



**UNIVERSITI PUTRA MALAYSIA**

***QUANTUM MECHANICS SIMULATION OF CADMIUM(II) TRIPEPTIDE  
COMPLEXES***

**SHAHO MOHAMMED ABDALLA**

**FS 2017 73**



**QUANTUM MECHANICS SIMULATION OF CADMIUM(II)  
TRIPETIDE COMPLEXES**

By

**SHAHO MOHAMMED ABDALLA**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfillments of the Requirements for the Degree of  
Master of Science**

**May 2017**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
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**May 2017**

**Chairman : Professor Mohd Basyaruddin Bin Abdul Rahman, PhD**  
**Faculty : Science**

Cadmium detection in aqueous medium is an important step in attempt to avoid human exposure to the extremely toxic metal. It is believed that one of the significant qualitative detection processes of cadmium(II) can be performed using a biosensor with the help of small peptides as the biological material. The current issue is that proper peptides to capture cadmium(II) are still unknown, since there is not enough information about the interaction between the metal ion and most of the small peptides in literature. Quantum mechanics methods, such as Density Functional Theory (DFT), can be employed to understand the electronic interaction between cadmium(II) and the peptides. Minnesota 06 functional (M06) in combination with Default 2 triple zeta plus polarization (Def2TZVP) basis set was denoted as the best method among those, which were employed in this research to describe properties of several cadmium complexes, such as Cd-S bond length and S-Cd-S bond angle. For this purpose, data on cadmium(II) benzethiolato was extracted from Cambridge Structural Database (CSD) and compared to the computed data on the same molecule achieved theoretically using Becke Three Parameter Hybrid Functional in combination with Double zeta split-valence plus polarization basis set (B3LYP/DGDZVP) and M06/Def2TZVP. The results from the second method yielded 2.66% of errors for Cd-S bond lengths and 2.87% of errors for S-Cd-S bond angles, while in the presence of DGDZVP basis set in combination with B3LYP the errors rose up to 5.17% in Cd-S bond lengths and 4.90% in S-Cd-S bond angles. Cadmium(II) complexes with the small peptides, such as dipeptide, tripeptide and tetrapeptide were optimized employing M06/Def2TZVP and Polarizable Continuum Method (PCM) to determine the peptide length effect on the Cd-S binding energy. Cd-S binding energy in the tripeptide, if compared with the dipeptide and the tetrapeptide, was bigger as much as 12.62 kJ and 5.82 kJ respectively. Therefore, Cd-S binding energy of different tripeptide sequences were screened by fixing cysteine in the terminals and changing the middle amino acid to each one of the twenty essential amino acids. The optimization was performed in

vacuum using B3LYP/DGDZVP and M06/Def2TZVP methods. The procedures were repeated using PCM, in which water was chosen as the solvent, to investigate dielectric effect of water molecules on the Cd-S moiety. Cd-S binding energy of Cysteine-Proline-Cysteine (CPC) is the highest, if compared to the other observed nineteen tripeptides with the binding energies of 285.98 kJ in the presence of PCM using M06/Def2TZVP. The computed bond lengths between the metal ion and the sulfur atoms, using M06/Def2TZVP, are between 2.353 Å to 2.476 Å in vacuum, and 2.434 Å to 2.451 Å with PCM. Thus, CPC peptide could serve as biological material in the cadmium(II) biosensor application.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

## **SIMULASI MEKANIK KUANTUM KOMPLEKS KADMIUM (II) TRIPTEPIDA**

Oleh

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Pengesanan logam kadmium dalam medium akueus adalah satu langkah penting dalam usaha untuk mengelakkan pendedahan logam yang sangat toksik kepada manusia. Penderia bio merupakan salah satu teknik pengesanan kualitatif bagi kadmium(II) boleh dilakukan dengan menggunakan peptida pendek sebagai bahan biologi. Walaubagaimanapun maklumat berkaitan peptida yang sesuai untuk mengesan kadmium(II) masih tidak diketahui, kerana tidak ada maklumat yang cukup tentang interaksi antara ion logam dan peptida kecil dalam rujukan ilmiah. Kaedah Kuantum Mekanik (QM), seperti Teori Fungsi Ketumpatan (DFT) boleh digunakan untuk memahami interaksi elektronik di antara kadmium(II) dan peptida. Pemfungsian Minnesota 06 (M06) dengan kombinasi set asas Lalai 2 Ganda Tiga Zeta Tambah Polarisasi (Def2TZVP) didapati merupakan kaedah terbaik dalam kajian ini untuk menggambarkan sifat-sifat kompleks kadmium, seperti panjang ikatan Cd-S dan sudut ikatan S-Cd-S. Untuk tujuan ini, data mengenai kadmium(II) benzeniolato daripada Pangkalan Data Struktur Cambridge (CSD) telah digunakan dan dibandingkan dengan perkiraan data pada molekul yang sama secara teori menggunakan Pemfungsian Hibrid Tiga Parameter Becke dengan kombinasi set asas Ganda Dua Zeta Belah-Valens Tambah Polarisasi (B3LYP/DGDZVP) dan M06/Def2TZVP. Keputusan daripada kaedah kedua memberikan ralat sebanyak 2.66% bagi panjang ikatan Cd-S dan 2.87% bagi sudut ikatan S-Cd-S, manakala dengan menggunakan set asas DGDZVP dengan kombinasi B3LYP pula ralat didapati telah meningkat sehingga 5.17% bagi panjang ikatan Cd-S dan 4.90% bagi sudut ikatan S-Cd-S. Kompleks Kadmium(II) dengan peptida kecil, seperti dipeptida, tripeptida dan tetrapeptida telah dioptimumkan menggunakan M06/Def2TZVP dan Kaedah Pengkutuban Berterusan (PCM) untuk menentukan kesan panjang ikatan peptida terhadap tenaga ikatan Cd-S. Tenaga pengikatan logam bagi tripeptida, jika dibandingkan dengan dipeptida dan tetrapeptida didapati lebih besar sehingga 12.62 kJ dan 5.82 kJ, masing-masing. Oleh itu, tenaga pengikatan logam Cd-S bagi tripeptida yang berbeza telah disaring dengan menetapkan sistina

pada kedua-dua terminal dan menukar asid amino tengah bagi tengah dengan 19 asid amino penting. Prosedur tersebut diulang menggunakan PCM, di mana air telah dipilih sebagai pelarut, untuk mengkaji kesan dielektrik molekul air pada moiety Cd-S. Tenaga pengikatan Cd-S bagi Sistina-Prolina-Sistina (CPC) adalah yang tertinggi jika dibandingkan dengan 19 tripeptida lain dengan tenaga ikatan 285.98 kJ dalam PCM dengan menggunakan M06/Def2TZVP. Pengiraan panjang ikatan di antara ion logam dan atom-atom sulfur menggunakan M06/Def2TZVP memberikan nilai di antara 2.353 Å hingga 2.476 Å dalam vakum, dan 2.434 Å hingga 2.451 Å dalam PCM. Justeru, peptida CPC boleh dijadikan sebagai bahan biologi dalam aplikasi penderia bio kadmium(II).



## ACKNOWLEDGEMENTS

I would first like to express my gratitude to my supervisory committee chairman, Professor Dr. Mohd Basyaruddin Abdul Rahman (Department of Chemistry, Faculty of Science, UPM) as well as to my respectable co-supervisor, Dr. Mohd Farid Bin Ismail (Department of Chemistry, Faculty of Science, UPM) for their expert, sincere, and valuable guidance during the process of writing this thesis. Their timely and scholarly advice, meticulous scrutiny and scientific approach have helped me to a very great extent to accomplish this research.

I wish to extend my deepest gratefulness to Dr. Muhammad Alif Mohammad Latif, Zalikha Binti Ibrahim, Siti Nor Zulaikha Binti Omar, Lim Wui Zhuan and the rest of my friends from the Macromolecular Simulation laboratory for their passionate participation and input throughout the course of study in UPM.

Finally, I must express my profound appreciation to my family for providing me with unceasing support and continuous encouragement throughout my years of study and through the process of researching and writing this thesis. Thank you.



I certify that a Thesis Examination Committee has met on 8 May 2017 to conduct the final examination of Shaho Mohammed Abdalla on his thesis entitled "Quantum Mechanics Simulation of Cadmium(II) Tripeptide Complexes" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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## LIST OF ABBREVIATIONS

ATSDR	Agency for Toxic Substances and Disease Registry
IARC	International Agency for Research on Cancer
ICP-MS	Inductively coupled plasma mass spectrometry
GFAAS	Graphite furnace atomic absorption spectroscopy
ESIMS	Electrospray Ionization Mass Spectrometry
NMR	Nuclear Magnetic Resonance
ROS	Reactive oxygen species
CD	Circular dichroism
MT	Metallothionein
DFT	Density functional theory
HF	Hartree-Fock
CPCM	Conductor-like Polarizable Continuum Model
PCM	Polarizable continuum model
CSD	Cambridge structural data base
IUPAC	International Union of Pure and Applied Chemistry
ISFET	Ion-sensitive field-effect transistor
ISE	Ion selective electrode
ENFET	Enzyme field-effect transistor
IMFET	Immunological field-effect transistor
B3LYP	Becke Three Parameter Hybrid Functional
M06	Minnesota 06 functional
Def2TZVP	Default 2 triple zeta plus polarization basis set
Def2SVP	Default 2 Split Valence plus Polarization
DGDZVP	Double zeta split-valence plus polarization basis set

Cys	Cysteine
His	Histidine
Ile	Isoleucine
Met	Methionine
Ser	Serine
Val	Valine
Ala	Alanine
Gly	Glycine
Leu	Leucine
Pro	Proline
Thr	Threonine
Arg	Arginine
Asn	Asparagine
Asp	Aspartate
Glu	Glutamate
Gln	Glutamine
Phe	Phenylalanine
Tyr	Tyrosine
Trp	Tryptophan
Lys	Lysine

## CHAPTER 1

### INTRODUCTION

Cadmium lies at the end of the second row of transition metals in the periodic table. Since it has filled 'd' shell orbitals, it has a variety of coordination systems such as three coordination numbers (Matzapetakis *et al.*, 2002), four coordination numbers (Shindo & Brown, 1965), and six coordination numbers (Barrie *et al.*, 1993). Cadmium(II) is the most abundant oxidation state of cadmium ions in nature, although cadmium(I) oxidation state is recorded by dissolving cadmium in a mixture of cadmium chloride and aluminum chloride (Holleman *et al.*, 1985). Cadmium is widely used by people due to its applicability, for instance, in industry as Ni-Cd batteries and coloring agents, and in agriculture as phosphate fertilizers (McLaughlin & Singh, 1999). In addition, semiconductor cadmium chalcogenide nanocrystals (CdS, CdSe, and CdTe) have recently emerged as attractive materials for biological probes, since they have unique photo-physical characteristics (Derfus *et al.*, 2004). Consequently, the number of sources of cadmium(II) exposure continues to increase around human, and the high level of cadmium(II) contamination in soil and crops has become a cause for concern, since it severely affects human health in case of overdose exposure.

Cadmium is ranked the seventh in the list of hazardous substances and environmental pollutants among 785 different chemicals (The ATSDR 2015 Substance Priority List, 2015). It is also classified by International Agency for Research on Cancer (IARC) as a human carcinogen causing tumors of lungs, prostate, injection site, and other tissues (Waalkes, 2003). Chronically it affects kidneys and damages their function (Lars Järup *et al.*, 1998). Human absorbs cadmium either by ingestion or inhalation. However, an average non-smoker German citizen has a daily intake of 30 – 35 µg cadmium; 95% of this intake comes from food and drinks, but this range increases in the case of smoking (Godt *et al.*, 2006). It enables us to draw a conclusion, that cadmium(II) detection in aqueous medium is a crucial step to avoid its toxicity toward human since human gets the toxic metal ion significantly from drinks. As it is suggested by scholars, one of the significant detection processes of cadmium(II) could be done by using biosensor with the help of peptides as the biological material. Cadmium(II) can be detected by several other methods as well, for instance, Graphite furnace-atomic absorption spectrometry (GFAAS) (Ashraf, 2012) and inductively coupled plasma mass spectrometry (ICP-MS) (A. Sigel *et al.*, 2013). However, biosensor detection technique is cheaper and faster in comparison with the previous methods, if a qualitative detection process is required.

Biosensor is composed mainly of three parts: a receptor, a transducer and a digital monitor. The one, which is dwelled upon in this research is the receptor. The receptor can be enzymes or nucleic acids or antibodies (Chambers *et al.*, 2002), or short peptides, used as the chelating agent to capture the substrates (Pavan & Berti, 2012).

However, proper biological materials to capture cadmium(II) are still under investigation. Steps in pursuit of such a chemical can give a significant result in serving the biosensor technology for detection of cadmium(II). In order to find a proper chemical for this purpose, a survey by the natural biological systems containing cadmium(II) can direct scientists toward prediction of a general structure to capture cadmium(II). Subsequently, computational chemistry methods can be employed to derive and predict the potential length and sequence of the detected structure from the natural systems theoretically.

Most of cadmium(II) in the living organs is bound to a small cysteine-rich, metal-binding protein called Metallothionein (MT) (Nordberg, 2004). MT was discovered in 1957 as a cadmium binding protein in horse kidney (Margoshes *et al.*, 1957). MT is known as a crucial molecule to protect human health from toxicity of cadmium due to its tendency to capture it (Klaassen *et al.*, 2009). MT is one of the existing proteins in human body. Cysteine amino acids frequently repeat in MT sequence. Since naturally it can capture cadmium via its sulfur atom in cysteine amino acids, a part of its sequence (including L-cysteine) as a small peptide can be observed theoretically as the base for deriving the most eligible biological materials to be used in the production of cadmium(II) biosensor technology.

### **1.1. Problem Statements**

Exact structural determination studies have not been published for the cadmium(II) complexes of the most common small peptide molecules (Sigel *et al.*, 2013). Therefore, the lack of information about the binding energy of cadmium(II) with small peptides and the cadmium(II) peptide complexes with the lowest energy conformation in literature demands further investigation to explain the probability of using small peptides in cadmium(II) biosensor technology. Cadmium(II) tendency to bind with cysteine amino acids in the peptides and proteins was proved experimentally (Sutherland *et al.*, 2011). Cysteine interacts with the metal ions via its sulfur atom, and the presence of cysteine in different peptides undoubtedly affects the metal binding energy with its sulfur atom. Thus, the process of binding of the metal with cysteine in different peptides needs further investigation and research.

### **1.2. Research Objectives**

The objectives of this research are as follows:

- To investigate the effect of the peptide length on Cd-S binding energy.
- To investigate the tripeptides sequence effect on Cd-S binding energy.
- To select a proper theoretical method for calculating bond length and bond angle of cadmium(II) complexes among those, which were used.

Having adequate information about this interaction in hand, the most feasible peptide can be proposed for further observation to be constructed to interact with cadmium(II) as the biological chelating agent in the biosensors for detection of cadmium(II). The information can also aid to identify some properties of proteins, which contain this metal ion. For these purposes, one needs to understand the stability of the cadmium(II) peptide complexes and to describe it based on the energy of the complex system. The questions, such as, how the cadmium(II)-peptide interaction region affects the whole complex system, how much the bond energy is and what the range of the bond length is, should be answered adequately and thoroughly.

To sum up, this research covers a theoretical study about the interaction between cadmium(II) and deprotonated cysteine using density functional theory (DFT) (Hohenberg & Kohn, 1964). Changes in the energy of the system can give us the idea of which reaction process is the most appropriate one by cadmium(II) in the medium. To denote the energy loss after forming the complex, the energy of cadmium(II), the energy of the peptide and the energy of the complex, in vacuum and considering water molecules' dielectric effect using polarizable continuum model (PCM), were calculated. The details about the mentioned methods are depicted and explained in chapter three.



## REFERENCES

- Aizawa, M. (1991). Principles and applications of electrochemical and optical biosensors. *Analytica Chimica Acta*, 250(C), 249–256. [https://doi.org/10.1016/0003-2670\(91\)85073-2](https://doi.org/10.1016/0003-2670(91)85073-2)
- Anna M. Carrie, M. D. W. and D. R. W. (1976). Thermodynamic considerations in co-ordination. Part XXII. Sequestering ligands for improving the treatment of plumbism and cadmiumism, (1012), 1012–1015.
- Arseniev, A., Schultze, P., Wörgötter, E., Braun, W., Wagner, G., Vasak, M., ... Wüthrich, K. (1988). Three-dimensional Structure of Rabbit Liver [Cd]Metallothionein-2a in Aqueous Solution Determined by Nuclear Magnetic Resonance. *Journal of Molecular Biology*, 201, 637–657. [https://doi.org/10.1016/0022-2836\(88\)90644-4](https://doi.org/10.1016/0022-2836(88)90644-4)
- Ashraf, M. W. (2012). Levels of heavy metals in popular cigarette brands and exposure to these metals via smoking. *TheScientificWorldJournal*, 2012, 729430. <https://doi.org/10.1100/2012/729430>
- Baldwin, G. S., Bailey, M. F., Shehan, B. P., Sims, I., & Norton, R. S. (2008). Tyrosine modification enhances metal-ion binding. *The Biochemical Journal*, 416(1), 77–84. <https://doi.org/10.1042/BJ20081059>
- Barrie, P. J., Gyani, A., Motevalli, M., & O'Brien, P. (1993). Solid-state cadmium-113 NMR studies on cadmium complexes with glycine, L-alanine, and L-cysteine. *Inorganic Chemistry*, 32(18), 3862–3867. <https://doi.org/10.1021/ic00070a016>
- Becke, A. D. (1993). Density-functional thermochemistry. III. The role of exact exchange. *The Journal of Chemical Physics*, 98(7), 5648. <https://doi.org/10.1063/1.464913>
- Belcastro, M., Marino, T., Russo, N., & Toscano, M. (2009). The role of glutathione in cadmium ion detoxification: coordination modes and binding properties--a density functional study. *Journal of Inorganic Biochemistry*, 103(1), 50–7. <https://doi.org/10.1016/j.jinorgbio.2008.09.002>
- Blindauer, C. A. (2008). Metallothioneins with unusual residues: Histidines as modulators of zinc affinity and reactivity. *Journal of Inorganic Biochemistry*, 102(3), 507–521. <https://doi.org/10.1016/j.jinorgbio.2007.10.032>
- Buck, R. P., & Lindner, E. (1994). Recommendations for nomenclature of ionselective electrodes. *Pure and Applied Chemistry*, 66(12), 2527–2536. <https://doi.org/10.1351/pac199466122527>

- Chambers, J. P., Arulanandam, B. P., Matta, L. L., Weis, A., & Valdes, J. J. (2002). <Biosensor Recognition Elements - Caister Academic Press.pdf>, 1–12.
- Chen, X., Chu, M., & Giedroc, D. P. (2000). Spectroscopic characterization of Co(II)-, Ni(II)-, and Cd(II)-substituted wild-type and non-native retroviral-type zinc finger peptides. *Journal of Biological Inorganic Chemistry*, 5(1), 93–101. <https://doi.org/10.1007/s007750050012>
- Cherifi, K., Reverend, B. D. Le, Loucheux, C., Varnagy, K., Kiss, T., Sovago, I., & Kozlowski, H. (1990). Transition metal complexes of L-cysteine containing Di- and tripeptides. *Journal of Inorganic Biochemistry*, 38(1), 69–80. [https://doi.org/10.1016/0162-0134\(90\)85008-K](https://doi.org/10.1016/0162-0134(90)85008-K)
- Covington, A. K. (1994). Terminology and conventions for microelectronic ion-selective field effect transistor devices in electrochemistry (IUPAC Recommendations 1994). *Pure and Applied Chemistry*, 66(3), 565–569. <https://doi.org/10.1351/pac199466030565>
- Cramer, C. J. (2004). DENSITY FUNCTIONAL THEORY. In *Essentials of Computational Chemistry* (p. 261). West Sussex: John Wiley & Sons Ltd.
- Cullen, D. C., Sethi, R. S., & Lowe, C. R. (1990). Multi-analyte miniature conductance biosensor. *Analytica Chimica Acta*, 231(C), 33–40. [https://doi.org/10.1016/S0003-2670\(00\)86394-1](https://doi.org/10.1016/S0003-2670(00)86394-1)
- Curtis D. Klaassen, Jie Liu, and B. A. D. (2009). Metallothionein Protection of Cadmium Toxicity. *Toxicol Appl Pharmacol.*, 100(2), 215–220. <https://doi.org/10.1016/j.pestbp.2011.02.012>. Investigations
- Derfus, a M., Chan, W. C. W., & Bhatia, S. N. (2004). Probing the Cytotoxicity of Semiconductor Quantum Dots, Supp. Info. *Nano Letters*, 4(1), 11–18. <https://doi.org/Doi 10.1021/Nl0347334>
- DeSilva, T. M., Veglia, G., Porcelli, F., Prantner, A. M., & Opella, S. J. (2002). Selectivity in heavy metal- binding to peptides and proteins. *Biopolymers*, 64(4), 189–197. article. <https://doi.org/10.1002/bip.10149>
- Dunbar, R. C., Steill, J. D., Polfer, N. C., & Oomens, J. (2009). Peptide length, steric effects, and ion solvation govern zwitterion stabilization in barium-chelated di- and tripeptides. *Journal of Physical Chemistry B*, 113(31), 10552–10554. <https://doi.org/10.1021/jp905060n>
- Erk, M., & Raspor, B. (2001). Anodic stripping voltammetry in the complexation study of the peptide Lys-Cys-Thr-Cys-Cys-Ala [56-61] MT I and cadmium: Application in determination of the complexing capacity and stability constant. *Journal of Electroanalytical Chemistry*, 502(1–2), 174–179. [https://doi.org/10.1016/S0022-0728\(01\)00356-4](https://doi.org/10.1016/S0022-0728(01)00356-4)

- Frisch M.J., Trucks G.W., Schlegel H.B., Scuseria G.E., Robb M.A., Cheeseman, J.R., Zakrzewski V.G., Montgomery J.A., Jr., Stratmann R.E, Burant J.C., Dapprich S., Millam J.M., Daniels A.D., K.N. Kudin, M.C. Strain, O. Farkas, J. Tomasi, V. Barone, M. Cossi, R. Cammi, B. Mennucci, C. Pomelli, C. Adamo, S. Clifford, J. Ochterski, G.A. Petersson, P.Y. Ayala, Q. Cui, K. Morokuma, D.K. Malick, A.D. Rabuck, K. Raghavachari, J.B. Foresman, J. Cioslowski, J.V. Ortiz, B.B. Stefanov, G. Liu, A. Liashenko, P. Piskorz, I. Komaromi, R. Gomperts, R.L. Martin, D.J. Fox, T. Keith, M.A. Al-Laham, C.Y. Peng, A. Nanayakkara, C. Gonzalez, M. Challacombe, P.M.W. Gill, B. Johnson, W. Chen, M.W. Wong, J.L. Andres, C. Gonzalez, M. Head-Gordon, E.S. Replogle and J.A. Pople, (2009). Gaussian 09W Tutorial. *An Introduction To Computational Chemistry Using G09W and Avogadro Software*, 34.
- Fuhr, B. J., & Rabenstein, D. L. (1973). Nuclear magnetic resonance studies of the solution chemistry of metal complexes. IX. Binding of cadmium, zinc, lead, and mercury by glutathione. *Journal of the American Chemical Society*, 95(21), 6944–6950. <https://doi.org/10.1021/ja00802a013>
- Glušič, M., Stare, J., Grdadolnik, J., & Vianello, R. (2013). Binding of cadmium dication to glutathione facilitates cysteine SH deprotonation: A computational DFT study. *Journal of Inorganic Biochemistry*, 119(NOVEMBER), 90–94. <https://doi.org/10.1016/j.jinorgbio.2012.11.004>
- Godbout, N., Salahub, D. R., Andzelm, J., & Wimmer, E. (1992). Optimization of Gaussian-type basis sets for local spin density functional calculations. Part I. Boron through neon, optimization technique and validation. *Canadian Journal of Chemistry*, 70(2), 560–571. <https://doi.org/10.1139/v92-079>
- Godt, J., Scheidig, F., Grosse-Siestrup, C., Esche, V., Brandenburg, P., Reich, A., & Groneberg, D. a. (2006). The toxicity of cadmium and resulting hazards for human health. *Journal of Occupational Medicine and Toxicology (London, England)*, 1, 22. <https://doi.org/10.1186/1745-6673-1-22>
- Hanwell, M. D., Curtis, D. E., Lonie, D. C., Vandermeersch, T., Zurek, E., & Hutchison, G. R. (2012). Avogadro: An advanced semantic chemical editor, visualization, and analysis platform. *Journal of Cheminformatics*, 4(8), 1–17. <https://doi.org/10.1186/1758-2946-4-17>
- Hohenberg, P., & Kohn, W. (1964). Inhomogeneous Electron Gas. *Phys. Rev.*, 136(3B), B864–B871. article. <https://doi.org/10.1103/PhysRev.136.B864>
- Holleman, A. F.; Wiberg, E; Wiberg, N. (1985). *Cadmium". Lehrbuch der Anorganischen Chemie*. Walter de Gruyter.
- Jacopo Tomasi, \*, †, Benedetta Mennucci, † and, & Cammi‡, R. (2005). Quantum Mechanical Continuum Solvation Models. <https://doi.org/10.1021/cr9904009>

- Jalilehvand, F., Leung, B. O., & Mah, V. (2009). Cadmium(II) complex formation with cysteine and penicillamine. *Inorganic Chemistry*, 48(13), 5758–5771. <https://doi.org/10.1021/ic802278r>
- Jiang, L. J., Vasák, M., Vallee, B. L., & Maret, W. (2000). Zinc transfer potentials of the alpha - and beta-clusters of metallothionein are affected by domain interactions in the whole molecule. *Proceedings of the National Academy of Sciences of the United States of America*, 97(6), 2503–2508. <https://doi.org/10.1073/pnas.97.6.2503>
- Kepp, K. P. (2012). Full quantum-mechanical structure of the human protein Metallothionein-2. *Journal of Inorganic Biochemistry*, 107(1), 15–24. <https://doi.org/10.1016/j.jinorgbio.2011.11.002>
- Krężel, A., & Bal, W. (1999). Coordination chemistry of glutathione. *Acta Biochimica Polonica*. p 567-580, Volume 46, Issue 3.
- Krzywoszynska K., Rowinska-Zyrek M., D. Witkowska, S. Potocki, M. L., & Kozłowski, H. (2011). Polythiol binding to biologically relevant metal ions. *Dalton Transactions*, 40(40), 10289–10303. <https://doi.org/10.1039/c1dt11184a>
- Lars Järup, Marika Berglund, Carl Gustaf Elinder, Gunnar Nordberg, & Marie Vanter. (1998). Health effects of cadmium exposure – a review of the literature and a risk estimate. *Scandinavian Journal of Work, Environment & Health*, 24(Supplement 1), 1–51.
- Lee, C., Yang, W., & Parr, R. G. (1988). Development of the Colle-Salvetti correlation-energy formula into a functional of the electron density. *Physical Review B*, 37(2), 785–789. <https://doi.org/10.1103/PhysRevB.37.785>
- Leszczyszyn, O. I., White, C. R. J., & Blindauer, C. A. (2010). The isolated Cys2His2 site in EC metallothionein mediates metal-specific protein folding. *Molecular bioSystems*, 6(9), 1592–603. <https://doi.org/10.1039/c002348e>
- Macias, M. J., Wiesner, S., & Sudol, M. (2002). WW and SH3 domains, two different scaffolds to recognize proline-rich ligands. *FEBS Letters*, 513(1), 30–37. [https://doi.org/10.1016/S0014-5793\(01\)03290-2](https://doi.org/10.1016/S0014-5793(01)03290-2)
- Margoshes, M., Vallee, B. L., Kojima, Y., Berger, C., Kägi, J. H. R., & Piscator. (1957). A cadmium protein from equine kidney cortex. *Proceedings of the »First International Meeting on Metallothionein and Other Low Molecular Weight Metal-Binding Proteins« Zürich, July 17–22, 1978*, 34(17), 378. <https://doi.org/10.1007/978-3-0348-6493-0>

- Matzapetakis, M., Farrer, B. T., Weng, T., Hemmingsen, L., Penner-hahn, J. E., Pecoraro, V. L., & Veterinary, R. (2002). Comparison of the Binding of Cadmium ( II ), Mercury ( II ), and Arsenic ( III ) to the de Novo Designed Peptides TRI L12C and, (Ii), 8042–8054.
- Matzapetakis, M., Ghosh, D., Weng, T. C., Penner-Hahn, J. E., & Pecoraro, V. L. (2006). Peptidic models for the binding of Pb(II), Bi(III) and Cd(II) to mononuclear thiolate binding sites. *Journal of Biological Inorganic Chemistry*, *11*(7), 876–890. <https://doi.org/10.1007/s00775-006-0140-7>
- McLaughlin, M. J., & Singh, B. R. (1999). Cadmium in soils and plants. In *Cadmium in soils and plants* (pp. 219–256). CHAP, Springer.
- Mendieta, J., Diaz-Cruz, M. S., Monjonell, A., Tauler, R., & Esteban, M. (1999). Complexation of cadmium by the C-terminal hexapeptide Lys-Cys-Thr-Cys-Cys-Ala from mouse metallothionein: Study by differential pulse polarography and circular dichroism spectroscopy with multivariate curve resolution analysis. *Analytica Chimica Acta*, *390*(1–3), 15–25. [https://doi.org/10.1016/S0003-2670\(99\)00181-6](https://doi.org/10.1016/S0003-2670(99)00181-6)
- Messerle, B. A., Schaffer, A., Vasak, M., Kagi, J. H. R., & Wuthrich, K. (1992). Comparison of the Solution Conformations of Human [Zn7]- Metallothionein-2 and [Cd7]-Metallothionein-2 Using Nuclear- Magnetic-Resonance Spectroscopy, *225*(2), 433–443. Retrieved from <Go to ISI>://1992HX08000014
- Mori, S., Endoh, T., Yaguchi, Y., Shimizu, Y., Kishi, T., & Yanai, T. K. (2011). Quantum chemical studies on the role of water microsolvation in interactions between group 12 metal species ( $\text{Hg}^{2+}$ ,  $\text{Cd}^{2+}$ , and  $\text{Zn}^{2+}$ ) and neutral and deprotonated cysteines. *Theoretical Chemistry Accounts*, *130*(2–3), 279–297. <https://doi.org/10.1007/s00214-011-0975-z>
- Muñoz, A., Laib, F., Petering, D. H., & Shaw, C. F. (1999). Characterization of the cadmium complex of peptide 49-61: A putative nucleation center for cadmium-induced folding in rabbit liver metallothionein IIA. *Journal of Biological Inorganic Chemistry*, *4*(4), 495–507. <https://doi.org/10.1007/s007750050335>
- Nambiar, S., & Yeow, J. T. W. (2011). Conductive polymer-based sensors for biomedical applications. *Biosensors and Bioelectronics*. Elsevier B.V. <https://doi.org/10.1016/j.bios.2010.09.046>
- Ngu, T. T., Sturzenbaum, S. R., & Stillman, M. J. (2006). Cadmium binding studies to the earthworm *Lumbricus rubellus* metallothionein by electrospray mass spectrometry and circular dichroism spectroscopy. *Biochemical and Biophysical Research Communications*, *351*(1), 229–233. <https://doi.org/10.1016/j.bbrc.2006.10.023>



- Nordberg, G. F. (2004). Cadmium and health in the 21st century--historical remarks and trends for the future. *Biometals : An International Journal on the Role of Metal Ions in Biology, Biochemistry, and Medicine*, 17(5), 485–489. <https://doi.org/10.1023/B:BIOM.0000045726.75367.85>
- Pavan, S., & Berti, F. (2012). Short peptides as biosensor transducers. *Analytical and Bioanalytical Chemistry*. <https://doi.org/10.1007/s00216-011-5589-8>
- Perrin, D. D., & Watt, A. E. (1971). Complex formation of zinc and cadmium with glutathione. *BBA - General Subjects*, 230(1), 96–104. [https://doi.org/10.1016/0304-4165\(71\)90057-2](https://doi.org/10.1016/0304-4165(71)90057-2)
- Poleć-Pawlak, K., Ruzik, R., & Lipiec, E. (2007). Investigation of Cd(II), Pb(II) and Cu(I) complexation by glutathione and its component amino acids by ESI-MS and size exclusion chromatography coupled to ICP-MS and ESI-MS. *Talanta*, 72(4), 1564–1572. <https://doi.org/10.1016/j.talanta.2007.02.008>
- Remko, M., Fitz, D., Broer, R., & Rode, B. M. (2011). Effect of metal Ions (Ni<sup>2+</sup>, Cu<sup>2+</sup> and Zn<sup>2+</sup>) and water coordination on the structure of L-phenylalanine, L-tyrosine, L-tryptophan and their zwitterionic forms. *Journal of Molecular Modeling*, 17(12), 3117–3128. <https://doi.org/10.1007/s00894-011-1000-0>
- Romero-Isart, N., Duran, N., Capdevila, M., González-Duarte, P., Maspoch, S., & Torres, J. L. (1998). Metal binding properties of three Cys2X2 (X = His, Asp) metallothionein-related peptides. *Inorganica Chimica Acta*, 278(1), 10–14. [https://doi.org/10.1016/S0020-1693\(97\)06168-9](https://doi.org/10.1016/S0020-1693(97)06168-9)
- Schulze-Gahmen, U., Bondt, H. L. De, & Kim, S.-H. (1996). High-Resolution Crystal Structures of Human Cyclin-Dependent Kinase 2 with and without ATP: Bound Waters and Natural Ligand as Guides for Inhibitor Design. *Chart*, 39, 4540–4546.
- Shindo, H., & Brown, T. L. (1965). Infrared Spectra of Complexes of L-Cysteine and Related Compounds With Zinc (II), Cadmium (II), Mercury (II), and Lead (II). *Journal of the American Chemical Society*, 87, 1904–1909.
- Shruthi GS, A. C., & Mathew, B. B. (2014). Biosensors: A Modern Day Achievement. *Journal of Instrumentation Technology*, 2(1), 26–39. <https://doi.org/10.12691/JIT-2-1-5>
- Sigel, A., Sigel, H., & Sigel, R. K. O. (2013). *Cadmium: From Toxicity to Essentiality*. Springer (Vol. 11). <https://doi.org/10.1007/978-94-007-5179-8>
- Sigel, H., & Martin, R. B. (1982). Coordinating properties of the amide bond. Stability and structure of metal ion complexes of peptides and related ligands. *Chemical Reviews*, 82, 385–426. <https://doi.org/10.1021/cr00050a003>

- Stellato, F., Menestrina, G., Serra, M. D., Potrich, C., Tomazzolli, R., Meyer-Klaucke, W., & Morante, S. (2006). Metal binding in amyloid ??-peptides shows intra- and inter-peptide coordination modes. *European Biophysics Journal*, 35(4), 340–351. <https://doi.org/10.1007/s00249-005-0041-7>
- Sutherland, D. E. K., & Stillman, M. J. (2011). The “magic numbers” of metallothionein. *Metallomics*, 3(5), 444. <https://doi.org/10.1039/c0mt00102c>
- Swenson, D., Baenziger, N. C., & Coucouvanis, D. (1978). Tetrahedral mercaptide complexes. Crystal and molecular structures of  $[(C_6H_5)_4P]_2M(SC_6H_5)_4$  complexes (M = cadmium(II), zinc(II), nickel(II), cobalt(II), and manganese(II)). *Journal of American Chemical Society*, 100(6), 1932–1934. <https://doi.org/10.1021/ja00474a053>
- The ATSDR 2015 Substance Priority List. (2015). Retrieved from <https://www.atsdr.cdc.gov/spl/>
- Thevenot Daniel R, Toth K., Durst R. A., W. G. S. (1999). ELECTROCHEMICAL BIOSENSORS : RECOMMENDED Electrochemical biosensors : Recommended de ® nitions and classi ® cation ( Technical Report ). *Pure Applied Chemistry*, 71(12), 2333–2348. [https://doi.org/10.1016/S0956-5663\(01\)00115-4](https://doi.org/10.1016/S0956-5663(01)00115-4)
- Varnagy Katalin. and Imre Sogova (2013). Cadmium(II) Complexes of Amino Acids and Peptides. In *Cadmium: From Toxicity to Essentiality* (pp. 296–297).
- Waalkes, M. P. (2003). Cadmium carcinogenesis. *Mutation Research - Fundamental and Molecular Mechanisms of Mutagenesis*. <https://doi.org/10.1016/j.mrfmmm.2003.07.011>
- Walker, M., Harvey, A. J. A., Sen, A., & Dessent, C. E. H. (2013). Performance of M06, M06-2X, and M06-HF density functionals for conformationally flexible anionic clusters: M06 functionals perform better than B3LYP for a model system with dispersion and ionic hydrogen-bonding interactions. *Journal of Physical Chemistry A*, 117(47), 12590–12600. <https://doi.org/10.1021/jp408166m>
- Weigend, F., & Ahlrichs, R. (2005). Balanced basis sets of split valence, triple zeta valence and quadruple zeta valence quality for H to Rn: Design and assessment of accuracy. *Physical Chemistry Chemical Physics*, 7(18), 3297–305. <https://doi.org/10.1039/b508541a>
- Wolfe, S. A., Grant, R. A., Elrod-Erickson, M., & Pabo, C. O. (2001). Beyond the “recognition code”: Structures of two Cys2His2 zinc finger/TaTa box complexes. *Structure*, 9(8), 717–723. [https://doi.org/10.1016/S0969-2126\(01\)00632-3](https://doi.org/10.1016/S0969-2126(01)00632-3)

Yue Yang, Michael N. Weaver, and Kenneth M. Merz Jr. (2010). Assessment of the '6-31+G\*\* + LANL2DZ Mixed Basis Set Coupled with Density Functional Theory Methods and Effective Core Potential: Prediction of Heats of Formation and Ionization Potentials for First Row Transition Metal Complexes. *J Phys Chem A*, 113(36), 9843–9851. <https://doi.org/10.1021/jp807643p>

Zhao, Y., & Truhlar, D. G. (2008). The M06 suite of density functionals for main group thermochemistry, thermochemical kinetics, noncovalent interactions, excited states, and transition elements: Two new functionals and systematic testing of four M06-class functionals and 12 other function. *Theoretical Chemistry Accounts*, 120(1–3), 215–241. <https://doi.org/10.1007/s00214-007-0310-x>