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PROTEOLYSIS DURING MANUFACTURE AND RIPENING/STORING OF “OLOMOUCKÉ TVARUŽKY” CHEESE (PGI)

Vendula Pachlová^{*1}, František Buňka¹, Leona Buňková²

Address(es): Ing. Vendula Pachlová, Ph.D.

¹Tomas Bata University in Zlín, Faculty of Technology, Department of Food Technology, nám. T. G. Masaryka 5555, 760 01, Zlín, Czech Republic, phone number: +420576033007.

²Tomas Bata University in Zlín, Faculty of Technology, Department of Environment Protect Engineering, nám. T. G. Masaryka 5555, 760 01, Zlín, Czech Republic.

*Corresponding author: pachlova@ft.utb.cz

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ABSTRACT

Twenty-two free amino acid (FAA) concentrations were observed during manufacture (1st, 3rd and 7th days of production) and ripening period (42 days storing at 8°C) of “Olomoucké tvaružky” (PGI, smear acid cheese). Sensory attributes were also analysed during ripening period. The free amino acids were determined by means of ion-exchange chromatography. The development of the individual FAA content positively correlated with the ripening period ($r = 0.7734-0.9229$; $P < 0.01$). The results gave information about the development precursors (FAA) of typically sensory active compound in “Olomoucké tvaružky” (PGI) during its production and especially ripening. In conclusion, we found that free amino acid concentration as finally products of proteolysis are positive with improved flavour.

Keywords: Proteolysis, free amino acid, cheese, ripening

INTRODUCTION

„Olomoucké tvaružky“ is a special type of Czech soft smear-ripening acid cheese with Protected Geographical Indication (PGI). “Olomoucké tvaružky” (PGI) gains its distinctive flavour by means of the microbial enzymatic apparatus. The intensity of the flavour depends on the maturity degree – ranging from mild to distinctive, piquant and even pungent. The microflora responsible for the above-mentioned changes in flavour also makes the typical gold to yellowish/orange smear on the surface of this cheese (Palencia *et al.* 2004, Williams, *et al.*, 2004, Rattray and Fox, 1999).

Standard curd (moisture content 68–66 % w/w) with modified acidity within (120–140°SH) is mixed with a pure dairy culture (complex consortium consisted especially of *Brevibacterium linens*, *Candida valida*, *Pediococcus acidilactici*) and formed into a desired shape. Within 2 to 4 days (called “drying” at ~20°C), the growth of microflora and formation of yeast film on the surface of the cheese can be observed. Subsequently, the surface of the cheese is rinsed with water. The cheeses treated this way are left to ripen for another 2 to 4 days to allow a sufficient growth of smear culture (mainly *Brevibacterium linens*). After the ripening, the cheeses are wrapped into semi-permeable clingfilm and are left to ripen at a temperature of <10°C (usually in chain stores).

Smear cheese typically ripens from the surface in towards the middle. Apart from the basic mesophilic culture of lactic acid bacteria (LAB – coming from the raw material – acid curd), aerophilic smear culture is also involved in the ripening (Fox *et al.*, 2004). The surface of the cheese is covered with a gold/yellowish smear coat which contains microorganisms of various origin (starters – SLAB and also non-starters – NSLAB), including coryneform bacteria (*Brevibacterium*), gram-positive cocci (*Lactococcus*) and yeasts (*Torulopsis*, *Candida*, *Oospora*). *Pediococcus acidilactici* is often used as protective culture which decreases the growth of many contaminating microorganisms primarily due to the production of lactic acid and secretion of bacteriocins. The growth of yeast within “drying” (2 to 4 days after the formation of curd) leads to neutralisation of the cheese surface (increase in pH) due to oxidation of lactic acid to CO₂ and water. The decrease in acidity of the cheese surface and removal of the yeast film (by rinsing with water) stimulate the subsequent growth of the desired smear bacteria (Irlinger *et al.*, 2012; Bockelmann *et al.*, 2005). During the ripening period of “Olomoucké tvaružky”, a significant increase in *Brevibacterium linens*, which forms a predominant part of the microflora, can be observed on the surface of the cheese (Rattray and Fox, 1999). *B. linens* produces orange-brown pigments and causes, together with other microorganisms (including NSLAB), the characteristic colouring of the cheese surface (Fox *et al.*,

2004). The proteolytic apparatus of the microflora present affects the release of free amino acids (FAA), which are important precursors for the subsequent flavour development. *B. linens* has a very active proteolytic system which is involved in cheese ripening (Williams *et al.*, 2004). By means of transamination, FAA can be changed into corresponding α -ketoacids and subsequently converted into carboxylic acids, methylaldehyde, 2- and 3-methylbutanal and 2-methylpropanal, which have a noticeable effect on the flavour of the final product (Yvon and Rijen, 2001). Moreover, *B. linens* can degrade amino acids while producing ammonia, which is a typical reaction in smear cheese. Also, the flavour of smear cheese is significantly influenced by elimination reactions, e.g. of methionin into methanethiol with a subsequent conversion (oxidation) to dimethyl disulfide and dimethyl trisulfide (Yvon and Rijen, 2001, Sablé and Cottencaeu, 1999, Smit *et al.*, 2000, Smit *et al.* 2005). *B. linens* is capable of metabolism of cysteine or cystine to hydrogen sulfides. However, this activity is lower in comparison with the formation of dimethyl disulfide and dimethyl trisulfide (Rattray and Fox, 1999). *B. linens* together with yeasts and micrococci converse tyrosine to cresol (Yvon and Rijen, 2001). Furthermore, some FAA can directly influence the flavour.

The aim of this study was (i) to describe of proteolytic changes (by means of free amino acid content) during the production phase (7 days) and ripening/storing (42 days) of smear cheese “Olomoucké tvaružky” (PGI) and (ii) to perform a correlation analysis of the dependence of free amino acid content and the data from the sensory analysis on the ripening period.

MATERIAL AND METHODS

Cheese production and sampling

The samples of “Olomoucké tvaružky” (PGI) were randomly taken from three batches made by a traditional manufacturer (A.W. Ltd., Loštice, Czech Republic) during standard production. The manufacture protocol was same as in Pachlová *et al.* (2013). The basic raw material for the production of “Olomoucké tvaružky” (PGI) was acid curd (quark) from skimmed milk. The acidity of the quark was modified by the mixture of sodium bicarbonate and calcium carbonate in order to reach the values ranging between 120–140°SH.

Subsequently, the acid curd (quark) was standardised to reach the desired moisture content (66–68%). The mixture of curd was stirred with the addition of pure dairy culture (bacteria *Brevibacterium linens* and *Pediococcus acidilactici* and yeast culture *Candida valida*). Later on, the curd was formed into the shape of sticks (approx. 15 mm height, 15 mm width and 80 mm depth). After the

forming, the mass was dried at $22\pm 2^\circ\text{C}$ (80–85% RH) for 2 days (first stage of ripening; target moisture content 64–65%). During the first stage of ripening, yeast culture appeared on the surface of the cheese. This culture decreases the acidity of curd and thus modifies the conditions for optimum growth of *Brevibacterium linens*. Subsequently, the oxidation yeast culture (film of the surface microbiota) was removed from the cheese blocks by means of rinsing (water temperature of $15\text{--}17^\circ\text{C}$) (Fox et al., 2004). The rinsed blocks of cheese ripened for another 5 days in ripening boxes at a temperature of $18\text{--}20^\circ\text{C}$. The blocks were turned over every day in order to ensure optimum growth of aerobic proteolytic microflora covering the whole surface of the cheese. The actual sampling of the intermediate products of the cheeses was performed on the 1st (after the forming of the blocks, sample code 1P), 3rd (after drying, sample code 3P) and 7th day (after packing, sample code 7P) after the start of the production. Furthermore, a ripening/storage experiment was started from the point of cheese packing. The cheeses ripened/were stored at $8\pm 2^\circ\text{C}$ (which corresponds to the standard process in retail). The sampling was performed on the 1st, 7th, 14th, 21st, 28th, 35th and 42nd day of storage (after production, sample codes 1S, 7S, 14S, 21S, 28S, 35S and 42S). The whole experiment was done with 3 batches which were the same as those in the sampling of intermediate products.

Determination of the free amino acid content

The content of twenty-two free amino acids and their derivatives (aspartic acid, threonine, serine, asparagine, glutamic acid, glutamine, proline, glycine, alanine, valine, methionine, cysteine, isoleucine, leucine, tyrosine, phenylalanine, lysine, histidine, arginine, ornithine, citrulline, γ -aminobutyric acid) was determined by ion-exchange chromatography (AAA400 Amino Acid Analyser; Ingos, Prague, Czech Republic) as reported by Buňková et al. (2009) and Pachlová et al. (2011). The samples were lyophilised on the day of the collection. Each sample was extracted three times. Each extract was analysed twice. The reagents for the preparation, separation and analysis of the samples were obtained from Ingos (Prague, Czech Republic). The standards were purchased from Sigma Aldrich (St. Louis, MO, USA).

Sensory analysis

The sensory evaluation of the samples was performed by 24 selected assessors (employees and students of the Faculty of Technology, Tomas Bata University in Zlín) trained according to ISO 8586(2012). The samples of cheeses were evaluated in a sensory laboratory with booths. All the samples were labelled with codes and served randomly at $20\pm 1^\circ\text{C}$. Nine sensory attributes (consistency of the surface, colour of the surface, flavour, taste, salty, off-flavour, bitterness, sourness and the overall profile of the product) were assessed on a five-point ordinal scale. The sensory analysis was performed only during the ripening period (from 7th to 42nd day of storage).

Statistical analysis of the data

The Kruskal-Wallis and Wilcoxon tests were used to evaluate the following data: the results of the sensory analysis and the content of free amino acids (Agresti, 1984). Pearson's correlation coefficients (r) between the ripening period and free amino acid content and data obtained from the sensory analysis were employed for the evaluation of the results (only the data from the storage period from 7th to 42nd day were evaluated). The data obtained in the experiment were processed by means of Unistat 5.5 statistical software (Unistat Ltd., London, UK).

RESULTS AND DISCUSSION

Proteolytic processes, including the release of free amino acids from the protein matrix, result mainly from the activity of the enzymatic apparatus of the SLAB

and NSLAB present. The total FAA content was slightly increased within the 7-day production (from 1.8 g.kg^{-1} in 1P day to 4.2 g.kg^{-1} in 7P; $P<0.05$). The growth dynamics of the total FAA content increased during the 42-day storage period, when the total FAA content was 30.3 g.kg^{-1} in 42S of storage ($P<0.05$). The development of the individual FAA content within the production and ripening is shown in Figure 1 and Figure 2. During the production, there was an even increase in the content of the individual FAA (Figure 1; $P<0.05$). Within the production period (from 1P to 7P), the strongest release of lysine, γ -aminobutyric acid, valine, leucine and proline occurred ($P<0.05$). On the other hand, the arginine content was decreasing during the production ($P<0.05$) (Pachlová et al., 2013).

The development of FAA content within the ripening period was very slow in the first month of storage (Figure 2; $P<0.05$). However, after 4 weeks of ripening the intensity of FAA releasing accelerated significantly (Figure 2; $P<0.05$). Compared with sampling in 28S of ripening ($P<0.05$), the content of some FAA doubled or even tripled in the end of experiment (sampling in 42S). The biggest increase in the individual FAA content was observed in lysine, methionine, asparagine, glutamine, glutamic acid, leucine and valine (Figure 2). The development of the individual FAA content positively correlated with the ripening period ($r = 0.8612\text{--}0.9229$ for most of the FAA; $r = 0.7734$ for arginine; $P<0.01$).

The different development of arginine content (within the production as well as the ripening period) in comparison with the other FAA could lie in the use of arginine by lactic acid bacteria (LAB) for the production of energy while producing ammonia, ornithine and CO_2 at the same time (McSweeney and Sousa, 2000). Thus it can be supposed that two reactions were in progress at the same time: (i) release of arginine by means of SLAB and/or NSLAB peptidases; and simultaneously (ii) conversion of arginine causing energy production (Tonon and Lonvaud-Funel, 2000). This statement can be supported by a significant increase in ornithine content not only during the production but also within the ripening period ($P<0.05$). Also, low concentrations of citrulline – an intermediate product of the conversion of arginine to ornithine (Christensen et al., 1999) – were detected during the production and ripening period (Figures 1 and 2). Towards the end of the ripening process (between sampling in 28S and 42S), a slight increase in arginine content was observed in comparison with 1S–21S. This could be explained by the accelerating cell lysis and lowering amount of the microflora present (both SLAB and NSLAB) at an advanced stage of ripening resulting in a lower need for energy by this metabolic pathway (Law, 2010, Komprda et al., 2008, Bergamini et al., 2006).

Secondary cultures on surface of cheese especially *Brevibacterium linens* are responsible for intensive proteolysis in smear cheese. Free amino acids (FAA) as the final products of proteolysis are important precursors of sensory active compounds (Smit et al., 2000; Ardö, 2006; Pachlová et al., 2013). The main precursors of sensory active compounds are phenylalanine, tyrosine, tryptophan and branched amino acids such as leucine, isoleucine and valine. Significant amounts of these FAA could lead to the development of the distinctive odour of “Olomoucké tvarůžky” (PGI). Moreover, methionine and its derivatives – sulphur compounds, play an important role in the production of soft smear cheese aroma (Yvon and Rijnen, 2001, Pachlová et al., 2013). The main sensory active compounds derived from methionine are methional, methanethiol and its oxidation products such as dimethyldisulphide and dimethyltrisulfide. *Brevibacterium linens* is responsible for production of volatile sulphur compounds in smear cheese (Marilley and Casey, 2004, Bockelmann et al., 2005). FAA themselves can also influence the organoleptic properties: phenylalanine, methionine, valine etc. by their bitterish flavour and aspartic or glutamic acids by their sour flavour. Amino acids such as leucine and glycine can also affect sweet flavour.

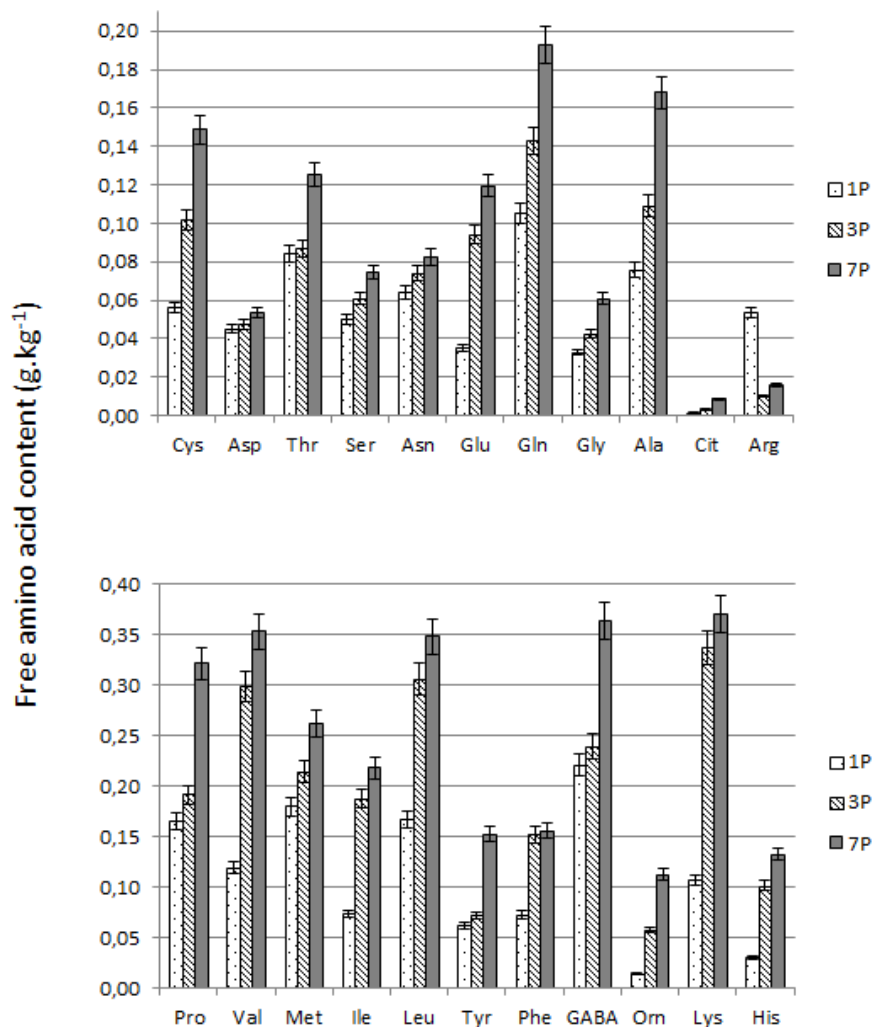


Figure 1 Development of individual free amino acid contents (g·kg⁻¹) during production of “Olomoucké tvarůžky” cheese (PGI): 1P – 1st day of production, 3P – 3rd day of production and 7P – 7th day of production. The values of amino acid content are expressed as means; the bars represented S.D. (n = 18).

The development of organoleptic properties (including the correlation coefficients *r*) during the ripening period of “Olomoucké tvarůžky” (PGI) is illustrated in Table 1. The intensity of flavour and taste was increasing during the storage period ($P < 0.05$). In the beginning of ripening (from sampling in 1S to 14S of storage), the cheese samples were most often evaluated as salty and sour. This evaluation corresponds to the character of “Olomoucké tvarůžky” (PGI) and production technology. During the ripening period, slight degradation in consistency of the surface occurred, which might have partly been caused by ripening in the wrapping foil. The micro-environmental conditions created under the wrapping within the ripening period might have influenced the smear culture on the surface of the cheese. On the other hand, the ripening wrapping adequately

protects the product against secondary contamination and water loss from the top layers of the cheese, which would otherwise get dry. The overall profile of the product was improving with the stage of ripening. Within the storage period of 21S to 35S, the product was evaluated as good and in the end of experiment (sampling in 42S) it was classified as very good. On the other hand, the assessors noticed slight bitterness within the ripening period. Mainly the low-molecular casein hydrolysates have a tendency to turn bitter, which is caused by a higher content of hydrophobic amino acids (Yvon and Rijen, 2001). The development of cheese bitterness could have also been affected by the content of phenylalanine, which was increasing dramatically from the 35th day of ripening (35S).

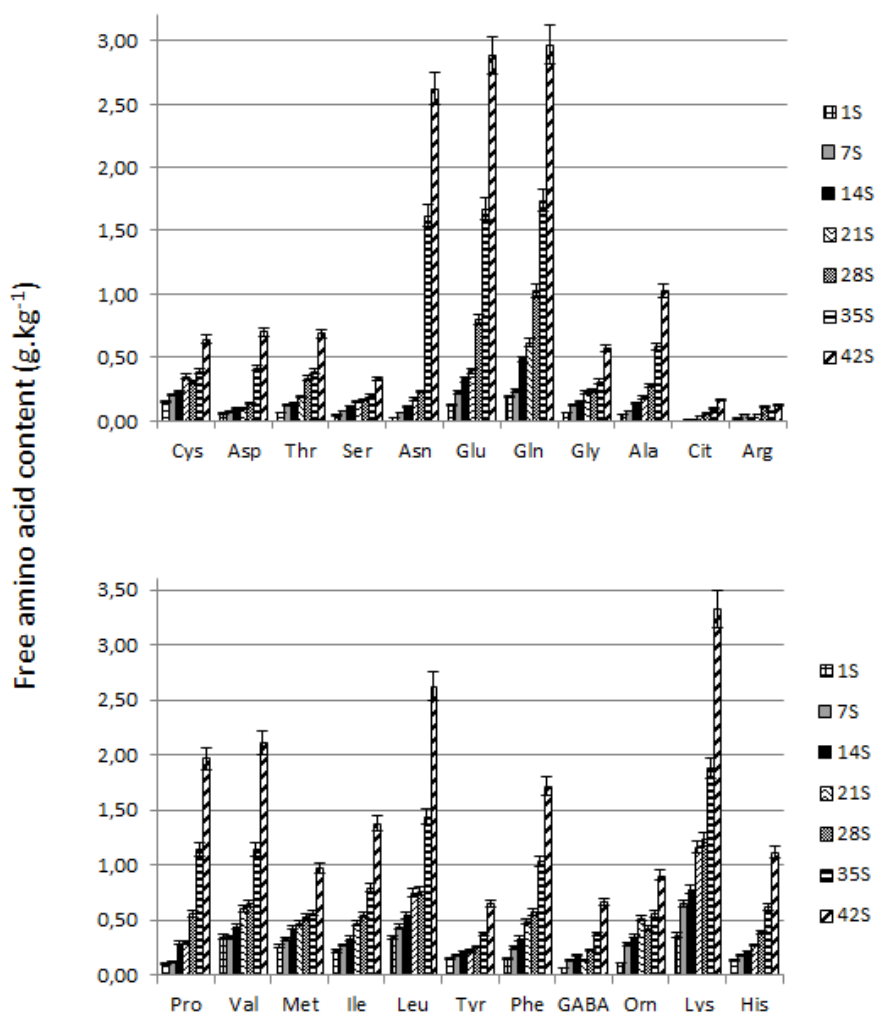


Figure 2 Development of individual free amino acid contents (g.kg⁻¹) during ripening of “Olomoucké tvarůžky” cheese (PGI; 0R–42R). The values of amino acid content are expressed as means; the bars represented S.D. (n = 18).

Table 1 Results (expressed as median) of the sensory analysis of tested “Olomoucké tvarůžky” cheese (PGI) during cheese ripening (7–42 days; 7R–42R)*

Sensory attribute	Storage period (days)						Correlation coefficient (r) **
	7S	14S	21S	28S	35S	42S	
Consistency of the surface	1	1.5	2	2	2	2	0.8305
Colour of the surface	2	1.5	2	2	2	2	0.3928
Flavour	2	2	2	3	3	3	0.8281
Taste	1	1.5	2	3	3	3	0.9411
Salty	2	1	1	1	1	2	-0.3928
Off-flavour	1	1	1	1	1	1	0.6547
Bitterness	1	1	1	1	1	2	0.6547
Sourness	3	3.5	3	2.5	2.5	3	-0.4971
Overall profile of the product	4	3.5	3	2.5	2.5	2	-0.9805

* Used ordinal scales: Consistency of the surface: 1 – smooth and uniform surface to 5 – absolutely unacceptable with large cracks or lumps. Colour of the surface: 1 – very light and white to 5 – very dark with atypical shades. Flavour: 1 – slight flavour to 5 – very strong flavour. Taste: 1 – insipid to 5 – very strong taste. Salty: 1 – very salty taste to 5 – salt-free. Off-flavour: 1 – no off-flavour to 5 – strong off-flavour. Bitterness: 1 – no bitterness to 5 – very strong intensity. Sourness: 1 – very sour taste to 5 – slightly acidic taste. Overall profile of the product: 1 – excellent to 5 – inconvenient.

** Correlation between the level of sensory attributes and the ripening period (in days).

CONCLUSION

This study describes the development of free amino acid content during the production and ripening of smear acid cheese “Olomoucké tvarůžky” (PGI). Moreover, changes in sensory attributes were observed during the ripening period. Within the production (sampling in 1P, 3P and 7P), the content of the individual FAA was increasing evenly. The only exception was arginine content, which was decreasing during the production (P<0.05). The development of the

individual FAA content was increasing during ripening and positively correlated with the ripening period. Changes in content of individual free amino acids are important due to possible formation of sensory active compounds during cheese ripening. The intensity of flavour and taste was increasing during the ripening period, mainly from the 28th day of ripening (28S). The overall profile of the product was improving with the stage of ripening, although the assessors noticed an increase in bitterness in the last week of ripening.

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