



UNIVERSITI PUTRA MALAYSIA

***EFFECTS OF Panax ginseng AND Eurycoma longifolia JACK
EXTRACTS
ON CHILLED AND FROZEN-THAWED CROSSBRED BULL SEMEN
COLLECTED USING MODIFIED ELECTRO-EJACULATION METHOD***

FALAH HASAN ALI BAIEE

FPV 2017 13



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By

FALAH HASAN ALI BAIEE

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of
Philosophy**

August 2017

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DEDICATION

I dedicate my dissertation work to my family.

In appreciation of their love, sacrifices, faith, and eternal goodness, I would like to dedicate my dissertation to my dear loving parents, Hasan and Salma.

I will always appreciate all they have done, especially my wife Ruaa' for helping me all the time throughout the entire doctoral program.

I dedicate this work and give special thanks to my best friends I mean my wonderful sons Ahmed and Mohammed Hasan for being there for me throughout the entire doctorate program.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Doctor of Philosophy

EFFECTS OF *Panax ginseng* AND *Eurycoma longifolia* JACK EXTRACTS ON CHILLED AND FROZEN-THAWED CROSSBRED BULL SEMEN COLLECTED USING MODIFIED ELECTRO-EJACULATION METHOD

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FALAH HASAN ALI BAIEE

August 2017

Chairman : Professor Abd Wahid b Haron, PhD
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Collection of semen from bulls can be done using different methods. Nowadays, electro-ejaculator technique is known as the most popular method to obtain semen samples from wild and domestic males. Chilled semen does not submit to the freeze-thaw procedure and suffers fewer damages, leading to greater capability and an improved capacity to fertilization. Cryopreservation of sperm plays a considerable role in economization of breeding programs in the cattle herd industry, genetic improvement of domestic animals, preservation of endangered species, and is clinically worthy in the controlling of infertility. Hence, the objectives of the present thesis were: to minimize the discomfort signs during semen collection using electro-ejaculator. Secondly, to evaluate the effect of *Panax ginseng* aqueous extract on the quality of chilled and frozen-thawed bull semen. Thirdly, to evaluate the effect of Tongkat Ali aqueous extract on the quality of chilled and frozen-thawed bull semen. Finally, to assess the synergistic effect of Tongkat Ali and *Panax ginseng* extracts on the quality of frozen-thawed bull semen. For all experiments, a total of 84 ejaculates were obtained from six crossbred bulls. The normal automatic electro-ejaculation method of semen collection (Method I) was compared to a modified method involving three stages (stage one, two and three) of gradation electrical stimulation (Method II). Discomfort signs, bulls' response to electro-ejaculator and fresh semen samples were assessed. Tris-egg yolk extender was used to dilute the semen sample. The extender was prepared into two parts, part one (P1) which did not contain glycerol while part two (P2) contained double amount of glycerol (12.8%). Each semen extender was divided into seven groups containing different concentrations either *Panax ginseng* aqueous extract, Tongkat Ali aqueous extract or combination between them. Chilled and frozen-thawed semen were carried out. Chilled semen groups were placed in test tube and kept in refrigerator at 5 °C for 6 days. The frozen semen were packaged in (0.25 mL French straws and cryopreserved in liquid nitrogen (-196 °C). Sperm motility, morphology, viability, membranes integrity,

DNA integrity, and lipid peroxidation were carried out to assess the extracts on chilled and frozen-thawed crossbred bulls. The data were analysed as mean \pm standard error of the mean and checked for normal distribution. Descriptive statistical analysis of data, independent samples *t*-test, one or two-way analysis of variance (ANOVA), Fisher's exact test, and chi square test were used to analyse the data. The results showed that the discomfort signs were reduced ($P < 0.05$), as well as using Method II than Method I. The total time taken for semen collection was similar in both methods. Also, there was no significant difference in fresh semen parameters. *Panax ginseng* aqueous extract did not improve the functional parameters of chilled and frozen-thawed bull semen. Moreover, sperm DNA integrity of frozen-thawed semen was not improved either. The low dosages (0.25 mg/mL) and (0.5 mg/mL) were not significant compared to control group. However, dosages more than (0.5 mg/mL) showed marked decrease in sperm characteristics ($P < 0.01$). Tongkat Ali aqueous extract significantly improved the chilled and frozen-thawed semen compared to control. The ideal dose of Tongkat Ali in chilled semen was (1 mg/mL) and (5 mg/mL) in frozen-thawed semen. DNA integrity and lipid peroxidation significantly improved in (5 mg/mL) compared to control group of frozen-thawed semen. In addition, different concentrations of combination between PGe and TAe into Tris-egg yolk extender did not improve the frozen-thawed bull semen quality. In conclusion, the discomfort signs prior and during semen collection were reduced by modification of the automatic mode of the electro-ejaculator device. *Panax ginseng* aqueous extract did not improve the quality of preserved semen. The quality of chilled and frozen-thawed crossbred bull semen was improved by adding Tongkat Ali aqueous extract to the semen diluent. The combination between *Panax ginseng* and Tongkat Ali extract also did not improve the quality of frozen-thawed bull semen.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**KESAN EKSTRAK *Panax ginseng* DAN *Eurycoma Longifolia*
KE ATAS SEMEN LEMBU JANTAN KACUKAN YANG DISEJUKKAN
DAN DIBEKU-CAIRKAN YANG DIKUMPULKAN SECARA
PENGUBAHSUAIAN KAEDAH ELEKTROEJAKULATOR**

Oleh

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Pengumpulan semen dari lembu jantan boleh dilakukan menggunakan kaedah yang berbeza. Pada masa kini, teknik elektro-ejakulator dikenali sebagai kaedah yang paling popular untuk mendapatkan sampel semen dari haiwan jantan liar dan belajinak. Semen yang disejukkan tidak melalui prosedur beku-cair dan mengalami kurang kerosakan, yang membawa kepada keupayaan yang lebih besar dan kapasiti yang lebih baik untuk persenyawaan. Kriopengawetan sperma memainkan peranan penting dalam mengekonomikan program pembiakan dalam industri kelompok lembu, pembaikan genetik haiwan domestik, pengawetan spesies terancam, dan secara klinikal adalah berguna untuk mengawal kemandulan. Oleh itu, objektif tesis ini pertamanya ialah untuk mengurangkan tanda ketidakselesaian semasa pengumpulan semen menggunakan elektro-ejakulator dan kedua, untuk menilai kesan ekstrak *Panax ginseng* (PGe) ke atas kualiti semen lembu jantan yang disejukkan dan dibeku-cairkan. Ketiga, ia adalah untuk menilai kesan ekstrak Tongkat Ali (TAe) ke atas kualiti semen lembu jantan yang disejukkan dan dibeku-cairkan. Akhirnya, ia adalah untuk menilai kesan sinergi ekstrak Tongkat Ali dan *Panax ginseng* ke atas kualiti semen lembu jantan yang dibeku-cairkan. Untuk semua eksperimen, sejumlah 84 hasil ejakulasi diperolehi daripada enam ekor lembu jantan kacukan. Kaedah automatik elektro-ejakulasi biasa untuk pengumpulan semen (Kaedah I) dibandingkan dengan suatu kaedah yang diubahsuai yang melibatkan tiga peringkat (peringkat satu, dua dan tiga) penggredan rangsangan elektrik (Kaedah II). Tanda-tanda ketidakselesaian, respons lembu jantan terhadap elektro-ejakulator dan sampel semen segar dinilai. Pengekal Tris-kuning telur digunakan untuk mencairkan sampel semen. Pengekal ini disediakan dalam dua bahagian, bahagian satu (P1) yang tidak mengandungi gliserol manakala bahagian dua (P2) mengandungi dua kali ganda jumlah gliserol (12.8%). Setiap pengekal semen dibahagikan kepada tujuh kumpulan yang mengandungi kepekatan yang berlainan sama ada ekstrak akueus

Panax ginseng, ekstrak akueus Tongkat Ali atau gabungan di antara kedua-duanya. Ujian ke atas semen yang disejukkan dan yang dibeku-cairkan telah dijalankan. Kumpulan semen sejuk diletakkan di dalam tabung uji dan disimpan di dalam peti sejuk pada 5 °C selama 6 hari. Semen beku disimpan di dalam straw Perancis 0.25 mL dan dikrioawetkan di dalam nitrogen cair (-196°C). Motiliti, morfologi, daya maju, integrity membran, dan integriti DNA sperma, serta ujian peroksidaan lipid telah dijalankan untuk menilai kesan ekstrak pada semen lembu jantan kacukan yang disejukkan dan yang dibeku-cairkan. Data dianalisis sebagai $\text{min} \pm \text{ralat piawai}$ min dan diperiksa untuk taburan normal. Analisis statistik deskriptif data, ujian-*t* sampel bebas, analisis varians satu atau dua-hala (ANOVA), ujian tepat Fisher, dan chi kuasa dua telah digunakan untuk menganalisis data. Hasil kajian menunjukkan bahawa tanda-tanda ketidakselesaian juga telah dikurangkan ($P < 0.05$) apabila menggunakan Kaedah II berbanding Kaedah I. Jumlah masa yang diambil untuk pengumpulan semen adalah sama bagi kedua-dua kaedah. Juga, tidak terdapat perbezaan yang signifikan bagi parameter semen segar. Ekstrak akueus *Panax ginseng* tidak memperbaiki parameter fungsi semen lembu jantan yang disejukkan dan dibeku-cairkan. Tambahan lagi, integriti DNA sperma semen yang dibeku-cairkan juga tidak bertambah baik. Dos yang rendah, 0.25 mg/mL dan 0.5 mg/mL tidak signifikan berbanding dengan kumpulan kawalan. Walau bagaimanapun, dos lebih daripada 0.5 mg/mL menunjukkan penurunan ketara dalam ciri-ciri sperma ($P < 0.01$). Ekstrak akueus Tongkat Ali dengan ketara memperbaiki semen yang disejukkan dan yang dibeku-cairkan berbanding kawalan. Dos Tongkat Ali yang terbaik bagi semen disejukkan adalah 1 mg/mL dan 5 mg/mL bagi semen dibeku-cairkan. Integriti DNA dan peroksidaan lipid bertambah baik dengan ketara di dalam 5 mg/mL berbanding dengan kumpulan kawalan semen dibeku-cairkan. Di samping itu, kepekatan yang berbeza bagi gabungan antara PGe dan TAe ke dalam pengekal Tris-kuning telur tidak memperbaiki kualiti semen dibeku-cairkan lembu jantan.

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I certify that a Thesis Examination Committee has met on 3 August 2017 to conduct the final examination of Falah Hasan Ali Baiee on his thesis entitled "Effects of *Panax ginseng* and *Eurycoma longifolia* Jack Extracts on Chilled and Frozen-Thawed Crossbred Bull Semen Collected using Modified Electro-Ejaculation Method" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

| | |
|--------|--|
| AI | Artificial insemination |
| ALH | The amplitude of lateral head displacement |
| ATP | Adenosine triphosphate |
| AV | Artificial vagina |
| BCF | The beat cross frequency |
| BHT | Butylated hydroxytoluene |
| CASA | Computer-assisted sperm analysis |
| CAT | Catalase |
| CFDA | Carboxyfluorescein di-acetate |
| COMET | Single cell gel electrophoresis |
| CR | Rate of calving |
| DHA | Docosahexaenoic acid |
| DMSO | Dimethyl sulphoxide |
| dsDNA | Double-strand DNA |
| EE | Electro-ejaculator |
| EG | Ethylene glycol |
| EL | <i>Eurycoma longifolia</i> jack |
| FCM | Flow-cytometry |
| FTAI | Fixed-time artificial insemination |
| GR | Glutathione reductase |
| GSH-Px | Glutathione peroxidase |
| h | Hour |
| HDL | High density lipoproteins |

| | |
|-------|---|
| HOST | Hypo-osmotic swelling test |
| IAV | Internal artificial vagina |
| IgY | Immunoglobulin Y |
| IVF | <i>In vitro</i> fertilization |
| LDL | Low-density lipoproteins |
| LIN | The line of the spermatozoa motility |
| LPO | Lipid peroxidation |
| m | Meter |
| MDA | Malondialdehyde |
| min | Minute |
| PAF | Platelet-activating factors |
| PAFA | Platelet activating factor acetyl-hydrolase |
| PGe | <i>Panax ginseng</i> aqueous extract |
| PUFAs | Poly unsaturated fatty acids |
| ROS | Reactive oxygen species |
| SCSA | Sperm chromatin structure assay |
| s | Second |
| SEM | Standard error of the mean |
| SOD | Super oxide dismutase |
| ssDNA | Single-strand DNA |
| STR | The straightness of the sperm's movement |
| TA | Tongkat Ali |
| TAe | Tongkat Ali aqueous extract |
| TBARS | Thiobarbituric acid reactive substance |

| | |
|-----|--------------------------|
| VAP | Velocity average pathway |
| VCL | Velocity curvilinear |
| V | Volume |
| VSL | Velocity straight line |
| ZP | Zona pellucida |



CHAPTER 1

INTRODUCTION

1.1 General

Bull semen can be collected by different methods. An artificial vagina (AV) is the commonest method to collect semen samples from bulls because AV is very close to natural mating (Palmer, 2016). However, AV requires bulls to be trained to mount a cow or a dummy (Palmer, 2005). Training of bull requires a substantial amount of time and it is unsuitable for aggressive bulls. Sometimes, a bull abstains to mount a cow or a dummy because the operator holding the AV is standing close to the bull. With this reason, an alternative method of collection uses an electro-ejaculator (EE). Nowadays, EE is the most popular method to obtain semen samples from wild and domestic animals due to its ease of use and reliability in obtaining a semen sample and safety (Palmer, 2016).

The earliest account on artificial insemination (AI) was in 1322 when an Arab chieftain pinched some semen from a stallion owned by his foe and used it to impregnate his prized mare. Whether that story is a fact or fiction is immaterial, but it does emphasise the fact that AI had been around for a good many years (Foote, 2002). Nowadays, artificial breeding through AI in dairy and beef cattle is practiced in many countries around the world (Thibier and Wagner, 2002). Artificial insemination uses diluted and extended semen which can be preserved by either chilled (4-5 °C) or frozen (-196 °C). Chilled semen can be used to inseminate females for a few days only, while frozen semen can be used for a very long period when stored properly. Thus, 95% of semen that are used in an AI programme is frozen-thawed semen. In addition, 95% of mature cows are artificially inseminated using either chilled or frozen semen, due to many factors, such as the usage of semen obtained from high-quality sires, safety purposes, as some bulls are known to be aggressive require careful handling. Moreover, genetic material of top sires can be imported or exported as frozen semen. In addition, AI also can reduce the risk of spreading diseases (Salisbury *et al.*, 1978). Although, the use of frozen-thawed semen in AI is highly popular, however, the fertility rate of frozen-thawed semen is quite low than fresh semen. This is because of the lower post-thawed survival rate of spermatozoa (Yimer *et al.*, 2014). Approximately, 50% of spermatozoa are dead during freezing-thawing course (Watson, 2000). Thus, it is still very important to enhance the method of semen collection and semen handling, improve the component of extenders and the process of freezing-thawing and achieve better quality of spermatozoa after thawing. Temperature variation, ice formation and oxidative stress are the main challenges in preparation of frozen-thawed semen (Watson, 2000; Sariözkan *et al.*, 2009).

In addition, semen extenders should contain buffers, source of energy, antibiotics to prevent the bacterial growth, and cryoprotectant to protect against physical and chemical changes that occur during chilling, freezing and thawing processes (Salisbury *et al.*, 1978). Egg yolk is an ingredient in the extender to reduce cold shock effect (Moussa *et al.*, 2002), while glycerol reduces the harmful effect of ice formation (Taşdemir *et al.*, 2013) Antibiotics prevent bacterial infection (Gloria *et al.*, 2014), and liquid nitrogen allows the sperm to be preserved infinity (Foote, 2002).

1.2 Problem statement

Currently, as it was mentioned earlier, EE is the most popular method to collect semen samples from domestic animals (Palmer, 2016). Reliability in obtaining semen samples, requires minimal facilities and safety were the main benefits of this method (Palmer, 2016). However, EE is accompanied with discomfort and stress because this method works on electrical stimulation in a very sensitive area (ductus deferens, accessory genital glands and nerves around). Thus, this action may be associated with struggling, vocalizing and attempting to lie down of bulls during collection (Ohl, 1993).

In spite of the aim of sperm cryopreservation is to preserve sperm functional parameters and fertility; the cryopreservation course surely causes impairment to spermatozoa (Tvrdá *et al.*, 2016), so that reducing fertility. To date, various amount of studies have been done to elevate and enhance chilled and frozen-thawed semen quality using some additives to the extender of semen such as, unsaturated fatty acid (Kaka *et al.*, 2015; Khoshvagh *et al.*, 2015), anti-oxidant (Khumran *et al.*, 2015; Eidan, 2016), or amino acids (Holt *et al.*, 2015; Kumar *et al.*, 2015; Sariözkan *et al.*, 2015). Furthermore, some plants' extract have demonstrated as anti-oxidant additives to semen or semen extender (Sapanidou *et al.*, 2015; Tvrdá *et al.*, 2016), to study the toxic effect of extract on sperm (Kaefer *et al.*, 2013) and to dilute semen (Abul Rashid and Nurin Qistina, 2015). However, most of these additives cannot achieve satisfactory results, and also some of these additives make the extender costly. Therefore, there is no natural material, rich with bioactive components has been investigated to improve chilled and frozen-thawed semen quality.

1.3 Justification

As mentioned above, there are some limitations to use AV for semen collection, such as bulls must be trained to mount a cow or a dummy, training of bull requires time, and AV is unsuitable for aggressive bulls. However, EE can overcome these limitations and has been reported that EE is associated with pain (Ohl, 1993). Few studies were conducted to reduce and relieve pain such as using a narcoleptic tranquilizer (pipothiazine palmitate) for Bison bulls (Toosi *et al.*, 2013), hormones such as oxytocin to reduce time of ejaculation (Palmer *et al.*, 2004) and segmented the probe of EE (Etson *et al.*, 2004). However, the use of a tranquilizer may cause animals to lie down, and heavy weight sires may be injured during the process.

Hormones, e.g. oxytocin may increase the pain because it increases the contractions of smooth muscles. Segmented probe has no significant effect compared to normal probes (Eton *et al.*, 2004). Thus, there is a need to find out a modified method that can be used to collect semen samples from bulls with minimal discomfort.

Panax ginseng and Tongkat Ali (*Eurycoma longifolia*) are very well studied medicinal herbal plants. They have many beneficial effects. *Panax ginseng* can improve the awareness in rising energy levels and overall strength (Won *et al.*, 2014). Principally, *Panax ginseng* gives an amusing diversity of energetic elements, minerals and nutrients for health benefits to the body (Kim *et al.*, 2006). Conventionally, *Panax ginseng* is recognized to fight weaknesses, provide additional energy, increase psychological effectiveness and relieve exhaustion (Yun, 2001). Numerous reports pointed out that *Panax ginseng* has anti-oxidative effects (Lim *et al.*, 1998; Yokozawa *et al.*, 1998; Hu and Kitts, 2001; Yokozawa *et al.*, 2004; Jung *et al.*, 2005; Li *et al.*, 2008; Kim *et al.*, 2011; Wei *et al.*, 2012; Yun *et al.*, 2016).

Tongkat Ali is presumed to be a treatment for numerous problems such as ulcer, fatigue, poor endurance, to improve physical and mental performance, enhance the immune system and boost energy levels (Hool *et al.*, 1997; Kuo *et al.*, 2003; Abdullah *et al.*, 2004; Ismail *et al.*, 2012; Tambi *et al.*, 2012; George and Henkel, 2014; Hai *et al.*, 2016; Han *et al.*, 2016). Many studies have been conducted to examine the benefits of Tongkat Ali, such as to improve overall well-being (Ismail *et al.*, 2012), anti-oxidant (Panjaitan *et al.*, 2013; Varghese *et al.*, 2013; Lulu *et al.*, 2015), and anti-stress benefits (Talbot *et al.*, 2010, 2013). Husen *et al.* (2004) reported a substantial anti-hyper glycaemic effect of Tongkat Ali in a rat model.

AI is widely used to inseminate females with diluted semen. Semen that is used for AI can be stored either chilled or frozen. Chilled semen should be used within three days because after three days of the quality of semen is diminished leading to a reduction in fertility rate (Bucher *et al.*, 2009; Crespilho *et al.*, 2014). Frozen-thawed semen can be stored for a very long time in liquid nitrogen (Vishwanath and Shannon, 2000). However, about 50% of spermatozoa die during the freezing-thawing process (Shannon and Vishwanath, 1995; Curry, 2000; Lessard *et al.*, 2000; Vishwanath and Shannon, 2000; Watson, 2000). Moreover, the lifespan of frozen-thawed semen is very short as compared to fresh and chilled semen. In addition, the motility of frozen-thawed semen is not as good as fresh semen (Thomas *et al.*, 1998; Watson, 2000; Chaveiro *et al.*, 2006; Yimer *et al.*, 2014, 2015; Papa *et al.*, 2015); and thus, the fertility rate of frozen-thawed semen is lesser than fresh semen (Shannon and Vishwanath, 1995; Holt, 2000; Amirat-Briand *et al.*, 2010; Büyükleblebici *et al.*, 2014).

At present, there is no plant, rich with bioactive components that has been investigated to improve chilled and frozen-thawed semen quality. Thus, it is important to investigate the effects of these two plants extracts (PGe and TAE) which are rich with bioactive components on chilled and frozen-thawed semen.

1.4 Hypothesis:

Ho:

- i. The modified EE method will not reduce the discomfort signs during semen collection in bulls.
- ii. There is no beneficial effect on chilled and frozen-thawed bull sperm quality that was supplemented with *Panax ginseng*, Tongkat Ali (*Eurycoma longifolia*) and their combination.

Ha:

- i. The modified EE method reduces the discomfort signs during semen collection in bulls.
- ii. Bioactive components of *Panax ginseng* and Tongkat Ali (*Eurycoma longifolia*) extender improve quality of chilled and/or frozen-thawed semen. Aqueous extraction of these plants used as supplements to semen extenders separately or in combination could protect spermatozoa and might increase the viability, provide energy for sperm motility, protect the membranes of spermatozoa, improve DNA integrity, and reduce lipid peroxidation.

1.5 Objectives:

Thus, the present study was carried out with the following objectives:

- i. To minimize the discomfort signs during semen collection using electro-ejaculator.
- ii. To evaluate the effect of *Panax ginseng* aqueous extract on the quality of chilled and frozen-thawed bull semen.
- iii. To evaluate the effect of Tongkat Ali aqueous extract on the quality of chilled and frozen-thawed bull semen.
- iv. To assess the synergistic effect of Tongkat Ali and *Panax ginseng* extracts on the quality of frozen-thawed bull semen.

REFERENCES

- Abavisani, A., Arshami, J., Naserian, A. A., Kandelousi, M. A. S., and Azizzadeh, M. (2013). Quality of bovine chilled or frozen-thawed semen after addition of omega-3 fatty acids supplementation to extender. *International Journal of Fertility and Sterility*, 7(3), 161.
- Abayasekara, D., and Wathes, D. (1999). Effects of altering dietary fatty acid composition on prostaglandin synthesis and fertility. *Prostaglandins, Leukotrienes and Essential Fatty Acids (PLEFA)*, 61(5), 275-287.
- Abdullah, M., Rahman, A., Shakaff, A., and Noor, A. (2004). Discrimination and classification of *Eurycoma longifolia* Jack in medicinal foods by means of a DSP-based electronic taste sensor. *Transactions of the Institute of Measurement and Control*, 26(1), 19-39.
- Abul Rashid, B., and Nurin Qistina, M. N. (2015). *Preliminary Evaluation on the Use of Coconut Water as Semen Diluent for Local Kampung Cockerel*. Paper presented at the Proc. 2nd ARACP and 36th MASP Ann. Conf., 1-3 June 2015, Port Dickson, Negeri Sembilan, Malaysia.
- Acker, J. P., Lu, X. m., Young, V., Cheley, S., Bayley, H., Fowler, A., and Toner, M. (2003). Measurement of trehalose loading of mammalian cells porated with a metal- actuated switchable pore. *Biotechnology and Bioengineering*, 82(5), 525-532.
- Ahmad, M., Ahmad, N., Riaz, A., and Anzar, M. (2015). Sperm survival kinetics in different types of bull semen: progressive motility, plasma membrane integrity, acrosomal status and reactive oxygen species generation. *Reproduction, Fertility and Development*, 27(5), 784-793.
- Aitken, R., Bronson, R., Smith, T., and De Iuliis, G. (2013). The source and significance of DNA damage in human spermatozoa; a commentary on diagnostic strategies and straw man fallacies. *Molecular Human Reproduction*, gat025.
- Aitken, R. J. (1995). Free radicals, lipid peroxidation and sperm function. *Reproduction, Fertility and Development*, 7(4), 659-668.
- Aitken, R. J., and De Iuliis, G. N. (2007). Origins and consequences of DNA damage in male germ cells. *Reproductive Biomedicine Online*, 14(6), 727-733.
- Akhter, S., Ansari, M., Rakha, B., Ullah, N., Andrabi, S., and Khalid, M. (2011). *In Vitro* Evaluation of Liquid- stored Buffalo Semen at 5° C Diluted in Soya Lecithin Based Extender (Bioxcell®), Tris- Citric Egg Yolk, Skim Milk and Egg Yolk- Citrate Extenders. *Reproduction in Domestic Animals*, 46(1), 45-49.

- Al-Daraji, H. J. (2015). The Use of Pomegranate Juice for Counteract Lipid Peroxidation that Naturally Occurred during Liquid Storage of Roosters' Semen. *Pharmacognosy Communications*, 5(1), 70.
- Al-Kanaan, A., König, S., and Brügemann, K. (2015). Effects of heat stress on semen characteristics of Holstein bulls estimated on a continuous phenotypic and genetic scale. *Livestock Science*, 177, 15-24.
- Al-Makhzoomi, A., Lundeheim, N., Håård, M., and Rodriguez-Martinez, H. (2008). Sperm morphology and fertility of progeny-tested AI dairy bulls in Sweden. *Theriogenology*, 70(4), 682-691.
- Al-Salahi, O. S. A., Kit-Lam, C., Majid, A. M. S. A., Al-Suede, F. S. R., Saghir, S. A. M., Abdullah, W. Z., Ahamed, M. B., Yusoff, N. M. (2013). Anti-angiogenic quassinoid-rich fraction from *Eurycoma longifolia* modulates endothelial cell function. *Microvascular Research*, 90, 30-39.
- Allen, B. J., Rogers, S. D., Ghilardi, J. R., Menning, P. M., Kuskowski, M. A., Basbaum, A. I., Simone, D. A. Mantyh, P. W. (1997). Noxious Cutaneous Thermal Stimuli Induce a Graded Release of Endogenous Substance P in the Spinal Cord: Imaging Peptide *Action In Vivo*. *The Journal of Neuroscience*, 17(15), 5921-5927.
- Almqvist, J., and Wickersham, E. (1962). Diluents for Bovine Semen. XII. Fertility and Motility of Spermatozoa in Skimmilk with Various Levels of Glycerol and Methods of Glycerolization 1, 2. *Journal of Dairy Science*, 45(6), 782-787.
- Álvarez, M., Tamayo-Canul, J., Martínez-Rodríguez, C., López-Urueña, E., Gomes-Alves, S., Anel, L., Martínez-Pastor, F., and De Paz, P. (2012). Specificity of the extender used for freezing ram sperm depends of the spermatozoa source (ejaculate, electroejaculate or epididymis). *Animal Reproduction Science*, 132(3), 145-154.
- Amann, R., Seidel, G., and Mortimer, R. (2000). Fertilizing potential *in vitro* of semen from young beef bulls containing a high or low percentage of sperm with a proximal droplet. *Theriogenology*, 54(9), 1499-1515.
- Amann, R. P. (1962). Reproductive capacity of dairy bulls. III. The effect of ejaculation frequency, unilateral vasectomy, and age on spermatogenesis. *American Journal of Anatomy*, 110(1), 49-67.
- Amann, R. P., and Waberski, D. (2014). Computer-assisted sperm analysis (CASA): capabilities and potential developments. *Theriogenology*, 81(1), 5-17. e13.

- Amirat-Briand, L., Bencharif, D., Vera-Munoz, O., Pineau, S., Thorin, C., Destrumelle, S., Desherces, S., Anton, M., Jouan, M., Shmitt, E. Tainturier, D. (2010). *In vivo* fertility of bull semen following cryopreservation with an LDL (low density lipoprotein) extender: Preliminary results of artificial inseminations. *Animal Reproduction Science*, 122(3), 282-287.
- Amirat, L., Anton, M., Tainturier, D., Chatagnon, G., Battut, I., and Courtens, J. L. (2005). Modifications of bull spermatozoa induced by three extenders: Biociphos, low density lipoprotein and Triladyl, before, during and after freezing and thawing. *Reproduction*, 129(4), 535-543.
- Andersson, M., Taponen, J., Koskinen, E., and Dahlbom, M. (2004). Effect of insemination with doses of 2 or 15 million frozen-thawed spermatozoa and semen deposition site on pregnancy rate in dairy cows. *Theriogenology*, 61(7), 1583-1588.
- Andraszek, K., Banaszewska, D., Czubaszek, M., Wójcik, E., and Szostek, M. (2014). Comparison of different chromatin staining techniques for bull sperm. *Archives Animal Breeding*, 57, 1-15.
- Ang, H., and Lee, K. (2002). Effect of *Eurycoma longifolia* Jack on orientation activities in middle-aged male rats. *Fundamental and Clinical Pharmacology*, 16(6), 479-483.
- Ang, H., Ngai, T., and Tan, T. (2003). Effects of *Eurycoma longifolia* Jack on sexual qualities in middle aged male rats. *Phytomedicine*, 10(6), 590-593.
- Ang, H. H., and Cheang, H. S. (1999). Studies on the Anxiolytic Activity of *Eurycoma longifolia* Jack Roots in Mice. *The Japanese Journal of Pharmacology*, 79(4), 497-500.
- Ang, H. H., Cheang, H. S., and Yusof, A. P. M. (2000). Effects of *Eurycoma longifolia* Jack (Tongkat Ali) on the initiation of sexual performance of inexperienced castrated male rats. *Experimental Animals*, 49(1), 35-38.
- Ang, H. H., and Sim, M. K. (1997). Effects of *Eurycoma longifolia* Jack on sexual behaviour of male rats. *Archives of Pharmacal Research*, 20(6), 656-658.
- Ang, H. H., and Sim, M. K. (1998). *Eurycoma longifolia* JACK and orientation activities in sexually experienced male rats. *Biological and Pharmaceutical Bulletin*, 21(2), 153-155.
- Ansari, M. S., Rakha, B. A., and Akhter, S. (2011). Effect of butylated hydroxytoluene supplementation in extender on motility, plasmalemma and viability of Sahiwal bull spermatozoa. *Pakistan Journal of Zoology*, 43(2), 311-314.

- Ansari, M. S., Rakha, B. A., Andrabi, S. M., and Akhter, S. (2010). Usefulness of powdered and fresh egg yolk for cryopreservation of Zebu bull spermatozoa. *Reproductive Biology*, 10(3), 235-240.
- Anton, M. (2007). Composition and structure of hen egg yolk *Bioactive egg compounds* (pp. 1-6): Springer.
- Anton, M., Martinet, V., Dalgalarondo, M., Beaumal, V., David-Briand, E., and Rabesona, H. (2003). Chemical and structural characterisation of low-density lipoproteins purified from hen egg yolk. *Food Chemistry*, 83(2), 175-183.
- Antończyk, A., Nizański, W., Twardoń, J., Kozdrowski, R., Ochota, M., Błasiak, K., . . . Stańczyk, E. (2010). Current views on computer assisted sperm analysis. *Medycyna Weterynaryjna*, 66(10), 663-667.
- Anzar, M., He, L., Buhr, M. M., Kroetsch, T. G., and Pauls, K. P. (2002). Sperm apoptosis in fresh and cryopreserved bull semen detected by flow cytometry and its relationship with fertility. *Biology of Reproduction*, 66(2), 354-360.
- Anzar, M., Kroetsch, T., and Boswall, L. (2011). Cryopreservation of bull semen shipped overnight and its effect on post-thaw sperm motility, plasma membrane integrity, mitochondrial membrane potential and normal acrosomes. *Animal Reproduction Science*, 126(1), 23-31.
- Arakawa, T., Carpenter, J. F., Kita, Y. A., and Crowe, J. H. (1990). The basis for toxicity of certain cryoprotectants: a hypothesis. *Cryobiology*, 27(4), 401-415.
- Asadpour, R., Jafari, R., and Tayefi-Nasrabadi, H. (2011). Effect of various levels of catalase antioxidant in semen extenders on lipid peroxidation and semen quality after the freeze-thawing bull semen. *Veterinary Research Forum*, 2(4), 218-221.
- Asadpour, R., Jafari, R., Tayefi-Nasrabadi, H., (2012). The effect of antioxidant supplementation in semen extenders on semen quality and lipid peroxidation of chilled bull spermatozoa. *Iranian Journal of Veerinary Research* 13, 246-249.
- Asiah, O., Nurhanan, M., and Ilham, A. M. (2007). Determination of bioactive peptide (4.3 kDa) as an aphrodisiac marker in six Malaysian plants. *Journal of Tropical Forest Science*, 19(1), 61-63.
- Attele, A. S., Wu, J. A., and Yuan, C.-S. (1999). Ginseng pharmacology: multiple constituents and multiple actions. *Biochemical Pharmacology*, 58(11), 1685-1693.
- Attia, S., Katila, T., and Andersson, M. (2016). The Effect of Sperm Morphology and Sire Fertility on Calving Rate of Finnish Ayrshire AI Bulls. *Reproduction in Domestic Animals*, 51(1), 54-58.

- Auger, J. (2010). Assessing human sperm morphology: top models, underdogs or biometrics. *Asian Journal Andrology*, 12(1), 36-46.
- Bag, S., Joshi, A., Naqvi, S., and Mittal, J. (2004). Effect of post-thaw incubation on sperm kinematics and acrosomal integrity of ram spermatozoa cryopreserved in medium-sized French straws. *Theriogenology*, 62(3), 415-424.
- Baghshahi, H., Riasi, A., Mahdavi, A., and Shirazi, A. (2014). Antioxidant effects of clove bud (*Syzygium aromaticum*) extract used with different extenders on ram spermatozoa during cryopreservation. *Cryobiology*, 69(3), 482-487.
- Bailey, J. L., Bilodeau, J., and Cormier, N. (2000). Semen cryopreservation in domestic animals: a damaging and capacitating phenomenon. *Journal of Andrology*, 21(1), 1-7.
- Ballester, J., Johannisson, A., Saravia, F., Håård, M., Gustafsson, H., Bajramovic, D., and Rodriguez-Martinez, H. (2007). Post-thaw viability of bull AI-doses with low-sperm numbers. *Theriogenology*, 68(6), 934-943.
- Banaszewska, D., Andraszek, K., Czubaszek, M., and Biesiada-Drzazga, B. (2015). The effect of selected staining techniques on bull sperm morphometry. *Animal Reproduction Science*, 159, 17-24.
- Bansal, A. K., and Bilaspuri, G. (2010). Impacts of oxidative stress and antioxidants on semen functions. *Veterinary Medicine International*, 2011.
- Barak, Y., Amit, A., Lessing, J. B., Paz, G., Homonnai, Z. T., and Yogev, L. (1992). Improved fertilization rate in an *in vitro* fertilization program by egg yolk-treated sperm. *Fertility and Sterility*, 58(1), 197-198.
- Barakat, I. A., Danfour, M. A., Galewan, F. A., and Dkhil, M. A. (2015). Effect of various concentrations of caffeine, pentoxifylline, and kallikrein on hyperactivation of frozen bovine semen. *BioMed Research International*, 2015.
- Barszcz, K., Wiesetek, D., Wasowicz, M., and Kupczynska, M. (2012). Bull Semen Collection and Analysis for Artificial Insemination. *Journal of Agricultural Science*, 4(3), 1.
- Barth, A. D., Arteaga, A. A., Brito, L. F., and Palmer, C. W. (2004). Use of internal artificial vaginas for breeding soundness evaluation in range bulls: an alternative for electroejaculation allowing observation of sex drive and mating ability. *Animal Reproduction Science*, 84(3), 315-325.
- Barth, A. D., Bowman, P. A., Bo, G. A., and Mapletoft, R. J. (1992). Effect of narrow sperm head shape on fertility in cattle. *The Canadian Veterinary Journal*, 33(1), 31.

- Batellier, F., Vidament, M., Fauquant, J., Duchamp, G., Arnaud, G., Yvon, J., and Magistrini, M. (2001). Advances in cooled semen technology. *Animal Reproduction Science*, 68(3), 181-190.
- Batista, M., Niño, T., Santana, M., Alamo, D., Castro, N., Reyes, R., González, F., Cabrera, F., and Gracia, A. (2011). Influence of the preservation temperature (37, 20, 4, – 196 C) and the mixing of semen over sperm quality of Majorera bucks. *Reproduction in Domestic Animals*, 46(2), 281-288.
- Bedford, J., and Nicander, L. (1971). Ultrastructural changes in the acrosome and sperm membranes during maturation of spermatozoa in the testis and epididymis of the rabbit and monkey. *Journal of Anatomy*, 108(Pt 3), 527.
- Belleannée, C., Labas, V., Teixeira-Gomes, A.-P., Gatti, J. L., Dacheux, J.-L., and Dacheux, F. (2011). Identification of luminal and secreted proteins in bull epididymis. *Journal of Proteomics*, 74(1), 59-78.
- Beran, J., Šimoník, O., Rajmon, R., Stádník, L., Doležalová, M., Krejčířková, A., Ducháček, J., and Šichtař, J. (2016). Effect of LDL Addition Into Selected Bull Sperm Diluters on Resistance of Spermatozoa Against Cold Shock. *Acta Universitatis Agriculturae et Silviculturae Mendelianae Brunensis*, 64(2), 395-399.
- Bergeron, A., and Manjunath, P. (2006). New insights towards understanding the mechanisms of sperm protection by egg yolk and milk. *Molecular Reproduction And Development*, 73(10), 1338-1344.
- Betancourt, M., and Reséndiz, A. (2006). Effect of two insecticides and two herbicides on the porcine sperm motility patterns using computer-assisted semen analysis (CASA) *in vitro*. *Reproductive Toxicology*, 22(3), 508-512.
- Bhat, R., and Karim, A. (2010). Tongkat Ali (*Eurycoma longifolia* Jack): a review on its ethnobotany and pharmacological importance. *Fitoterapia*, 81(7), 669-679.
- Blackshaw, A., and Salisbury, G. (1957). Factors influencing metabolic activity of bull spermatozoa. II. Cold-shock and its prevention. *Journal of Dairy Science*, 40(9), 1099-1106.
- Blässe, A.-K., Oldenhof, H., Ekhlasi-Hundrieser, M., Wolkers, W., Sieme, H., and Bollwein, H. (2012). Osmotic tolerance and intracellular ion concentrations of bovine sperm are affected by cryopreservation. *Theriogenology*, 78(6), 1312-1320.
- Bó, G., Baruselli, P., and Martinez, M. (2003). Pattern and manipulation of follicular development in *Bos indicus* cattle. *Animal Reproduction Science*, 78(3), 307-326.

- Bochenek, M., Smorag, Z., and Pilch, J. (2001). Sperm chromatin structure assay of bulls qualified for artificial insemination. *Theriogenology*, 56(4), 557-567.
- Boe-Hansen, G., Rego, J., Crisp, J., Moura, A., Nouwens, A., Li, Y., Venus, B., Burns, B.M., McGowan, M. (2015). Semen plasma proteins and their relationship with percentage of morphologically normal sperm in 2-year-old Brahman (*Bos indicus*) bulls. *Animal Reproduction Science*, 162, 20-30.
- Boersma, A., Braun, J., and Stolla, R. (1999). Influence of Random Factors and Two Different Staining Procedures on Computer- assisted Sperm Head Morphometry in Bulls. *Reproduction in Domestic Animals*, 34(2), 77-82.
- Bollwein, H., Fuchs, I., and Koess, C. (2008). Interrelationship between plasma membrane integrity, mitochondrial membrane potential and DNA fragmentation in cryopreserved bovine spermatozoa. *Reproduction in Domestic Animals*, 43(2), 189-195.
- Borges-Silva, J. C., Silva, M. R., Marinho, D. B., Nogueira, E., Sampaio, D. C., Oliveira, L. O. F., Abreu, U. G. P., Mourão, G. B. and Sartori, R. (2016). Cooled semen for fixed-time artificial insemination in beef cattle. *Reproduction, Fertility and Development*, 28(7), 1004-1008.
- Brito, L., Silva, A., Rodrigues, L., Vieira, F., Deragon, L., and Kastelic, J. (2002). Effect of age and genetic group on characteristics of the scrotum, testes and testicular vascular cones, and on sperm production and semen quality in AI bulls in Brazil. *Theriogenology*, 58(6), 1175-1186.
- Briton-Jones, C., Yeung, Q. S. Y., Tjer, G. C. C., Chiu, T. T. Y., Cheung, L. P., Yim, S. F., Lok, I. H., and Haines, C. (2001). The Effects of Follicular Fluid and Platelet-Activating Factor on Motion Characteristics of Poor-Quality Cryopreserved Human Sperm. *Journal of Assisted Reproduction and Genetics*, 18(3), 165-170.
- Bucher, A., Kasimanickam, R., Hall, J., Dejarnette, J., Whittier, W., Kähn, W., and Xu, Z. (2009). Fixed-time AI pregnancy rate following insemination with frozen-thawed or fresh-extended semen in progesterone supplemented CO-Synch protocol in beef cows. *Theriogenology*, 71(7), 1180-1185.
- Buranaamnuay, K., Seesuan, K., and Saikhun, K. (2016). Preliminary study on effects of bovine frozen semen storage using a liquid nitrogen-independent method on the quality of post-thaw spermatozoa. *Animal Reproduction Science*, 172, 32-38.
- Büyükblebici, S., Tuncer, P. B., Bucak, M. N., Eken, A., Sarıözkan, S., Taşdemir, U., and Endirlik, B. Ü. (2014). Cryopreservation of bull sperm: Effects of extender supplemented with different cryoprotectants and antioxidants on sperm motility, antioxidant capacity and fertility results. *Animal Reproduction Science*, 150(3), 77-83.

- Caballero, I., Parrilla, I., Almiñana, C., del Olmo, D., Roca, J., Martínez, E., and Vázquez, J. (2012). Seminal plasma proteins as modulators of the sperm function and their application in sperm biotechnologies. *Reproduction in Domestic Animals*, 47(s3), 12-21.
- Calvin, H., and Bedford, J. (1971). Formation of disulphide bonds in the nucleus and accessory structures of mammalian spermatozoa during maturation in the epididymis. *Journal of Reproduction and Fertility. Supplement*, 13, Suppl 13: 65.
- Celeghini, E. C. C., de Arruda, R. P., de Andrade, A. F. C., Nascimento, J., Raphael, C. F., and Rodrigues, P. H. M. (2008). Effects that bovine sperm cryopreservation using two different extenders has on sperm membranes and chromatin. *Animal Reproduction Science*, 104(2), 119-131.
- Chan, K.-L., Choo, C.-Y., Abdullah, N. R., and Ismail, Z. (2004). Antiplasmodial studies of *Eurycoma longifolia* Jack using the lactate dehydrogenase assay of *Plasmodium falciparum*. *Journal of Ethnopharmacology*, 92(2), 223-227.
- Chaveiro, A., Machado, L., Frijters, A., Engel, B., and Woelders, H. (2006). Improvement of parameters of freezing medium and freezing protocol for bull sperm using two osmotic supports. *Theriogenology*, 65(9), 1875-1890.
- Chelucci, S., Pasciu, V., Succu, S., Addis, D., Leoni, G. G., Manca, M. E., Naitana, S., and Berlinguer, F. (2015). Soybean lecithin-based extender preserves spermatozoa membrane integrity and fertilizing potential during goat semen cryopreservation. *Theriogenology*, 83(6), 1064-1074.
- Chen, J., Xu, M., Chen, L., Chen, Y., and Chiu, T. (1998). Effect of *Panax notoginseng* saponins on sperm motility and progression *in vitro*. *Phytomedicine*, 5(4), 289-292.
- Chen, Y., Phang, W.-M., Mu, A. K.-W., Chan, C.-K., Low, B.-S., Sasidharan, S., and Chan, K.-L. (2015). Decreased expression of alpha-2-HS glycoprotein in the sera of rats treated with *Eurycoma longifolia* extract. *Frontiers in Pharmacology*, 6.
- Cheng, Y., and Zhang, J. t. (2005). Anti- amnestic and anti- aging effects of ginsenoside Rg1 and Rb1 and its mechanism of action. *Acta Pharmacologica Sinica*, 26(2), 143-149.
- Choi, H., Seong, D., and Rha, K. (1995). Clinical efficacy of Korean red ginseng for erectile dysfunction. *International Journal of Impotence Research*, 7(3), 181-186.
- Choi, K.-t. (2008). Botanical characteristics, pharmacological effects and medicinal components of Korean *Panax ginseng* CA Meyer. *Acta Pharmacologica Sinica*, 29(9), 1109.

- Ciftci, H., and Zulkadir, U. (2010). The correlation between bull sperm head dimensions and mitochondrial helix length. *Journal of Animal and Veterinary Advances*, 7, 1169-1172.
- Contri, A., Gloria, A., Robbe, D., Valorz, C., Wegher, L., and Carluccio, A. (2013). Kinematic study on the effect of pH on bull sperm function. *Animal reproduction science*, 136(4), 252-259.
- Contri, A., Valorz, C., Faustini, M., Wegher, L., and Carluccio, A. (2010). Effect of semen preparation on casa motility results in cryopreserved bull spermatozoa. *Theriogenology*, 74(3), 424-435.
- Coulter, G. H., and Kozub, G. (1989). Efficacy of methods used to test fertility of beef bulls used for multiple-sire breeding under range conditions. *Journal of Animal Science*, 67(7), 1757-1766.
- Crespilho, A., Nichi, M., Guasti, P., Freitas-Dell'Aqua, C., Sá Filho, M., Maziero, R., Dell'Aqua Jr, J. A., Papa, F. O. (2014). Sperm fertility and viability following 48h of refrigeration: evaluation of different extenders for the preservation of bull semen in liquid state. *Animal Reproduction Science*, 146(3), 126-133.
- Crespilho, A., Sá Filho, M., Dell'Aqua, J., Nichi, M., Monteiro, G., Avanzi, B., Martins, A. and Papa, F. O. (2012). Comparison of *in vitro* and *in vivo* fertilizing potential of bovine semen frozen in egg yolk or new lecithin based extenders. *Livestock Science*, 149(1), 1-6.
- Crespilho, A. M., Papa, F. O., de Paula Santos, M., and de Sa Filho, M. F. (2012). Use of cooled bull semen as a strategy to increase the pregnancy rate in fixed-time artificial insemination programs-case report. *American Journal of Animal and Veterinary Sciences*, 7(4), 175.
- Curry, M. R. (2000). Cryopreservation of semen from domestic livestock. *Reviews of reproduction*, 5(1), 46-52.
- Cyranoski, D. (2005). Malaysian researchers bet big on home-grown Viagra. *Nature Medicine*, 11(9), 912-912.
- D'Occhio, M., Hengstberger, K., and Johnston, S. (2007). Biology of sperm chromatin structure and relationship to male fertility and embryonic survival. *Animal Reproduction Science*, 101(1), 1-17.
- Darszon, A., Labarca, P., Nishigaki, T., and Espinosa, F. (1999). Ion channels in sperm physiology. *Physiological Reviews*, 79(2), 481-510.
- Davis, I., Bratton, R., and Foote, R. (1963). Livability of bovine spermatozoa at 5C in tris-buffered and citrate-buffered yolk-glycerol extenders. *Journal of Dairy Science*, 46(1), 57-60.

- De Andrade, A., De Arruda, R., Celeghini, E., Nascimento, J., Martins, S., Raphael, C., and Moretti, A. (2007). Fluorescent stain method for the simultaneous determination of mitochondrial potential and integrity of plasma and acrosomal membranes in boar sperm. *Reproduction in Domestic Animals*, 42(2), 190-194.
- De Leeuw, F., De Leeuw, A., Den Daas, J., Colenbrander, B., and Verkleij, A. (1993). Effects of various cryoprotective agents and membrane-stabilizing compounds on bull sperm membrane integrity after cooling and freezing. *Cryobiology*, 30(1), 32-44.
- Den Daas, J., De Jong, G., Lansbergen, L., and Van Wagtenonk-De Leeuw, A. (1998). The relationship between the number of spermatozoa inseminated and the reproductive efficiency of individual dairy bulls. *Journal of Dairy Science*, 81(6), 1714-1723.
- DeVane, C. L. (2001). Substance P: a new era, a new role. *Pharmacotherapy: The Journal of Human Pharmacology and Drug Therapy*, 21(9), 1061-1069.
- Dietrich, G., Szyrka, A., Wojtczak, M., Dobosz, S., Goryczko, K., and Ciereszko, A. (2005). Effects of UV irradiation and hydrogen peroxide on DNA fragmentation, motility and fertilizing ability of rainbow trout (*Oncorhynchus mykiss*) spermatozoa. *Theriogenology*, 64(8), 1809-1822.
- Dobrinski, I., Hughes, H., and Barth, A. (1994). Flow cytometric and microscopic evaluation and effect on fertility of abnormal chromatin condensation in bovine sperm nuclei. *Journal of Reproduction And Fertility*, 101(3), 531-538.
- Drobnis, E. Z., Crowe, L. M., Berger, T., Anchordoguy, T. J., Overstreet, J. W., and Crowe, J. H. (1993). Cold shock damage is due to lipid phase transitions in cell membranes: a demonstration using sperm as a model. *Journal of Experimental Zoology*, 265(4), 432-437.
- Du, G.-J., Dai, Q., Williams, S., Wang, C.-Z., and Yuan, C.-S. (2011). Synthesis of protopanaxadiol derivatives and evaluation of their anticancer activities. *Anti-Cancer Drugs*, 22(1), 35.
- Ebrahimi, F., Ibrahim, B., Teh, C.-H., Murugaiyah, V., and Chan, K.-L. (2016). Urinary NMR-based metabolomic analysis of rats possessing variable sperm count following orally administered *Eurycoma longifolia* extracts of different quassinoid levels. *Journal of Ethnopharmacology*, 182, 80-89.
- Eidan, S. M. (2016). Effect on post-cryopreserved semen characteristics of Holstein bulls of adding combinations of vitamin C and either catalase or reduced glutathione to Tris extender. *Animal Reproduction Science*, 167, 1-7.

- Ejaz, R., Ansari, M., Rakha, B., Husna, A., Qadeer, S., Iqbal, R., Ullah, N., and Akhter, S. (2016). Cholesterol Supplementation in Extender Improves Quality of Frozen-Thawed Nili-Ravi Buffalo Bull Semen. *Buffalo Bulletin (June 2016)*, 35(2).
- Enciso, M., Cisale, H., Johnston, S., Sarasa, J., Fernández, J., and Gosálvez, J. (2011). Major morphological sperm abnormalities in the bull are related to sperm DNA damage. *Theriogenology*, 76(1), 23-32.
- Erasmus, N., Solomon, M., Fortuin, K., and Henkel, R. (2012). Effect of *Eurycoma longifolia* Jack (Tongkat ali) extract on human spermatozoa *in vitro*. *Andrologia*, 44(5), 308-314.
- Esterbauer, H., and Cheeseman, K. H. (1990). [42] Determination of aldehydic lipid peroxidation products: Malonaldehyde and 4-hydroxynonenal. *Methods in Enzymology*, 186, 407-421.
- Etson, C. J., Waldner, C. L., and Barth, A. D. (2004). Evaluation of a segmented rectal probe and caudal epidural anesthesia for electroejaculation of bulls. *The Canadian Veterinary Journal*, 45(3), 235.
- Evenson, D., Jost, L., Marshall, D., Zinaman, M., Clegg, E., Purvis, K., De Angelis, P., and Claussen, O. (1999). Utility of the sperm chromatin structure assay as a diagnostic and prognostic tool in the human fertility clinic. *Human Reproduction*, 14(4), 1039-1049.
- Evenson, D. P., Darzynkiewicz, Z., and Melamed, M. R. (1980). Comparison of human and mouse sperm chromatin structure by flow cytometry. *Chromosoma*, 78(2), 225-238.
- Fahy, G. M. (1986). The relevance of cryoprotectant "toxicity" to cryobiology. *Cryobiology*, 23(1), 1-13.
- Fahy, G. M., Lilley, T. H., Linsdell, H., Douglas, M. S. J., and Meryman, H. T. (1990). Cryoprotectant toxicity and cryoprotectant toxicity reduction: in search of molecular mechanisms. *Cryobiology*, 27(3), 247-268.
- Farouk, A.-E., and Benafri, A. (2007). Antibacterial activity of *Eurycoma longifolia* Jack. A Malaysian medicinal plant. *Saudi Medical Journal*, 28(9), 1422-1424.
- Farrell, P., Presicce, G., Brockett, C., and Foote, R. (1998). Quantification of bull sperm characteristics measured by computer-assisted sperm analysis (CASA) and the relationship to fertility. *Theriogenology*, 49(4), 871-879.
- Fatehi, A., Bevers, M., Schoevers, E., Roelen, B., Colenbrander, B., and Gadella, B. (2006). DNA damage in bovine sperm does not block fertilization and early embryonic development but induces apoptosis after the first cleavages. *Journal of Andrology*, 27(2), 176-188.

- Fawcett, D. W. (1970). A comparative view of sperm ultrastructure. *Biology of Reproduction*, 2(Supplement 2), 90-127.
- Fawcett, D. W. (1975). The mammalian spermatozoon. *Developmental Biology*, 44(2), 394-436.
- Felipe-Pérez, Y. E., de Lourdes Juárez-Mosqueda, M., Hernández-González, E. O., & de Jesús Valencia, J. (2008). Viability of fresh and frozen bull sperm compared by two staining techniques. *Acta Veterinaria Brasilica*, 2(4), 123-130.
- Fernández-Santos, M., Martínez-Pastor, F., Matias, D., Domínguez-Rebolledo, A., Estesó, M., Montoro, V., and Garde, J. (2009). Effects of long-term chilled storage of red deer epididymides on DNA integrity and motility of thawed spermatozoa. *Animal Reproduction Science*, 111(1), 93-104.
- Ferraz, M., Morató, R., Yeste, M., Arcarons, N., Pena, A., Tamargo, C., Hidalgo, C. O., Muiño, R. and Mogas, T. (2014). Evaluation of sperm subpopulation structure in relation to *in vitro* sperm-oocyte interaction of frozen-thawed semen from Holstein bulls. *Theriogenology*, 81(8), 1067-1072.
- Filho, I. B., Pederzoli, C., Sgaravatti, A., Gregory, R., Filho, C. D., Jobim, M., and Mattos, R. (2009). Skim milk-egg yolk based semen extender compensates for non-enzymatic antioxidant activity loss during equine semen cryopreservation. *Animal Reproduction*, 6(2), 392-399.
- Flesch, F. M., and Gadella, B. M. (2000). Dynamics of the mammalian sperm plasma membrane in the process of fertilization. *Biochimica et Biophysica Acta (BBA)-Reviews on Biomembranes*, 1469(3), 197-235.
- Foote, R. (1970). Fertility of bull semen at high extension rates in Tris-buffered extenders. *Journal of Dairy Science*, 53(10), 1475-1477.
- Foote, R. (2002). The history of artificial insemination: Selected notes and notables. *Journal of Animal Science*, 80(2), 1-10.
- Foote, R., and Kaproth, M. (1997). Sperm numbers inseminated in dairy cattle and nonreturn rates revisited. *Journal of Dairy Science*, 80(11), 3072-3076.
- Foote, R., and Kaproth, M. (2002). Large batch freezing of bull semen: effect of time of freezing and fructose on fertility. *Journal of Dairy Science*, 85(2), 453-456.
- Foote, R. H., Brockett, C. C., and Kaproth, M. T. (2002). Motility and fertility of bull sperm in whole milk extender containing antioxidants. *Animal Reproduction Science*, 71(1), 13-23.

- Fukuda, M., Sakase, M., Fukushima, M., and Harayama, H. (2016). Changes of IZUMO1 in bull spermatozoa during the maturation, acrosome reaction, and cryopreservation. *Theriogenology*, 86(9), 2179-2188. e2173.
- Furman, J., Ball, L., and Seidel, G. (1975). Electroejaculation of bulls using pulse waves of variable frequency and length. *Journal of Animal Science*, 40(4), 665-670.
- Gadea, J., Gumbao, D., Canovas, S., García- Vázquez, F. A., Grullón, L. A., and Gardón, J. C. (2008). Supplementation of the dilution medium after thawing with reduced glutathione improves function and the *in vitro* fertilizing ability of frozen- thawed bull spermatozoa. *International Journal of Andrology*, 31(1), 40-49.
- Gamal, A., El-Nattat, W. S., El-Sheshtawy, R. I., and El-Maaty, A. M. A. (2016). Substitution of egg yolk with different concentrations of soybean lecithin in tris-based extender during bulls' semen preservability. *Asian Pacific Journal of Reproduction*, 5(6), 514-518.
- Gandini, L., Lombardo, F., Paoli, D., Caponecchia, L., Familiari, G., Verlengia, C., . . . Lenzi, A. (2000). Study of apoptotic DNA fragmentation in human spermatozoa. *Human Reproduction*, 15(4), 830-839.
- García- Macías, V., De Paz, P., Martínez- Pastor, F., Álvarez, M., Gomes- Alves, S., Bernardo, J., Anel, E. and Anel, L. (2007). DNA fragmentation assessment by flow cytometry and Sperm-Bos-Halomax (bright- field microscopy and fluorescence microscopy) in bull sperm. *International Journal of Andrology*, 30(2), 88-98.
- Garner, D., Thomas, C., Gravance, C., Marshall, C., DeJarnette, J., and Allen, C. (2001). Seminal plasma addition attenuates the dilution effect in bovine sperm. *Theriogenology*, 56(1), 31-40.
- Garrido, N., Meseguer, M., Simon, C., Pellicer, A., and Remohi, J. (2004). Pro-oxidative and anti-oxidative imbalance in human semen and its relation with male fertility. *Asian Journal of Andrology*, 6(1), 59-66.
- George, A., and Henkel, R. (2014). Phytoandrogenic properties of *Eurycoma longifolia* as natural alternative to testosterone replacement therapy. *Andrologia*, 46(7), 708-721.
- Gholami, H., Chamani, M., Towhidi, A., and Fazeli, M. (2010). Effect of feeding a docosahexaenoic acid-enriched nutraceutical on the quality of fresh and frozen-thawed semen in Holstein bulls. *Theriogenology*, 74(9), 1548-1558.
- Gillis, C. N. (1997). *Panax ginseng* pharmacology: a nitric oxide link? *Biochemical Pharmacology*, 54(1), 1-8.

- Gloria, A., Contri, A., Wegher, L., Vignola, G., Dellamaria, D., and Carluccio, A. (2014). The effects of antibiotic additions to extenders on fresh and frozen-thawed bull semen. *Animal Reproduction Science*, 150(1), 15-23.
- Gomendio, M., and Roldan, E. R. (2004). Implications of diversity in sperm size and function for sperm competition and fertility. *International Journal of Developmental Biology*, 52(5-6), 439-447.
- Goovaerts, I., Hoflack, G., Van Soom, A., Dewulf, J., Nichi, M., de Kruif, A., and Bols, P. (2006). Evaluation of epididymal semen quality using the Hamilton-Thorne analyser indicates variation between the two caudae epididymides of the same bull. *Theriogenology*, 66(2), 323-330.
- Gosz, E., Mirny, Z., Horbowy, J., and Ziętara, M. (2010). Morphometry of turbot spermatozoa in relation to the location and time of capture during the spawning season. *Journal of Applied Ichthyology*, 26(5), 784-788.
- Grassi, G., Cappello, N., Gheorghe, M., Salton, L., Di Bisceglie, C., Manieri, C., and Benedetto, C. (2010). Exogenous platelet-activating factor improves the motility of human spermatozoa evaluated with CASA: optimal concentration and incubation time. *Journal of Endocrinological Investigation*, 33(10), 684-690.
- Gray, S. L., Lackey, B. R., and Boone, W. R. (2015). Effects of *Panax ginseng*, zearalenol, and estradiol on sperm function. *Journal of Ginseng Research*.
- Griveau, J., and Lannou, D. (1997). Reactive oxygen species and human spermatozoa: physiology and pathology. *International Journal of Andrology*, 20(2), 61-69.
- Gualtieri, R., Barbato, V., Fiorentino, I., Braun, S., Rizos, D., Longobardi, S., and Talevi, R. (2014). Treatment with zinc, d-aspartate, and coenzyme Q10 protects bull sperm against damage and improves their ability to support embryo development. *Theriogenology*, 82(4), 592-598.
- Gürler, H., Calisici, O., and Bollwein, H. (2015). Inter- and intra-individual variability of total antioxidant capacity of bovine seminal plasma and relationships with sperm quality before and after cryopreservation. *Animal Reproduction Science*, 155, 99-105.
- Guthrie, H., Liu, J., and Critser, J. (2002). Osmotic tolerance limits and effects of cryoprotectants on motility of bovine spermatozoa. *Biology of Reproduction*, 67(6), 1811-1816.
- Guthrie, H., and Welch, G. (2006). Determination of intracellular reactive oxygen species and high mitochondrial membrane potential in Percoll-treated viable boar sperm using fluorescence-activated flow cytometry. *Journal of Animal Science*, 84(8), 2089-2100.

- Hai Dang, N., Choo, Y. Y., Tien Dat, N., Hoai Nam, N., Van Minh, C., and Lee, J. H. (2016). 7-Methoxy-(9H- β -Carboline-1-yl)-(E)-1-Propenoic Acid, a β -Carboline Alkaloid From *Eurycoma longifolia*, Exhibits Anti-Inflammatory Effects by Activating the Nrf2/Heme Oxygenase-1 Pathway. *Journal of Cellular Biochemistry*, 117(3), 659-670.
- Hall, S. H., Hamil, K. G., and French, F. S. (2002). Host defense proteins of the male reproductive tract. *Journal of Andrology*, 23(5), 585-597.
- Hammerstedt, R., Graham, J. K., and Nolan, J. P. (1990). Cryopreservation of mammalian sperm: what we ask them to survive. *J Androl*, 11(1), 73-88.
- Hammerstedt, R., and Parks, J. (1987). Changes in sperm surfaces associated with epididymal transit. *Journal of Reproduction and Fertility*, 34: 133-149.
- Han, Y. M., Woo, S.-U., Choi, M. S., Park, Y. N., Kim, S. H., Yim, H., and Yoo, H. H. (2016). Antiinflammatory and analgesic effects of *Eurycoma longifolia* extracts. *Archives of Pharmacal Research*, 39(3), 421-428.
- He, L., Bailey, J., and Buhr, M. (2001). Incorporating lipids into boar sperm decreases chilling sensitivity but not capacitation potential. *Biology of Reproduction*, 64(1), 69-79.
- Henkel, R. R., Wang, R., Bassett, S. H., Chen, T., Liu, N., Zhu, Y., and Tambi, M. I. (2014). Tongkat Ali as a potential herbal supplement for physically active male and female seniors—a pilot study. *Phytotherapy Research*, 28(4), 544-550.
- Hib, J. (1976). *The 'in vivo' effects of oxytocin and vasopressin on spontaneous contractility of the rat epididymis.* *International Journal of Fertility*, 22(1), 63-64.
- Hirai, M., Cerbito, W., Wijayagunawardane, M., Braun, J., Leidl, W., Ohosaki, K., Matsuzawa, T., Miyazawa, K., and Sato, K. (1997). The effect of viscosity of semen diluents on motility of bull spermatozoa. *Theriogenology*, 47(7), 1463-1478.
- Holroyd, R., Doogan, V., De Faveri, J., Fordyce, G., McGowan, M., Bertram, J., Vankan, D. M., Fitzpatrick, L. A., Jayawardhana, G. A., and Miller, R. (2002). Bull selection and use in northern Australia: 4. Calf output and predictors of fertility of bulls in multiple-sire herds. *Animal Reproduction Science*, 71(1), 67-79.
- Holt, W. (2000). Basic aspects of frozen storage of semen. *Animal Reproduction Science*, 62(1), 3-22.
- Holt, W., Del Valle, I., and Fazeli, A. (2015). Heat shock protein A8 stabilizes the bull sperm plasma membrane during cryopreservation: Effects of breed, protein concentration, and mode of use. *Theriogenology*, 84(5), 693-701.

- Holt, W., and North, R. (1984). Partially irreversible cold- induced lipid phase transitions in mammalian sperm plasma membrane domains: Freeze-fracture study. *Journal of Experimental Zoology*, 230(3), 473-483.
- Hong, B., Ji, Y. H., Hong, J. H., Nam, K. Y., and Ahn, T. Y. (2002). A double-blind crossover study evaluating the efficacy of Korean red ginseng in patients with erectile dysfunction: a preliminary report. *The Journal of Urology*, 168(5), 2070-2073.
- Hoogewijs, M., De Vliegher, S., Govaere, J., De Schauwer, C., de Kruif, A., and Van Soom, A. (2012). Influence of counting chamber type on CASA outcomes of equine semen analysis. *Equine Veterinary Journal*, 44(5), 542-549.
- Hu, C., and Kitts, D. D. (2001). Free radical scavenging capacity as related to antioxidant activity and ginsenoside composition of Asian and North American ginseng extracts. *Journal of the American Oil Chemists' Society*, 78(3), 249-255.
- Hu, J.-H., Zan, L.-S., Zhao, X.-L., Li, Q.-W., Jiang, Z.-L., Li, Y.-K., and Li, X. (2010). Effects of trehalose supplementation on semen quality and oxidative stress variables in frozen-thawed bovine semen. *Journal of Animal Science*, 88(5), 1657-1662.
- Hu, J.-H., Zhao, X.-L., Tian, W.-Q., Zan, L.-S., and Li, Q.-W. (2011). Effects of vitamin E supplementation in the extender on frozen-thawed bovine semen preservation. *Animal*, 5(01), 107-112.
- Humphries, S., Evans, J. P., and Simmons, L. W. (2008). Sperm competition: linking form to function. *BMC Evolutionary Biology*, 8(1), 1.
- Husen, R., Pihie, A. H. L., and Nallappan, M. (2004). Screening for antihyperglycaemic activity in several local herbs of Malaysia. *Journal of Ethnopharmacology*, 95(2), 205-208.
- Hwang, S.-Y., Sohn, S.-H., Wee, J.-J., Yang, J.-B., Kyung, J.-S., Kwak, Y.-S., Kim, S.-W., and Kim, S.-K. (2010). *Panax ginseng* improves senile testicular function in rats. *Journal Ginseng Research*, 34, 327-335.
- Hwang, S. H., Lee, B.-H., Kim, H.-J., Cho, H.-J., Shin, H.-C., Im, K.-S., Choi, S.-H., Shin, T.-J., Lee, S.-M., and Nam, S. W. (2013). Suppression of metastasis of intravenously-inoculated B16/F10 melanoma cells by the novel ginseng-derived ingredient, gintonin: involvement of autotaxin inhibition. *International Journal of Oncology*, 42(1), 317-326.
- Hwang, S. H., Shin, E.-J., Shin, T.-J., Lee, B.-H., Choi, S.-H., Kang, J., Kim, H.-J., Kwon, S.-H., Jang, C.-G., and Lee, J.-H. (2012). Gintonin, a ginseng-derived lysophosphatidic acid receptor ligand, attenuates Alzheimer's disease-related neuropathies: involvement of non-amyloidogenic processing. *Journal of Alzheimer's Disease*, 31(1), 207-223.

- Hyakutake, T., Suzuki, H., and Yamamoto, S. (2015). Effect of non-Newtonian fluid properties on bovine sperm motility. *Journal of Biomechanics*, 48(12), 2941-2947.
- Ibănescu, I., Leiding, C., Ciornei, Ș. G., Roșca, P., Sfartz, I., and Drugociu, D. (2016). Differences in CASA output according to the chamber type when analyzing frozen-thawed bull sperm. *Animal Reproduction Science*, 166, 72-79.
- Imoedemhe, D. A., Sigue, A. B., Pacpaco, E. L., and Olazo, A. B. (1992). The effect of caffeine on the ability of spermatozoa to fertilize mature human oocytes. *Journal of Assisted Reproduction and Genetics*, 9(2), 155-160.
- Iqbal, S., Riaz, A., Andrabi, S., Shahzad, Q., Durrani, A., and Ahmad, N. (2016). L-Cysteine improves antioxidant enzyme activity, post-thaw quality and fertility of Nili-Ravi buffalo (*Bubalus bubalis*) bull spermatozoa. *Andrologia*, 48(9) 943-949.
- Ismail, S. B., Wan Mohammad, W. M. Z., George, A., Nik Hussain, N. H., Musthapa Kamal, Z. M., and Liske, E. (2012). Randomized clinical trial on the use of PHYSTA freeze-dried water extract of *Eurycoma longifolia* for the improvement of quality of life and sexual well-being in men. *Evidence-Based Complementary and Alternative Medicine*, ID 429268.
- James, P., Wolfe, C., Ladha, S., and Jones, R. (1999). Lipid diffusion in the plasma membrane of ram and boar spermatozoa during maturation in the epididymis measured by fluorescence recovery after photobleaching. *Molecular Reproduction And Development*, 52(2), 207-215.
- Jamil-ur-Rahman, H., Ahmad, N., Najib-ur-Rahman, S. W., Ahmad, M., Younis, M., and Ahmad, T. (2012). Effects of different levels of pigeon egg yolk in extenders on the post-thaw semen quality of Sahiwal bulls. *Pakistan Veterinary Journal*, 32(3), 315-318.
- Jang, D. J., Lee, M. S., Shin, B. C., Lee, Y. C., and Ernst, E. (2008). Red ginseng for treating erectile dysfunction: a systematic review. *British Journal of Clinical Pharmacology*, 66(4), 444-450.
- Januskauskas, A., Gil, J., Söderquist, L., Hrd, M., Hrd, M. C., Johannisson, A., and Rodriguez-Martinez, H. (1999). Effect of cooling rates on post-thaw sperm motility, membrane integrity, capacitation status and fertility of dairy bull semen used for artificial insemination in Sweden. *Theriogenology*, 52(4), 641-658.
- Januskauskas, A., Johannisson, A., Söderquist, L., and Rodriguez-Martinez, H. (2000). Assessment of sperm characteristics post-thaw and response to calcium ionophore in relation to fertility in Swedish dairy AI bulls. *Theriogenology*, 53(4), 859-875.

- Jeyendran, R., Van der Ven, H., Perez-Pelaez, M., Crabo, B., and Zaneveld, L. (1984). Development of an assay to assess the functional integrity of the human sperm membrane and its relationship to other semen characteristics. *Journal of Reproduction and Fertility*, 70(1), 219-228.
- Jiménez-Rabadán, P., Ramón, M., García-Álvarez, O., Maroto-Morales, A., Del Olmo, E., Pérez-Guzmán, M., Bisbal, A., Fernández-Santos, M. R., Garde, J. J., and Soler, A. (2012). Effect of semen collection method (artificial vagina vs. electroejaculation), extender and centrifugation on post-thaw sperm quality of Blanca-Celtibérica buck ejaculates. *Animal Reproduction Science*, 132(1), 88-95.
- Jones, J. M., and Bavister, B. D. (2000). Acidification of intracellular pH in bovine spermatozoa suppresses motility and extends viable life. *Journal of Andrology*, 21(5), 616-624.
- Jung, C.-H., Seog, H.-M., Choi, I.-W., and Cho, H.-Y. (2005). Antioxidant activities of cultivated and wild Korean ginseng leaves. *Food Chemistry*, 92(3), 535-540.
- Kaefer, C., Komninou, E. R., Campos, V. F., de Leon, P. M., Arruda, F. V. S., Nascimento, K. S., Teixeira, E. H., Stefanello, F. M., Barschak, A. G., and Deschamps, J. C. (2013). Binding pattern and toxicological effects of lectins from genus *Canavalia* on bovine sperm. *Reproductive Toxicology*, 38, 72-80.
- Kaka, A., Haron, W., Leghari, R. A., Memon, M. I., Kaka, U., Mirani, A. H., Naeem, M., and Kalwar, Q. (2016). Effect of in-vitro supplementation of polyunsaturated fatty acids on frozen-thawed bull sperm characteristics using Bioxcell[®] extender. *Pure and Applied Biology*, 5 (3), 399-405.
- Kaka, A., Wahid, H., Rosnina, Y., Yimer, N., Khumran, A., Behan, A. A., and Ebrahimi, M. (2015). Alpha- Linolenic Acid Supplementation in Tris Extender Can Improve Frozen–Thawed Bull Semen Quality. *Reproduction in Domestic Animals*, 50(1), 29-33.
- Kaka, A., Wahid, H., Rosnina, Y., Yimer, N., Khumran, A., Sarsaifi, K., Kaka, U., and Ebrahimi, M. (2015). α -Linolenic acid supplementation in BioXcell[®] extender can improve the quality of post-cooling and frozen-thawed bovine sperm. *Animal Reproduction Science*, 153, 1-7.
- Kandelousi, M. S., Arshami, J., Naserian, A., and Abavisani, A. (2013). The effects of addition of omega-3, 6, 9 fatty acids on the quality of bovine chilled and frozen-thawed sperm. *Open Veterinary Journal*, 3(1), 47.
- Kasimanickam, R., Kasimanickam, V., Thatcher, C., Nebel, R., and Cassell, B. (2007). Relationships among lipid peroxidation, glutathione peroxidase, superoxide dismutase, sperm parameters, and competitive index in dairy bulls. *Theriogenology*, 67(5), 1004-1012.

- Kathiravan, P., Kalatharan, J., Karthikeya, G., Rengarajan, K., and Kadirvel, G. (2011). Objective Sperm Motion Analysis to Assess Dairy Bull Fertility Using Computer- Aided System—A Review. *Reproduction in Domestic Animals*, 46(1), 165-172.
- Kavitha, N., Noordin, R., Chan, K.-L., and Sasidharan, S. (2012). *In vitro* anti-toxoplasma gondii activity of root extract/fractions of eurycoma longifolia jack. *BMC Complementary and Alternative Medicine*, 12(1), 1.
- Kazi, J. A., and Yaakob, N. A. (2015). Eurycoma longifolia Jack (Tongkat Ali) induced c-Fos expression in sensory and motor neurons of the rat brain nervous system. *Advances in Environmental Biology*, 9(17 S3), 24-32.
- Khalifa, T., Rekkas, C., Lymberopoulos, A., Sioga, A., Dimitriadis, I., and Papanikolaou, T. (2008). Factors affecting chromatin stability of bovine spermatozoa. *Animal Reproduction Science*, 104(2), 143-163.
- Khanam, Z., Wen, C. S., and Bhat, I. U. H. (2015). Phytochemical screening and antimicrobial activity of root and stem extracts of wild Eurycoma longifolia Jack (Tongkat Ali). *Journal of King Saud University-Science*, 27(1), 23-30.
- Khanijo, T. and Jiraungkoorskul, W. (2016). Review ergogenic effect of long jack, Eurycoma Longifolia. *Pharmacognosy Reviews*, 10(20): 139–142.
- Khoshvaght, A., Towhidi, A., Zare-shahneh, A., Noruozi, M., Zhandi, M., Davachi, N. D., and Karimi, R. (2015). Dietary n-3 PUFAs improve fresh and post-thaw semen quality in Holstein bulls via alteration of sperm fatty acid composition. *Theriogenology*, 65(5), 807-812.
- Khumran, A., Yimer, N., Rosnina, Y., Ariff, M., Wahid, H., Kaka, A., Ebrahimi, M., and Sarsaifi, K. (2015a). Butylated hydroxytoluene can reduce oxidative stress and improve quality of frozen–thawed bull semen processed in lecithin and egg yolk based extenders. *Animal Reproduction Science*, 163, 128-134.
- Kim, D. H., Jung, J. S., Suh, H. W., Huh, S. O., Min, S.-K., Son, B. K., Park, J. H., Kim, N. D., Kim, Y. H., and Song, D. K. (1998). Inhibition of stress-induced plasma corticosterone levels by ginsenosides in mice: involvement of nitric oxide. *Neuroreport*, 9(10), 2261-2264.
- Kim, H.-J., Lee, S.-G., Chae, I.-G., Kim, M.-J., Im, N.-K., Yu, M.-H., Lee, E.-J., and Lee, I.-S. (2011). Antioxidant effects of fermented red ginseng extracts in streptozotocin-induced diabetic rats. *Journal of Ginseng Research*, 35(2), 129-137.
- Kim, J.-H., Lee, J.-H., Jeong, S. M., Lee, B.-H., Yoon, I.-S., Lee, J.-H., Choi, S.-H., Kim, D.-H., Park, T.-K., and Kim, B.-K. (2006). Stereospecific effects of ginsenoside Rg3 epimers on swine coronary artery contractions. *Biological and Pharmaceutical Bulletin*, 29(2), 365-370.

- Kim, N. D., Kim, E. M., Kang, K. W., Cho, M. K., Choi, S. Y., and Kim, S. G. (2003). Ginsenoside Rg3 inhibits phenylephrine- induced vascular contraction through induction of nitric oxide synthase. *British Journal of Pharmacology*, 140(4), 661-670.
- Kim, S.-H., Shim, S.-H., Choi, D.-S., Kim, J.-H., Kwon, Y.-B., and Kwon, J.-K. (2011). Modulation of LPS-stimulated astroglial activation by ginseng total saponins. *Journal of Ginseng Research*, 35(1), 80-85.
- Kim, S.-J., and Kim, A. K. (2015). Anti-breast cancer activity of Fine Black ginseng (*Panax ginseng* Meyer) and ginsenoside Rg5. *Journal of Ginseng Research*, 39(2), 125-134.
- Kondracki, S., Banaszewska, D., Wysokińska, A., and Iwanina, M. (2012). The effect of sperm concentration in the ejaculate on morphological traits of bull spermatozoa. *Folia Biologica*, 60(1-2), 85-91.
- Kopalli, S. R., Cha, K.-M., Jeong, M.-S., Lee, S.-H., Sung, J.-H., Seo, S.-K., and Kim, S.-K. (2015a). Pectinase-treated *Panax ginseng* ameliorates hydrogen peroxide-induced oxidative stress in GC-2 sperm cells and modulates testicular gene expression in aged rats. *Journal of Ginseng Research*, 40(2), 185-195.
- Kopalli, S. R., Hwang, S.-Y., Won, Y.-J., Kim, S.-W., Cha, K.-M., Han, C.-K., Hong, J.-Y., and Kim, S.-K. (2015b). Korean red ginseng extract rejuvenates testicular ineffectiveness and sperm maturation process in aged rats by regulating redox proteins and oxidative defense mechanisms. *Experimental Gerontology*, 69, 94-102.
- Kopalli, S. R., Won, Y.-J., Hwang, S.-Y., Cha, K.-M., Kim, S.-Y., Han, C.-K., Lee, S.-H., Hong, J.-Y., and Kim, S.-K. (2016). Korean red ginseng protects against doxorubicin-induced testicular damage: An experimental study in rats. *Journal of Functional Foods*, 20, 96-107.
- Koprowski, E. (2017). Systematic Review of *Eurycoma longifolia* Reports Herb Improves Health, Vigor for Male and Female Subjects. *Journal of Nutraceuticals and Food Science*, 2 (1), 1-3.
- Kotirum, S., Ismail, S. B., and Chaiyakunapruk, N. (2015). Efficacy of Tongkat Ali (*Eurycoma longifolia*) on erectile function improvement: Systematic review and meta-analysis of randomized controlled trials. *Complementary Therapies in Medicine*, 23(5), 693-698.
- Kumar, P., Kumar, D., Sikka, P., and Singh, P. (2015). Sericin supplementation improves semen freezability of buffalo bulls by minimizing oxidative stress during cryopreservation. *Animal Reproduction Science*, 152, 26-31.

- Kumar, R., Mohanarao, G. J., and Atreja, S. (2011). Freeze-thaw induced genotoxicity in buffalo (*Bubalus bubalis*) spermatozoa in relation to total antioxidant status. *Molecular Biology Reports*, 38(3), 1499-1506.
- Kundu, C., Chakraborty, J., Dutta, P., Bhattacharyya, D., Ghosh, A., and Majumder, G. (2000). Development of a simple sperm cryopreservation model using a chemically defined medium and goat cauda epididymal spermatozoa. *Cryobiology*, 40(2), 117-125.
- Kuo, P.-C., Damu, A. G., Lee, K.-H., and Wu, T.-S. (2004). Cytotoxic and antimalarial constituents from the roots of *Eurycoma longifolia*. *Bioorganic and Medicinal Chemistry*, 12(3), 537-544.
- Kuo, P.-C., Shi, L.-S., Damu, A. G., Su, C.-R., Huang, C.-H., Ke, C.-H., Wu, J.-B., Lin, A.-J., Bastow, K. F., and Lee, K.-H. (2003). Cytotoxic and antimalarial β -carboline alkaloids from the roots of *Eurycoma longifolia*. *Journal of Natural Products*, 66(10), 1324-1327.
- Lasiene, K., Gedrimas, V., Vitkus, A., Glinskyte, S., Lasys, V., Valanciute, A., and Sienkiewicz, W. (2013). Evaluation of morphological criteria of sperm quality before *in vitro* fertilization and intracytoplasmic sperm injection. *Polish Journal of Veterinary Sciences*, 16(4), 773-785.
- Leboeuf, B., Guillouet, P., Batellier, F., Bernelas, D., Bonne, J., Forgerit, Y., Renaud, G., and Magistrini, M. (2003). Effect of native phosphocaseinate on the *in vitro* preservation of fresh semen. *Theriogenology*, 60(5), 867-877.
- Lecewicz, M., Kordan, W., Majewska, A., Kamiński, S., Dziekońska, A., and Mietelska, K. (2016). Effects of the platelet-activating factor (PAF) on selected quality parameters of cryopreserved bull semen (AI) with reduced sperm motility. *Polish Journal of Veterinary Sciences*, 19(1), 147-158.
- Lee, C. H., and Kim, J.-H. (2014). A review on the medicinal potentials of ginseng and ginsenosides on cardiovascular diseases. *Journal of Ginseng Research*, 38(3), 161-166.
- Lee, H.-U., Bae, E.-A., Han, M. J., and Kim, D.-H. (2005). Hepatoprotective effect of 20 (S)-ginsenosides Rg3 and its metabolite 20 (S)-ginsenoside Rh2 on tert-butyl hydroperoxide-induced liver injury. *Biological and Pharmaceutical Bulletin*, 28(10), 1992-1994.
- Leibo, S., McGrath, J., and Cravalho, E. (1978). Microscopic observation of intracellular ice formation in unfertilized mouse ova as a function of cooling rate. *Cryobiology*, 15(3), 257-271.
- Lenz, R., Kjelland, M., VonderHaar, K., Swannack, T., and Moreno, J. (2011). A comparison of bovine seminal quality assessments using different viewing chambers with a computer-assisted semen analyzer. *Journal of Animal Science*, 89(2), 383-388.

- Lenzi, A., Picardo, M., Gandini, L., and Dondero, F. (1996). Lipids of the sperm plasma membrane: from polyunsaturated fatty acids considered as markers of sperm function to possible scavenger therapy. *Human Reproduction Update*, 2(3), 246-256.
- Lessard, C., Parent, S., Leclerc, P., Baileys, J. L., and Sullivan, R. (2000). Cryopreservation alters the levels of the bull sperm surface protein P25b. *Journal of Andrology*, 21(5), 700-707.
- Leung, K. W., and Wong, A. S. (2013). Ginseng and male reproductive function. *Spermatogenesis*, 3(3), e26391.
- Lewis, S., and Aitken, R. (2005). DNA damage to spermatozoa has impacts on fertilization and pregnancy. *Cell and Tissue Research*, 322(1), 33-41.
- Lewis, S. E., and Simon, L. (2010). Clinical implications of sperm DNA damage. *Human Fertility*, 13(4), 201-207.
- Li, H., Lee, J.-H., and Ha, J.-M. (2008). Effective purification of ginsenosides from cultured wild ginseng roots, red ginseng, and white ginseng with macroporous resins. *Journal of Microbiology and Biotechnology*, 18(11), 1789-1791.
- Lim, B. O., Yu, B. P., Oh, J. H., and Park, D. K. (1998). The inhibitory effects of ginsenoside and quercetin on oxidative damage by puromycin aminonucleoside in rat. *Phytotherapy Research*, 12(5), 375-377.
- Low, B.-S., Choi, S.-B., Wahab, H. A., Das, P. K., and Chan, K.-L. (2013). Eurycomanone, the major quassinoid in *Eurycoma longifolia* root extract increases spermatogenesis by inhibiting the activity of phosphodiesterase and aromatase in steroidogenesis. *Journal of Ethnopharmacology*, 149(1), 201-207.
- Low, B.-S., Das, P. K., and Chan, K.-L. (2013). Standardized quassinoid-rich *Eurycoma longifolia* extract improved spermatogenesis and fertility in male rats via the hypothalamic–pituitary–gonadal axis. *Journal of Ethnopharmacology*, 145(3), 706-714.
- Lulu, T., Park, S.-Y., Ibrahim, R., and Paek, K.-Y. (2015). Production of biomass and bioactive compounds from adventitious roots by optimization of culturing conditions of *Eurycoma longifolia* in balloon-type bubble bioreactor system. *Journal of Bioscience and Bioengineering*, 119(6), 712-717.
- Lymberopoulos, A., and Khalifa, T. (2010). Sperm chromatin stability during *in vitro* manipulation of beef bull semen. *Reproduction in Domestic Animals*, 45(2), 307-314.

- Makker, K., Agarwal, A., and Sharma, R. (2009). Oxidative stress and male infertility. *Indian Journal of Medical Research*, 129, 357-367.
- Makler, A., David, R., Blumenfeld, Z., and Better, O. S. (1981). Factors affecting sperm motility. VII. Sperm viability as affected by change of pH and osmolarity of semen and urine specimens. *Fertility and Sterility*, 36(4), 507-511.
- Malik, A., Laily, M., and Zakir, M. I. (2015). Effects of long term storage of semen in liquid nitrogen on the viability, motility and abnormality of frozen thawed Frisian Holstein bull spermatozoa. *Asian Pacific Journal of Reproduction*, 4(1), 22-25.
- Malik, A., Yayan, M., and Djaya, M. S. (2016). Effects of Addition of Juice Date Palm to the Extender on the Semen Qualities of Frozen Thawed in Bull Spermatozoa. *Global Veterinaria*. 16(1),100-104.
- Manicardi, G., Bianchi, P., Pantano, S., Azzoni, P., Bizzaro, D., Bianchi, U., and Sakkas, D. (1995). Presence of endogenous nicks in DNA of ejaculated human spermatozoa and its relationship to chromomycin A3 accessibility. *Biology of Reproduction*, 52(4), 864-867.
- Manjunath, P., and Thérien, I. (2002). Role of seminal plasma phospholipid-binding proteins in sperm membrane lipid modification that occurs during capacitation. *Journal of Reproductive Immunology*, 53(1), 109-119.
- Marco- Jiménez, F., Vicente, J., and Viudes- de- Castro, M. (2008). Seminal plasma composition from ejaculates collected by artificial vagina and electroejaculation in Guirra ram. *Reproduction in Domestic Animals*, 43(4), 403-408.
- Marin-Guzman, J., Mahan, D., and Whitmoyer, R. (2000). Effect of dietary selenium and vitamin E on the ultrastructure and ATP concentration of boar spermatozoa, and the efficacy of added sodium selenite in extended semen on sperm motility. *Journal of Animal Science*, 78(6), 1544-1550.
- Maroto-Morales, A., Ramón, M., Garcia-Alvarez, O., Soler, A., Estesó, M., Martínez-Pastor, F., Pérez-Guzmán, M. D., and Garde, J. (2010). Characterization of ram (*Ovis aries*) sperm head morphometry using the Sperm-Class Analyzer. *Theriogenology*, 73(4), 437-448.
- Martin, G., Sabido, O., Durand, P., and Levy, R. (2004). Cryopreservation induces an apoptosis-like mechanism in bull sperm. *Biology of Reproduction*, 71(1), 28-37.
- Martinez-Pastor, F., Guerra, C., Kaabi, M., Diaz, A., Anel, E., Herraéz, P., De Paz, P., and Anel, L. (2005). Decay of sperm obtained from epididymes of wild ruminants depending on postmortem time. *Theriogenology*, 63(1), 24-40.

- Martins, C., Driessen, K., Costa, P. M., Carvalho-Neto, J., De Sousa, R., Rumpf, R., and Dode, M. (2009). Recovery, cryopreservation and fertilization potential of bovine spermatozoa obtained from epididymides stored at 5 C by different periods of time. *Animal Reproduction Science*, 116(1), 50-57.
- Martins, C., Rumpf, R., Pereira, D., and Dode, M. (2007). Cryopreservation of epididymal bovine spermatozoa from dead animals and its uses *in vitro* embryo production. *Animal Reproduction Science*, 101(3), 326-331.
- Martins, S. G., Miranda, P. V., and Brandelli, A. (2003). Acrosome reaction inhibitor released during *in vitro* sperm capacitation. *International Journal of Andrology*, 26(5), 296-304.
- Mazur, P., and Rigopoulos, N. (1983). Contributions of unfrozen fraction and of salt concentration to the survival of slowly frozen human erythrocytes: influence of warming rate. *Cryobiology*, 20(3), 274-289.
- Medeiros, C., Forell, F., Oliveira, A., and Rodrigues, J. (2002). Current status of sperm cryopreservation: why isn't it better? *Theriogenology*, 57(1), 327-344.
- Meng, D., Li, X., Han, L., Zhang, L., An, W., and Li, X. (2014). Four new quassinoids from the roots of *Eurycoma longifolia* Jack. *Fitoterapia*, 92, 105-110.
- Métayer, S., Dacheux, F., Dacheux, J.-L., and Gatti, J.-L. (2002). Comparison, characterization, and identification of proteases and protease inhibitors in epididymal fluids of domestic mammals. Matrix metalloproteinases are major fluid gelatinases. *Biology of Reproduction*, 66(5), 1219-1229.
- Mittal, P. K., Anand, M., Madan, A., Yadav, S., and Kumar, J. (2014). Antioxidative capacity of vitamin E, vitamin C and their combination in cryopreserved Bhadavari bull semen. *Veterinary World*, 7(12), 1127-1131.
- Miyake, K., Tezuka, Y., Awale, S., Li, F., and Kadota, S. (2009). Quassinoids from *Eurycoma longifolia*. *Journal of Natural Products*, 72(12), 2135-2140.
- Moce, E., and Graham, J. (2006). Cholesterol-loaded cyclodextrins added to fresh bull ejaculates improve sperm cryosurvival. *Journal of Animal Science*, 84(4), 826-833.
- Mohri, H., and Masaki, J. (1967). Glycerokinase and its possible role in glycerol metabolism of bull spermatozoa. *Journal of Reproduction And Fertility*, 14(2), 179-194.
- Moore, A., Squires, E., Bruemmer, J., and Graham, J. (2006). Effect of cooling rate and cryoprotectant on the cryosurvival of equine spermatozoa. *Journal of Equine Veterinary Science*, 26(5), 215-218.

- Mostafa, T., Elbadry, D., and Anour, A. M. (2014). Freezability and Dna Integrity of Dromedary Camel Spermatozoa in Semen Collected by Artificial Vagina and Electro-Ejaculator. *Egyptian Journal of Animal Production*, 51(2), 145-155.
- Motlagh, M. K., Sharafi, M., Zhandi, M., Mohammadi-Sangcheshmeh, A., Shakeri, M., Soleimani, M., and Zeinoaldini, S. (2014). Antioxidant effect of rosemary (*Rosmarinus officinalis* L.) extract in soybean lecithin-based semen extender following freeze–thawing process of ram sperm. *Cryobiology*, 69(2), 217-222.
- Moura, A. A., Souza, C. E., Stanley, B. A., Chapman, D. A., and Killian, G. J. (2010). Proteomics of cauda epididymal fluid from mature Holstein bulls. *Journal of proteomics*, 73(10), 2006-2020.
- Moussa, M., Martinet, V., Trimeche, A., Tainturier, D., and Anton, M. (2002). Low density lipoproteins extracted from hen egg yolk by an easy method: cryoprotective effect on frozen–thawed bull semen. *Theriogenology*, 57(6), 1695-1706.
- Moustafa, M. H., Sharma, R. K., Thornton, J., Mascha, E., Abdel- Hafez, M. A., Thomas, A. J., and Agarwal, A. (2004). Relationship between ROS production, apoptosis and DNA denaturation in spermatozoa from patients examined for infertility. *Human Reproduction*, 19(1), 129-138.
- Muhamad, S., Pihie, A. H. L., Latif, J., Rha, C., and Sambandan, T. (2011). Induction of apoptosis in MCF-7 via the Caspase pathway by longilactone from *Eurycoma longifolia* Jack. *Research in Pharmaceutical Biotechnology*, 3(1), 1-10.
- Muldrew, K., and McGann, L. E. (1990). Mechanisms of intracellular ice formation. *Biophysical Journal*, 57(3), 525.
- Munsi, M., Bhuiyan, M., Majumder, S., and Alam, M. (2007). Effects of exogenous glutathione on the quality of chilled bull semen. *Reproduction in Domestic Animals*, 42(4), 358-362.
- Murphy, C., Fahey, A., Shafat, A., and Fair, S. (2013). Reducing sperm concentration is critical to limiting the oxidative stress challenge in liquid bull semen. *Journal of Dairy Science*, 96(7), 4447-4454.
- Naaby-Hansen, S., Flickinger, C. J., and Herr, J. C. (1997). Two-dimensional gel electrophoretic analysis of vectorially labeled surface proteins of human spermatozoa. *Biology of Reproduction*, 56(3), 771-787.
- Nagy, S., Johannisson, A., Wahlsten, T., Ijäs, R., Andersson, M., and Rodriguez-Martinez, H. (2013). Sperm chromatin structure and sperm morphology: Their association with fertility in AI-dairy Ayrshire sires. *Theriogenology*, 79(8), 1153-1161.

- Nair, S. J., Brar, A., Ahuja, C., Sangha, S., and Chaudhary, K. (2006). A comparative study on lipid peroxidation, activities of antioxidant enzymes and viability of cattle and buffalo bull spermatozoa during storage at refrigeration temperature. *Animal Reproduction Science*, 96(1), 21-29.
- Najafi, A., Daghigh-Kia, H., Dodaran, H. V., Mehdipour, M., and Alvarez-Rodriguez, M. (2016). Ethylene glycol, but not DMSO, could replace glycerol inclusion in soybean lecithin-based extenders in ram sperm cryopreservation. *Animal Reproduction Science*, 177, 35-41.
- Nasiri, A., Towhidi, A., and Zeinoaldini, S. (2012). Combined effect of DHA and α -tocopherol supplementation during bull semen cryopreservation on sperm characteristics and fatty acid composition. *Andrologia*, 44(s1), 550-555.
- Nocerino, E., Amato, M., and Izzo, A. A. (2000). The aphrodisiac and adaptogenic properties of ginseng. *Fitoterapia*, 71, S1-S5.
- Noiles, E., Ruffing, N., Kleinhans, F., Mark, L., Watson, P., Critser, J., Horstman, L., and Mazur, P. (1990). Critical tonicity determination of sperm using fluorescent staining and flow cytometry: Oak Ridge National Lab., TN (USA).
- Nur, Z., Dogan, I., Soylu, M., and Ak, K. (2003). Effect of different thawing procedures on the quality of bull semen. *Revue de Médecine Vétérinaire*, 154(7), 487-490.
- Nurhanan, M., Hawariah, L., Ilham, A. M., and Shukri, M. (2005). Cytotoxic effects of the root extracts of *Eurycoma longifolia* Jack. *Phytotherapy Research*, 19(11), 994-996.
- O'Kelly, F., Manecksha, R. P., Cullen, I. M., McDermott, T. E., Flynn, R., and Grainger, R. (2011). Electroejaculatory stimulation and its implications for male infertility in spinal cord injury: a short history through four decades of sperm retrieval (1975-2010). *Urology*, 77(6), 1349-1352.
- O'Hara, L., Hanrahan, J., Richardson, L., Donovan, A., Fair, S., Evans, A., and Lonergan, P. (2010). Effect of storage duration, storage temperature, and diluent on the viability and fertility of fresh ram sperm. *Theriogenology*, 73(4), 541-549.
- Oh, K. J., Chae, M. J., Lee, H. S., Hong, H. D., and Park, K. (2010). Effects of Korean red ginseng on sexual arousal in menopausal women: Placebo- controlled, double- blind crossover clinical study. *The journal of Sexual Medicine*, 7(4), 1469-1477.
- Ohl, D. (1993). Electroejaculation. *The Urologic clinics of North America*, 20(1), 181-188.

- Ostermeier, G. C., Sargeant, G. A., Yandell, B. S., Evenson, D. P., and Parrish, J. J. (2001). Relationship of bull fertility to sperm nuclear shape. *Journal of Andrology*, 22(4), 595-603.
- Ostermeier, G. C., Sargeant, G. A., Yandell, B. S., and Parrish, J. J. (2001). Measurement of bovine sperm nuclear shape using Fourier harmonic amplitudes. *Journal of Andrology*, 22(4), 584-594.
- Palmer, C., Amundson, S., Brito, L., Waldner, C., and Barth, A. (2004). Use of oxytocin and cloprostenol to facilitate semen collection by electroejaculation or transrectal massage in bulls. *Animal Reproduction Science*, 80(3), 213-223.
- Palmer, C., Brito, L., Arteaga, A., Söderquist, L., Persson, Y., and Barth, A. (2005). Comparison of electroejaculation and transrectal massage for semen collection in range and yearling feedlot beef bulls. *Animal Reproduction Science*, 87(1), 25-31.
- Palmer, C. W. (2005). Welfare aspects of theriogenology: investigating alternatives to electroejaculation of bulls. *Theriogenology*, 64(3), 469-479.
- Palmer, C. W. (2016). Management and Breeding Soundness of Mature Bulls. *Veterinary Clinics of North America: Food Animal Practice*, 32(2), 479-495.
- Panjaitan, R. G. P., Handharyani, E., Chairul, and Manalu, W. (2013). Hepatoprotective activity of *Eurycoma longifolia* Jack. roots. *INDIAN Journal of Traditional Knowledge*, 12(2), 225-230.
- Papa, P. M., Maziero, R. D., Guasti, P. N., Junqueira, C. R., Freitas-Dell'Aqua, C. P., Papa, F. O., Vianna, F. P., Alvarenga, M. A., Crespilho, A. M. and Dell'Aqua, J. A. (2015). Effect of glycerol on the viability and fertility of cooled bovine semen. *Theriogenology*, 83(1), 107-113.
- Papa, P. M., Papa, F. O., Oliveira, L. A., Guasti, P. N., Castilho, C., and Giometti, I. C. (2015). Different extenders in the cryopreservation of bovine epididymal spermatozoa. *Animal Reproduction Science*, 161, 58-63.
- Park, B. G., Jung, H. J., Cho, Y. W., Lim, H. W., and Lim, C. J. (2013). Potentiation of antioxidative and anti-inflammatory properties of cultured wild ginseng root extract through probiotic fermentation. *Journal of Pharmacy and Pharmacology*, 65(3), 457-464.
- Park, S., Nhiem, N. X., Van Kiem, P., Van Minh, C., Tai, B. H., Kim, N., Yoo, H. H., Song, J-H., Ko, H-J., and Kim, S. H. (2014). Five new quassinoids and cytotoxic constituents from the roots of *Eurycoma longifolia*. *Bioorganic and Medicinal Chemistry Letters*, 24(16), 3835-3840.
- Parkinson, T. (2004). Evaluation of fertility and infertility in natural service bulls. *The Veterinary Journal*, 168(3), 215-229.

- Parks, J., and Graham, J. (1992). Effects of cryopreservation procedures on sperm membranes. *Theriogenology*, 38(2), 209-222.
- Parks, J. E., and Lynch, D. V. (1992). Lipid composition and thermotropic phase behavior of boar, bull, stallion, and rooster sperm membranes. *Cryobiology*, 29(2), 255-266.
- Patel, M. K., Cheema, R. S., Bansal, A. K., and Gandotra, V. (2016). A 31-kDa Seminal Plasma Heparin Binding Protein reduces cold shock stress during Cryo-preservation of Cross-bred Cattle Bull Semen. *Theriogenology*, 86(6), 1599-1606.
- Peña, F. J., Saravia, F., García- Herreros, M., Núñezmartínez, I., Tapia, J. A., Johannisson, A., Wallgre, M., and Rodríguez- Martínez, H. (2005). Identification of sperm morphometric subpopulations in two different portions of the boar ejaculate and its relation to postthaw quality. *Journal of Andrology*, 26(6), 716-723.
- Perumal, P. (2014). Effect of Superoxide Dismutase on semen parameters and antioxidant enzyme activities of liquid stored (5 C) Mithun (*Bos frontalis*) semen. *Journal of Animals*, ID 821954.
- Phillips, N., McGowan, M., Johnston, S., and Mayer, D. (2004). Relationship between thirty post-thaw spermatozoal characteristics and the field fertility of 11 high-use Australian dairy AI sires. *Animal Reproduction Science*, 81(1), 47-61.
- Phillips, P. H., and Lardy, H. A. (1940). A yolk-buffer pabulum for the preservation of bull semen. *Journal of Dairy Science*, 23(5), 399-404.
- Pihie, A. (2002). *Current Status on the Effect of Eurycoma longifolia (Tongkat Ali) Extracts as a Sexual Stimulant Agent*. Paper presented at the Tongkat Ali, Kacip Fatimah and Pegaga: New Dimensions in Complementary Health Care. Proceedings of the Seminar on Medicinal Plants.
- Piomboni, P., Focarelli, R., Stendardi, A., Ferramosca, A., and Zara, V. (2012). The role of mitochondria in energy production for human sperm motility. *International Journal of Andrology*, 35(2), 109-124.
- Prisant, N., Escalier, D., Soufir, J., Morillon, M., Schoevaert, D., Misrahi, M., and Tachdjian, G. (2007). Ultrastructural nuclear defects and increased chromosome aneuploidies in spermatozoa with elongated heads. *Human Reproduction*, 22(4), 1052-1059.
- Purdy, P., Tharp, N., Stewart, T., Spiller, S., and Blackburn, H. (2010). Implications of the pH and temperature of diluted, cooled boar semen on fresh and frozen-thawed sperm motility characteristics. *Theriogenology*, 74(7), 1304-1310.

- Qadeer, S., Khan, M., Ansari, M., Rakha, B., Ejaz, R., Husna, A., Ashiq, M., Iqbal, R., Ullah, N., and Akhter, S. (2014). Evaluation of antifreeze protein III for cryopreservation of Nili-Ravi (*Bubalus bubalis*) buffalo bull sperm. *Animal Reproduction Science*, 148(1), 26-31.
- Ramesh, T., Kim, S.-W., Sung, J.-H., Hwang, S.-Y., Sohn, S.-H., Yoo, S.-K., and Kim, S.-K. (2012). Effect of fermented *Panax ginseng* extract (GINST) on oxidative stress and antioxidant activities in major organs of aged rats. *Experimental Gerontology*, 47(1), 77-84.
- Rego, J., Crisp, J., Moura, A., Nouwens, A., Li, Y., Venus, B., Corbet, N. J., Corbet, D. H., Burns, B. M., and Boe-Hansen, G. (2014). Seminal plasma proteome of electroejaculated *Bos indicus* bulls. *Animal Reproduction Science*, 148(1), 1-17.
- Rehman, F., Qureshi, M., and Khan, R. (2014). Effect of soybean based extenders on sperm parameters of holstein-friesian bull during liquid storage at 4°C. *Pakistan Journal of Zoology*, 46(1), 185-189.
- Revell, S., and Mrode, R. (1994). An osmotic resistance test for bovine semen. *Animal Reproduction Science*, 36(1), 77-86.
- Ribas-Maynou, J., García-Peiró, A., Fernandez-Encinas, A., Amengual, M. J., Prada, E., Cortes, P., Navarro, J., and Benet, J. (2012). Double stranded sperm DNA breaks, measured by Comet assay, are associated with unexplained recurrent miscarriage in couples without a female factor. *PloS One*, 7(9), e44679.
- Rodríguez- Martínez, H., Kvist, U., Ernerudh, J., Sanz, L., and Calvete, J. J. (2011). Seminal plasma proteins: what role do they play? *American Journal of Reproductive Immunology*, 66(s1), 11-22.
- Rubio- Guillén, J., González, D., Garde, J., Estesó, M., Fernández- Santos, M., Rodríguez- Gil, J., Madrid-Bury, N., and Quintero- Moreno, A. (2007). Effects of cryopreservation on bull spermatozoa distribution in morphometrically distinct subpopulations. *Reproduction in Domestic Animals*, 42(4), 354-357.
- Saacke, R., and Almquist, J. (1964). Ultrastructure of bovine spermatozoa. I. The head of normal, ejaculated sperm. *American Journal of Anatomy*, 115(1), 143-161.
- Saacke, R., Dalton, J., Nadir, S., Nebel, R., and Bame, J. (2000). Relationship of seminal traits and insemination time to fertilization rate and embryo quality. *Animal Reproduction Science*, 60, 663-677.
- Sakkas, D., Mariethoz, E., Manicardi, G., Bizzaro, D., Bianchi, P. G., and Bianchi, U. (1999). Origin of DNA damage in ejaculated human spermatozoa. *Reviews of Reproduction*, 4(1), 31-37.

- Sakkas, D., Moffatt, O., Manicardi, G. C., Mariethoz, E., Tarozzi, N., and Bizzaro, D. (2002). Nature of DNA damage in ejaculated human spermatozoa and the possible involvement of apoptosis. *Biology of Reproduction*, 66(4), 1061-1067.
- Salisbury, G., Fuller, H., and Willett, E. (1941). Preservation of bovine spermatozoa in yolk-citrate diluent and field results from its use. *Journal of Dairy Science*, 24(11), 905-910.
- Salisbury, G. W., VanDemark, N. L., and Lodge, J. R. (1978). *Physiology of Reproduction and Artificial Insemination of Cattle*: San Francisco (and London): W. H. Freeman and Company.
- Salvati, G., Genovesi, G., Marcellini, L., Paolini, P., De Nuccio, I., Pepe, M., and Re, M. (1996). Effects of *Panax Ginseng* CA Meyer saponins on male fertility. *Panminerva Medica*, 38(4), 249-254.
- Sambandan, T., Rha, C., Kadir, A. A., Aminudim, N., and Saad, J. M. (2006). Bioactive fraction of *Eurycoma longifolia*: Google Patents.
- Santiago-Moreno, J., Castaño, C., Toledano-Díaz, A., Estesó, M., López-Sebastián, A., Guerra, R., Ruiz, M. J., Mendoza, N., Luna, C., and Cebrián-Pérez, J. (2013). Cryopreservation of aoudad (*Ammotragus lervia sahariensis*) sperm obtained by transrectal ultrasound-guided massage of the accessory sex glands and electroejaculation. *Theriogenology*, 79(2), 383-391.
- Sapanidou, V., Taitzoglou, I., Tsakmakidis, I., Kourtzelis, I., Fletouris, D., Theodoridis, A., Zervos, I., and Tsantarliotou, M. (2015). Antioxidant effect of crocin on bovine sperm quality and *in vitro* fertilization. *Theriogenology*, 84(8), 1273-1282.
- Sariözkan, S., Bucak, M. N., Tuncer, P. B., Büyükleblebici, S., Eken, A., and Akay, C. (2015). Influence of fetuin and hyaluronan on the post-thaw quality and fertilizing ability of Holstein bull semen. *Cryobiology*, 71(1), 119-124.
- Sariözkan, S., Bucak, M. N., Tuncer, P. B., Ulutaş, P. A., and Bilgen, A. (2009). The influence of cysteine and taurine on microscopic-oxidative stress parameters and fertilizing ability of bull semen following cryopreservation. *Cryobiology*, 58(2), 134-138.
- Sariözkan, S., Tuncer, P. B., Bucak, M. N., and Ulutaş, P. A. (2009). Influence of various antioxidants on microscopic-oxidative stress indicators and fertilizing ability of frozen-thawed bull semen. *Acta Veterinaria Brno*, 78(3), 463-469.
- Sarsaifi, K., Rosnina, Y., Ariff, M., Wahid, H., Hani, H., Yimer, N., Vejayan, J., Win N. S., and Abas, M. (2013). Effect of Semen Collection Methods on the Quality of Pre- and Post- thawed Bali Cattle (*Bos javanicus*) Spermatozoa. *Reproduction in Domestic Animals*, 48(6), 1006-1012.

- Sarsaifi, K., Vejayan, J., Haron, A. W., Yusoff, R., Hani, H., Rasoli, M., Ariff, M. O., and Othman, A. M. (2015). Protein profile and functionality of spermatozoa from two semen collection methods in Bali bulls. *Livestock Science*, 172, 96-105.
- Sathananthan, A., Ratnam, S., Ng, S., Tarin, J., Gianaroli, L., and Trounson, A. (1996). The sperm centriole: its inheritance, replication and perpetuation in early human embryos. *Human Reproduction*, 11(2), 345-356.
- Schuffner, A., Morshedi, M., Vaamonde, D., Duran, E. H., And Oehninger, S. (2002). Effect of different incubation conditions on phosphatidylserine externalization and motion parameters of purified fractions of highly motile human spermatozoa. *Journal of Andrology*, 23(2), 194-201.
- Seshoka, M. M., Mphaphathi, M. L., and Nedambale, T. L. (2016). Comparison of four different permitting and combination of two best cryoprotectants on freezing Nguni sperm evaluated with the aid of computer aided sperm analysis. *Cryobiology*, 72(3), 232-238.
- Shannon, P., and Vishwanath, R. (1995). The effect of optimal and suboptimal concentrations of sperm on the fertility of fresh and frozen bovine semen and a theoretical model to explain the fertility differences. *Animal Reproduction Science*, 39(1), 1-10.
- Sharma, M., Singh, M., Kapoor, S., and Jasial, S. (2012). Inter relationship between some routine semen evaluation parameters in Jersey X local hill cattle crossbred bulls. *Open Veterinary Journal*, 2(1), 26-31.
- Sharma, O., and Hays, R. (1973). Release of an oxytocic substance following genital stimulation in bulls. *Journal of Reproduction and Fertility*, 35(2), 359-362.
- Sheikh, N., Amiri, I., Farimani, M., Najafi, R., and Hadeie, J. (2008). Correlation between sperm parameters and sperm DNA fragmentation in fertile and infertile men. *International Journal of Reproductive BioMedicine*, 6(1), 13-18.
- Shinkai, K., Akedo, H., Mukai, M., Imamura, F., Isoai, A., Kobayashi, M., and Kitagawa, I. (1996). Inhibition of *in vitro* tumor cell invasion by ginsenoside Rg3. *Japanese Journal of Cancer Research*, 87(4), 357-362.
- Shojaei, H., Kroetsch, T., Wilde, R., Blondin, P., Kastelic, J., and Thundathil, J. (2012). Moribund sperm in frozen-thawed semen, and sperm motion end points post-thaw and post-swim-up, are related to fertility in Holstein AI bulls. *Theriogenology*, 77(5), 940-951.
- Shuid, A. N., Abu Bakar, M. F., Abdul Shukor, T. A., Muhammad, N., Mohamed, N., and Soelaiman, I. N. (2011). The anti-osteoporotic effect of *Eurycoma longifolia* in aged orchidectomised rat model. *The Aging Male*, 14(3), 150-154.

- Simões, R., Feitosa, W. B., Siqueira, A. F. P., Nichi, M., Paula-Lopes, F. F., Marques, M. G., Peres, M. A., Barnabe, V. H., Visintin, L. A., and Assumpção, M. E. O. (2013). Influence of bovine sperm DNA fragmentation and oxidative stress on early embryo *in vitro* development outcome. *Reproduction*, 146(5), 433-441.
- Simon, L., and Lewis, S. E. (2011). Sperm DNA damage or progressive motility: which one is the better predictor of fertilization *in vitro*? *Systems Biology in Reproductive Medicine*, 57(3), 133-138.
- Simonik, O., Sichtar, J., Krejcarkova, A., Rajmon, R., Stadnik, L., Beran, J., Dolezalva, M., and Biniova, Z. (2015). Computer assisted sperm analysis—the relationship to bull field fertility, possible errors and their impact on outputs: A review. *Indian Journal of Animal Sciences*, 85(1), 3-11.
- Sin, S., Kim, S. Y., and Kim, S. S. (2012). Chronic treatment with ginsenoside Rg3 induces Akt-dependent senescence in human glioma cells. *International Journal of Oncology*, 41(5), 1669-1674.
- Singh, M., Sinha, A., and Singh, B. (1995). Effect of cryoprotectants on certain seminal attributes and on the fertility of buck spermatozoa. *Theriogenology*, 43(6), 1047-1053.
- Smith, D., Gaffney, E., Gadêlha, H., Kapur, N., and Kirkman- Brown, J. (2009). Bend propagation in the flagella of migrating human sperm, and its modulation by viscosity. *Cell Motility and The Cytoskeleton*, 66(4), 220-236.
- Smith, R., Berndtson, W., Unal, M., and Pickett, B. (1979). Influence of percent egg yolk during cooling and freezing on survival of bovine spermatozoa. *Journal of Dairy Science*, 62(8), 1297-1303.
- Soldati, F., and Sticher, O. (1980). HPLC separation and quantitative determination of ginsenosides from *Panax ginseng*, *Panax quinquefolium* and from ginseng drug preparations. *Planta Medica*, 39(08), 348-357.
- Songsasen, N., Tong, J., and Leibo, S. (1998). Birth of live mice derived by *in vitro* fertilization with spermatozoa retrieved up to twenty- four hours after death. *Journal of Experimental Zoology*, 280(2), 189-196.
- Sotolongo, B., Lino, E., and Ward, W. S. (2003). Ability of hamster spermatozoa to digest their own DNA. *Biology of Reproduction*, 69(6), 2029-2035.
- Stambaugh, R., and Buckley, J. (1970). Comparative studies of the acrosomal enzymes of rabbit, rhesus monkey, and human spermatozoa. *Biology of Reproduction*, 3(3), 275-282.
- Stradaoli, G., Noro, T., Sylla, L., and Monaci, M. (2007). Decrease in glutathione (GSH) content in bovine sperm after cryopreservation: comparison between two extenders. *Theriogenology*, 67(7), 1249-1255.

- Suarez, S., and Pacey, A. (2006). Sperm transport in the female reproductive tract. *Human Reproduction Update*, 12(1), 23-37.
- Summerfield, F. W., and Tappel, A. L. (1981). Determination of malondialdehyde-DNA crosslinks by fluorescence and incorporation of tritium. *Analytical Biochemistry*, 111(1), 77-82.
- Talbott, J., Christopoulos, A.-M., and Ekberg, E. (2010). Ancient wisdom meets modern ailment—traditional Asian medicine improves psychological vigor in stressed subjects. *Progress in Nutrition*, 12(1), 64-69.
- Talbott, S. M., Talbott, J. A., George, A., and Pugh, M. (2013). Effect of Tongkat Ali on stress hormones and psychological mood state in moderately stressed. *Journal of the International Society of Sports Nutrition*, 10, 28.
- Tambi, M. (2002). *Glycoprotein water-soluble extract of Eurycoma Longifolia Jack as a health supplement in management of Health aging in aged men*. Paper presented at the Proceedings of the 3rd World Congress on the Aging Male.
- Tambi, M., Imran, M., and Henkel, R. (2012). Standardised water- soluble extract of Eurycoma longifolia, Tongkat ali, as testosterone booster for managing men with late- onset hypogonadism? *Andrologia*, 44(s1), 226-230.
- Tambi, M., and Imran, M. K. (2010). Eurycoma longifolia Jack in managing idiopathic male infertility. *Asian Journal of Andrology*, 12(3), 376-380.
- Tambi, M., and Kadir, A. (2006). Eurycoma longifolia jack: a potent adaptogen in the form of water-soluble extract with the effects of maintaining men's health. *Asian Journal of Andrology*, 8(Suppl 1), 49-50.
- Taşdemir, U., Büyükleblebici, S., Tuncer, P. B., Coşkun, E., Özgürtaş, T., Aydın, F. N., Büyükleblebici, O., and Gürcan, İ. S. (2013). Effects of various cryoprotectants on bull sperm quality, DNA integrity and oxidative stress parameters. *Cryobiology*, 66(1), 38-42.
- Taşdemir, U., Tuncer, P. B., and Büyükleblebici, S. (2014). Effects of various antioxidants on cryopreserved bull sperm quality. *Kafkas University of Veterinary Faculty Derg*, 20(2), 253-258.
- Tee, T. T., and Azimahtol, H. L. P. (2005). Induction of apoptosis by Eurycoma longifolia Jack extracts. *Anticancer Research*, 25(3B), 2205-2213.
- Tee, T. T., Cheah, Y. H., and Hawariah, L. P. A. (2007). F16, a fraction from Eurycoma longifolia jack extract, induces apoptosis via a caspase-9-independent manner in MCF-7 cells. *Anticancer Research*, 27(5A), 3425-3430.

- Tejada, R., Mitchell, J. C., Norman, A., Marik, J., and Friedman, S. (1984). A test for the practical evaluation of male fertility by acridine orange (AO) fluorescence. *Fertility and Sterility*, 42(1), 87-91.
- Therrien, A., Manjunath, P., and Lafleur, M. (2013). Chemical and physical requirements for lipid extraction by bovine binder of sperm BSP1. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1828(2), 543-551.
- Thibier, M., and Wagner, H.-G. (2002). World statistics for artificial insemination in cattle. *Livestock Production Science*, 74(2), 203-212.
- Thun, R., Hurtado, M., and Janett, F. (2002). Comparison of Biociphos-Plus® and TRIS-egg yolk extender for cryopreservation of bull semen. *Theriogenology*, 57(3), 1087-1094.
- Thundathil, J., Meyer, R., Palasz, A., Barth, A., and Mapletoft, R. (2000). Effect of the knobbed acrosome defect in bovine sperm on IVF and embryo production. *Theriogenology*, 54(6), 921-934.
- Thundathil, J., Palasz, A., Mapletoft, R., and Barth, A. (1999). An investigation of the fertilizing characteristics of pyriform-shaped bovine spermatozoa. *Animal Reproduction Science*, 57(1), 35-50.
- Tian, J., Fu, F., Geng, M., Jiang, Y., Yang, J., Jiang, W., Wang, C., and Liu, K. (2005). Neuroprotective effect of 20 (S)-ginsenoside Rg 3 on cerebral ischemia in rats. *Neuroscience Letters*, 374(2), 92-97.
- Tong, K. L., Chan, K. L., AbuBakar, S., Low, B. S., Ma, H. Q., and Wong, P. F. (2015). The *In Vitro* and *In Vivo* Anti-Cancer Activities of a Standardized Quassinoids Composition from *Eurycoma longifolia* on LNCaP Human Prostate Cancer Cells. *PloS One*, 10(3), e0121752.
- Toosi, B., Gratton, G., McCorkell, R., Wynne-Edwards, K., Woodbury, M., and Lessard, C. (2013). Effects of pipothiazine palmitate on handling stress and on the characteristics of semen collected by electroejaculation in bison (*Bison bison*) bulls. *Animal Reproduction Science*, 138(1), 55-63.
- Towhidi, A., and Parks, J. (2012). Effect of n-3 fatty acids and α -tocopherol on post-thaw parameters and fatty acid composition of bovine sperm. *Journal of Assisted Reproduction and Genetics*, 29(10), 1051-1056.
- Trzcińska, M., Bryła, M., and Smoraż, Z. (2008). Effect of liquid storage on membrane integrity and mitochondrial activity: a new diagnostic method of evaluating boar sperm quality. *Journal of Animal Feed Science*, 17, 372-380.
- Tvrda, E., Tušimová, E., Kováčik, A., Paál, D., Greifová, H., Abdramanov, A., and Lukáč, N. (2016). Curcumin has protective and antioxidant properties on bull spermatozoa subjected to induced oxidative stress. *Animal Reproduction Science*, 172, 10-20.

- Valeanu, S., Johannisson, A., Lundeheim, N., Morrell, J. (2015). Seasonal variation in sperm quality parameters in Swedish red dairy bulls used for artificial insemination. *Livestock Science*, 173: 111-118.
- Van Tilburg, M., Salles, M., Silva, M., Moreira, R., Moreno, F., Monteiro-Moreira, A., Martins, J. A. M., Cândido, M. J. D., Araújo, A. A., and Moura, A. (2015). Semen variables and sperm membrane protein profile of Saanen bucks (*Capra hircus*) in dry and rainy seasons of the northeastern Brazil (3° S). *International Journal of Biometeorology*, 59(5), 561-573.
- Varghese, C., Ambrose, C., Jin, S., Lim, Y., and Keisaban, T. (2013). Antioxidant and anti-inflammatory activity of *Eurycoma longifolia* Jack, a traditional medicinal plant in Malaysia. *International Journal of Pharmaceutical Sciences and Nanotechnology*, 5(4), 1875-1878.
- Verberckmoes, S., Van Soom, A., Dewulf, J., and de Kruif, A. (2005). Comparison of three diluents for the storage of fresh bovine semen. *Theriogenology*, 63(3), 912-922.
- Verstegen, J., Iguer-Ouada, M., and Onclin, K. (2002). Computer assisted semen analyzers in andrology research and veterinary practice. *Theriogenology*, 57(1), 149-179.
- Visconti, P. E., Krapf, D., de la Vega-Beltrán, J. L., Acevedo, J. J., and Darszon, A. (2011). Ion channels, phosphorylation and mammalian sperm capacitation. *Asian Journal of Andrology*, 13(3), 395-405.
- Vishwanath, R., and Shannon, P. (1996). Do sperm cells age? A review of the physiological changes in sperm during storage at ambient temperature. *Reproduction, Fertility, and Development*, 9(3), 321-331.
- Vishwanath, R., and Shannon, P. (2000). Storage of bovine semen in liquid and frozen state. *Animal Reproduction Science*, 62(1), 23-53.
- Voces, J., Alvarez, A., Vila, L., Ferrando, A., De Oliveira, C. C., and Prieto, J. (1999). Effects of administration of the standardized *Panax ginseng* extract G115 on hepatic antioxidant function after exhaustive exercise. *Comparative Biochemistry and Physiology Part C: Pharmacology, Toxicology and Endocrinology*, 123(2), 175-184.
- Wahab, N. A., Mokhtar, N. M., Halim, W. N. H. A., and Das, S. (2010). The effect of *Eurycoma longifolia* Jack on spermatogenesis in estrogen-treated rats. *Clinics*, 65(1), 93-98.
- Wang, T., Yu, X., Qu, S., Xu, H., Han, B., and Sui, D. (2010). Effect of ginsenoside Rb3 on myocardial injury and heart function impairment induced by isoproterenol in rats. *European Journal of Pharmacology*, 636(1), 121-125.

- Ward, W. S., and Coffey, D. (1991). DNA packaging and organization in mammalian spermatozoa: comparison with somatic cells. *Biology of Reproduction*, 44(4), 569-574.
- Waterhouse, K., Haugan, T., Kommisrud, E., Tverdal, A., Flatberg, G., Farstad, W., Evenson, D. P., and De Angelis, P. (2006). Sperm DNA damage is related to field fertility of semen from young Norwegian Red bulls. *Reproduction, Fertility and Development*, 18(7), 781-788.
- Watson, P. (1995). Recent developments and concepts in the cryopreservation of spermatozoa and the assessment of their post-thawing function. *Reproduction, Fertility and Development*, 7(4), 871-891.
- Watson, P. (2000). The causes of reduced fertility with cryopreserved semen. *Animal Reproduction Science*, 60, 481-492.
- Wei, X., Su, F., Su, X., Hu, T., and Hu, S. (2012). Stereospecific antioxidant effects of ginsenoside Rg3 on oxidative stress induced by cyclophosphamide in mice. *Fitoterapia*, 83(4), 636-642.
- Wernsdorfer, W. H., Ismail, S., Chan, K. L., Congpuong, K., and Wernsdorfer, G. (2009). Activity of *Eurycoma longifolia* root extract against *Plasmodium falciparum* *in vitro*. *Wiener klinische Wochenschrift*, 121(3), 23-26.
- Whitlock, B. K., Coffman, E. A., Coetzee, J. F., and Daniel, J. A. (2012). Electroejaculation increased vocalization and plasma concentrations of cortisol and progesterone, but not substance P, in beef bulls. *Theriogenology*, 78(4), 737-746.
- Woelders, H. (1991). Overview of *in vitro* methods for evaluation of semen quality. *Reprod Domestic Animal Supplement*, 1, 145-164.
- Woelders, H., and Malva, A. (1998). How important is the cooling rate in cryopreservation of (bull) semen, and what is its relation to thawing rate and glycerol concentration. *Reproduction in Domestic Animals*, 33(3- 4), 299-305.
- Woelders, H., Matthijs, A., and Engel, B. (1997). Effects of trehalose and sucrose, osmolality of the freezing medium, and cooling rate on viability and intactness of bull sperm after freezing and thawing. *Cryobiology*, 35(2), 93-105.
- Won, Y.-J., Kim, B.-k., Shin, Y.-K., Jung, S.-H., Yoo, S.-K., Hwang, S.-Y., Song, J.-H., and Kim, S.-K. (2014). Pectinase-treated *Panax ginseng* extract (GINST) rescues testicular dysfunction in aged rats via redox-modulating proteins. *Experimental Gerontology*, 53, 57-66.

- Xiong, Y., Shen, L., Liu, K. J., Tso, P., Xiong, Y., Wang, G., Woods, S. C., and Liu, M. (2010). Antiobesity and antihyperglycemic effects of ginsenoside Rb1 in rats. *Diabetes*, 59(10), 2505-2512.
- Yayeh, T., Jung, K.-H., Jeong, H.-Y., Park, J.-H., Song, Y.-B., Kwak, Y.-S., Kang, H.-S., Cho, J.-Y., Oh, J.-W., and Kim, S.-K. (2012). Korean red ginseng saponin fraction downregulates proinflammatory mediators in LPS stimulated RAW264. 7 cells and protects mice against endotoxic shock. *Journal of Ginseng Research*, 36(3), 263-269.
- Yimer, N., Muhammad, N., Sarsaifi, K., Rosnina, Y., Wahid, H., Khumran, A., and Kaka, A. (2015). Effect of honey supplementation into Tris Extender on Cryopreservation of Bull Spermatozoa. *Malaysian Journal of Animal Science*, 18(2), 47-54.
- Yimer, N., Noraisyah, A., Rosnina, Y., Wahid, H., Sarsaifi, K., and Hafizal, A. (2014). Comparison of cryopreservative effect of different levels of omega-3 egg yolk in citrate extender on the quality of goat spermatozoa. *Pakistan Veterinary Journal*, 34(3), 347-350.
- Yokozawa, T., Satoh, A., and Cho, E. J. (2004). Ginsenoside- Rd attenuates oxidative damage related to aging in senescence- accelerated mice. *Journal of Pharmacy and Pharmacology*, 56(1), 107-113.
- Yokozawa, T., Wu Liu, Z., and Dong, E. (1998). A study of ginsenoside-Rd in a renal ischemia-reperfusion model. *Nephron*, 78(2), 201-206.
- Yoshida, M. (2000). Conservation of sperms: current status and new trends. *Animal Reproduction Science*, 60, 349-355.
- Yudin, A. I., Generao, S. E., Tollner, T. L., Treece, C. A., Overstreet, J. W., and Cherr, G. N. (2005). Beta-defensin 126 on the cell surface protects sperm from immunorecognition and binding of anti-sperm antibodies. *Biology of Reproduction*, 73(6), 1243-1252.
- Yun, S. J., Bae, G.-S., Park, J. H., Song, T. H., Choi, A., Ryu, B.-Y., Pang, M-G., Kim, E. J., Yoon, M., and Chang, M. B. (2016). Antioxidant effects of cultured wild ginseng root extracts on the male reproductive function of boars and guinea pigs. *Animal Reproduction Science*, 170, 51-60.
- Yun, T.-K. (2001). *Panax ginseng*—a non-organ-specific cancer preventive? *The Lancet Oncology*, 2(1), 49-55.
- Yusuf, H., Mustofa, M., Susidarti, R. A., Asih, P. B. S., and Suryawati, S. (2013). A new quassinoid of four isolated compounds from extract *Eurycoma longifolia* Jack roots and their in-vitro antimalarial activity. *International Journal of Research in Pharmaceutical and Biomedical Science*, 4(3), 728-734.

Zanoli, P., Zavatti, M., Montanari, C., and Baraldi, M. (2009). Influence of *Eurycoma longifolia* on the copulatory activity of sexually sluggish and impotent male rats. *Journal of Ethnopharmacology*, 126(2), 308-313.

Zheng, G.-q., Cheng, W., Wang, Y., Wang, X.-m., Zhao, S.-z., Zhou, Y., Liu, S.-J., and Wang, X.-t. (2011). Ginseng total saponins enhance neurogenesis after focal cerebral ischemia. *Journal of Ethnopharmacology*, 133(2), 724-728.

Zhmakin, A. I. (2008). *Fundamentals of Cryobiology*: Springer Berlin Heidelberg.

