

Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxxx

### **ARTICLE TYPE**

# Metal dyshomeostasis based biomarkers for lung cancer diagnosis using human biofluids

Belén Callejón Leblica, José Luis Gómez Arizaa\*, Antonio Pereira Vegab and Tamara García Barrera\*a

s <sup>a</sup> Department of Chemistry, Faculty of Experimental Sciences, University of Huelva, Campus de El Carmen, 21007-Huelva, Spain. Research Center on Health and Environment (RENSMA)

10 AUTHOR E-MAIL ADDRESS: T. García-Barrera (tamara@dqcm.uhu.es); J.L, Gómez-Ariza (ariza@uhu.es)

Received (in XXX, XXX) Xth XXXXXXXXX 20XX, Accepted Xth XXXXXXXXX 20XX DOI: 10.1039/b000000x

- Lung cancer (LC) is one of the most common causes of cancer-related deaths on the world and it is well known that trace elements play important roles in the carcinogenic process activating and inhibiting enzymatic reactions and metalloproteins, in which they usually participate as cofactors. Eleven elements have been included in the study: V, Cr, Mn, Fe, Co, Cu, Zn, Se, Mo, Cd, and Pb, some of them considered toxic (V, Cd, Cr and Pb), while others are essential (Co, Mo, Se, Fe and Zn), and they have been analyzed
- 20 by ICP-QQQ-MS in serum, urine and for the first time in bronchoalveolar lavage fluid (BALF). In order to deepen the involvement of metal in this process, an analytical metallomic approach based on non-denaturing precipitation of proteins (NDPP) has been optimized for the fractionation of high molecular mass (HMM) and low molecular mass (LMM) metal species, in order to distinguish between metal species that affect the biological activity and toxicological potential of the elements. In this work, the NDPP/ICP-
- 25 QQQ-MS metallomic approach has been applied for the first time to serum, urine and BALF samples from lung cancer patients and controls in order to get metal-size molecule profiles (MSMP), which can be used as metal-based biomarkers of altered metabolic processes as oxidative stress and homeostasis. In this sense, we have demonstrated that several metals are good biomarkers when they are related to the labile ions, complexed with low molecular mass ligands, or in the form of metalloproteins (i.e. V and Cr in HMM and
- <sup>30</sup> Cu in LMM), which has been described for the first time. On the other hand, metal dyshomeostasis biomarkers are proposed using metals ratios and correlations. Finally, ratios between elements resulted to be important biomarkers for lung cancer disease in serum (V/Mn, V/Pb, V/Zn, Cr/Pb), urine (Cr/Cd, Mn/Cd, V/Cd, Co/Cd, Cd/Pb) and BALF (V/Cu), which reflect the dyshomeostasis of metals in lung cancer disease. In this sense, several metals are correlated to others suggesting also the existence of an interconnected homeostasic in lung cancer.

35 homeostasis in lung cancer.

Keywords: Lung cancer, metal homeostasis, inter-element interactions, bronchoalveolar lavage fluid, serum, urine, ICP-QQQ-MS.

#### Introduction

40 Elements are essential to regular human homeostasis and play significant roles in biological systems participating in many cellular processes. These essential metals, which are only required in trace amounts, are crucial for the function of numerous enzymes required for fundamental biochemical processes<sup>1</sup>. For instance, iron is essential for the function of ribonucleotide reductase<sup>2</sup> and zinc is a constituent of over 300 enzymes that play vital roles in gene expression<sup>3</sup>. Apart from their role in enzyme function, metals are also required for numerous biological processes such as the

<sup>&</sup>lt;sup>b</sup> Pneumonology Area of Juan Ramón Jiménez Hospital, Huelva, Spain

transport of oxygen in the blood, which is mediated by hemoglobin containing iron<sup>4</sup>.

Deficiency or excess of any of these elements can lead to disease (e.g. anemia) or deleterious toxic effects, inflammation<sup>5</sup> and <sup>5</sup> cancer<sup>6</sup>. As a consequence the levels of these essential metals have to be carefully balanced and a homeostatic state is maintained within the body<sup>5</sup>.

On the other hand, a number of non-essential elements can also have important implications on human health. In this way, environmental exposure to arsenic, cadmium, lead and nickel has carcinogenic consequences<sup>6</sup> due to activation of oncogenic signaling pathways<sup>7,8</sup>, oxidative stress<sup>6,9,10</sup> or inhibition of DNA repairing system by Ni<sup>11</sup>. Many of these non-essential metals can also alter some enzymes function, as is the case of the competitive interaction of zinc and cadmium for many enzymes as consequence of their very similar atomic structure<sup>12</sup>, which has dramatic effects on many zinc-containing enzymes involved in important biological processes leading to cancer onset<sup>13</sup>. Many metals also contribute to cancer progression and metastasis<sup>14–18</sup>.

<sup>20</sup> Therefore, the study of the elements levels in human tissues and particularly in serum can provide interesting information about the changes occurring during the biological processes involved in the progression of diseases such as lung cancer (LC), which is the second most frequent cancer in humans and is the common cause of cancer deaths in the world<sup>19</sup>. Thus, the changes in the presence of some elements and the profiling of their chemical forms can reflect the status of human nutrition and metabolism and can assist to a possible early prediction of cancer onset and development.

There are two different points of view to relate the behavior of the 30 elements in the organism during the carcinogenic process: the disturbance of the natural chemical form of the essential element in the metabolism due to cancer onset and progression and the consideration of elements involved in the carcinogenic process as a consequence of their high exposure. On the other hand, the 35 majority of works focus on the estimation of a deficiency state or excess and into a lesser extent about the unbalance episodes in which the excess of one element affects the function of other. However, the importance of the chemical form of elements in biology is well-known. Elements can be mainly present as labile 40 ions or complexed with low molecular mass ligands, or in the form of metalloproteins. This difference between low molecular mass (LMM) and high molecular mass (HMM) species is very important, since it finally affects to the biological activity or toxicological potential of the element and their mobility across 45 different biological compartments. On the other hand, the importance of metal homeostasis and metals interactions in biology has also been demonstrated<sup>20</sup> and the ratio of metals (Cu/Zn) resulted to be different in serum and whole blood21 and pleural effusion<sup>22</sup> of lung cancer patients, but the interplay of 50 elements are rarely reported.

Many authors have reported concentrations of essential and nonessential elements in human biological samples from patients with lung cancer, such as serum<sup>21,23-25</sup>, plasma<sup>26,27</sup>, urine<sup>28,29</sup>, pleural efussion<sup>22</sup> or hair<sup>24,30</sup>. In addition, there are several papers 55 describing the analysis of metals in bronchoalveolar lavage fluid<sup>31,32</sup>, but they are not related with lung cancer patients. BALF is obtained during the exploratory study of patients with lung diseases and provides constituents information on cellular and biochemical epithelial surface of the lower respiratory tract 60 through instillation and later aspiration of liquid in one or more lung segments. It is estimated that BALF samples take a million cells (1% of the lung surface) to yield about 1 ml of pulmonary secretions in the actual total recovered liquid<sup>33</sup>. Because of BALF is in close interaction with lung tissue is a more representative 65 sample of lung status than other biofluids as blood or urine. Other authors have also classify lung cancer patients and healthy people using metals content in serum and hair<sup>24,30</sup> and urine<sup>28</sup> in order to use the alteration of elements for the diagnosis of LC.

In addition most elemental determinations in several biofluids have been performed by techniques such as atomic absorption spectroscopy (AAS)<sup>31,34–37</sup>, particle induced X-ray emission<sup>38</sup>, energy dispersive X-ray fluorescence<sup>39</sup>, inductive coupled plasma atomic emission spectroscopy (ICP-AES)<sup>24,28,29</sup> or inductive coupled plasma mass spectrometry (ICP-MS)<sup>22,26,27,30,32</sup>. The main advantage of using an ICP-MS equipped with a triple quadrupole is the elimination of interferences by operating in either standard single quadrupole (SQ) mode or tandem MS/MS. A significant difference is the measurement of selenium in biological samples. In this sense, the signal of <sup>80</sup>Se in oxygen mode (<sup>96</sup>SeO<sup>+</sup>) would overlap with signals like <sup>96</sup>Zr<sup>+</sup>, <sup>96</sup>Mo<sup>+</sup> or <sup>96</sup>Ru<sup>+</sup> in a conventional ICP-MS, but with the ICP-QQQ-MS this drawback is eliminated because these interferences would be rejected in the first quadrupole.

The aim of this work is to propose metal based biomarkers for lung cancer disease in serum, urine and for the first time in BALF, using a powerful analytical tool like ICP-QQQ-MS, that has not been previously used in biofluids from LC patients. In addition, the fractionation of the blood serum for the analysis of metals in HMM and LMM fractions provide for the first time new contributions in the field of lung cancer using a procedure based on protein precipitation in non-denaturing conditions. Finally, some of these biomarkers are based in the homeostasis of metals using their ratios and correlations.

#### Materials and methods

#### 95 Instrumentation

Elemental analysis was performed by inductively coupled plasma mass spectrometry equipped with an triple quadrupole, using the Agilent 8800 Triple Quad (Agilent Technologies, Tokyo, Japan), with helium of high-purity grade (>99.999%) and oxygen of high purity (>99.999%) and H<sub>2</sub> gas. Instrumental conditions were optimized using a Tuning aqueous solution containing Li, Co, Y and Tl at 1µg L<sup>-1</sup>. Nickel sampling and skimmer cones were employed, with a sampling depth of 10 mm. The forward power

was set at 1550 W, and the gas flow rates were fixed at 15 L min<sup>-1</sup> for plasma gas and 1.08 L min<sup>-1</sup> for carrier gas. To analyze most elements a flow of 4.5 mL min<sup>-1</sup> of helium was used. For selenium, a flow of 2 mL min<sup>-1</sup> of H<sub>2</sub> with 40% of O<sub>2</sub> was used in MS/MS mode. Isotopes monitored were <sup>51</sup>V, <sup>53</sup>Cr, <sup>55</sup>Mn, <sup>57</sup>Fe, <sup>63</sup>Cu, <sup>64</sup>Zn, <sup>65</sup>Cu, <sup>66</sup>Zn, <sup>78</sup>Se, <sup>80</sup>Se, <sup>95</sup>Mo, <sup>98</sup>Mo, <sup>103</sup>Rh, <sup>112</sup>Cd, <sup>114</sup>Cd and <sup>208</sup>Pb with dwell time of 0.3 s per isotope.

A MARS microwave oven (CEM Matthrews, NC, USA) was used for the mineralization of samples in PFA Teflon vessels.

#### 10 Standard solutions and reagents

Acetone (Trace Analysis Grade), nitric acid (purity 67-69%, Trace Metal Grade) and hydrogen peroxide (purity 30-35%, Optima Grade) were purchased from Fisher Scientific (Leicestershire, UK). Water was purified with a Milli-Q Gradient system (Millipore, Watford, UK).

Blood samples were collected from lung cancer patients (LC), patients with non-cancerous lung diseases (NCC) and healthy people used as control (C), at the Pneumonology Area of Juan Ramón Jiménez Hospital (Huelva, Spain). The blood samples were

- obtained by venipuncture of the antecubital region, after 8 hours of fasting, and collected in BD Vacutainer SST II tubes with gel separator and Advance vacuum system. The samples were immediately cooled and protected from light for 30 minutes to allow clot retraction. After centrifugation (2000 g for 10 minutes)
- 25 serum samples were frozen at -80°C until analysis.

  Bronchoalveolar fluid samples were obtained by instillation and subsequent aspiration of saline solution into one or more pulmonary segments or subsegments through a bronchoscope from patients with LC and NCC. It is not possible to obtain
- <sup>30</sup> bronchoalveolar lavage samples from healthy individuals as it is an invasive technique. The resulting liquid was aliquoted into eppendorf tubes and stored at -80 ° C until analysis. Finally, urine samples were collect in sterile urine vessels and were aliquoted into eppendorf tubes and stored at -80 ° C until analysis.
- 35 The study was performed in accordance with the principles contained in the Declaration of Helsinki and approved by the Ethical Committee from Juan Ramón Jiménez Hospital and University of Huelva. Table 1 showed clinical characteristics of patients (LC and NCC) and healthy controls (HC). In addition, all
- 40 people gave informed consent for the extraction of peripheral venous blood and BALF.

#### Fractionation of samples and total elements determination

45 For non-denaturing protein precipitation of samples we used a method previously developed by the authors with some modifications <sup>40</sup>. This fractionation procedure was only applied to serum samples since the precipitation of proteins was negligible in bronchoalveolar lavage fluid and urine samples. Briefly, 600 μl of <sup>50</sup> cold acetone (-20°C) was dropwise added to 300 μl of serum and kept for 10 min in an ice bath. The mixture was vortexed and the precipitate removed by centrifugation (12857 g, 4°C, 5 min). The supernatant, containing LMM species, was taken to dryness under nitrogen stream and reconstituted in 1 ml of ultrapure water with <sup>55</sup> 0.1 μg L<sup>-1</sup> of Rh as internal standard. On the other hand, the precipitate was subjected to microwave assisted acid digestion for the determination of metal content in the HMM fraction. To this end, the precipitate was introduced into the microwave vessel together with 500 μl of a mixture containing nitric acid and

- <sup>60</sup> hydrogen peroxide (4:1 v/v). Mineralization was carried at 400 W, ramping the temperature to 150°C in 10 min. Then, extracts were made up to 3 ml with ultrapure water, adding 0.1  $\mu$ g L<sup>-1</sup> of Rh. Before analysis, samples were filtered through 0.45  $\mu$ m pore size filters of PTFE.
- 65 The fractionation procedure was validated using an aqueous solution of bovine serum albumin standard containing Cu and Zn, in order to ensure the integrity of the metal protein binding during sample treatment as the same procedure used by Gónzalez-Domínguez et al<sup>41</sup>. In addition, a reference material of serum and <sup>70</sup> urine (Clinchek, Serum Control lyophylised for trace elements, level II, Recipe, and Clinchek, Urine Control lyophylised for trace elements, level II, Recipe) were used to validate the analytical method by ICP-QQQ-MS including the reproducibility.

Finally, total metal content of serum, urine and BALF (TOTAL) vas determined in diluted samples previously described<sup>42</sup>. In this way, serum, urine and BALF samples were five-fold diluted with ultrapure water and 0.1 µg L<sup>-1</sup> of Rh was added as internal standard

Table 1. Clinical Characteristics of patients (LC and NCC) and healthy 80 controls (HC)

| Samples   | Characteristics   | LC          | NCC     | HC      |  |  |  |
|-----------|-------------------|-------------|---------|---------|--|--|--|
|           | Number of samples | n=48        | -       | n=39    |  |  |  |
|           | Age (years)       | $65 \pm 11$ | -       | 58 ± 14 |  |  |  |
| SERUM     | Sex (M/W)         | 39/9        | -       | 17/22   |  |  |  |
| and URINE | Histology         |             |         |         |  |  |  |
|           | NSCLC 42          |             | -       | -       |  |  |  |
|           | SCLC              | 6           | -       | -       |  |  |  |
|           | Number of samples | n=24        | n=31    |         |  |  |  |
|           | Age (years)       | $65 \pm 13$ | 54 ± 14 | -       |  |  |  |
|           | Sex (M/W)         | 20/4        | 27/4    | -       |  |  |  |
|           | Histology         |             |         |         |  |  |  |
|           | NSCLC             | 22          | -       | -       |  |  |  |
| BALF      | SCLC              | 2           | -       | -       |  |  |  |
|           | DILD              | -           | 11      | -       |  |  |  |
|           | Haemoptysis       | -           | 5       | -       |  |  |  |
|           | Bronchiectasis    | -           | 5       | -       |  |  |  |
|           | SPN               | -           | 3       | -       |  |  |  |
|           | Others*           | -           | 7       | -       |  |  |  |

NSCLC: Non-small cell lung cancer, SCLC: Small cell lung cancer, DILD: Diffuse Interstitial Lung Disease, SPN: solitary Pulmonary Nodule, M: Men, W: Women, LC: Lung Cancer. \*Lung Contusions

#### Statistical analysis

Statistical calculations were made in STATISTICA 8.0 software (StatSoft, Tulsa, USA). Non parametric methods were used since most of the variables showed a skewed distribution (checked by normal probability plots) and variances were not homogeneous (checked by Levene's test). Thus, group comparison was conducted using Krustal-Wallis one-way analysis of variance, and when significance effects were observed, Mann Whitney U test was carried out for pairwise comparisons to find differences between groups. Only p values below 0.05 were regarded as statistically significant. Finally, to evaluate the specificity and

sensitivity of metabolites altered by the disease, ROC (receiver operator characteristic) curves were applied to the dataset and metabolites with "area under the curve" (AUC) higher than 0.75 were considered as relevant in the progression of LC.

#### Results and discussion

## Classification analysis using elements concentrations in urine, BALF, serum and serum fractions

In this work, the total concentration of 11 elements in 87 serum samples, 87 urine samples and 55 BALF samples from patients with LC and controls (HC or NCC) were determined by ICP-QQQ-MS. In addition, the analysis of metal content by size-fractionation in serum (HMM and LMM fractions) provided new contributions to the characterization of the metal profile of patients with LC.

In order to establish a classification of the study groups on the basis of their elements concentrations, statistical discriminant analysis of partial least squares (PLS-DA) was performed. In addition, the weight of each variable (elements concentrations and inter-element ratios) of each biofluid (Total serum, HMM and LMM fractions, urine and BALF) in the classification (VIP value, variable importance on the projection) was used together with other criteria 25 (see next section) for the selection of biomarkers for lung cancer diagnosis.

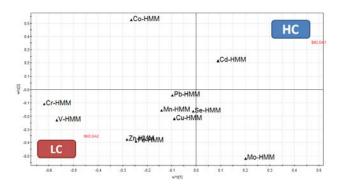
PLS-DA is a supervised method that provides statistical models that allow visualizing groupings and trends between different groups of samples through representation of score plots. Figure 1A represents the 3D score plot of serum samples from LC and C groups. A clear separation of the groups can be observed indicating that there are elements whose concentrations are clearly altered in lung cancer, or maybe that when a threshold

35 concentration in the body is exceeded, lung cancer is induced.

40 Figure 1. 3D score plot obtained by PLS-DA in serum samples using the concentration of elements as variables. A) TOTAL serum, B) HMM fraction, C) LMM fraction. Red triangles: Cancer group, Black triangles: Control group.

45 Similar graphs were constructed for urine and BALF samples (Supplementary Material, Figure 1), which give also a good separation between LC and HC or NCC group.

In addition, the position of signals on the "loading" chart indicates if the element concentration is overexpressed or inhibited in lung cancer. In this way, Figure 2 shows the representation of the loadings plot obtained using the elements concentration in the HMM fraction of serum samples. Then, the signals clustered in the left side of the graph represent metals overexpressed in lung cancer, while those found in the opposite corner are reduced in these samples. Thus, the concentration of vanadium and chromium in the HMM of serum are higher in lung cancer patients, which confirm results obtained by Mann-Whitney tests. The loadings plots were also obtained for the concentration of elements in the LMM fraction of serum, serum, urine and BALF (Supplementary Material, figure 2, 3, 4 and 5).



65 Figure 2. Loadings plot for comparison of elements concentration in the HMM fraction of serum obtained from LC and HC.

### Elements concentrations and ratios. Proposal of lung cancer biomarkers for Diagnosis

Tables 2, 3 and 4 shows the concentration of elements in the serum and serum fractions, urine and BALF, respectively, as well as VIP values (variable importance on the projection), *p*-value (Mann-Whitney U test) and AUC ("area under the curve", ROC "Receiver operation curve"). Only those elements or inter-elements ratios that have a VIP value greater than 1, fold change lower than 0.5 or greater than 2, *p*-value<0.05 and AUC>0.75 (see next section) are included in the tables in order to select the most powerful lung cancer biomarkers in this paper.

Concentration of elements for LC and HC patients in *serum* samples and serum fractions (HMM and LMM) are summarized in Table 2. Total levels of vanadium in serum, vanadium and chromium in HMM fraction of serum and copper in LMM fraction of serum were statistically different between LC and C and all of them were increased in LC patients from 2 to 5-fold (V-HMM).

In this study, **vanadium** concentration was 3.77 and 5.03-fold higher in LC patients in serum and HMM fraction of serum, respectively. There is very scarce information about the role of vanadium in the body. Lin et al., analyzed vanadium in plasma from women with LC and found no significant differences between cancer and control groups<sup>26</sup>, that has also been concluded in other works<sup>23</sup>. On the other hand, some authors have reported the anticancer properties of vanadium and showed that complexes of these metals are the new metal-based drugs used in the treatment of several cancers, such us lung cancer<sup>43</sup>. Other authors have

demonstrated the influence of vanadium compounds in the cytotoxicity of some ligands in human lung cancer cultured cells<sup>44</sup>. **Chromium** is associated to glucose and lipid metabolism, protein synthesis and other important physiological functions<sup>45</sup>. Hexavalent form of chromium and its containing compounds are well-established lung carcinogens. Chronic exposure of the normal human epithelial cells is able to induce malignant cell transformation, the first stage of metal carcinogenesis<sup>46</sup>. Chromium has been previously found to be increased in serum samples of LC patients<sup>24,26</sup>. In this work chromium is 2.11-fold increase in the HMM of serum of LC patients. Lin et al reported an increase of this element in serum of LC group, but they do not found significant statistical differences between control and LC groups.

Copper is primarily found in serum (95% as part of the oxidative enzyme ceruloplasmin and the remainder is present in an anionic form loosely bound to albumin<sup>47</sup>. Copper, zinc and manganese regulate the levels and activities of antioxidants, especially enzymatic ones, and the disturbed redox status may be critical to lung carcinogenesis. These metals are cofactors or ions stabilizing the molecular structure of superoxide dismutase (SOD), an endogenous antioxidant<sup>48</sup>. The concentration of copper has been reported to be increased in LC patients and its concentration is related to cancer state and localization<sup>21,23,24,26,27,35–37,47</sup>. In this study, the copper concentration in the LMM fraction of serum (labile copper) is 2.29-fold higher than in control patients.

The main altered metal in the *urine* of LC patients of this study is <sup>30</sup> **cadmium** (Table 3). Cadmium is recognized as a human

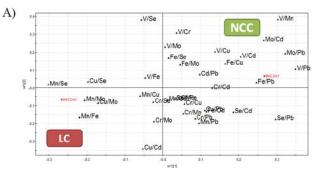
carcinogen, a classification mainly based on occupational studies of lung cancer<sup>49</sup>. Cadmium levels in urine, serum and blood of smokers has previously proposed as a marker for the development and progression of lung diseases 50 and it has also found to be 35 increased in the urine of cancer patients <sup>28,29</sup>. The National Report on Human Exposure to Environmental Chemicals from the Centers for Disease Control and Prevention (CDC) reported a mean urine cadmium concentration of 0.172 µg/L (0.199 µg/g creatinine) for non-smokers versus 0.308 µg/L (0.336 µg/g creatinine) for 40 smokers in the U.S. population (not stratified by age)<sup>51</sup>. On the other hand, lung cancer patients have manganese concentrations in bronchoalveolar lavage fluid 1.5-fold higher than control group (Table 4). Manganese superoxide dismutase (Mn-SOD), as a single superoxide radical scavenger in 45 mitochondria, may have a big role in preventing cells as an antioxidant and tumor suppressor<sup>52</sup>. In the lungs, Mn-SOD is considered to be of critical importance for antioxidant defense<sup>53</sup>. A number of studies have defined associations between the Mn-SOD Ala16Val polymorphism and different cancer types<sup>54–56</sup>. Lin et al<sup>26</sup> 50 found higher levels of Mn in serum from LC group according our results. In the same way, Tan et al. reported a decrease of this element in urine of LC patients<sup>29</sup> . Some authors reported decreased levels of manganese in BALF from patients with diffuse lung diseases<sup>34</sup> or in ill calves<sup>57</sup>, but there is no antecedent about 55 the concentration of manganese in BALF samples from LC patients.

#### Metal dyshomeostasis in lung cancer

100

60 As has been commented, lung cancer is associated with imbalances in the levels of elements that are reflected in the results from serum, urine, BALF and/or fractions (Table 2, 3 and 4). In addition, the concentrations of elements were further analyzed to obtain **inter- element ratios** in order to understand the interrelations of
65 elements. Those ratios that showed significant changes between
the study groups are listed in Table 2, 3 and 4 for serum, urine and
BALF, respectively, which allows discovering the effect of
alterations of single elements on the homeostasis of the rest in each
level of structural organization. Good separations between LC and
70 C or NCC group were also obtained using the inter-element ratios
by PLS-DA in BALF (Figure 3), urine and serum samples
(Supplementary Material, figure 6 and 7).

Results obtained shows clear interactions of V with Mn, Pb and 75 Zn, Cr with Pb in serum, Cd with Cr, Mn, V, Co and Pb in urine and V with Cu in BALF. Several papers describes the interactions between Cu and Zn, concluding that high Cu:Zn ratios are common in serum of patients with lung cancer<sup>21–23,58–60</sup>, but we do not find this interaction to be significate using VIP, *p*-value, AUC and fold change criteria. The ratio between Cd and Zn has also been found to be different between smokers and nonsmokers and it is also different in smokers for several diseases and cancers<sup>50</sup>. On the other hand, the inter-element ratios that resulted to be significative different in lung cancer patients against healthy people in this paper have not been previously described and show high values of VIP, fold changes AUC and p-value.



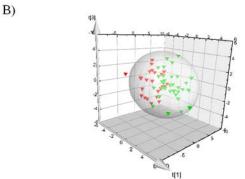


Figure 3. A) Loadings plot for comparison of inter-element ratios in BALF obtained from LC and NCC. B) 3D score plot obtained by PLS-DA in BALF samples using the inter-element ratios as variables. Red triangles: Cancer group, Green triangles: Control group.

Table 2. Total elements concentration (µg g<sup>-1</sup>) in serum samples (TOTAL) and serum fractions (HMM and LMM)

|                               | V<br>(TOTAL) | V<br>(HMM) | Cr<br>(HMM) | Cu<br>(LMM) | V/Mn     | V/Pb     | V/Zn     | Cr/Pb    |
|-------------------------------|--------------|------------|-------------|-------------|----------|----------|----------|----------|
| Medium concentration HC       | 0.05         | 0.04       | 0.39        | 33.33       | 0.02     | 0.03     | 0.00004  | 0.35     |
| <sup>a</sup> SEM in HC group  | 0.02         | 0.02       | 0.17        | 2.78        | 0.01     | 0.01     | 0.00002  | 0.14     |
| Medium concentration LC       | 0.18         | 0.19       | 0.83        | 76.29       | 0.08     | 0.14     | 0.00164  | 0.96     |
| <sup>b</sup> SEM in LC group  | 0.04         | 0.05       | 0.12        | 2.58        | 0.01     | 0.02     | 0.00004  | 0.19     |
| Fold Change (LC/HC)           | 3.77         | 5.03       | 2.11        | 2.29        | 3.55     | 4.27     | 4.10     | 2.79     |
| VIP                           | 1.82         | 1.72       | 1.89        | 2.08        | 1.427    | 1.472    | 1.50     | 1.641    |
| p-value (Mann Whitney U test) | 0.000039     | 0.000007   | 0.00003     | 0.00000     | 0.000033 | 0.000015 | 0.000048 | 0.000026 |
| AUC                           | 0.76         | 0.78       | 0.78        | 0.92        | 0.77     | 0.76     | 0.76     | 0.76     |

<sup>&</sup>lt;sup>a</sup>Standard error of the mean (n=39), <sup>b</sup>Standard error of the mean (n=48),

Fold changes (Lung cancer vs healthy people), Variable importance on the projection values (VIP), p-value (statistical pairwise comparisons by Mann-Whitney U test) and AUC values of ROC curves

Table 3. Total elements concentration (μg g<sup>-1</sup>) in urine samples)

|                               | Cd       | Cr/Cd    | Mn/Cd    | V/Cd     | Co/Cd | Cd/Pb    |
|-------------------------------|----------|----------|----------|----------|-------|----------|
| Medium concentration HC       | 0.67     | 6.38     | 10.62    | 3.83     | 1.56  | 0.09     |
| <sup>a</sup> SEM in HC group  | 0.07     | 1.47     | 1.66     | 0.73     | 0.24  | 0.01     |
| Medium concentration LC       | 1.55     | 2.33     | 3.95     | 1.6      | 0.42  | 0.23     |
| <sup>b</sup> SEM in LC group  | 0.21     | 0.47     | 0.48     | 1.43     | 0.05  | 0.03     |
| Fold Change (LC/HC)           | 2.32     | 0.37     | 0.37     | 0.42     | 0.27  | 2.47     |
| VIP                           | 1.703    | 1.813    | 2.07     | 1.484    | 1.897 | 1.574    |
| p-value (Mann Whitney U test) | 0.000014 | 0.000006 | 0.000002 | 0.000021 | 0     | 0.000955 |
| AUC                           | 0.79     | 0.76     | 0.77     | 0.75     | 0.81  | 0.74     |

<sup>&</sup>lt;sup>a</sup>Standard error of the mean (n=39), <sup>b</sup>Standard error of the mean (n=48),

Fold changes (Lung cancer vs healthy people), Variable importance on the projection values (VIP), p-value (statistical pairwise comparisons by Mann-Whitney U test) and AUC values of ROC curves

Table 4. Total elements concentration (µg g-1) in bronchoalveolar lavage fluid)

|                               | Mn    | V/Cu    |
|-------------------------------|-------|---------|
| Medium concentration NCC      | 0.46  | 1.03    |
| <sup>a</sup> SEM in NCC group | 0.06  | 0.10    |
| Medium concentration LC       | 0.69  | 0.51    |
| <sup>a</sup> SEM in LC group  | 0.09  | 0.11    |
| Fold Change (LC/NCC)          | 1.51  | 0.49    |
| VIP                           | 1.29  | 1.22    |
| p-value (Mann Whitney U test) | 0.003 | 0.01811 |
| AUC                           | 0.75  | 0.76    |

<sup>&</sup>lt;sup>a</sup> Standard error of the mean ( (n=31), <sup>b</sup>Standard error of the mean ( (n=24)

**Table 5.** Correlations between metals in biofluids and fractions (p<0.05)

Fold changes (Lung cancer vs non-cancerous control), Variable importance on the projection values (VIP), p-value (statistical pairwise comparisons by Mann-Whitney U test) and AUC values of ROC curves

| URINE         |                                    |
|---------------|------------------------------------|
| Spearman test | Correlation coefficients (r)       |
| Cr            | Mn (r=0.58), Fe (r=0.58), Pb(0.63) |
| Mn            | Fe (r= 0.7), Pb (r=0.58)           |
| Fe            | Pb (r=0.53)                        |
| SERUM         |                                    |
| V-total       | Cr-total (r=0.62), Cr-HMM (r=0.58) |
| V-HMM         | Cr-total (r=0.62), Cr-HMM (r=0.59) |
| Mn-total      | Zn-HMM (r=0.5), Zn-total (r=0.44)  |
| Zn-total      | Mn-HMM (r=0.34)                    |
| BALF          |                                    |
| V-total       | Pb(r=0.5)                          |
| Cr-total      | Zn(r=0.55), Se (r=0.52)            |
| Mn-total      | Cu(r=0.56)                         |
| Co-total      | Cu(r=0.55), Mo(r=0.53)             |
| Zn-total      | Cr(r=0.55), Se(r=0.68)             |
| Se-total      | Zn(r=0.68)                         |

Moreover, Table 5 shows Spearman's correlation coefficients calculated to evaluate the interdependence of metals in the organism, suggesting the existence of an interconnected 5 homeostasis. In this sense, levels of Cr, Mn, Fe, Pb, V, Zn, Co and Se in the different biofluids and/or HMM fraction present a correlation coefficient among them higher than 0.5, indicating a possible common route for their biological regulation. Among these elements, Cr, Mn and V also show significate imbalances in the analyzed biofluids (Table 2). On the other hand, the interrelations between Cr and Pb (urine) and V and Pb (BALF) were also important as can be concluded by the inter-elements ratios (serum, Table 2).

Other elements that are interconnected only by the correlation coefficients are selenium and zinc (BALF). The antioxidant properties of selenium are well-known and alterations of this element can be related with oxidative stress. In this work, selenium is positively correlated with zinc in BALF samples, which is implicated in glucometabolic disorders<sup>61</sup>. Cobalt and copper are also positively correlated in BALF. The function of cobalt in the body is to be a cofactor of vitamin B12, but in the form of labile ions is able to generate reactive oxygen species, such as copper and iron<sup>41</sup>. On the other hand, other correlations have not been previously described.

#### **Conclusions**

The present paper describes for the first time the use of triple quadrupole ICP-MS for the analysis of metals in biofluids from lung cancer patients and healthy people. This powerful analytical methodology has been combined with a careful approach based

in the use of VIP values, fold changes, *p*-value<0.05 and AUC values to propose the most important lung cancer biomarkers. This work shows that metal contents in serum, urine and for the

- 35 first time in BALF can be used to distinguish healthy people and lung cancer patients. In addition, several metals are good biomarkers when they are related to the labile ions, complexed with low molecular mass ligands, or in the form of metalloproteins (i.e. V and Cr in HMM and Cu in LMM), which 40 has been described for the first time.
- On the other hand, taking into account the complexity of biological systems, some important effects may be caused by the interplay of more than two elements, but such interactions are rarely reported. Finally, ratios between elements resulted to be
- 45 important biomarkers for lung cancer disease (in serum: V/Mn, V/Pb, V/Zn, Cr/Pb, urine: Cr/Cd, Mn/Cd, V/Cd, Co/Cd, Cd/Pb and BALF: V/Cu), which reflect the dyshomeostasis of metals in lung

cancer disease. In this sense, several metals are correlated to others suggesting also the existence of an interconnected 50 homeostasis in lung cancer.

#### Acknowledgements

The authors thank to Spanish Ministry of Economy and Scompetitiveness (CTM2015-67902-C2-1-P) and Regional Ministry of Economy, Innovation, Science and Employment (Andalusian Government. P12-FQM-0442. B. Callejón-Leblic thanks the Ministerio de Educación for a predoctoral scholarship FPU13/03615. Finally, the authors are grateful to FEDER (European Community) (UNHU13-1E-1611 and UNHU15-CE-3140).

#### References

- 65 1 W. Maret, Int. J. Mol. Sci., 2016, 17, 1–8.
- S. J. Elledge, Z. Zhou and J. B. Allen, *Trends Biochem. Sci.*, 1992, 17, 119–123.
- 3 A. F. Parisi and B. L. Vallee, *Am. J. Clin. Nutr.*, 1969, **22**, 1222–1239.
- 70 4 C. C. W. Hsia, N. Engl. J. Med., 1998, 338, 239–247.
- D. S. Kalinowski, C. Stefani, S. Toyokuni, T. Ganz, G. J. Anderson, N. V. Subramaniam, D. Trinder, J. K. Olynyk, A. Chua, P. J. Jansson, S. Sahni, D. J. R. Lane, A. M. Merlot, Z. Kovacevic, M. L. H. Huang, C. S. Lee and D. R. Richardson, *Biochim. Biophys. Acta Mol. Cell Res.*, 2016, 1863, 727–748.
- H. S. Kim, Y. J. Kim and Y. R. Seo, *J. Cancer Prev.*, 2015, 20, 232–240.
- 7 M. O. Huff, S. L. Todd, A. L. Smith, J. T. Elpers, A. P. Smith, R. D. Murphy, A. S. Bleser-Shartzer, J. E. Hoerter, B. N. Radde and C. M. Klinge, *Toxicol. Sci.*, 2016, **152**, 62–71.
- 8 W. H. Watson and J. D. Yager, *Toxicol. Sci.*, 2007, **98**, 1–4
- 85 9 Y. Wang, J. Fang, S. S. Leonard and K. M. K. Rao, Free Radic. Biol. Med., 2004, 36, 1434–1443.
  - T. K. Hei and M. Filipic, Free Radic. Biol. Med., 2004, 37, 574–581.
- 11 A. Hartwig, M. Asmuss, I. Ehleben, U. Herzer, D.
- Kostelac, A. Pelzer, T. Schwerdtle and A. Bürkle, Environ. Health Perspect., 2002, 110, 797–799.
  - 12 A. Lützen, S. E. Liberti and L. J. Rasmussen, Biochem. Biophys. Res. Commun., 2004, 321, 21–25.
- 13 A. Hartwig, Cadmium and cancer, 2013, vol. 11.
- 95 14 G. Y. L. Lui, Z. Kovacevic, V. Richardson, A. M. Merlot, D. S. Kalinowski and D. R. Richardson, Oncotarget, 2015, 6, 18748–18779.
  - D. J. R. Lane, T. M. Mills, N. H. Shafie, A. M. Merlot, R. Saleh Moussa, D. S. Kalinowski, Z. Kovacevic and D. R. Richardson, *Biochim. Biophys. Acta - Rev.*
- Cancer, 2014, **1845**, 166–181.

  N. Déliot and B. Constantin, *Biochim. Biophys. Acta*
- Biomembr., 2015, **1848**, 2512–2522.

  V. Thakur and B. Bedogni, *Pharmacol. Res.*, 2016,
- 17 V. Thakur and B. Bedogni, *Pharmacol. Res.*, 2016, 105 111, 17–22.
  - L. Fouani, S. V. Menezes, M. Paulson, D. R. Richardson and Z. Kovacevic, *Pharmacol. Res.*, 2017, 115, 275–287.
- 19 A. Jemal, F. Bray, M. M. Center, J. Ferlay, E. Ward and D. Forman, *CA. Cancer J. Clin.*, 2011, **61**, 69–90.

- 20 T. García-Barrera, J. L. Gómez-Ariza, M. González-Fernández, F. Moreno, M. A. García-Sevillano and V. Gómez-Jacinto, *Anal. Bioanal. Chem.*, 2012, 403, 2237–2253.
- 5 21 K. Zabłocka-Słowińska, S. Płaczkowska, A. Prescha, K. Pawełczyk, I. Porębska, M. Kosacka, L. Pawlik-Sobecka and H. Grajeta, J. Trace Elem. Med. Biol., 2018, 45, 78–84.
- 22 K.-Y. Lee, P.-H. Feng, H.-C. Chuang, S.-M. Wu, W.-T. Liu, K.-Y. Chen, C.-Y. Liu and S.-C. Ho, *Biol. Trace Elem. Res.*, 2017, **182**, 14–20.
- P. Sarita, G. J. N. Raju, M. R. Kumar, A. S. Pradeep and S. B. Reddy, *J. Radioanal. Nucl. Chem.*, 2013, 297, 431–436.
- 15 24 Y. Ren, Z. Zhang, Y. Ren, W. Li, M. Wang and G. Xu, *Talanta*, 1997, **44**, 1823–1831.
  - Y. Jin, C. Zhang, H. Xu, S. Xue, Y. Wang, Y. Hou, Y. Kong and Y. Xu, *Cancer Epidemiol.*, 2011, 35, 182–187.
- 20 26 L.-L. Zhang, F.-S. Wei and G.-P. Wu, in 3rd International Conference on Bioinformatics and Biomedical Engineering, iCBBE 2009, 2009.
- J. Y. Kim, H. B. Lim and M. H. Moon, *Anal. Chem.*,
   2016, 88, 10198–10205.
- 25 28 C. Tan, H. Chen and T. Wu, Biol. Trace Elem. Res., 2011, 142, 18–28.
  - C. Tan, H. Chen and C. Xia, J. Pharm. Biomed. Anal., 2009, 49, 746–752.
- Y. Benderli Cihan and S. Öztürk Yildirim, *Biol. Trace Elem. Res.*, 2011, **144**, 272–294.
- 31 C. Harlyk, J. Mccourt, G. Bordin, A. R. Rodriguez and A. Van Der Eeckhout, *J. Trace Elem. Med. Biol.*, 1997, **11**, 137–142.
- P. Censi, P. Zuddas, L. A. Randazzo, E. Tamburo, S. Speziale, A. Cuttitta, R. Punturo, P. Aricò and R. Santagata, *Environ. Sci. Technol.*, 2011, **45**, 6262–6267.
- 33 A. Escribano Montaner and A. Moreno Galdó, *An. Pediatría*, 2005, **62**, 352–366.
- 40 34 E. Bargagli, F. Monaci, N. Bianchi, C. Bucci and P. Rottoli, *Biol. Trace Elem. Res.*, 2008, **124**, 225–235.
- S. Atukorala, T. K. Basu, J. W. Dickerson, D. Donaldson and A. Sakula, *Br. J. Cancer*, 1979, 40, 927–931.
- 45 36 B. F. Issell, B. V. Macfadyen, E. T. Gum, M. Valdivieso, S. J. Dudrick and G. P. Bodey, *Cancer*, 1981, 47, 1845–1848.
  - 37 M. Zowczak, M. Iskra, J. Paszkowski, M. Manczak, L. Torlinski and E. Wysocka, J. Trace Elem. Med. Biol., 2001, 15, 193–196.
- 38 K. Suzuki, Y. Yamaya, N. Kanzawa, M. Chiba, K. Sera and R. Asano, *Biol. Trace Elem. Res.*, 2008, **124**, 92, 96
- E. A. Maier, A. Dietemann-Molard, F. Rastegar, R. Heimburger, C. Ruch, A. Maier, E. Roegel and M. J. Leroy, *Clin. Chem.*, 1987, **33**, 2234–2239.
- 40 R. González-Domínguez, T. García-Barrera and J. L. Gómez-Ariza, *BioMetals*, 2014, **27**, 539–549.
- 41 R. González-Domínguez, T. García-Barrera and J. L. Gómez-Ariza, *Metallomics*, 2014, **6**, 292–300.
- 42 C. S. Muñiz, J. L. Fernández-Martin, J. M. Marchante-Gayón, J. I. G. Alonso, J. B. Cannata-Andía and A. Sanz-Medel, *Biol. Trace Elem. Res.*, 2001, **82**, 259–272.

- 65 43 I. E. León, J. F. Cadavid-Vargas, A. L. Di Virgilio and S. Etcheverry, *Curr. Med. Chem.*, 2016, **23**.
  - 44 M. Le, O. Rathje, A. Levina and P. A. Lay, *J. Biol. Inorg. Chem.*, 2017, 22, 663–672.
- 45 G. N. Schrauzer, D. A. White and C. J. Schneider, *Bioinorg. Chem.*, 1977, **7**, 35–56.
- 46 M. Clementino, X. Shi and Z. Zhang, *Curr. Opin. Toxicol.*, 2018, **8**, 20–27.
- 47 M. K. Schwartz, Cancer Res., 1975, 35, 3481–3487.
- 48 J. D. Aguirre and V. C. Culotta, *J. Biol. Chem.*, 2012, **287**, 13541–13548.
- 49 G. F. Nordberg, A. Bernard, G. L. Diamond, J. H. Duffus, P. Illing, M. Nordberg, I. A. Bergdahl, T. Jin and S. Skerfving, *Pure Appl. Chem.*, 2018, 90, 755–808.
- 80 50 P. Richter, O. Faroon and R. S. Pappas, *Int. J. Environ. Res. Public Health*, 2017, **14**, 1–18.
  - 51 (2009 Rep. Updat. Febr. 2015), 2009.
  - 52 L. W. Oberley, *Biomed. Pharmacother.*, 2005, **59**, 143–148.
- 85 53 V. L. Kinnula and J. D. Crapo, *Am. J. Respir. Crit. Care Med.*, 2003, **167**, 1600–1619.
  - 54 C. B. Ambrosone, J. L. Freudenheim, P. A. Thompson, E. Bowman, J. E. Vena, J. R. Marshall, S. Graham, R. Laughlin, T. Nemoto and P. G. Shields, *Cancer Res.*, 1999, 59, 602–606.
- 55 K. Mitrunen, P. Sillanpää, V. Kataja, M. Eskelinen, V.-M. Kosma, S. Benhamou, M. Uusitupa and A. Hirvonen, *Carcinogenesis*, 2001, **22**, 827–829.
- M. Bergman, M. Ahnström, P. P. Wegman and S.
   Wingren, J. Cancer Res. Clin. Oncol., 2005, 131, 439–444.
- 57 K. Suzuki, H. Higuchi, H. Iwano, J. Lakritz, K. Sera, M. Koiwa and K. Taguchi, *Biol. Trace Elem. Res.*, 2012, 145, 166–171.
- 100 58 G. S. Andrews, *J. Clin. Pathol.*, 1979, **32**, 325–333.
  - 59 M. Díez, F. J. Cerdà, M. Arroyo and J. L. Balibrea, Cancer, 1989, 63, 726–730.
  - 60 V. Voyatzoglou, T. Mountokalakis, V. Tsata-Voyatzoglou, A. Koutselinis and G. Skalkeas, Am. J. Surg., 1982, 144, 355–358.
  - 61 N. Wiernsperger and J. Rapin, *Diabetol. Metab.* Syndr., 2010, **2**.

110