesjournals.org/content/46/8/1354.full-text.pdf +</u>

121 <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4587882/).</u>

These plasma lipid changes in the elderly can cause an increase in plasma free fatty acid levels (<u>https://eds-</u>
b-ebscohost-com.ezproxy.usherbrooke.ca/eds/pdfviewer/pdfviewer?vid=1&sid=3f462f39-8acd-4cbf-

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b113-b378e62a44c1%40sessionmgr103). Increasing plasma FFA may result in increased plasma glucose 124 by decreasing glucose uptake into the cells. The enzymes responsible for the oxidative cascade of GLA are 125 intimately related to that of glycolysis. Thus, increased lipid oxidation inhibits glucose metabolism, 126 uptake in cells, and impairs 127 decreases glucose glycogen storage http://diabetes.diabetesjournals.org/content/diabetes/37/6/667.full.pdf.. This promotes hyperinsulinemia 128 ultimate insulin resistance 129 and https://eds-b-ebscohostcom.ezproxy.usherbrooke.ca/eds/pdfviewer/pdfviewer?vid=1&sid=3f462f39-8acd-4cbf-b113-130

131 <u>b378e62a44c1%40sessionmgr103</u>.

Insulin resistance, often seen in the elderly (https://www.jci.org/articles/view/110908/pdf), will also cause 132 an increase in VLDL and blood triglycerides. Insulin resistance impairs the metabolism of chylomicrons, 133 VLDL, LDL and HDL ³¹ since a lack of insulin or a lower sensitivity to insulin will reduce the catabolism 134 of chylomicrons and VLDL by LPL. During aging, there is also a higher level of LDL which remains 135 transient for a longer period of time in the plasma (Einarsson K, Nilsell K, Leijd B, Angelin B. Influence of 136 age on secretion of cholesterol and synthesis of bile acids by the liver. N Engl J Med 1985;313(5):277-82. 137 doi: 10.1056/NEJM198508013130501.) as a reference of this statement. = REF 30). In the long term, these 138 LDLs are more likely to be oxidized ³¹. The higher concentration of VLDL and chylomicrons in addition to 139 oxidized LDL accumulation in older insulin-resistant individuals would increase the risk of developing 140 cardiovascular disease (CVD) ³². Furthermore, the increase of LDL may be due to the diminution of bile 141 synthesis from cholesterol by the liver during aging ^{30, 33}. The decrease in bile acid synthesis is due to the 142 decrease in the expression of "cholesterol 7-alpha hydroxylase" (CYP7A1) during aging. This cytochrome 143 is one of the CYP450 and regulates the formation of bile acids ³⁴. This causes a decrease in the use of 144 cholesterol by the liver as well as a reduction in LDLR expression with age. Thus, plasma LDL will have 145 lower clearance with age resulting in an increase in plasma LDL concentration in the elderly ²⁹. In the end, 146 147 it is possible that deregulation of LDL in the elderly is due to several different phenomena stemming from the large amount of change that occurs with age. The decrease in LDL in the elderly has shown a reduction 148 in the incidence of CVD ³⁵. In particular, a study showed that long chain polyunsaturated fatty acids 149 (PUFAs) allowed an increase in LDLR expression ³⁶, which could increase the clearance rate of plasma 150 LDL in the elderly and reduce the incidence of CVD. These are some of the modification of the lipid 151 metabolism occurring during aging. Overall, there are usually higher TG and LDL levels in the blood of 152 153 older adults and it is important to consider these modifications in the prevention of chronic diseases but also when interpreting results pertaining to fatty acid metabolism. 154

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157 **2.3 Omega-3 fatty acid metabolism during aging**

Over the last 10 years, our group has worked on omega-3 metabolism with a focus on modifications that occur during aging. This section will report the evidence of omega-3 fatty acid metabolism in three different conditions: before supplementation with omega-3, during or after supplementation with omega-3 fatty acid, and kinetics studies using uniformly labeled carbon 13 fatty acids (¹³C-).

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2.3.1 Without an omega-3 fatty acid supplementation

163 To our knowledge, there are ~ 24 studies that have reported the level of omega-3 fatty acids or the omega-3 index in young versus old adults (Table 1). Most of the studies reported the fatty acid profile 164 165 in red blood cells or in plasma/serum phospholipids (PL). Among the 24 studies, 7 studies reported the omega-3 index only and showed that it was higher in older participants ³⁷⁻⁴³. Two studies on the 166 omega-3 index reported an increase of about 5-7% of the omega-3 index every decade ^{37, 41}. Eleven 167 studies reported the fatty acid profile in red blood cells (RBC) ^{37, 38, 41-50}. For most of the studies, it is 168 difficult to compare the results since the data were not expressed the same way. For instance, two 169 studies reported that the participants having the highest level of omega-3 were on average 8-10 years 170 older than those with the lowest omega-3 fatty acid levels in erythrocytes ^{43, 44}. Other studies reported 171 the level of increase in omega-3 fatty acids for each increasing decade. Hence, it is difficult to draw a 172 clear conclusion for the omega-3 fatty acid results in RBC but it appears that at older ages, there is 173 174 more omega-3 in RBC. It is important to note that these papers did not include a complete fatty acid profile of the RBC as it was recently recommended in a paper describing the best practices for the 175 design, laboratory analysis and reporting of clinical trials involving fatty acids ⁵¹, hence limiting 176 comparisons between studies. With respect to plasma/serum PL, there were eight studies ^{45, 52-58}. Six 177 of these studies reported on average a 1.5 fold higher level of DHA in the plasma PL of older 178 participants, aged between 50-88 years old compared with younger participants, aged between 20-49 179 years old ^{45, 53-57}. One study reported a 2 fold higher level of EPA in plasma PL but there were no 180 difference between ages for DHA ⁵². Yet another study reported only a positive correlation between 181 age and EPA+DHA in plasma PL but it was not possible to quantify the magnitude of the difference 182 between young and older adults ⁵⁸. Overall, there is generally good evidence supporting the idea that 183 during aging, the relative % of omega-3 fatty acids or its concentration in RBC and plasma/serum are 184

higher in the oldest participants compared to the youngest. Some of the proposed mechanism includes a reduction of omega-3 fatty acids in cell membranes, higher intestinal absorption during aging, higher availability and release of adipose tissue stocks. Hence, the exact mechanism behind this higher level of blood omega-3 in older individuals might be multi-level but the important point here is that they might be associated to longevity.

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2.3.2 With an omega-3 fatty acid supplementation

To our knowledge, there are nine published studies specifically addressing EPA and DHA responses to an 191 omega-3 fatty acid supplement with participants of different ages (Table 2). Supplementation doses range 192 from 300 mg/d to more than 4 g/d and lasted between 6 weeks and twelve months. Seven studies evaluated 193 the fatty acid profile in the plasma whereas one study evaluated the fatty acid profile in erythrocytes only ⁵⁹ 194 and another did so in platelets and adipose tissues only ⁶⁰. One study reported the omega-3 index pre- and 195 post-supplementation ⁵⁹ and showed that a low omega-3 index at baseline and an older age predicted those 196 with a greater increase of the omega-3 index after supplementation ⁵⁹. This study had similar results to 197 Vandal et al., ⁶¹ which showed that the oldest had a higher increase in DHA compared to the youngest after 198 the supplementation, but in their study, Vandal had similar DHA levels in young and old participants at 199 baseline. 200

The other studies investigated the plasma level of omega-3 fatty acids. One study reported that older 201 participants had higher omega-3 levels at baseline but after the supplementation, the increase was similar in 202 both groups ⁶². The six other studies reported a higher increase in EPA ⁶³⁻⁶⁷ and/or DHA ⁶⁸ in older 203 participants compared to younger. The exact mechanism explaining this effect is unclear. Most of the studies 204 reported that it is unlikely that the age-related differences in EPA and DHA at baseline are due to differences 205 206 in intake of omega-3 PUFA with age. Rather it seems to be related to age differences in endogenous production and incorporation of EPA and DHA due to hormones and hormone sensitivity, body 207 composition, and physical activity, all of which change with age ⁶⁷. The study of Walker et al. also showed 208 that the adipose tissue stores less DHA with age in response to EPA + DHA supplementation, hence 209 210 suggesting that age-related differences in the handling and storage of exogenous supplied DHA may be related to impaired insulin sensitivity with aging or to differences in body composition with aging ⁶⁷. The 211 adipose tissue represents a significant store of EPA and DHA, containing the equivalent of several hundred 212 days of the fatty acid content of a typical diet. Altogether, these results support that providing a supplement 213 214 of omega-3 fatty acid to older adults increases their blood levels when compared to younger individuals. These results may be caused by the fact that older individuals have shown to be more compliant to treatments 215

than younger people (REF = https://onlinelibrary.wiley.com/doi/full/10.1046/j.1365-2710.2000.00315.x),
 causing a higher level of omega-3 in their blood. But despite that fact, those results brings into question
 whether this type of supplementation is useful to them in the prevention of chronic diseases since they may
 not be able to use it. Another important point is that it might also be due to their lower turnover of circulating
 TG, hence contributing to their higher omega-3 levels, since omega-3 fatty acid levels are esterified in TG.
 To answer some of these questions, employing ¹³C-fatty acids is useful.

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2.3.3 Using ¹³C-fatty acid to evaluate their kinetics during aging

Tracing metabolism of ¹³C-fatty acids may provide some insight into possible age-related changes in 223 fatty acid metabolism in humans. Metabolism of ¹³C-DHA has been investigated in humans ⁶⁹⁻⁷¹. In 224 young adults given an oral dose of 250-280 mg ¹³C-DHA, ¹³C enrichment peaked at 2 h post-dose in 225 plasma triglycerides when the tracer was given in the triglyceride form, but at 6 h post-dose when the 226 tracer was esterified to phosphatidylcholine ^{69,71}. Brossard *et al.* have reported a 1.4% apparent retro-227 conversion of ¹³C-DHA to ¹³C-docosapentaenoate (22:5 omega-3) and ¹³C-EPA 3 d after giving the 228 tracer ⁷⁰. These first results showed the feasibility of tracing DHA metabolism in humans. However, 229 neither the impact of aging on ¹³C-DHA metabolism nor its β -oxidation were investigated, although 230 both may influence the somewhat higher blood levels of EPA and DHA commonly seen in healthy 231 elderly ^{54, 65, 66, 68, 72}. Our group are pioneers in this field as we investigated the kinetics of ¹³C-DHA 232 in six young and six elderly participants ⁷³. We found that, in the elderly, ¹³C-DHA was 4 times higher 233 in plasma triglycerides and NEFA at 4 h post-dose, β -oxidation was 1.9 times higher, whereas 234 apparent retro-conversion of ¹³C-DHA to other ¹³C-omega-3 fatty acids was 2.1 times higher 24 h and 235 7 d after tracer intake compared to the young adults ⁷³. Hence, because DHA seems to remain 236 transiently for longer periods of time in the blood of the elderly compared to the young, it may thus 237 238 indicate that efficiency to remove DHA from the blood is lower in the elderly than in the young, resulting in lower incorporation of DHA in the membrane of cells that serve to initiate signalization 239 ^{65, 66, 68, 72}. This result is consistent with the transient slower metabolism of TG and LDL in older as 240 compared to young adults and this was described in a previous section. 241

Our most recent work with tracers between old and young men was conducted with ¹³C-EPA or arachidonic acid (¹³C-ARA), two key fatty acids that are precursors of anti- and pro-inflammatory cytokines, respectively. Surprisingly, the kinetics of ¹³C-EPA and ¹³C-ARA was quite similar between young and old men despite a time x age interaction for ¹³C-EPA kinetics where the postprandial shape

of the curve was steeper in old vs young men ⁷⁴. One intriguing result we obtained was that in old 246 247 men, synthesis of DHA from EPA started 2 h after tracer intake whereas it was delayed to 1 d in young 248 men. This result suggests that old adults might need more DHA than what was actually provided in their diet compared to the young men. However, newly synthesized DHA accumulated in the plasma 249 of old men for 7 d and this might be because it remains for a longer period in the plasma as suggested 250 by our previous study with ¹³C-DHA. Therefore, there might be a defect in old adults to uptake DHA 251 in the tissues. We also calculated that plasma half-life of ¹³C-EPA was 2 d whereas that of ¹³C-ARA 252 was 4 d, similar to that of DHA. DHA and ARA are the two most concentrated long chain 253 polyunsaturated fatty acids in brain membranes. With our β -oxidation measures using breath samples, 254 we calculated ¹³C-EPA whole-body half-life to be \sim 14 days in old men whereas in the younger group 255 it was \sim 21 days ⁷⁴. This result indicates that older adults turn over EPA \sim 7 days faster than the younger 256 adults. This is an intriguing result since epidemiological studies and results we obtained in previous 257 studies ^{62, 65} support that old adults have twice as much plasma EPA, hence one would anticipate a 258 lower whole-body turnover in old vs young adults. Therefore, it seems that there is somehow a 259 260 disconnect between plasma levels of EPA and perhaps DHA and their kinetics, thus more studies are 261 needed to understand the mechanism of these modifications and their possible consequences such as 262 potential higher risk of cognitive decline.

3 Brain modifications during aging:

The brain is composed of 60% fat with one third of its content being ARA and DHA. The brain is hence the second most rich tissue in fat after adipose tissue. The brain fatty acids are however mostly PLs unlike the adipose tissue that is mainly composed of TGs. Because DHA is an important constituent of brain structure, there has been much interest in the association between the level of DHA in brain membranes, brain function and brain volume and losses during aging. Therefore, this section will summarize the evidence about morphological, functional, and content modifications of the brain during aging and whether dietary omega-3 intake can improve brain structure and function.

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3.1. Morphological modifications of the brain during aging

There are a number of morphological modifications of the brain that occur during aging. Several studies have indicated that brain volume decreases over the course of the human lifespan. A review conducted by Hedman et al. ⁷⁵ compiled the results of 56 longitudinal magnetic resonance imaging (MRI) studies on whole brain volumes in healthy individuals and concluded that the rate of total brain volume loss

is not constant throughout aging. For instance, the rate of brain volume loss after 35 years of age is approximately 0.2% per year. Between 35 and 60 years of age, the volume loss rate slowly increases to 0.5% followed by a steady volume loss of over 0.5% per year over 60 years of age ⁷⁵. Furthermore, other studies have indicated that volume loss in the whole brain is greater in males than in females ^{76, 77}.

Several studies demonstrate a reduction of gray matter volume during aging ⁷⁸⁻⁸⁴. More specifically, 280 the volume of gray matter in the cortex and the cerebellum of older individuals is 18% and 13% smaller, 281 respectively, than those of their younger counterparts⁸¹. There is also a significant loss of gray matter in the 282 frontal, limbic, temporal, and parietal lobes but not in the occipital lobe ^{78, 83}. Similarly, studies have also 283 indicated that there is a decrease of white matter volume in the brains of older individuals ^{81, 85-87}. According 284 to Jäncke et al.⁸¹, there is a decrease in white matter volume in the cortex and cerebellum of older individuals 285 by 5% and ~9%, respectively, compared to younger adults. Moreover, one study indicated that the rate of 286 decrease of white matter is not constant during aging ⁸⁷. Instead, white matter volume slowly increases 287 before the age of 40, peaks at approximately 50 years of age, and then quickly decreases after the age of 60 288 ⁸⁷. As well, white matter hyperintensity lesions increase in size with age in the frontal, temporal, and parietal 289 lobes but not in the occipital lobes ⁸⁶. 290

291 In addition to age-related changes in the volume of the whole brain, gray matter, and white matter, there are also differences in the volume of specific brain structures. There seems to be a general decrease in the 292 volume of the following brain structures in older individuals compared to younger individuals: cerebral 293 hemisphere ⁷⁶, frontal lobe ^{88, 89}, parietal lobe ^{77, 88, 89}, temporal lobe ^{88, 89}, thalamus ^{81, 90}, basal ganglia ⁸⁹, 294 and the cerebellum ⁸⁹. Notably, there is atrophy of the hippocampus during aging ^{77, 81, 91-93}. A meta-analysis 295 by Fraser et al. ⁹³ detailed hippocampal atrophy rates according to 28 studies. They determined that the 296 overall rate of atrophy for the entire sample was 0.85% per year ⁹³. However, the rate of hippocampal 297 298 atrophy reported in the studies differed based on mean age of the participants: rate of atrophy was 0.38% per year in studies with a mean age of 55, 0.98% per year for a mean age of 55 to 70 years, and 1.12% per 299 year for a mean age of greater than 70 years. In contrast to the aforementioned structures, the ventricles of 300 the brain increase in volume during aging ^{76, 91}. Altogether, there is generally good evidence supporting loss 301 of matter in many brain structures, including loss in white and gray matter. These losses of brain matter can 302 contribute to lower cognitive functions during aging. 303

304 3.2 Modification of brain functions during aging

In addition to the many structural changes that occur during aging, brain functions are also modified during 305 306 this period. For instance, there is an age-related decrease in glucose metabolism in the whole brain and the frontal, parietal, and temporal lobes as well as in Broca's and Wernicke's areas ⁷⁷. It also seems that brain 307 activation during the execution of motor functions is modified in older adults. For example, there is a 308 309 decrease in blood-oxygen level dependent (BOLD) signals in multiple brain regions (sensorimotor cortex, cerebellum and thalamus) of older adults during mastication and an increase in BOLD signal in the 310 prefrontal area ⁹⁴. Another study showed that classical motor coordination regions were activated during 311 complex inter-limb coordination tasks, but that there was also increased activation of higher-level 312 sensorimotor and frontal regions in older individuals ⁹⁵. Similarly, other studies have demonstrated that the 313 performance of motor tasks result in increased activation of additional brain areas such as the basal ganglia, 314 prefrontal cortex, precuneus, and the cerebellum ⁹⁶⁻⁹⁸ in older invividuals. 315

Moreover, cognitive functions are modified as a result of changes in the volume of various brain structures. 316 317 For instance, a meta-analysis of 57 publications from the years 1984 to 1998 concluded that white matter hyperintensities are linked with poorer performance on cognitive tests for processing speed, immediate and 318 delayed memory, executive functions, and global cognitive functioning ⁹⁹. Further, a decrease in the 319 thalamus volume in older individuals is associated with attenuated performance on tests assessing cognitive 320 speed ⁹⁰. An additional meta-analysis of 33 studies concluded that larger prefrontal cortex volume and 321 thickness is correlated with better executive functioning ¹⁰⁰. In regard to hippocampus volume and memory, 322 Van Petten¹⁰¹ reported in a meta-analysis of 33 studies that the positive correlation between hippocampus 323 size and episodic memory in older adults was weaker than expected. However, a more recent study 324 demonstrated that smaller hippocampus size is significantly associated with lower performance in episodic 325 memory, working memory, processing speed, and executive function tasks ¹⁰². Similarly to motor function, 326 it has been shown that older adults recruit additional brain regions during memory tasks ¹⁰³⁻¹⁰⁵. 327

Aging is also associated with changes in the activity of brain structures involved in sensation and perception. For instance, there are less areas activated in older versus younger adults in response to various odors ¹⁰⁶. A meta-analysis of 105 studies concluded that the activation of the fusiform gyrus, cerebellum, and hippocampus is elevated in elderly versus younger individuals during the processing of emotional faces ¹⁰⁷. Moreover, older individuals had greater activation of the prefrontal cortex during more difficult perceptual tasks compared to younger individuals ¹⁰⁸. The brains of older adults are also less responsive to blue light stimulation compared to younger adults ¹⁰⁹.

More recent studies have shed light on the changes that occur in the functional neural networks of the brain. 335 336 It seems that aging is associated with weaker connectivity in long-range connections and stronger connectivity of short-range connections ^{110, 111}. Elderly individuals also have less intra-network and greater 337 inter-network connectivity ^{112, 113}. More specifically, older individuals have less connectivity within the 338 default mode network (DMN) and somatomotor network ¹¹³, as well as greater connectivity between the 339 salience network and the executive control network (ECN) and the DMN ¹¹². Moreover, age seems to shift 340 dynamic functional connectivity from posterior to anterior regions, which is also reflected in the decreased 341 activation of posterior regions during the decline of episodic memory in older individuals ¹¹⁴. 342

Overall, there are several morphological and functional modifications within the brain during aging and understanding how these modifications manifest could be helpful to limit the rate at which these declines occur.

346 3.3 Modifications of brain content during aging

347 The number of studies, particularly those that use neuroimaging techniques, that have evaluated the change in human brain content during aging is limited. Post-mortem examinations of the human brain have 348 indicated that there is a change in protein and lipid content during aging. With regard to protein, one study 349 indicated that there is a 5-15% decrease in total protein content of the brain between 30 and 90 years of age 350 ¹¹⁵. A decrease in protein content in the substantia nigra, hippocampus, caudate nucleus, and gray matter 351 has also been reported ^{116, 117}. However, Söderberg et al. ¹¹⁶ found that protein content remained unchanged 352 in the cerebellum, pons, and medulla oblongata of older individuals. Similarly, a number of post-mortem 353 354 studies have demonstrated changes in the lipid content of older brains. For instance, Svenerholm et al. ¹¹⁸ reported that there is a linear decrease in cholesterol and phospholipids in the frontal and temporal cortices 355 and a curvilinear decrease in cholesterol, PLs, cerebrosides, and sulfatides in frontal and temporal white 356 matter between the ages of 20 and 100. In terms of PLs, Söderberg et al. ¹¹⁶ found that they were relatively 357 unchanged during aging with only a 5-10% decrease in the oldest age group. A more recent study conducted 358 by Hancock et al.¹¹⁹ reported that PL content in the entorhinal cortex of older individuals is relatively stable 359 during aging, but there is an increase in mitochondrial phosphatidylcholine (PC) and a decrease in 360 mitochondrial phosphatidylethanolamine (PE). The same group reported that age is associated with an 361 increase in mitochondrial PE containing DHA, but said the increase is not large enough to increase total 362 DHA in the mitochondria. Norris et al. ¹²⁰ examined phospholipid composition in the dorsolateral prefrontal 363 cortex in individuals aged 20-100 years. They found that there is a general age-related increase in 364 phospholipids containing DHA and decrease in PLs containing ARA and docosatetraenoic acid ¹²⁰. 365

A recent study used positron emission tomography to assess the incorporation of DHA from plasma to the 366 brain using carbon-11 ([1-C¹¹])-DHA in apolipoprotein E epsilon 4 allele (APOE4) carriers versus non-367 carriers ¹²¹. APOE4 is the most important genetic risk of late-onset Alzheimer's disease ¹²². Yassine et al. 368 found that the mean global gray matter DHA incorporation coefficient was 16% higher in APOE4 carriers 369 vs non-carriers ¹²¹. A higher DHA incorporation coefficient was also observed in other regions including 370 the entorhinal cortex ¹²¹. However, the whole-brain DHA incorporation rate was not significantly different 371 between APOE groups ¹²¹. They also did not observe any age-related effects on DHA incorporation, but this 372 may be due to the fact that only 4 of their 23 participants were over 50 years old ¹²¹. The authors 373 hypothesized that increased DHA incorporation in the brains of APOE4 carriers could be a compensatory 374 mechanism to counteract brain DHA loss ¹²¹. Our group also documented that the metabolism of DHA is 375 imbalanced in APOE4 carriers ¹²³⁻¹²⁶ and that they are perhaps more vulnerable to DHA deficiency ¹²⁷. 376

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3.4 Does omega-3 fatty acid consumption improve brain structure and function?

378 There are a number of studies that have examined the relationship between omega-3 fatty acid consumption and brain structure and function. For instance, Gu et al. ¹²⁸ evaluated the link between white matter integrity 379 and dietary nutrient intake in 239 elderly participants. They assessed white matter integrity using fractional 380 anisotropy measured by diffusion tensor imaging (DTI). They found that the nutrient pattern characterized 381 by high consumption of omega-3 and omega-6 fatty acids and vitamin E was positively correlated with 382 fractional anisotropy which corresponds to better white matter integrity ¹²⁸. Another group examined the 383 relationship between dietary fish consumption and brain structural integrity in 260 cognitively normal adults 384 aged 65 years or older ¹²⁹. Fish intake was measured using the National Cancer Institute Food Frequency 385 Questionnaire and the gray matter volume of various brain regions was measured with MRI¹²⁹. They found 386 387 that eating baked or broiled fish weekly is positively associated with higher gray matter volume in several brain regions, including the hippocampus, posterior cingulate, precuneus, and the orbital frontal cortex ¹²⁹. 388 Samieri and colleagues ¹³⁰ evaluated the association between plasma EPA and DHA concentrations and 389 390 gray matter atrophy in the medial temporal lobe in 281 individuals aged 65 years or older. The authors 391 compared fatty acid plasma concentrations at baseline to the results of MRI examinations from baseline and four years after baseline ¹³⁰. They observed that greater levels of plasma EPA was associated with lower 392 atrophy of the gray matter of the right amygdala and the hippocampal/parahippocampal region; this same 393 association was not observed for plasma DHA levels ¹³⁰, which is counterintuitive. Samieri et al. ¹³⁰ also 394 found that increased amygdala gray matter atrophy was linked with more depressive symptoms and poorer 395 semantic memory performances compared to baseline. Lastly, Witte et al. ¹³¹ assessed the connection 396

between fish oil supplement consumption and brain structure and function in 65 participants aged 50 to 75 397 398 years. Participants consumed either fish oil, which contained 2.2 grams of omega-3 fatty acids, or a placebo 399 daily for 26 weeks. Neuropsychological testing and MRI examinations were performed before and after the intervention period. The investigators found that after the 26-week intervention period, the fish oil group 400 had better white matter structural integrity in selective white matter tracts in the frontal, temporal, parietal, 401 and limbic areas ¹³¹. They also observed that the fish oil group had significant increases in gray matter 402 volume in the left hippocampus, precuneus, the superior temporal, inferior parietal and postcentral gyri, and 403 in the right middle temporal gyrus ¹³¹. In terms of performance on cognitive measures, they found that the 404 fish oil group had an improvement of 26% on executive function scores compared to no improvement in the 405 placebo group ¹³¹. In addition, they found a positive correlation between verbal fluency scores and EPA 406 percentage in red blood cell membranes in the fish oil group after intervention ¹³¹. Although for many years 407 408 it was thought that an intake of fish throughout life protects against cognitive decline, the recent evidence 409 suggests that fish intake might not be required throughout life to improve brain structure and function. Hence, starting an EPA+DHA supplementation after 50 years old might benefit older individuals with 410 411 respect to prevention of brain volume and function losses.

412 4 Are we ready for updated recommendations on dietary omega-3 fatty acids intake during aging?

In this review paper, we have outlined that there are many physiological modifications occurring during 413 aging with respect to lipid metabolism and brain volume and function losses and that an omega-3 fatty 414 acid intake might help to support the brain throughout aging. It is important to note that life expectancy 415 416 is longer, which means that older adults may live longer with their chronic diseases. A major concern 417 regarding old age is a decline in health, especially if this entails a loss of self-sufficiency and independence caused by a decline in cognition. A decline in working memory appears to be one of the 418 major consequences of normal aging ^{132, 133}. As outlined in the previous sections, the brain undergoes 419 physiological change during aging. While age is one risk of cognitive decline, this multifactorial disease 420 is also increased by a complex interaction between both genetic and environmental risk factors ¹³⁴⁻¹³⁶. 421

We believe nutrition has a role to play in the prevention of cognitive decline but nutrition alone might not be as efficient as a multidomain intervention. Recent evidence from the FINGER trials ¹³⁷ reported that combining physical exercise, personalized nutritional recommendations to avoid nutrient deficiencies, controlling cardiovascular risks and having cognitive stimulation prevented cognitive decline. However, they recently refocus their message by showing that dietary changes initiated early in the intervention was the most influential for global cognition improvement over two years of follow-up

¹³⁸. Therefore, nutrition might have a key role to play in the prevention of cognitive decline. In the case 428 429 of the FINGER study, dietary recommendations were not focussed on the consumption of fish oil but 430 were either focused to alleviate nutritional deficiency including low blood levels of DHA. It also has to be emphasized that there is currently no drug to prevent, cure or delay the progression of dementia and 431 that some pharmaceutical companies have shut down their research laboratories in this area. Therefore, 432 prevention strategies are currently the most efficient means since once the disease process has started, 433 there is no available drug for limiting its progression. However, there is one group in Canada working 434 on a nutritional strategy, a ketogenic beverage. They reported that a ketogenic beverage increases brain 435 energy metabolism in Alzheimer's patients ^{139, 140}. 436

Returning to the question of if we are ready to change recommendations on omega-3 fatty acids, we think that we are not there yet. However, working on the biology of aging might greatly improve our understanding about what are «normal» modifications occurring during aging and what can become pathological. Seizing this opportunity, we might contribute to the prevention of cognitive decline in the future with nutrition playing a vital role in this process.

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450 **Conflicts of Interest**

451 MP received funding from Neptune Wellness Solutions but the funding did not funded studies cited in 452 this review and did not funded any of the work of this review.

453 **References**

454 1. Barberger-Gateau P, Letenneur L, Deschamps V, Peres K, Dartigues JF, Renaud S. Fish, meat, and risk of 455 dementia: cohort study. Bmj. 2002;325(7370):932-3.

456 2. Holub DJ, Holub BJ. Omega-3 fatty acids from fish oils and cardiovascular disease. Mol Cell Biochem.
457 2004;263(1-2):217-25.

458 3. Morris MC, Evans DA, Tangney CC, Bienias JL, Wilson RS. Fish consumption and cognitive decline with age 459 in a large community study. Arch Neurol. 2005;62(12):1849-53.

460 4. Arterburn LM, Hall EB, Oken H. Distribution, interconversion, and dose response of n-3 fatty acids in 461 humans. Am J Clin Nutr. 2006;83(6):S1467-76.

Vidgren HM, Agren JJ, Schwab U, Rissanen T, Hanninen O, Uusitupa MI. Incorporation of n-3 fatty acids
into plasma lipid fractions, and erythrocyte membranes and platelets during dietary supplementation with fish,
fish oil, and docosahexaenoic acid-rich oil among healthy young men. Lipids. 1997;32(7):697-705.

Calder PC. Polyunsaturated fatty acids and inflammation. Prostaglandins Leukot Essent Fatty Acids.
2006;75(3):197-202. Epub 2006/07/11.

Plourde M, Cunnane SC. Extremely limited synthesis of long chain polyunsaturates in adults: Implications
for their dietary essentiality and use as suppements. Appl Physiol Nutr Metab. 2007;32(4):619-34.

8. Blasbalg TL, Hibbeln JR, Ramsden CE, Majchrzak SF, Rawlings RR. Changes in consumption of omega-3 and
omega-6 fatty acids in the United States during the 20th century. The American Journal of Clinical Nutrition.
2011;93(5):950-62.

472 9. Cunnane SC, Plourde M, Pifferi F, Begin M, Feart C, Barberger-Gateau P. Fish, docosahexaenoic acid and
473 Alzheimer's disease. Prog Lipid Res. 2009;48(5):239-56. Epub 2009/04/14.

474 10. Salem N, Jr., Vandal M, Calon F. The benefit of docosahexaenoic acid for the adult brain in aging and
475 dementia. Prostaglandins Leukot Essent Fatty Acids. 2015;92:15-22.

476 11. Barberger-Gateau P, Samieri C, Feart C, Plourde M. Dietary omega 3 polyunsaturated fatty acids and
477 Alzheimer's disease: interaction with apolipoprotein E genotype. Curr Alzheimer Res. 2011;8(5):479-91.

- 12. Loef M, Walach H. The Omega-6/Omega-3 Ratio and Dementia or Cognitive Decline: A Systematic Review
- on Human Studies and Biological Evidence. Journal of Nutrition in Gerontology and Geriatrics. 2013;32(1):1-23.

van der Lee SJ, Teunissen CE, Pool R, Shipley MJ, Teumer A, Chouraki V, et al. Circulating metabolites and
general cognitive ability and dementia: Evidence from 11 cohort studies. Alzheimer's & dementia : the journal of
the Alzheimer's Association. 2018;14(6):707-22. Epub 2018/01/10.

483 14. Mc Auley MT, Mooney KM. Computationally Modeling Lipid Metabolism and Aging: A Mini-review.
484 Computational and structural biotechnology journal. 2015;13:38-46. Epub 2015/03/10.

Schwenk RW, Holloway GP, Luiken JJ, Bonen A, Glatz JF. Fatty acid transport across the cell membrane:
regulation by fatty acid transporters. Prostaglandins, leukotrienes, and essential fatty acids. 2010;82(4-6):149-54.
Epub 2010/03/09.

488 16. Mu H, Hoy CE. The digestion of dietary triacylglycerols. Prog Lipid Res. 2004;43(2):105-33.

489 17. Mattson FH, Volpenhein RA. The Digestion and Absorption of Triglycerides. J Biol Chem. 1964;239:2772-7.

490 18. Kunisaki S, C. M. Ultrasound growth patterns of fetal lung malformations: Implications on prenatal care
 491 and postnatal outcome. Prenat Diagn. 2015;35(24):89-90.

492 19. D'Aquila T, Hung YH, Carreiro A, Buhman KK. Recent discoveries on absorption of dietary fat: Presence,
493 synthesis, and metabolism of cytoplasmic lipid droplets within enterocytes. Biochimica et biophysica acta.
494 2016;1861(8 Pt A):730-47. Epub 2016/04/25.

495 20. Bisgaier CL, Glickman RM. Intestinal synthesis, secretion, and transport of lipoproteins. Annu Rev Physiol.
496 1983;45:625-36.

497 21. Mansbach CM, 2nd, Nevin P. Intracellular movement of triacylglycerols in the intestine. J Lipid Res.
498 1998;39(5):963-8.

499 22. Cooper AD. Hepatic uptake of chylomicron remnants. J Lipid Res. 1997;38(11):2173-92.

50023. Redgrave TG, Small DM. Quantitation of the transfer of surface phospholipid of chylomicrons to the high501density lipoprotein fraction during the catabolism of chylomicrons in the rat. J Clin Invest. 1979;64(1):162-71.

502 24. Gruffat D, Durand D, Graulet B, Bauchart D. Regulation of VLDL synthesis and secretion in the liver. Reprod
 503 Nutr Dev. 1996;36(4):375-89.

504 25. Havel RJ. The formation of LDL: mechanisms and regulation. J Lipid Res. 1984;25(13):1570-6.

505 26. Brown MS, Goldstein JL. A receptor-mediated pathway for cholesterol homeostasis. Science.
506 1986;232(4746):34-47.

50727.Zhang Y, Ma KL, Ruan XZ, Liu BC. Dysregulation of the Low-Density Lipoprotein Receptor Pathway Is508Involved in Lipid Disorder-Mediated Organ Injury. Int J Biol Sci. 2016;12(5):569-79.

509 28. B. B. Stress oxydant et pathologies cardiovasculaires. . Médecine Thérapeutique Cardiol. . 2006;2(1):43-52.

Fortier M, Tremblay-Mercier J, Plourde M, Chouinard-Watkins R, Vandal M, Pifferi F, et al. Higher plasma
 n-3 fatty acid status in the moderately healthy elderly in southern Quebec: higher fish intake or aging-related
 change in n-3 fatty acid metabolism? Prostaglandins, leukotrienes, and essential fatty acids. 2010;82(4-6):277-80.

513 Epub 2010/03/09.

514 30. Einarsson K, Nilsell K, Leijd B, Angelin B. Influence of age on secretion of cholesterol and synthesis of bile 515 acids by the liver. The New England journal of medicine. 1985;313(5):277-82. Epub 1985/08/01.

516 31. Verges B. Pathophysiology of diabetic dyslipidaemia: where are we? Diabetologia. 2015;58(5):886-99.

Austin MA, Hokanson JE, Edwards KL. Hypertriglyceridemia as a cardiovascular risk factor. Am J Cardiol.
1998;81(4A):7B-12B.

519 33. Ericsson S, Berglund L, Frostegard J, Einarsson K, Angelin B. The influence of age on low density lipoprotein
520 metabolism: effects of cholestyramine treatment in young and old healthy male subjects. J Intern Med.
521 1997;242(4):329-37.

522 34. Russell DW, Setchell KD. Bile acid biosynthesis. Biochemistry. 1992;31(20):4737-49.

52335.Grundy SM, Cleeman JI, Rifkind BM, Kuller LH. Cholesterol lowering in the elderly population. Coordinating524Committee of the National Cholesterol Education Program. Arch Intern Med. 1999;159(15):1670-8.

525 36. Fernandez ML, West KL. Mechanisms by which dietary fatty acids modulate plasma lipids. J Nutr. 526 2005;135(9):2075-8.

527 37. Block RC, Harris WS, Pottala JV. Determinants of Blood Cell Omega-3 Fatty Acid Content. The open 528 biomarkers journal. 2008;1:1-6. 529 38. Gellert S, Schuchardt JP, Hahn A. Low long chain omega-3 fatty acid status in middle-aged women. 530 Prostaglandins Leukot Essent Fatty Acids. 2017;117:54-9. 531 39. Harris WS, Luo J, Pottala JV, Espeland MA, Margolis KL, Manson JE, et al. Red blood cell polyunsaturated fatty acids and mortality in the Women's Health Initiative Memory Study. J Clin Lipidol. 2017;11(1):250-9 e5. 532 533 40. Harris WS, Pottala JV, Lacey SM, Vasan RS, Larson MG, Robins SJ. Clinical correlates and heritability of 534 erythrocyte eicosapentaenoic and docosahexaenoic acid content in the Framingham Heart Study. Atherosclerosis. 535 2012;225(2):425-31. 536 Harris WS, Pottala JV, Varvel SA, Borowski JJ, Ward JN, McConnell JP. Erythrocyte omega-3 fatty acids 41. 537 increase and linoleic acid decreases with age: observations from 160,000 patients. Prostaglandins Leukot Essent 538 Fatty Acids. 2013;88(4):257-63. 539 42. Sands SA, Reid KJ, Windsor SL, Harris WS. The impact of age, body mass index, and fish intake on the EPA 540 and DHA content of human erythrocytes. Lipids. 2005;40(4):343-7. 541 43. Aarsetoey H, Ponitz V, Grundt H, Staines H, Harris WS, Nilsen DW. (n-3) Fatty acid content of red blood 542 cells does not predict risk of future cardiovascular events following an acute coronary syndrome. J Nutr. 543 2009;139(3):507-13. 544 44. Block RC, Harris WS, Reid KJ, Sands SA, Spertus JA. EPA and DHA in blood cell membranes from acute 545 coronary syndrome patients and controls. Atherosclerosis. 2008;197(2):821-8. 546 45. Caprari P, Scuteri A, Salvati AM, Bauco C, Cantafora A, Masella R, et al. Aging and red blood cell 547 membrane: a study of centenarians. Exp Gerontol. 1999;34(1):47-57. 548 Carver JD, Benford VJ, Han B, Cantor AB. The relationship between age and the fatty acid composition of 46. 549 cerebral cortex and erythrocytes in human subjects. Brain Res Bull. 2001;56(2):79-85. 550 47. Farzaneh-Far R, Harris WS, Garg S, Na B, Whooley MA. Inverse association of erythrocyte n-3 fatty acid 551 levels with inflammatory biomarkers in patients with stable coronary artery disease: The Heart and Soul Study. 552 Atherosclerosis. 2009;205(2):538-43. 553 48. Itomura M, Fujioka S, Hamazaki K, Kobayashi K, Nagasawa T, Sawazaki S, et al. Factors influencing 554 EPA+DHA levels in red blood cells in Japan. In vivo. 2008;22(1):131-5. 555 Thorlaksdottir AY, Skuladottir GV, Petursdottir AL, Tryggvadottir L, Ogmundsdottir HM, Eyfjord JE, et al. 49. 556 Positive association between plasma antioxidant capacity and n-3 PUFA in red blood cells from women. Lipids. 557 2006;41(2):119-25. 558 50. Yanagisawa N, Shimada K, Miyazaki T, Kume A, Kitamura Y, Ichikawa R, et al. Polyunsaturated fatty acid 559 levels of serum and red blood cells in apparently healthy Japanese subjects living in an urban area. Journal of 560 atherosclerosis and thrombosis. 2010;17(3):285-94. 561 51. Brenna JT, Plourde M, Stark KD, Jones PJ, Lin YH. Best practices for the design, laboratory analysis, and 562 reporting of trials involving fatty acids. Am J Clin Nutr. 2018;108(2):211-27. 563 52. Asciutti-Moura LS, Guilland JC, Fuchs F, Richard D, Klepping J. Fatty acid composition of serum lipids and its 564 relation to diet in an elderly institutionalized population. Am J Clin Nutr. 1988;48(4):980-7. Crowe FL, Skeaff CM, Green TJ, Gray AR. Serum n-3 long-chain PUFA differ by sex and age in a population-565 53. 566 based survey of New Zealand adolescents and adults. Br J Nutr. 2008;99(1):168-74. 54. 567 de Groot RH, van Boxtel MP, Schiepers OJ, Hornstra G, Jolles J. Age dependence of plasma phospholipid 568 fatty acid levels: potential role of linoleic acid in the age-associated increase in docosahexaenoic acid and 569 eicosapentaenoic acid concentrations. Br J Nutr. 2009;102(7):1058-64. 570 Dewailly E, Blanchet C, Gingras S, Lemieux S, Holub BJ. Cardiovascular disease risk factors and n-3 fatty 55. 571 acid status in the adult population of James Bay Cree. Am J Clin Nutr. 2002;76(1):85-92. 572 56. Dewailly E, Blanchet C, Lemieux S, Sauve L, Gingras S, Ayotte P, et al. n-3 Fatty acids and cardiovascular 573 disease risk factors among the Inuit of Nunavik. Am J Clin Nutr. 2001;74(4):464-73. 574 Dewailly EE, Blanchet C, Gingras S, Lemieux S, Sauve L, Bergeron J, et al. Relations between n-3 fatty acid 57. 575 status and cardiovascular disease risk factors among Quebecers. Am J Clin Nutr. 2001;74(5):603-11. 576 Ogura T, Takada H, Okuno M, Kitade H, Matsuura T, Kwon M, et al. Fatty acid composition of plasma, 58. 577 erythrocytes and adipose: their correlations and effects of age and sex. Lipids. 2010;45(2):137-44.

578

59.

579 erythrocyte omega-3 fatty acid content in response to fish oil supplementation: a dose-response randomized 580 controlled trial. Journal of the American Heart Association. 2013;2(6):e000513. 581 Witt PM, Christensen JH, Ewertz M, Aardestrup IV, Schmidt EB. The incorporation of marine n-3 PUFA into 60. 582 platelets and adipose tissue in pre- and postmenopausal women: a randomised, double-blind, placebo-controlled 583 trial. Br J Nutr. 2010;104(3):318-25. 584 Vandal M, Freemantle E, Tremblay-Mercier J, Plourde M, Fortier M, Bruneau J, et al. Plasma omega-3 fatty 61. acid response to a fish oil supplement in the healthy elderly. Lipids. 2008;43(11):1085-9. 585 586 62. Fortier M, Tremblay-Mercier J, Plourde M, Chouinard-Watkins R, Vandal M, Pifferi F, et al. Higher plasma 587 n-3 fatty acid status in the moderately healthy elderly in southern Quebec: higher fish intake or aging-related 588 change in n-3 fatty acid metabolism? Prostaglandins Leukot Essent Fatty Acids. 2010;82(4-6):277-80. 589 63. Meydani M, Natiello F, Goldin B, Free N, Woods M, Schaefer E, et al. Effect of long-term fish oil 590 supplementation on vitamin E status and lipid peroxidation in women. J Nutr. 1991;121(4):484-91. 591 Meydani SN, Endres S, Woods MM, Goldin BR, Soo C, Morrill-Labrode A, et al. Oral (n-3) fatty acid 64. 592 supplementation suppresses cytokine production and lymphocyte proliferation: comparison between young and 593 older women. J Nutr. 1991;121(4):547-55. 594 Plourde M, Tremblay-Mercier J, Fortier M, Pifferi F, Cunnane SC. Eicosapentaenoic acid decreases 65. 595 postprandial beta-hydroxybutyrate and free fatty acid responses in healthy young and elderly. Nutrition. 596 2009;25(3):289-94. 597 Rees D, Miles EA, Banerjee T, Wells SJ, Roynette CE, Wahle KW, et al. Dose-related effects of 66. 598 eicosapentaenoic acid on innate immune function in healthy humans: a comparison of young and older men. Am J 599 Clin Nutr. 2006;83(2):331-42. 600 67. Walker CG, Browning LM, Mander AP, Madden J, West AL, Calder PC, et al. Age and sex differences in the 601 incorporation of EPA and DHA into plasma fractions, cells and adipose tissue in humans. Br J Nutr. 2014;111(4):679-89. 602 603 68. Vandal M, Freemantle E, Tremblay-Mercier J, Plourde M, Fortier M, Bruneau J, et al. Plasma omega-3 fatty 604 acid response to a fish oil supplement in the healthy elderly. Lipids. 2008;43(11):1085-9. 605 69. Brossard N, Croset M, Normand S, Pousin J, Lecerf J, Laville M, et al. Human plasma albumin transports 606 [13C]docosahexaenoic acid in two lipid forms to blood cells. J Lipid Res. 1997;38(8):1571-82. 607 Brossard N, Croset M, Pachiaudi C, Riou JP, Tayot JL, Lagarde M. Retroconversion and metabolism of 70. 608 [13C]22:6n-3 in humans and rats after intake of a single dose of [13C]22:6n-3-triacylglycerols. Am J Clin Nutr. 609 1996;64(4):577-86. 610 71. Lemaitre-Delaunay D, Pachiaudi C, Laville M, Pousin J, Armstrong M, Lagarde M. Blood compartmental 611 metabolism of docosahexaenoic acid (DHA) in humans after ingestion of a single dose of [(¹³)C]DHA in 612 phosphatidylcholine. J Lipid Res. 1999;40(10):1867-74. 613 72. Fortier M, Tremblay-Mercier J, Plourde M, Chouinard-Watkins R, Vandal M, Pifferi F, et al. Higher plasma n-3 fatty acid status in the moderately healthy elderly in southern Quebec: higher fish intake or aging-related 614 615 change in n-3 fatty acid metabolism? Prostaglandins Leukot Essent Fatty Acids. ;82(4-6):277-80. Epub 2010/03/09. 616 73. Plourde M, Chouinard-Watkins R, Vandal M, Zhang Y, Lawrence P, Brenna JT, et al. Plasma incorporation, 617 apparent retroconversion and beta-oxidation of 13C-docosahexaenoic acid in the elderly. Nutrition & metabolism. 618 2011;8:5. Epub 2011/01/29. 619 Leveille P, Chouinard-Watkins R, Windust A, Lawrence P, Cunnane SC, Brenna JT, et al. Metabolism of 74. 620 uniformly labeled 13C-eicosapentaenoic acid and 13C-arachidonic acid in young and old men. Am J Clin Nutr. 621 2017. 622 75. Hedman AM, van Haren NE, Schnack HG, Kahn RS, Hulshoff Pol HE. Human brain changes across the life 623 span: a review of 56 longitudinal magnetic resonance imaging studies. Hum Brain Mapp. 2012;33(8):1987-2002. 624 76. Coffey CE, Lucke JF, Saxton JA, Ratcliff G, Unitas LJ, Billig B, et al. Sex differences in brain aging: a 625 quantitative magnetic resonance imaging study. Arch Neurol. 1998;55(2):169-79.

Flock MR, Skulas-Ray AC, Harris WS, Etherton TD, Fleming JA, Kris-Etherton PM. Determinants of

Murphy DG, DeCarli C, McIntosh AR, Daly E, Mentis MJ, Pietrini P, et al. Sex differences in human brain
morphometry and metabolism: an in vivo quantitative magnetic resonance imaging and positron emission
tomography study on the effect of aging. Arch Gen Psychiatry. 1996;53(7):585-94.

78. Bourisly AK, El-Beltagi A, Cherian J, Gejo G, Al-Jazzaf A, Ismail M. A voxel-based morphometric magnetic
resonance imaging study of the brain detects age-related gray matter volume changes in healthy subjects of 21-45
years old. The neuroradiology journal. 2015;28(5):450-9.

632 79. Draganski B, Ashburner J, Hutton C, Kherif F, Frackowiak RS, Helms G, et al. Regional specificity of MRI
633 contrast parameter changes in normal ageing revealed by voxel-based quantification (VBQ). Neuroimage.
634 2011;55(4):1423-34.

80. Hafkemeijer A, Altmann-Schneider I, de Craen AJ, Slagboom PE, van der Grond J, Rombouts SA.
Associations between age and gray matter volume in anatomical brain networks in middle-aged to older adults.
Aging Cell. 2014;13(6):1068-74.

638 81. Jancke L, Merillat S, Liem F, Hanggi J. Brain size, sex, and the aging brain. Hum Brain Mapp.
639 2015;36(1):150-69.

640 82. Minkova L, Habich A, Peter J, Kaller CP, Eickhoff SB, Kloppel S. Gray matter asymmetries in aging and 641 neurodegeneration: A review and meta-analysis. Hum Brain Mapp. 2017;38(12):5890-904.

83. Peng F, Wang L, Geng Z, Zhu Q, Song Z. A Cross-Sectional Voxel-Based Morphometric Study of Age- and
Sex-Related Changes in Gray Matter Volume in the Normal Aging Brain. Journal of computer assisted tomography.
2016;40(2):307-15.

84. Tremblay P, Dick AS, Small SL. Functional and structural aging of the speech sensorimotor neural system:
functional magnetic resonance imaging evidence. Neurobiol Aging. 2013;34(8):1935-51.

647 85. Callaghan MF, Freund P, Draganski B, Anderson E, Cappelletti M, Chowdhury R, et al. Widespread age648 related differences in the human brain microstructure revealed by quantitative magnetic resonance imaging.
649 Neurobiol Aging. 2014;35(8):1862-72.

650 86. Honda Y, Noguchi A, Maruyama K, Tamura A, Saito I, Sei K, et al. Volumetric analyses of cerebral white 651 matter hyperintensity lesions on magnetic resonance imaging in a Japanese population undergoing medical check-652 up. Geriatrics & gerontology international. 2015;15 Suppl 1:43-7.

Eiu H, Wang L, Geng Z, Zhu Q, Song Z, Chang R, et al. A voxel-based morphometric study of age- and sexrelated changes in white matter volume in the normal aging brain. Neuropsychiatric disease and treatment.
2016;12:453-65.

656 88. Coffler MS, Patel K, Dahan MH, Yoo RY, Malcom PJ, Chang RJ. Enhanced granulosa cell responsiveness to
657 follicle-stimulating hormone during insulin infusion in women with polycystic ovary syndrome treated with
658 pioglitazone. J Clin Endocrinol Metab. 2003;88(12):5624-31. Epub 2003/12/13.

65989.Xu J, Kobayashi S, Yamaguchi S, Iijima K, Okada K, Yamashita K. Gender effects on age-related changes in660brain structure. AJNR Am J Neuroradiol. 2000;21(1):112-8.

90. Van Der Werf YD, Tisserand DJ, Visser PJ, Hofman PA, Vuurman E, Uylings HB, et al. Thalamic volume
 predicts performance on tests of cognitive speed and decreases in healthy aging. A magnetic resonance imaging based volumetric analysis. Brain research. Cognitive brain research. 2001;11(3):377-85.

664 91. Erten-Lyons D, Dodge HH, Woltjer R, Silbert LC, Howieson DB, Kramer P, et al. Neuropathologic basis of 665 age-associated brain atrophy. JAMA Neurol. 2013;70(5):616-22.

Fjell AM, Westlye LT, Grydeland H, Amlien I, Espeseth T, Reinvang I, et al. Critical ages in the life course of
 the adult brain: nonlinear subcortical aging. Neurobiol Aging. 2013;34(10):2239-47.

668 93. Fraser MA, Shaw ME, Cherbuin N. A systematic review and meta-analysis of longitudinal hippocampal 669 atrophy in healthy human ageing. Neuroimage. 2015;112:364-74.

670 94. Onozuka M, Fujita M, Watanabe K, Hirano Y, Niwa M, Nishiyama K, et al. Age-related changes in brain

regional activity during chewing: a functional magnetic resonance imaging study. Journal of dental research.2003;82(8):657-60.

Heuninckx S, Wenderoth N, Swinnen SP. Systems neuroplasticity in the aging brain: recruiting additional
neural resources for successful motor performance in elderly persons. J Neurosci. 2008;28(1):91-9.

675 96. Holtzer R, Epstein N, Mahoney JR, Izzetoglu M, Blumen HM. Neuroimaging of mobility in aging: a targeted 676 review. The journals of gerontology. Series A, Biological sciences and medical sciences. 2014;69(11):1375-88. 677 97. Kim JH, Lee YS, Lee JJ, Song HJ, Yoo DS, Lee HJ, et al. Functional magnetic resonance imaging reveals agerelated alterations to motor networks in weighted elbow flexion-extension movement. Neurological research. 678 679 2010;32(9):995-1001. 680 98. Linortner P, Jehna M, Johansen-Berg H, Matthews P, Schmidt R, Fazekas F, et al. Aging associated changes 681 in the motor control of ankle movements in the brain. Neurobiol Aging. 2014;35(10):2222-9. 682 Gunning-Dixon FM, Raz N. The cognitive correlates of white matter abnormalities in normal aging: a 99. 683 quantitative review. Neuropsychology. 2000;14(2):224-32. Yuan P, Raz N. Prefrontal cortex and executive functions in healthy adults: a meta-analysis of structural 684 100. 685 neuroimaging studies. Neurosci Biobehav Rev. 2014;42:180-92. 686 101. Van Petten C. Relationship between hippocampal volume and memory ability in healthy individuals across the lifespan: review and meta-analysis. Neuropsychologia. 2004;42(10):1394-413. 687 O'Shea A, Cohen RA, Porges EC, Nissim NR, Woods AJ. Cognitive Aging and the Hippocampus in Older 688 102. 689 Adults. Front Aging Neurosci. 2016;8:298. 690 103. Gonneaud J, Lecouvey G, Groussard M, Gaubert M, Landeau B, Mezenge F, et al. Functional 691 dedifferentiation and reduced task-related deactivations underlie the age-related decline of prospective memory. 692 Brain imaging and behavior. 2017;11(6):1873-84. 693 Meusel LA, Grady CL, Ebert PE, Anderson ND. Brain-behavior relationships in source memory: Effects of 104. age and memory ability. Cortex. 2017;91:221-33. 694 Rieckmann A, Pudas S, Nyberg L. Longitudinal Changes in Component Processes of Working Memory. 695 105. 696 eNeuro. 2017;4(2). 697 106. Suzuki Y, Critchley HD, Suckling J, Fukuda R, Williams SC, Andrew C, et al. Functional magnetic resonance 698 imaging of odor identification: the effect of aging. The journals of gerontology. Series A, Biological sciences and 699 medical sciences. 2001;56(12):M756-60. 700 107. Fusar-Poli P, Placentino A, Carletti F, Landi P, Allen P, Surguladze S, et al. Functional atlas of emotional 701 faces processing: a voxel-based meta-analysis of 105 functional magnetic resonance imaging studies. Journal of 702 psychiatry & neuroscience : JPN. 2009;34(6):418-32. 108. 703 Grady CL. Age-related differences in face processing: a meta-analysis of three functional neuroimaging 704 experiments. Canadian journal of experimental psychology = Revue canadienne de psychologie experimentale. 705 2002;56(3):208-20. 706 109. Daneault V, Hebert M, Albouy G, Doyon J, Dumont M, Carrier J, et al. Aging reduces the stimulating effect 707 of blue light on cognitive brain functions. Sleep. 2014;37(1):85-96. Sala-Llonch R, Junque C, Arenaza-Urquijo EM, Vidal-Pineiro D, Valls-Pedret C, Palacios EM, et al. Changes in 708 110. 709 whole-brain functional networks and memory performance in aging. Neurobiol Aging. 2014;35(10):2193-202. 710 111. Tomasi D, Volkow ND. Aging and functional brain networks. Molecular psychiatry. 2012;17(5):471, 549-58. 711 112. Archer JA, Lee A, Qiu A, Chen SH. A Comprehensive Analysis of Connectivity and Aging Over the Adult Life 712 Span. Brain connectivity. 2016;6(2):169-85. 713 113. Geerligs L, Maurits NM, Renken RJ, Lorist MM. Reduced specificity of functional connectivity in the aging 714 brain during task performance. Hum Brain Mapp. 2014;35(1):319-30. 715 Zhang H, Lee A, Qiu A. A posterior-to-anterior shift of brain functional dynamics in aging. Brain structure & 114. 716 function. 2017;222(8):3665-76. 717 Naber D, Dahnke HG. Protein and nucleic acid content in the aging human brain. Neuropathology and 115. 718 applied neurobiology. 1979;5(1):17-24. 719 116. Soderberg M, Edlund C, Kristensson K, Dallner G. Lipid compositions of different regions of the human 720 brain during aging. J Neurochem. 1990;54(2):415-23. Epub 1990/02/01. 721 117. Surowka AD, Adamek D, Radwanska E, Szczerbowska-Boruchowska M. Variability of protein and lipid 722 composition of human subtantia nigra in aging: Fourier transform infrared microspectroscopy study. Neurochem 723 Int. 2014;76:12-22.

118. Svennerholm L, Bostrom K, Jungbjer B, Olsson L. Membrane lipids of adult human brain: lipid composition
of frontal and temporal lobe in subjects of age 20 to 100 years. J Neurochem. 1994;63(5):1802-11. Epub
1994/11/01.

Hancock SE, Friedrich MG, Mitchell TW, Truscott RJ, Else PL. The phospholipid composition of the human
entorhinal cortex remains relatively stable over 80 years of adult aging. GeroScience. 2017;39(1):73-82.

120. Norris SE, Friedrich MG, Mitchell TW, Truscott RJW, Else PL. Human prefrontal cortex phospholipids

containing docosahexaenoic acid increase during normal adult aging, whereas those containing arachidonic acid
 decrease. Neurobiol Aging. 2015;36(4):1659-69.

Yassine HN, Croteau E, Rawat V, Hibbeln JR, Rapoport SI, Cunnane SC, et al. DHA brain uptake and APOE4
status: a PET study with [1-(11)C]-DHA. Alzheimers Res Ther. 2017;9(1):23.

Coon KD, Myers AJ, Craig DW, Webster JA, Pearson JV, Lince DH, et al. A high-density whole-genome
association study reveals that APOE is the major susceptibility gene for sporadic late-onset Alzheimer's disease. J
Clin Psychiatry. 2007;68(4):613-8.

123. Chouinard-Watkins R, Conway V, Minihane AM, Jackson KG, Lovegrove JA, Plourde M. Interactive impact
 of BMI and APOE genotype on the plasma long chain polyunsaturated fatty acid response to a fish oil supplement
 in healthy participants Am J Clin Nutr. 2015;102:505-13.

740 124. Chouinard-Watkins R, Rioux-Perreault C, Fortier M, Tremblay-Mercier J, Zhang Y, Lawrence P, et al.

Disturbance in uniformly 13C-labelled DHA metabolism in elderly human subjects carrying the apoE epsilon4
allele. Br J Nutr. 2013;110(10):1751-9. Epub 2013/05/02.

- 125. Conway V, Allard MJ, Minihane AM, Jackson KG, Lovegrove JA, Plourde M. Postprandial enrichment of
 triacylglycerol-rich lipoproteins with omega-3 fatty acids: lack of an interaction with apolipoprotein E genotype?
 Lipids Health Dis. 2014;13(1):148. Epub 2014/09/18.
- Plourde M, Vohl MC, Vandal M, Couture P, Lemieux S, Cunnane SC. Plasma n-3 fatty acid response to an nfatty acid supplement is modulated by apoE epsilon4 but not by the common PPAR-alpha L162V polymorphism
 in men. Br J Nutr. 2009;102(8):1121-4. Epub 2009/10/16.

Nock TG, Chouinard-Watkins R, Plourde M. Carriers of an apolipoprotein E epsilon 4 allele are more
 vulnerable to a dietary deficiency in omega-3 fatty acids and cognitive decline. Biochim Biophys Acta.

751 2017;1862(10 Pt A):1068-78.

Gu Y, Vorburger RS, Gazes Y, Habeck CG, Stern Y, Luchsinger JA, et al. White matter integrity as a mediator
 in the relationship between dietary nutrients and cognition in the elderly. Ann Neurol. 2016;79(6):1014-25.

129. Raji CA, Erickson KI, Lopez OL, Kuller LH, Gach HM, Thompson PM, et al. Regular fish consumption and agerelated brain gray matter loss. American journal of preventive medicine. 2014;47(4):444-51.

130. Samieri C, Maillard P, Crivello F, Proust-Lima C, Peuchant E, Helmer C, et al. Plasma long-chain omega-3
 fatty acids and atrophy of the medial temporal lobe. Neurology. 2012;79(7):642-50.

Witte AV, Kerti L, Hermannstadter HM, Fiebach JB, Schreiber SJ, Schuchardt JP, et al. Long-chain omega-3
fatty acids improve brain function and structure in older adults. Cereb Cortex. 2014;24(11):3059-68.

132. Salthouse TA, Mitchell DR, Palmon R. Memory and age differences in spatial manipulation ability. Psychol
Aging. 1989;4(4):480-6. Epub 1989/12/01.

762 133. Salthouse TA, Mitchell DR, Skovronek E, Babcock RL. Effects of adult age and working memory on

reasoning and spatial abilities. J Exp Psychol Learn Mem Cogn. 1989;15(3):507-16. Epub 1989/05/01.

134. Blennow K, de Leon MJ, Zetterberg H. Alzheimer's disease. Lancet. 2006;368(9533):387-403.

135. Deary IJ, Whiteman MC, Pattie A, Starr JM, Hayward C, Wright AF, et al. Cognitive change and the APOE
epsilon 4 allele. Nature. 2002;418(6901):932.

767136.Hoyer S. The aging brain. Changes in the neuronal insulin/insulin receptor signal transduction cascade

trigger late-onset sporadic Alzheimer disease (SAD). A mini-review. J Neural Transm. 2002;109(7-8):991-1002.

137. Rosenberg A, Ngandu T, Rusanen M, Antikainen R, Backman L, Havulinna S, et al. Multidomain lifestyle

intervention benefits a large elderly population at risk for cognitive decline and dementia regardless of baseline

characteristics: The FINGER trial. Alzheimers Dement. 2018;14(3):263-70.

138. Lehtisalo J, Levalahti E, Lindstrom J, Hanninen T, Paajanen T, Peltonen M, et al. Dietary changes and
cognition over 2 years within a multidomain intervention trial-The Finnish Geriatric Intervention Study to Prevent
Cognitive Impairment and Disability (FINGER). Alzheimers Dement. 2018.

139. Croteau E, Castellano CA, Fortier M, Bocti C, Fulop T, Paquet N, et al. A cross-sectional comparison of brain
glucose and ketone metabolism in cognitively healthy older adults, mild cognitive impairment and early
Alzbeimer's disease. Exp. Corontel. 2019;107:18-26

Alzheimer's disease. Exp Gerontol. 2018;107:18-26.

140. Croteau E, Castellano CA, Richard MA, Fortier M, Nugent S, Lepage M, et al. Ketogenic Medium Chain

779 Triglycerides Increase Brain Energy Metabolism in Alzheimer's Disease. J Alzheimers Dis. 2018;64(2):551-61.

141. Nogi A, Yang J, Li L, Yamasaki M, Watanabe M, Hashimoto M, et al. Plasma n-3 polyunsaturated fatty acid
and cardiovascular disease risk factors in Japanese, Korean and Mongolian workers. Journal of occupational
health. 2007;49(3):205-16.

142. Skuladottir GV, Gudmundsdottir S, Olafsson GB, Sigurdsson SB, Sigfusson N, Axelsson J. Plasma fatty acids
 and lipids in two separate, but genetically comparable, Icelandic populations. Lipids. 1995;30(7):649-55.

143. Stark KD, Park EJ, Holub BJ. Fatty acid composition of serum phospholipid of premenopausal women and
 postmenopausal women receiving and not receiving hormone replacement therapy. Menopause. 2003;10(5):448 55.

Stark KD, Beblo S, Murthy M, Whitty JE, Buda-Abela M, Janisse J, et al. Alcohol consumption in pregnant,
 black women is associated with decreased plasma and erythrocyte docosahexaenoic acid. Alcoholism, clinical and
 experimental research. 2005;29(1):130-40.

791 145. Yesilyurt B, Whittingstall K, Ugurbil K, Logothetis NK, Uludag K. Relationship of the BOLD signal with VEP 792 for ultrashort duration visual stimuli (0.1 to 5 ms) in humans. J Cereb Blood Flow Metab. 2010;30(2):449-58. Epub 793 2009/10/22.

794

Table 1: Cross-sectional modulation by age of blood fatty acid 795

				Age-increasing e	effects at baseline	e in blood pool
Reference	n, sex and age	Blood pool	Age-increasing effects	Omega-3 index	EPA	DHA
43	n=460, 299 men and 161 women, 29-97 y (~72 y)	RBC	9.8 y older in Higher Omega-3 Index Quartile compared to Lower Quartile	Higher Omega- 3 Index quartile were 9.8 y older vs lower quartile		
52	53 institutionalized elderly subjects (24 men and 29 women), \ge 60 y (~ 79 y); 24 young healthy adults, 20 – 42 y (~29 y)	Plasma NEFA, TG, CE and PL	In plasma PL: EPA higher in elderly; DHA and DPA: appear lower in the elderly but non- significantly different		PL: 2.1 fold higher	
44	768 acute coronary syndrome patients and 768 matched controls (66 % male, ~61 y)	RBC membranes	Positive relation between age and EPA and DHA levels: 8 years older in those with higher EPA + DHA levels vs those with lower group		Higher RBC EPA + DHA group: 8 y older compared to Lower group	Higher RBC EPA + DHA group: 8 y older compared to Lower group
37	704 outpatients (67% male), ~62 y	RBC	RBC Omega-3 Index increases with age	5.3% increase by 10 years increase		
45	15 centenarians (12 females and 3 males),~ 103 y (101–107 y), living in a family unit, self- sufficient and without major illnesses and 13 normal subjects (6 males and 7 females), ~65 y (6.0 -69 y)	RBC-PL	Increased DHA in RBC-PC and in RBC-PE, and increased DPA in RBC-PS and RBC-PE;			PC: 2.2 fold higher PE: 1.6 fold higher
53	2793 New Zealanders ≥15 y (men and women)	Serum PL, CE and TG	Serum PL: EPA and DHA increase with age in both sexes while DPA increases with age only in women aged between of 20 and 73 y		PL: in both sexes, increased by 0.3 mol% between 20 and 73 y	PL: in both sexes, increased by 0.3 mol% between 20 and 73 y

54	234 men and women (Dutch: low fish consumption), 36 to 88 years (~60 y)	Plasma PL	Significant positive relationship between age and plasma PL concentrations of DHA and EPA.		PL: ~1.5 fold increased between 36 to 88 y	PL: ~1.3 fold increased between 36 to 88 y
56	426 Inuits, 18 to 74 years: 179 men (~38.7 y) and 247 women (~37.8 y), n=254 in 18-39 y and n=172 in \geq 40 y	Plasma PL	Concentrations of EPA, DHA and EPA + DHA increased significantly with age		2.4 fold higher in ≥40 y group compared to 18-39 y group	r 1.4 fold higher in
57	1460 subjects, 18–74 years: 722 men (~40.6 y) and 738 women (~39.6 y), n=784 in 18-34 y, n=432 in 35-49 y and n=244 in 50- 74 y	Plasma PL	Older persons had higher EPA, DHA, EPA+DHA, EPA: AA and n- 3: n-6 ratio in older vs younger individuals		1.1 fold highe in 50-74 y compared to 18-34 y	 1.2 fold higher in 50-74 y compared to 18- 34 y
55	917 subjects, 18-74 y: 422 men (~36.0 y) and 495 women (~35.6 y), n=536 in 18-34 y, n=220 in 35-49 y and n=161 in 50-74 y	Plasma PL	EPA: AA, n-3: n-6 FA, and concentrations of EPA, DHA, and EPA+DHA did not vary according to sex, but there was a significant increase in the concentrations with age		2.5 fold highe in 50-74 y compared to 18-34 y	 1.7 fold higher in 50-74 y compared to 18- 34 y
47	992 participants (mainly men: >80%), age: early 50s to late 70s	RBC membranes	Lower levels EPA + DHA were significantly associated with younger age			
38	446 women, ~48,5 y (40–60 y)	RBC membrane	In women aged ≥50 years, EPA and DPA levels and omega-3 index were significantly higher compared to women under the age of 50 years.	4% higher in ≥50 y compared to <50 y	13% higher ir ≥50 y compared to <50 y) /
40	n= 3196, 55 % women, ~66 y (40-74 y)	RBC	RBC Omega-3 Index increases with age	5% increase every decade		

41	159 771 patients (48% males, 52% females) being evaluated by their physicians for CVD risk	RBC	Increases in EPA and DHA each decade. A <u>fter age 70, significant</u> decrease in EPA while DHA remain high	7% increase by decade until 70 y, stable thereafter	13% increase by decade up to 70 y, then 9% decrease by decade	6% increase by decade until 70 y, stable thereafter
39	6501 women aged 65–80, ~15 years follow-up	RBC	RBC Omega-3 Index increases with age:	Higher Omega- 3 index quartile: 0.6 y older compared to lower quartile		
48	n=456, 320 men and 136 women, 18 to 70 y (~42.5 y)	RBC-PL	EPA+DHA: ~1.4 fold increase in both gender between 18-20 vs 60+ years			
141	411 Japaneses (194 men and 217 women), 418 Koreans (240 men and 178 women) and 252 Mongolians (100 men and 152 women) aged 30-60 y	Plasma	EPA and DHA increase with age in Japanese and Koreans.		Japaneses: 1.4 fold increase Koreans: 1.9 fold increase	Japaneses: 1.2 fold increase Koreans: 1.3 fold increase Mongolians: 1.1 fold <u>decrease</u>
58	75 adults admitted for elective surgery: 48 men (~58 y: 27-81 y) and 27 women (~58 y: 33-74 y)	Plasma PL, RBC-PL and AT	Positive correlation between EPA+DHA and age, in plasma and RBC-PL but not in AT			
42	163 adults, 74 men and 89 women, 20 to 80 years	RBC	Omega-3 Index increased each decade but decreased by 0.3 units with each 3-unit increase in BMI	0.5 unit increase by 10 years of age		
142	119 subjects for each population, Icelandic (59 males and 60 females) and Icelandic-Canadians	Plasma PL	Young Icelandic-Canadians had lower levels of EPA than the middle and oldest age groups		1.8foldincreaseinoldestgroupcomparedtothe youngest	

	(60 males and 59 females), 20-69 years				
143	54 women, 43-60 years: 19 premenopausal (~48 y), 16 postmenopausal not receiving HRT (~52 y) and 19 postmenopausal receiving HRT (~51 y)	Serum PL	DHA levels were significantly lower in premenopausal women than postmenopausal women. Those receiving HRT had significantly lower levels of DPA.		1.3 fold increase in postmenopausal women without HRT vs premenopausal
144	338 women; alcohol intake: abstainers (n=254, ~24,2 y), occasional (n=45, ~27,9 y) and habitual (n=8, ~30,5 y)	Plasma and RBC	DHA and AA correlates positively with maternal age		↑ in plasma (µg/ml et %) ↑ in RBC (%)
49	99 Icelandic women, 18 to 73 y (~45.8 y)	RBC	Proportions of total n-3 PUFA, EPA, and DHA correlated positively with age	\uparrow	\uparrow
145	200 Japaneses, 126 males and 74 females, ~50 y (<35 to ≥65 y)	Serum and RBC total lipids	EPA, DHA, n-3: n-6 ratio and EPA: AA ratio increased with age (stronger effect in serum):	Group ≥65 y compared to group <35 y: 2.3 fold increase in serum and 2 fold increase in RBC	Group ≥65 y compared to group <35 y: 1.7 fold increase in serum and 1,2 fold increase in RBC

AA: arachidonic acid, EPA: eicosapentaenoic acid: DHA, docosahexaenoic acid: DPA: docosapentaenoic acid, AT: adipose tissue, PUFA:
 polyunsaturated fatty acids, FA, fatty acids, PC: phosphatidylcholine, PE: phosphatidylethanolamine, PS: phosphatidylserine, CE: cholesteryl
 esters, NEFA: non-esterified fatty acids, RBC: red blood cells, HRT: Hormone receiving therapy, BMI: body mass index,

799Table 2: Blood fatty acid modulation by age after an omega-3 fatty acid supplementation

Reference	n, sex and age	Blood pool	Omega-3	Age effects
			supplementation	
59	n=115, 60 men and 55 women, 20 to 45 years	RBC	5 doses (0, 300, 600, 900, 1800 mg) of EPA+DHA (fish oil) for ~5 months	Lower Omega-3 Index (O3I) status (P<0.0001) and older age (P=0.02) each predicted greater increases in O3I with supplementation
62	24–28 participants in each age group (except as noted in the tables), young adult = 18-34 y (~23 y) and elderly group = ≥65 y (~74 y)	Plasma	Two supplementations: n- 3 supplement enriched in DHA (680 mg DHA/d plus 323 mg EPA/d) for 3 weeks, or a supplement enriched in EPA (1480 mg EPA/d plus 250 mg DHA/d) for 6 weeks	Expressed as % of total fatty acids: At baseline, total n- 3 PUFA, EPA and DPA higher in elderly (32%, 100% and 25% respectively); Expressed as concentration (mg/L): At baseline, total n-3 PUFA, 18:3n-3, DHA, DPA and EPA higher in elderly (74%, 40%, 63%, 85% and 142% respectively); After supplementation: no higher effect with increasing age
63	15 young (22-35 y) and 10 older (51-71 y) women	Plasma	Daily 1680 mg EPA and 720 mg DHA for 3 months	Older women had a significantly higher increase in EPA and DHA than did young women (EPA: 10-fold vs 8-fold and DHA: 2.5-fold vs 2-fold)
64	6 young (23-33 y) and 6 older (51-68 y) women	Plasma	Daily 1680 mg EPA and 720 mg DHA for 3 months	At baseline there was no difference in percentage of EPA and DHA between young and older women; however, after 3 mo of (n-3) fatty acid supplementation, older women had a significantly higher percentage of EPA and DHA: EPA: 10-fold vs 5- fold and DHA: 2.5-fold vs 1.6-fold
65	10 young (5 men and 5 women, ~22 y) and 10 elderly (5 men and 5 women, ~75 y)	Plasma	EPA-enriched supplement (1.4 g/d of EPA and 0.2 g/d of DHA) for 6 wk	Before and after the EPA supplement, fasting plasma EPA was higher in the elderly (by 85% and 67% respectively)

66	Young (18-42 y; n=93) and old (53-70 y; n=62) men	Plasma and MNC PL	Placebo (corn oil) or 1.35, 2.7, or 4.05 g EPA/day for 12 wks	In both plasma and MNC PL : at baseline, EPA and DPA increase with age while after supplementation, only EPA increases in old men; at baseline, EPA, DPA and DHA respectively ~1.3, ~1.1 and ~1.4 higher in older in plasma and EPA and DHA respectively ~1.3 and ~1.25 higher in older in MNC; with High-EPA supplementation: EPA and DPA respectively ~1.6 and ~1.3 higher in plasma and EPA ~1.4 higher in MNC
61	Elderly (n=9, 5 males and 4 females, 74 y) and young (n=10, 5 males and 5 females, 24 y)	Plasma	680 mg/day of DHA and 320 mg/day of EPA for 3 weeks, followed by 2 weeks of wash-out	Higher baseline plasma EPA in elderly group; In response to the supplement, plasma DHA rose 42% more in the elderly but EPA responded similarly in both groups
67	n=193 (101 women, 92 men), 20–79 y	Plasma PC, CE, NEFA and TG; MNC; RBC; PLAT; BU; AT	EPA+DHA equivalent to 0, 1, 2 or 4 portions of oily fish per week, for 12 months	At baseline, EPA in AT and DHA in plasma TG and AT higher with increasing age; Following supplementation, EPA in plasma TAG higher with increasing age while DHA in AT smaller with increasing age
60	92 Danish women: half premenopaused (~43 y) and half postmenopaused (~56 y), 18-70 y	PLAT, AT	2,2 g of marine n-3 PUFA (38,5% EPA, 25,9% DHA and 6,0% DPA) or control oil (thistle oil) daily for 12 weeks	Baseline contents of EPA, DPA and DHA were all significantly lower (P<0.05) in premenopausal group both in platelets and adipose tissue, except for EPA in platelets (P=0.05); After supplementation, increase in platelets and adipose tissue was, however, the same in both groups

in both groups800EPA: eicosapenaenoic acid, DHA, docosahexaenoic acid, DPA: docosapentaenoic acid, PLAT: platelets, AT: adipose tissue, PUFA:801polyunsaturated fatty acids, PC: phosphatidylcholine, CE: cholesteryl esters, NEFA: non-esterified fatty acids, MNC: mononuclear cells, RBC:

802 red blood cells, BU, buccal cells,