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Aldehydes gas ozonation monitoring: Interest of SIFT/MS *versus* GC/FID

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HIGHLIGHTS

- Ozone interferes on aldehyde analysis by GC/FID with silicone-based column.
- Aldehyde-ozone reaction in GC system could lead to a process misinterpretation.
- SIFT/MS is a reliable technique to monitor VOCs in presence of ozone.

GRAPHICAL ABSTRACT
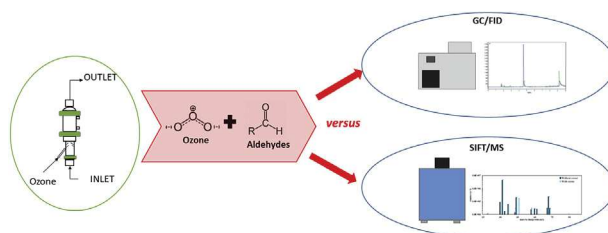
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ABSTRACT

Two analytical techniques — online Gas Chromatography coupled with Flame Ionization Detector (often used method for VOCs monitoring) *versus* Selected Ion Flow Tube coupled with Mass Spectrometry (a more recent technique based on direct mass spectrometry) — were compared in association to an ozone-based gas treatment. Selecting aldehydes as the representative VOCs, their concentrations were monitored during ozonation experiments by both techniques in parallel. Contradictory results were obtained in the presence of ozone. Aldehydes were up to 90% removed due to a reaction with ozone according to GC/FID analysis, whereas with SIFT/MS, aldehydes concentration remained at the same level during the experiments regardless of the ozone presence. In addition, it was demonstrated that the apparent aldehydes removal was affected by GC injector temperature, varying from 90% (when it was at 250 °C) to 60% (at 100 °C). Meanwhile, even when the ozonation reactor was heated to 100 °C, no aldehydes conversion was evidenced by SIFT/MS, suggesting that the GC injector temperature was not the only interference-causing parameter. The ozone-aldehyde reaction is probably catalyzed by some material of GC injector and/or column. An ozone-GC interference was therefore confirmed, making unsuitable the use of GC/FID with silicone stationary phase to monitor aldehydes in presence of high concentrations of ozone (at least 50 ppmv). On the other hand, SIFT/MS was validated as a reliable technique, which can be employed in order to measure VOCs concentrations in ozonation processes.

1. Introduction

Environmental chemistry field pays particular attention to volatile organic compounds (VOCs). Emitted from variety of sources,

such as motor vehicles and chemical plants, these carbon-containing molecules play an important role in atmospheric photochemical reactions, in aerosol formation and some are harmful to human health (Parmar and Rao, 2009). In addition, some oxygenated volatile organic compounds (OVOCs) — such as aldehydes — are often associated to odor nuisance (Anet et al., 2013; Fang et al., 2012; Kolar and Kastner, 2010).

Among several treatments to reduce the environmental impact caused by VOCs, oxidation processes are of major concern. They are destructive processes, indicated for low pollutant concentrations and are often based on radical mechanisms. Plasma treatment, photocatalysis, catalytic oxidation and oxidative wet scrubbers are some examples of oxidation technologies (Martinez et al., 2014; Parmar and Rao, 2009; Thevenet et al., 2014; Vega et al., 2014). Thanks to its high oxidizing potential (Oyama, 2000), ozone (O_3) has been applied as oxidant in many VOCs abatement studies, such as: (*i*) ozonation of slurries (Bildsoe et al., 2012; Liu et al., 2011); (*ii*) chemical absorption/wet scrubber coupled with ozone as advanced oxidation process (Domeno et al., 2010; Kerc and Olmez, 2010; Vega et al., 2014) (*iii*) adsorption with or without regeneration (Brosillon et al., 2001; Monneyron et al., 2007); (*iv*) catalytic processes (Brodu et al., 2012; Kastner et al., 2008; Kolar and Kastner, 2010); (*v*) non-thermal plasma — which produces ozone as a by-product (Abou Saoud et al., 2018; Roland et al., 2005) and (*vi*) homogeneous gas ozonation (Zhang and Pagilla, 2013).

In order to correctly evaluate a VOCs abatement, the analytical technique must be carefully selected. It is even more a critical choice in case of complex systems, such as those in which ozone is present. VOCs monitoring/detection has mainly been carried out by gas chromatography (GC) (Aragón et al., 2000; Dewulf et al., 2002; Helmig, 1999; Woolfenden, 2010), including VOCs abatement studies that employ ozone (Abou Saoud et al., 2018; Bildsoe et al., 2012; Domeno et al., 2010; Huang et al., 2016; Kolar and Kastner, 2010; Li et al., 2018; Vega et al., 2014). Associating a chromatography column type with a GC detector — such as flame ionization (FID), sulfur chemiluminescence (SCD), mass spectrometry (MS), flame photometric (FPD) —, GC shows a high adaptability, making possible the detection of all classes of VOCs (sulfides, oxygenates, hydrocarbons, aromatics and nitriles) (Aragón et al., 2000; Dewulf et al., 2002). GC analysis can be carried out in two different sampling configurations: direct system, when the analytical system is connected online with the gas matrix by a sampling loop (Dewulf et al., 2002) or indirect system, by syringe/bag sampling or by a pre-concentration step based on thermally desorbable solid sorbents (Woolfenden, 2010). Besides the use as sampling method, the pre-concentration step, such as cryogenic trapping/cold trap and thermodesorption coupled with GC, have also been applied in order to analyze low pollutant concentrations or to lower detection limits (Pal and Kim, 2008; Tuduri et al., 2001; Woolfenden, 2010). Carbon or polymer-containing sorbent tubes — like GCB, Tenax GC, Tenax TA, Carbopack B, Carbopack X — and solid phase micro-extraction (SPME) with polydimethylsiloxane (PDMS), carbon molecular sieve (Carboxen), divinylbenzene (DVB) fibers are examples of pre-concentration techniques (Lee et al., 2006; Nicolas et al., 2007; Pal and Kim, 2008; Tuduri et al., 2001). In case of OVOC analysis — more specifically of carbonyl compounds, such as aldehydes and ketones —, derivatization techniques can also be used as an alternative. The most common derivatization agent is 2,4-dinitrophenylhydrazine (DNPH) cartridges. After reaction with carbonyl, DNPH generates hydrazones, which can be analyzed by high pressure liquid chromatography coupled with UV detector. Derivatization agent can also be impregnated on SPME fiber. For example, O-(2,3,4,5,6-pentafluorobenzyl)hydroxylamine hydrochloride (PFBHA) forms volatile oxime derivatives which can be analyzed by GC/MS (Bourdin and Desauziers, 2014; Szulejko and Kim, 2015; Zhu et al., 2015).

Nonetheless, ozone artifacts generation has been reported in literature when the use of sorbent tubes, SPME fibers or derivatization agents is associated to the presence of ozone in gas matrix (Kahnt et al., 2011; Lee et al., 2006; McClenny et al., 2001; Nicolas et al., 2007; Szulejko and Kim, 2015; Uchiyama et al., 2012). Ozone reacts with sorbent material (Tenax TA, Tenax GR, Carbopack B, GCB) — promoting its chemical decomposition — and/or reacts with the previously adsorbed analytes, causing substantial loss or even gain of VOC concentration (Lee et al., 2006; McClenny et al., 2001; Nicolas et al., 2007). In order to reduce the interference, a previous ozone removal step is indicated, applying scrubbers or denuders such as potassium iodide (KI) filters and 1,2-bis-(4-pyridyl) ethylene (BPE) cartridges. However, these materials can also adsorb VOCs, and so, preliminary tests are mandatory (Lee et al., 2006; Nicolas et al., 2007). The negative ozone interference due to artifacts generation on VOC monitoring by indirect GC system (using pre-concentration or sampling devices) has been well studied and highlighted in the literature. However, a potential ozone disturbance on VOC analysis by direct GC system has not been discussed, as far we know. Only one study (Klasson et al., 2003) has indicated an ozone reactivity with one specific type of GC column and a positive ozone interference on CO_2 analysis. This previous study has proposed to measure ozone — in an extremely high concentration range (1400−32600 ppmv of O_3) — by GC associated to thermal conductivity detector (TCD) through quantification of CO_2 generated from the reaction of ozone and the column coating (porous divinylbenzene homopolymer GS-Q column).

Techniques based on chemical ionization coupled with mass spectrometry (ICMS), such as proton transfer reaction (PTR/MS) and selected ion flow tube (SIFT/MS), have emerged as alternatives to GC methods. They are direct mass spectrometry techniques, in which no separation step is necessary, enabling instantaneous quantification of most volatile compounds (Majchrzak et al., 2018; Perraud et al., 2016; Smith and Spanel, 2005; Vitola Pasetto et al., 2019; Volckaert et al., 2016). Particularly, SIFT/MS is able to carry out real-time analysis, even in humid air samples, and can also separate isobaric compounds at wide concentration ranges (from pptv to ppmv), showing low detection limits and high sensitivity (Olivares et al., 2011; Smith and Spanel, 2005). As chemical ionization techniques are softer and therefore more selective processes (compared to electron impact usually applied in mass spectrometry), SIFT/MS is used to analyze complex gas mixtures composed by several VOCs (Huffel et al., 2012; Noseda et al., 2010; Olivares et al., 2011; Prince et al., 2010). Moreover, SIFT/MS is also able to monitor inorganic compounds, such as ozone (Hera et al., 2017; Williams et al., 2002).

In this work, two analytical techniques — GC/FID and SIFT/MS — were compared in association to a gas ozonation process applied as an odor treatment. Considering aldehydes as VOCs representative odorous compounds, both analytical methods were installed in parallel at the outlet of an ozonation reactor to monitor aldehyde concentration in high ozone content (compared to ppbv level of standard tropospheric ozone concentration (Sicard et al., 2018)). In order to avoid a potential ozone interference caused by sorbent material or ozone scrubber, we have selected operating conditions in which pre-concentration step was not necessary, and thus, a direct GC sampling configuration was adopted.

2. Material and methods

2.1. Chemicals

Propanal (PA) and butanal (BA) have been selected as the

representative VOCs because they are common pollutants with low odor threshold limits (1 ppbv for PA (Nagata, 2003) and 0.5 ppbv for BA (Blazy et al., 2015)), often detected in gas emitted by odor sensitive sites (Anet et al., 2013; Kolar and Kastner, 2010). They were purchased from Sigma-Aldrich Inc. (USA), with ≥97% (PA) and ≥99% (BA) of purity.

Methanol, used as solvent for analytical calibration by liquid standard injection, was purchased from Panreac Química SLU (Spain) with ≥99.9% of purity.

2.2. Experimental set-up

Fig. 1 shows an overview of the experimental set-up that is composed of four sections: ozone generation; gaseous effluent generation; ozonation reactor and analytical methods.

In the ozone generation section, the ozone generator (HTU500 AZCO Industries Limited, Canada) was fed with dry air (dew point equal to −40 °C at 101.3 kPa), which gas flow was controlled by a mass flowmeter (SLA 5850S-B Brooks Instruments, USA).

In the gaseous effluent generation section, the aldehyde was injected and controlled via a syringe pump (PH, 2000Harvard Apparatus, USA) and using a 10 mL (SGE, Australia) or a 250 μL glass gastight syringe (Hamilton, USA). The aldehydes were vaporized by the dry air stream, which temperature and relative humidity were measured by a transmitter (HMT333, Vaisala, Finland) and gas flow controlled by a mass flowmeter (SLA 5850S-B Brooks Instruments, USA).

In order to study the reaction at higher temperatures (from 25 °C to 100 °C), a heated circulating oil bath (Model 1160S, VWR, USA) composed of a stainless steel smooth-coil immersed in a synthetic thermoliquid (Ultra 350, Lauda, Germany) bath was installed at the air stream before the vaporization system.

In the ozonation reactor section, the gas phase reaction was performed in a continuous flow, using a jacketed glass tubular reactor (250 mL). The reactor was kept at atmospheric pressure during all experiments (101.3 kPa) and heated using the synthetic thermoliquid from the heated circulating oil bath.

In the analytical methods section, ozone concentrations were measured by a UV analyzer (BMT 964, BMT MESSTECHNIK GMBH, Germany) directly after its production by the generator and by SIFT/MS (Voice 200*ultra*, Syft Technologies Ltd, New Zealand) after dilution at the ozonation reactor. The aldehydes were measured using GC/FID (Varian 3800, USA) and SIFT/MS, both analytical equipment being installed in parallel and directly to the reactor outlet in an online system.

All ozonation experiments were carried out in two steps: (*i*) stabilization step (only BA or PA and dry air were presented, *i.e.* with ozone generator off), during which the aldehyde concentration at the inlet of the reactor was measured (called [aldehyde]-without ozone) and (*ii*) ozonation step (under stable aldehydes-dry air matrix, the ozone generator was switched on and ozone was presented at the system), during which the aldehyde concentration at the outlet of the reactor was monitored (called [aldehyde]with ozone).

2.3. Analytical methods

2.3.1. GC/FID

The compounds were separated on a non-polar (5% phenyl) methylpolysiloxane Varian CP-SIL8 capillary column (30 m, 320 μm, 1 μm), using helium as carrier gas at a flow rate of 2 mL min⁻¹. The 1000 μL sample loop was kept at 50 °C and 101.3 kPa. The injector temperature was initially set at 250 °C using a deactivated and inert liner with volume equal to 0.9 mL (5190-2293 model, Agilent Technologies, Inc., USA) and with a split flow ratio of 20:1 (leading to a residence time in the injector equals to 1.3 s). The oven temperature was kept at 40 °C for 6 min, ramped to 200 °C at 25 °C min⁻¹ and held at 200 °C for 2 min, which resulted in a analysis time equals to 15 min. The FID detector temperature was

Fig. 1. Experimental set-up.
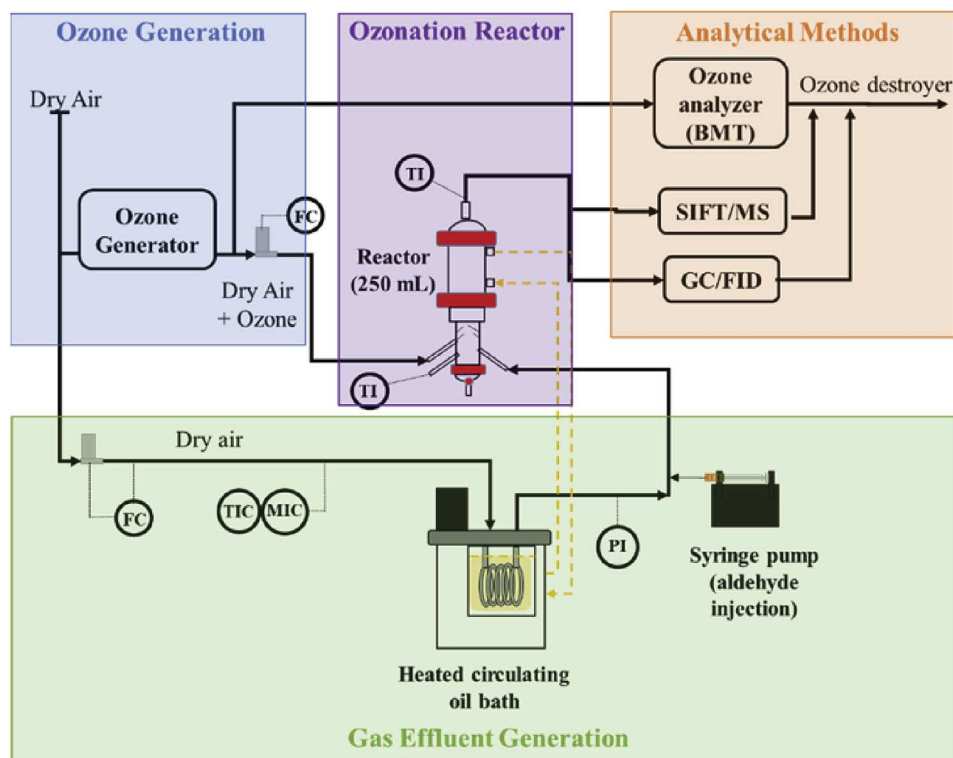
maintained at 250 °C, with gas flows equal to 25 mL min^{-1} of He, 30 mL min^{-1} of H$_2$ and 300 mL min^{-1} of air.

2.3.2. SIFT/MS

In SIFT/MS, the analyte (neutral compound) can be chemically ionized by different precursor ions: three positive ions (H$_3$O$^+$, NO$^+$, O$_2^+$) and five negative ions (NO$_2^-$, NO$_3^-$, O$_2^-$, HO$^-$, O$^-$). These precursor ions were produced by microwave discharge and then selected by a first quadrupole mass filter so that only one precursor at a time was injected to the reaction chamber (flow tube) by a nitrogen flow (180 NmL min^{-1}) as carrier gas. The sample was introduced to the flow tube (kept at 115 °C and 0.07 kPa) by a calibrated capillary at 20 NmL min^{-1}. Even when several VOCs are present in sample gas, each compound reacts with a judiciously selected precursor ion and generates product ions with specific mass-to-charge ratios (*m/z*) that are quantified by a second quadrupole mass spectrometer (Michel et al., 2005; Smith and Spanel, 2005).

As in classical mass spectrometry devices, SIFT/MS operates in two modes: multiple ion monitoring (MIM) and full mass scans (FS). During FS mode − for each selected precursor ion − the second quadrupole spectrometer scans over a mass range covering from 15 to 250 *m/z*, calculating a count rate (signal intensity in s^{-1}) for each unit of *m/z*. Along with information about sample composition, a semi-quantitative analysis can also be obtained in FS mode if kinetic parameters are known. In MIM mode, only the precursor and characteristic product ions of the target compounds are monitored, following their count rates over a time interval (Olivares et al., 2011; Smith and Spanel, 2005).

In SIFT/MS, the analyte concentration is calculated by Equation (1) based on the rate coefficient (k) of the reaction between the neutral compound and the precursor ion, the ratio between product ion count rate (P) and precursor ion count rate (I), and the reaction time in the flow tube (t) (Guimbaud et al., 2007; Smith and Spanel, 2005), which was set at 5 ms in the Syft model used for this project. During sampling, the analyte present in the sample ([A]$_{sample}$) is diluted by the carrier gas in the flow tube, whose analyte concentration ([A]$_{ft}$) depends on the operating conditions of the flow tube − temperature (T$_{ft}$), pressure (P$_{ft}$), sample flow (φ_s), carrier gas flow (φ_c) − and on the Boltzmann constant (k$_b$), as described by Equation (2) (Smith and Spanel, 2005). At low analyte concentrations, Equation (1) can be approximated to a linear correlation between [A]$_{ft}$ and P/I ratio (Equation (3)), if the limit of k [A]$_{ft}$t approaching zero is considered in the exponential expression

(Smith and Spanel, 2005). This linear approximation is only valid when P/I is less than 0.4, *i.e.* the product ion count rate (P) does not represent more than 40% of its precursor count rate (I) and it only describes primary ionization reactions (without clusters formation). When more than one product ion is generated, P represents the sum of all product ion count rates (Smith and Spanel, 2005).

$$\frac{P}{I} = \frac{\left(1 - e^{-k[A]_{ft}t}\right)}{e^{-k[A]_{ft}t}} \tag{1}$$

$$[A]_{sample} = [A]_{ft} \frac{T_{ft}k_b}{P_{ft}} \frac{(\varphi_s + \varphi_c)}{\varphi_s} \tag{2}$$

$$\frac{P}{I} = k[A]_{ft}t \tag{3}$$

Based on the reactional mechanism proposed by Martinez (1982) and Voukides et al. (2009), carboxylic acids are potential by-products of aldehydes gas ozonation. Hence, butanoic acid was also monitored during the ozonation experiments along with BA and O$_3$. As shown in Table 1, Butanoic acid and BA are ionized by all positive precursor ions (H$_3$O$^+$, NO$^+$ and O$_2^+$) (Michalcikova and Spanel, 2014; Spanel et al., 2002, 1997; Spanel and Smith, 1998). Ozone is ionized by three negative precursor ions (NO$_2^-$, O$_2^-$ and HO$^-$) (Williams et al., 2002) and NO$_2^-$ is the most indicated precursor ion for high concentrations (until 100 ppmv of ozone). In order to avoid overlapping of product ions, butanoic acid concentrations were quantified by NO$^+$ spectra and a product ion with a *m/z* ratio of 118, whereas butanal was monitored by product ions with *m/z* ratios equal to 44 and 72 in O$_2^+$ spectra. Ozone was determined with a *m/z* ratio of −62 in NO$_2^-$ spectra.

2.3.3. Method performance

In GC/FID, the limit of detection (LOD) and quantification (LOQ) were calculated as three and ten times the background level, respectively, resulting in 0.02 ng for LOD and 0.07 ng for LOQ. In concentration units, LOQ obtained for BA was 0.5 ppmv. The gas calibration *via* syringe pump was performed with 5 levels of BA concentrations, between LOQ and 3200 times the LOQ, with a coefficient of determination (R^2) equal to 0.9962. The repeatability was of 15% for concentrations up to 100 times the LOQ and equal to 5% for the rest of the domain. The gas calibration was validated by direct injection of liquid standards using methanol as solvent. The difference between both calibration methodologies was of less than

Table 1
Product ions of the reactions between BA, O$_3$ and butanoic acid with H$_3$O$^+$, NO$^+$, O$_2^+$ and NO$_2^-$, with their respective rate coefficients and branching ratios in SIFT/MS.

Precursor ion	Compound	Butanal	Ozone	Butanoic acid
	Molar mass (g/mol)	72	48	88
H$_3$O$^+$	k[a]	3.8[c]	−	2.9[d]
	Product ion [*m/z*] (BR[b])	C$_4$H$_9$O$^+$ [73] (95%)[c] C$_4$H$_7^+$ [55] (5%)[c]		C$_4$H$_9$O$_2^+$ [89] (91%)[d] C$_4$H$_7$O$^+$ [71] (9%)[d]
NO$^+$	k[a]	3.1	−	1.9[d]
	Product ion [*m/z*] (BR[b])	C$_4$H$_7$O$^+$ [71] (100%)[c]		NO$^+$.C$_4$H$_8$O$_2$ [118] (77%)[d] C$_4$H$_7$O$^+$ [71] (23%)[d]
O$_2^+$	k[a]	2.8	−	2.1[d]
	Product ion [*m/z*] (BR[b])	C$_2$H$_4$O$^+$ [44] (48%)[c] C$_4$H$_8$O$^+$ [72] (52%)[c]		C$_2$H$_4$O$_2^+$ [60] (78%)[d] C$_3$H$_5$O$_2^+$ [73] (17%)[d] C$_4$H$_8$O$_2^+$ [88] (5%)[d]
NO$_2^-$	k[a]	−	0.17	−
	Product ion [*m/z*] (BR[b])		NO$_3^-$ [-62] (100%)	

[a] Rate coefficient given in units of 10^{-9} cm^{3}molecule^{-1}s^{-1}.
[b] Branching ratio (BR) represents the product ion distribution (appearance percentage) when more than one precursor is generated.
[c] Kinetic parameters from Spanel et al. (2002).
[d] Kinetic parameters from Michalcíková and Spanel (2014).

6.5% for BA.

In SIFT/MS, the linearity is valid when the precursor ion is in excess compared to product ions. In order to verify the linearity range of SIFT/MS measurements for each selected precursor ion, a calibration procedure was carried out varying BA concentration from 0.15 ppmv to 15 ppmv and ozone from 1 ppmv to 100 ppmv, both in 4 levels. The coefficient of determination (R^2) of P/I ratio *versus* BA concentration at the flow tube was equal to 0.9852 (where P was the sum of product ions O_2^+ [44] and O_2^+ [72] and I was O_2^+ [32]), whereas for O_3 R^2 was equal to 0.9527 (considering P as NO_2^- [-62] and I as NO_2^- [-46]). From the linear regression of P/I ratio *versus* $[A]_{ft}$, it was also possible to obtain k, which was determined dividing the slope by the reaction time in the flow tube. For BA, k_{NO+} and k_{O2+} obtained by this procedure were equal to $(3.1 \pm 0.9) \times 10^{-9}$ cm^3 molecule^{-1} s^{-1} and $(2.8 \pm 0.8) \times 10^{-9}$ cm^3 molecule^{-1} s^{-1} respectively, which are similar to those reported by Spanel et al. (2002, 1997). In the case of ozone, k_{NO2-} was equal to $(1.7 \pm 0.4) \times 10^{-10}$ cm^3molecule^{-1} s^{-1}, which are also similar to previously reported values (Williams et al., 2002). In SIFT/MS, LOD was defined as three times the standard deviation of the mean in the background measurements, which were calculated by blank spectra in absence of BA and O_3, according to the expression proposed by Milligan et al. (2007). LOD depends on the mean background count rate of the product ion at specific *m/z* ratio; on the time of measurement and on the sensitivity of SIFT/MS device (which represents how many product ions at the specific *m/z* ratio were produced for a given concentration of analyte) (Milligan et al., 2007; Ross, 2008). Considering 100 s of time analysis, the LOD for BA was 0.3 ppbv by O_2^+ [44] and O_2^+ [72] and 0.2 ppbv for NO^+ [71]. For ozone, LOD was equal to 40 ppbv by NO_2^- for 60 s of time analysis [-62].

In both analytical techniques, blank chromatograms and spectra were continuously obtained in order to verify the presence of pollutants in dry air and the absence of BA and O_3 in the system before the experiments. The parameters associated to analytical performance of PA are reported in the Support *Information*.

The uncertainties of ozone concentration measured by UV analyzer were calculated by the method of error propagation, considering the standard deviation of a measurement time of 15 min (data recorded each 30 s) and uncertainty of the analytical system (estimated at 25% for 100 ppmv of O_3 and <1% for ozone concentration superior to 4000 ppmv). The uncertainties from the SIFT/MS quantification were obtained from LabSyft$^®$ Data Analysis and were equal to the standard deviation of the mean concentration for a measurement time of 300 s), whereas those from GC/FID were calculated by the standard deviation of at least three chromatograms.

3. Results and discussion

3.1. Ozonation of aldehydes

Preliminary experiments were carried out to verify if SIFT/MS and GC/FID would result to similar removal efficiencies of aldehydes. Initially, ozonation of BA was conducted considering a reactor temperature ($T_{reactor}$) equal to 25 °C, relative humidity in the reactor (RH) at 0.1%, a residence time in the reactor (t_{RES}) of 2 s and ozone concentration at the reactor inlet equal to 69 ± 4 ppmv ($[O_3]_{in}$). These operating conditions were compatible with those applied by Kolar and Kastner (2010), who have reported a removal efficiency of 80% through homogeneous reaction of aldehydes with ozone.

With a BA inlet concentration set point at 5 ppmv, *i.e.* when ozone was not fed into the reactor, concentrations measured by GC/FID and by SIFT/MS in absence of ozone ($[BA]_{without\ ozone}$) were respectively 5.5 ± 0.3 ppmv and 5.7 ± 0.1 ppmv, according to Fig. 2. However, when ozone was sent into the reactor, BA concentration in presence of ozone ($[BA]_{with\ ozone}$) measured by GC/FID was 0.6 ± 0.1 ppmv, whereas by SIFT/MS was 5.3 ± 0.1 ppmv. GC/FID results suggest a removal efficiency of BA up to 90%, which did not tally with SIFT/MS measurements. Similar conflicting results were obtained for PA ozonation, as shown in *Figure SI-1* in Support *Information*.

3.2. Increasing of ozonation reactor temperature

Due to the contradictory results of outlet aldehyde concentrations obtained by GC/FID and SIFT/MS devices when $T_{reactor}$ was 25 °C, complementary experiments were conducted only focusing on ozonation of BA, since the similarity of BA and PA behaviors have already been confirmed in the preliminary tests. By varying the $T_{reactor}$ from 25 °C to 100 °C, two aspects were simultaneously investigated: (*i*) the repeatability of conflicting results and (*ii*) the impact of reactor conditions on removal efficiency of BA, *i.e.* if ozone-aldehydes reaction (seen by GC/FID) really happens in the gas ozonation reactor.

BA concentrations were monitored by SIFT/MS and by GC/FID simultaneously and in the same operating conditions applied previously ($[BA]_{without\ ozone}$ set point at 5 ppmv, RH at 0.1% and t_{RES} equal to 2 s). As it is shown in Fig. 2, in addition to 25 °C, 4 more levels of $T_{reactor}$ were investigated (45 °C; 65 °C; 85 °C and 100 °C) and with $[O_3]_{in}$ equal to 58 ± 6 ppmv in average. The ozonation of

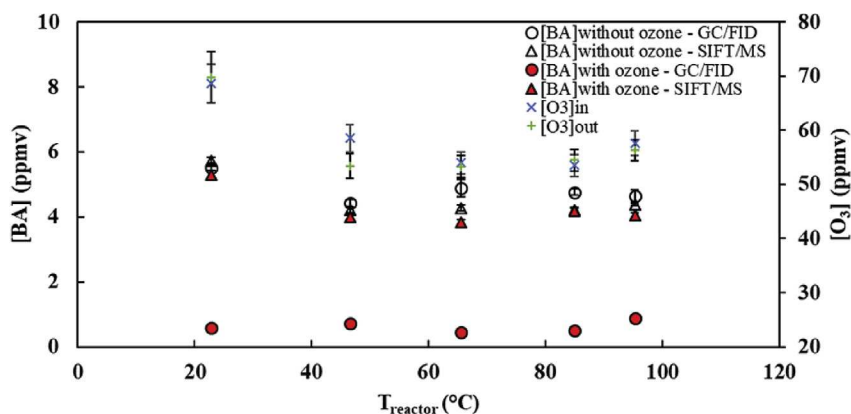**Fig. 2.** BA concentration measured by GC/FID and by SIFT/MS and ozone concentration measured by UV analyzer ($[O_3]_{in}$) and by SIFT/MS ($[O_3]_{out}$) for a range of reactor temperature with the same ozonation conditions: dry atmosphere (RH = 0.1%) and t_{RES} = 2 s.

BA has shown to be independent of the reactor temperature, since BA concentration during ozonation step ([BA]$_{with\ ozone}$) stayed at a constant level even with temperature increase. In absence of ozone, GC/FID and SIFT/MS measured similar BA concentrations ([BA]$_{without\ ozone}$ equal to 4.6 ± 0.7 ppmv). However, when the reactor was fed by ozone, the measurements were discordant whatever the temperature considered. According to GC/FID results, [BA]$_{with\ ozone}$ was 0.6 ± 0.2 ppmv for all reactor temperatures, while [BA]$_{with\ ozone}$ measured by SIFT/MS was 4.3 ± 0.6 ppmv. No variation between ozone concentration at the inlet ([O$_3$]$_{in}$) and outlet of the reactor ([O$_3$]$_{out}$) could be detected, due to its large excess (Fig. 2).

In Fig. 3, GC/FID and SIFT/MS analyses are compared at the extremes of T$_{reactor}$ range: 25 °C (Fig. 3A and B) and 100 °C (Fig. 3C and D). In presence of 58 ppmv of ozone, BA peak intensity (retention time of 4.6 min) in the GC chromatogram (Fig. 3A and C) decreased compared to the chromatogram profile without ozone, suggesting that around 85% of BA was removed from gas matrix through reaction with ozone. In addition to BA peak reducing, new chromatographic peaks also appeared in the chromatogram profile in presence of ozone for both reactor temperatures, which may indicate by-products generation. The biggest by-product peak (retention time of 9.2 min) was identified as butanoic acid by injection of liquid standards.

Regarding the SIFT/MS results, both characteristic product ions of BA at m/z 44 and 72 in O$_2^+$ spectra have shown the same count rate in presence or absence of ozone (Fig. 3B and D), which is also valid for precursor ion (O$_2^+$ [32]) and product ion of butanoic acid (O$_2^+$ [60] and O$_2^+$ [73]). From O$_2^+$ spectra, the ozone occurrence did not dramatically modify m/z and count rate distributions of BA-air matrix, except for m/z 46, which only appeared when ozone was present for both T$_{reactor}$. Peaks at m/z 46 and m/z 64 have also been

detected in presence of ozone in H$_3$O$^+$ spectra (*Figure SI-2A and Figure SI-3A in Support Information*), probably due to NO$_2$ production in the ozone generator, since NO$_2^+$ (m/z 46) is its characteristic product ion. The non-variability of m/z and count rate distribution, and consequently, of chemical composition in BA-air matrix was also confirmed by H$_3$O$^+$ spectra, NO$^+$ spectra and NO$_2^-$ spectra for T$_{reactor}$ of 25 °C and of 100 °C (in Figure SI-2 and Figure SI-3 in Support Information, respectively).

Therefore, four aspects can be highlighted regarding the conflicting results: (*i*) the decrease of aldehyde concentrations and by-products generation according to GC/FID; (*ii*) the non-removal of aldehydes and no by-product generation based on SIFT/MS measurements; (*iii*) the repeatability of GC/FID and SIFT/MS results in presence of ozone and (*iv*) the stable apparent removal of BA for T$_{reactor}$ varying from 25 °C to 100 °C. The association of these aspects suggests that the ozone-aldehydes reaction − previously assumed to be located in the reactor − occurs elsewhere between the GC inlet and the GC system.

3.3. Varying GC parameters

Some GC parameters were tested to verify a probable influence on aldehyde analysis, *i.e.* on the apparent aldehyde removal. As an aldehyde-ozone reaction seems to happen, the GC injector temperature (T$_{GC\ inj.}$) − set initially at 250 °C − showed the highest potential to be the interference-causing parameter. In addition, residence time in the injector was 1.3 s, based on the GC operating conditions (split ratio, column gas flow and liner volume).

The removal efficiency of BA was monitored when T$_{GC\ inj.}$ was varied from 250 °C to 100 °C, as shown in Fig. 4. BA concentrations were measured by GC/FID with operating conditions kept constant ([BA]$_{without\ ozone}$ at 22 ppmv; T$_{reactor}$ at 25 °C; RH at 0.1% and [O$_3$]$_{in}$
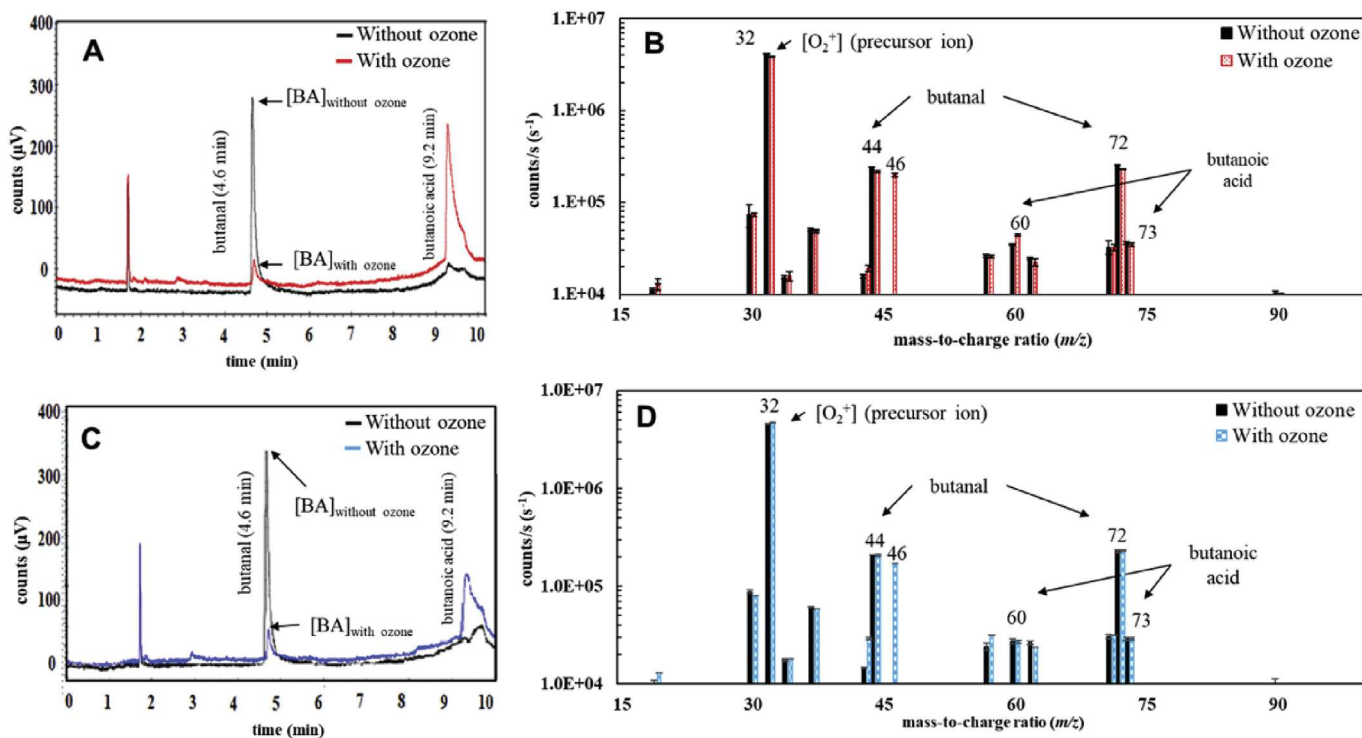

Fig. 3. A) GC/FID chromatogram of BA detection in absence of ozone (black line) and in presence of ozone (red line) with T$_{reactor}$ = 25 °C. **B)** SIFT/MS full mass scan O$_2^+$ spectra of BA-air matrix without ozone (dark bars) and in presence of ozone (red dotted bars) with T$_{reactor}$ = 25 °C. **C)** GC/FID chromatogram of BA detection in absence of ozone (black line) and in presence of ozone (blue line) with T$_{reactor}$ = 100 °C. **D)** SIFT/MS full mass scan O$_2^+$ spectra of BA-air matrix without ozone (dark bars) and in presence of ozone (blue and striped bars) with T$_{reactor}$ = 100 °C. (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)
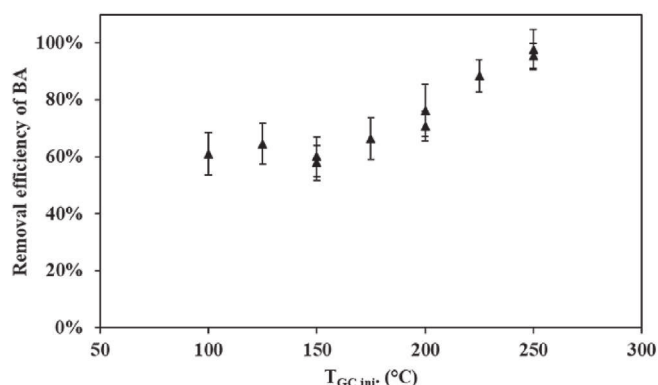
Fig. 4. Removal efficiency of BA (▲) profile according to a range of $T_{GC\ inj.}$ under similar operating conditions in the reactor: $[BA]_{without\ ozone} = 22 \pm 1$ ppmv; $[O_3]_{in} = 163 \pm 15$ ppmv; dry atmosphere (RH = 0.1%); $T_{reactor}$ at 25 °C and $t_{RES} = 2$ s.

at 163 ppmv). The attenuation of removal efficiency with decreasing of GC injector temperature suggests that GC operating conditions indeed affect the BA ozonation.

In the GC chromatograms in Fig. 5A, the overlapped profiles for $T_{GC\ inj.}$ at 250 and 100 °C demonstrate that when ozone was not in the gas matrix, BA detection by GC/FID was not impacted by $T_{GC\ inj.}$ changing, which confirms that no thermal degradation of BA occurred due to high injector temperatures. However, in presence of ozone (63 ± 4 ppmv), the BA peaks in Fig. 5B for $T_{GC\ inj}$ at 250 and 100 °C were no longer overlapped, but they were less intense than those without ozone. BA concentration decreased from 25 ± 2 ppmv to 4.2 ± 0.2 ppmv when $T_{GC\ inj}$ was set at 250 °C, whereas at $T_{GC\ inj}$ of 100 °C, the concentration decrease was smaller (from 25 ± 2 ppmv to 14 ± 1 ppmv).

The gain in BA concentration with $T_{GC\ inj}$ reduction only when ozone was added confirms that BA reacts with ozone somewhere in the GC system, and not in the reactor as it could be wrongly supposed if SIFT/MS analysis was not available.

Nonetheless, $T_{GC\ inj}$ may not be the only GC parameter that causes the interference on aldehydes analysis, since even when $T_{reactor}$ was at 100 °C, no removal efficiency was detected by SIFT/MS whereas it was observed by GC/FID at $T_{GC\ inj}$ equal to 100 °C.

Complementary experiments were conducted to better understand the role of the GC injector (detailed in Figure SI-4 in Support *Information*). It was observed that the removal efficiency of BA measured by GC/FID mainly depended on ozone concentration, but

it showed a light influence of the residence time in the GC injector.

Different models of inlet liner (model Topaz 23301, Restek Corporation, USA) and of GC capillary column (non-polar; 100% PDMS Rtx-1; 60 m, 320 μm, 1 μm) were tested using the same operating conditions ($T_{GC\ inj.}$ at 250 °C; split ratio equal to 20:1; $[BA]_{without\ ozone}$ at 30 ppmv; $[O_3]_{in}$ at 60 ppmv and RH equal to 0.1%), but similar removal efficiencies were achieved compared to the models described in *Analytical Methods*. In addition, BA removal did not change even when a longer capillary column was used, in which the dead time and the retention time of BA (9.15 min) were higher. It suggests that aldehyde-ozone reactions may not be affected by a longer analysis time, and consequently, the reaction does not happen during the passage of the gas through the capillary column. Furthermore − based on Figs. 3 and 5 and Figure SI-1, Support *Information* − the chromatographic peaks related to PA and BA were still present (at a smaller intensity) and narrow and symmetric peaks of the reaction product have appeared. According to Dettmer-Wilde and Engewald (2014), these chromatogram characteristics suggest that the aldehyde-ozone reactivity probably takes place in the injector or at column head, through a fast reaction that is completed before entering into the column.

Therefore, the ozonation of aldehydes in GC may occur due to a catalytic pathway. Some GC material could act as a catalyst, as it was already observed for metal oxides, zeolites and carbon-based materials which can enhance the ozone-oxidation of VOCs (Brodu et al., 2012; Kastner et al., 2008; Masuda et al., 2001; Oyama, 2000). On the surface of these catalysts, the ozone reacts with the active sites (Lewis acid site) to form secondary oxidizing agents (atomic oxygen, peroxide ions, hydroxyl radicals), which subsequently react with the pollutants (Abou Saoud et al., 2019; Brodu et al., 2018; Wang et al., 2018).

As far we know, the ozone decomposition on PDMS stationary phase has not been discussed in the literature. There are only studies that investigate the PDMS decomposition − in form of membranes or microfluidic systems − through reaction with ozone or in contact of non-thermal plasma (Alves dos Santos et al., 2015; Duffy et al., 1998; Fu et al., 2009). The aim of these studies was to explore the changing of material properties, which was not related to catalysis or degradation of VOCs. Nevertheless, they have evidenced the reactivity between ozone and PDMS. Duffy et al. (1998), for example, have demonstrated that the attack of the non-thermal plasma was at the silicon atoms, converting the $-OSi(CH_3)_2O-$ groups on the surface to $-O_nSi(OH)_{4-n}O-$ groups. Another example is the studies conducted by Fu et al. (2009) that have investigated the attack of ozone (generated by UV light) on membranes made by
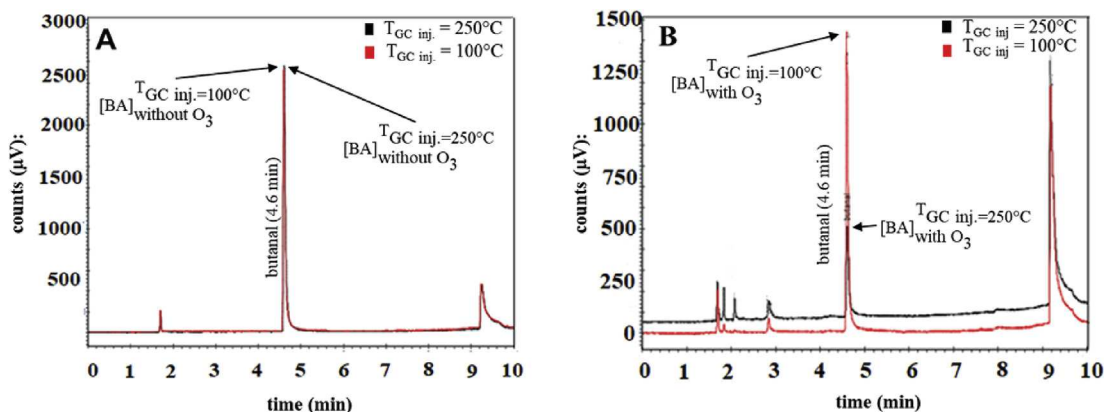

Fig. 5. GC/FID chromatograms of BA detection carried out with $[BA]_{without\ ozone}$ at 25 ± 2 ppmv; $T_{reactor}$ at 25 °C; RH at 0.1% and $[O_3]_{in}$ at 63 ± 4 ppmv. **A)** Chromatogram profiles during stabilization time of BA-dry air matrix (without ozone) when $T_{GC\ inj.}$ was at 250 °C (dark line) and at 100 °C (red line). **B)** Chromatogram profile with ozone when $T_{GC\ inj.}$ was at 250 °C (dark line) and at 100 °C (red line). (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

PDMS. Besides the generation of silanol groups (SiOH) on the polymeric chain − in accordance with Duffy et al. (1998) − the authors have also identified the formation of an inorganic SiO_2 layer over the surface. In addition, the UV/ozone treatment has been applied to remove layer of organic compound present at the surface of PDMS membranes, due to generation of radicals and oxygen atom. Furthermore, these silanol groups exhibit acid properties, which could act as active sites and interact with polar analytes (Dettmer-Wilde and Engewald, 2014) and also decompose ozone, as demonstrated by Monneyron et al. (2003b)in their studies of catalytic ozonation using zeolites.

In summary, we could explain the conversion of aldehyde by a reaction of ozone with the silicone-based material of the GC column, which is enhanced by the high temperature of the GC injector and generates silanol groups. This silanol group could act as active sites and lead, in the presence of ozone, to the formation of radical species that react with the aldehydes.

4. Conclusion

An ozone-GC interference was evidenced in the operating conditions applied in this study (using silicone stationary phase), leading to falsely low aldehyde concentration in presence of ozone. Since GC injector temperature was not the only interference-causing parameter, the ozone-aldehyde reaction is probably catalyzed by some material of GC injector and/or column. In the case when GC/FID must be applied to analyze an ozone-containing gas matrix, a preliminary ozone-removal step is thereby required.

The untrue aldehydes quantification by GC/FID could only be verified by comparison with a second analytical approach − SIFT/MS −, which is based on a different operation principle, and thereby, does not present the same drawbacks as GC. SIFT/MS is the most indicated analytical device to monitor VOCs in presence of ozone, since it has proven to yield reliable results and to be indifferent to ozone disturbance. In addition, SIFT/MS is a faster and more sensitive technique, which is an interesting advantage in the environmental odor context that often faces limitations related to low odor threshold limits.

The major outcome of this study is the warning of applying GC/FID for aldehyde detection in presence of ozone. The use of unsuitable analytical devices could lead to unreliable results, and if this analytical device is associated to chemical processes, it could result in a process misinterpretation. For example, in case of this study, due to the use of GC/FID using silicone stationary phase, aldehyde removal values could be wrongly attributed to gas ozonation treatment.

Acknowledgements

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.chemosphere.2019.06.186.

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